Prevalent sex ratio bias in *Caenorhabditis* nematodes

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Abstract

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Although equal sex ratio represents an equilibrium in evolutionary theory, cases of sex ratio bias are accumulating, mostly reported in single species. Here, we surveyed progeny sex ratios in 23 species of the nematode genus Caenorhabditis. In experiments where males and females had unlimited access to each other (unlimited mating), we found 15 species out of the 23 species had female(hermaphrodite)-biased sex ratios. Phylogenetic mapping indicates female-bias to represent the ancestral state, with the occurrence of seven independent transitions from femalebias to non-bias along the phylogeny. As sperm competition could underlie the female bias, we also assayed progeny sex ratios in an experiment where mating was limited to a few hours. Of the 15 species that showed female-biased ratios under unlimited mating, six species showed no sex ratio bias when mating was limited. This result is consistent with sperm competition whereby Xbearing sperm take precedence over nullo-X during fertilization, yielding more female progeny when sperm are unlimited but equal sex ratio when sperm are limited. The other nine species showed sex ratio bias in both experiments, but the day-by-day profiles suggest sperm competition may also play a role. Our study shows that sex ratio evolution within Caenorhabditis nematodes is dynamic and that sex ratio bias is common not only in parasites as previously found but also in free-living nematodes. Our results also suggest that sperm competition could be a mechanism underlying sex ratio bias.

Key words:

sex ratio bias, sperm competition, mating, Caenorhabditis

Introduction

Sex ratio theory is at the core of evolutionary biology. First proposed by Charles Darwin (1, 2), and later formulated by Ronald Fisher (3), sex ratio theory posits that in a large, random mating population, when the sex ratio departs from equality, the rarer sex would have better mating prospects. Thus, genetic elements that favor the rarer sex would be favored by natural selection. Consequently, equal sex ratio should be restored and represents an evolutionary stable strategy. However, as many organisms have unique life histories, social structures and parental behaviors that do not meet Fisher's assumptions, biased sex ratios exist and are intriguing examples to study the evolution of sex ratio (4).

Theories have been proposed to explain sex ratio bias based on the asymmetry of sex allocation and reproductive return between the two sexes in different scenarios (5, 6). Hamilton's theory of local mate competition (LMC) predicts that in cases of highly structured populations and local mating, sex ratios would be biased toward females so that just enough males are produced to inseminate their female siblings (4). Similarly, theories predict that sex ratio bias may arise due to local resource competition, where the sex that only consumes local resources is reduced (7). In the case of cooperative breeding, sex ratio would be biased toward the helping sex (local resource enhancement, LRE) (8, 9). Moreover, Trivers and Willard proposed that females in good condition or of high social ranking would produce more male offspring as their sons would inherit their advantages and have above-average breeding success (10). These theories have successfully explained many empirical findings of sex ratio bias.

Examples of sex ratio bias across the unicellular and metazoan world are plentiful. For instance, female-biased sex ratios are commonly found in Apicomplexan parasites such as *Plasmodium malariae* (11-13) and *Toxoplasma gondii* (14), as well as in intestinal parasitic nematodes of *Heligmosomidae* (15, 16). In Hymenoptera insects, female-biased sex ratios are found and/or tested in fig wasps (17), parasitoid wasps of *Nasonia vitripennis* (18, 19), and the *Bethylidae* family(20). These female-biased sex ratios are good examples of LMC as these parasites are confined within their hosts. In birds, the Seychelles warbler shows facultative sex ratio bias in their offspring as they adjust production of helpers in relation to the quality of their territories (21, 22). Male-biased sex ratio was found in African wild dogs because male helpers

contribute to raising pups in the dens and increase pup survivorship (23). These are two typical examples of LRE. Female-biased sex ratios have also been observed in red deer and wild spider monkeys with subordinate females producing more daughters but high-ranking females producing more sons (24, 25), which fits the Trivers-Willard hypothesis.

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Despite the mature theories for sex ratio bias and plenty of empirical examples, the evolutionary dynamics of sex ratio change above the species level is less understood. Because sex ratio is generally selected to maximize reproductive success, it can be regarded as an adaptive trait of sexually reproducing organisms. But how often and how fast do sex ratios evolve among diverging species, especially when they adapt to new environments and adopt new life histories? Does sex ratio bias evolve reversibly and repeatedly? To answer these questions, one needs to examine sex ratios across a lineage.

The genus Caenorhabditis provides a good opportunity to study the evolution of sex ratios as it is a species-rich genus (26), comprising ecologically diverse species with various life histories, population structures, and even reproductive modes (27). Most Caenorhabditis species have standard female-male reproduction (dioecy), however, three species, C. elegans, C. briggsae and C. tropicalis, have independently evolved reproduction through hermaphrodites and facultative males (androdioecy) (28, 29). Species of both reproductive modes in Caenorhabditis have the same chromosomal sex determination system with females (or hermaphrodites) being XX and males being XO (30). But unlike the male-female species that have obligate out-crossing, the androdioecious species have two ways of reproduction: a hermaphrodite can either self or outcross with a male. Self-fertilization produces mostly hermaphrodites while males are produced by spontaneous nondisjunction of the X chromosome during meiosis at very low rates (31). On the other hand, outcrossing with a male should produce hermaphrodites and males in a 1:1 ratio, according to Mendel's first rule. The common lab model, C. elegans, progenies derived from outcrossing display an equal sex ratio (selfing produces >99.5% hermaphrodites)(31, 32). In contrast, C. briggsae, yields a hermaphrodite-biased sex ratio upon outcrossing, putatively due to sperm competition where the male X-bearing sperm outcompeting the nullo-X counterpart (33). Knowledge of sex ratios and the underlying mechanisms in the other species of this genus is so far scarce.

In this study, we examined differences in progeny sex ratio across the genus *Caenorhabditis*. We first assayed the sex ratios where males and females (or hermaphrodites) have continuous access to each other throughout their lifetime ("unlimited mating experiment"). We mapped these observed sex ratios onto the phylogeny and inferred the ancestral state of sex ratio within this genus. As LaMunyon and Ward (1997) found sperm competition between male X-bearing and nullo-X sperm as a putative mechanism underlying the hermaphrodite-biased sex ratios in *C. briggsae* (33), we also investigated the role of sperm competition in sex ratio bias in the other *Caenorhabditis* species. In contrast to the unlimited mating experiment, we assayed the sex ratios by limiting mating to a short time interval so that with limited amounts of sperm, when X-bearing sperm are used up, the nullo-X sperm have a chance to catch up. Species exhibiting female-biased sex ratios in the unlimited mating experiment but no bias in the limited mating experiment suggest sperm competition as a mechanism explaining sex ratio bias. These surveys and analyses would enhance our understanding of the evolutionary dynamics of sex ratio within this diverse genus and also shed light on the mechanisms underlying sex ratio bias.

Results

Sex ratio bias is common in *Caenorhabditis*

To survey sex ratio bias within the *Caenorhabditis* genus, we conducted crosses in 23 species where females and males had continuous access to each other (unlimited mating experiment) and counted the numbers of female (hermaphrodite) versus male progeny produced. Fifteen of the 23 species tested exhibited sex ratio bias in this experiment (combined and adjusted P < 0.05, binomial test, Figure 1a, Table 1), all of which were female(hermaphrodite)-biased, with the median sex ratios ranging from 0.524 (*C. latens*) to 0.708 (*C. doughertyi*). In contrast, the eight species that did not show sex ratio bias had the median sex ratios ranging from 0.493 (*C. elegans*) to 0.550 (*C. castelli*).

Species with sex ratio bias do not cluster by phylogeny or reproductive mode

The species with sex ratio bias were scattered across the phylogeny, i.e., across the *Elegans* group, the *Japonica* group, and the *Drosophilae* supergroup. Ancestral trait reconstruction analysis assigned female-biased sex ratio as the ancestral states of the *Elegans* group, *Japonica* group, and the *Drosophilae* supergroup. Furthermore, the ancestral state of the *Caenorhabditis* genus was also assigned as female-biased (Figure 1b). Across the phylogeny, there was a total of seven transitions from female-biased to non-biased sex ratios, with four transitions in the *Elegans* group, one in the *Japonica* group, and two in the *Drosophilae* supergroup (Figure 1b). A single transition in the *Elegans* group resulted in one monophyletic clade of equal sex ratio, i.e., clade of *C. wallacei* and *C. tropicalis*. The other six transitions resulted in singleton species with equal sex ratio: *C.* sp. 41, *C. brenneri*, and *C. elegans* (*Elegans* group), *C. panamensis* (*Japonica* group), and *C. virilis* and *C. castelli* (*Drosophilae* supergroup).

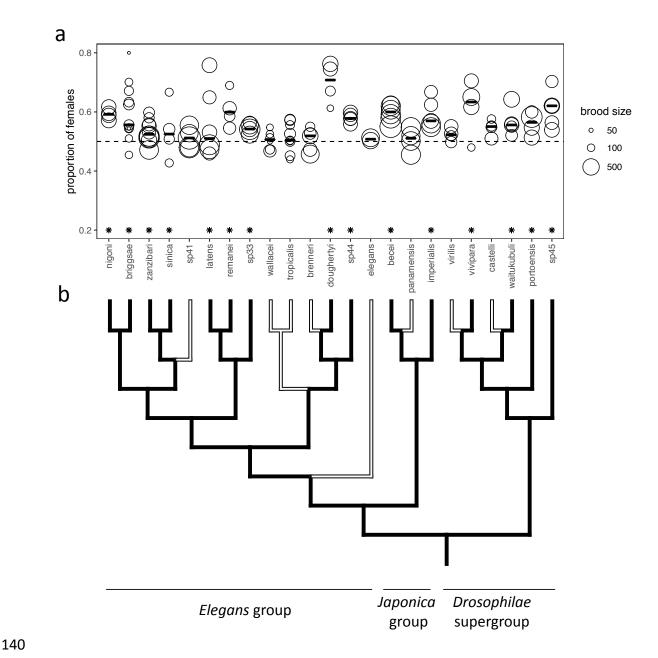


Figure 1. Progeny sex ratios and the ancestral states of 23 *Caenorhabditis* species. (a) The proportion of females (hermaphrodites) in total progeny in the unlimited mating assay. For the species with multiple strains, only one tester strain is included in this figure. Each circle represents a replicate mating pair of the tester strain and the circle size denotes the total number of progeny. Bars represent median sex ratios. The asterisks indicate significant female (hermaphrodite) bias (combined and adjusted P < 0.05, binomial test). (b) The ancestral states of sex ratio bias constructed based on the phylogeny of the 23 species. The black branches represent sex ratio bias in the child nodes whereas the white branches represent equal sex ratio.

Furthermore, the sex ratio bias was not consistent across the hermaphroditic species. Of the three hermaphroditic species that independently evolved in the *Elegans* group (29), *C. briggsae* showed a hermaphrodite-biased sex ratio whereas *C. elegans* and *C. tropicalis* did not show sex ratio bias.

For the species with multiple strains, the sex ratios from different strains showed

Testing multiple strains within species reveals consistent sex ratios

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consistent bias or non-bias within the species. For example, for the four female-male species (C. nigoni, C. latens, C. remanei, and C. sp. 33), all strains from the respective species showed consistently biased sex ratios (P < 0.05 for the strains), except the C. remanei strain JU1084, which had a marginally non-biased sex ratio (P = 0.058). However, C. nigoni, C. remanei, and C. sp. 33 had significantly different sex ratios between strains (P < 0.05, ANOVA test), whereas C. latens did not have significantly different sex ratios between strains (P = 0.824, ANOVA test). To test the possibility that sex ratio bias was due to sex-specific consequences of inbreeding depression (34), for the four female-male species, for which we had multiple strains, (C. nigoni, C. latens, C. remanei, and C. sp. 33), we performed inter-stain crosses. The inter-strain crosses yielded consistent female-biased sex ratios, congruent with the intrastrain experiments above (combined P < 0.05, Table S1). For the three androdioecious species, the multiple tester strains also showed consistent bias (for C. briggsae) or non-bias (for C. elegans and C. tropicalis). All the strains from C. elegans and C. tropicalis, respectively, did not have hermaphrodite-biased sex ratios (P > 0.05 for the strains), and the sex ratios were not significantly different between strains for both of these two species (P = 0.658 for C. elegans and P = 0.441 for C. tropicalis, ANOVA test). For 31 C. briggsae strains we surveyed in addition to the common lab strain AF16, 19 strains showed significant hermaphrodite-biased sex ratios (combined and adjusted P < 0.05, binomial test,

Table S2), consistent with AF16. For the other 12 strains that did not have significant sex ratio bias, most of them still had more hermaphrodites than males for all replicates, except 1 out of 4 replicates in BRC20095, 1 out of 5 in BRC20299, 1 out of 5 in BRC20324, 1 out of 5 in BRC20334, 2 out of 5 in BRC20234 and 4 out of 8 in BRC20339. Thus, the majority of the *C. briggsae* strains had significantly hermaphrodite-biased sex ratios, whereas for the other strains, although there was no significant bias, there was a general tendency toward bias, except for BRC20234 and BRC20339.

Sex ratios in the limited mating experiment

To test whether sperm competition between X and nullo-X plays a role in progeny sex ratio bias, we conducted limited mating experiments to assay sex ratios. When exposed to males for only up to 5 hours, the females (or hermaphrodites) sired on average 51% (range 20-80%) fewer progeny (outcrossed progeny for androdioecious species) compared to unlimited mating, except for C. sinica and C. latens, which had comparable numbers of progeny between the two mating experiments (Table 1 and 2). Ten of the 23 species examined showed female(hermaphrodite)-biased sex ratios in the limited mating experiment (combined and adjusted P < 0.05, binomial test; Figure 2a, Table 2). Out of the 15 species that showed female-biased sex ratio in the unlimited mating experiment, nine were also found with female(hermaphrodite)-biased sex ratios in the limited mating experiment. The other six species (C. nigoni, C. zanzibari, C. sinica, C. remanei, C. becei, and C. portoensis) did not show significant sex ratio bias in the limited mating experiment. In contrast, C. sp. 41, which did not show sex-ratio bias in the unlimited mating experiment, showed significant female-biased sex ratio in the limited mating experiment, showed significant female-biased sex ratio in the limited mating experiment, showed significant female-biased sex ratio in the limited mating experiment, showed significant female-biased sex ratio in the limited mating experiment, showed significant female-biased sex ratio in the limited mating experiment, showed significant female-biased sex ratio in the limited mating experiment, showed significant female-biased sex ratio in the limited mating experiment (adjusted P = 0.0002). The remaining seven species consistently showed no bias in both the unlimited and limited mating experiments.

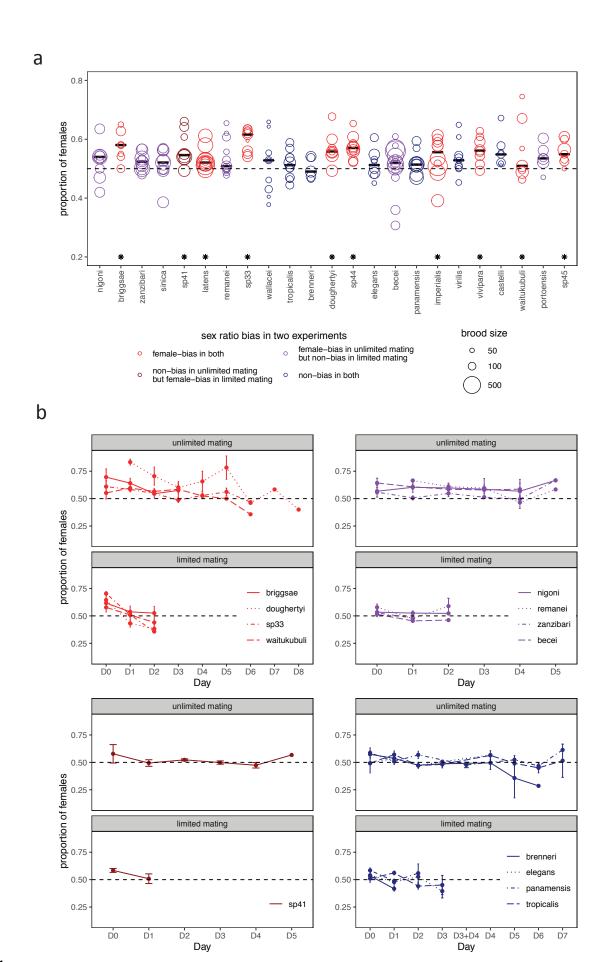


Figure 2. Progeny sex ratios in the limited mating assay. (a) The proportion of females (hermaphrodites) in total progeny after limited mating. Each circle represents a replicate mating pair of the tester strain of the species and the circle size denotes the total number of progeny. Black bars represent median sex ratios. The asterisks indicate significant female (hermaphrodite) bias (combined and adjusted P < 0.05, binomial test). The 23 species are categorized into 4 groups: significant female bias in both unlimited mating and limited mating experiments (red), significant female bias in unlimited mating but not in limited mating (purple), significant female bias in limited mating but not in unlimited mating (brown), and no bias in both experiments (blue). (b) Day-by-day sex ratios of species of the 4 categories. Each line represents a species. The points represent mean sex ratios per day across replicates and the error bars represent standard error of the mean. The day-by-day sex ratios calculated from less than 10 progeny are excluded. Note that for the red, purple and blue panels, only 4 exemplary species are displayed. Other species are presented in Figure S1.

Examining the sex ratios by day shows heterogenous profiles across species. For the nine species with female(hermaphrodite)-bias in both the unlimited mating and limited mating experiments, the sex ratios per day show strongest female(hermaphrodite)-bias at the beginning and then a diminishing bias (with some fluctuations) to about or below 50% at the end of reproduction in both experiments (Figure 2b red panel). For the six species with female bias in the unlimited mating experiment but not in the limited mating experiment, the sex ratios were female-biased throughout the reproductive period under unlimited mating but fluctuating around 50% under limited mating (Figure 2b purple panel). Amongst the species with no sex ratio bias in unlimited mating, *C.* sp. 41 was the only species that had female bias in limited mating. The unlimited mating of *C.* sp. 41 showed a fluctuating sex ratio around 50% throughout the reproductive period, while the two-day profile of 5-hour mating showed a female bias on the first day and a decrease to 50% on the second day (Figure 2b brown panel). The seven species with no sex ratio bias in either experiment had sex ratios fluctuating around 50% throughout the reproductive period in both experiments (Figure 2b blue panel).

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Discussion We found prevalent female sex ratio bias (15 out of 23) among the Caenorhabditis species and none in the other direction. The ancestral state was inferred to be female biased, with seven transitions from female bias to no bias in the phylogeny. While most species (16 out of 23) showed consistent sex ratio bias or no bias between the unlimited mating and limited mating experiments, six species had a female(hermaphrodite)-biased sex ratio in the unlimited experiment but no bias in the limited mating experiment, consistent with sperm competition as a possible explanation. Our genus-wide survey of progeny sex ratios sheds light on the evolution of sex ratio bias in this genus. Female-biased sex ratios are frequently found in parasitic nematodes (15, 35-37). Theory predicts female-biased sex ratios in parasites due to confined dispersion and high inbreeding (4, 38, 39). We found that the species of the free-living nematode Caenorhabditis also show prevalent female-biased sex ratios. Though being free-living and dwelling in diverse habitats, many Caenorhabditis species probably share the features of life history such as boom-andbust population growth in ephemeral habitats, active dispersal seeking, and strong founder effect followed by population re-expansion, such as found in extensive sampling of *C. elegans* (40-42). These life history features may result in high inbreeding rates and intense local competition for mating between kin. Hence, a female(hermaphrodite)-biased sex ratio may be favored, according to the theory of LMC. Compared to their parasitic relatives, despite the very distinct life styles, sex ratio bias may have evolved in parallel in the free-living Caenorhabditis species. Alternatively, sex ratio bias is widely conserved across diverse nematode taxa. This hypothesis could be tested with a broader survey of sex ratios in freeliving nematode species outside of *Caenorhabditis* genus. Based on the prevalent sex ratio bias detected in these 23 species, the ancestral state was

inferred to be female-biased, with seven transitions from female bias to equal sex ratio along

the phylogeny. This suggests that sex ratio bias could be a phenotype that can be frequently gained or lost, possibly reflecting the adaptation to the respective habitats and life histories in these species. Despite common features of their life styles, *Caenorhabditis* species dwell in ecologically diverse habitats, ranging from cattle auditory canals to rotting fruits and manmade compost (27). Except for a few species that have been sampled extensively, such as *C. elegans*, *C. briggsae*, and *C. remanei*, (40, 43-45), the natural habitats of most *Caenorhabditis* species are largely unknown, mainly because they have been sampled very rarely. More knowledge about the ecology and natural history of *Caenorhabditis* species might thus be able to explain the driving forces of sex ratio evolution in these species.

Of the three androdioecious *Caenorhabditis* species, *C. elegans*, *C. briggsae*, and *C. tropicalis*, only *C. briggsae* showed a hermaphrodite-biased sex ratio in out-crossed progeny. As the hermaphrodites can self-fertilize, the role of males in these species is obscure (46) but is likely important for rapid adaptation, such as to pathogens (47). Production of male progeny would take up brood "quota" but does not directly contribute to population growth (48). Field studies of *C. elegans* have rarely found males in the wild, and wild *C. elegans* largely suffer from outcrossing depression (49-51). Thus, a hermaphrodite-biased sex ratio may be favored in androdioecious species. However, *C. elegans* and *C. tropicalis* showed no hermaphrodite-biased sex ratio, suggesting unknown ecological factors or historical contingencies that may be contributing to offspring sex ratios in these hermaphroditic species.

Our strategy of a broad survey across species with limited diversity of strains within species assumes that the sex ratio status of the tester strain is representative for each species. An alternative scenario is that sex ratio is a trans-species polymorphism in the genus. As we used a single strain for the sex ratio assays for most species (16 of 23), we cannot exclude the possibility that, for some of these species, the sex ratio bias we detected was specific to those

strains. However, for the four female-male species for which we did have multiple strains, we tested sex ratios for multiple strains as well as for inter-strain crosses. We found that both the intra-strain and inter-strain crosses yielded qualitatively consistent sex ratio bias within species. For the three androdioecious species, the sex ratios of multiple strains also showed consistent non-bias in *C. elegans* and *C. tropicalis*. With an extensive survey of *C. briggsae* strains, we found that most of the strains had significantly hermaphrodite-biased sex ratios, consistent with the common lab strain AF16. Although the other *C. briggsae* strains did not have significant sex ratio bias, they mostly had more hermaphrodites than males in the replicates, except a few outlier strains. These results together suggest that the prevalent sex ratio bias in *Caenorhabditis* nematodes is rather a stable trait within species rather than a trans-species polymorphic trait.

Despite the qualitative consistency in female bias, *C. nigoni*, *C. remanei*, and *C.* sp. 33 showed quantitative differences in sex ratios between strains, as indicated by ANOVA tests, suggesting a contribution of genetic differences in the female-biased sex ratios. On the other hand, *C. latens*, *C. elegans*, and *C. tropicalis* had quantitatively constant sex ratios among strains, suggesting genetic constraints that govern the bias or non-bias.

To investigate sperm competition as a potential mechanism underlying the sex ratio bias, as previously found in *C. briggsae* (33), we conducted mating experiments where mating was limited for a few hours as opposed to the unlimited mating experiments where mating was allowed for the entirety of adulthood. A contrast of female biased sex ratios in unlimited mating experiment and an equal sex ratio in limited mating experiment suggests sperm competition. In the unlimited mating experiments, the couples probably mated repeatedly and the X-bearing sperm would be refilled and therefore X-bearing sperm would always take precedence over the nullo-X sperm, resulting in an overall female-biased sex ratio. In contrast,

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in the limited mating experiments, the amount of sperm transferred was limited and all sperm were presumably used, resulting in an overall equal sex ratio. Consistent with this sperm competition model, six of the 15 species that showed significant female-biased sex ratio in unlimited mating did not have a sex ratio bias in limited mating. The day-by-day sex ratios of these six species also show female bias in unlimited mating throughout the reproductive period and equal sex ratio in limited mating, consistent with the sperm competition scenario. On the other hand, nine species showed female bias in both mating experiments. The day-byday profile of these nine species in both experiments showed female bias at the beginning and then declined to equal sex ratio or male bias in the following days. These observations suggest that sperm competition plays, at least, a partial role in contributing to female-biased sex ratios in both experiments. In C. briggsae, LaMunyon and Ward (1997) conducted 3-hour mating as well as 8-hour mating experiments and found the overall sex ratio was equal after 3-hour mating whereas it was hermaphrodite-biased after 8-hour mating (33). Here, we found a hermaphrodite-biased sex ratio after 5 hours of mating. These together suggest that the amount of sperm ejaculated into the hermaphrodite is a limiting factor for sperm competition and hence sex ratio bias. In addition, we found one species, C. sp. 41, which had no sex ratio bias under unlimited mating conditions while limited mating yielded a female bias. The dayby-day sex ratios show a slightly female-biased sex ratio on the first day in both experiments, suggesting sperm competition may be present but very weak in this species. Finally, the remaining nine species did not have sex ratio bias in either mating experiments, and the dayby-day sex ratios fluctuated around 50:50. Thus, there is no indication of sperm competition in these species. The presence of selfish genetic elements could provide an alternative mechanism for sex ratio bias where the proportion of X-bearing sperm is selfishly enhanced in some species. Examples include segregation distorter on the X chromosome of Drosophila simulans (52) and

asymmetric division in spermatocytes in a *Rhabditis* nematode (53). So far, several selfish elements have been discovered in *Caenorhabditis* nematodes, but they all reside on autosomes (54-57).

While equal sex ratios are presumably predominant in nature, biased sex ratios may be largely underappreciated. In this study, we carried out a broad survey of sex ratio bias in outcrossed progeny across *Caenorhabditis* nematodes. Our findings of prevalent sex ratio bias add to the limited knowledge about sex ratio bias in the animal kingdom, and provide evidence that sex ratio bias can evolve rapidly within a single genus.

Materials and Methods

Species and culture

We examined 23 *Caenorhabditis* species for progeny sex ratio in this study (Table 1), including four new species: *C.* sp. 33, *C.* sp. 41, *C.* sp. 44, and *C.* sp. 45. These four new species were placed onto the phylogenetic tree based on their ITS2 sequences (the intergenic region between the 5.8S and LSU rRNA genes) (58). These 23 species comprise 17 species from the *Elegans* supergroup and 6 from the *Drosophila* supergroup. Of the 17 species from the *Elegans* supergroup, three were from the *Japonica* group and the rest were from the *Elegans* group, including the three androdioecious species (*C. elegans, C. briggsae,* and *C. tropicalis*). All species were grown at room temperature (23-24°C) on nematode growth media agar plates seeded with OP50 *Escherichia coli* bacteria.

Sex ratio assay

Mating experimental design

For the female-male species, sex ratios were assayed by intra-strain crosses, i.e., females and males from the same wild-type isofemale strains (tester strains). For most of the

female-male species, we had only one tester strain, except for four of the species (*C. nigoni*, *C. latens*, *C. remanei*, and *C.* sp. 33). For these four species we had more than one strain available at the start of this experiment, so the sex ratios of intra-strain crosses were tested for multiple strains. We also conducted inter-strain crosses for these four species to test the possibility that sex ratio bias was due to sex-specific consequences of inbreeding depression (34). For *C. nigoni*, for which we had two strains, we intercrossed the two strains and then crossed the heterozygous F1 males with the maternal strain. For the three species, *C. latens*, *C. remanei*, and *C.* sp. 33, for which we had three strains, we first crossed two strains and then crossed the heterozygous F1 males with the third strain (Table S1). We scored the two sexes in the F2 progeny.

For the androdioecious species, we crossed males from wild-type strains (tester strain) to hermaphrodite strains carrying a recessive mutation, so that the outcrossed progeny were visually identifiable. The recessive morphological mutant strains had Uncoordinated (Unc) or Dumpy (Dpy) phenotypes: *C. elegans* (BRC0189, *unc-119(ed9))*; *C. briggsae* (BRC0258, *unc(ant10)*); and *C. tropicalis* (BRC0419, *dpy(ant23)*). For each of these androdioecious species, we had multiple tester strains, especially for *C. briggsae*, for which we had many isolates collected in Taiwan. Because androdioecious species are normally inbred, we tested sex ratios using males from different strains but did not generate heterozygous F1 males to test an inter-strain effect.

Time limitations for mating

We set up mating experiments to assay sex ratio in the same manner across all the species tested. In the "unlimited mating" experiment, we placed one L4 female (or hermaphrodite) and one L4 male on a fresh 55 mm diameter Petri plate and then transferred them together every one or two days to fresh plates until they died or produced no more eggs. When the progeny reached the L4 or adult stage, the numbers of outcrossed females (or hermaphrodites) and males per plate were manually scored under the microscope. For the male-female species, all progeny were counted, while for the androdioecious species,

only wild-type cross progeny were counted whereas selfed Unc or Dpy progeny were ignored. Mating pairs with total number of out-crossed progeny smaller than 40 were excluded from further analyses to ensure adequate statistical power. For each test of the strains, we had at least three replicate mating pairs. The counts of progeny of the two sexes for each replicate were used for further statistical analyses (see below).

In addition to the unlimited mating experiment, to interrogate the potential role of competition between male X-bearing sperm and the nullo-X counterpart in progeny sex ratio bias, we conducted sex ratio assays by "limited mating" for one tester strain per species. To do so, L4 females (hermaphrodites) and L4 males were isolated one day prior to the cross to ensure their virginity and matured singly overnight. The next day, one male was added to one isolated female (hermaphrodite). The male was removed when mating plugs were observed on the females (hermaphrodites) or after five hours. Mated females (hermaphrodites) were transferred daily to new plates. For *C. tropicalis*, all pairs failed to mate within five hours, so we crossed one hermaphrodite with three males to increase the chance of mating. The progeny sex ratios were scored as above.

Statistical analysis

For each of the sex ratio assays, unlimited mating or limited mating, we tested whether the progeny sex ratio was biased. For each replicate within the tester strain, the counts of total females (hermaphrodites) and total males were used for the binomial test. As the majority of crosses yielded more female (hermaphrodite) than male progeny, we tested whether the proportions of females (hermaphrodites) significantly exceeded equality (R, binom.test, alternative = "greater"). The *P*-values of the replicates within a strain were corrected for multiple testing for the numbers of replicates using the Benjamini-Hochberg method (59) and then combined to yield the overall *P*-value for the strain (Fisher's method, R package metaseqR (60)). For species with only one strain, the strain *P*-value was used to represent the species. For species with multiple strains, the *P*-values of the strains were again

corrected for multiple strains and then combined to yield the *P*-value for the species. The species *P*-values were corrected a final time for multiple testing for the 23 species. Species with the corrected *P*-values smaller than 0.05 were defined as having sex ratio bias. Based on the states of sex ratio bias or non-bias of the 23 species (unlimited mating) and the phylogenetic tree (61), we inferred the ancestral state of sex ratio bias and the evolutionary transitions between bias and non-bias among these species, using the maximum parsimony method in MESQUITE v. 3.10 (62). In addition, for species with multiple strains, we applied ANOVA to test if sex ratios are significantly different between strains. All statistical analyses were performed in R v.3.2.3 except otherwise indicated (63).

Acknowledgements

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Author contributions

Y.-H.L., and J.W. conceived and designed the study. Y.-H.L, T.S.L, J.-C.H., F.-J.Y., T.C., and J.W. conducted experiments. C.B. contributed strains and help revised the manuscript. Y.H. and Y.-H.L. analyzed the data. Y.-H.L., Y.H., and J.W wrote the manuscript with input from others.

Competing interest

The authors declare that they have no conflict of interest.

Tables

Table 1. Sex ratios in the unlimited mating experiment

	Table 1. Ock	Tatios III tile	diminited in	ating expens				
species	strain	no. of replicates	median brood size	median sex ratio	binomial test p value (strain)	sex ratio (species)	combined p value (species)	female bias
nigoni	BRC20235 BRC20079	4 5	379.5 695	0.590 0.534	0.000 0.004	0.562	0.000	yes
briggsae	AF16	5	107	0.656	0.000	0.656	0.000	yes
zanzibari	BRC20266	7	396.5	0.517	0.150	0.517	0.216	no
sinica	BRC20243	4	149	0.525	0.037	0.525	0.061	no
sp41	BRC20276	9	647	0.523	0.372	0.523	0.475	no
•	NIC1200	6	687.5	0.525	0.000			
latens	NIC1201	5	400	0.538	0.000	0.524	0.000	yes
	NIC1207	6	392.5	0.509	0.000			•
	BRC20108	4	232	0.600	0.000			
remanei	JU1084	5	854	0.511	0.058	0.556	0.000	yes
	MY31	5	563	0.559	0.000			•
	BRC20005	5	539	0.540	0.002			
sp33	BRC20258	5	497	0.635	0.000	0.578	0.000	yes
	BRC20273	5	344	0.558	0.000			
wallacei	JU1904	6	109	0.516	0.978	0.516	1.000	no
	BRC20400	8	211	0.545	0.097			
tropicalis	JU1373	11	162	0.503	0.877	0.533	0.652	no
	NIC58	3	60	0.550	0.235			
brenneri	JU1397	5	326	0.519	0.820	0.519	0.993	no
doughertyi	JU1333	1	265.5	0.745	0.000	0.745	0.000	yes
sp44	BRC20300	5	342	0.576	0.000	0.576	0.000	yes
	AB1	4	243	0.487	1.000			
elegans	BRC20067	4	123	0.490	0.997	0.493	1.000	no
	N2	3	549	0.503	0.828			
becei	QG704	6	706	0.600	0.000	0.600	0.000	yes
panamensis	QG702	4	681.5	0.510	0.222	0.510	0.407	no
imperialis	JU1905	5	402	0.570	0.000	0.570	0.000	yes
virilis	JU1968	4	274	0.521	0.337	0.521	0.521	No
vivipara	NIC1070	4	392	0.633	0.000	0.633	0.000	yes
castelli	JU1427	4	193.5	0.550	0.071	0.550	0.167	no
waitukubuli	NIC564	5	341	0.558	0.000	0.558	0.000	yes
portoensis	EG4788	8	419	0.591	0.000	0.591	0.000	yes
sp45	NIC759	5	369	0.618	0.000	0.618	0.000	yes

Table 2. Sex ratios in the limited mating experiment

					binomial	binomial	
species	strain	no. of	median	median	test	test	female
Species	Strain	replicates	brood size	sex ratio	combined	adjusted p	bias
					p value	value	
nigoni	BRC20235	9	162	0.54	1.34E-01	2.57E-01	no
briggsae	AF16	8	81	0.58	1.35E-02	3.44E-02	yes
zanzibari	BRC20266	10	219.5	0.524	7.24E-01	9.80E-01	no
sinica	BRC20243	9	217	0.521	5.14E-01	7.88E-01	no
sp41	BRC20276	8	191.5	0.546	1.42E-04	1.09E-03	yes
latens	NIC1207	10	378	0.521	6.20E-03	2.04E-02	yes
remanei	BRC20108	11	90	0.508	6.50E-01	9.34E-01	no
sp33	BRC20005	9	134	0.616	1.26E-07	2.90E-06	yes
wallacei	JU1904	9	53	0.528	9.62E-01	9.88E-01	no
tropicalis	JU1373	10	118.5	0.512	9.88E-01	9.88E-01	no
brenneri	JU1397	5	150	0.49	9.69E-01	9.88E-01	no
doughertyi	JU1333	8	190.5	0.558	4.87E-04	2.80E-03	yes
sp44	BRC20300	10	171.5	0.57	5.69E-05	6.54E-04	yes
elegans	N2	8	101.5	0.512	9.36E-01	9.88E-01	no
becei	QG704	15	139	0.519	1.87E-01	3.30E-01	no
panamensis	QG702	9	286	0.514	8.23E-01	9.88E-01	no
imperialis	JU1905	10	203	0.556	1.29E-03	4.93E-03	yes
virilis	JU1968	9	72	0.529	8.25E-01	9.88E-01	no
vivipara	NIC1070	8	145.5	0.561	7.31E-03	2.10E-02	yes
castelli	JU1427	7	64	0.548	2.20E-01	3.62E-01	no
waitukubuli	NIC564	7	95	0.509	9.45E-04	4.35E-03	yes
portoensis	EG4788	6	171.5	0.535	1.21E-01	2.52E-01	no
sp45	NIC759	7	105	0.549	1.68E-02	3.87E-02	yes

- 452 Supplementary Figures and Tables
- 454 Figure S1. Day-by-day sex ratios of species not included in Figure 2
- 455 Table S1. Sex ratios of inter-stain crosses
- 456 Table S2. Sex ratios of 31 *C. briggsae* strains
- Table S3. Day-by-day sex ratios by replicates of each species (unlimited mating)
- Table S4. Day-by-day sex ratios by replicates of each species (limited mating)

References

- 462 1. Darwin C. The descent of man and selection in relation to sex. 2nd ed: John Murray,
- 463 London; 1874.

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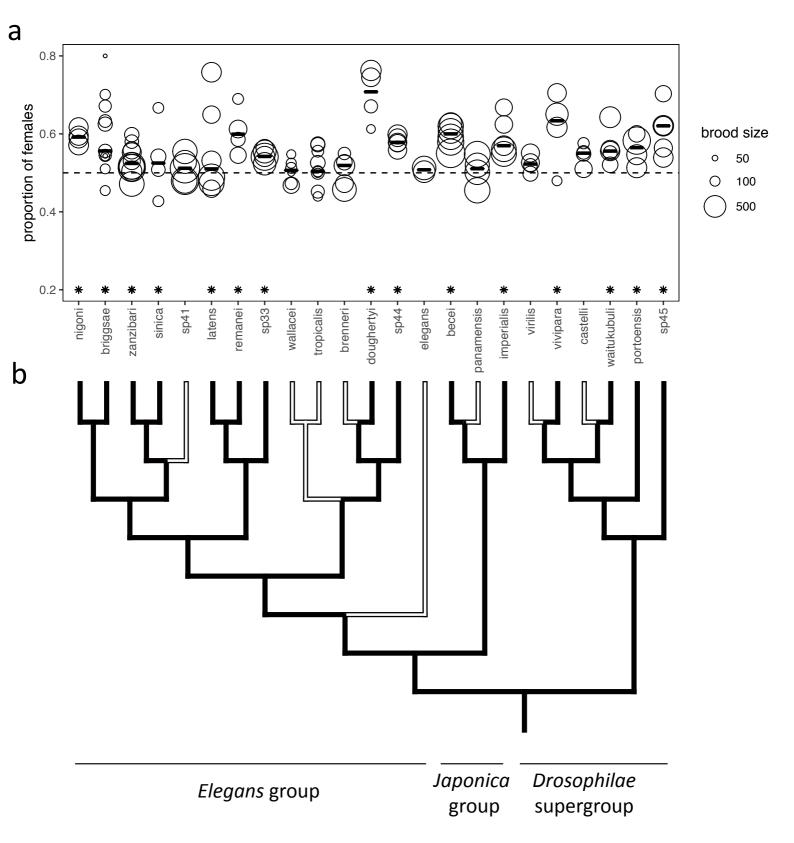
- 464 2. Edwards AWF. Natural Selection and the Sex Ratio: Fisher's Sources. The American
- 465 Naturalist. 1998;151(6):564-9.
- 466 3. Fisher RA. The genetical theory of natural selection: Clarendon, Oxford; 1930.
- 467 4. Hamilton WD. Extraordinary sex ratios. Science. 1967;156(3774):477-88.
- 468 5. West SA, Herre EA, Sheldon BC. The Benefits of Allocating Sex. Science.
- 469 2000;290(5490):288-90.
- 470 6. Charnov EL. The theory of sex allocation: Princeton University Press, Princeton, NJ.;
- 471 1982.
- 472 7. Clark AB. Sex ratio and local resource competition in a prosimian primate. Science.
- 473 1978;201(4351):163-5.
- 474 8. Gowaty PA, Lennartz MR. Sex Ratios of Nestling and Fledgling Red-Cockaded
- Woodpeckers (*Picoides borealis*) Favor Males. The American Naturalist. 1985;126(3):347-53.
- 476 9. Emlen ST, Emlen JM, Levin SA. Sex-Ratio Selection in Species with Helpers-At-The-
- 477 Nest. The American Naturalist. 1986;127(1):1-8.
- 478 10. Trivers RL, Willard DE. Natural Selection of Parental Ability to Vary the Sex Ratio of
- 479 Offspring. Science. 1973;179(4068):90-2.
- 480 11. Paul RE, Raibaud A, Brey PT. Sex ratio adjustment in *Plasmodium gallinaceum*.
- 481 Parassitologia. 1999;41(1-3):153-8.
- 482 12. Osgood SM, Eisen RJ, Schall JJ. Gametocyte sex ratio of a *malaria* parasite:
- 483 experimental test of heritability. The Journal of parasitology. 2002;88(3):494-8.
- 484 13. Reece SE, Drew DR, Gardner A. Sex ratio adjustment and kin discrimination in malaria
- 485 parasites. Nature. 2008;453(7195):609-14.
- 486 14. Ferguson DJ. *Toxoplasma gondii* and sex: essential or optional extra? Trends in
- 487 parasitology. 2002;18(8):355-9.
- 488 15. Haukisalmi V, Henttonen H, Vikman P. Variability of sex ratio, mating probability and
- 489 egg production in an intestinal nematode in its fluctuating host population. International

- 490 Journal for Parasitology. 1996;26(7):755-64.
- 491 16. Kloch A, Michalski A, Bajer A, Behnke J. Biased sex ratio among worms of the family
- 492 Heligmosomidae--searching for a mechanism. Int J Parasitol. 2015;45(14):939-45.
- 493 17. Herre EA. Sex ratio adjustment in fig wasps. Science. 1985;228(4701):896-8.
- 494 18. Werren JH. Sex Ratio Adaptations to Local Mate Competition in a Parasitic Wasp.
- 495 Science. 1980;208(4448):1157-9.
- 496 19. David M. Shuker, Ido Pen, Alison B. Duncan, Sarah E. Reece, Stuart A. West. Sex
- 497 Ratios under Asymmetrical Local Mate Competition: Theory and a Test with Parasitoid
- 498 Wasps. The American Naturalist. 2005;166(3):301-16.
- 499 20. Griffiths NT, Godfray HCJ. Local mate competition, sex ratio and clutch size in bethylid
- wasps. Behavioral Ecology and Sociobiology. 1988;22(3):211-7.
- 501 21. Komdeur J. Facultative sex ratio bias in the offspring of Seychelles warblers.
- 502 Proceedings of the Royal Society B: Biological Sciences. 1996;263(1370):661-6.
- 503 22. Komdeur J, Daan S, Tinbergen J, Mateman C. Extreme adaptive modification in sex
- ratio of the Seychelles warbler's eggs. Nature. 1997;385(6616):522-5.
- 505 23. Malcolm JR, Marten K. Natural selection and the communal rearing of pups in African
- wild dogs (Lycaon pictus). Behavioral Ecology and Sociobiology. 1982;10(1):1-13.
- 507 24. Clutton-Brock TH, Albon SD, Guinness FE. Maternal dominance, breeding success and
- 508 birth sex ratios in red deer. Nature. 1984;308(5957):358-60.
- 509 25. Symington MM. Sex ratio and maternal rank in wild spider monkeys: when daughters
- disperse. Behavioral Ecology and Sociobiology. 1987;20(6):421-5.
- 511 26. Kiontke K, Fitch DH. The phylogenetic relationships of *Caenorhabditis* and other
- 512 rhabditids. WormBook: the online review of C elegans biology. 2005:1-11.
- 513 27. Kiontke K, Sudhaus W. Ecology of Caenorhabditis species. WormBook: the online
- review of C elegans biology. 2006:1-14.
- 515 28. Cutter AD, Payseur BA. Rates of deleterious mutation and the evolution of sex in
- 516 *Caenorhabditis*. Journal of evolutionary biology. 2003;16(5):812-22.
- 517 29. Kiontke K, Gavin NP, Raynes Y, Roehrig C, Piano F, Fitch DHA. Caenorhabditis
- 518 phylogeny predicts convergence of hermaphroditism and extensive intron loss. Proceedings
- of the National Academy of Sciences. 2004;101(24):9003-8.
- 520 30. Haag ES. The evolution of nematode sex determination: *C. elegans* as a reference
- 521 point for comparative biology. WormBook: the online review of C elegans biology. 2005:1-
- 522 14.
- 523 31. Hodgkin J, Horvitz HR, Brenner S. Nondisjunction Mutants of the Nematode
- 524 Caenorhabditis elegans. Genetics. 1979;91(1):67-94.
- 525 32. Teotónio H, Manoel D, Phillips PC. Genetic variation for outcrossing among
- 526 *Caenorhabditis elegans* isolates. Evolution. 2006;60(6):1300-5.
- 527 33. LaMunyon CW, Ward S. Increased competitiveness of nematode sperm bearing the

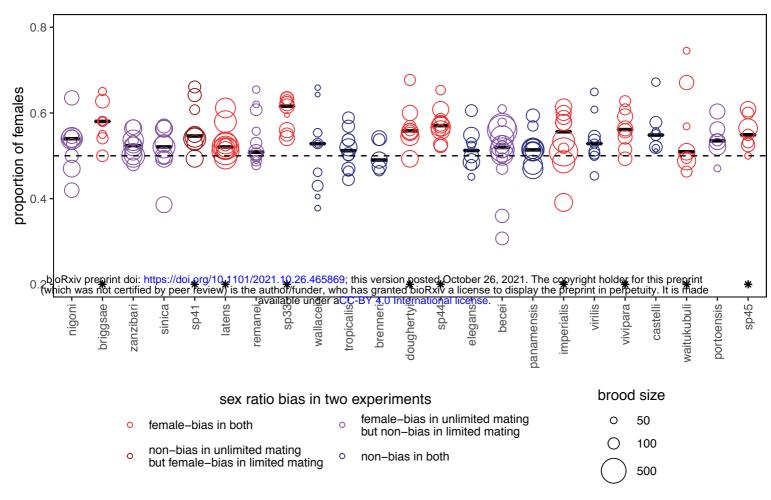
- male X chromosome. Proceedings of the National Academy of Sciences. 1997;94(1):185-9.
- 529 34. Ebel ER, Phillips PC. Intrinsic differences between males and females determine sex-
- specific consequences of inbreeding. BMC Evol Biol. 2016;16:36.
- 531 35. D'Ávila S, Bessa ECA, Souza-Lima S, Rodrigues MLA. Biased sex ratio and niche
- restriction in Baruscapillaria obsignata (Madsen 1945) (Nematoda, Capillariidae) from
- 533 *Columba livia* (Aves, Columbidae). Journal of helminthology. 2012;86(4):401-5.
- 534 36. Craig BH, Pilkington JG, Pemberton JM. Sex ratio and morphological polymorphism in
- an isolated, endemic *Teladorsagia circumcincta* population. Journal of helminthology.
- 536 2010;84(2):208-15.
- 537 37. Seidenberg AJ, Kelly PC, Lubin ER, Buffington JD. Helminths of the Cotton Rat in
- 538 Southern Virginia, with Comments on the Sex Ratios of Parasitic Nematode Populations.
- 539 American Midland Naturalist. 1974;92:320.
- 540 38. West SA, Reece SE, Read AF. Evolution of gametocyte sex ratios in malaria and
- related apicomplexan (protozoan) parasites. Trends in parasitology. 2001;17(11):525-31.
- 542 39. Nee S, West SA, Read AF. Inbreeding and parasite sex ratios. Proceedings Biological
- 543 sciences. 2002;269(1492):755-60.
- 544 40. Petersen C, Dirksen P, Prahl S, Strathmann EA, Schulenburg H. The prevalence of
- 545 Caenorhabditis elegans across 1.5 years in selected North German locations: the importance
- of substrate type, abiotic parameters, and *Caenorhabditis* competitors. BMC ecology.
- 547 2014;14:4.
- 548 41. Félix M-A, Braendle C. The natural history of *Caenorhabditis elegans*. Current Biology.
- 549 2010;20(22):R965-R9.
- 550 42. Frézal L, Félix MA. C. elegans outside the Petri dish. eLife. 2015;4.
- 551 43. Cook DE, Zdraljevic S, Roberts JP, Andersen EC. CeNDR, the Caenorhabditis elegans
- natural diversity resource. Nucleic Acids Res. 2017;45(D1):D650-D7.
- 553 44. Crombie TA, Zdraljevic S, Cook DE, Tanny RE, Brady SC, Wang Y, et al. Deep sampling
- of Hawaiian Caenorhabditis elegans reveals high genetic diversity and admixture with global
- 555 populations. eLife. 2019;8:e50465.
- 556 45. Félix M-A, Duveau F. Population dynamics and habitat sharing of natural populations
- of Caenorhabditis elegans and C. briggsae. BMC Biology. 2012;10(1):59.
- 558 46. Cutter AD, Morran LT, Phillips PC. Males, Outcrossing, and Sexual Selection in
- 559 Caenorhabditis Nematodes. Genetics. 2019;213(1):27-57.
- 560 47. Morran LT, Schmidt OG, Gelarden IA, Parrish RC, 2nd, Lively CM. Running with the
- Red Queen: host-parasite coevolution selects for biparental sex. Science.
- 562 2011;333(6039):216-8.
- 563 48. Yin D, Haag ES. Evolution of sex ratio through gene loss. Proceedings of the National
- 564 Academy of Sciences. 2019;116(26):12919-24.
- 565 49. Barrière A, Félix MA. High local genetic diversity and low outcrossing rate in

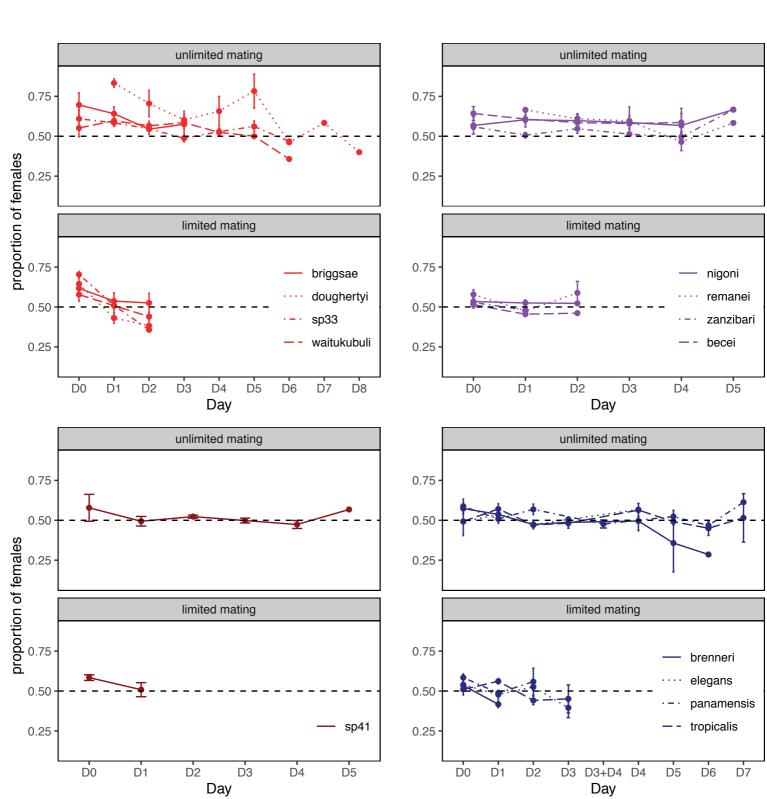
- 566 Caenorhabditis elegans natural populations. Current biology: CB. 2005;15(13):1176-84.
- 567 50. Barrière A, Félix MA. Temporal dynamics and linkage disequilibrium in natural
- 568 Caenorhabditis elegans populations. Genetics. 2007;176(2):999-1011.
- 569 51. Gimond C, Jovelin R, Han S, Ferrari C, Cutter AD, Braendle C. Outbreeding depression
- 570 with low genetic variation in selfing Caenorhabditis nematodes. Evolution.
- 571 2013;67(11):3087-101.
- 572 52. Tao Y, Araripe L, Kingan SB, Ke Y, Xiao H, Hartl DL. A sex-ratio Meiotic Drive System in
- 573 Drosophila simulans. II: An X-linked Distorter. PLOS Biology. 2007;5(11):e293.
- 574 53. Shakes DC, Neva BJ, Huynh H, Chaudhuri J, Pires-daSilva A. Asymmetric spermatocyte
- 575 division as a mechanism for controlling sex ratios. Nature Communications. 2011;2(1):157.
- 576 54. Ben-David E, Burga A, Kruglyak L. A maternal-effect selfish genetic element in
- 577 *Caenorhabditis elegans*. Science. 2017;356(6342):1051-5.
- 578 55. Ben-David E, Pliota P, Widen SA, Koreshova A, Lemus-Vergara T, Verpukhovskiy P, et
- al. Ubiquitous Selfish Toxin-Antidote Elements in Caenorhabditis Species. Current Biology.
- 580 2021;31(5):990-1001.e5.
- 581 56. Noble LM, Yuen J, Stevens L, Moya N, Persaud R, Moscatelli M, et al. Selfing is the
- safest sex for *Caenorhabditis tropicalis*. eLife. 2021;10:e62587.
- 583 57. Seidel HS, Rockman MV, Kruglyak L. Widespread Genetic Incompatibility in C. elegans
- Maintained by Balancing Selection. Science. 2008;319(5863):589-94.
- 585 58. Félix M-A, Braendle C, Cutter AD. A Streamlined System for Species Diagnosis in
- 586 Caenorhabditis (Nematoda: Rhabditidae) with Name Designations for 15 Distinct Biological
- 587 Species. PLOS ONE. 2014;9(4):e94723.
- 588 59. Benjamini Y, Hochberg Y. Controlling the False Discovery Rate: A Practical and
- 589 Powerful Approach to Multiple Testing. Journal of the Royal Statistical Society Series B
- 590 (Methodological). 1995;57(1):289-300.
- 591 60. Moulos P, Hatzis P. Systematic integration of RNA-Seq statistical algorithms for
- accurate detection of differential gene expression patterns. Nucleic Acids Res.
- 593 2014;43(4):e25-e.
- 594 61. Kiontke KC, Félix M-A, Ailion M, Rockman MV, Braendle C, Pénigault J-B, et al. A
- 595 phylogeny and molecular barcodes for Caenorhabditis, with numerous new species from
- rotting fruits. BMC Evolutionary Biology. 2011;11(1):339.
- 597 62. Maddison WP, Maddison DR. Mesquite: a modular system for evolutionary analysis.
- 598 3.70 ed2021.

- 599 63. R Development Core Team. A language and environment for statistical computing.
- 600 Vienna, Austria: R Foundation for Statistical Computing; 2020.



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Prevalent sex ratio bias in Caenorhabditis nematodes

Supplementary Information

Figure S1. Day-by-day sex ratios of species not included in Figure 2

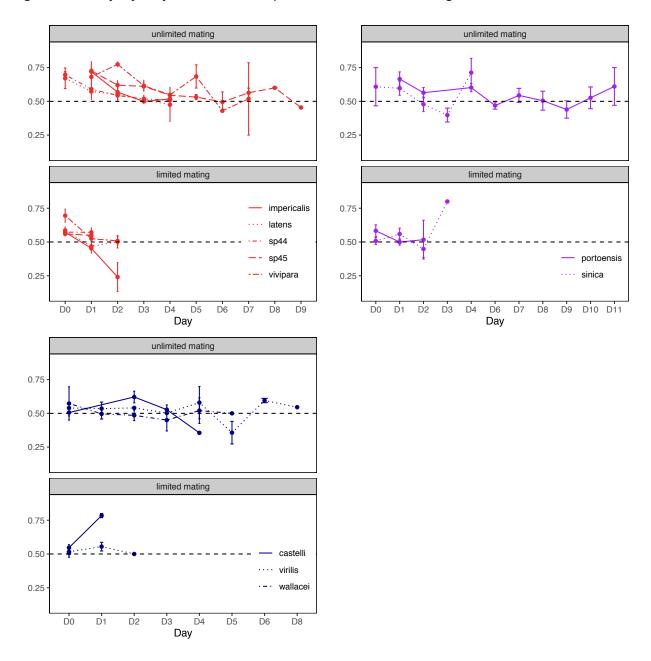


Table S1. Sex ratios of inter-strain crosses

species	rplc	paternal_grandma	paternal_grandpa	maternal_stain	total	proportion_female	p.binomial	p.adjusted
nigoni	NN	BRC20235	BRC20079	BRC20235	410	0.495	0.598	0.598
nigoni	00	BRC20235	BRC20079	BRC20235	296	0.551	0.046	0.069
nigoni	PP	BRC20235	BRC20079	BRC20235	321	0.564	0.013	0.025
nigoni	KK	BRC20079	BRC20235	BRC20079	523	0.528	0.110	0.132
nigoni	LL	BRC20079	BRC20235	BRC20079	565	0.577	0.000	0.000
nigoni	НН	BRC20079	BRC20235	BRC20079	572	0.601	0.000	0.000
latens	E	NIC1201	NIC1207	NIC1200	95	0.589	0.050	0.060
latens	D	NIC1201	NIC1207	NIC1200	482	0.541	0.038	0.057
latens	F	NIC1201	NIC1207	NIC1200	469	0.563	0.004	0.007
latens	Α	NIC1207	NIC1201	NIC1200	303	0.584	0.002	0.006
latens	В	NIC1207	NIC1201	NIC1200	590	0.603	0.000	0.000
latens	С	NIC1207	NIC1201	NIC1200	33	0.485	0.636	0.636
sp33	Α	BRC20258	BRC20005	BRC20273	237	0.599	0.001	0.003
sp33	D	BRC20258	BRC20005	BRC20273	112	0.669	0.000	0.001
sp33	E	BRC20005	BRC20258	BRC20273	90	0.611	0.022	0.033
sp33	F	BRC20005	BRC20258	BRC20273	284	0.679	0.000	0.000
remanei	Α	my31	BRC20108	JU2084	417	0.535	0.085	0.102
remanei	В	my31	BRC20108	JU2084	570	0.565	0.001	0.002
remanei	GG	my31	BRC20108	JU2084	656	0.550	0.006	0.008
remanei	JJ	BRC20108	my31	JU2084	250	0.656	0.000	0.000
remanei	KK	BRC20108	my31	JU2084	346	0.604	0.000	0.000

Table S2. Sex ratios of 31 C. briggsae strains

strain	no.rplc	median brood size	median sex ratio	p.combined	p.adjusted
BRC20069	5	145	0.592	0.000	0.001
BRC20076	6	108.5	0.529	0.000	0.000
BRC20085	4	71	0.594	0.015	0.024
BRC20088	3	117	0.548	0.045	0.061
BRC20092	4	122	0.542	0.213	0.236
BRC20093	4	156	0.589	0.000	0.001
BRC20095	4	102.5	0.545	0.396	0.423
BRC20096	4	78	0.618	0.044	0.061
BRC20099	3	81	0.636	0.000	0.001
BRC20102	3	66	0.621	0.058	0.074
BRC20103	6	113	0.616	0.000	0.000
BRC20105	5	146	0.607	0.000	0.000
BRC20115	4	158.5	0.569	0.018	0.028
BRC20120	5	160	0.539	0.101	0.120
BRC20228	4	92.5	0.601	0.001	0.003
BRC20234	5	100	0.505	0.659	0.659
BRC20242	2	128	0.609	0.002	0.004
BRC20244	5	169	0.613	0.000	0.001
BRC20246	4	95.5	0.576	0.000	0.000
BRC20255	4	98.5	0.584	0.013	0.022
BRC20269	3	103	0.576	0.000	0.000
BRC20294	5	138	0.569	0.037	0.054
BRC20299	5	200	0.542	0.105	0.120
BRC20312	6	133	0.609	0.000	0.000
BRC20324	5	139	0.538	0.410	0.424
BRC20334	5	219	0.544	0.072	0.089
BRC20339	8	208	0.498	0.000	0.000
BRC20341	5	244	0.592	0.010	0.018
BRC20345	5	193	0.597	0.001	0.002
BRC20347	5	306	0.510	0.003	0.007
BRC20348	5	244	0.717	0.000	0.000