

1
2
3
4
5 **Sex and Estrous Cycle in Memory for Sequences of Events in Rats**

6 Abbreviated Title: SEX AND ESTROUS IN SEQUENCE MEMORY

7
8 M. Jayachandran¹, P. Langius¹, F. Pazos Rego¹, R. P. Vertes², T. A. Allen^{1,2,3}

9
10 ¹Cognitive Neuroscience Program, Florida International University, Miami FL, 33199

11 Correspondence: tallen@fiu.edu

12 ²Center for Complex Systems and Brain Sciences, Florida Atlantic University, Boca Raton, FL 33431, USA

13 ³Department of Environmental Health Sciences, Robert Stempel College of Public Health, Florida International
14 University, Miami, FL 33199, USA

15
16
17
18
19 Key words: sequence memory, episodic memory, temporal context, sex hormones, learning and memory

20
21
22
23 Number of Figures: 5
24 Number of tables: 0
25 Number of Text Pages: 31
26 Title: 15
27 Abstract: 248
28 Introduction: 784
29 Methods: 1622
30 Results: 2931
31 Discussion: 1233
32 Acknowledgement: 59
33 Author Contributions: 38
34 Figure Captions: 864
35 Total: 7780

36
37 Corresponding Author:

38 Timothy A. Allen, PhD
39 Department of Psychology
40 Florida International University
41 11200 SW 8th Street
42 Miami, FL 33199
43 email: tallen@fiu.edu
44
45

ABSTRACT

The ability to remember sequences of events is fundamental to episodic memory. While rodent studies have examined sex and estrous cycle in episodic-like spatial memory tasks, little is known about these biological variables in memory for sequences of events that depend on representations of temporal context. We investigated the role of sex and estrous cycle in rats during all training and testing stages of a cross-species validated sequence memory task (Jayachandran et al., 2019). Rats were trained on a task composed of two sequences, each with four unique odors delivered on opposite ends of a linear track. Training occurred in six successive stages starting with learning to poke in a nose port for ≥ 1.2 s; eventually demonstrating sequence memory by holding their nose in the port for ≥ 1 s for in-sequence odors and < 1 s for out-of-sequence odors in order to receive a water reward. Performance was analyzed across sex and estrous cycle (proestrus, estrus, metestrus, and diestrus), the latter being determined by the cellular composition of a daily vaginal lavage. We found no evidence of sex differences in asymptotic sequence memory performance, similar to published data in humans performing the analogous task (Reeders et al., 2021). Likewise, we found no differences in performance across the estrous cycle. One minor difference was that female rats tended to have slightly longer poke times, while males had slightly more short poke times but this did not affect their decisions. These results suggest sex and estrous cycle are not major factors in sequence memory capacities.

Word Count: 249 (max: 250)

INTRODUCTION

Episodic memory is the ability to encode and recollect our daily personal experiences (Tulving, 2002; Allen & Fortin, 2013). A fundamental feature of episodic memory is the ability to remember sequences of events as they occurred across time (Allen et al., 2020; Allen et al., 2014, 2016; Eichenbaum & Fortin, 2005; Jayachandran et al., 2019). While rodent studies have examined sex and estrous cycle as variables in spatial and episodic-like memory tasks, little is known about these biological variables in a sequence memory task that depends on representations of temporal contexts. The study of sex and sex hormones in sequence memory is particularly important in understanding the neurobiology of mental health disorders because of known sex differences and the prevalence of deficits in temporal cognition (Li & Singh, 2014; Postma et al., 2004; Valera et al., 2010) in disorders such as schizophrenia and Alzheimer's disease (Kessler et al., 2005; Launer, 1999; Pigott, 2003).

Sex differences in memory have been widely reported in human and nonhuman animals alike (Levy et al., 2005; Seymoure et al., 1996; Williams et al., 1990). For example, rodent studies have shown that sex impacts both spatial cognition and object recognition (Barha et al., 2010; Hamson et al., 2016; Koss & Frick, 2017; Luine et al., 2017; Spritzer et al., 2008). Most commonly, evidence suggests that human and rodent males have an advantage in tests of spatial memory (Driscoll, 2005; Jonasson, 2005; Nowak et al., 2014; Steimer & Woolley et al., 2010). Importantly, when strategy is considered or a landmark, such as a wall cue is included, these sex differences disappear (Chai & Jacobs, 2010; Saucier et al., 2008). Alternatively, females tend to outperform males in object memory (Bettis & Jacobs, 2012; Levy et al., 2005; Sutcliffe et al., 2007). However, not every study finds sex differences in spatial learning and memory tasks (Bucci et al., 2021; Frick et al., 1999; Healy et al., 1999).

When looking at sex differences in memory, it is important to consider the bioavailability of key sex hormones. For example, exogenous ovarian hormones have been shown to enhance performance in object memory in both rodent and human females (Walf & Frye, 2006). This suggests the sex differences in memory can be dependent on the levels of circulating gonadal hormones. Important to our experiments here, hormone levels change reliably during the estrous cycle in females and different phases have been linked to differences in memory. For example, Healy and colleagues, (1999) showed that female rodents

00 were slowest to locate the platform in a Morris water maze task during estrus and were fastest during
01 proestrus which was attributed to a core spatial memory difference. Likewise, in other spatial tasks, females
02 demonstrate the use of allocentric strategies during proestrus and egocentric/mixed strategies during
03 diestrus (Korol, 2004; Koss & Frick, 2017). In object memory tasks, rats in proestrus outperformed rats in
04 diestrus or metestrus (Koss & Frick, 2017; Van Goethem et al., 2012). By contrast to these aforementioned
05 studies not all studies find differences in memory across the estrous cycle. For example, Stackman et al.,
06 (1997) did not observe significant differences in acquisition or performance levels across the estrous cycle
07 when testing working memory using a radial arm maze; however, they showed that rats performed the task
08 significantly slower during proestrus than on any other estrous cycle phase. This suggests that while the
09 underlying working memory process remained stable, other aspects of performance can vary across the
10 estrous cycle.

11 Here, we investigated the role of sex and estrous cycle (proestrus, estrus, metestrus, diestrus) in rats
12 during training and testing stage of an odor sequence memory task (Jayachandran et al., 2019). Rats were
13 trained for several months on the sequence memory task composed of two four-odor sequences delivered
14 at opposite ends of a linear track. Training occurred progressively across six major stages starting with
15 learning to poke and hold for ≥ 1.2 s for a small water reward to eventually differentiating in-sequence (InSeq)
16 and out-of-sequence (OutSeq) odors, one at a time, in four odor sequence sets. Performance across
17 training stages was analyzed across sex and estrous cycle phases, the latter of which was determined by
18 the cellular composition of a vaginal lavage. We conducted an extensive analysis of behavior at each stage
19 including trials to criterion, self-paced inter-odor interval times, self-paced inter-sequence interval times,
20 poke times under different sequential and accuracy conditions, and overall sequence memory. We found no
21 evidence that males and females differed in learning, remembering, or performing the sequence memory
22 task. Moreover, we did not find any differences across the estrous cycle in sequence memory. The results
23 suggest sex and estrous cycle are not major factors in memory for sequences of events.

24

25

METHODS

26 **Subjects:** Twenty Long-Evans rats (10 females; Charles River Laboratories; weighing 250-350g upon
27 arrival) were used. Two male rats failed to progress past the TwoOdor stage of training and were excluded
28 from all analysis. Rats were individually housed and maintained on a 12-hr inverse light/dark cycle (lights
29 were turned off at 10 AM). Rats had *ad libitum* access to food, but access to water was limited to 3 - 5min
30 each day, depending on how much water they received as a reward during behavioral training (6 - 9mL). All
31 training sessions and lavages were conducted during the dark phase (active period) of the light cycle. All
32 experimental procedures using animals were conducted in accordance with the Florida International
33 University Institutional Animal Care and Use Committee (FIU IACUC).

34

35 **Task Apparatus:** Rats were tested in a noise-attenuated experimental room. The behavioral apparatus
36 consisted of a linear track (length, 183cm; width, 10cm; height, 43cm) with walls angled outward at 15° and
37 nose ports centered at each end through which repeated deliveries of multiple distinct odors could be
38 presented. Photobeam sensors were used to detect nose port entries. Each nose port was connected to an
39 odor delivery system (Med Associates; Fairfax, VT). Odor deliveries were initiated by a nose-poke entry and
40 terminated either when the rat withdrew their poke or after 1s had elapsed. Water ports were positioned
41 under each nose port for reward delivery. Timing boards (Plexon; Dallas, TX) and digital input/output
42 devices (National Instruments; Austin, TX) were used to measure all event times and control the hardware.
43 All aspects of the task were automated using custom MATLAB scripts (MathWorks 2021a; Natick, MA). A
44 256-channel OmniPlex with video tracking and CinePlex behavioral software (Plexon; Dallas, TX) were used
45 to interface with the hardware in real time and record behavioral data. Odors were organic odorants
46 contained in glass jars (Sigma-Aldrich; St. Louis, MO, A1: 1-octanol, CAS: 111-87-5; B1: (-) - limonene,
47 CAS: 5989-54-8; C1: l-menthone, CAS: 14073-97-3; D1: isobutyl alcohol, CAS: 78-83-1; A2: acetophenone,
48 CAS: 98-86-2; B2: (1S) - (-) – beta-pinene, CAS: 18172-67-3; C2: L (-) - carvone, CAS: 6485-40-1; D2: 5-
49 methyl-2-hexanone, CAS: 110-12-3) that were volatilized with nitrogen air (flow rate, 2L/min) and diluted
50 with ultrapure air (flow rate, 1L/min). To prevent cross-contamination, separate Teflon tubing lines were
51 used for each odor. These lines converged into a single channel at the bottom of the odor port. In addition, a

52 vacuum located at the top of the odor port provided constant negative pressure to quickly evacuate odor
53 traces with a matched flow rate.

54

55 **Sequence Memory Task:** The sequence memory task (Jayachandran et al., 2019; Allen et al., 2014)
56 involves repeated presentations of odor sequences and requires the rat to determine whether each item
57 (odor) presented is in-sequence (InSeq) or out-of-sequence (OutSeq). Rats were trained on two sequences,
58 each comprised of four distinct odors (e.g., Seq1: A₁B₁C₁D₁, Seq2: A₂B₂C₂D₂). Each sequence was
59 presented at either end of a linear track maze. Odor presentations were initiated by a nose-poke, and each
60 trial was terminated after the rat either held the nose-poke response for ≥1s (InSeq) or withdrew its nose-
61 poke response <1s (OutSeq). Trials were separated by 1s intervals. Water rewards (20μl; diluted at 1g of
62 aspartame for every 500mL of water) were delivered below the odor port after each correct response.
63 Incorrect responses triggered a buzzer sound and the termination of the sequence. Each sequence was
64 presented 50-100 times per session; approximately half the presentations included all items InSeq (ABCD)
65 and half included one item OutSeq (e.g., ABAD, odor A repeated in the 3rd position). Note that OutSeq
66 items could be presented in any sequence position except the first position (i.e., sequences always began
67 with an InSeq item). Sequence memory was probed with OutSeq trials (e.g., ABAD; one OutSeq trial
68 randomly presented per sequence).

69

70 **Sequence Memory Task Training:** Naive rats were initially trained on a series of incremental stages over
71 20-30 weeks. Each rat was trained to poke and hold its nose in an odor port to receive a small water reward.
72 The minimum required nose-poke duration started at 50ms and was gradually increased (in 15ms
73 increments) until the rat held the nose-poke position for ≥1.2s ~ 75% of the time over three sessions (75-100
74 nose-pokes per session). The rats were then trained to poke on side 2 until reaching criterion; poking for
75 ≥1.2s with ~75% accuracy for three consecutive sessions. The rats were then habituated to odor
76 presentations in the port (odor A₁ and A₂). The hold time was reduced down to 1 s and rats were required to
77 alternate between sides for a small water reward at each side for ~75% accuracy for three consecutive
78 sessions before moving on. Then a second set of odors was introduced (Seq1: A₁B₁ and Seq2: A₂B₂) and

79 each rat was required to maintain its nose-poke response for ≥ 1 s to receive a reward and needed to
80 achieve $\sim 70\%$ accuracy before moving on. The rats were next trained to identify InSeq and OutSeq items.
81 Rats began this stage trained on a two-item sequence: they were presented with “AB” and “AA”
82 sequences. The correct response to the first odor was to hold their nose-poke for ≥ 1 s (Odor A always being
83 the first item). For the second odor, rats were required to determine whether the item was InSeq (AB; hold
84 for ≥ 1 s to receive reward) or OutSeq (AA; withdraw before 1s to receive a reward). After reaching criterion
85 on the two-item sequence, the number of items per sequence was increased to three odors each side
86 (Seq1: A₁B₁C₁ and Seq2: A₂B₂C₂), and four odors each side (Seq1: A₁B₁C₁ D₁ and Seq2: A₂B₂C₂D₂) in
87 successive stages. The criterion for these stages set percentages as indicated in the results or the ability
88 differentiate InSeq and OutSeq items above a sequence memory index (SMI; see quantitative and statistical
89 analysis) of 0.2 over three consecutive sessions. Once rats reached criterion for the last stage of training, 15
90 consecutive sessions were collected to examine both sex and estrous cycle differences in overall SMI.

91

92 **Lavaging:** Phases of the estrus cycle were determined in each female rat using vaginal lavage within four
93 hours of each behavioral session. Rats were restrained using a designated towel and a drop of sterilized
94 saline (NaCl, 0.85%) was placed over the vaginal opening to clean the area. The tip of a sterilized transfer
95 pipette filled with sterilized saline ($\sim 0.25 - 0.5$ mL) was inserted into the vaginal opening to extract vaginal
96 cells. All vaginal smears were placed on glass slides and photomicrographs were taken under brightfield
97 illumination using an Olympus BX41 following guidelines from Westwood (2008). To control for
98 experimenter handling, male rats were handled similarly and poked around the anal opening with a clean
99 transfer pipette.

00

01 **Estrous Cycle Cytology:** Estrous cycles in rats typically last four to five days and have four distinct phases:
02 proestrus, estrus, metestrus and diestrus; each phase corresponds with specific cell types, i.e., nucleated
03 epithelial cells, amorphous cornified cells, and small round leukocytes. Proestrus is characterized
04 predominantly by nucleated epithelial cells, estrus predominantly amorphous cornified cells, metestrus by a
05 mixture of nucleated epithelial cells, cornified cells and leukocytes, and diestrus predominantly leukocytes.

06 All vaginal cell samples were obtained and imaged, then the images were blindly classified by three
07 investigators.

08

09 **Quantification and Statistical Analysis:**

10 All data was analyzed in MATLAB 2021a (MathWorks; Natick, MA), SPSS 20.0.0, and Excel 2016 using
11 custom scripts and functions. Performance on the task can be analyzed using a number of measures (Allen
12 et al., 2014). The first position of each sequence was excluded from all analysis as these items are always
13 InSeq. Expected vs. observed frequencies were analyzed with G tests to determine whether the observed
14 frequencies of InSeq and OutSeq responses for a given session were significantly different from the
15 frequency expected by chance. G tests provide a measure of performance that controls for response bias
16 and is a robust alternative to the χ^2 test, especially for datasets that include cells with smaller frequencies
17 (Sokal & Rohlf, 1995). To compare performance across sessions or animals, a SMI was calculated (Allen et
18 al., 2014) as shown in the following equation:

$$19 \quad SMI = \frac{(0.9 * IN_{out})(0.1 * OUT_{cor}) - (0.9 * IN_{inc})(0.1 * OUT_{inc})}{\sqrt{(0.9 * IN_{cor} + 0.9 * IN_{inc})(0.1 * OUT_{cor} + 0.1 * OUT_{inc}) * (0.9 * IN_{cor} + 0.1 * OUT_{inc})(0.9 * IN_{inc} + 0.1 * OUT_{cor})}}$$

20 The SMI normalizes the proportion of InSeq and OutSeq items presented during a session and reduces
21 sequence memory performance to a single value ranging from -1 to 1. A score of 1 represents a perfect
22 sequence memory performance in which a subject would have correctly held their nose-poke response to all
23 InSeq items and correctly withdrawn on all OutSeq items. A score of 0 indicates chance performance.
24 Negative SMI scores represent performance levels below that expected by chance. SMI was calculated for
25 males and females as well as between estrous phases during the two, three, and four odor stages of the
26 sequence task. Using SPSS, an independent t-test was used to determine if males and females differed in
27 their overall SMI. We went on to examine the differences between sequence 1 and sequence 2 within sex
28 and estrous cycle by running a two-way ANOVA. A one-way ANOVA was used to determine if overall SMI
29 differed within female estrous cycle. Linear regressions were performed in increments of pre-determined set
30 trials or sessions to compare males and females in order to determine learning at each stage of training on
31 the sequence task. Independent sample t-tests were used to analyze whether males and females differed in

32 self-paced inter-trial-interval (ITI), inter-odor-interval (IOI), and inter-sequence interval (ISI). Nose-poke
33 duration was analyzed using independent t-tests to determine whether rats held their responses significantly
34 longer in InSeq_{correct} than in OutSeq_{correct} trials. General poke distributions were created through MATLAB
35 using the session data.

36 RESULTS

37 Overall Sequence Memory Task and Training

38 We trained male ($n = 10$) and female ($n = 10$) rats in an odor sequence memory task (Jayachandran et
39 al., 2019). The behavioral setup was automated and allowed repeated delivery of pure chemical odorants
40 between two odor ports located at opposite ends of a linear track (Figure 1A). The rats were trained to
41 indicate if an odor is InSeq by holding their nose in the nose-port for ≥ 1 s (Figure 1Bi) or OutSeq by
42 withdrawing their nose prior to 1 s for a small water reward (Figure 1Bii). Rats were trained in progressive
43 stages over several months on the sequence task, as depicted in Figure 1C. At each stage of training on the
44 sequence task, rats had to reach a specific performance criterion in order to progress to the next stage.
45 Once they reached the final stage (FourOdors) and behavioral criterion, we collected 15 consecutive
46 sessions to analyze overall sequence memory. Two male rats were not able to continue past TwoOdors and
47 therefore we removed them from the overall analysis. We calculated an SMI to measure overall sequence
48 memory while controlling for individual differences in poke-hold behavior (Allen et al., 2014). We averaged
49 the overall number of sessions per stage of training between males (Poke&Hold_{Side1}: 28.125 ± 2.601 ;
50 Poke&Hold_{Side2}: 8.625 ± 1.731 ; OneOdor: 15.500 ± 2.706 ; TwoOdors: 49.375 ± 8.268 ; ThreeOdors: 33.500
51 ± 8.203 ; FourOdors: 30.625 ± 1.499) and females (Poke&Hold_{Side1}: 22.800 ± 1.504 ; Poke&Hold_{Side2}: $8.400 \pm$
52 1.962 ; OneOdor: 13.300 ± 1.506 ; TwoOdors: 40.600 ± 4.861 ; ThreeOdors: 30.800 ± 3.782 ; FourOdors:
53 30.400 ± 0.957). There were no significant differences in the number of sessions to criterion for stage when
54 comparing males and females (Poke&Hold_{Side1}: $t_{(16)} = -1.861$, $p = 0.081$; Poke&Hold_{Side2}: $t_{(16)} = -0.084$, $p =$
55 0.934 ; OneOdor: $t_{(16)} = -0.749$, $p = 0.465$; TwoOdors: $t_{(16)} = -0.959$, $p = 0.352$; ThreeOdors: $t_{(16)} = -0.320$, $p =$
56 0.753 ; FourOdors; $t_{(16)} = -0.131$, $p = 0.897$).

58 Males and Females Parallel in Poking Behaviors

59 Rats were first trained to poke and hold in the nose port located on side 1 of the sequence memory task
60 (Figure 2Ai). During this stage, the rat is trained to hold their nose in the nose port with a start hold time of
61 50ms. With each correct trial, the hold time was increased by 15ms until reaching a 1.2s hold time. The rats
62 were then required to poke for ≥ 1.2 s at $\sim 75\%$ accuracy for 3 consecutive sessions. We first examined the
63 learning rates of males and females by calculating the slope over 200 trial increments (Figure 2Aii). Males
64 and females did not show any significant differences in learning during $\text{Poke\&Hold}_{\text{side1}}$ (Trials_{1-200} : $t_{(16)} =$
65 1.075 , $p = 0.299$; $\text{Trials}_{200-400}$: $t_{(16)} = 0.850$, $p = 0.262$; $\text{Trials}_{400-600}$: $t_{(16)} = 0.336$, $p = 0.741$; $\text{Trials}_{600-800}$: $t_{(16)} =$
66 0.840 , $p = 0.413$; $\text{Trials}_{800-1000}$: $t_{(16)} = 0.754$, $p = 0.462$). A few rats exceeded 1000 trials to progress to the
67 next stage of the task; however, due to too few values we could not calculate an average past 1000 trials.
68 We then looked at the poke distributions between males and females for all trials. Overall, the proportion of
69 nosepokes remained similar between males and females with a slight increase for males on short pokes
70 (Figure 2Aiii). We performed several analyses to test the alternative hypothesis that non-memory-related
71 behavioral effects account for impaired performance in the sequence task. We measured the time it took
72 rats to initiate each poke (inter-trial-interval; ITI). The ITI did not differ significantly between males and
73 females (Figure 2Aiv; $t_{(410)} = -0.174$, $p = 0.862$).

74 Once reaching criterion for $\text{Poke\&Hold}_{\text{side1}}$ rats were then trained on $\text{Poke\&Hold}_{\text{side2}}$ where rats were
75 familiarized with the odor port on side 2 and trained to poke for ≥ 1.2 s at $\sim 75\%$ accuracy for 3 consecutive
76 sessions (Figure 2Bi). We calculated the proportion correct for each session after poke hold times became
77 consistent. We examined learning between males and females by measuring proportion correct and
78 calculating the rate of change in proportion correct (Figure 2Bii; 5 session increments). A few rats continued
79 past 10 sessions; however we were not able to calculate an average due to the low sample sizes after this
80 point. Males and females did not show any significant differences in learning (Sessions_{1-5} : $t_{(16)} = -0.545$, $p =$
81 0.593 ; Sessions_{5-10} : $t_{(7)} = -0.124$, $p = 0.905$). When we looked at poke distributions, the proportion of nose-
82 pokes between males and females remained similar (Figure 2Biii). However, we do note that males seem to
83 have more short pokes (~ 0.1 s) while females tended to hold longer (~ 1.4 s). We then examined ITI and
84 found no significant differences between males and females (Figure 2Biv; $t_{(142)} = -0.258$, $p = 0.797$).

85 Once past Poke&Hold_{side2}, rats were then moved to OneOdor which began with habituating to one odor
86 presentation on either end of the linear track. The hold time criterion was also reduced down to 1 s and rats
87 were required to alternate between sides (Figure 2Ci). Rats were required to reach ~75% accuracy for 3
88 consecutive sessions at this stage. We found no significant differences in learning between males and
89 females (Sessions₁₋₅: $t_{(16)} = -1.123$, $p = 0.278$; Sessions₅₋₁₀: $t_{(16)} = 1.034$, $p = 0.317$; Sessions₁₀₋₁₅: $t_{(7)} = 0.774$,
90 $p = 0.464$). We then looked at poke distributions between males and females; overall, we found they were
91 very similar. Again, we observed males' tendencies towards short pokes (~0.1 - 0.2s) with females'
92 tendencies towards longer pokes (~1.2 - 1.5s). At this stage we measured the time it took the rats to run
93 between sequences which we refer to as inter-sequence-interval (ISI). We found no significant differences in
94 ISI between males and females (Figure 2Civ; $t_{(224)} = 0.655$, $p = 0.513$). Overall, these results suggest that
95 males and females show little to no differences in their poking and timing behaviors.

96

97 **Males and Females Perform Similarly on the Sequence Memory Task**

98 After reaching criterion for the OneOdor stage of training, rats are then moved onto TwoOdors where
99 they are trained to poke for two separate odors on each side (Figure 3Ai; Seq1: A₁B₁ and Seq2: A₂B₂). Once
00 rats are able to poke twice on each side with ~70% accuracy, they are then trained to identify InSeq and
01 OutSeq odors. The correct response to the first odor is always to hold the nose-poke for ≥1s (Odor A was
02 always the first item). For the second odor, rats were required to determine whether the item was InSeq
03 (AB; hold for 1 s to receive reward) or OutSeq (AA; withdraw before 1s to receive a reward). Rats were
04 trained until they demonstrated sufficient performance (≥ 0.2 SMI) for three consecutive sessions. We
05 calculated the SMI once OutSeq trials were introduced and examined the overall learning between males
06 and females. Although we refer to memory in this stage as sequence memory, it is more equivalent to a
07 delayed-to-non-match sample task which exclusively measures working memory. Position 1 was excluded
08 because an OutSeq item was never presented in that position. Twenty sessions were used to measure
09 learning at this stage. While some rats continued to learn past 20 sessions, there were not enough subjects
10 to calculate an average. Generally, we found no significant differences in learning between males and
11 females (Sessions₁₋₅: $t_{(16)} = 1.140$, $p = 0.227$; Sessions₅₋₁₀: $t_{(16)} = 0.685$, $p = 0.503$; Sessions₁₀₋₁₅: $t_{(15)} = -$

12 0.938, $p = 0.363$; Sessions₁₅₋₂₀: $t_{(10)} = -0.394$, $p = 0.702$). We also examined the overall SMI of the 3
13 consecutive stages in which criterion was met. We found no significant difference between the sexes
14 (Figure 3Aiii; $t_{(52)} = 1.134$, $p = 0.262$, Male: 0.347 ± 0.022 , Female: 0.386 ± 0.026). Further, we examined
15 whether there were differences between sequence 1 and sequence 2. A two-way ANOVA revealed no
16 statistically significant interaction between sex and sequence ($F_{(3,104)} = 0.031$, $p = 0.860$). Simple main effect
17 analysis showed that sex and sequence did not have a statistically significant effect on SMI (Sex: $p = 0.304$;
18 Sequence: $p = 0.643$).

19 We examined the nose-poke distributions for both InSeq and OutSeq trials (Figures 3Av and 3Avi). The
20 InSeq distribution showed the proportion of nose-pokes in males and females remained similar. On OutSeq
21 trials, however, we see a modest difference between male and female poke times. Females tended to poke
22 for OutSeq trials close to the 1s threshold ($\sim 0.8 - 0.99$ s) whereas males continued to show short pokes
23 ($\sim 0.1 - 0.4$ s) in both InSeq and OutSeq trials. We evaluated whether males and females differed in nose-
24 poke times on InSeq_{correct} and OutSeq_{correct} trials (Figure 3Avii). No significant differences were detected
25 between males and females for InSeq_{correct} trials ($t_{(373)} = 1.195$, $p = 0.233$). However, in OutSeq_{correct} trials we
26 found a significant difference, with females showing a slightly longer hold time ($t_{(373)} = 9.609$, $p = 1.119 \times 10^{-19}$;
27 Male: 0.446 ± 0.020 ; Female: 0.678 ± 0.014) which corresponds with what we observed in the poke
28 distribution. We then looked at the time rats spent between each odor trial (inter-odor-interval; IOI) and ISI.
29 The IOI did not differ significantly between males and females (Figure 3Aviii; $t_{(767)} = 1.495$, $p = 0.135$),
30 suggesting rats collected water rewards and engaged with the odors at similar rates. Furthermore, we found
31 no effect by group on the ISI (Figure 3Aix; $t_{(757)} = 0.312$, $p = 0.755$), suggesting rats in both groups
32 alternated at similar rates between sequences.

33 Once rats reach criterion in the TwoOdor training stage, they are moved to ThreeOdors. In this stage,
34 one more odor is added to each sequence (Figure 3Bi; Seq1: A₁B₁C₁ and Seq2: A₂B₂C₂). Rats were trained
35 to poke three times on either side with at least $\sim 65\%$ accuracy before being introduced to the OutSeq probe
36 trials. With this three-item sequence the number of possible OutSeq configurations has expanded, making
37 this the first stage to test sequence memory. Rats were trained up until they demonstrated sufficient
38 sequence memory (≥ 0.2 SMI) for three consecutive trials to move onto the final training stage. We

39 averaged the first twenty sessions to measure learning. We found a significant difference in the first five
40 sessions where males seemed to learn at a faster rate compared to females (Figure 3Bii; Sessions₁₋₅: $t_{(16)} =$
41 -2.471 , $p = 0.025$). However, this difference diminished with the subsequent sessions (Sessions₅₋₁₀: $t_{(14)} =$
42 1.131 , $p = 0.277$; Sessions₁₀₋₁₅: $t_{(12)} = -0.037$, $p = 0.971$; Sessions₁₅₋₂₀: $t_{(9)} = -0.206$, $p = 0.842$).

43 We then examined the overall SMI using data from after subjects had reached the behavioral criterion.
44 We found no significant differences between males and females (Figure 3Biii; $t_{(52)} = -1.755$, $p = 0.085$, Male:
45 0.428 ± 0.015 , Female: 0.394 ± 0.012). We also observed no significant interactions between sex and
46 sequence ($F_{(3,104)} = 0.059$, $p = 0.808$). A simple main effect analysis showed that sex and sequence did not
47 have a statistically significant effect on SMI (Sex: $p = 0.094$; Sequence: $p = 0.182$).

48 We then evaluated the nose-poke distributions for both InSeq and OutSeq trials (Figure 3Bv and 3Bvi).
49 The InSeq distribution shows a similar proportion of nose-pokes in males and females. The OutSeq trials
50 showed that males poked between $\sim 0.4 - 0.7$ s whereas females tended to poke between $\sim 0.8 - 0.95$ s. We
51 also saw more short pokes from males in both InSeq and OutSeq trials. We evaluated non-mnemonic
52 behaviors by examining nose-poke behaviors, IOI, and ISI. Additionally, InSeq_{correct} nose-poke times
53 between males and females did not significantly differ (Figure 3Bvii; $t_{(344)} = 1.718$, $p = 0.087$). There was a
54 significant difference in OutSeq_{correct} trials with females holding longer (Figure 3Bvii; $t_{(344)} = 5.619$, $p = 3.965$
55 $\times 10^{-8}$; Males: 0.501 ± 0.021 ; Females: 0.643 ± 0.015). The IOI and ISI did not differ significantly between
56 males and females (Figure 3Bviii and 3Bix; IOI: $t_{(562)} = 1.568$, $p = 0.118$; ISI: $t_{(562)} = 1.431$, $p = 0.153$).
57 Overall, these results suggest that males and females do not differ in working memory. However, males
58 show a slight advantage in learning the OutSeq rule during the ThreeOdor training stage.

59 Finally, upon successfully completing the ThreeOdor training stage, an additional odor is introduced to
60 each sequence (Seq1: A₁B₁C₁D₁ and Seq2: A₂B₂C₂D₂). Rats were trained until they achieved $\sim 65\%$
61 accuracy in differentiating between InSeq and OutSeq (≥ 0.2 SMI). Once this criterion was met, we then ran
62 the rats for 15 consecutive sessions (Figure 4). We took the averaged results from 25 sessions after the
63 task was learned in order to describe overall learning and performance. We found no significant difference
64 in learning between males and females. (Figure 4B; Sessions₁₋₅: $t_{(16)} = 0.221$, $p = 0.828$; Sessions₅₋₁₀: $t_{(16)} = -$
65 0.176 , $p = 0.862$; Sessions₁₀₋₁₅: $t_{(16)} = -0.214$, $p = 0.833$; Sessions₁₅₋₂₀: $t_{(16)} = -0.437$, $p = 0.668$; Sessions₂₀₋₂₅:

66 $t_{(14)} = 1.701, p = 0.111$). By session ten, most rats reached behavioral criterion (asymptotic sequence
67 memory performance levels over multiple sessions). Overall, rats demonstrated strong sequence memory
68 (Figure 4C; Male: $0.262 \pm 8.690 \times 10^{-3}$, Female: $0.272 \pm 8.120 \times 10^{-3}$) and performance did not differ
69 significantly between males and females (Figure 4Ci; SMI: $t_{(268)} = 0.784, p = 0.434$). Both males and females
70 performed well above chance levels on sequence 1 (Male: 0.268 ± 0.017 ; Female: 0.280 ± 0.018) and
71 sequence 2 (Male: 0.279 ± 0.016 ; Female: 0.318 ± 0.018), with no significant interactions between
72 sequences or sex (Figure 4Cii; $F_{(1,479)} = 0.608, p = 0.436$). Simple main effect analysis showed that sex and
73 sequence did not have a statistically significant effect on SMI (Sex: $p = 0.160$; Sequence: $p = 0.177$),
74 indicating rats successfully switched between the two sequences.

75 We evaluated non-mnemonic effect of sex by examining IOI and ISI. The IOI and ISI did not differ
76 significantly between males and females (Figure 4D and 4E; IOI: $t_{(278)} = -0.569, p = 0.570$; ISI: $t_{(278)} = 0.162,$
77 $p = 0.871$). We analyzed the poke distribution between males and females (Figure 4Fi and Fii). The InSeq
78 and OutSeq distribution showed the proportion of nose-pokes remained similar between males and females.
79 However, there was a slight increase in the proportion of InSeq and OutSeq nose-pokes near the short
80 distribution peak for males ($\sim 0.1 - 0.2$ s). We also examined whether sex affected nose-poke times on
81 InSeq_{correct} and OutSeq_{correct} trials (Figure 4G). No significant differences were detected in InSeq_{correct} trials
82 ($t_{(268)} = -1.925, p = 0.055$). In the OutSeq_{correct} trials there was a significant increase in nose-poke time for
83 females ($t_{(268)} = 3.617, p = 3.560 \times 10^{-4}$; Male: 0.443 ± 0.020 ; Female: 0.544 ± 0.019). Overall, these results
84 demonstrate that males and females do not differ in their overall sequence memory. However, there are
85 slight differences in their hold times for OutSeq trials, which may indicate uncertainty regarding whether a
86 trial was InSeq or OutSeq rather than a deficit related to basic nose-poke behavior or sequence memory.

87

88 **Estrous Cycle in Sequence Memory**

89 Generally, the results demonstrate that males and females do not differ in learning and performance on
90 the sequence memory task. We then looked at estrous cycle phases within females, as the fluctuations of
91 female hormone levels are important factors to consider when working with female animals. Figure 5A
92 illustrates the hormone level in female rats during each of the four phases of the estrous cycle. In the

93 proestrus phase, ovulation begins with the sharp increase and decrease of progesterone, increase of
94 estradiol, and decrease of luteinizing hormone (LH). In the estrus phase, known as the “heat phase”,
95 progesterone remains steady, estradiol peaks and begins to decrease while LH decreases. In the metestrus
96 phase, all hormones remain steady. Finally, in the diestrus phase LH begins to increase slowly.

97 We first examined the overall SMI once rats reach asymptotic levels on the fully learned sequence
98 memory task. We found no significant differences between the estrous cycle phases (Figure 5Bi; $F_{(3, 146)} =$
99 $1.405, p = 0.244$). We also observed no significant differences between estrous and sequence (Figure 5Bii:
00 $F_{(7, 270)} = 3.56, p = 0.078$). Simple main effect analysis showed that estrous and sequence were not
01 statistically significant (Estrous: $p = 0.433$; Sequence: $p = 0.322$). We then observed the general poke
02 distributions for InSeq and OutSeq trials between the estrous cycle phases. For InSeq trials, all estrous
03 cycle phases showcased a cluster of pokes slightly after the 1 s decision threshold (Figure 5Ci). For OutSeq
04 trials, there are two main peaks of nose-pokes around 0.2 s and 0.8 s (Figure 5Cii). We did observe that
05 there was a slight increase in poke times during estrus (~0.2 – 0.3s) in comparison to the other three
06 phases. Next, we evaluated the nose-poke times for InSeq and OutSeq correct trials. There were no
07 significant differences in InSeq_{correct} trials between the estrous cycle phases (Figure 5D; $F_{(3, 149)} = 0.667, p =$
08 0.573). There was a significant difference in OutSeq_{correct} trials (Figure 5D; $F_{(3, 149)} = 3.301, p = 0.022$). Post
09 hoc comparisons using Bonferroni test revealed a significant difference between estrus and metestrus ($p =$
10 0.033) with a mean difference poke time of 0.177s. We explored non-mnemonic effects relevant to the
11 sequence task. We looked at IOI and ISI and found no significant differences between the estrous cycle
12 phases within those conditions (Figure 5E and 5F: IOI: $F_{(3, 149)} = 0.676, p = 0.568$; ISI: $F_{(3, 149)} = 0.416, p =$
13 0.742). These results that the estrous cycle does not strongly affect overall sequence memory performance.

14

15

DISCUSSION

16

Summary of Main Findings

17

18

19

We evaluated the role of sex in rats during training and testing phases of a sequence memory task. Although this task has been used in previous studies (Allen et al., 2014, 2016a; Jayachandran et al., 2019). here we describe the stages of training and its performance criterion for the first time. Overall, both male

20 and female rats learned the sequence memory task at relatively the same rate. However, there were a few
21 differences between males and females. Males tended to have a modest number of shorter poke times
22 compared to females, which was more evident once OutSeq trials were introduced, but this did not affect
23 overall accuracy. Moreover, males seemed to learn at a somewhat faster rate once the sequence was
24 expanded to include more than one OutSeq position (ThreeOdor) but this effect did not extend to the full
25 sequences. In general, there were no noticeable sex differences in sequence memory once the task was
26 fully learned. This finding alone, however, did not address whether the lack of sex differences could have
27 been confounded by female rats' estrous cycle. Therefore, we looked at estrous cycle specifically and found
28 that regardless of phase, females performed similarly well on the sequence memory task.

29 **No Evidence that Sequence Memory Differs in Males and Females**

30 To investigate sex differences in sequence memory, we examined males and females at each training
31 stage of the sequence task. One main difference we observed was with the introduction of the three-item
32 sequence. At this stage, we found males had a faster learning curve compared to females. This advantage
33 could be due to males' tendency for short pokes, allowing them more opportunities to have chance correct
34 responses for OutSeq trials and thus providing a modest boost in discovering the rules. This may not reflect
35 sex differences in learning per se, but rather sex differences in strategies (Gruene et al., 2015). This has
36 been suggested before for spatial learning paradigms, in which male rats outperform females by using a
37 more direct strategy (McCarthy & Konkle, 2005). This difference could also be due to Type I family-wise
38 error. As multiple tests were run at multiple stages and therefore these differences could be minor.
39 Moreover, this advantage was not observed once the rats moved to the final stage of training (FourOdors),
40 where overall sequence memory did not differ between sexes. This is further supported by Reeders et al.,
41 (2021) who did not observe sex differences in humans performing an analogous task. Importantly, sequence
42 memory effects were consistent across two different sequences. This eliminated the possibility that males
43 and females show preference for a specific sequence or odors throughout the entirety of a session.

44 Apart from examining the general learning rates at each stage, we also performed several analyses to
45 test whether males and females differed in non-memory related behaviors in the sequence task. We saw no
46 effects between males and females on reward retrieval activity (ITI & IOI), on the time it took to run between

47 sequence (ISI), or on the overall frequency of nose pokes in which rats held the nose poke response for ≥ 1
48 s (InSeq trials). We did observe females had modestly longer hold times compared to males for OutSeq
49 trials. This was more clearly revealed in the analysis of poke time histogram where males showed a
50 tendency to have shorter poke times whereas females showed a tendency to have longer poke times.

51 Although, several studies have shown a robust male advantage in working and reference memory (Roof
52 et al., 1993; Veng et al., 2003), other studies indicate no differences or a female advantage (Bucci et al.,
53 2021; Healy et al., 1999; Lamberty & Gower, 1988). These confounding observations in sex differences
54 have been associated with different factors within the task parameters. For example, a consistent start
55 position for each trial has been associated with a lack of sex differences (Roof & Stein, 1999). Pre-training
56 has also been shown to reduce sex differences (Jonasson, 2005a; Perrot-Sinal et al., 1996). Generally
57 speaking, male and female rodents typically reach identical levels of performance on most cognitive tasks
58 although their rates and strategies of learning may differ.

59 **No Evidence Sequence Memory Differs Across Estrous Cycle**

60 Several studies have postulated that hormone fluctuations across the estrus cycle may account for sex
61 differences (McEwen & Milner, 2017; Sherry & Hampson, 1997; Warren & Juraska, 1997). However, no
62 clear consensus has emerged regarding the influence of estrous cycle on learning and memory. It has been
63 argued that estrous cycle cannot be causally linked to more variability in females relative to males
64 (Prendergast et al., 2014). Pompili et al. (2010) observed no variation in performance associated with cycle
65 phase when using the radial-arm maze. Stackman et al. (1997), using a delayed nonmatching-to-sample
66 paradigm in the radial-arm maze, found no effect of estrous cycle. Nevertheless, it is important to consider
67 sex hormones in behavior. Therefore, we examined sequence memory across phases of estrous cycle. In
68 general, we did not find differences between the phases of estrous cycle in sequence memory. It is possible
69 that pre-training might have reduced the sensitivity of this assessment (Bannerman et al., 1995; D. Saucier
70 & Cain, 1995). However, effects of the cycle on memory are somewhat subtle and inconsistent, so they may
71 wash out when averaging across multiple females.

72 Interestingly, while sequence memory was not affected by the day of estrous cycle, performance on
73 estrus was characterized by longer OutSeq poke times. One natural issue presented by measuring estrous

74 cycle for this task is the asymmetry of the length of stages; because each stage averages a different length
75 of time, we would expect to see far more trials occurring during these longer stages than the shorter ones.
76 To the extent that proestrus, metestrus and diestrus is over-represented compared to estrus, which could
77 have skewed the group mean. Several studies report that spatial memory tested in the Morris water maze or
78 object placement tasks was enhanced during proestrus relative to estrus and/or diestrus in mice and rats
79 (Frick & Berger-Sweeney, 2001; Frye, 1995; Paris & Frye, 2008; Pompili et al., 2010). However, the
80 reported proestrus advantage in spatial tasks is inconsistent with other data in rats showing enhanced
81 spatial reference memory during estrus relative to proestrus (Frye, 1995; Sutcliffe et al., 2007a; Warren &
82 Juraska, 1997) or no detectable effect of the cycle on memory (Berry et al., 1997; Bucci et al., 2008;
83 Cimadevilla et al., 2000; Conrad et al., 2004; Pompili et al., 2010; Stackman et al., 1997). While the exact
84 cognitive bases are not yet known, Korol & Kolo (2002) have suggested that the direction of estrogen effects
85 on performance depends upon the availability of strategies or solutions that match the participation of neural
86 or cognitive systems. Thus, the general effects of estrogen on memory are somewhat ambiguous
87 (Dohanich, 2002). The sequence task used here is known to involve multiple strategies including temporal
88 contexts, ordinal representations and working memory (e.g., Reeders et al., 2021; Jayachandran et al.,
89 2019)

90 **Conclusions**

91 We present evidence that sex and estrous cycle do not influence memory for sequences of events
92 suggesting that this core aspect of episodic memory does not differ between the sexes. These data are
93 consistent with few other studies that have investigated sex differences and estrous cycle as it pertains to
94 behavior in rodents as well as in humans (Bucci et al., 2021; Healy et al., 1999; Reeders et al., 2021;
95 Schmidt et al., 2009; Stackman et al., 1997). However, further studies are needed to fully characterize the
96 effects of sex hormones on sequence memory in both male and female rats by comparing the effects of
97 gonadectomy and possible hormone replacement. For now, the consideration of sex and estrous cycle as a
98 biological variable will have significantly advanced our understanding of the basic mechanisms of learning
99 and memory.

01
02
03
04
05
06
07
08
09
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27

ACKNOWLEDGMENTS

This work was supported by NIH grants R01 MH113626 and F99 NS119001. A special thanks to all the members of the Allen Lab, specifically our undergraduate research assistants, Andy Garcia, Sofia Levya, and Chiara Pavon, who helped with task training and data collection. Also, we would like to thank the Animal Care Facility and Dr. H. Vinerean DVM, DACLAM.

AUTHOR CONTRIBUTIONS

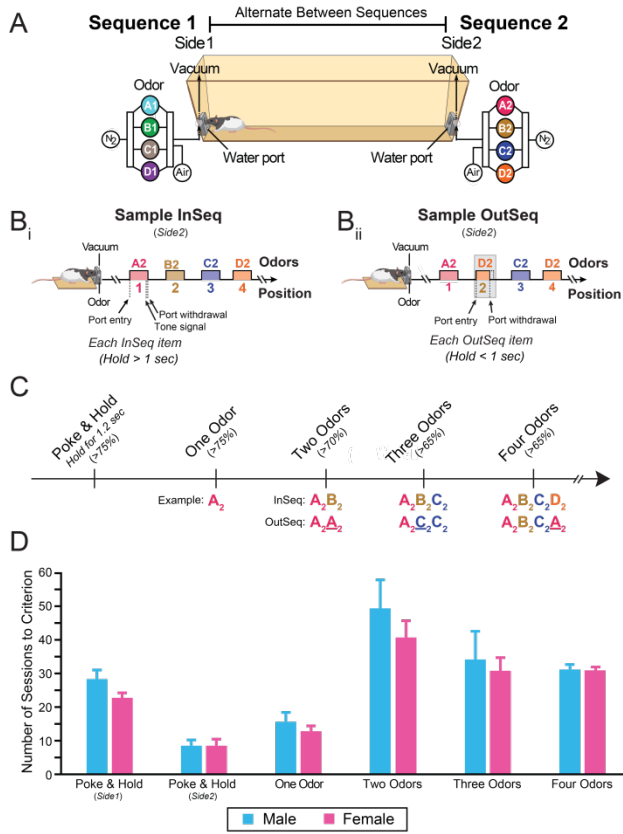
Conceptualization, T.A.A. and M.J.; Methodology, T.A.A. and M.J.; Investigation, M.J., P.L., F.P.R.; Writing – Original Draft, M.J., P.L., T.A.A.; Writing – Review and Editing, M.J., P.L., F.P.R., T.A.A., R.P.V.; Funding Acquisition, M.J., T.A.A. and R.P.V.; Resources, T.A.A., R.P.V.; Supervision, T.A.A.

DECLARATION OF INTERESTS

The authors declare no competing interests.

28

FIGURE CAPTIONS



29

30

Figure 1. Sequence Memory Task

(A) A 2m linear track was used with odor ports located at opposite ends where two four-odor sequences were presented (Seq1: A₁B₁C₁D₁ and Seq2: A₂B₂C₂D₂).

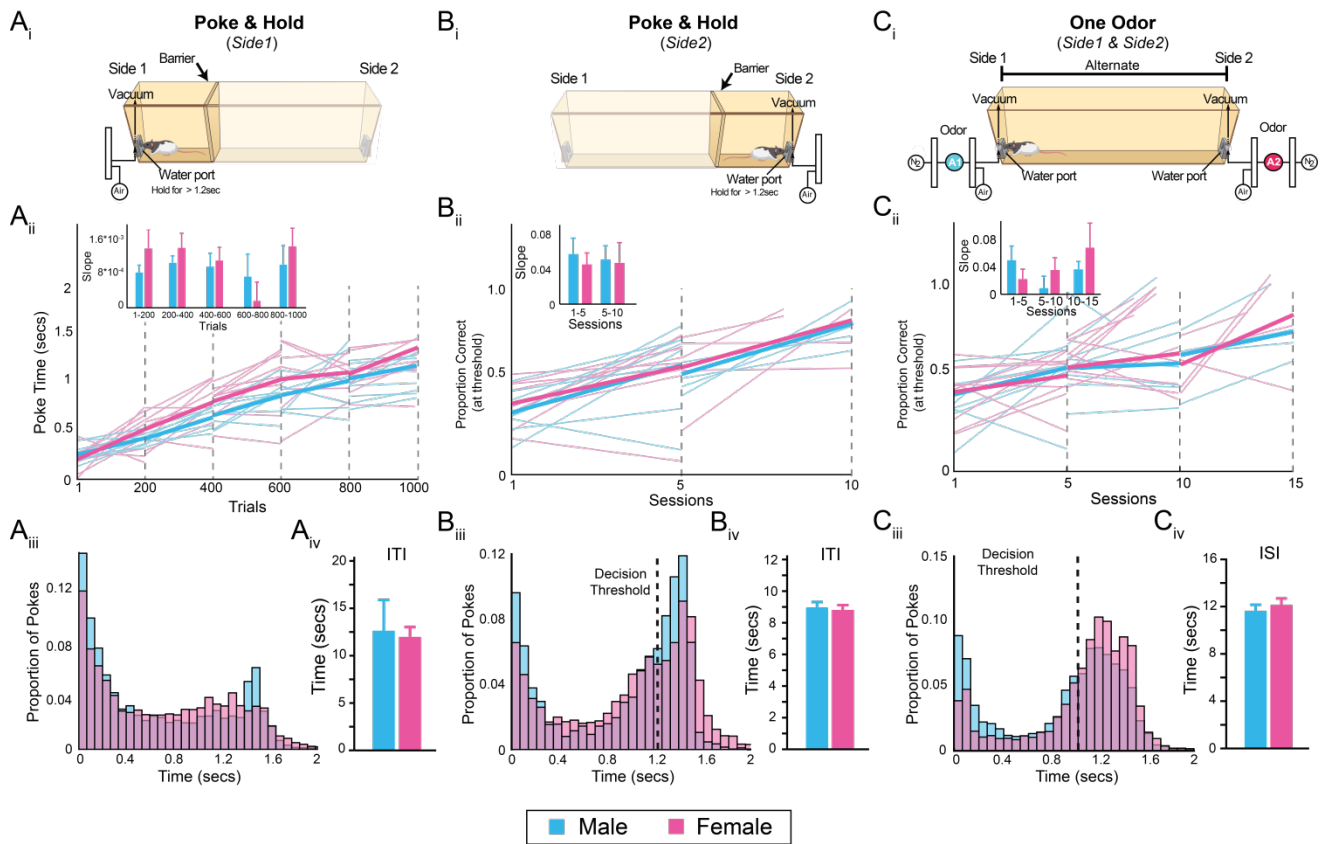
(B) Rats had to correctly identify the odors as either InSeq by holding their nose in the port for > 1 s (Bi) or OutSeq where the rat withdrew their nose prior to 1 s (Bii).

(C) Timeline showing the criterion for each training stage for the sequence memory task.

(D) Males and females did not differ in the average number of sessions per training stage.

37

All data are represented as mean ± SEM. *p < 0.05; **p < 0.01; ***p < 0.001



38
39

Figure 2. Males and Females Do Not Differ in Poking Behaviors

(A_i) Rats were trained to hold their nose in the nose-port on side 1 for 1.2 s.
 (A_{ii}) Males and females show no significant differences in the rate of learning across trials.
 (A_{iii}) Males and females poke times were relatively similar.
 (A_{iv}) Inter-trial-interval (ITI) was not significantly different between males and females
 (B_i) Rats were trained to hold their nose in the nose-port on side 2 for 1.2 s.
 (B_{ii}) No significant differences found between males and females in the rate of learning across sessions.
 (B_{iii}) Males and females show subtle shifts in behavior where males show a tendency to have short poke times. Both males and females had increased number of pokes at the decision threshold (1.2 s).
 (B_{iv}) ITI between sexes were not significantly different.
 (C_i) Rats were introduced to one odor on either side of the maze and were trained to alternate holding their nose in the pose-port on both sides of the maze for 1 s.
 (C_{ii}) No significant differences found between males and females in the rate of learning across sessions.
 (C_{iii}) Male and female rats show subtle shifts in behavior where males show a tendency to have shorter poke times.
 (C_{iv}) ISI between males and females was not significantly different.
 All data are represented as mean ± SEM. *p < 0.05; **p < 0.01; ***p < 0.001

56

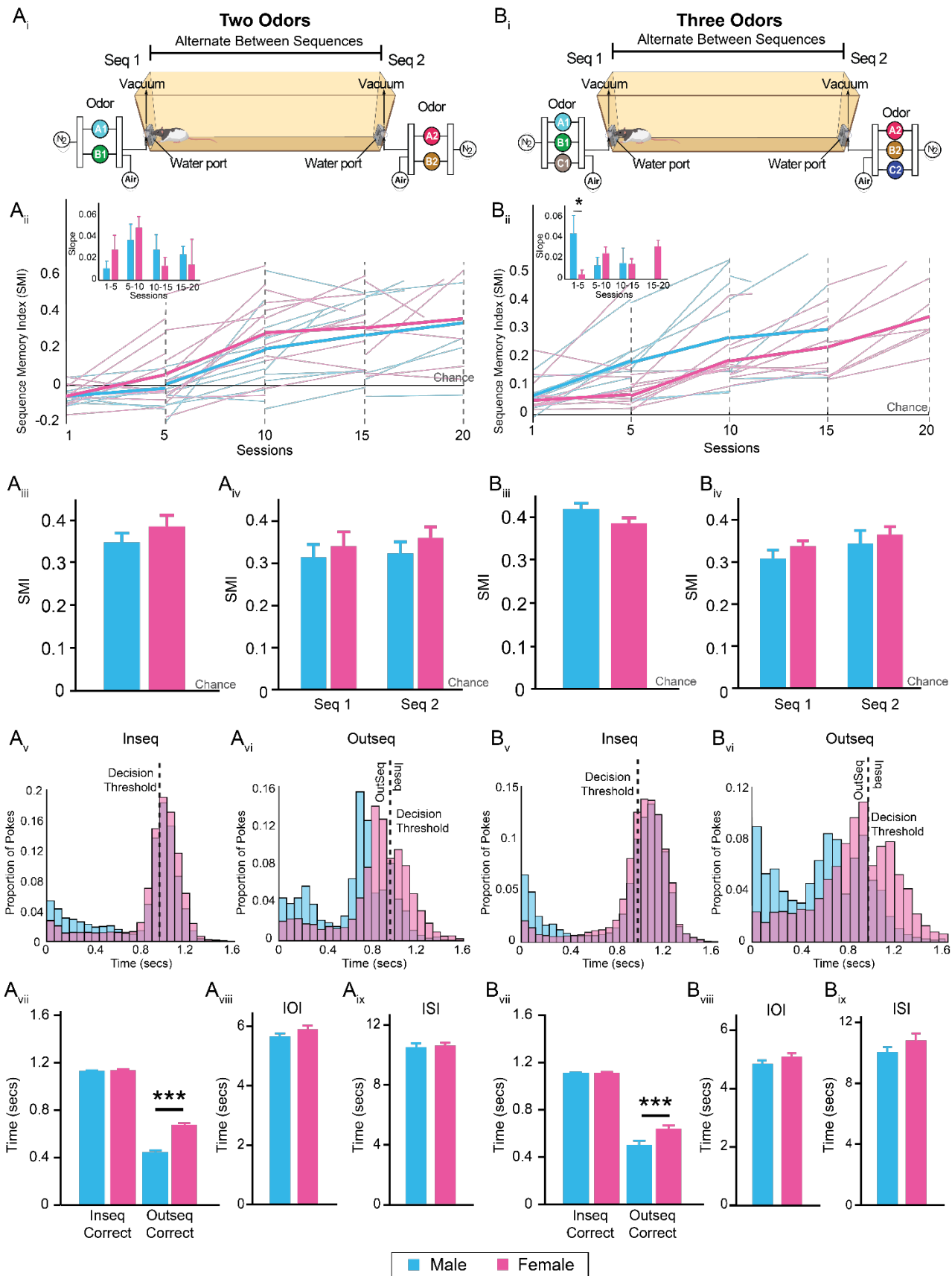


Figure 3. Learning Differs Between Males and Females

(A_i) A second odor was added to either side of the maze (Seq1: A₁B₁ and Seq2: A₂B₂).

- 61 (A_{ii}) Performance did not differ between males and females in the rate of learning across sessions.
62 (A_{iii}) SMI was not significantly different between males and females.
63 (A_{iv}) SMI was not significantly different between sequence 1 and sequence 2
64 (A_v) Males and females poke times were relatively similar.
65 (A_{vi}) Female nose-poke times show an increase in longer hold times while male poke times distribute
66 between short and long poke times
67 (A_{vii}) InSeq_{correct} trials did not differ between males and females. OutSeq_{correct} nose-poke times were
68 significantly different between males and females.
69 (A_{viii}) Inter-odor-interval (IOI) was not significantly different between sexes
70 (A_{ix}) Inter-sequence-interval (ISI) was not significantly different between males and females
71 (B_i) A third odor was added to both sides of the maze (Seq1: A₁B₁C₁ and Seq2: A₂B₂C₂).
72 (B_{ii}) Learning rate between males and females was statistically significant during the first five sessions. As
73 sessions continued, males and females no longer differed.
74 (B_{iii}) SMI was not significantly different between males and females.
75 (B_{iv}) SMI was not significantly different between sequence 1 and sequence 2.
76 (B_v) Male and female poke times for InSeq trials were relatively similar. Males have an increase in shorter
77 poke times.
78 (B_{vi}) Males show an increase number of short pokes for OutSeq trials while females showed longer hold
79 times.
80 (B_{vii}) InSeq_{correct} trials did not differ between males and females. OutSeq_{correct} poke times were significantly
81 different between males and females.
82 (B_{viii}) IOI did not differ between males and females
83 (B_{ix}) ISI did not differ between males and females.
84 All data are represented as mean ± SEM. *p < 0.05; **p < 0.01; ***p < 0.001
85

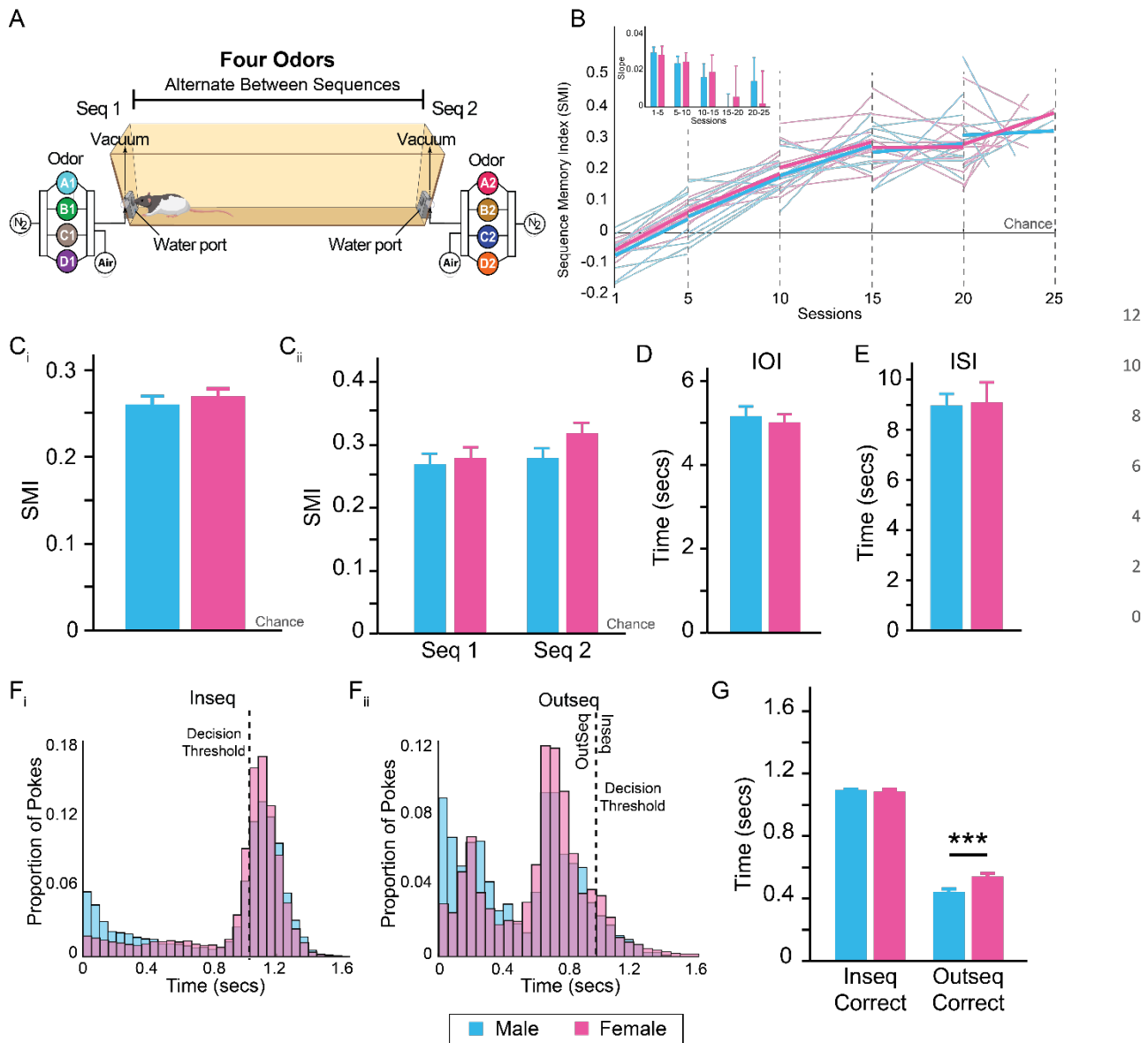


Figure 4. Males and Females Perform Similarly on the Sequence Memory Task

(A) Rats were trained in the final stage of the sequence memory task with four odors in each sequence (Seq1: A₁B₁C₁D₁ and Seq2: A₂B₂C₂D₂).

(B) Males and females showed no significant differences in the rate of learning across sessions.

(C_i) SMI was not significantly different between males and females.

(C_{ii}) SMI did not show any interaction effects between sequence 1 and sequence 2 and sex.

(D) IOI was not statistically different between males and females.

(E) ISI was not statistically different between males and females.

(F_i) Male and female poke distributions are relatively similar for InSeq trials

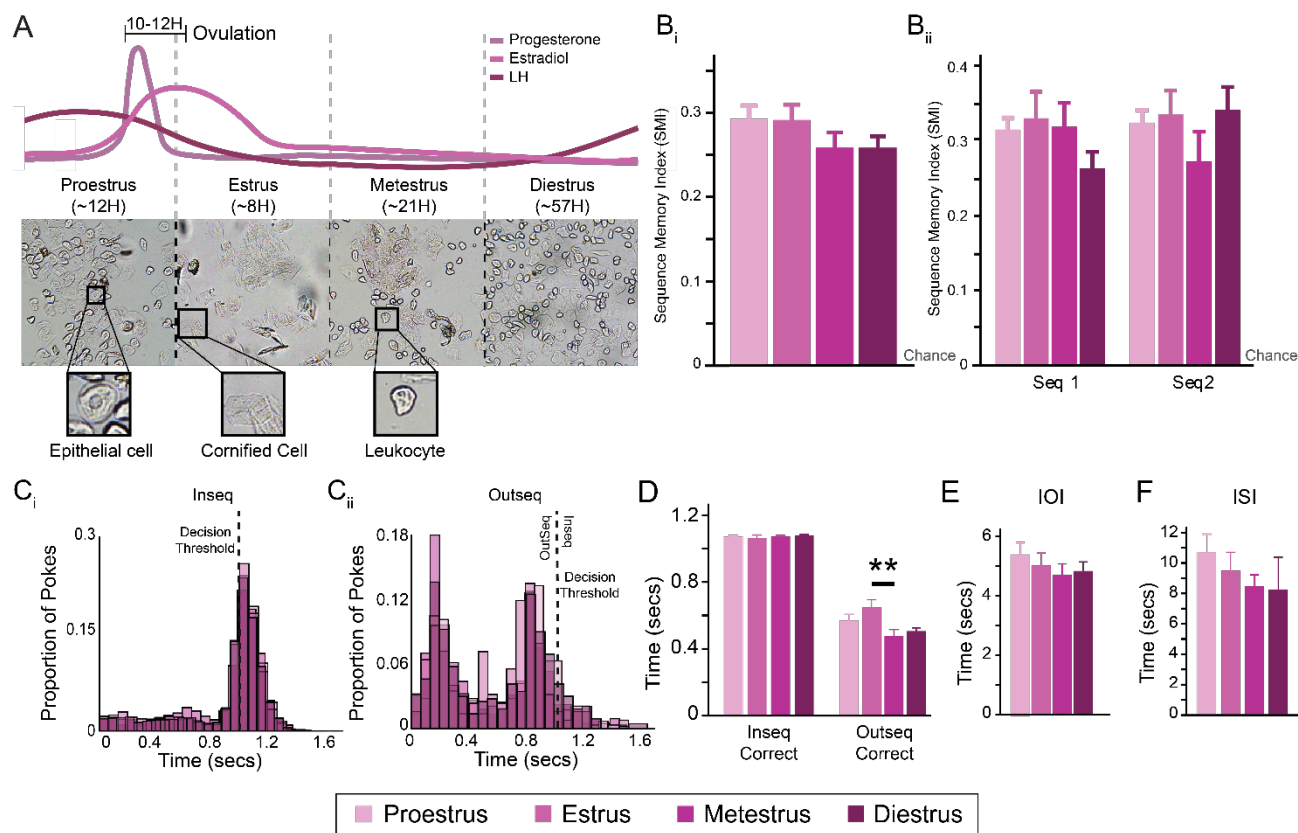
(F_{ii}) OutSeq poke distributions show subtle changes with males demonstrating an increase in shorter pokes compared to females.

(G) InSeq_{correct} trials did not differ between males and females. OutSeq_{correct} trials showed a significant difference between sexes.

All data are represented as mean ± SEM. *p < 0.05; **p < 0.01; ***p < 0.001

86
87

01



02
03

04

05

06

07

08

09

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51

REFERENCES

- Allen, L. M., Lesyshyn, R. A., O'Dell S. J., Allen, T. A., Fortin, N. J. (2020). The hippocampus, prefrontal cortex, and perirhinal cortex are critical to incidental order memory. *Behavioural Brain Research*, 379, 112215. <https://doi.org/10.1016/j.bbr.2019.112215>
- Allen, T. A., Morris, A. M., Mattfeld, A. T., Stark, C. E. L., & Fortin, N. J. (2014). A sequence of events model of episodic memory shows parallels in rats and humans. *Hippocampus*, 24(10), 1178–1188. <https://doi.org/10.1002/hipo.22301>
- Allen, T. A., Morris, A. M., Stark, S. M., Fortin, N. J., & Stark, C. E. L. (2015). Memory for sequences of events impaired in typical aging. *Learning & Memory*, 22(3). <https://doi.org/10.1101/lm.036301.114>
- Allen, T. A., Salz, D. M., McKenzie, S., & Fortin, N. J. (2016). Nonspatial Sequence Coding in CA1 Neurons. *The Journal of Neuroscience*, 36(5). <https://doi.org/10.1523/JNEUROSCI.2874-15.2016>
- Bannerman, D. M., Good, M. A., Butcher, S. P., Ramsay, M., & Morris, R. G. M. (1995). Distinct components of spatial learning revealed by prior training and NMDA receptor blockade. *Nature*, 378(6553). <https://doi.org/10.1038/378182a0>
- Barha, C. K., Dalton, G. L., & Galea, L. A. (2010). Low Doses of 17 α -Estradiol and 17 β -Estradiol Facilitate, Whereas Higher Doses of Estrone and 17 α - and 17 β -Estradiol Impair, Contextual Fear Conditioning in Adult Female Rats. *Neuropsychopharmacology*, 35(2). <https://doi.org/10.1038/npp.2009.161>
- Berry, B., McMahan, R., & Gallagher, M. (1997). Spatial learning and memory at defined points of the estrous cycle: Effects on performance of a hippocampal-dependent task. *Behavioral Neuroscience*, 111(2). <https://doi.org/10.1037/0735-7044.111.2.267>
- Bettis, T., & Jacobs, L. F. (2012). Sex differences in object recognition are modulated by object similarity. *Behavioural Brain Research*, 233(2), 288–292. <https://doi.org/10.1016/j.bbr.2012.04.028>
- Blanchard, R. J., Carol, D., & Blanchard, V. (1968). Passive avoidance: A variety of fear conditioning? In *Psych on. Sci* (Vol. 13). DOI:10.3758/BF03342386
- Bucci, D. J., Chiba, A. A., & Gallagher, M. (1995). Spatial learning in male and female Long-Evans rats. *Behavioral Neuroscience*, 135(1). <https://doi.org/10.1037/bne0000437>

- 52 Bucci, D. J., Hopkins, M. E., Nunez, A. A., Breedlove, S. M., Sisk, C. L., & Nigg, J. T. (2008). Effects of sex
53 hormones on associative learning in spontaneously hypertensive rats. *Physiology & Behavior*, 93(3).
54 <https://doi.org/10.1016/j.physbeh.2007.11.005>
- 55 Chai, X. J., & Jacobs, L. F. (2010). Effects of cue types on sex differences in human spatial memory.
56 *Behavioural Brain Research*, 208(2), 336–342. <https://doi.org/10.1016/j.bbr.2009.11.039>
- 57 Chesler, E. J., & Juraska, J. M. (2000). Acute Administration of Estrogen and Progesterone Impairs the
58 Acquisition of the Spatial Morris Water Maze in Ovariectomized Rats. *Hormones and Behavior*,
59 38(4). <https://doi.org/10.1006/hbeh.2000.1626>
- 60 Cimadevilla, J. M., Fenton, A. A., & Bures, J. (2000). Continuous place avoidance task reveals differences in
61 spatial navigation in male and female rats. *Behavioural Brain Research*, 107(1–2).
62 [https://doi.org/10.1016/S0166-4328\(99\)00128-X](https://doi.org/10.1016/S0166-4328(99)00128-X)
- 63 Conrad, C. D., Jackson, J. L., Wiczorek, L., Baran, S. E., Harman, J. S., Wright, R. L., & Korol, D. L.
64 (2004). Acute stress impairs spatial memory in male but not female rats: influence of estrous cycle.
65 *Pharmacology Biochemistry and Behavior*, 78(3). <https://doi.org/10.1016/j.pbb.2004.04.025>
- 66 Davis, D. M., Jacobson, T. K., Aliakbari, S., & Mizumori, S. J. Y. (2005). Differential effects of estrogen on
67 hippocampal- and striatal-dependent learning. *Neurobiology of Learning and Memory*, 84(2), 132–
68 137. <https://doi.org/10.1016/j.nlm.2005.06.004>
- 69 Dohanich, G. (2002). Gonadal Steroids, Learning, and Memory. In *Hormones, Brain and Behavior*. Elsevier.
70 <https://doi.org/10.1016/B978-012532104-4/50024-X>
- 71 Eichenbaum, H., & Fortin, N. J. (2005). Bridging the gap between brain and behavior: cognitive and neural
72 mechanisms of episodic memory. *Journal of the Experimental Analysis of Behavior*, 84(3).
73 <https://doi.org/10.1901/jeab.2005.80-04>
- 74 Frick, K. M., & Berger-Sweeney, J. (2001). Spatial reference memory and neocortical neurochemistry vary
75 with the estrous cycle in C57BL/6 mice. *Behavioral Neuroscience*, 115(1).
76 <https://doi.org/10.1037/0735-7044.115.1.229>

- 77 Frick, K. M., Burlingame, L. A., Arters, J. A., & Berger-Sweeney, J. (1999). Reference memory, anxiety and
78 estrous cyclicity in c57bl/6nia mice are affected by age and sex.
79 www.elsevier.com/locate/neuroscience
- 80 Frye, C. A. (1995). Estrus-associated decrements in a water maze task are limited to acquisition. *Physiology*
81 *& Behavior*, 57(1). [https://doi.org/10.1016/0031-9384\(94\)00197-D](https://doi.org/10.1016/0031-9384(94)00197-D)
- 82 Frye, C. A., Duffy, C. K., & Walf, A. A. (2007). Estrogens and progestins enhance spatial learning of intact
83 and ovariectomized rats in the object placement task. *Neurobiology of Learning and Memory*, 88(2).
84 <https://doi.org/10.1016/j.nlm.2007.04.003>
- 85 Goldman, J. M., Murr, A. S., & Cooper, R. L. (2007). The rodent estrous cycle: Characterization of vaginal
86 cytology and its utility in toxicological studies. In *Birth Defects Research Part B - Developmental and*
87 *Reproductive Toxicology* (Vol. 80, Issue 2, pp. 84–97). <https://doi.org/10.1002/bdrb.20106>
- 88 Gruene, T. M., Flick, K., Stefano, A., Shea, S. D., & Shansky, R. M. (2015). Sexually divergent expression of
89 active and passive conditioned fear responses in rats. *ELife*, 4. <https://doi.org/10.7554/eLife.11352>
- 90 Hamson, D. K., Roes, M. M., & Galea, L. A. M. (2016). Sex Hormones and Cognition: Neuroendocrine
91 Influences on Memory and Learning. In *Comprehensive Physiology* (Vol. 6, Issue 3, pp. 1295–1337).
92 <https://doi.org/10.1002/cphy.c150031>
- 93 Healy, S. D., Braham, S. R., & Braithwaite, V. A. (1999). Spatial working memory in rats: no differences
94 between the sexes. *Proceedings of the Royal Society of London. Series B: Biological Sciences*,
95 266(1435). <https://doi.org/10.1098/rspb.1999.0923>
- 96 Jayachandran, M., Linley, S. B., Schlecht, M., Mahler, S. v., Vertes, R. P., & Allen, T. A. (2019). Prefrontal
97 Pathways Provide Top-Down Control of Memory for Sequences of Events. *Cell Reports*, 28(3), 640-
98 654.e6. <https://doi.org/10.1016/j.celrep.2019.06.053>
- 99 Jonasson, Z. (2005). Meta-analysis of sex differences in rodent models of learning and memory: a review of
00 behavioral and biological data. *Neuroscience & Biobehavioral Reviews*, 28(8).
01 <https://doi.org/10.1016/j.neubiorev.2004.10.006>

- 02 Kessler, R. C., Berglund, P., Demler, O., Jin, R., Merikangas, K. R., & Walters, E. E. (2005). Lifetime
03 Prevalence and Age-of-Onset Distributions of DSM-IV Disorders in the National Comorbidity Survey
04 Replication. *Archives of General Psychiatry*, 62(6). <https://doi.org/10.1001/archpsyc.62.6.593>
- 05 Korol, D. L. (2004). Role of estrogen in balancing contributions from multiple memory systems.
06 *Neurobiology of Learning and Memory*, 82(3). <https://doi.org/10.1016/j.nlm.2004.07.006>
- 07 Korol, D. L., & Kolo, L. L. (2002). Estrogen-induced changes in place and response learning in young adult
08 female rats. *Behavioral Neuroscience*, 116(3). <https://doi.org/10.1037/0735-7044.116.3.411>
- 09 Koss, W. A., & Frick, K. M. (2017). Sex differences in hippocampal function. In *Journal of Neuroscience*
10 *Research* (Vol. 95, Issues 1–2, pp. 539–562). John Wiley and Sons Inc.
11 <https://doi.org/10.1002/jnr.23864>
- 12 Lamberty, Y., & Gower, A. J. (1988). Investigation into sex-related differences in locomotor activity, place
13 learning and passive avoidance responding in NMRI mice. *Physiology & Behavior*, 44(6).
14 [https://doi.org/10.1016/0031-9384\(88\)90063-7](https://doi.org/10.1016/0031-9384(88)90063-7)
- 15 Launer, J. (1999). Narrative based medicine: A narrative approach to mental health in general practice.
16 *BMJ*, 318(7176). <https://doi.org/10.1136/bmj.318.7176.117>
- 17 Levy, L. J., Astur, R. S., & Frick, K. M. (2005). Men and Women Differ in Object Memory but Not
18 Performance of a Virtual Radial Maze. *Behavioral Neuroscience*, 119(4).
19 <https://doi.org/10.1037/0735-7044.119.4.853>
- 20 Luine, V., Gomez, J., Beck, K., & Bowman, R. (2017). Sex differences in chronic stress effects on cognition
21 in rodents. *Pharmacology Biochemistry and Behavior*, 152, 13–19.
22 <https://doi.org/10.1016/j.pbb.2016.08.005>
- 23 McCarthy, M. M., & Konkle, A. T. M. (2005). When is a sex difference not a sex difference? *Frontiers in*
24 *Neuroendocrinology*, 26(2). <https://doi.org/10.1016/j.yfrne.2005.06.001>
- 25 McEwen, B. S., & Milner, T. A. (2017). Understanding the broad influence of sex hormones and sex
26 differences in the brain. *Journal of Neuroscience Research*, 95(1–2).
27 <https://doi.org/10.1002/jnr.23809>

- 28 Nowak, N. T., Diamond, M. P., Land, S. J., & Moffat, S. D. (2014). Contributions of sex, testosterone, and
29 androgen receptor CAG repeat number to virtual Morris water maze performance.
30 *Psychoneuroendocrinology*, 41. <https://doi.org/10.1016/j.psyneuen.2013.12.003>
- 31 Paris, J. J., & Frye, C. A. (2008). Estrous cycle, pregnancy, and parity enhance performance of rats in object
32 recognition or object placement tasks. *Reproduction*, 136(1). <https://doi.org/10.1530/REP-07-0512>
- 33 Perrot-Sinal, T. S., Kostenuik, M. A., Ossenkopp, K.-P., & Kavaliers, M. (1996). Sex differences in
34 performance in the Morris water maze and the effects of initial nonstationary hidden platform training.
35 *Behavioral Neuroscience*, 110(6). <https://doi.org/10.1037/0735-7044.110.6.1309>
- 36 Pigott, T. A. (2003). Anxiety disorders in women. *Psychiatric Clinics of North America*, 26(3).
37 [https://doi.org/10.1016/S0193-953X\(03\)00040-6](https://doi.org/10.1016/S0193-953X(03)00040-6)
- 38 Pompili, A., Tomaz, C., Arnone, B., Tavares, M. C., & Gasbarri, A. (2010). Working and reference memory
39 across the estrous cycle of rat: A long-term study in gonadally intact females. *Behavioural Brain*
40 *Research*, 213(1). <https://doi.org/10.1016/j.bbr.2010.04.018>
- 41 Prendergast, B. J., Onishi, K. G., & Zucker, I. (2014). Female mice liberated for inclusion in neuroscience
42 and biomedical research. *Neuroscience & Biobehavioral Reviews*, 40.
43 <https://doi.org/10.1016/j.neubiorev.2014.01.001>
- 44 Reeders, P. C., Hamm, A. G., Allen, T. A., & Mattfeld, A. T. (2021). Medial prefrontal cortex and
45 hippocampal activity differentially contribute to ordinal and temporal context retrieval during
46 sequence memory. *Learning & Memory*, 28(4). <https://doi.org/10.1101/lm.052365.120>
- 47 Roof, R. L., Zhang, Q., Glasier, M. M., & Stein, D. G. (1993). Gender-specific impairment on Morris water
48 maze task after entorhinal cortex lesion. *Behavioural Brain Research*, 57(1).
49 [https://doi.org/10.1016/0166-4328\(93\)90060-4](https://doi.org/10.1016/0166-4328(93)90060-4)
- 50 Roof, R., & Stein, D. (1999). Gender differences in Morris water maze performance depend on task
51 parameters. *Physiology & Behavior*, 68(1–2). [https://doi.org/10.1016/S0031-9384\(99\)00162-6](https://doi.org/10.1016/S0031-9384(99)00162-6)
- 52 Saucier, D., & Cain, D. P. (1995). Spatial learning without NMDA receptor-dependent long-term potentiation.
53 *Nature*, 378(6553). <https://doi.org/10.1038/378186a0>

- 54 Saucier, D. M., Shultz, S. R., Keller, A. J., Cook, C. M., & Binsted, G. (2008). Sex differences in object
55 location memory and spatial navigation in Long-Evans rats. *Animal Cognition*, 11(1), 129–137.
56 <https://doi.org/10.1007/s10071-007-0096-1>
- 57 Schmidt, B., Jacobson, T. K., & Markus, E. (2009). Hippocampal and striatal dependent navigation: Sex
58 differences are limited to acquisition. *Hormones and Behavior*, 56(2), 199–205.
59 <https://doi.org/10.1016/j.yhbeh.2009.04.004>
- 60 Seymoure, P., Dou, H., & Juraska, J. M. (1996). Sex differences in radial maze performance: Influence of
61 rearing environment and room cues. In *Psychobiology* (Vol. 24, Issue 1).
62 <https://doi.org/10.3758/BF03331950>
- 63 Sherry, D. F., & Hampson, E. (1997). Evolution and the hormonal control of sexually-dimorphic spatial
64 abilities in humans. *Trends in Cognitive Sciences*, 1(2). [https://doi.org/10.1016/S1364-](https://doi.org/10.1016/S1364-6613(97)01015-2)
65 [6613\(97\)01015-2](https://doi.org/10.1016/S1364-6613(97)01015-2)
- 66 Sokal, Robert. R., & Rohlf, J. F. (1995). *Biometry: The principles and practice of statistics in Biological*
67 *Research* (Vol. 2nd). Freeman.
- 68 Spritzer, M. D., Gill, M., Weinberg, A., & Galea, L. A. M. (2008). Castration Differentially Affects Spatial
69 Working and Reference Memory in Male Rats. *Archives of Sexual Behavior*, 37(1).
70 <https://doi.org/10.1007/s10508-007-9264-2>
- 71 Stackman, R. W., Blasberg, M. E., Langan, C. J., & Clark, A. S. (1997). Stability of Spatial Working Memory
72 across the Estrous Cycle of Long–Evans Rats. *Neurobiology of Learning and Memory*, 67(2).
73 <https://doi.org/10.1006/nlme.1996.3753>
- 74 Steimer, T., & Driscoll, P. (2005). Inter-individual vs line/strain differences in psychogenetically selected
75 Roman High-(RHA) and Low-(RLA) Avoidance rats: Neuroendocrine and behavioural aspects. In
76 *Neuroscience and Biobehavioral Reviews* (Vol. 29, Issue 1 SPEC. ISS., pp. 99–112). Elsevier Ltd.
77 <https://doi.org/10.1016/j.neubiorev.2004.07.002>
- 78 Sutcliffe, J. S., Marshall, K. M., & Neill, J. C. (2007). Influence of gender on working and spatial memory in
79 the novel object recognition task in the rat. *Behavioural Brain Research*, 177(1).
80 <https://doi.org/10.1016/j.bbr.2006.10.029>

- 81 Tulving, E. (2002). Episodic Memory: From Mind to Brain. *Annual Review of Psychology*, 53(1).
82 <https://doi.org/10.1146/annurev.psych.53.100901.135114>
- 83 van Goethem, N. P., Rutten, K., van der Staay, F. J., Jans, L. A. W., Akkerman, S., Steinbusch, H. W. M.,
84 Blokland, A., van't Klooster, J., & Prickaerts, J. (2012). Object recognition testing: Rodent species,
85 strains, housing conditions, and estrous cycle. *Behavioural Brain Research*, 232(2).
86 <https://doi.org/10.1016/j.bbr.2012.03.023>
- 87 Veng, L. M., Granholm, A.-C., & Rose, G. M. (2003). Age-related sex differences in spatial learning and
88 basal forebrain cholinergic neurons in F344 rats. *Physiology & Behavior*, 80(1).
89 [https://doi.org/10.1016/S0031-9384\(03\)00219-1](https://doi.org/10.1016/S0031-9384(03)00219-1)
- 90 Walf, A. A., & Frye, C. A. (2006). A Review and Update of Mechanisms of Estrogen in the Hippocampus and
91 Amygdala for Anxiety and Depression Behavior. *Neuropsychopharmacology*, 31(6).
92 <https://doi.org/10.1038/sj.npp.1301067>
- 93 Warren, S. G., & Juraska, J. M. (1997). Spatial and nonspatial learning across the rat estrous cycle.
94 *Behavioral Neuroscience*, 111(2). <https://doi.org/10.1037/0735-7044.111.2.259>
- 95 Williams, C. L., Barnett, A. M., & Meck, W. H. (1990). Organizational effects of early gonadal secretions on
96 sexual differentiation in spatial memory. *Behavioral Neuroscience*, 104(1).
97 <https://doi.org/10.1037/0735-7044.104.1.84>
- 98 Woolley, D. G., Vermaercke, B., de Beeck, H. O., Wagemans, J., Gantois, I., D'Hooge, R., Swinnen, S. P.,
99 & Wenderoth, N. (2010). Sex differences in human virtual water maze performance: Novel measures
00 reveal the relative contribution of directional responding and spatial knowledge. *Behavioural Brain*
01 *Research*, 208(2), 408–414. <https://doi.org/10.1016/j.bbr.2009.12.019>
- 02 Zurkovsky, L., Brown, S. L., Boyd, S. E., Fell, J. A., & Korol, D. L. (2007). Estrogen modulates learning in
03 female rats by acting directly at distinct memory systems. *Neuroscience*, 144(1), 26–37.
04 <https://doi.org/10.1016/j.neuroscience.2006.09.002>
05
06