- 1 Global ecological and biogeochemical impacts of pelagic tunicates 2 3 4 Jessica Y. Luo¹, Charles A. Stock¹, Natasha Henschke², John P. Dunne¹, Todd D. O'Brien³ 5 6 ¹NOAA Geophysical Fluid Dynamics Laboratory, 201 Forrestal Rd, Princeton NJ 08540 7 ²School of Biological, Earth and Environmental Sciences, University of New South Wales, 8 Sydney, NSW 2052 9 ³NOAA NMFS Office of Science and Technology, Silver Spring MD, USA 10 11 Manuscript correspondence: Jessica.Luo@noaa.gov 12 13
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- 15 export, global model

16 Abstract

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18 The pelagic tunicates, gelatinous zooplankton that include salps, doliolids, and appendicularians,

- 19 are filter feeding grazers thought to produce a significant amount of particulate organic carbon
- 20 (POC) detritus. However, traditional sampling methods (i.e., nets), have historically
- 21 underestimated their abundance, yielding an overall underappreciation of their global biomass
- 22 and contribution to ocean biogeochemical cycles relative to crustacean zooplankton. As climate
- change is projected to decrease the average plankton size and POC export from traditional
- 24 plankton food webs, the ecological and biogeochemical role of pelagic tunicates may increase;
- 25 yet, pelagic tunicates were not resolved in the previous generation of global earth system climate
- 26 projections. Here we present a global ocean study using a coupled physical-biogeochemical
- model to assess the impact of pelagic tunicates in the pelagic food web and biogeochemical
- 28 cycling. We added two tunicate groups, a large salp/doliolid and a small appendicularian to the
- 29 NOAA-GFDL Carbon, Ocean Biogeochemistry, and Lower Trophics version 2 (COBALTv2)
- 30 model, which was originally formulated to represent carbon flows to crustacean zooplankton.
- The new GZ-COBALT simulation was able to simultaneously satisfy new pelagic tunicate
- 32 biomass constraints and existing ecosystem constraints, including crustacean zooplankton
- 33 observations. The model simulated a global tunicate biomass of 0.10 Pg C, annual production of
- 0.49 Pg C y^{-1} in the top 100 m, and export flux of 0.7 Pg C y $^{-1}$, representing 11% of the total
- 35 export flux past 100 m. Overall export from the euphotic zone remained largely constant, with
- the GZ-COBALT pe-ratio only increasing 5.3% (from 0.112 to 0.118) compared to the
- 37 COBALTv2 control. While the bulk of the tunicate-mediated export production resulted from the
- 38 rerouting of phytoplankton- and mesozooplankton-mediated export, tunicates also shifted the
- 39 overall balance of the upper oceans away from recycling and towards export. Our results suggest
- 40 that pelagic tunicates play important trophic roles in both directly competing with
- 41 microzooplankton and indirectly shunting carbon export away from the microbial loop.

42 1. Introduction

43

44 In recent decades, there has been a growing recognition of the prevalence and ecological 45 importance of gelatinous zooplankton (GZ), which include the cnidarian jellyfish, ctenophores, and pelagic tunicates (Hays et al., 2018; Henschke et al., 2016). While they have been a natural 46 47 component of marine ecosystems extending back to the Cambrian (Hagadorn et al., 2002), their 48 abundance and distributions have been largely overlooked among the public and non-GZ 49 specialists (Condon et al., 2012). This may be due to a combination of the erroneous perception 50 of GZ being "trophic dead-ends" within marine food webs (Hays et al., 2018; Lynam et al., 51 2006; Verity and Smetacek, 1996), as well as systematic biases in sampling leading to overall 52 under-sampling of their biomass. Net-based sampling has been prevalent for over a century, and 53 is very effective for sampling fish and hard-bodied, crustacean zooplankton (Wiebe and 54 Benfield, 2003). Unfortunately, the fragile gelatinous zooplankton are often broken apart in nets, 55 yielding a *ca*. 3 fold underestimation of their abundance and a *ca*. 10 fold (range: 5-15) 56 underestimation of their carbon biomass relative to non-extractive, optical sampling (Remsen et 57 al., 2004). The rise of in-situ plankton imaging systems have resulted in improved estimates of GZ abundance and distribution (e.g., Luo et al., 2014), yielding advances in understanding of 58 59 their food-web interactions and biogeochemical impacts (Greer et al., 2021; Robison, 2005; 60 Smith Jr et al., 2014). Combined with the increase in ecosystem-level studies of GZ (e.g., Stukel 61 et al., 2021) and technological advances revealing the importance of GZ as food for higher 62 trophic levels (Hays et al., 2018), there has been an overall paradigm shift in our understanding 63 of the importance and role of gelatinous zooplankton within marine ecosystems. 64 Amongst zooplankton, GZ are notable for their high clearance rates and boom-and-bust 65 population dynamics, which yield mass mortality events ("jelly-falls") that sink rapidly through the water column (Acuña et al., 2011; Billett et al., 2006; Lebrato et al., 2012; Lucas and 66 67 Dawson, 2014). As a result, models have estimated, based on population densities and allometric scaling of ecological and physiological rates, a large contribution (e.g., 1.6-5.2 Pg C y⁻¹ in Luo et 68 69 al. 2020) of GZ-mediated carbon in the global biological pump, with their relative impact 70 increasing with depth (Lebrato et al., 2019; Luo et al., 2020). However, these studies have been 71 done independently of other biogeochemical and ecological constraints (i.e., "offline" 72 calculations), which may yield unrealistic estimates of GZ contributions to marine ecosystems. 73 Indeed, in a new model that includes cnidarian jellyfish as a plankton functional type 74 (PlankTOM11), global export production exhibited very modest increases (+0.1 Pg C y⁻¹), 75 suggesting that the online inclusion of a jellyfish class does not by itself substantially increase 76 total export production (Wright et al., 2021). Of the three major groups of GZ considered by Luo

et al. (2020), cnidarian jellyfish had the largest standing stock biomass but pelagic tunicates,

despite their much lower biomass, over two times more sinking detritus than cnidarians and
 ctenophores combined.

80 These pelagic tunicates, small filter-feeders including appendicularians, salps, doliolids, 81 and pyrosomes, are less conspicuous than the cnidarians and ctenophores, but are highly 82 significant components of marine ecosystems due to their low trophic position, high clearance 83 rates, and fast sinking fecal pellets (Andersen, 1998; Berline et al., 2011; Henschke et al., 2016; 84 Hopcroft and Roff, 1998). Compared to crustacean mesozooplankton such as copepods which 85 feed at predator to prey size ratios ranging from 5:1 to 100:1 (Hansen et al., 1994), pelagic 86 tunicates can feed at predator to prey size ratios ranging from 10:1 to 10000:1 (Conley et al., 87 2018). Salps pump water through their fine mucous meshes that can filter submicron particles

88 such as bacteria and picoplankton; they are able to sustain the entirety of their energetic demands

89 by grazing on these size classes alone (Sutherland et al., 2010). Using both external and internal

90 filters to feed, appendicularians have the widest range of predator to prev size ratios (exceeding

91 2500:1 to lower than 10:1), and thus can feed on organisms ranging from 0.2-20 µm in size

92 (Conley et al., 2018; Deibel and Lee, 1992; Fernández et al., 2004). The offline Luo et al. (2020)

93 model estimated that, due to these feeding characteristics, these pelagic tunicates consume 94 between 3.8-8.3 Pg C y⁻¹ in prey. Of this consumption, approximately 12-17% were later

95 consumed by higher trophic level predators, and 55-60% became detritus.

96 Global ocean biogeochemical and marine ecosystem models typically represent marine 97 food webs with roughly linear food chains of phytoplankton to zooplankton to (implicit) fish

98 (Kearney et al., 2021). The traditional inclusion of multiple zooplankton groups has been to

99 distinguish between zooplankton size, such as microzooplankton and mesozooplankton, with the

100 latter parameterized using crustacean zooplankton measurements (Aumont et al., 2015;

101 Buitenhuis et al., 2006; Stock and Dunne, 2010; Ward et al., 2012). Even in models with

102 complex food-webs, predator to prey size ratios beyond \sim 50:1 (c.f. Hansen et al., 1994) are

103 rarely considered. As such, current ocean biogeochemical models typically represent a marine

104 ecosystem in which crustaceans dominate zooplankton ecology. While this view represents

105 certain ecosystems well (Pershing and Stamieszkin, 2020), globally, there is a tension between

106 the traditional, crustacean-dominated zooplankton view of marine ecosystems and a shifting

107 paradigm that emphasizes the role of GZ. Unfortunately, GZ-focused offline models are unable

108 to reconcile this tension, as evidenced by high GZ-mediated global ingestion and production

109 rates (Luo et al., 2020) that may not support primary and secondary production rates for

110 crustacean zooplankton consistent with observations. Additionally, offline studies are limited in

111 the capacity to explore factors underlying observed GZ niches, and how GZ impacts emergent food web patterns. These challenges are compounded by stubborn limitations in GZ observations

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which are patchy and inconsistently sampled.

114 In this study, we added two explicit zooplankton functional types that represent 115 thaliaceans (salps, doliolids, pyrosomes) and appendicularians into Carbon, Ocean 116 Biogeochemistry, and Lower Trophics version 2 (COBALTv2; Stock et al., 2020), a global

117 model designed to represent a "traditional" marine ecosystem dominated by crustacean 118 zooplankton. We ask the following four questions:

119 1) Can simulations capture the magnitude and gradients of observed GZ biomass across 120 ocean biomes and along productivity gradients, after accounting for approximately an order of 121 magnitude under-sampling by nets?

122 2) Can simulations reconcile recent evidence for the importance of GZ with established 123 evidence for the prominence of crustacean zooplankton in biogeochemical cycles and the 124 plankton food web?

125 3) How does a simulation of GZ-modulated export that satisfies multiple food web 126 constraints compare with offline estimates?

127 4) What is the net impact of GZ zooplankton on the partitioning of carbon flows between recycling, carbon export and energy flows to higher trophic levels? 128

130 **2. Methods**

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132 As a brief overview of the methods, we begin with a description of plankton food web dynamics 133 within the original COBALTv2 marine ecosystem model (Stock et al., 2014a, 2020), followed by 134 the GZ additions that comprise GZ-COBALT: small and large pelagic tunicates. We first 135 describe the baseline parameterization of GZ-COBALT, and then a few sensitivity experiments 136 that explore parts of the parameter space and particular elements of the tunicate groups that make 137 them distinct from crustacean zooplankton. Next, we detail the physical framework of the model. 138 Finally, we describe the construction of a validation dataset for the two new GZ groups and the 139 identification of an emergent relationship that contrasts gelatinous against crustacean 140 zooplankton. The model is validated against multiple constraints, comprising new and 141 established ecological and biogeochemical datasets.

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143 2.1 COBALT Ecosystem Model

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145 We use the COBALTv2 marine ecosystem model (Stock et al., 2020) as our baseline 146 model configuration, with slight modifications. COBALTv2 is a 33-tracer, intermediate 147 complexity model, representing biogeochemical cycles of carbon, alkalinity, oxygen, nitrogen, 148 phosphorus, iron, silica, calcium carbonate, and lithogenic materials. The food web consists of 149 three phytoplankton and three zooplankton functional types, as well as a free-living heterotrophic 150 bacteria group. Two phytoplankton size classes (small and large) are represented, as well as 151 diazotrophs, parameterized as a large *Trichodesmium* group. The small phytoplankton type 152 includes cyanobacteria and other phytoplankton, up to 10 µm in equivalent spherical diameter 153 (ESD), and the large phytoplankton type represents diatoms and other large phytoplankton from 154 10-100 µm in ESD. The different sized phytoplankton are parametrized to capture size-based 155 contrasts in nutrient uptake, light harvesting, carbon to chlorophyll ratios, and susceptibility to 156 microzooplankton grazing (Edwards et al., 2015, 2012; Hansen et al., 1994; Munk and Riley, 157 1952), such that the small phytoplankton are more successful in the low nutrient, seasonally 158 stable subtropical gyres, and large phytoplankton are more competitive in the highly seasonal, 159 high nutrient oceans (Stock et al., 2014a, 2020). 160 The base configuration of COBALTv2 contains three zooplankton size classes: a

161 microzooplankton and two size classes of crustacean mesozooplankton. Microzooplankton (< 200 µm ESD) include ciliates and heterotrophic nanoflagellates, medium zooplankton (i.e, small 162 163 mesozooplankton; 200-2000 µm ESD) represent small to medium-bodied copepods, and large 164 zooplankton (2 – 20 mm ESD) represent large copepods and krill (Stock et al., 2014a). Predator-165 prey relationships are also largely based on size, with microzooplankton predating on bacteria 166 and small phytoplankton, small mesozooplankton predating on diazotrophs, large phytoplankton, 167 and microzooplankton, and large mesozooplankton predating on diazotrophs, large phytoplankton, and small mesozooplankton (Fig. 1a). Grazing is modeled as a Hollings Type II 168 169 function with density-dependent switching (Stock et al., 2008), with maximum biomass specific 170 grazing rates decreasing with increasing zooplankton size (Hansen et al., 1997). Grazing half-171 saturation constants do not vary between the zooplankton classes, and are tuned to reproduce 172 observed patterns in phytoplankton biomass and turnover rates (Stock and Dunne, 2010). 173 Zooplankton grazing was assimilated at an assimilation efficiency (AE) of 0.7, with the 174 non-assimilated grazing partitioned into dissolved and particulate (detritus) matter, depending on

175 size. Detritus partitioning of non-assimilated matter were: 1/6 for microzooplankton, 2/3 for

176 medium, and 1 for large zooplankton, and the remainder separated between labile (70%), semi-177 labile (20%) and semi-refractory (10%) dissolved matter. Assimilated matter was partitioned into 178 respiration (basal and active) and zooplankton production. Basal respiration is proportional to 179 biomass, whereas active respiration is proportional to ingestion rates. When respiration rates exceed assimilated ingestion, production becomes negative and recontributes to zooplankton 180 181 mortality. The temperature dependence of biological rates (phytoplankton nutrient uptake and growth, zooplankton grazing) is determined by a common Eppley (1972) curve with a Q_{10} factor

- 182 183 of 1.88, representing a near doubling of rates for every 10°C temperature increase.
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186 2.2 Gelatinous Zooplankton in COBALT (GZ-COBALT) 187

188 Following Luo et al. (2020), we introduced two new pelagic tunicate groups into

189 COBALTv2 (Fig. 1b): small tunicates, which represents Appendicularians, and large tunicates,

190 which represents Thaliaceans (i.e., salps and doliolids). Appendicularians are small, free-

191 swimming organisms that produce gelatinous houses for filter-feeding, which are discarded when

192 clogged and re-created multiple times per day. Salps and doliolids are also filter-feeders, but

193 unlike appendicularians, are colonial (though salps and doliolids have solitary life stages), form

- 194 rapidly sinking fecal pellets (Perissinotto and Pakhomov, 1998), and exhibit mass die-offs (jelly-
- 195 falls; Henschke et al., 2013).
- 196





Figure 1. Food web structure of (a) COBALTv2 base model, and (b) GZ-COBALT model. Additional functional 199 groups in GZ-COBALT include a small and large tunicate. Bact = free living bacteria, SP = small phytoplankton, 200 Diazo = diazotrophs, LP = large phytoplankton, SZ = small zooplankton, MZ = medium zooplankton, LZ = large 201 zooplankton, and HP = higher trophic-level predators. Note that within typical zooplankton size categories, small 202 zooplankton would be considered microzooplankton, and medium and large zooplankton would be considered 203 mesozooplankton.

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205 2.2.1 GZ food web structure

206 The pelagic tunicates were inserted into the COBALTv2 model as summarized in Fig. 1. 207 Pelagic tunicates have the widest prey-to-predator size ratios among zooplankton (Conley et al., 208 2018), with appendicularians predating on organisms ranging from 0.04 - 20% of its size,

209 equivalent to $0.2 - 20 \,\mu\text{m}$ in length (Deibel and Lee, 1992; Fernández et al., 2004; Lombard et 210 al., 2011). Small tunicates in the baseline model setting are thus able to consume bacteria, small phytoplankton, large phytoplankton, and microzooplankton. This is consistent with previous 211 212 efforts that included appendicularians in a NPZD-type model (Berline et al., 2011) wherein small tunicates consumed small phytoplankton, which lie near the center of their prev kernel, with high 213 214 preference (100% prey availability), and all others with low preference (25% prey availability). 215 Appendicularians are important prey for many invertebrates and fish, often contributing large 216 proportions of the diets of medusae jellyfish and many larval fish species (Gorsky and Fenaux, 217 1998; Llopiz et al., 2010; Purcell et al., 2005). Thus, small tunicates were predated upon by 218 medium and large zooplankton and higher trophic level predators with high preference (Fig. 1b).

219 While they have relatively large body sizes (e.g., the solitary form of Salpa thompsoni 220 can exceed 10 cm; Dubischar et al., 2006), salps have fine mucous meshes that can filter 221 submicron particles, and have a preference for grazing on small algae such as picoplankton 222 (Sutherland et al., 2010). Salps are able to consume diatoms and bacteria, but with relatively low 223 efficiency (Dadon-Pilosof et al., 2019) and with many diatoms passing through salp guts 224 undigested (Harbison et al., 1986). Additionally, salps have been shown to be dominant grazers 225 on Trichodesmium in certain regions (Post, 2002). While, the prey of doliolids is less studied, 226 Walters et al. (2019) found using genetic approaches that doliolids preferentially consume 227 diatoms (particularly in their early life stages) and ciliates. Thus, in the present model, the large 228 tunicates were able to feed on small phytoplankton and diazotrophs with high preference (100% 229 prey availability), microzooplankton at medium preference (50% prey availability), and bacteria 230 and large phytoplankton at low preference (25% prey availability).

231 Large tunicate predators and parasites were traditionally thought to be primarily 232 sapphirinid copepods and large hyperiid amphipods, including the predatory and parasitic 233 *Phronima* spp., which consume salp tissue and live in their cleared-out barrels (Laval, 1980; 234 Madin and Harbison, 1977; Takahashi et al., 2013). Restricted emphasis on these relatively rare 235 crustacean predators and parasites has yielded the misconception of gelatinous zooplankton, 236 particularly salps, as trophic dead-ends. However, increasing evidence has highlighted the role of 237 thaliaceans as food for fish and other higher trophic levels. Over 200 species of fish, turtles, 238 corals, and echinoderms consume salps, doliolids, and pyrosomes, with many predators filling 239 their guts with thaliaceans during bloom periods (Harbison, 1998; Henschke et al., 2016; 240 Mianzan et al., 2001). Therefore, in our model, large tunicates are predated upon by medium and 241 large zooplankton at low preference (25% prey availability), reflecting the specialized nature of 242 the copepod and amphipod predators relative to the broader crustacean zooplankton population, 243 and by higher trophic level predators at medium preference (50% prev availability).

244 The diets of small and large tunicates are fairly similar in the model, as they are both 245 microphagous generalists. Notably, there was no size scaling in the tunicates' diets relative to 246 their body size (i.e., smaller tunicates did not consume smaller prey), which is in contrast to 247 crustacean mesozooplankton; large tunicates have larger predator-to-prey size ratios than small 248 tunicates (Conley et al., 2018). Rather, the key distinction (with respect to food web dynamics) 249 between the tunicates is the level of predation. Small tunicates have very strong levels of top-250 down control, exerted by all mesozooplankton and higher trophic level predators. While large 251 tunicates experience predation by similar predators as small tunicates in the model, the strength 252 of that predation is reduced to account for their larger size. Other distinctions between the two 253 tunicates, including ingestion rates, metabolic scalings, and susceptibility to jelly-falls are

described in the next sections.

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256 2.2.2 Ingestion and assimilation

As filter feeders, the prey consumption rates of pelagic tunicates are typically measured as biomass-specific clearance (or filtration) rates, yielding ingestion rates (I, mg C m⁻³ d⁻¹) as the product of the specific clearance rate (C_b, m³ mg C⁻¹ d⁻¹), prey biomass (P, mg C m⁻³), and predator biomass (Z, mg C m⁻³)(Acuña et al., 2011):

$$I = C_b P Z$$
 (eq. 1)

262 In contrast, COBALTv2 and other NPZD-type models utilize saturating functional response

263 (Hollings Type II in the case of one prey) for zooplankton grazing (Fasham et al., 1990;

264 Gentleman et al., 2003; Stock and Dunne, 2010):

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 $I = \frac{I_{max}PZ}{K_i + P}$ (eq. 2)

266 Where I_{max} is the maximum specific ingestion rate (d⁻¹), and K_i is the ingestion half-saturation 267 constant (mg C m⁻³). At $K_i >> P$, eq. (2) reduces to $I = (I_{max}/K_i)*P*Z$, and C_b becomes equivalent 268 to I_{max}/K_i (Acuña and Kiefer, 2000).

269 For small tunicate clearance rates, we used allometric relationships from Lombard et al. 270 (2009), which measured physiological rates for a common appendicularian, Oikopleura dioica. 271 Using a characteristic individual body size of 1 mm and associated biomass of 6.68 µg C, 272 specific filtration rates at 0°C (converted using a Q10 of 1.88) range from 0.010-0.017 (mean: 0.013) m³ mg C⁻¹ d⁻¹ (Lombard et al., 2009a). Considering that the clearance rates of both small 273 274 and large tunicates do not significantly change with low to medium food concentrations (Gibson 275 and Paffenhöfer, 2000; Paffenhöfer and Köster, 2011), we opted for a higher tunicate half-276 saturation constant ($K_i = 250 \text{ mg C m}^{-3}$) than that of crustacean zooplankton (102 mg C m⁻³; 277 Table 1), consistent with the estimated range of K_i for O. dioica of 20-500 mg C m⁻³ (Acuña and 278 Kiefer, 2000). The equivalent I_{max} at low prey concentrations would be 2.50-4.25 (mean: 3.25) d⁻ 279 ¹. Following model tuning, we used an I_{max} value on the low end of the range (1.875 d⁻¹) given a 280 mean K_i, to avoid overconsumption by small tunicates. However, considering the wide variation 281 in K_i, these values were well within the observational bounds. The tradeoffs between tunicates and crustacean zooplankton were visualized in a plot of specific ingestion at 25°C (Fig. 2a): the 282 283 small tunicate I_{max} and K_i results in a specific ingestion in between small/micro-zooplankton and 284 medium/crustacean zooplankton. Additionally, a sensitivity run was conducted to illustrate the 285 effect of the I_{max} tuning choice (Section 2.3).

For large tunicates, we were able to use an allometric scaling relationship from a prior 286 287 effort (Luo et al., 2020) that compiled length, carbon biomass, and clearance rate relationships 288 from various studies (see Madin and Deibel, 1998) into a single equation. Using a characteristic 289 biomass of 1.5 mg C, which corresponds to a 20-38 mm length individual, specific clearance 290 rates at 0°C range from 4.2E-4 to 7.4E-3 (mean: 1.8E-3) $m^3 mg C^{-1} d^{-1}$. Using a K_i of 250 mg C m^{-3} , and the same relationship between clearance rates, K_i , and I_{max} as above, we estimated large 291 tunicate I_{max} values to be between 0.105-1.85 d⁻¹. In the model, we used a value in the lower half 292 293 of the reported range (0.55 d⁻¹), tuned in conjunction with other variables with wide uncertainty 294 bounds to match observed biomass concentrations. At the lowest prey concentrations, the I_{max}/K_i 295 of large tunicates matched that of the large crustacean zooplankton. As prey concentrations 296 increased, large tunicate ingestion fell between that of the medium and large crustacean

297 zooplankton and reached its maximum ingestion rate much slower than either crustacean group298 (Fig. 2a).

Assimilation efficiency (AE) for zooplankton is typically set to be a fixed fraction of ingested material in models (between 0.6-0.8; Carlotti et al., 2000), and for crustacean

301 zooplankton in COBALTv2, it is set to 0.7, allowing for 70% of all food consumed to be

302 assimilated independent of prey concentration. However, for pelagic tunicates, in particular

303 appendicularians, there is evidence of AE declines as prey concentration increases, due to

- development of tears of their pharyngeal filter and active prey rejection with increasing food
- 305 (Acuña and Kiefer, 2000; Lombard et al., 2009a, 2011). Retention and assimilation efficiencies
- for salps and doliolids also vary widely, from 28-90%, which may be due to prey selectivity for optimal sizes and preferred taxa (Andersen, 1986; Dadon-Pilosof et al., 2019; Pakhomov, 2004;
- 308 Pakhomov et al., 2006; Vargas and Madin, 2004; Walters et al., 2019). While we have
- 309 implemented prey selectivity in the feeding relationships, there is still evidence for feeding
- 310 apparatus clogging at high food concentrations due to the formation of a food bolus (Harbison et
- apparatus clogging at high food concentrations due to the formation of a food bolds (Harbison of 311 al., 1986). Therefore, we implemented varying assimilation efficiencies for pelagic tunicates:
- 312

$$AE = AE_{max} - \left((AE_{max} - AE_{min}) \left(\frac{P}{K_{AE} + P} \right) \right)$$
(eq. 3)

313 where AE_{max} and AE_{min} are maximum and minimum assimilation efficiencies, respectively, and 314 K_{AE} is the half-saturation constant for AE (all dimensionless).

This AE equation is a desaturating functional form (Hollings Type II subtracted from AE_{max})(see also Berline et al., 2011). For small tunicates, AE_{max} and AE_{min} were 0.7 and 0.25, respectively, and K_{AE} was 110 mg C m⁻³, which is at the low end of the range for the appendicularian *O. dioica* (145.52 +/- 33.36 std. err.) as measured by Lombard et al. (2009). For

appendicularian *O. alolca* (145.52 \pm /- 55.56 std. err.) as measured by Lombard et al. (2009). For 319 large tunicates, to simulate the clogging response, we also used the same AE bounds as small

tunicates but with a K_{AE} of 215 mg C m⁻³, a value at which approximately half of the *Pegea*

confoederata salps studied by Harbison et al. (1986) would form boluses. The difference in K_{AE}
 between the tunicates results in the AE declining faster for small tunicates compared to their
 larger counterparts (Fig. 2b).

Non-assimilated egestion losses for tunicates were parameterized similar to large zooplankton with 100% of the detritus losses going towards sinking particulate organic matter and no dissolved organic matter generated through grazing. This assumption is consistent with the representation of small tunicates in Berline et al. (2011) and the observation that tunicate detritus, representing the sinking houses of appendicularians and salp fecal pellets, are an important source of zooplankton detritus (Alldredge and Silver, 1988), and contribute significantly to carbon export fluxes (Alldredge, 2005; Andersen, 1998; Luo et al., 2020;

331 Robison, 2005).

In GZ-COBALT, we did not modify the sinking speed of GZ-mediated detritus, which is pooled together with non-gelatinous detritus and sinks at a rate of 100 m d⁻¹ (Alldredge and Silver, 1988; Stock et al., 2014a; Turner, 2015). While studies have shown that GZ detritus can sink at rates greatly exceeding that of marine aggregates and crustacean zooplankton fecal pellets (Lebrato et al., 2013), we opted to focus this study on the impact of gelatinous zooplankton,

337 specifically tunicates, within the euphotic zone and leave the assessment of GZ-mediated export

- 338 on biogeochemical cycles at depth to future work.
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Figure 2. Specific ingestion (a), assimilation efficiency (b), and specific production (c) as a function of generalized prey biomass at 25°C for zooplankton in GZ-COBALT. The three original COBALTv2 zooplankton types (small, medium, and large zooplankton; dashed lines) are unchanged in all GZ-COBALT simulations from the COBALTv2 control model.

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347 2.2.3 Metabolism and respiration

348 In the COBALTv2 base model, the total zooplankton respiration rate is a sum of the basal 349 and active respiration rate, with the former the resting metabolic rate that is proportional to 350 biomass and the latter a fixed fraction of the ingestion rate (Flynn, 2005; Stock et al., 2014a). 351 Pelagic tunicates are true filter feeders, and in contrast with crustacean zooplankton, are 352 constantly in motion regardless of the food concentration, pumping water via tail oscillations 353 (appendicularians), muscle contractions (salps), and ciliary action (doliolids and pyrosomes) 354 (Deibel, 1998; Madin and Deibel, 1998). Therefore, a truly "basal" respiration rate only occurs 355 when the tunicate is anesthetized. For Salpa fusiformis, swimming accounts for over half of total 356 oxygen demand (Trueman et al., 1984), while for the appendicularian O. dioica, swimming 357 accounted for roughly 34% of active respiration (Lombard et al., 2005). Furthermore, Lombard 358 et al. (2005) found that there was no significant difference between the respiration levels of 359 appendicularians at various food levels (no food, low, and high food), implying that the energy 360 required to digest food was small compared to the energetic requirements of swimming. In 361 contrast, the basal respiration rates for crustacean zooplankton are relatively low compared to 362 respiration rates while feeding, which increase linearly with ingestion (Kiørboe et al., 1985). 363 Thus, a key contrast that is built into GZ-COBALT is the difference between crustacean and 364 gelatinous zooplankton respiration tradeoffs: crustaceans have relatively high active respiration 365 rate (30% of ingestion) but low basal respiration, whereas GZ have low active respiration (15% 366 of ingestion) but high basal respiration rates. Consequently, compared to crustacean zooplankton, tunicates incur a higher metabolic cost in low food concentrations, which prevents them from 367 368 accumulating biomass in large portions of the subtropical gyres. These tradeoffs can be seen on a 369 plot of specific production as a function of food concentration (Fig. 2c), and is most obvious 370 when comparing small tunicates vs. small mesozooplankton, as the tunicate specific production 371 remains negative for a greater portion of the low food concentrations. A full comparison of 372 parameters is found in Table 1.

Biomass-specific basal respiration for small tunicates was calculated using relationships for *O. dioica* (Lombard et al., 2005). A laboratory study in the absence of food found that a 6.68 µg C individual at 0°C has a weight-specific oxygen consumption of 0.068 (0.055-0.085) µmol

376 $O_2 \ (\mu mol \ C)^{-1} \ d^{-1} \ (Lombard et al. 2005)$. Assuming a general zooplankton respiratory quotient of 377 0.87 (Mayzaud et al., 2005), this translates to a basal respiration of 0.047-0.074 d^{-1} . We opted to 378 use a value at the low end of the range (0.047 d^{-1}), so as to avoid complete elimination of their 379 biomass in subtropical environments, where they are found in low concentrations (Steinberg et 380 al., 2008).

381 For large tunicates, GZ-COBALT takes advantage of the mean allometric respiration 382 relationship compiled by Luo et al. (2020) from observations in Madin and Deibel (1998; and 383 references therein). With an average large tunicate of 1.5 mg C in that study respiring 2.5E-3 (8.0E-4 - 8.1E-3) mmol O₂ mg C⁻¹ d⁻¹, and using a salp-specific respiratory quotient of 1.16 384 385 (Mayzaud et al., 2005), basal respiration varied by an order of magnitude (0.011-0.11 d⁻¹) with 386 mean of 0.035 d^{-1} . However, even with this large uncertainty range, Luo et al. (2020) found that 387 under average conditions in the pelagic oceans, even the lower bound of these respiration rates 388 were too high, such that metabolic demands exceeded available food resources, yielding 389 unfavorable conditions for survival, particularly in the subtropical gyres. Therefore, in the 390 baseline model configuration, we used a basal respiration rate (0.008 d^{-1}) that was slightly below 391 the lower bound, tuned to ensure realistic gelatinous zooplankton in oligotrophic ecosystems. 392 This choice is consistent with the strategy enlisted for calibrating the basal metabolic costs of the 393 crustacean zooplankton (Stock and Dunne, 2010) wherein highly uncertain basal metabolic rates 394 were calibrated to ensure that the simulated biomass was consistent with observations in the 395 oligotrophic gyres where the impact of basal metabolic costs are most prominent. In nature, To 396 explore the effect of this tuning, a sensitivity case was run where large tunicate basal respiration 397 was set to the mean value from allometry (Section 2.3).

398

399 2.2.4 Other sources of mortality

400 Another key difference between the small and large tunicates is in the additional sources 401 of mortality. While small tunicates primarily experience mortality through predation, large 402 tunicates experience cold temperature reproductive failures (Henschke and Pakhomov, 2019) as 403 well as mass mortality events (jelly-falls) (Henschke et al., 2015; Lebrato and Jones, 2009). 404 Large tunicate aggregation losses, representing jelly-falls, were parameterized as a quadratic loss 405 that is suppressed when food is plentiful, following the same functional form as the 406 phytoplankton aggregation losses as a function of nutrient limitation in COBALTv2 (Stock et al., 407 2020; Waite et al., 1992) and other global biogeochemical models (e.g., PISCES; Aumont et al., 2015). The aggregation losses are controlled by two parameters: f_{agg} , which represents the 408 409 fraction of the maximum ingestion rate above which aggregation losses are suppressed, and m_{agg} , 410 or a maximum aggregation loss rate. f_{agg} was set to 0.1 to account for the salp ability to tolerate low food concentrations, and m_{agg} was set to 1.0E-3 m³ mg C⁻¹ d⁻¹ to achieve jelly-falls 411 412 representing approximately 35% of total large tunicate mortality following Luo et al. (2020). 413

415	Table 1. GZ-COBALT zooplankton ingestion, respiration, and aggregation parameters. Plankton functional types
416	(PFT)s: smt = small tunicates, lgt = large tunicates, smz = small zooplankton, mdz = medium zooplankton, and lgz =
417	large zooplankton.

Parameter	Name (Units)	PFT	Values	References
	Maximum ingestion rate (day ⁻¹)	smt	1.875	(Lombard et al., 2009a)
I _{max}		lgt	0.55	(Luo et al., 2020; Madin and Deibel, 1998)
		smz	1.42	

		mdz	0.57	(II		
		107	0.23	(Hansen et al., 1997; Stock et al., 2020; Stock and Dunne, 2010)		
	In partice, half activities	smt, lgt	250	(Acuña and Kiefer, 2000; Gibson		
K _i	constant (mg C m ⁻³)	smz, mdz,	102	(Stock et al., 2014a, 2020)		
		smt	0.06	(Lombard et al., 2005)		
bresp	Basal respiration rate	lgt	0.008	(Luo et al., 2020; Madin and Deibel, 1998); tuned to allow for sufficient biomass in subtropical gyres		
	(day ')	smz	0.018			
		mdz	0.008	(Hansen et al., 1997; Stock et al., 2020: Stock and Dunne, 2010)		
		lgz	0.0032	2020, 5000k and Damie, 2010)		
faresp	Fraction of ingestion rate for active respiration	smt, lgt	0.15	See text; also tuned to allow for sufficient biomass in subtropical gyres		
	(dimensionless)	smz, mdz, lgz	0.3	(Stock et al., 2014a, 2020)		
AE_{min}, AE_{max}	Minimum and maximum assimilation efficiency	smt, lgt	0.25, 0.8	(Lombard et al., 2009a; Madin and Deibel, 1998; Pakhomov et al., 2006)		
	(dimensionless)	smz, mdz, lgz	0.7, 0.7	(Stock et al., 2014a, 2020)		
		smt	110	(Berline et al., 2011; Lombard et al., 2009a)		
K_{AE}	Assimilation efficiency half- saturation constant (mg C m ⁻³)	lgt	215	(Harbison et al., 1986)		
		smz, mdz, lgz	N/A	N/A		
f_{agg}	Fraction of maximum ingestion at which aggregation mortality is suppressed (dimensionless)	lgt	0.1	See text		
m _{agg}	Maximum aggregation loss rate $(m^3 mg C^{-1} d^{-1})$	lgt	1.0E-3	Calibrated to achieve jelly-falls representing 35% of total mortality		

418 419

420 **2.4 Physical Framework**

421

422 The GZ-COBALT model with 35 tracers was run in a global ocean-ice configuration 423 using the GFDL models Modular Ocean Model 6 (MOM6) and Sea Ice Simulator 2 (SIS2) in a 424 nominal 0.5°, or roughly 50 km, horizontal resolution (OM4p5, Adcroft et al., 2019). The 0.5° 425 horizontal grid improves the resolution of boundary currents compared to earlier generations of 1° MOM models. The vertical coordinate in MOM6 is a hybrid z*-isopycnal vertical coordinate 426 427 system implemented using an Arbitrary Lagrangian-Eulerian (ALE) method, such that isopycnal 428 coordinates are used in the ocean interior and a z* coordinate is used in the mixed layer. OM4p5 429 uses 75 vertical layers, which allows for finer resolution at the ocean surface (~ 2 m) compared to 430 earlier model configurations with 10 m surface resolution and 50 vertical layers (Adcroft et al., 431 2019). The ocean and ice model configurations are also equivalent to those components used

432 within the fully-coupled ESM4.1 model (Dunne et al., 2020).

Model simulations were forced using Common Ocean-Ice Reference Experiment II 433 434 (CORE-II) (Large and Yeager, 2009), a 60-year interannually varying dataset representing 435 atmospheric forcings from 1948-2007. The model was initialized similar to that of the fully-436 coupled model (Stock et al., 2020): from World Ocean Atlas 2013 (WOA13) data for temperature, salinity, oxygen, and dissolved inorganic nutrients (Garcia et al., 2013a, 2013b; 437 438 Locarnini et al., 2013; Zweng et al., 2013), and from Global Ocean Data Analysis Project 439 (GLODAPv2) for dissolved inorganic carbon and alkalinity (Lauvset et al., 2016). Other tracers 440 were initialized from outputs of a previous version of COBALT (Stock et al., 2014a), and the 441 two new gelatinous zooplankton tracers were initialized with biomass concentrations similar to 442 medium and large zooplankton. Additional sources of nutrients include atmospheric deposition 443 of NH₄ and NO₃ (Horowitz et al., 2003), dust from Zhao et al. (2018) with soluble Fe calculated 444 in accordance with Baker and Croot (2010), as well as coastal iron and river nutrients from the 445 GlobalNEWS dataset (Seitzinger et al., 2005), as described in (Stock et al., 2020) 446 GZ-COBALT was run for one 60-year interannual forcing cycle. Results are reported 447 from a climatology computed from the last 20 years of model simulation, representing 1988-448 2007. A COBALTv2 control simulation with the same exact model setup, but with tunicates 449 turned off, was also run for 60 years. 450 451 **2.5 Parameter Sensitivity Runs** 452 453 To understand the impact of the unique aspects of GZ physiology and ecology as 454 described above on their emergent distribution and productivity, we considered a number of 455 perturbations around the baseline settings described above. These sensitivity runs (Table 2) 456 examine both the effect of our tuning choices (cases 1-2) as well as some key tradeoffs relative 457 to crustacean zooplankton (cases 3-5). All parameter sensitivity runs were conducted following 458 the same physical forcing as the GZ-COBALT base simulation. 459 460 1) For large tunicates, the basal respiration rate was adjusted to be slightly below the 461 lower bound from the literature. Sensitivity case 1 examines the effects of using the 462 mean basal respiration rate from allometric relationships from the literature. 463 2) For the baseline GZ-COBALT model, we used a maximum ingestion rate of small 464 tunicates at the lower bound of the range. For sensitivity case 2, we test a case where 465 the small tunicates' I_{max} is higher, set to the mean of the literature-based range. 466 3) Sensitivity case 3 explores the impact of the unique feeding behavior of GZ relative 467 to crustaceans. For the calibrated model (base case), we used a mean K_i value from a 468 wide range measured by Acuña and Kiefer (2000). However, models are known to be 469 quite sensitive to this relatively unconstrained parameter (Stock and Dunne, 2010). 470 Thus, we ran a sensitivity test where the tunicate K_i values were the same as that of 471 the crustacean zooplankton. Since the maximum ingestion rate (I_{max}) was set in 472 combination with the K_i values to achieve specific filtration rates at low prey concentrations consistent with observations, Imax was also modified accordingly to 473 474 preserve the observational constraint. 475 4) Sensitivity case 4 examines another unique aspect of pelagic tunicates feeding 476 relative to that of crustaceans. In the baseline GZ-COBALT, we implemented varying 477 assimilation efficiency (AE) following Berline et al. (2011). To explore the 478 uncertainty associated with this assumption, we ran a sensitivity test where the

479 tunicates' AE were constant, but set at a slightly lower value than other zooplankton, 480 to account for their comparatively lower retention rates.

- 5) Finally, a case was run to explore the impacts of ignoring the role of large tunicate aggregation mortality (representing jelly-falls).
- 482 483

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484 To illustrate the impact of these various parameter modifications, we computed the 485 specific production of all zooplankton groups under an idealized condition of 25°C with a 486 generalized prey biomass (Fig 3), which can be contrasted against the GZ-COBALT baseline

- 487 (Fig. 2c).
- 488

Table 2. Parameters tested in sensitivity tests, showing the values in the base code as well as the permutation for the
 individual sensitivity case. For detailed names of parameters, see Table 1. Plankton functional types (PFTs): smt =
 small tunicates; lgt = large tunicates.

Case #	Parameter	PFT	Base code	Permutation
1	bresp (d ⁻¹)	lgt	0.008	0.035
2	I_{max} (d ⁻¹)	smt	1.875	3.25
2	$I = (1^{-1}) K (m - C m^{-3})$	smt	1.875, 250	0.765, 102
3	I_{max} (d ⁻), K_i (mg C m ⁻)	lgt	0.55, 250	0.22, 102
4	AE_{min} , AE_{max}	smt & lgt	0.25, 0.8	0.6, 0.6
5	$m_{agg} ({ m m}^3{ m mg}{ m C}^{-1}{ m d}^{-1})$	lgt	1.0E-3	0





in all GZ-COBALT simulations from the COBALTv2 control. The specific production in sensitivity case 5 is the
 same as the base case (Fig. 2c).

499 500

501 2.6 Validation data

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As described in the introduction, a central question in our analysis is whether the model can simultaneously reconcile recent measurements suggesting that GZ carbon biomass is *ca*. 10x greater than previously thought while also satisfying core observational constraints on crustacean biomass, nutrients, chlorophyll and net primary production (NPP). The subsections that follow describe the datasets compiled and/or enlisted for each of these tasks, and the analyses used to assess model-data consistency across trophic gradients and ocean biomes.

509

510 2.6.1 Pelagic tunicate dataset and model-data comparison

511 To generate biomass validation data for pelagic tunicates, we updated the gridded 512 tunicate data (primarily salps) from Luo et al. (2020) with additional data on both thaliaceans and 513 appendicularians from the NOAA Coastal and Oceanic Plankton Ecology, Production, and 514 Observation Database (COPEPOD) (O'Brien, 2014). From the COPEPOD database, raw data on 515 urochordates were extracted and divided into small tunicates (all appendicularians) and large 516 tunicates (salps, doliolids, and pyrosomes). With the exception of data from the ECOSAR-II 517 cruise (Muxagata, 1999), all other data were in numeric density only (# individuals m⁻³). Numeric density data were first converted to a common 330 µm mesh size (Moriarty and 518 519 O'Brien, 2013; O'Brien, 2005). Second, since the geometric mean cannot handle zeros, zero 520 numeric density values were modified to be a non-zero value slightly below the minimum value 521 for both size fractions (small: 0.0008 ind. m⁻³, large: 0.001 ind. m⁻³). Next, numeric density was converted to carbon biomass using the characteristic individual biomass values defined in section 522 523 2.2.3 (appendicularians: 6.7 µg C, salps: 1.5 mg C). Characteristic biomass values for pyrosomes 524 and doliolids were 22.9 mg C ind⁻¹ (100 mm individual) and 19.2 µg C ind⁻¹ (5 mm individual), respectively, following Lucas et al. (2014), which used regression conversions from Gibson and 525 526 Paffenhöfer (2000) and Mayzaud et al. (2007). Using the geometric mean, 1° gridded values were averaged by month, then year, for an annual mean biomass. Finally, the Jellyfish Database 527 528 Initiative (JeDI; Condon et al., 2015) database was additionally queried for appendicularian data. 529 Data from 90 additional 1° grid cells, primarily from the North Atlantic and Eastern Equatorial 530 Pacific, were present in the JeDI dataset but not in the COPEPOD database. These data were 531 added to our validation dataset. Appendicularian data were present in a total of 3.914 1° grid 532 cells (Fig. 4a).

533 Thaliacean data from the COPEPOD database were combined with the Luo et al. (2020) 534 gridded salp data, which primarily included gridded biomass data from Lucas et al. (2014), with updates from JeDI the Palmer LTER site at the Western Antarctic Peninsula (Steinberg et al., 535 536 2015), and KRILLBASE (Atkinson et al., 2017). Out of the 5,468 grid cells with data, there were 537 1,481 cells where COPEPOD data were only present, 1,952 cells where the Luo et al. (2020) data 538 were only present, and overlap at 2,035 grid cells (Fig. 4b). Because the raw data with assumed 539 lengths and carbon conversions from Lucas et al. (2014) were not available, we were unable to 540 examine individual data points for overlap and cross-validation. However, a broad examination of the two datasets revealed that at the areas of overlap, carbon biomass compiled from 541 542 COPEPOD was 1.4x (geometric mean) that of Luo et al. (2020), with variations likely due to the 543 finer taxonomic detail of the Lucas et al. (2014) effort. This was within the uncertainty bounds

544 that we considered acceptable (roughly 2x uncertainty). Ultimately, due to discrepancies in 545 classification and specificity over time (e.g., broad categories such as "Tunicata" and "Salps and 546 doliolids" were dominant in classifications from the 1950's and 1960's, but not later), we 547 decided that using a single characteristic carbon biomass conversion for each broad taxonomic 548 category in the COPEPOD data gave a taxonomic specificity consistent with the coarsest 549 taxonomic specificity in the data. Further, this single biomass conversion removed a persistent 550 discontinuity in the Luo et al. (2020) carbon biomass values in the Indian Ocean south of 5°N 551 that we were previously unable to resolve. Since the vast majority of the JeDI data sources in the 552 Indian Ocean were from the 1959-1965 International Indian Ocean Expedition (IIOE), which are 553 also present in COPEPOD (Condon et al., 2015), we opted to replace the Luo et al. (2020) Indian 554 Ocean data with the COPEPOD data during our merge. For the rest of the oceans, we merged the 555 two datasets by taking the geometric mean at every grid cell where there was overlap.

556 Ultimately, the discrepancies between the datasets were quite low compared to the 557 differences in biomass due to sampling type, particularly when comparing extractive (nets) vs. 558 non-extractive (imaging) methods. Since traditional, net-based sampling systems break apart 559 fragile organisms such as pelagic tunicates and other gelatinous zooplankton, a biomass 560 adjustment is necessary to account for the reduced sampling from nets. Remsen et al. (2004) 561 used concurrent sampling with an imaging system and a 162 µm mesh plankton net, and found 562 that for pelagic tunicates, their abundance was undersampled by nets by a factor of 3-4x, and 563 their carbon biomass undersampled by a factor of 5-15x. Therefore, we considered an additional 564 "adjusted biomass" from samples with a 10x increase relative to the unadjusted biomass. We 565 focus mainly on this adjusted biomass, which is indicative of nascent appreciation of the likely 566 broader importance of gelatinous zooplankton as revealed by optical instruments. This is also 567 consistent with our intent to assess whether these high values can be reconciled with the overall 568 high abundance of mesozooplankton in many regions.

569 We complement our assessment of the simulated magnitude of tunicate biomass with one 570 of the relationship between tunicate biomass and other ecosystem properties spanning 571 oceanographic gradients. As the spatial gradients in tunicate biomass span 5 orders of magnitude, 572 this assessment provides a second metric less sensitive to the adjustments above. To do this, we 573 considered the GZ biomass as a function of chlorophyll concentration. The resultant, large-scale 574 relationship allowed for contrasts between large and small tunicates, and between tunicates and 575 crustaceans. For chlorophyll, we used the GlobColour merged satellite chlorophyll product (from 576 MERIS, MODIS-Aqua, and SeaWiFS) monthly climatology for case 1 waters using the weighted 577 averaging method, blended at latitudes south of 50°S with the Southern Ocean algorithm of 578 Johnson et al. (2013). We computed a growing season mean, define as all months for latitudes 579 between 30°N and 30°S, and spring and summer only for latitudes poleward of 30°N/S. The 580 slope of the log-log relationships between chlorophyll and the biomass of small tunicates, large 581 tunicates, and crustacean mesozooplankton (Moriarty and O'Brien, 2013; more details in Section 582 2.6.3) were established as emergent relationships for validation purposes.

583

584 2.6.2 Biome definitions

Finally, we assess gelatinous zooplankton simulation as a function of ocean biome,
adjusting for any systematic biases in the model by referencing biome locations to chlorophyll,
light, and temperature thresholds. We used the three major ocean biomes of Stock et al. (2014),
following Banse (1992): 1) Low Chlorophyll (LC), which encompasses the subtropical gyres, 2)
High Chlorophyll Seasonally Stratified (HCSS), which encompasses the high latitudes, and 3)

590 High Chlorophyll Permanently Stratified (HCPS), which includes the coastal and equatorial 591 upwelling regions. Stock et al. (2014) used a threshold of 0.125 mg Chl m⁻³ to separate between 592 the low vs. high chlorophyll regions in observational chlorophyll datasets. In our biome 593 definition, we first calculated the total ocean area with observational chlorophyll values lower 594 than that threshold (approximately 40% of the world's oceans), then found the model chlorophyll 595 threshold that resulted in a model LC area that most closely matched the LC surface area from 596 observations. For the COBALTv2 control and GZ-COBALT, this threshold was 0.162 and 0.184 597 mg Chl m⁻³, respectively. Next, to distinguish between the seasonally vs. permanently stratified 598 regions, we used the minimum of the mixed layer irradiance climatology (light averaged over the 599 mixed layer). HCSS regions were demarcated as those with minimum mixed layer irradiances 600 lower than 5 W m⁻², while the opposite was true of HCPS. Using mixed layer irradiance more 601 accurately defined the seasonal seas vs. upwelling areas, preventing HCPS areas from occurring 602 in Arctic regions with shallow maximum mixed layers (Stock et al., 2014a). Biomes for both 603 GZ-COBALT and the COBALTv2 control are shown in Fig. S1.

604

605 2.6.3: Crustacean zooplankton dataset and model-data comparison

606 To assess whether pelagic tunicate biomass magnitude and cross-biome gradients can be 607 represented while maintaining crustacean zooplankton populations consistent with observations, 608 we used the 2012 gridded carbon biomass data compilation from the COPEPOD database 609 (Moriarty and O'Brien, 2013). The entire COPEPOD database (O'Brien, 2014) consists of 610 multiple types of data products, including the raw, taxonomic data as used above for pelagic 611 tunicates, as well as the Moriarty and O'Brien (2013) carbon biomass compilation, which is the 612 more commonly used dataset for mesozooplankton model validation. In total, the COPEPOD 613 global carbon biomass compilation includes over 150,000 data points that were converted to an 614 equivalent 333 um mesh net size, with each gridded value representing multiple data points. 615 Given the net-based sampling and the historical focus on crustacean zooplankton, the vast 616 majority of the individual data points consisted of hard-bodied mesozooplankton. Thus, we used 617 the COPEPOD global carbon biomass compilation as a proxy of crustacean mesozooplankton to 618 compare against the small and large crustacean mesozooplankton in GZ-COBALT. 619 Similar to the GZ data, the crustacean observations were also scaled with chlorophyll on 620 a log-log scale. This enabled us to make comparisons along trophic gradients and across biomes

for crustaceans and gelatinous zooplankton.
 Finally, in addition to GZ and crustacean constraints, we include a suite of standard
 biogeochemical metrics to ensure that the model solution satisfies large-scale productivity and

nutrient patterns. The data we used were the dissolved inorganic nutrient concentrations (NO₃, $PO = and SiO_3$) at the accurate from the Wealth Ocean Atlas (WOA) 2018 (Carrie et al.

625 PO₄, and SiO₃) at the ocean surface from the World Ocean Atlas (WOA) 2018 (Garcia et al.,

- 626 2019).
- 627
- 628

629 **3. Results**

630

631 3.1 Global distribution and biomass-chlorophyll scaling

The GZ-COBALT simulation produced values consistent with the adjusted biomass of small and large pelagic tunicates, while also reproducing observed crustacean biomass and satisfying ocean biogeochemical constraints (Figs. 4-5, Tables 3-4). Global NPP was 53.7 Pg C y^{-1} and export flux at 100 m was 6.36 Pg C y^{-1} in GZ-COBALT, compared to 55.4 and 6.23 Pg C y^{-1} in COBALTv2. Surface chlorophyll and macronutrient concentrations in GZ-COBALT also compared well with the COBALTv2 control and observational constraints (Fig. 5).

638 The modeled global mean annual biomass integrated over the top 100 m was 5.8 Tg C for 639 small tunicates and 81.5 Tg C for large tunicates, yielding a total 100 m biomass of 87 Tg C. A 640 small but non-negligible fraction of tunicate biomass was below 100 m, even with the model 641 lacking vertical migration, such that the water column integrated biomass was 102 Tg C. These 642 values are within the adjusted mean and uncertainty of the observations. In comparison, the 643 medium/large crustacean mesozooplankton biomass (representing the size fraction most closely 644 comparable to the values in COPEPOD database) in GZ-COBALT was 205 Tg C in the top 100 645 m, which was slightly lower than the COBALTv2 value of 220 Tg C. Observational estimates of 646 large mesozooplankton biomass from COPEPOD, using a biome-specific geometric mean and standard deviation to extrapolate globally, was 133 (+/- 209) Tg C over the top 200 m. See Table 647

- 648 3 for additional comparisons by major ocean biome.
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Figure 4. Global mean distributions of small and large tunicates and crustacean mesozooplankton, comparing tunicate adjusted observations (a,b), large crustacean biomass from COPEPOD (c) with results from the top 100 m of the model (d,e,f). Medium/large crustacean mesozooplankton model values are given as the large mesozooplankton plus 0.5*small mesozooplankton. Model data show the time-average of the growing season only (fall and winter months excluded poleward of 30°N/S).

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666 Table 3. Global and biome-specific biomass comparison of the observations and the GZ-COBALT model. Model 667 results are the annual area weighted mean of the top 100 m carbon biomass. Observational values are given as the 668 geometric mean and geometric standard deviation of values within the biomes, with global geometric standard 669 deviation calculated following the procedure in Luo et al. (2020). Biomes: LC = Low Chlorophyll; HCPS = High

670 Chlorophyll Permanently Stratified; HCSS = High Chlorophyll Seasonally Stratified. See Fig. S1 for biome maps.

	Large Tunica	tes (mg C m ⁻³)	Small Tunicat	tes (mg C m ⁻³)	m ⁻³) Med/Large Crustacean mesozooplankton (mg C m ⁻³)		
Biome	Adj. Obs. mean (stdev)	Model annual mean	Adj. Obs. mean (stdev)	Model annual mean	Obs. mean (stdev)	Model annual mean	
LC	1.4 (10)	1.4	0.05 (16)	0.06	0.98 (2.3)	2.6	
HCPS	1.9 (8.0)	3.2	0.11 (15)	0.16	3.6 (2.5)	6.6	
HCSS	2.9 (11)	2.4	0.08 (23)	0.28	2.9 (3.2)	8.3	
Global	2.0 (9.9)	1.6	0.07 (18)	0.11	2.0 (3.1)	4.1	







Figure 5. Surface chlorophyll, log₁₀ scale (mg Chl m⁻³, a-c) and nutrients (mmol NO₃ m⁻³, d-f; mmol PO₄ m⁻³, g-i;
mmol SiO₃ m⁻³, j-i) from GZ-COBALT (left column; a, d, g, j), the COBALTv2 Control (center column; b, e, h, k),
and observations (right column; c, f, i, l). Model bias, root mean squared error (RMSE), and Pearson's correlation
coefficient (r) are also reported.

In the observations, we found that there was a contrast in the slope and intercept of
biomass-chlorophyll scaling relationship between small tunicates, large tunicates, and crustacean
mesozooplankton (Fig. 6). The small tunicates had significantly less biomass and a steeper log-

682 log slope (0.63 ± 0.045) residual std. err.; Fig. 6a) than the large tunicates, which was much 683 flatter (0.22 ± 0.036 ; Fig. 6b). The crustacean mesozooplankton data had much less variability, 684 a mean biomass similar to that of the large tunicates, and a biomass-chlorophyll scaling slope a 685 little shallower than the small tunicates (slope: 0.57 +/- 0.009; Fig. 6c). GZ-COBALT 686 successfully captured the differences in mean biomass across all three groups, as well as the 687 contrast in slope between the three groups, though admittedly the modeled slopes were all 688 slightly steeper than the observational slopes. The large tunicates had the shallowest biomass-689 chlorophyll scaling slope (0.43, Fig. 6b), followed by the crustacean mesozooplankton (0.71, 690 Fig. 6c) and the small tunicates (0.89, Fig. 6a).

691



692 693 Figure 6. Log-log relationship between tunicate biomass and surface chlorophyll from adjusted observations 694 (blue) and model data (black), sampled at the same locations as the observational dataset, for (a) small 695 tunicates, (b) large tunicates, and (c) crustacean mesozooplankton. The observations were adjusted from the 696 data compilation to account for the systematic low sampling bias from nets (10-fold adjustment), with the grey 697 bars around the observational regression line showing the 5-15x adjustment range. Model values are from the 698 top 100-m, and the crustacean mesozooplankton biomass was computed as large mesozooplankton + 0.5*small 699 mesozooplankton. Observational surface chlorophyll, as well as model data, were time means from the 700 growing season. 701

702 The sensitivity tests illustrated the impact of various aspects of the base GZ-COBALT 703 parameter choices, as well as the distinct physiology of tunicates relative to crustacean 704 mesozooplankton. For large tunicates, the reduced basal respiration rate, relative to the literature-705 based mean, in the base GZ-COBALT simulation was key for achieving a mean biomass 706 consistent with observations (see case 1, Fig. S2a-c). Similarly, for small tunicates, the lower 707 maximum ingestion rate relative to the literature-based values, was also essential for achieving a 708 mean biomass consistent with observations, and a biomass-chlorophyll scaling slope that did not 709 deviate too far from the observational constraints (case 2, Fig. S2d-f). In case 2, total tunicate 710 production also doubled, despite modest increases in biomass, largely due to the role of the small 711 tunicates (Table 4). The tunicate biomass-chlorophyll scaling slope was a result of many factors, 712 including competition between tunicate size classes, as the relative increase in small tunicate 713 biomass resulted in a shallower scaling slope for large tunicates (case 2, Fig. S2e).

Sensitivity cases 3-5 focused on model formulations distinct to pelagic tunicates relative to the original crustacean mesozooplankton in COBALTv2. In case 3, where the ingestion halfsaturation constant (with associated adjustments to the maximum ingestion rate) was set to the same value as that of the crustaceans, the resultant mean biomass and biomass-chlorophyll scaling for both tunicates matched the observations more closely (Fig. S2g-i). This was an interesting result and could have been a tuning choice; however, doing so would have negated a

key criterion (of the half-saturation constant, K_i, being much greater than the prey concentration) in converting measurements of clearance rate to ingestion rate. Setting the assimilation efficiency to a constant value (acce 4) resulted in small tunicates being clearents charged by the set of the set

to a constant value (case 4) resulted in small tunicates being closer to observations, but large

tunicates dropping significantly in biomass, particularly in the low productivity areas (Fig. S2j-l).
 This suggests that the variable assimilation efficiency was one factor in allowing large tunicates

to survive in the subtropical gyres. Finally, in the case where large tunicate aggregation mortality

- was removed, this resulted in large tunicate biomass greatly increasing in the high chlorophyll
- areas, with associated increases in the biomass scaling slope (Fig. S2m-o).
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729 *3.2 Seasonal cycle*

730 All zooplankton exhibited a stronger seasonal cycle in the high chlorophyll seasonally 731 stratified (HCSS) biome compared to the high chlorophyll permanently stratified (HCPS) and 732 low chlorophyll (LC) biomes, with the biomass peak shifting later in the summer as zooplankton 733 size increases. GZ exhibited a late summer (August-September) peak for both small and large 734 tunicates. The large tunicates were also unique amongst zooplankton in that their biomass in the 735 HCSS biome did not exceed that of the HCPS biome (Fig. 7). Results from the sensitivity cases 736 showed that this is largely due to the large tunicate aggregation mortality, or jelly-falls (case 5, 737 Fig. S3k-o), which serves to strongly dampen blooms. Additionally, reductions in the ingestion 738 half-saturation constant (and associated maximum ingestion rate; case 3, Fig. S3a-e) and the 739 constant assimilation efficiency (case 4) also reduced the magnitude of the blooms. Additionally, 740 in case 4, the small tunicates' bloom timing was also shifted to be slightly earlier (Fig. S3f-j). In 741 the base GZ-COBALT configuration, the biomass of the non-GZ zooplankton were overall 742 reduced compared to the COBALTv2 control (-7%, -6%, and -7.2% for small, medium, and 743 large zooplankton, respectively), with the biggest difference seen in the summer 744 microzooplankton biomass in the HCPS biome. Other substantial differences included the

745 wintertime biomass of small crustacean mesozooplankton in the LC biome, due to the low 746 starting biomass in the control (Fig. 7).

747



748MonthMonthMonthMonth749Figure 7. Seasonal cycle of modeled (a) microzooplankton, (b) medium zooplankton, (c) large zooplankton,750(d) small tunicates, and (e) large tunicates, separated by biome. Biome definitions: LC = low chlorophyll,751HCSS = high chlorophyll seasonally stratified, HCPS = high chlorophyll permanently stratified. Southern752Hemisphere values were shifted six months such that Austral summer is represented by months 6-8. Solid lines753indicate the GZ-COBALT simulation, and dashed lines show zooplankton values from the COBALTv2 control754simulation.

756 3.3. Biogeochemical impacts

757 The overall impact of gelatinous zooplankton on the partitioning of energy between the 758 microbial food web, export to depth, and energy available to higher trophic levels through 759 mesozooplankton was assessed via the difference between GZ-COBALT and the control 760 formulation (Fig. 8). This comparison suggests that the two tunicate classes have a competitive 761 interaction with microzooplankton (Fig. 8c) and a small, but net negative impact of the total 762 combined production of mesozooplankton (i.e., GZ and crustaceans, Fig. 8f). This is in spite of a 763 competitive impact on crustacean zooplankton, which was greater for the small crustaceans, 764 particularly in the upwelling zones, compared to the large crustaceans (Fig. S4).

765 The differences between the simulations becomes more pronounced when considering 766 plankton functional types that dominate recycling vs. export processes. With the addition of 767 pelagic tunicates, the routing of carbon to the microbial food-web decreased, as indicated by 768 declines in both the heterotrophic bacteria production ratio and the microzooplankton production 769 ratio (Fig. 8c, 7l). Meanwhile, the particle export ratio (pe-ratio, defined as the export flux at 100 770 m divided by NPP) increased globally (Fig. 8i). This comes as small and large tunicates 771 contributed 0.19 and 0.79 Pg C y⁻¹, respectively, of total export production in the top 100 m (of 772 which 72% sinks past 100 m). This increase in gelatinous-mediated export reflects a 773 redistribution of export production from existing sources, with the largest coming from small 774 mesozooplankton (Fig. 9a,c), as well as a reduction in the dissolved pool.







781 production to NPP (d-f), POC export past 100 m to NPP, or pe-ratio (g-i), and free-living heterotrophic

782 bacteria production to NPP (j-l). The plots show the GZ-COBALT simulation (left column; a,d,g,j), the 783 COBALTv2 control (center column; b,e,h,k), and the difference between the two (right column; c,f,i,l).

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Figure 9. Top 100-m production of sinking detritus (a, c) and loss to higher trophic level (HTL) predators (b, 787 d) in the COBALTv2 control (a, b) and GZ-COBALT (c, d) simulations. Of the total export production,

788 approximately 84% of it sinks below 100 m. All values are from the top 100 m, and are in Pg C y⁻¹.

790 791 792 **Table 4.** Comparison of the major results from the COBALTv2 control, GZ-COBALT base simulation, and the sensitivity experiments. 'z100' refers to the top 100 m of the water column.

E. 11	COBALT	GZ- COBALT Base	GZ-COBALT Sensitivity Experiments				
Field	Control		Exp. 1	Exp. 2	Exp. 3	Exp. 4	Exp. 5
NPP (Pg C y^{-1})	55.4	53.7	55	51	54.5	54.5	53.6
Export at 100 m (Pg C y ⁻¹)	6.23	6.36	6.25	6.37	6.23	6.31	6.39
Total Grazing (Pg C y ⁻¹)	39.9	38.8	39.7	37.5	39.3	39.3	38.8
Zooplankton Ingestion, z100 (Pg C y ⁻¹)	53.6	51.1	53.1	47.9	52.2	52.4	50.9
HP Ingestion, z100 (Pg C y ⁻¹)	2.39	2.18	2.33	1.96	2.26	2.25	2.27
Total Phytoplankton Biomass (Pg C)	0.488	0.482	0.486	0.466	0.485	0.486	0.48
Total Zooplankton Biomass (Pg C)	0.659	0.719	0.659	0.676	0.706	0.677	0.766
Total Tunicate Biomass (Tg C)		102	8.69	112	73	36.9	161
Sm. Tunicate Biomass, z100 (Tg C)		5.78	6.13	37	3.73	3.2	5.53
Lg. Tunicate Biomass, z100 (Tg C)		81.5	1.08	61.5	58.1	26.8	132
Crustacean Mesozoop. Biomass (Pg C)	0.378	0.356	0.373	0.331	0.365	0.368	0.352
Tunicate Prod., z100 (Pg C y ⁻¹)		0.49	0.0361	1	0.259	0.0884	0.0724
Crust. Mesozoo. Prod., z100 (Pg C y ⁻¹)	4.54	3.89	4.39	3.31	4.14	4.21	2.77
Z-ratio	0.082	0.072	0.08	0.065	0.076	0.077	0.07
Sm. Tunicate Detritus Prod., z100 (Pg C y ⁻¹)		0.19	0.21	1.58	0.11	0.14	0.18
Lg. Tunicate Detritus Prod., z100 (Pg C y ⁻¹)		0.79	0.01	0.51	0.5	0.38	0.96
Tunicate Prod./Biomass, z100 (y ⁻¹)		5.61	5.02	10.2	4.19	2.95	5.25
Mesozoop Prod./Biomass, z100 (y ⁻¹)	14.70	13.4	14.3	12.3	13.9	14	13.1
Lg. Tunicate Prod. lost to HTL Pred.		57%	0%	56%	56%	51%	91%
Lg. Tunicate Prod. lost to Jelly-Falls		35%	4%	33%	34%	34%	0%
Slope of Sm. Tun/Chlorophyll scaling		0.89	0.9	1.1	0.67	0.79	0.88
Slope of Lg. Tun/Chlorophyll scaling		0.43	0.26	0.29	0.39	0.62	0.58

795 **4. Discussion**

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797 We have added a simple representation of two pelagic tunicate groups, representing 798 appendicularians and thaliaceans, into the GFDL COBALTv2 ocean biogeochemistry model 799 (GZ-COBALT) that captures large-scale patterns in tunicate distribution consistent with the 800 emerging recognition of their importance to marine ecosystems, while maintaining a skillful 801 representation of crustacean mesozooplankton, surface chlorophyll, and macronutrient 802 concentrations. The GZ-COBALT simulation achieved a reasonable match between the 803 modelled mean tunicate biomass and a global observational dataset, compiled from a range of 804 sources including the COPEPOD database (O'Brien, 2014, 2005), Jellyfish Environmental 805 Database Initiative (JeDI) (Condon et al., 2015), and KRILLBASE (Atkinson et al., 2017). 806 Notably, GZ-COBALT captured a contrast between gelatinous and crustacean zooplankton types 807 in their emergent relationship between biomass and surface chlorophyll (Fig. 6). These results 808 confirm that it is possible to reconcile GZ biomass an order of magnitude above previous 809 estimates (Remsen et al., 2004) with prevalent crustacean zooplankton populations: carbon flows 810 through planktonic food-webs are sufficient to support both GZ and crustacean populations.

811 Observations of tunicate biomass exhibited high variability, even when compared with 812 crustacean zooplankton observations gridded to the same horizontal resolution (Moriarty and 813 O'Brien, 2013), which indicates either large sampling variability (e.g., from inconsistency in 814 sampling effort and/or gear), or unresolved physical or biological dynamics (Andersen, 1998; 815 Boero et al., 2008; d'Ovidio et al., 2010; Greer et al., 2020; Lévy et al., 2018; Luo et al., 2014), 816 or a combination thereof. The modelled variability for tunicates was lower than the observations 817 may suggest, even when daily rather than monthly outputs were sampled (Fig. S5). This was also 818 apparent for crustacean zooplankton, as it is a near-universal outcome when comparing global 819 biogeochemical fields against point measurements or averages of small numbers of point 820 measures (e.g., Krumhardt et al., 2017; Martiny et al., 2019; Saba et al., 2011; Usbeck et al., 821 2003). The discrepancy is admittedly acute for the tunicates, which was not unexpected given the 822 sparsity and difficulty of measurements. A more complete understanding the drivers of this 823 patchiness and their implications will likely require high resolution physical simulations and GZ 824 models capable of better resolving unique aspects of GZ life cycles and ecology conducive to 825 patch formation (Groeneveld et al., 2020; Henschke et al., 2018a, 2018b)

826 In our analysis, we found a strong contrast in the biomass-chlorophyll relationship 827 between crustacean zooplankton, small tunicates, and large tunicates, wherein the large tunicates 828 exhibited a flatter scaling relationship compared to the steeper scalings of the small tunicates and 829 crustaceans (Fig. 6). After incorporating an expanded view of GZ biomass considering 830 undersampling by nets, the resultant observational biomass-chlorophyll scaling became one of 831 our primary validation tools, as this emergent relationship can capture the mean biomass 832 responses across productivity gradients. These relationships become important as steep spatial 833 gradients in the contemporary ocean generally translates to amplified trends with climate change 834 (Stock et al., 2017, 2014b). The shallower slope for large tunicates relative to crustacean 835 zooplankton, in contrast, is indicative of less sensitivity to NPP, and suggestive of greater 836 resilience to NPP declines projected by the majority of models under high emissions scenarios 837 (Kwiatkowski et al., 2020) than their crustacean competitors. This would be consistent with 838 current hypotheses for increased prevalence of GZ under climate change (Henschke et al., 2016). 839

841 4.1 Marine food web and biogeochemical impacts

842 Pelagic tunicates have long been identified as a potentially important source of carbon 843 export, due to fecal pellets from salps (Iversen et al., 2017; Madin et al., 2006; Ramaswamy et 844 al., 2005; Smith Jr et al., 2014; Urrère and Knauer, 1981) and appendicularians (Wilson et al., 845 2013), discarded appendicularian houses (Berline et al., 2011; Lombard and Kiørboe, 2010; 846 Robison, 2005), and salp and pyrosome carcasses from jelly-falls (Henschke et al., 2013; Lebrato 847 et al., 2013; Lebrato and Jones, 2009). Given the boom-and-bust population dynamic of pelagic 848 tunicates, they can often be found to dominate POC export when present (Madin et al., 2006; 849 Smith Jr et al., 2014). Indeed, a recent study from a NASA EXPORTS cruise found that salp 850 fecal pellets comprised up to 80% of the detrital production in the upper 100 m in the NE Pacific 851 when present, though they contributed an average of 28% of fecal pellet carbon production over 852 a month-long sampling period (Stamieszkin et al., 2021). In our 20-year model climatology, 853 large tunicate detritus production comprises 20% of the total detritus production in the top 100 m 854 from July-September in the same region (Fig. S6). These values are a bit lower, but still fairly 855 consistent with the sampled cruise mean, though the high observed variability in Stamieszkin et 856 al. (2021) highlights the challenge in model-observation comparisons with snapshot studies at a 857 single time point. Model comparisons with GZ-COBALT and sediment trap data, which 858 integrates observations over longer time scales, will need to incorporate tunicate-specific POC 859 sinking speeds and is a target for future work.

One common implication of observations of pelagic tunicate-mediated carbon export is 860 861 that they would add to the existing POC export out of the surface ocean, often attributed to a 862 combination of phyto-detritus and crustacean zooplankton fecal pellets (Buesseler et al., 2008; De La Rocha and Passow, 2007). Here, we found that when considering the upper oceans (top 863 864 100 m), the integration of pelagic tunicates with a "traditional" food-web model did not 865 substantially increase total export flux past 100 m, which was 6.36 Pg C y^{-1} compared with 6.23 Pg C y⁻¹ in the COBALTv2 control despite GZ accounting for 0.7 Pg C y⁻¹. The integration of 866 867 GZ thus led primarily to a redistribution of fluxes away from those previously attributed to 868 crustacean zooplankton, rather than a creation of a new additive flux. The modest increase in 869 particle export that did occur is consistent with compensation for reductions in dissolved organic 870 carbon export arising from GZ-induced redirection of carbon flows away from the microbial food-web (Fig. 8, 9). 871

872 Compared to the offline estimates of tunicate export (1.3-3.9 Pg C y⁻¹ at 100 m; Luo et 873 al., 2020), the online GZ-COBALT model was lower, suggesting that food web and 874 biogeochemical feedbacks decreased the overall export contribution of tunicates. Rather than 875 relying on the direct application of GZ data, GZ-COBALT accomplishes this correspondence by 876 mechanistically representing the primary observational features and satisfying myriad additional 877 physical, biogeochemical, and food-web constraints. Large tunicates contributed about four 878 times more export production than small tunicates in GZ-COBALT (0.79 vs. 0.19 Pg C v^{-1}), with 879 approximately 0.16 Pg C y⁻¹ from jelly-falls, which was only slightly lower than the offline estimates of 0.3-0.7 Pg C y⁻¹. Tunicates in GZ-COBALT also contributed 0.1 Pg C y⁻¹ to higher 880 881 trophic level predators (Fig. 9d), which was much lower than the offline estimates of 0.8-1.1 Pg 882 C y⁻¹ (Luo et al., 2020). The higher trophic level predation in the offline model was one of the 883 least constrained parameters, as Luo et al. (2020) extracted a total GZ ecotrophic efficiency 884 (fraction of production to predation) from a combination of EcoPath models (e.g., Ruzicka et al., 885 2020), and tuned this term for individual GZ groups to achieve a global fraction consistent with 886 EcoPath estimates. Future observational and experimental work aimed to increasing our

understanding of GZ predation by higher trophic levels should reduce the uncertaintiesassociated with these global models.

889 The GZ-COBALT simulation showed that, compared with the COBALTv2 control, the 890 largest impact of pelagic tunicates to ocean biogeochemical cycles is in the partitioning between 891 the biological pump and the microbial food web. In GZ-COBALT, the impact of tunicates served 892 to reduce microzooplankton and bacterial production as a function of NPP by 14% and 4%, 893 respectively (Fig. 8). Pelagic tunicates, unlike other gelatinous zooplankton, are notable for 894 primarily grazing on small particles and their high predator to prey size ratios (Conley et al., 895 2018; Sutherland et al., 2010), though some exceptions exist (Post, 2002; Walters et al., 2019). 896 Recent work from Stukel et al. (2021) showed that in the Southern Ocean, the dominant salp, S. 897 thompsoni, most strongly competed with protistan grazers instead of with krill due to the large 898 size-based overlap between the salp and protistan diets. This is in contrast to previous 899 speculation that salps are a dominant competitor of the Antarctic krill, *Euphausia superba*, and 900 can be implicated as a factor in its long-term decline (Atkinson et al., 2004) and is consistent 901 with recent evidence that this decline can be attributed to positive anomalies in the Southern 902 Annular Mode (SAM) and loss of sea ice in the Southern Ocean (Atkinson et al., 2019). Our 903 results indicate that while tunicates do compete with large crustacean mesozooplankton for prey, 904 namely through the grazing of large phytoplankton and diatoms by appendicularians and 905 doliolids, tunicates also serve as a source of food for both small and large crustacean 906 mesozooplankton. Instead of competing with crustaceans, the magnitude of decline of 907 microzooplankton and heterotrophic bacteria production in GZ-COBALT compared to the 908 control and agreement with observations indicates that the microphagous tunicates serve as a 909 trophic and carbon export shunt away from the microbial loop and towards the mesozooplankton 910 food web and biological pump.

911

912 *4.2 Model limitations*

913 Amongst the marine zooplankton, thaliaceans are notable for their complex life cycles 914 which include the ability to reproduce asexually, alternation between sexual and asexual 915 reproductive phases (salps and doliolids), and hermaphroditism (pyrosomes), all of which can 916 yield large, transient, blooms under the right conditions (Andersen, 1998). Here, we have opted 917 against modeling the complex life cycle of pelagic tunicates (Henschke et al., 2018a, 2015; 918 Lombard et al., 2009b) for a more simple representation (Berline et al., 2011) aimed at capturing 919 their mean state, seasonal fluctuations, and long-term trends that can be run in an Earth System 920 Model over O(100) years. As such, there were a number of necessary simplifications, and 921 associated insights.

The model suggests that the mean turnover time scale, as measured by the ratio of production over biomass, or P/B, for pelagic tunicates is overall lower (implying slower growth) than microzooplankton and crustacean mesozooplankton (Table 4, Fig. S7). While some shallow coastal areas exhibited P/B exceeding $10\% d^{-1}$ in the summer for both small and large tunicates, the majority of the oceans had turnover rates $< 3\% d^{-1}$, even in the summer months. In contrast, the turnover timescales for large tunicates as reported in the literature were 15-71% d⁻¹ (Deibel, 1982; Gibson and Paffenhöfer, 2000; Madin and Purcell, 1992). While there may be some

averaging due to the model's monthly output, not even daily data captured the range of

variability in the observations (Fig. S5). Future efforts may focus on determining whether the

931 model's inability to reproduce observed variability is due to its coarse horizontal resolution

relative to the scales of observed variations in tunicate distributions (Greer et al., 2021; Luo et

al., 2014), or due to the representation of the simplified life cycle. For some gelatinous
zooplankton populations, a representation of the complex life cycle may be key for reproducing
interannual and multi-decadal climate fluctuations (e.g., Henschke et al., 2018b).

937 *4.3 Future outlook*

936

938 Gelatinous zooplankton (GZ) are ubiquitous throughout the world's oceans and a key 939 contributor to marine food webs (Hays et al., 2018). Of the GZ, pelagic tunicates are likely the 940 most important group in terms of carbon fluxes, due to their low trophic position and 941 microphagous diet. We demonstrate, through a new model with food-web and biogeochemical 942 feedbacks incorporated, that it is possible to reconcile an enhanced role of GZ in marine food 943 webs with the established importance of crustacean mesozooplankton and other ocean 944 biogeochemical constraints. Simulation results provide GZ flux estimates arising from a self-945 consistent physical-biological model satisfying myriad physical, biogeochemical and plankton 946 food-web constraints, a unique contribution relative to previous "offline" estimates. Climate 947 change is projected to drive decreases in NPP; coupled Model Intercomparison Project Phase 6 948 (CMIP6) models under the Shared Socioeconomic Pathway 5 (SSP5; fossil-fueled development) 949 project a 3-9% decline in NPP by the year 2100 (Kwiatkowski et al. 2020). Associated with 950 climate-induced NPP decreases, models also project a shift in mean pelagic body size: the 951 abundance of large autotrophic phytoplankton will likely be reduced relative to their smaller 952 counterparts due to increased warming, stratification, and subsequent nutrient limitation (Peter 953 and Sommer 2013). Consequently, as evidenced by the shallow scaling between biomass and 954 chlorophyll, the role of large pelagic tunicates (thaliaceans) in marine food webs may further 955 increase under climate change. 956 In this study, we have focused primarily on the upper ocean impacts of GZ, both to the

957 food web and to the balance between recycling and export. Omitted in this work are 958 considerations of the impact of fast sinking GZ export on the remineralization length scale and transfer efficiency to "sequestration depths", which may have further impacts on benthic fluxes 959 960 and air-sea CO₂ exchange (Kwon et al., 2009; Lebrato et al., 2019; Luo et al., 2020; Sweetman et 961 al., 2014; Titelman et al., 2006). In particular, there may be important feedbacks between 962 climate-induced stratification and tunicate-mediated increases in export. Our results indicate that 963 total carbon export was not significantly increased with inclusion of GZ in an ocean 964 biogeochemical model. However, these tunicate fluxes are globally quite significant and are 965 associated with a redistribution of export from existing phytoplankton and mesozooplankton 966 sources. As climate change will have differing impacts by taxonomic group, better understanding 967 of the sources of carbon export and the mechanisms that drive their variation will improve our 968 ability to project changes in the future.

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- 975

976 Author contributions

- 977 JYL and CAS conceived and designed the study, and evaluated the model. JYL wrote the code,
- 978 carried out simulations, analyzed data, and led the manuscript writing. JYL and TOB compiled
- 979 data. All authors contributed critical feedback and edits to the final manuscript.
- 980

981 **Competing Interests**

982 The authors have no competing interests to declare.

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Supplemental Tables and Figures



Figure S1. Major ocean biomes for the COBALTv2 control (a) and GZ-COBALT (b). The chlorophyll thresholds delineating the low chlorophyll (LC) regions were 0.162 and 0.184 mg Chl m⁻³ for the control and GZ-COBALT, respectively. The annual minimum of the mixed layer depth irradiance climatology ($< 5 \text{ W m}^{-2}$) delineated the high chlorophyll seasonally stratified (HCSS) biome from the high chlorophyll permanently stratified (HCPS) biome.



Figure S2. Biomass chlorophyll scaling for small tunicates (left column), large tunicates (center column) and crustacean mesozooplankton (right column) for the five sensitivity cases: case 1 (a-c), where large

tunicate basal respiration is increased; case 2 (d-f), where small tunicate maximum ingestion rate is increased; case 3 (g-i), where the tunicate ingestion half-saturation constant is the same as crustaceans, and maximum ingestion is adjusted accordingly; case 4 (j-l), where tunicate assimilation efficiency is set to be a constant; and case 5 (m-o), where large tunicate aggregation mortality is turned off. See Table 2, and the caption on Fig. 6 for further details.



Figure S3. Zooplankton seasonal cycle differences between the GZ-COBALT base case (dashed) and three of the sensitivity cases: case 3 (a-e); case 4 (f-j); case 5 (k-o). See caption on Fig. 7 for further details.



Figure S4. Differences in annual mean productivity ratios at the top 100 m between GZ-COBALT and the COBALTv2 control, showing ratios of small mesozooplankton (= medium zooplankton) production to NPP (a-c) and large mesozooplankton (= large zooplankton) production to NPP (d-f). The plots show the GZ-COBALT simulation (left column; a, d), the COBALTv2 control (center column; b, e), and the difference between the two (right column; c, f). Colorbar is set to be the same as in Fig. 8 to allow for a direct comparison.



Figure S5. Biomass chlorophyll scaling for small (a) and large (b) tunicates, using daily output from the GZ-COBALT model, showing a randomly selected day within the growing season from the last year of the model simulation (2007). See also the caption in Fig. 6 for more details.



Figure S6. Fraction of late summertime detritus production in the top 100 m from (a) all tunicates, and (b) large tunicates only. Late summer is defined as July-September in the Northern Hemisphere, and January-March in the Southern Hemisphere.

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Figure S7. Mean daily Production/Biomass (P/B, d⁻¹) ratios for small zooplankton (a), medium zooplankton (b), large zooplankton (c), small tunicates (d), and large tunicates (e). Months in the Southern Hemisphere are shifted such that Austral summer occurs during months 6-8, and Austral winter occurs during months 12, 1, and 2.