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4 **Improvement of beef cattle cow's pregnancy rate using**  
5 **an effective dose of Galectin-1**

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9 Erika da Silva Carvalho Morani<sup>1¶</sup> (<https://orcid.org/0000-0001-6862-1776>),

10 Helen Alves Penha<sup>1¶</sup> (<https://orcid.org/0000-0002-7156-444X>),

11 Fernando Sebastián Baldi Rey<sup>2¶</sup> (<https://orcid.org/0000-0003-4094-2011>)

12 Marcelo Rácoletta<sup>1\*¶</sup> (<https://orcid.org/0000-0002-3341-4993>)

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15 <sup>1</sup>Yoni Group, Inpreha Biotecnologia, Jaboticabal, São Paulo, Brazil.

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17 <sup>2</sup> Faculdade de Ciências Agrárias e Veterinárias, Universidade Estadual Paulista –

18 FCAV/UNESP, Departamento de Zootecnia, Jaboticabal, São Paulo, Brazil.

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20 **\*Corresponding Author** - Marcelo Roncoletta, email: [mroncoletta@inpreha.com.br](mailto:mroncoletta@inpreha.com.br)

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22 <sup>¶</sup> These authors contributed equally to this work

## 23 **Abstract**

24           Galectins are mentioned in the literature as multifunctional molecules that  
25 participate in several biological processes such as adhesion, cell proliferation, and cycle,  
26 apoptosis, RNA processing, inflammatory process control, and reproductive  
27 physiological mechanisms. Galectin-1 has been referred to as a mediator involved in the  
28 prevention of early embryonic mortality in mammals. Exogenous GAL-1 (eGAL-1) can  
29 be found in Tolerana®. The objective of the study was to evaluate whether eGAL-1 can  
30 increase the pregnancy rate when used in an AI procedure (in a complementary artificial  
31 insemination procedure, using a second AI gun). The pregnancy rate was determined by  
32 the pregnancy condition through an ultrasound exam performed 25 to 35 days after the  
33 fixed-time artificial insemination (FTAI) of breeding cows (n=3,469 beef cows). The  
34 efficacy of GAL-1 was evaluated by comparing the pregnancy rate of the two groups  
35 (Treatment and Control Groups) in 107 contemporary groups (YG) established by the  
36 created statistical model. Based on the obtained results, it can be confirmed that the  
37 correct administration of a single dose of eGAL-1 can increase the probability of  
38 obtaining pregnancy in beef cows by up to 8.68% ( $p < 0.0001$ ). The recommendation of  
39 the use of eGAL-1 during the FTAI procedure was reasonable in the beef cattle AI  
40 routine. On average, the complete procedure, using eGAL-1, took about  $10 \pm 5$  seconds  
41 more time than the conventional procedure.

42

## 43 **Introduction**

44

45 Cattle pregnancy loss can be considered one of the major challenges in the handling  
46 and cause of economic losses to the producer.

47 The average pregnancy rate in beef cattle, considering a single service, varies  
48 widely in the available literature. Now the fertilization rate in beef cattle can be high,  
49 reaching up to 95% in some scenarios, however, calculating the pregnancy rate by  
50 considering the complete breeding season, where 2 to 3 Artificial Insemination (AI)  
51 procedures and more exposure to the bulls are carried out. Embryonic mortality occurs  
52 mainly during the first 30 days of pregnancy, considerably reducing reproductive  
53 efficiency (Diskin and Morris, 2008; Diskin et al., 2016). Estimates indicate that  
54 pregnancy loss can reach 48% with one insemination during this period (Reese et al.,  
55 2020), which compromises the possibility of obtaining high pregnancy rates.

56 Several causes explain the reduction in the pregnancy rate, including early  
57 embryonic mortality (Diskin and Morris, 2008); diseases (Cheng et al., 2016), uterine  
58 asynchrony inducing failure of maternal recognition of pregnancy (Pope, 1988; Pohler et  
59 al., 2016), nutrition and milk production (Abdalla et al., 2017), placental homeostasis and  
60 uterine environment (Farin et al., 2006; Pohler et al., 2016), embryo-lethal genetic  
61 mutations (Pohler et al., 2016), and unbalanced immunological factors at the maternal-  
62 fetal interface (Bidarimath and Tayade, 2017).

63 The global livestock industry depends on the successful use of reproductive  
64 techniques to increase the productivity of herds, especially of animals with superior  
65 genetics. One of the most useful tools for this purpose is Artificial Insemination (AI),  
66 which allows for the maximization of reproductive performance (Baruselli et al., 2012).

67 Currently, it is recognized that Fixed-Time Artificial Insemination (FTAI) is an even  
68 more efficient tool to increase the reproductive performance of beef cattle (Baruselli et  
69 al., 2019). The efficacy of AI or FTAI procedures can be measured through the pregnancy  
70 rate. Therefore, why not associate eGAL with FTAI protocols.

71 In this context, the prevention of embryonic loss is still a challenge, given the  
72 complexity of mechanisms involved in the development and maintenance of pregnancy.  
73 Thus, new technologies have been developed, based on knowledge of the cattle's  
74 reproductive physiology, to achieve better reproductive results and reduce production  
75 costs, for instance, the hypothesis of using eGAL-1 as presented herein.

76 During pregnancy, uterine vascularization undergoes dramatic changes with a  
77 remodeling of existing vessels and the formation of a new network through angiogenesis  
78 that is stimulated (Cross et al., 2002) to provide an adequate supply of oxygen and  
79 nutrients for the developing embryo. At the same time, at the embryonic implantation  
80 site, the placentation process depends on a complex interaction between invasive  
81 trophoblasts and maternal immune cells, involving periods controlled by the development  
82 of branched angiogenesis, trophoblast differentiation, and syncytia formation.  
83 Interruption/alteration of this pattern of placental development can directly affect its  
84 function and result in pregnancy loss in humans (Blois et al., 2019). There are important  
85 differences in the degree of invasiveness of blood vessels during placentation of primate  
86 and rodent species (greater invasiveness) and ruminants (less invasiveness) and there is  
87 still no way to prove that in ruminants, GAL-1 acts on angiogenesis, however it is  
88 believed to have an important role in modulating maternal recognition.

89 Embryos generated by AI procedures consist of 50% genetic material from sperm  
90 deposited in the uterus. According to Hide and Schust (2016), alloantigen's derived from

91 parents or partners are present in the maternal environment at various times during the  
92 reproductive process. Embryos originating from embryo transfer procedures, when  
93 inoovulated in the recipient's uterus, have a greater chance of maternal-fetal  
94 incompatibility, as it is an embryo composed of 100% different genetic material. They  
95 are first exposed in semen deposition (either in the natural mating or AI conception) and  
96 continue during implantation and placentation. Changes in maternal immune responses  
97 allow fertilization and survival/development of the semiallogeneic conceptus until  
98 delivery. This immunomodulation must be balanced and continuous, responding  
99 appropriately not only to situations of invasion of commensal pathogens in the uterus,  
100 cell or tissue damage and any tendency to malignant transformation, as well as against  
101 the paternal alloantigen of the conceptus, making it “strange ” to the uterine environment  
102 and consequently attacked by the maternal immune system. The absence of this maternal-  
103 fetal tolerance or modulation of maternal immunity (the process by which the maternal  
104 organism recognizes/accepts the fetus without its immune system attacking it) can be  
105 considered as the main cause of embryonic loss (Hyde and Schust, 2016).

106 Thus, a complex network of metabolic, immunological, and endocrine interactions  
107 is activated during the gestational process. Such interactions are necessary to maintain  
108 pregnancy and occur at the maternal-fetal interface (on both sides), including cell  
109 signaling pathways related to differentiation and growth, vascular development, and  
110 immune regulation.

111 Complex immunoregulatory mechanisms at the maternal-fetal interface must be  
112 balanced to activate maternal tolerance against fetal alloantigens and protection against  
113 infections and inflammation (Hyde and Schust, 2016). Many mediators are involved in  
114 these mechanisms, and galectins, including GAL-1, play a key role, generating great

115 interest in the field of reproductive medicine due to their unique ability to modulate  
116 various processes of gestational development and their potential use as biomarkers for  
117 gestational disorders (Blois et al., 2019). These facts support the hypothesis for testing  
118 the effect of eGAL-1 administration in increasing the pregnancy rate in cattle production.

119 The present work explored the potential of the administration of exogenous GAL-  
120 1 (eGAL-1) to increase the pregnancy rate in beef cattle, combining the administration of  
121 a single dose of eGAL-1 with the FTAI technique, and the results represented a significant  
122 increase when using this tool. As mentioned in the literature, this experiment remarkably  
123 elucidates the effect of GAL-1 on reproductive physiology, positively impacting the  
124 development of pregnancy, and consequently, under the item reproductive efficiency in  
125 production farms.

126

## 127 **Material and Methods**

### 128 **Study design**

129 This study analyzed the increase in the pregnancy rate with the administration of  
130 a dose of exogenous GAL-1 (eGAL-1), combined with the technique of fixed-time  
131 artificial insemination (FTAI) in beef cattle. An effective dose of eGAL-1 means 1 (one)  
132 dose of Tolerana® (Inpreha Biotecnologia) whose administration is “extra” but similar  
133 to the application of a dose of semen, during the FTAI procedure. One effective dose of  
134 eGAL-1 contains  $200 \pm 10 \mu\text{g}$  of recombinant protein (GAL-1), diluted in  $200 \mu\text{L}$  of sterile  
135 PBS 1X pH 7.0 buffer solution (Phosphate Buffered Saline with kanamycin sulfate)  
136 present in a 0.25mL French-style straw. The definition of what represents an effective  
137 dose of eGAL-1 was established in previous experiments (unpublished data), where 5

138 different doses were tested, always considering use in the same presentation and  
139 administration model cited in the present experiment.

140 Recombinant GAL-1 was obtained through the construction of a heterologous  
141 expression vector containing the gene (pET-29a(+)+lgals-1 gene) and purification to  
142 obtain active protein, sterile, in its alkylated form and free of endotoxins. The Tolerana®  
143 a veterinary product is duly registered with MAPA [Ministry of Agriculture, Livestock  
144 and Food Supply] (MAPA/SP 000104-0.000001), and has intellectual property  
145 protection, with deposits at INPI and PCT (several countries), both in partnership with  
146 the Faculty of Pharmaceutical Sciences of Ribeirão Preto – University of São Paulo,  
147 Brazil.

148 The verification of the effectiveness of GAL-1 was performed by comparing the  
149 pregnancy rate in bovine females combining the administration of semen and eGAL-1  
150 (Treated Group - TG) versus the administration of only one dose of semen (Control  
151 Group) in the procedure of IA. Thus, in TG, the dose of eGAL-1 is deposited in the lumen  
152 of the uterus after the deposition of the semen dose, therefore, there are two procedures  
153 for passing the applicators through the cervix. Pregnancy rates for each group (TG and  
154 CG) were determined by ultrasound diagnosis (between 28 and 3 days after the FTAI  
155 procedure) and submitted to statistical analysis.

## 156 **GALECTIN-1 (GAL-1) production and purification**

157 GAL-1 can be obtained from mammalian genomes (from species such as human,  
158 bovine, ovine, caprine, equine and porcine) through heterologous expression systems, in  
159 the form of active protein, sterile, alkylated, and free of endotoxins. The method for  
160 obtaining recombinant Galectin -1 is determined by the manufacturing process of  
161 Tolerana® (Inpreha Biotecnologia®) and involves the following steps: (i) obtaining a

162 crude extract of bacteria cultivated to express Galectin-1; (ii) purification of Galectin-1;  
163 (iii) preservation of the lectin activity of Galectin-1 by alkylation; (iv) removal of  
164 bacterial endotoxin (LPS) from the alkylated Galectin-1 solutions; (v) adjustment of  
165 protein concentration; (vi) filling and (vii) quality control. Among the possibilities  
166 disclosed in the literature for the upstream and downstream steps, Galectin-1 was  
167 produced based on the following procedures, including particularities of the  
168 manufacturer's process. *Subcloning of Gal-1 into pET-29a(+) expression vector*

169 *Gal-1* Consensus Coding Sequence (CCDS) CCDS13954.1 (length 408nt) was  
170 synthesized and subcloned, with juxtaposed insertion of the desired sequence,  
171 immediately after the RBS Ribosome binding site sequence of a pET-29a (+) expression  
172 vector cut in NdeI / HindIII (GenScript®). This construct was then used for competent  
173 transformation of the Rosetta strain of *Escherichia coli*, maintained in a cell bank.

## 174 **Bacterial culture, expression, and Lysis**

175 Aliquots of *E. coli* strains transformed with the insertion of the vector containing  
176 the GAL-1 gene (pET-29a(+)+lgals-1 gene) were grown in systems with LB Broth Base  
177 medium containing kanamycin sulfate until obtaining optimal bacterial growth rate,  
178 demonstrated by optical density. Induction of expression is done with the addition of  
179 Isopropyl-D-Thiogalactopyranoside (Sigma-Aldrich) to the culture. After the induced  
180 growth period, the bacterial suspension is retained by microfiltration on a Hollow Fiber  
181 membrane (0.22µm, Cytiva) and centrifuged at 5000 g for 15-20 minutes at 4°C, always  
182 with the supernatant being discarded and the "bacterial crude = pellet", which were then  
183 subjected to bacterial lysis.

184 For Bacterial lysis, the crude or bacterial pellet was resuspended in Phosphate  
185 Saline Lysis buffer (1X PBS - 136.8 mM NaCl, 2.7mM KCl, 6.4 mM Na<sub>2</sub>HPO<sub>4</sub>, 0.9 mM



186 KH<sub>2</sub>PO<sub>4</sub>, pH 7.4), containing 14 mM Mercaptoethanol, protease inhibitor EDTA-free,  
187 lysozyme-1, RNase A-Type 3A, and DNase I Type IV-10. All components are Sigma-  
188 Aldrich. The pellet diluted in Lysis buffer (Chemical Lysis) was subjected to constant  
189 homogenization for 70 minutes and then sonicated for 3 cycles of 15 seconds each in a  
190 Vibra-Cells Sonicator, Sonics (Mechanical Lysis), with intervals of 20 seconds between  
191 each cycle. The bacterial lysate was then clarified by centrifugation at 7,000 g for 20  
192 minutes at 4°C and filtered through a 1.0 µm filter (Whatman) with the aid of a peristaltic  
193 pump (maximum pressure of 4 BAR).

## 194 **Purification steps**

195 After the Chemical and Mechanical Lysis process, the lysate was submitted to 3  
196 steps of purification by chromatography in an AKTA Protein Purification System  
197 (Cytiva) to obtain a buffered protein solution containing only Galectin-1.

198 The first step is based on affinity chromatography on agarose-lactose columns  
199 (Sigma-Aldrich), previously equilibrated with equilibration buffer (1XPBS, 14mM 2-  
200 ME, pH 7.4). After injection of the protein solution, the affinity column “binders” were  
201 washed and eluted with elution buffer (1X PBS containing lactose and 2-ME pH 7.4).  
202 The protein peak was collected and 20µM of iodoacetamide (Sigma-Aldrich; I1149) was  
203 added to the solution, keeping it under incubation at 4°C, protected from light, overnight.  
204 After this incubation, the solution was subjected to “size exclusion” chromatography  
205 (Sephadex G-25, Cytiva) to remove the free salts of iodoacetamide and lactose. The last  
206 chromatographic step was the removal of bacterial endotoxins (LPS). To this end, the  
207 preparations were subjected to chromatography using LPS affinity resin (PIERCE High-  
208 Capacity Endotoxin Removal Resin column - Thermo Scientific). After all the  
209 chromatographic steps, the protein concentration was determined by spectrometry (Abs

210 280nm) and expressed in milligrams of protein per milliliter (mg/mL) and were submitted  
211 to sterilizing filtration (0.22  $\mu$ m PES membrane).

212 Purified protein batches were submitted to the last stage of industrialization only  
213 if they reached compliance with the quality standard predetermined by the company,  
214 including protein concentration, microbiological status, protein bioactivity  
215 (Hemagglutination test), molecular weight analysis by SDS-PAGE, and SEC (size  
216 exclusion chromatography), protein secondary structure analysis (Circular Dichroism  
217 Analysis), aggregate detection and molecular size by DLS (Dynamic Light Scattering)  
218 analysis and endotoxin quantification (LPS). Protein identity was confirmed by LCMS  
219 (Liquid Chromatography Mass Spectrometry) and nucleotide sequence confirmation of  
220 human galectin-1 cDNA - galectin-1 [Homo sapiens] Consensus Coding Sequence  
221 (CCDS) CCDS13954.1 ([https://www.  
222 ncbi.nlm.nih.gov/projects/CCDS/CcidsBrowse.cgi?REQUEST=ALLFIELDS&DATA=  
223 CCDS13954.1&ORGANISM=0&BUILDS=CURRENTBUILDS](https://www.ncbi.nlm.nih.gov/projects/CCDS/CcidsBrowse.cgi?REQUEST=ALLFIELDS&DATA=CCDS13954.1&ORGANISM=0&BUILDS=CURRENTBUILDS)).

224 One dose of eGAL-1 translates to  $200 \pm 10$   $\mu$ g of purified protein diluted in 200  
225  $\mu$ L of Sterile Phosphate Buffer solution (1X PBS pH 7.0) containing 50  $\mu$ g/mL of  
226 kanamycin sulfate. The commercial presentation of eGAL-1 (Tolerana) is in a paper box  
227 containing 50 straws stored in vacuum-sealed plastic containers and kept at  $5 \pm 3^{\circ}\text{C}$  until  
228 the moment of use. The material was transported in isothermal boxes containing hard ice  
229 ( $5 \pm 3^{\circ}\text{C}$ ).

## 230 **Field experiment**

### 231 **Location**

232 The experiments were conducted in 17 commercial beef cattle farms located in  
233 different Brazilian municipalities (Campo Grande - MS; Naviraí - MS; Água Clara - MS;

234 Formoso do Araguaia - TO; Gurupi - TO; Paragominas - PA; Uberaba - MG , Uberaba –  
235 MG; Pedregulho – SP; São Gotardo – MG; Prata – MG; Água Clara – MS and Cuiabá -  
236 MT). It should be noted that the farms selected to participate in this experiment were  
237 farms that have a history of working with FTAI procedures for at least 2 years.

## 238 **Animals**

239 The experiments were carried out on female bovine animals (cows conventionally  
240 managed as dams and not intended for slaughter) managed in extensive beef cattle rearing  
241 systems. The dams were kept in an extensive rearing system, under native and/or  
242 cultivated pasture, with mineral supplementation. All cows underwent an FTAI  
243 procedure. It should be noted that only cows diagnosed as empty (by ultrasonography, 28  
244 to 35 days after the first service - 1<sup>st</sup> FTAI) were worked on in a second FTAI protocol.  
245 Fifteen days from the 2<sup>nd</sup>. FTAI, bulls were introduced for transfer with natural breeding.  
246 It is important to remember that the experiment and the statistical model considered only  
247 1<sup>st</sup> service results to ascertain the effectiveness of the dose of eGAL-1. Prophylactic  
248 management with annual vaccinations against BHV-1, BVDV, and BL (dose and booster)  
249 of cows before the start of the breeding season was implemented in some farms.

250 In total, 3469 beef cows (Nelore and crossbred dams) were considered in the  
251 statistical model, which were divided into 2 treatment groups (TG and CG) and for  
252 statistical analysis divided into 107 contemporary groups (YG) as described below. The  
253 experiment was designed with 4730 dams at the time of insemination, distributed equally  
254 (same number of cows in each group n= 2365) and randomly (without the previous choice  
255 of the TG or CG group that would be part of). However, 1261 cows were excluded from  
256 the experiment, because different reasons, including (i) dams did not maintain BSC  
257 between 3.5 and 2.5; the dams' body condition score (BSC) was observed in two

258 situations – at the time of the FTAI and the day before the pregnancy diagnosis. Only  
259 dams that maintained a BSC between 3.5 and 2.5 in the 02 situations mentioned above  
260 were approved to participate in the statistical analysis (ii) dams died during this interval;  
261 (iii) became ill during this interval (e. g., hoof, mastitis, diarrhea, pneumonia); (iv) those  
262 who had problems with the synchronization protocol (e. g. loss of CIDR); (v) who  
263 changed management lot); and for these reasons, it is noted that in some farms the number  
264 of dams mentioned in table 03 differs between the TG and CG groups. The numerical  
265 decompensation between groups was corrected in the construction of contemporary  
266 groups (YG), as described in the item below (Contemporary Groups and Statistical  
267 Analysis). An important detail is that the dams were submitted to BSC classification  
268 before the pregnancy diagnosis.

269         The criteria for defining the BSC used were based on the descriptions by Machado  
270 et al.(2008), who empirically determined the separation of dams into 5 BS classifications:  
271 1 (cachectical): complete visualization of the ribs, exposures of ileum bones and ischium,  
272 and pronounced muscle atrophy (skin and bones apparent); 2 (thin): very prominent bones  
273 with visible dorsal, iliac and ischial processes; 3 (great): light muscle coverage and no fat  
274 accumulation; 4 (fat): good muscle coverage and fat deposition at tail insertion; 5 (obese):  
275 all body angles covered, including protruding skeletal parts and overall animal  
276 appearance.

277         The experiment considered 3 different animal categories in the work lots – heifers,  
278 multiparous and primiparous. Multiparous and Primiparous cows had calves on their feet,  
279 at 60 to 100 days of lactation. These categories defined differences in the estrus  
280 synchronization protocols used for the categories.

281

## 282 **FTAI and eGAL-1 administration**

283 The breeding cows were kept in management batches on the farms. Each batch  
284 was submitted to FTAI after estrus synchronization protocols were performed. These  
285 synchronization protocols were decided by each farm, as described in Table 01. We did  
286 not interfere in these protocols and within each batch, there were no changes in the  
287 protocols. No imposition was imposed on participating farms in the choice of estrus  
288 synchronization protocols. Other decisions by the participating farms were (i) regarding  
289 the selection of the “bull” (semen doses) selected for use in the FTAI procedure and (ii)  
290 the choice and training of the inseminator who would inseminate each batch of breeding  
291 cows. A total of 46 bulls were used, selected by the partner farm and the semen doses of  
292 each bull, distributed in the Treated (TG) and Control (CG) groups. In total, 23  
293 inseminators participated in the experiment carried out on these 17 farms. There was no  
294 previous selection for the dam to receive the dose of eGAL-1 during the AI procedure. If  
295 the first dam that entered the containment trunk received the dose, the second did not  
296 receive it, thus continuing until the end of the insemination of the batch in question.

297

298 **Table 01. Farm (in letter codes), category of cows (H = heifers, P = primiparous or**  
299 **M = multiparous), days of estrus synchronization protocols (D0= day zero, D7 = day**  
300 **seven, D8= da eight, D9 = day nine, TAI day = day of the AI procedure (CG) and**  
301 **eGAL-1 and respective hormones applied, with amounts administrated, according**  
302 **to the estrus synchronization protocol adopted by the partner farm.**

Farms Codes	Category	D0	D7	D8	D9	TAI day
A	P	EB 2.0mL	-	-	PGF <sub>2α</sub> 2.0mL	D11

					ECP 1.0mL eCG 1.5 mL	
K	M	EB 2.0mL	-	-	PGF <sub>2α</sub> 2.0mL ECP 1.0mL eCG 1.5 mL	D11
B	M	EB 2.0mL	-	PGF <sub>2α</sub> 1.5mL ECP 0.5mL eCG 1.5 mL	-	D10
C	M	EB 2.0mL	PGF <sub>2α</sub> 2.0mL	-	ECP 0.5mL eCG 1.5 mL	D11
F J L K M N O	H	EB 2.0mL	-	PGF <sub>2α</sub> 2.0mL ECP 1.0mL eCG 0.5 mL	-	D10
G H I J L M N O P Q R T	M	EB 2.0mL	-	PGF <sub>2α</sub> 2.0mL ECP 1.0mL eCG 1.5 mL	-	D10
J	P	EB 2.0mL	-	PGF <sub>2α</sub> 2.0mL ECP 1.0mL eCG 1.5 mL	-	D10

303 EB- Estradiol Benzoate. PGF<sub>2α</sub> -Prostaglandin F<sub>2</sub>alpha. EC- Estradiol Cypionate. eCG

304 – Equine Chorionic Gonadotropin

305

306

307           The procedure in the treated group TG was to inseminate the breeding cows using  
308 a conventional semen applicator, followed by the administration of the eGAL-1 dose  
309 using a second applicator (identical to the semen), which represents that breeding cows  
310 in the treated group were at disadvantage compared to the CG, as they had 2 events of the  
311 transgression of the cervical rings. The deposition of the eGAL-1 dose in the uterine  
312 lumen was performed as the second insemination where after removal of the semen  
313 applicator, a second applicator mounted with a straw containing the protein dose was re-  
314 introduced as shown in Fig 1.

315           As a procedure in group CG, the females were inseminated according to the  
316 standard procedure, the same one recommended by (Brazilian Association of Artificial  
317 Insemination, 2018), with a single dose of semen (which represents 1 applicator being  
318 passed through the cervical rings). The time spent for the insemination procedure in the  
319 females of the CG and TG groups was considered a point of attention to the method. In  
320 this experimental model, we worked with the prerogative that dams belonging to the CG  
321 were at an advantage compared to the TG, as they received only "one act" to transverse  
322 the cervical rings during the procedure, and that it is also known that this "act " can  
323 negatively affect the pregnancy rate.

### 324 **Obtainment of the pregnancy rate in the groups**

325           The experimental results of pregnancy rates obtained in the breeding cows of the  
326 TG and CG groups obtained by ultrasonography (28 to 35 days after FTAI), and the  
327 increase in the rate obtained using the eGAL-1 was calculated based on statistical  
328 methodology considering dams, which maintained a BSC between 3.5 and 2.5, divided  
329 into 2 experimental groups (TG and CG) compared within the same contemporary groups

330 formed, as described below. The diagnosis was performed by a technician with  
331 experience in ultrasonography and without knowledge of the division of dams into TG  
332 and CG groups.

333

### 334 **Contemporary groups and statistical analysis**

335 To define and compare the pregnancy rates between breeding cows inseminated  
336 with and without Tolerana®, they were grouped into contemporary groups (YG). Each  
337 contemporary YG group was composed of dams inseminated by the same inseminator  
338 (identified by a letter code), belonging to the same farm (identified by a letter code), of  
339 the same animal category (which codes were used to facilitate separation (M =  
340 multiparous, P = primiparous or H = heifers), from the same management group  
341 (identified by the FTAI date + Farm code + management lot code); inseminated with the  
342 same semen batch (identified by the name of the Bull) and being of the same breed (N =  
343 Nellore and CB = crossbreed) A minimum number of at least 5 dams was considered to  
344 form a YG or those groups that did not show variation in the pregnancy rate (100 or 0%)  
345 were also discarded. To perform the analysis, the Generalized Linear Model (GLM) was  
346 applied, with the GENMOD procedure of the SAS (version 9.3), assuming a binomial  
347 distribution (pregnant or not pregnant) with residual effect and a logarithmic function  
348 (PROBIT). The model included the fixed effect of YG and treatment (dose = 0 of eGAL-1  
349 in the CG and dose = 200 that means  $200 \pm 10 \mu\text{g}$  of eGAL-1 in the TG).

350 Altogether, 107 YG were formed with the arrangement of these 3469 breeding  
351 cows, distributed among the 3 animal categories, inseminated with doses of semen from  
352 46 bulls, by 23 inseminators and distributed among the farms and management batches.



353 Remind that 1,261 dams were excluded to the statistical analyses as mentioned  
354 previously.

355 The PROC GENMOD is modeling the probability that Pre='2' = pregnant, using  
356 Information (Prm) and Effect parameters, where Prm1 = Interception; Prm2 = Dose0;  
357 Prm3 =Dose200; Prm4 = YG#1; Prm5 = YG#2; .....Prm10 = YG#107. Dose200 means  
358 administration of eGAL-1 in TG dams.

359 The pregnancy rate obtained in these 107 YG was determined considering the  
360 diagnosis of pregnancy by ultrasound between 28 and 35 days after the FTAI procedure.

361

362

### 363 **Ethical Statement**

364 This study complied with the ethical requirements for the use of animals in  
365 experiments and was approved by the CEUA/USP, protocol number 11.1.95.53.5.

366

### 367 **Results**

368 The administration of the recommended dose of eGAL-1 for the FTAI procedure  
369 took about  $10 \pm 5$  seconds longer than a conventional procedure. The probability of  
370 positive pregnancy in the CG group was 49.4% while in the TG group it was 58.08%  
371 ( $p < 0.0001$ ), as detailed in Table 2 . The mean obtained with Dose0 = 0.491, equivalent to  
372 49.41% of probability of obtaining a positive pregnancy in the CG, while the average  
373 obtained with Dose200 = 0.5808, equivalent to 58.08% probability of obtaining a positive  
374 pregnancy in the TG, which represents 8.68% difference between the treatment groups,  
375 when compared within of each YG.

376 **Table 2. Dose Least Squares Means, using Generalized Linear Model (GLM), with**  
377 **GENMOD procedure, under binomial distribution (pregnant/ not pregnant) and in**  
378 **logarithmic function (PROBIT), by source as “dose of eGAL-1”, that means dose 0**  
379 **= GC and dose 200 = TG, using [SAS] software, Version [6.9].**

Dose	Estimate	SE <sup>a</sup>	z Value <sup>b</sup>	Pr >  z	Mean <sup>c</sup>	SE of Mean <sup>d</sup>
0	-0.02368	0.07005	-0.34	0.7353	0.4941	0.01751
200	0.3259	0.06969	4.68	<.0001	0.5808	0.01697

380 (a) SE = Standard error; (b) z Value = z-score also called a standard score, gives an idea  
381 of how far from the mean a data point is. It is a measure of how many standard deviations  
382 below or above the population mean a raw score is. (c) Mean = probability of success of  
383 pregnancy rate; (d) SE of means = Standard error of the probability of average pregnancy  
384 rate.

385

386 The “YGs effects”, under binomial distribution (“pregnant” and “not pregnant”),  
387 did not present statistical significance ( $p = 0.1787$ ), perhaps because the variables that  
388 made up the construction of the YG greatly interfere in the pregnancy rate.

389 Table 03 describes the simple average obtained in each group (TG and CG) in the  
390 different farms, within each animal category, and for each inseminator who performed  
391 the procedures. Based on the comparison between the simple means of the CG (48.58%)  
392 and the TG (58.34%), a 9.76 percentage point difference was obtained between the  
393 groups. We are aware that the pregnancy rate can be interfered with by several factors or  
394 variables, going well beyond "just the location of the farm", so we proposed to discuss a  
395 discussion based on results based on this proposed statistical model, which considers

396 “product dose-effect” within contemporary groups, grouping all impacting variables for  
397 “pregnancy rate” within each of the YG created. There were so many variables that 107  
398 YG were created in the established statistical model. Importantly, among the calves born  
399 in this experiment, more than 900 of them conceived and gestated in the uterus that had  
400 contact with the eGAL-1 protein (dams belonging to the TG group) during the FTAI  
401 procedure, no congenital defect or stillbirths were observed.

402

403 **Table 03 – Average of pregnancy rate (%P) and number of cows (N) on the Control**  
404 **Group (CG) and Treated Group (TG) by each farm (identified by code name) and**  
405 **by all farms. Font in bolt format indicate higher %P in TG.**

	Groups			
	CG		TG	
Code name of Farms	NCG	%PGC	NTG	%PTG
A	52	38.46	54	48.15
B	100	50.00	617	60.62
C	169	46.75	281	57.65
F	18	50.00	19	73.68
G	25	60.00	20	70.00
H	22	59.09	20	85.00
I	29	62.07	18	61.11
J	337	47.48	215	53.95
K	80	55.00	82	50.00
L	274	50.36	197	56.35

M	42	40.48	24	45.83
N	74	47.30	65	46.15
O	66	57.58	66	65.15
P	90	35.56	90	53.33
Q	45	37.78	40	65.00
R	92	55.43	93	68.00
ALL	1515	48.58	1901	58.34

406

## 407 **Discussion**

408

### 409 **Efficacy of exogenous Galectin-1 on the increase of pregnancy rate**

410 The aim of this study was to explore the potential effect of eGAL-1 in increasing  
411 the pregnancy rate in beef cattle, combining the FTAI technique, the administration of a  
412 single dose of eGAL-1, measuring the difference in efficacy obtained, through the  
413 designed statistical model (comparison of the pregnancy rate between TG and CG within  
414 the formed YG), and thus it was observed that the probability of the pregnancy rate was  
415 58.08% in TG and 49.4% in CG ( $p < 0.0001$ ), demonstrating a positive efficacy of eGAL-1  
416 on the pregnancy rate (Table 2).

417 YG fixed several factors that can interfere with the pregnancy rate in beef cattle,  
418 keeping only the variable dose of eGAL-1 as an extra variable in the statistical model.  
419 YG considered the grouping of breeding cows from both groups (TG and CG) equally  
420 distributed (before dams' exclusion) within the same batch, within the same farm, within  
421 the same category, inseminated with doses from the same bull and by the same

422 inseminator. Particulars of sanitary and nutritional management are also being considered  
423 in the YG grouping (batch). The negative effect on the pregnancy rate (or increased  
424 probability of pregnancy loss) due to nutritional deficiency was controlled in the  
425 experiment, excluding dams that did not maintain their body score, as previously  
426 mentioned.

427 Thus, the pregnancy rate obtained on TG was different, (in this case, 8.68%  
428 higher), only because of the eGAL-1 administration ( $p < 0.0001$ ). Remembering yet, the  
429 YG effect, under binomial distribution (pregnant or not pregnant), was not statistically  
430 significant ( $p = 17.87$ ). However, in the scenery with 3469 dams, 3 animal categories, 46  
431 different bulls, 23 inseminators, several batches, and 2 treatment groups, it is reasonable  
432 to consider the YG effect as a biological effect. The recommendation to use a dose of  
433 eGAL-1 during a FTAI procedure was reasonable in the beef cattle routine. On average,  
434 the whole procedure, when we administrate de eGAL-1, spent only  $10 \pm 5$  seconds more  
435 than the conventional procedure - ten seconds as a price to get 8.68% more chance to  
436 pregnant a dam, is reasonable in the animal production systems.

437 The form of eGAL-1 administration is not foolproof to improve the pregnancy  
438 rate in beef cattle but showed that can help. eGAL-1 means Tolerana® administration and  
439 it is an innovative technology and its efficacy experimentation model was executed  
440 between the manufacturing company and partner farms. However, as in any product  
441 development process, we go through stages and technological challenges, such as (1) the  
442 ideal dosage for different cows categories, (2) the ideal eluent for maintaining the stability  
443 of the active protein in the product, (3) ideal packaging to facilitate the procedure and to  
444 not harm the product, (4) determination of the best form and moment of application, (5)  
445 the application procedure definition, (6) the interference of synchronization protocols,

446 among others. In this scenario, before obtaining this 8.68% higher pregnancy rate  
447 achieved in this experiment, using eGAL-1 administration, some procedures might not  
448 work well.

449         Also, there were technological challenges in manufacturing the active ingredient  
450 - a recombinant protein that has intrinsic particularities to the molecular structure, and it  
451 can suffer the methodology influence and the manufacturing processes. It should be noted  
452 that, like any biotechnological product, "product = process", while the product had no  
453 commercial registration, optimizations were performed in the process, aiming for greater  
454 yield and scale-up manufacturing. In some situations, with certain partners, such changes  
455 had a negative impact on the stability and effectiveness of the technology, resulting in no  
456 or small increase in the pregnancy rate. However, the manufacturing process is currently  
457 consolidated and robust on homogeneity, stability, and efficiency. In fact, during the  
458 experimental phase of technology development (Tolerana®), it should be noted that it  
459 was used more than 12 thousand times in AI procedures of bovine dams, and there were  
460 no undesired biological effects. There were no reports of discomfort, pain, and irritation  
461 with the administration of the product, except for those already known in artificial  
462 insemination procedures. It is also important to note that stillbirths, malformations, and/or  
463 neonatal complications were not verified. There are no reports of intoxication in humans  
464 using GAL-1 as an active ingredient in drugs. In the current literature, galectins (soluble  
465 in blood serum or expressed in tissues) have generally been used as biological markers of  
466 several pathological events (He et al. 2017; Vergetaki et al., 2014; Vasta, 2012), directing  
467 treatments, but still in experimental stages.

468         Another important note about interference on results falls on semen doses. The  
469 doses of semen used in this experiment were a "farm decision", even that we observed 46

470 different bulls, used. eGAL-1 is a protein and is feasible occur an interaction between it  
471 and some components of the semen extender, including lactose, a carbohydrate used in  
472 some recipes of semen extender that present a high interaction with galectin 1. If the  
473 protein binds with the semen extender lactose, it will be interacting with the endometrium  
474 at the time of administration of eGAI-1? This is a hypothesis does not respond by the  
475 authors yet. Difficult to know every semen extender present on semen doses used. Centers  
476 that industrialized semen doses do not share this information easily. For this reason, YG  
477 was considered as statistical analysis.

478           The efficacy of the technology should also be observed carefully. If it has not been  
479 performed consistently, considering the experimental and statistical model (as described  
480 in item *Methods - Contemporary groups and statistical analysis*), the effect of  
481 effectiveness can be masked. Thus, when used correctly, the results are quite promising.  
482 The indication of Tolerana® is NOT for the treatment of infertile nor sick animals, but it  
483 is indicated as a health catalyst of animal fertility, being, therefore, a tool to increase the  
484 reproductive/productive efficacy, impacting economically on the productive cattle chain.

485

486 **The economic impact of reproductive efficiency must be calculated**  
487 **and/or considered**

488

489           In the beef cow-calf system, the number of calves on the final breeding season,  
490 number of days up to the time of a new pregnancy (feeding cost versus open days), and  
491 cow reposition taxes must be considered the principal goals for a bio-economic  
492 evaluation. Moreira (2019) said that it is possible to reduce the investment cost of the  
493 FTAI program by 10% for each percentage point added to the pregnancy rate. If true,

494 using eGAL-1 administration on FTAI procedure the consumer can reduce the cost with  
495 the FTAI program per cow considerably and, consequently, can also reduce the cost of  
496 calves' birth. The cost benefits of eGAL-1 are interesting, considering that a dose of  
497 Tolerana® means 10% plus on the cost of the FTAI program per dam. More details of the  
498 economic impact of using eGAL-1 on the pregnancy rates rise in FTAI programs will be  
499 discussed in another paper.

500 The data presented in this study corroborate with several authors, as described  
501 below, and support the innovative hypothesis and the new product presented.

502

### 503 **Why galectin-1 could improve the pregnancy rate?**

504

505 Galectins are a family of evolutionarily conserved proteins distributed from lower  
506 invertebrates to mammals (Cummings & Liu, 2009; Modenutti et al., 2019). Thus, the  
507 efficacy of recombinant human Gal-1 under assisted reproduction procedures was  
508 evaluated, besides bovine females (this work), ovine, and equine (unpublished data). All  
509 evaluations presented in species different from bovine promising results only with dosage  
510 adjustment because of the area (cm<sup>2</sup>) of lumen uterus of each species.

511 From the mid-1970s onwards, several findings of animal lectins and  $\beta$ -galactoside  
512 ligands have been described. Barondes et al. (1994a) proposed the creation of the galectin  
513 family to group these proteins. "Electrolectin" was the first member of that family,  
514 isolated from tissues of electric fish (Teichberg et al., 1975). Even the first  $\beta$ -galactoside-  
515 binding lectins derived from mammals were described and were later defined as galectin-  
516 1 and 3 (Barondes et al., 1994b).



517           There is enormous structural diversity of glycoconjugates in living beings, which  
518 can be associated with a significant biological diversity since these glycostructures can  
519 encode various biological information decoded by lectins (Sharon and Lis, 1989).  
520 Therefore, the carbohydrates recognition by lectins is a biochemical phenomenon  
521 associated with several physiological and/or pathological processes such as cell  
522 fertilization, embryogenesis, migration, proliferation, and differentiation, immune  
523 defense, infection by microorganisms, and cancer (Sharon & Lis, 1986; Sharon & Lis,  
524 1989; Santos-de-Oliveira et al., 1994; Dias-Baruffi et al., 2003; Liu & Rabinovich, 2005).

525           Fifteen mammalian-derived galectins have been described, and all have a CRD  
526 with approximately 130 amino acid residues. Galectins were classified into three  
527 categories: “proto-type”, chimera, and “tandem repeat-type”, the first being those with a  
528 single CRD type and with identical monomers or dimers with CRD associated non-  
529 covalently (galectins: 1, 2, 5, 7, 10, 11, 13, 14 and 15). Galectin-3 represents the chimera  
530 type, has a CRD, and a non-lectin domain involved in its oligomerization (Cummings &  
531 Liu, 2009; Modenutti et al., 2019). Galectins 4, 6, 8, 9, and 12 are known as “Tandem  
532 repeat-type” because they have two distinct CDRs joined by a small binding peptide  
533 (Rubinstein et al., 2004a).

534           Most galectins have characteristics of cytoplasmic proteins such as acetylated N-  
535 terminal region, non-oxidized (free) sulfhydryl groups, and absence of glycosylation  
536 (Rubinstein et al., 2004a and b). However, galectins can be located on the cell surface,  
537 extracellular matrix, cytoplasm, and cell nucleus (Rubinstein et al., 2004a). Although  
538 these proteins can be detected in the extracellular environment, they do not present signal  
539 peptides, being secreted by the cells by a non-classical mechanism and independent of  
540 the endoplasmic reticulum and the Golgi complex (Hughes, 1999). Literature data suggest

541 that GAL-1 can be secreted by direct translocation of cytosol through the plasma  
542 membrane with the aid of cytosol and membrane factors, as described for fibroblast  
543 growth factor-2 (Schäfer et al., 2004; Nickel, 2005).

544 Since galectins are bivalent, they can promote the intercrossing of  
545 glycoconjugates on the cell surface in the extracellular environment and induce signal  
546 transduction events by forming clusters of receptors and a mesh (galectin-receptor) on the  
547 cell surface (Brewer et al., 2002). In the intracellular environment, the biological events  
548 of galectins do not seem to depend on their lectin properties, and they can participate in  
549 the processing of RNA and the regulation of cellular homeostasis (Liu et al., 2002; Wang  
550 et al., 2004). Interestingly, galectins can exert antagonistic effects depending on these  
551 proteins' location in the intra- or extracellular environment (Yang et al., 1996; Fukumori  
552 et al., 2003). Then et al. (2008) showed that LGALS1 has a high degree of structural  
553 conservation, dimerization, and binding properties with carbohydrates and integrins  
554 (adhesion proteins), suggesting that these properties are conserved among vertebrates and  
555 that they maintain a pattern of gene expression among the different types of the placenta  
556 (deciduous or not).

557 Galectins are multifunctional molecules that participate in several biological  
558 processes such as adhesion, proliferation, and cell cycle, apoptosis, RNA processing,  
559 control of the inflammatory process, and physiological mechanisms of reproduction  
560 (Perillo et al., 1995; Liu et al., 2002; Dias-Baruffi et al., 2003; Rubinstein et al., 2004b;  
561 Stowell et al., 2007; Ramhorst et al., 2012; Barrientos et al., 2014; Blois et al., 2019).

562 The galectin's maternal-fetal tolerance role, both innate and adaptive, is associated  
563 with regulating and modulating the embryo elongation events' immunological responses  
564 and adherence to the endometrium. Besides GAL-15 and GAL-1, other galectins can be

565 expressed by the endometrium and the placenta of mammals, presenting essential  
566 functions in differentiating the endometrium implanting the blastocyst, and differentiating  
567 the trophoblast (Farmer et al., 2008). They also contribute to placentation as they regulate  
568 the development, migration, and trophoblastic invasion, essential in early gestational  
569 development (Barrientos et al., 2014; Blois et al., 2019, 2007; Freitag et al., 2013). Even,  
570 they act in the maternal immunological tolerance mechanism to fetal alloantigen's,  
571 regulating the Natural Killer uterus cells and modulating T cells, which are mainly  
572 responsible for cellular immunity (Than et al., 2008).

573         The endometrial expression of GAL-1 fluctuates during the estrous cycle of  
574 different phases because steroidal hormones influence it. GAL-1 has been detected in 3-  
575 to 5-day old human embryos, acting on trophoblasts differentiation in the fetus's placenta  
576 and internal cell mass. The interaction of GAL-1 with integrins suggests participation in  
577 the extracellular matrix and placentation events, either in the oxygen exchange and/or  
578 nutrients or by the vessels formation (angiogenesis), showing that GAL-1 plays a vital  
579 role in interface signaling maternal-fetal since it has multiple biological functions (Choe  
580 et al., 1997).

581         Blois et al. (2007) demonstrated high pregnancy loss rates in mice in which the  
582 *Lgals1* gene was deficient (knockout mice). When treating deficient mice with  
583 recombinant GAL-1, there was a decrease in fetal loss and the restoration of tolerance  
584 through several mechanisms, including the induction of tolerogenic dendritic cells, which  
585 in turn promoted the expansion of regulatory T cells secreting interleukin-10 (IL-10) *in*  
586 *vivo*. Consequently, the protective effects of GAL-1 have been revoked in mice depleted  
587 of regulatory or IL-10 deficient T cells. Thus, they (Blois et al. 2007) demonstrated the

588 fundamental importance of GAL-1 in fetomaternal tolerance and the synergy between  
589 GAL-1 and progesterone in maintaining pregnancy.

## 590 **Conclusion**

591 This study showed the eGAL-1's effectiveness in improving the beef cattle  
592 pregnancy rate and by administering the recommended dose, the procedure may take 5 to  
593 10 seconds longer than the conventional procedure, however, the statistics show a  
594 considerable increase in the pregnancy rate. Considering the "eGAL-1 administration  
595 effect" it is possible to improve in 8.68% the chances of pregnancy in an inseminated  
596 cow.

597

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609

## 610 **References**

- 611 1. Abdalla, H., Elghafghuf, A., Elsohaby, I., Nasr, M. A. (2017). Maternal and non-  
612 maternal factors associated with late embryonic and early fetal losses in dairy  
613 cows. *Theriogenology*, 100, 16–23. doi: 10.1016/j.theriogenology.2017.04.005
- 614 2. Associação Brasileira de Inseminação Artificial. (2018). Manual de inseminação  
615 artificial em bovinos (??rd ed.), ASBIA
- 616 3. Barondes, S. H., Castronovo, V., Cooper, D. N. W., Cummings, R. D., Drickamer,  
617 K., Felzi, T., et al. (1994a). Galectins: a family of animal beta-galactoside-binding  
618 lectins. *Cell*, 76, 597-598. doi: 10.1016/0092-8674(94)90498-7
- 619 4. Barondes, S. H., Cooper, D. N. W., Gitt, M. A., Leffler, H. (1994b). Galectins.  
620 Structure and function of a large family of animal lectins. *Journal of Biological*  
621 *Chemistry*, 269, 20807-20810. doi: 10.1016/S0021-9258(17)31891-4
- 622 5. Barrientos, G., Freitag, N., Tirado-González, I., Unverdorben, L., Jeschke, U.,  
623 Thijssen, V. L. J. L., et al. (2014). Involvement of galectin-1 in reproduction: Past,  
624 present and future. *Human Reproduction Update*, 20:175-193. doi:  
625 10.1093/humupd/dmt040
- 626 6. Baruselli, P. S., Sales, J. N. S., Sala, R., Vieira, L. M., Filho, M. F. S. (2012).  
627 History, evolution and perspectives of timed artificial insemination programs in  
628 Brazil. *Animal Reproduction*, 9, 139-152.
- 629 7. Baruselli, P. S., Catussi, B. L.C., de Abreu, L. A., Elliff, F. M., da Silva, L. G.,  
630 Batista, E. S., et al.. (2019). Evolução e perspectivas da inseminação artificial em  
631 bovinos. *Revista Brasileira de Reprodução Animal*, 43, 308-314.  
632 [http://cbra.org.br/portal/downloads/publicacoes/rbra/v43/n2/p308-](http://cbra.org.br/portal/downloads/publicacoes/rbra/v43/n2/p308-314%20(RB812).pdf)  
633 [314%20\(RB812\).pdf](http://cbra.org.br/portal/downloads/publicacoes/rbra/v43/n2/p308-314%20(RB812).pdf)

- 634 8. Bidarimath, M., Tayade, C. (2017). Pregnancy and spontaneous fetal loss: A pig  
635 perspective. *Molecular Reproduction and Development*, 84(9), 856-869. doi:  
636 10.1002/mrd.22847
- 637 9. Blois, S. M., Dveksler, G., Vasta, G. R., Freitag, N., Blanchard, V., Barrientos, G.  
638 (2019). Pregnancy galectinology: Insights into a complex network of glycan  
639 binding proteins. *Frontiers in Immunology*, 10, 1166. doi:  
640 10.3389/fimmu.2019.01166
- 641 10. Blois, S. M., Ilarregui, J. M., Tometten, M., Garcia, M., Orsal, A. S., Cordo-  
642 Russo, R., et al. (2007). A pivotal role for galectin-1 in fetomaternal tolerance.  
643 *Nature Medicine*, 13(12), 1450-1457. doi: 10.1038/nm1680
- 644 11. Brewer, C. F., Miceli, M. C., Baum, L. G. (2002). Clusters, bundles, arrays and  
645 lattices: novel mechanisms for lectin-saccharide-mediated cellular  
646 interactions. *Current Opinion in Structural Biology*, 12, 616-623. doi:  
647 10.1016/S0959-440X(02)00364-0
- 648 12. Cheng, Z., Abudureyimu, A., Oguejiofor, C. F., Ellis, R., Barry, A. T., Chen, X.,  
649 et al.. (2016). BVDV alters uterine prostaglandin production during pregnancy  
650 recognition in cows. *Reproduction*, 151, 605-614. doi: 10.1530/REP-15-0583
- 651 13. Choe, Y. S., Shim, C., Choi, D., Lee, C. S., Lee, K. K., Kim, K. (1997). Expression  
652 of galectin-1mRNA in the mouse uterus is under the control of ovarian steroids  
653 during blastocyst implantation. *Molecular Reproduction and Development*, 48,  
654 261-266. doi: 10.1002/(SICI)1098-2795(199710)48:2<261::AID-  
655 MRD14>3.0.CO;2-0

- 656 14. Cobuci, J. A., de Abreu, U. G. P., Torres, R. D. A. (2006). *Formação de grupos*  
657 *contemporâneos em bovinos de corte*. Embrapa Pantanal-Documentos. ISSN:  
658 1517-1973
- 659 15. Cross, J. C., Hemberger, M., Lu, Y., Nozaki, T., Whiteley, K., Masutani, M., et  
660 al. (2002). Trophoblast functions, angiogenesis and remodeling of the maternal  
661 vasculature in the placenta. *Molecular and Cellular Endocrinology*, 187, 207-212.  
662 doi: 10.1016/s0303-7207(01)00703-1.
- 663 16. Cummings, R. D., Liu, F. T. (2009). Galectins. In A. C. Varki, R. D. Cummings,  
664 J. D. Esko, H. H. Freeze, P. Stanley, C. R. Bertozzi, G. W. Hart & M. E. Etzler  
665 (Eds.), *Essentials of Glycobiology* (2th ed., chapter 33). Cold Spring Harbor  
666 Laboratory Press. PMID: 20301239
- 667 17. Dias-Baruffi, M., Zhu, H., Cho, M., Karmakar, S., McEver, R. P., Cummings, R.  
668 D. (2003). Dimeric galectin-1 induces surface exposure of phosphatidylserine and  
669 phagocytic recognition of leukocytes without inducing apoptosis. *Journal of*  
670 *Biological Chemistry*, 278, 41282-41293. doi: 10.1074/jbc.M306624200
- 671 18. Diskin, M. G., Morris, D. G. (2008). Embryonic and early fetal losses in cattle  
672 and other ruminants. *Reproduction in Domestic Animals*, 43 (Suppl 2), 260-267.  
673 doi: 10.1111/j.1439-0531.2008.01171.x
- 674 19. Diskin, M., Waters, S., Parr, M., Kenny, D. (2016). Pregnancy losses in cattle:  
675 potential for improvement. *Reproduction, Fertility and Development*, 28, 83-93.  
676 doi: 10.1071/RD15366
- 677 20. Farin, P. W., Piedrahita, J. A., Farin, C. E. (2006). Errors in development of  
678 fetuses and placentas from in vitro-produced bovine embryos. *Theriogenology*,  
679 65, 178-191. doi: 10.1016/j.theriogenology.2005.09.022

- 680 21. Farmer, J. L., Burghardt, R.C., Jousan, F. D., Hansen, P. J., Bazer, F. W., Spencer,  
681 T. E. (2008). Galectin 15 (LGALS15) functions in trophoctoderm migration and  
682 attachment. *FASEB Journal*, 22, 548-560. doi:10.1096/fj.07-9308com
- 683 22. Freitag, N., Tirado-González, I., Barrientos, G., Herse, F., Thijssen, V. L. J. L.,  
684 Weedon-Fekjær, S. M., et al. (2013). Interfering with Gal-1-mediated  
685 angiogenesis contributes to the pathogenesis of preeclampsia. *Proceedings of the*  
686 *National Academy of Sciences of the United States of America*, 110, 11451-11456.  
687 doi: 10.1073/pnas.1303707110
- 688 23. Fukumori, T., Takenaka, Y., Yoshii, T., Kim, H. R. C., Hogan, V., Inohara, H., et  
689 al. (2003). CD29 and CD7 mediate galectin-3-induced type II T-cell  
690 apoptosis. *Cancer Research*, 63, 8302-8311. PMID: 14678989
- 691 24. He, J., Li, X., Luo, H., Li, T., Zhao, L., Qi, Q., et al. (2017). Galectin-3 mediates  
692 the pulmonary arterial hypertension–induced right ventricular remodeling through  
693 interacting with NADPH oxidase 4. *Journal of the American Society of*  
694 *Hypertension*, 11, 275-289. doi: 10.1016/j.jash.2017.03.008
- 695 25. Hughes, R. C. (1999). Secretion of the galectin family of mammalian  
696 carbohydrate-binding proteins. *Biochimica et Biophysica Acta*, 1473, 172-185.  
697 doi: 10.1016/s0304-4165(99)00177-4
- 698 26. Hyde, K. J., Schust, D. J. (2016). Immunologic challenges of human reproduction:  
699 an evolving story. *Fertility and Sterility*, 106, 499-510. doi:  
700 10.1016/j.fertnstert.2016.07.1073
- 701 27. Lamb, G. C., Dahlen, C. R., Larson, J. E., Marquezini, G., Stevenson, J. S. (2010).  
702 Control of the estrous cycle to improve fertility for fixed-time artificial



- 703           insemination in beef cattle: a review. *Journal of Animal Science*, 88 (13 Suppl),  
704           E181–E192. doi: 10.2527/jas.2009-2349
- 705           28. Liu, F. T., Patterson, R. J., Wang, J. L. (2002). Intracellular functions of  
706           galectins. *Biochimica et Biophysica Acta*, 572, 263-273. doi: 10.1016/s0304-  
707           4165(02)00313-6
- 708           29. Liu, F. T., Rabinovich, G. A. (2005). Galectins as modulators of tumor  
709           progression. *Nature Reviews Cancer*, 5, 29-41. doi: 10.1038/nrc1527
- 710           30. Machado, R., Corrêa, R. F., Barbosa, R. T., Bergamaschi, M. A. C. M. (2008).  
711           *Escore da condição corporal e sua aplicação no manejo reprodutivo de*  
712           *ruminantes*. EMBRAPA São Carlos, Brasil Circular Técnica 57. ISSN 1981-2086
- 713           31. Marques, M. O., Morotti, F., Lorenzetti, E., Bizarro-Silva, C., Seneda, M. M.  
714           (2018). Intensified use of TAI and sexed semen on commercial farms. *Animal*  
715           *Reproduction*, 15, 197-203. doi: 10.21451/1984-3143-AR2018-0070
- 716           32. Modenutti, C. P., Capurro, J. I. B., di Lella, S., Martí, M. A. (2019). The Structural  
717           Biology of Galectin-Ligand Recognition: *Current Advances in Modeling Tools,*  
718           *Protein Engineering, and Inhibitor Design*. *Frontiers in Chemistry*, 7, 823.
- 719           33. Moreira, R. (2019, january 01). Aumentar em 1% a taxa de prenhez reduz em 10%  
720           o custo da IATF. *Giro do boi*. [https://www.girodobo.com.br/videos/aumentar-](https://www.girodobo.com.br/videos/aumentar-em-1-na-taxa-de-prenhez-reduz-em-10-o-custo-da-iatf/)  
721           *em-1-na-taxa-de-prenhez-reduz-em-10-o-custo-da-iatf/*
- 722           34. Nickel, W. (2005). Unconventional secretory routes: direct protein export across  
723           the plasma membrane of mammalian cells. *Traffic*, 6, 607-614. doi:  
724           10.1111/j.1600-0854.2005.00302.x
- 725           35. Perillo, N. L., Pace, K. E., Seilhamer, J. J., Baum, L. G. (1995). Apoptosis of T  
726           cells mediated by galectin-1. *Nature*, 378, 736-739. doi: 10.1038/378736a0

- 727 36. Pohler, K., Peres, R., Green, J., Graff, H., Martins, T., Vasconcelos, J., Smith, M.  
728 (2016). Use of bovine pregnancy-associated glycoproteins to predict late  
729 embryonic mortality in postpartum Nelore beef cows. *Theriogenology*, 85, 1652-  
730 1659. doi: 10.1016/j.theriogenology.2016.01.026
- 731 37. Pope, W. (1988). Uterine asynchrony: a cause of embryonic loss. *Biology of*  
732 *Reproduction*, 39, 999-1003. doi: 10.1095/biolreprod39.5.999
- 733 38. Ramhorst, R. E., Giribaldi, L., Fraccaroli, L., Toscano, M. A., Stupirski, J. C.,  
734 Romero, M.D., et al. (2012). Galectin-1 confers immune privilege to human  
735 trophoblast: implications in recurrent fetal loss. *Glycobiology*, 22, 1374-1386.  
736 doi: 10.1093/glycob/cws104
- 737 39. Reese, S. T., Franco, G. A., Poole, R. K., Hood, R., Fernandez-Montero, L.,  
738 Oliveira Filho, R., et al. (2020). Pregnancy loss in beef cattle: A meta-analysis.  
739 *Animal Reproduction Science*, 212, 106251. doi:  
740 10.1016/j.anireprosci.2019.106251
- 741 40. Rubinstein, N., Ilarregui, J. M., Toscano, M. A., Rabinovich, G. A. (2004a). The  
742 role of galectins in the initiation, amplification and resolution of the inflammatory  
743 response. *Tissue Antigens*, 64(1):1-12. doi: 10.1111/j.0001-2815.2004.00278.x
- 744 41. Rubinstein, N., Alvarez, M., Zwirner, N. W., Toscano, M. A., Ilarregui, J. M.,  
745 Bravo, A., et al. (2004b). Targeted inhibition of galectin-1 gene expression in  
746 tumor cells results in heightened T cell-mediated rejection: a potential mechanism  
747 of tumor-immune privilege. *Cancer cell*, 5, 241-251. doi:10.1016/S1535-  
748 6108(04)00024-8

- 749 42. Santos-de-Oliveira, R., Dias-Baruffi, M., Thomaz S. M., Beltramini, L. M.,  
750 Roque-Barreira, M. C. (1994). A neutrophil migration-inducing lectin from  
751 *Artocarpus integrifolia*. *The Journal of Immunology*, 153, 1798-1807.  
752 <https://www.jimmunol.org/content/153/4/1798>
- 753 43. Sharon, N., Lis, H. (1986). Lectin biochemistry. New way of protein maturation.  
754 *Nature*, 323, 203-204. doi: 10.1038/323203a0
- 755 44. Sharon, N., Lis, H. (1989). Lectins as cell recognition molecules. *Science*, 246,  
756 227-234. doi: 10.1126/science.2552581
- 757 45. Schäfer, T., Zentgraf, H., Zehe, C., Brügger, B., Bernhagen, J., Nickel, W. (2004).  
758 Unconventional secretion of fibroblast growth factor 2 is mediated by direct  
759 translocation across the plasma membrane of mammalian cells. *Journal of*  
760 *Biological Chemistry*, 279, 6244-6251. doi: 10.1074/jbc.M310500200
- 761 46. Stowell, S. R., Karmakar, S., Stowell, C. J., Dias-Baruffi, M., McEver, R. P.,  
762 Cummings, R. D. (2007) Human galectin-1,-2, and -4 induce surface exposure of  
763 phosphatidylserine in activated human neutrophils but not in activated T cells.  
764 *Blood*, 109, 219-227. doi: 10.1182/blood-2006-03-007153.
- 765 47. Teichberg, V. I., Silman, I., Beitsch, D. D., Resheff, G. (1975). A beta-D-  
766 galactoside binding protein from electric organ tissue of *Electrophorus electricus*.  
767 *Proceedings of the National Academy of Sciences*, 72, 1383-1387. doi:  
768 10.1073/pnas.72.4.1383
- 769 48. Than, N. G., Romero, R., Erez, O., Weckle, A., Tarca, A. L., Hotra, J., et al.  
770 (2008). Emergence of hormonal and redox regulation of galectin-1 in placental  
771 mammals: implication in maternal–fetal immune tolerance. *Proceedings of the*

- 772            *National Academy of Sciences*, 105, 15819-15824. doi:  
773            10.1073/pnas.0807606105
- 774            49. Vasta, G. R. (2012). Galectins as pattern recognition receptors: structure, function,  
775            and evolution. *Advances in Experimental Medicine and Biology*, 946 21-36. doi:  
776            10.1007/978-1-4614-0106-3\_2
- 777            50. Vergetaki, A., Jeschke, U., Vrekoussis, T., Taliouri, E., Sabatini, L.,  
778            Papakonstanti, E. A., et al. (2014). Galectin-1 overexpression in endometriosis  
779            and its regulation by neuropeptides (CRH, UCN) indicating its important role in  
780            reproduction and inflammation. *Plos One*, 9, e114229. doi:  
781            10.1371/journal.pone.0114229
- 782            51. Yang, R. Y., Hsu, D. K., Liu, F. T. (1996). Expression of galectin-3 modulates T-  
783            cell growth and apoptosis. *Proceedings of the National Academy of Sciences*, 93,  
784            6737-6742. doi: 10.1073/pnas.93.13.6737
- 785            52. Wang, J. L., Gray, R. M., Haudek, K. C., Patterson, R. J. (2004).  
786            Nucleocytoplasmic lectins. *Biochimica et Biophysica Acta*, 1673, 75-93. doi:  
787            10.1016/j.bbagen.2004.03.013
- 788





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