Muscle systems and motility of early animals highlighted by cnidarians from the basal Cambrian

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Abstract

Although fossil evidence suggest that various animal groups were able to move actively through their environment in the early stages of their evolution, virtually no direct information is available on the nature of their muscle systems. The origin of jellyfish swimming, for example, is of great concern to biologists. Exceptionally preserved muscles are described here in benthic peridermal olivooid medusozoans from the basal Cambrian of China (Kuanchuanpu Formation, ca. 535 Ma) that have direct equivalent in modern medusozoans. They consist of circular fibers distributed over the bell surface (subumbrella) and most probably have a myoepithelial origin. This is the oldest record of a muscle system in cnidarians and more generally in animals. This basic system was probably co-opted by younger early Cambrian jellyfish to develop capacities for the jet-propelled swimming within the water column. Additional lines of fossil evidence obtained from ecdysozoans (worms and panarthropods) show that the muscle systems of early animals underwent a rapid diversification through the early Cambrian and increased their capacity to colonize a wide range of habitats both within the water column and sediment at a critical time of their evolutionary radiation.

Introduction

Cnidarians are generally seen as the sister group to bilaterians (Brusca et al., 2016; Erwin et al., 2011;

Leclère and Röttinger, 2017; Zapata et al., 2015) and are represented by a huge variety of jellyfish, sea anemones, corals, sea fans, hydrozoans (including the colonial siphonophores) and less familiar parasitic groups (Raikova, 1988). Although often sessile (polyps) or parasitic, many of them are motile animals and a large proportion of them (such as jellyfish) use their muscles to move very actively through the water column. In contrast with bilaterians, cnidarians owe most of their contractile power to epitheliomuscular or myoepithelial cells that line both epithelial body layers (Brusca et al., 2016; Schmidt-Rhaesa, 2007). These specialized cells contain interconnected contractile basal extensions (myonemes or myofilaments) that altogether form longitudinal or circular sheets and play virtually the same role as the muscle layers of other animals. Epitheliomuscular cells are connected to nerve cells via chemical synapses (Westfall et al. 1971). Cnidarian muscles are characterized by multifunctional capacities and plasticity and perform key functions in locomotion, defense from predators, feeding and digestion at all stages (planula, polyp and medusa stages; see ref Leclère and Röttinger, 2017). In medusae, locomotion is performed by the rhythmic pulsation of circular sheets of epithelial striated muscles located around the bell margins and over the subumbrellar surface. These contractions are counteracted by the elastic properties and antagonistic force of the mesoglea and result in expelling water from beneath the bell and thus moving the medusa via jet propulsion (Brusca et al., 2016).

The Precambrian fossil record of cnidarians remain patchy and controversial although molecular models often predict a very ancient (e.g. pre-Ediacaran) origin of the group (Erwin et al., 2011). For example, *Haootia quadriformis* from the Ediacaran (Fermeuse Formation; ca. 560 Ma; Newfoundland, eastern Canada) roughly resembles modern stalked jellyfish, such as staurozoans, and bears very fine wrinkles interpreted as putative coronal muscles (Liu et al., 2014; 2015). Whether this external network corresponds to underlying muscles is uncertain and seems at odd with the assumed sessile lifestyle of the animal. Numerous circular forms with a radial pattern have been described in the Ediacaran (e.g. *Cyclomedusa*; Ukraine, Russia; see refs. Zaika-Novatskii et al., 1968; Fedonkin, 1981). Although some of them may potentially represent bell imprints of jellyfish, others probably have a different origin (e.g. circular holdfasts of non-cnidarian sessile organisms or possible gas-escape sedimentary structures; see ref. Sun, 1986). In contrast, convincing evidence for ancestral medusozoans comes from the early Cambrian Chengjiang Lagerstätte (ca. 521 Ma; Yunnan Province China) and is best exemplified by *Yunnanoascus haikouensis* that shares a set of morphological features with modern jellyfish such as a tetramerous symmetry, rhopalia, long tentacles around the bell margin, and a possible manubrium in the central part of the bell (Han et al., 2016a). Although Chengjiang fossils usually show extremely fine details of soft animal tissues and organs (including digestive, nervous and reproductive systems), muscles remain elusive, and no trace of possible coronal muscles can be seen in *Yunnanoascus*. Other jellyfish from the mid-Cambrian Marjum Formation (ca. 505 Ma, Utah, USA; see ref. Cartwright et al., 2007) display fine recognizable anatomical details such as the exumbrella and subumbrella, tentacles and relatively well-preserved coronal muscles that suggest swimming capacities.

The Kuanchuanpu Formation (ca. 535 Ma, lowermost Cambrian Fortunian Stage; Shaanxi Province, south China) yields a great variety of three-dimensionally preserved microfossils including cnidarians such as Olivooidae (Dong et al. 2013, 2016; Han et al. 2013, 2016b; Liu et al. 2014; 2017; Steiner et al., 2014). The developmental sequence of *Olivooides* starts with a spherical embryo that, after hatching,

gives rise to conical juveniles (Bengtson and Yue, 1997), suggesting a direct development (Han et al., 2013; Steiner et al., 2014; Wang et al., 2020) with no counterpart among modern cnidarians that all grow via planula larval stage. However, microtomography clearly show that post-embryonic *Olivooides* do have anatomical features typical of modern cnidarians, a radial symmetry, single body opening, exumbrella and subumbrella, interradial septa (internal ridges), gonads, manubrium, oral lips, apertural lobes, tentacles, frenula and velaria (Dong et al. 2013, 2016; Han et al. 2013, 2016b; Wang et al., 2020), which collectively support a placement of *Olivooides* and its related forms within Medusozoa (Dong et al. 2013, 2016; Han et al. 2013, 2016b; Liu et al. 2014; 2017; Wang et al., 2020). Wang et al. (2017) reported possible coronal muscles around the aperture of *Sinaster* (Olivooidae) but did not investigate their organization and possible function. We describe here secondarily phosphatized muscle fibers preserved in three dimensions, in post-embryonic stages of olivooids from the early Cambrian (Fortunian) Kuanchuanpu Formation. They represent the oldest occurrence of muscle tissue in cnidarians and more generally in animals. We also address the nature (e.g. myoepithelial) and function of this muscle system through detailed comparisons with modern jellyfish.

These new findings prompted us to re-examine and integrate fossil data obtained from other early Cambrian groups such as ecdysozoans (e.g. worms, lobopodians; see Budd, 1998; Vannier and Martin 2017; Young and Vinther, 2017; Zhang et al., 2016) that altogether shed light on the diversity and functions of muscle systems in early animals.

Results

The twelve fossil specimens studied here have the diagnostic features of post-embryonic olivooids (Dong et al., 2013; 2016; Han et al., 2013; 2016b; Wang et al., 2017; 2020), such as an ovoid shape with a pentaradial symmetry and the presence of a periderm, apertural lobes, exumbrella, perradial ridges and interradial furrows (Figures. 1 and 2A-E—figure supplements 1-3). They also display a well-preserved network of circular fibers (Figures. 1 and 2A-E), tentatively interpreted as coronal muscles by Wang et al. (2017) in a pilot study.

The body has a consistent ovoid shape and size (diameter between 560 and 580 µm) and is enclosed by a smooth periderm (5-10 µm in thickness) (Figure 1A—figure supplements 2A and 3C, E-H). Centripetal, triangular projections, termed perradial apertural lobes (see refs. Han et al., 2013; Wang et al., 2020) can be seen around the oral aperture of most specimens. They correspond to perradii and display a pentaradial symmetry (Figures 1A and 2A, D—figure supplements 1-3). The partial loss of the periderm and perradial apertural lobes in numerous specimens reveals a fine network of underlying closely packed, circular fibers, that are best developed around the oral aperture where they form four or five separate concentric bundles (individual thickness between 9 and 15 µm), each consisting of numerous (possibly up to eight) individual fibers (Figures 1B, C and 2B, C, E, F). These fiber rings run around the oral aperture, have a consistent thickness and do not seem to be interrupted (Figures 1A and 2A, D—figure supplements 1 and 2A, B). Comparable fibers occur all over the body, but seem to be sparser towards the aboral pole (figure supplements 1B, C and 3A-F) and not organized in well-defined bundles (Figures 1D-G and 2A—figure supplements 1 and 3). Individual fibers are cylindrical (diameter ca. 2 µm), lying mostly parallel to each other although oblique

V-shaped interconnections (angle ca. 20-30°) may occur locally (Figure 2C). Fibers are finely and evenly coated with microcrystalline (ca 0.4 µm) calcium phosphate (Figure 1B, C, E). Circular fibers clearly extend into the triangular perradial apertural lobes (figure supplement 1A, E, F). The circular fiber network seems to be overprinted by faint longitudinal depressions in the aboral half (figure supplement 1D) that may correspond to interradial furrows.

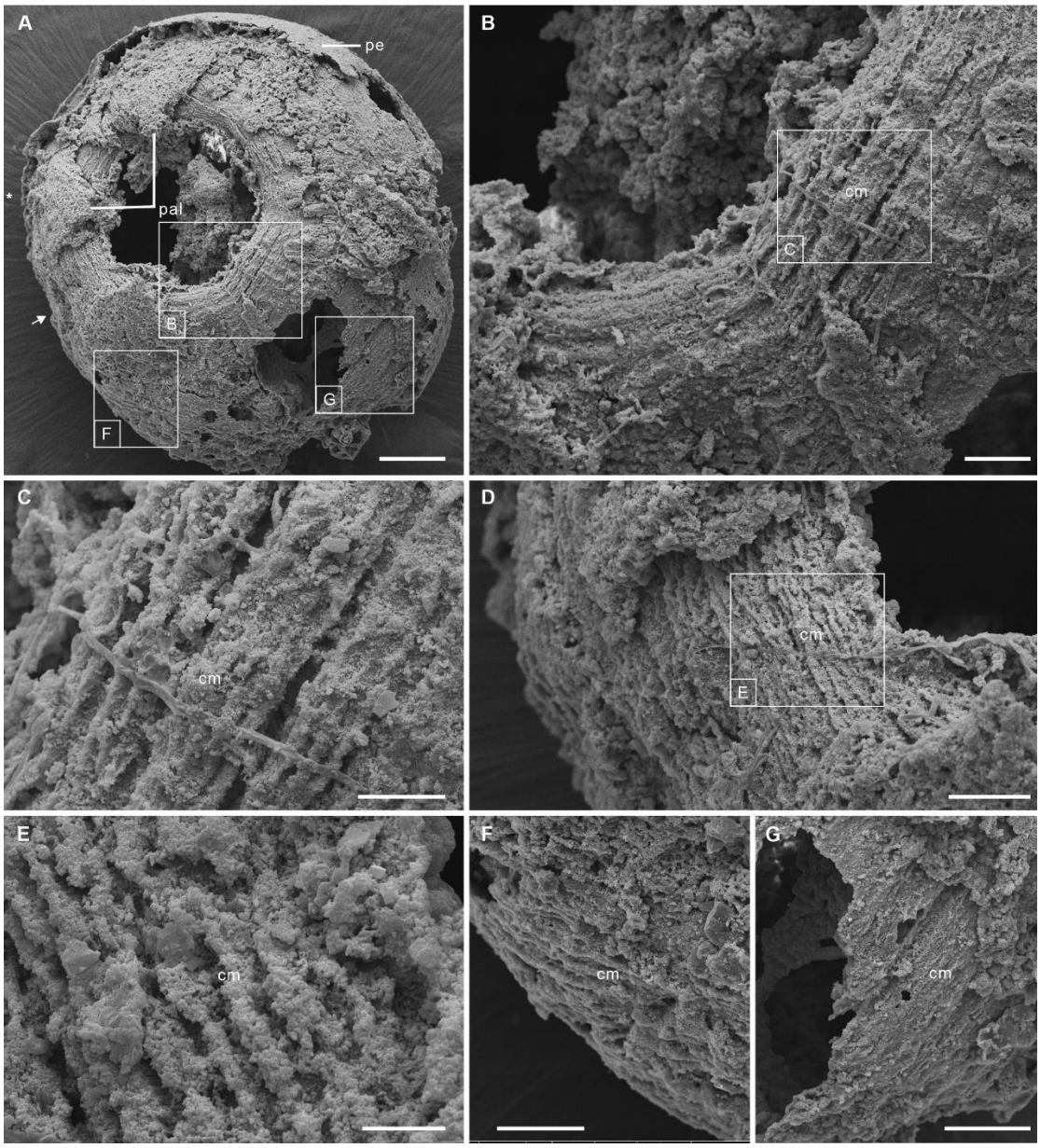

Figure 1. Post-embryonic stage of *Olivooides* sp. from the early Cambrian Kuanchuanpu Formation (ca. 535 Ma; South China), showing exposed muscle fibers. ELISN150-278. (**A**) General view of oral side. (**B**) Details of fiber bundles around aperture (see close-up location in **A**). (**C**) Close-up showing individual fibers within each bundle. (**D**) Dense network of fibers (see location in **A**). (**E**) Close-up of individual fibers coated with fine grains of calcium phosphate. (**F**), (**G**) Circular fibers approximately half way between the oral and aboral poles. All Scanning electron micrographs. Abbreviations: cm, circular muscle; pal, perradial apertural lobe; pe, periderm; *, perradii; →, interradii. Scale bars: 100 µm in (**A**); 20 µm in (**B**); 10 µm in (**C**), (**E**); 20 µm in (**D**), (**F**) and (**G**).

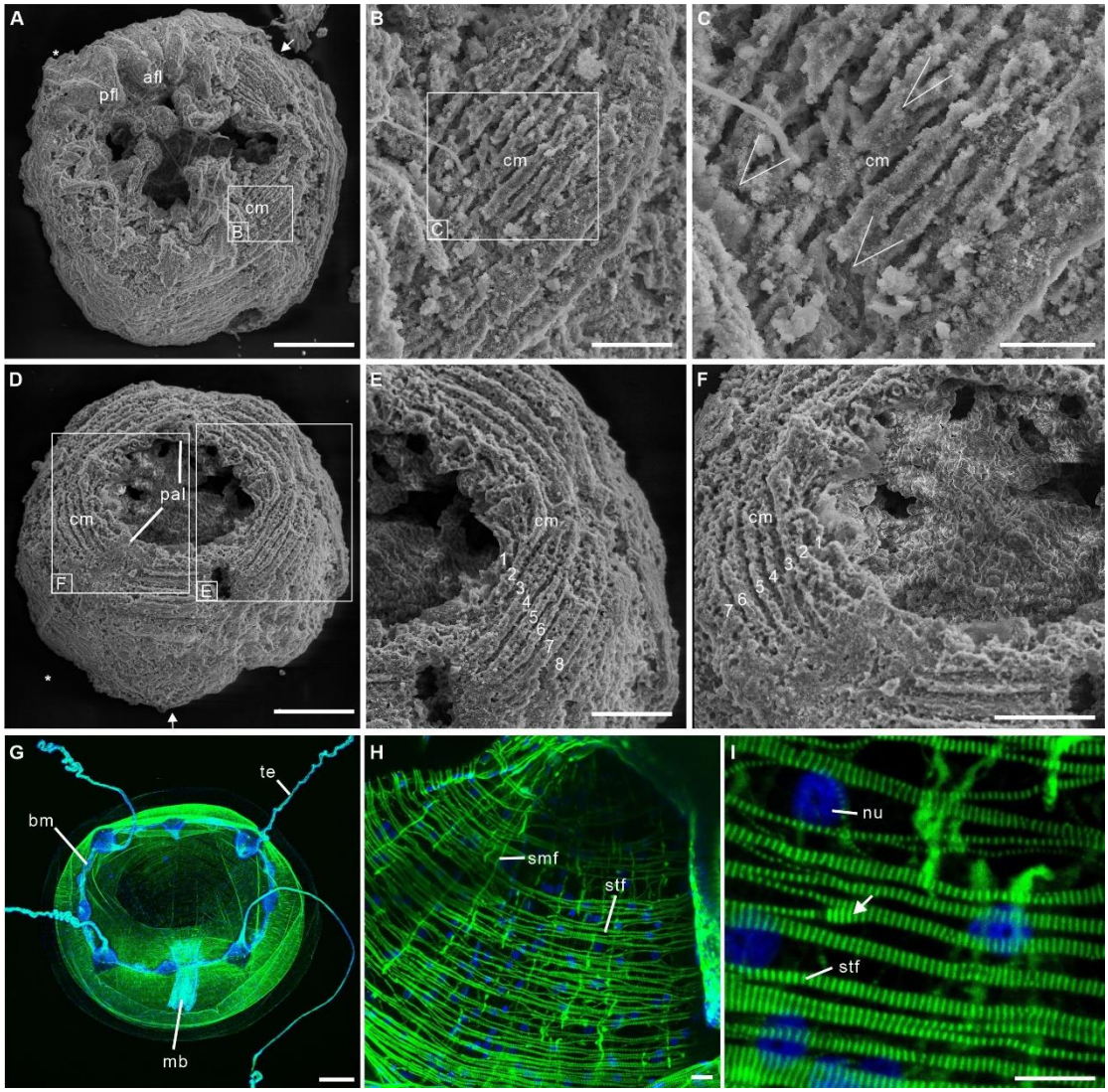

Figure 2. Post-embryonic stage of *Olivooides* sp. from the early Cambrian Kuanchuanpu Formation (ca. 535 Ma; South China), showing exposed muscle fibers (A-F) and myoepithelial muscle network in extant hydrozoan jellyfish (G-I). (**A-C**) ELISN052-33. General view of oral side and details of apertural circular muscle fibers and the V-shaped interconnection between the fibers in (**C**). (**D-F**) ELISN061-19. General view of oral side and details of apertural, circular muscles fibers. (**G-I**) *Eirene sp.* (Hydrozoa) young medusa, general oral view, circular muscles along subumbrella and details of striated fibers; white arrow (**I**) indicates bifurcating fibers. Green and blue colours in (**G-I**) correspond to actin (phalloidin) and DNA (Hoechst) staining. (**A-F**) Scanning electron micrographs. Abbreviations: afl, adradial fold lappet; bm, bell margin; cm, circular muscle; mb, manubrium; nu, nucleus; pal, perradial apertural lobe; pfl, perradial fold lappet; smf, smooth (radial) muscle fiber; stf, striated (circular) muscle fiber; te, tentacle *, perradii; →, interradii. Scale bars: 200 µm in (**A**), (**D**); 100 µm in (**G**); 20 µm in (**B**), (**E**) and (**F**); 10 µm in (**C**), (**H**) and (**I**).

Discussion

Myoepithelial muscles in cnidarians from the basal Cambrian

The close-knit circular fibers found in the bell of Olivooidae sp. can be confidently interpreted as epithelial muscle fibers based on their individual cylindrical shape, size range (ca. 2 µm in diameter), regular arrangement in bundles (e.g. possibly five more around bell margin), and closely-packed distribution over a single anatomical surface represented by the subumbrella (Figures 1 and 2—figure supplement 4). Our interpretation is also strongly supported by close similarities with the muscular system of modern cnidarians (myoepithelial cells; MEC). For example, the medusae of hydrozoans (Figure 2G-I) display a continuous network of circular striated and radial smooth muscles over the inner layer of their bell (subumbrella). These ca. 1-to-4-µm-thick individual fibers show oblique interconnections (Figure 2G-I—figure supplement 4), and sparser radial smooth fibers run perpendicular to them (Leclère and Röttinger, 2017). A very similar configuration can be seen in early Cambrian olivooids that display both continuous circular fibers (transverse markings imprinted on circular bundles; see Figures 1, 2 and 3A-C). Microscopic series of functional units (sarcomeres) that characterize striated muscles and give them a typical striated appearance (Figure 2G-I—figure supplement 4) are not discernible in the muscle network of Cambrian olivooids, making it impossible to distinguish their original nature, either striated or smooth (Schmidt-Rhaesa, 2007). The muscles of olivooids seem to be more developed and concentrated around the bell margin, as seen in modern jellyfish, such as *Clytia*, *Pelagia* and *Chrysaora* (Figure 2G-I—figure supplement 4). Other olivooids from the Kuanchuanpu Formation such as *Sinaster* have a comparable concentration of strong muscles around the oral aperture (possibly five bundles; see Wang et al., 2017) but, unlike those of the present specimens, seem to be interrupted by interradial structures. The muscle fibers of olivooids distribute over a surface interpreted as the inner layer of the bell (subumbrella), as in modern medusae. In contrast with bilaterians, modern cnidarians are characterized by myoepithelial cells that are fully integrated into the ectodermal and endodermal epithelial tissues. Although the cellular organization cannot be seen in Cambrian olivooids we hypothesize that their muscular system was similarly composed of myoepithelial cells that emitted myofilaments from their basal part. The circular network of olivooids seems to have been supplemented by possibly longitudinal muscles accommodated within adradial furrows (figure supplement 2B). However, no clear individual fiber can be seen in these adradial areas. Paired features interpreted as tentacular buds occur around the oral rim of some olivooids (Han et al., 2013; Wang et al., 2020). Their external annulations may represent underlying muscle fibers (Wang et al., 2020), or, more likely, anchoring features of nematocysts. The tentacles of modern cnidarians have longitudinal muscles but lack circular fibers (Hyman, 1940).

The current well-accepted hypothesis is that olivooids developed from an ovoid post-embryonic form (present material) into a conical corrugated polyp-like structure (Bengtson ang Yue, 1997; Dong et al., 2016; Han et al., 2013; Liu et al., 2014; Steiner et al., 2014; Wang et al., 2020). The transition to polyps is characterized by the gradual increase of external ornamented ridges that most probably resulted in a complete anatomical reorganization, as commonly seen in the development cycle of modern cnidarians. Unfortunately, very little is known of the internal anatomy of these polyps, except that they secreted a tubular feature (periderm) comparable with that of some extant scyphozoans (figure supplement 6) and had possible oral lobes. Although rare, clear traces of circular fibers do occur in the polyps of *Olivooides mirabilis* (see ref. Steiner et al., 2014; fig. 12), suggesting that key features of the muscle system were conserved through the lifecycle.

In summary, the close-knit fibrous network described here in post-embryonic olivooids is the oldest record of a muscular system in cnidarians and more generally in animals.

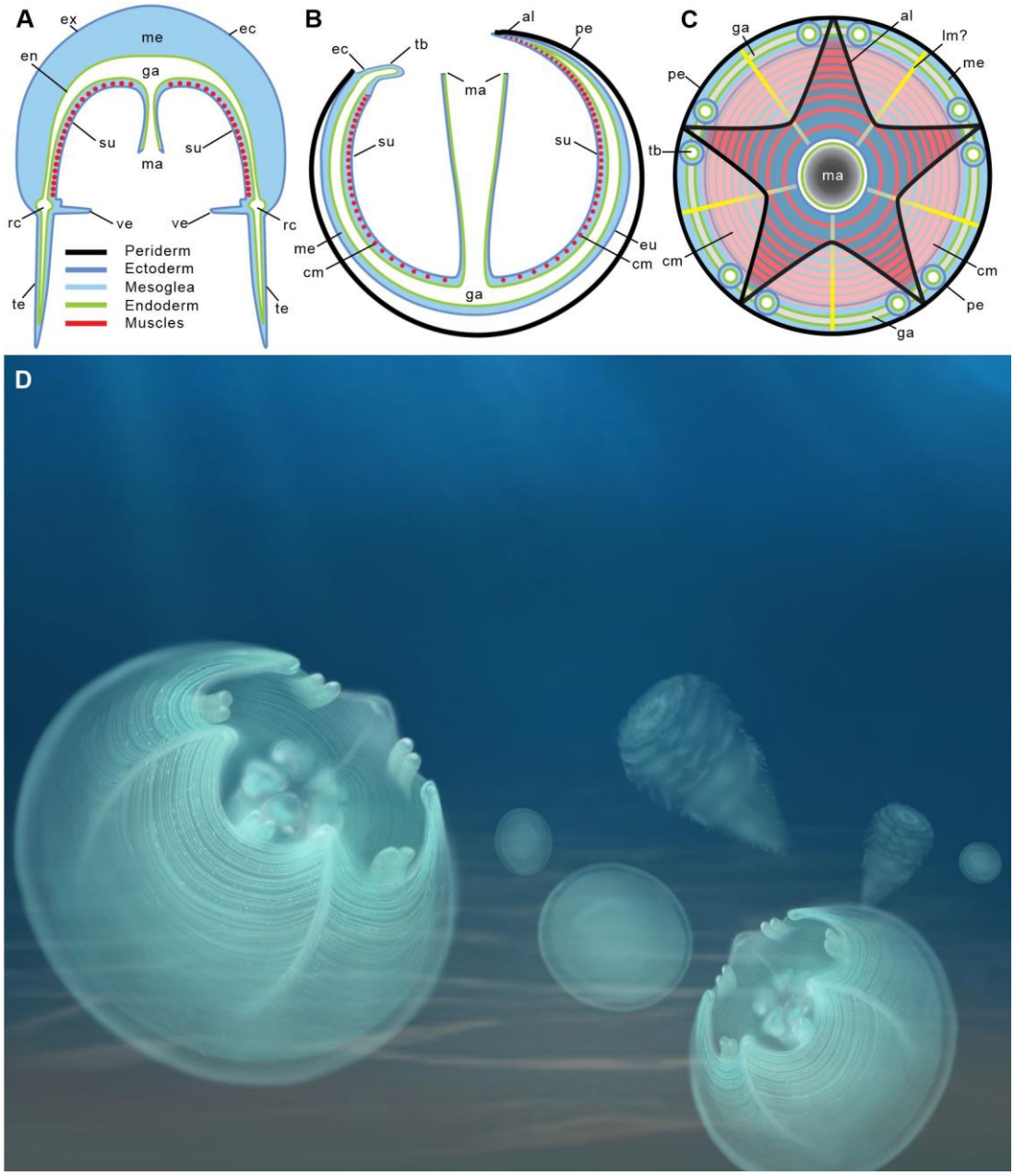

Figure 3. Location of epithelial muscles in extant hydromedusae (A) and early Cambrian Olivooidae medusozoans (B, C). (**A**), (**B**) are simplified radial sections through body. (**C**) is seen in oral view. (**D**) Artistic reconstruction of 535-million-year-old olivooid cnidarians showing eggs (no opening, background), post-embryonic (foreground) and polyp (background) stages. The circular muscle system is visible through the translucent periderm.Abbreviations: al, apertural lobe; cm, circular (coronal) muscle; ec, ectoderm; en, endoderm; ga, gastrovascular cavity; lm?, possible longitudinal muscle; ma, manubrium; me, mesoglea; pe, periderm; rc, radial canal; su, subumbrella; tb, tentacular bud; te, tentacle; ve, velum. Not to scale.

Functions of muscles in post-embryonic olivooids

The occurrence of strong muscles around the bell aperture and inside the perradial apertural lappets suggests that olivooids could contract their bell as do modern medusae (Figures 3A-C and 4A-C). In extant jellyfish, these contractions are counteracted by the elastic properties and antagonistic force of the adjacent mesoglea. As a result, water is rhythmically expelled from beneath the bell and drives the medusa through the water column via jet propulsion (Brusca et al., 2016). Although olivooids share important external and internal morphological features with medusozoans (Dong et al., 2013; Han et al., 2013; 2016b; Wang et al., 2017; 2020), they are distinguished by an unusual pentaradial symmetry and life cycle (ovoid post-embryonic stage to sessile polyp crest-bearing conical polyp). This life cycle has no direct counterpart in modern medusozoans (e.g. ref. Brusca et al., 2016) that develop from a motile planula larva into a polyp and eventually a juvenile medusa through various processes (e.g. strobilation generating ephyrae in scyphozoans; see ref. Gershwin, 1999). It has been suggested (Wang et al., 2020) that the post-embryonic stage of olivooids combined the characters of a medusa with those of a polyp and thus would resemble a juvenile sessile jellyfish almost encased within a periderm, with its bell aperture facing upwards. At first sight, these circular muscles may suggest a role in locomotion, as in modern medusae. However major structural differences separate modern jellyfish from Cambrian olivooids (see above). Whereas the movement of modern medusae is unconstrained, that of olivooids was most probably strongly hindered by its periderm (Bengtson and Yue, 1997; Steiner et al., 2014). The assumed mesoglea layer of olivooids seems to have been very thin (narrow gap between ex- and subumbrella; see Han et al., 2013, 2016b; Dong et al., 2016; Wang et al., 2017, 2020) thus limiting its dynamic capacity. Swimming efficiency of modern medusae depends on the power and distribution of muscles but also largely on the flexibility of the bell margin, a condition that is not found in olivooids. Powerful muscle contractions may have been able to propel the animal over a very short distance but are unlikely to have sustained dynamic locomotion through the water column. This hypothetical “clumsy” locomotion would have probably created more disadvantages (e.g. energy cost) than advantages to the animal. Moreover, it is unlikely to have generated any adequate escape reaction (e.g. from predators) or effective feeding technique. We favour an alternative option. One of the most vital requirements for post-embryonic olivooids was probably to obtain sufficient food. Modern medusae feed on small soft-bodied prey by using nematocyst-laden tentacles and oral arms that convey food to the mouth (Brusca et al., 2016). Post-embryonic olivooids had tentacular buds that were not enough developed to capture food. Feeding may have been achieved by a different method, such as the rhythmic contractions of their circular muscles, especially those bordering the bell aperture (Figure 4A-C). Such contractions may have resulted in pumping and engulfing sea water containing potential food particles. Extant jellyfish such as *Aurelia* feed by using a comparable mechanism (Costello and Colin, 1994). The aperture lappets of olivooids may have played the key roles in closing the aperture after contraction in locking away food particles within the bell cavity before being transferred to the mouth and preventing foreign matters from entering the bell. Besides, the remarkable plasticity of modern cnidarians to transformation and rearrange their muscle system (e.g. transition from polyp to medusa stages; Brusca et al., 2016) may have played in this evolution and diversification. Interestingly, muscle fibers primarily assigned to feeding (olivooids) may have been used for other functions such as active propulsion, escape reaction during evolution while maintaining an important role in prey capture. In

this context, we could make a bold speculation that the origin of the medusae swimming, associated by the subsequent loss of periderm, is an evolutionary inheritance of rhythmic feeding and respiration among benthic periderm-dwelling medusozoans, probably during Ediacarian-Cambrian period (Figure 3D).

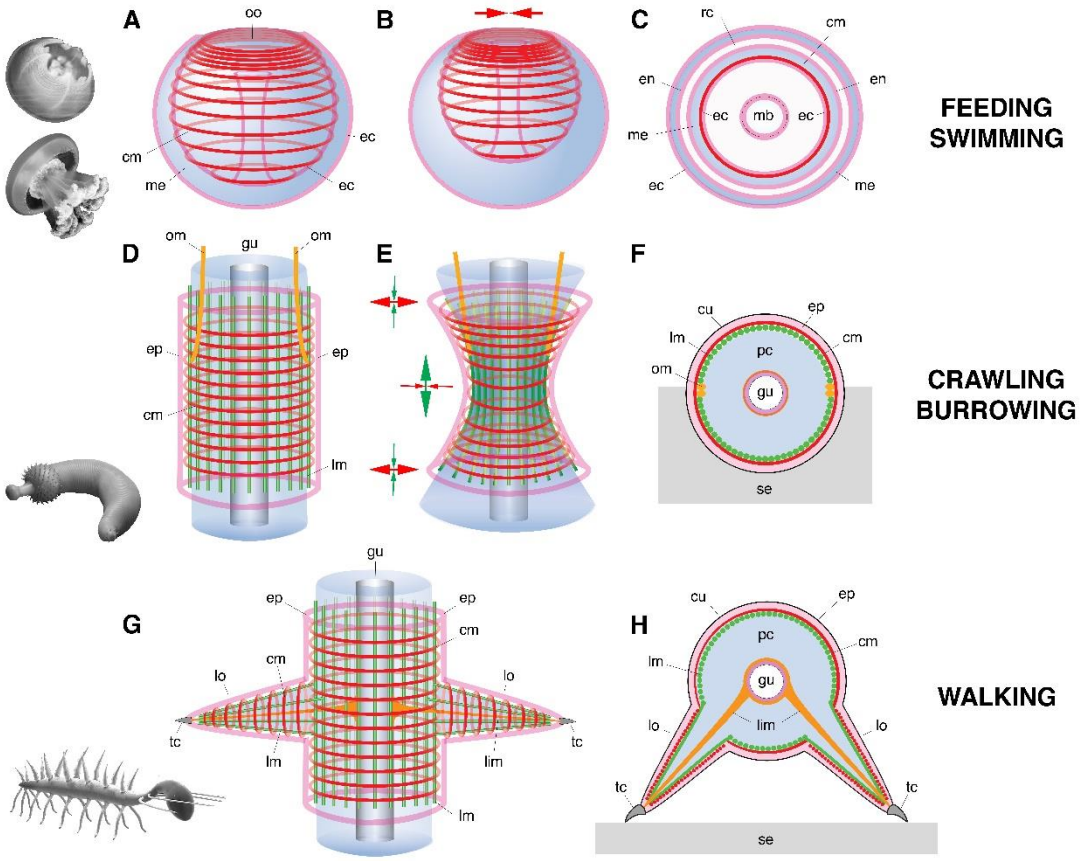

Figure 4. Basic muscle systems in early Cambrian animals and their main functions. (**A-C**) Contractile epithelial muscles (MEC, myoepithelial, predominantly circular) and antagonistic mesoglea exemplified by olivooiids and free-swimming jellyfish; idealized relaxed (**A**) and contracted (**B**) states and simplified transverse section (**C**). (**D-F**) Grid-like network of circular and longitudinal subepidermal muscle fibers (HMS) around cylindrical body filled with antagonistic internal fluid (hydroskeleton) exemplified by scalidophoran worms; idealized relaxed state (**D**), peristaltic contractions along body (**E**) and transverse section (**F**). (**G-H**) Longitudinal and circular muscles and extrinsic retractor muscles in limbs; idealized relaxed state (**G**); transverse section (**H**). Images (from top to bottom) represent an olivooid cnidarian, an extant jellyfish, the scalidophoran worm *Ottoia* (see ref. Vannier, 2012) and the lobopodian *Hallucigenia* (see refs. Smith and Caron, 2015; Smith and Ortega-Hernández, 2014). Drawings and images not to scale. Abbreviations: cm, circular muscle; cu, cuticle; ec, ectoderm; en, endoderm; ep, epidermal layer; gu, gut; lc, lobopod claw; lim, limb muscle; lm, longitudinal muscle; lo,

lobopod (soft leg); mb, manubrium; me, mesoglea; om, oblique muscle; oo, oral opening (bell margin); pc, primary cavity filled with fluid; rc, radial canal; se, sediment; tc, terminal claw.

Early evolution of muscle systems

The shift of the actin-myosin system from strictly intracellular to intercellular functions seems to have occurred in the early stages of metazoan evolution (Schmidt-Rhaesa, 2007). Although the most basal metazoans lack true muscles they do have the capacity to contract their body or part of it. For example, sponges have myocytes around the osculum that play a role in expelling wastes (Bagby, 1966). The fiber cells of placozoans such as *Trichoplax adherens* have contractile extensions packed with actin filaments that link the ventral and dorsal epithelia (Armon et al., 2018). Although the gliding motility of placozoans is mainly performed by ciliated epithelial cells, fiber cells seem to be involved in active body deformation and invagination possibly related to feeding (Smith and Mayorova, 2019). The integration of the actin-myosin system within a dense network of myoepithelial cells as seen in Cambrian and modern cnidarian is likely to have been a major innovation that provided animals with new capacities for powerful movements (e.g. swimming, feeding, respiration, etc.). We suggest that the myoepithelial fiber network seen in post-embryonic olivooids may represent one of the oldest and basic types of animal muscle systems that probably finds its origin in the late Precambrian. Larger jellyfish from the early (e.g. Chengjiang, Han et al., 2016a) and mid-Cambrian (Marjum Lagerstätte; coronal muscles, Cartwright et al., 2007) are likely to have co-opted this basic system for more diverse functions such as swimming within the water column (e.g. active jet propulsion) and prey capture as seen in modern jellyfish such as *Aurelia*.

Diversity of muscle systems and locomotion modes in early Cambrian animals

Ecdysozoans (worms and panarthropods) provide additional evidence on the diversity of muscle systems in early Cambrian animals.

Scalidophoran worms were diverse and numerically abundant in the Cambrian (e.g. Burgess-shale-type Lagerstätten). Modern representatives of the group such as *Priapulus* have a close-knit network of circular and longitudinal muscle linning the inner surface of the body wall, that is surrounded by ECM (Figure 4D-F—figure supplement 7) and often termed “Hautmuskelschlauch” (HMS; see Schmidt-Rhaesa, 2007). The enclosed mass of incompressible fluid (hydroskeleton) that fills the primary cavity of such worms is the principal antagonist for muscular action (Figure 4). Dynamic interactions between HMS and hydroskeleton allow them to perform repeated body contraction and extension for burrowing (Figure 4D-F) (e.g. ref. Vannier et al., 2010) and feeding. The eversion of the introvert is a process of pumping body fluids into the anterior body region whereas its invagination is performed by extrinsic oblique retractor muscles. Based on remarkable anatomical and functional similarities with modern priapulids (Figure 4D-F—figure supplement 7), we posit that Cambrian scalidophorans also had a HMS-type musculature and possible retractors. This hypothesis is supported by abundant crawling and burrowing traces found in the basal Cambrian (e.g. treptichnid burrow systems; see refs. Kesidis et al., 2019; Vannier et al., 2010) and the late Precambrian (e.g. refs. Evans et al., 2020; Gehling et al., 2001) that could not have been made without the action of HMS on

a hydrostatic skeleton (Figure 4D-F—figure supplement 7). HMS clearly differs from the musculature of olivooid cnidarians in at least two key features: 1) it does not consist of myoepithelial cells and 2) the antagonist onto which muscular force is transferred is not the primary cavity filled with fluid but the mesoglea (Leclère and Röttinger, 2017).

Lobopodians (e.g. Ortega-Hernandez, 2015) is an informal group of ecdysozoans with an annulated cuticle and paired soft legs (lobopods), that is crucial for understanding the remote ancestry of euarthropods. They are best exemplified by iconic Cambrian forms such as *Hallucigenia* (Smith and Caron, 2015; Smith and Ortega-Hernández, 2014) and *Microdictyon* (Pan et al. 2018). Most recent phylogenetic trees (Aria et al., 2021; Giribet and Edgecombe, 2019) have resolved Cambrian lobopodians as an "intermediate" group between scalidophoran worms and arthropods with an arthrodized exoskeleton. The inner surface of their body wall was lined with closely packed circular and longitudinal muscle fibers that seem to have extended into the limbs (see Tritonychus in Zhang et al., 2016; Figure 4G, H—figure supplement 5A, B). This configuration strongly recalls that of ecdysozoan worms (see above). In *Paucipodia* from the Chengjiang Lagerstätte (see Vannier and Martin 2017; see supplement figure 5C-E) a connecting strand runs between the terminal claw of the limbs and the area surrounding the gut, and is interpreted here as a possible retractor muscle (see analogues in extant onychophorans (De Sena Oliveira and Mayer, 2013). Although unsegmented as in smaller lobopodians, larger Cambrian lobopodians such as *Pamdelurion* from the Sirius Passet Lagerstätte (ca. 520 Ma) are characterized by a more complex musculature with paired, lateral, ventral and dorsal longitudinal muscles. Well-developed bundles of extrinsic and intrinsic limb muscles presumably controlled leg motion as in modern onychophorans (Budd, 1998; Hoyle and Williams, 1980; Young and Vinther, 2017; see Figure 4G, H).

Early euarthropods that co-existed with lobopodians and scalidophoran worms (e.g. Burgess-shale-type Lagerstätten) had already acquired rigid exoskeletal elements (sclerotized cuticular elements jointed by an arthrodial membrane, such as body sclerites and appendage podomeres) that were operated by a lever-like system of segmentally arranged antagonistic muscles as seen in *Kiisortoqia* and *Camparamuta* from the Sirius Passet Lagerstätte (Young and Vinther, 2017). This suggests that the rise of euarthropods was associated with a profound rearrangement of the muscle system inherited from lobopodian ancestors, such as the reduction of the circular HMS musculature that lost its primary hydrostatic function and peristaltic capabilities because of exoskeletal rigidity (Young and Vinther, 2017).

In summary, both fossil and indirect evidence presented here indicate that different types of musculature co-existed among early Cambrian animals: (1) myoepithelial circular (MEC) muscles in cnidarians, (2) grid-like and subepidermal (HMS) muscles in scalidophoran worms, (3) HMS and extrinsic muscles to control leg motion in lobopodians and (4) segmentally arranged muscles tightly integrated to exoskeletal elements in early euarthropods.

Whereas the myoepithelial system appears as the most basic one, that of ecdysozoans seems to have undergone considerable changes and diversification over a relatively short time interval during the early Cambrian. This remarkable diversity and plasticity of muscle systems allowed a great variety of animals to explore and colonize new environments and can be seen as one of the driving forces of the animal radiation.

Geological setting, materials and methods

All studied fossils come from phosphatic limestones collected from the Kuanchuanpu Formation at the Shizhonggou section, (Ningqiang County, Shaanxi Province, China; see figure supplement 8). Biostratigraphy (*Anabarites – Protohertzina – Arthrochities* zone, the *Siphogonuchites – Paragloborilus* zone and *Lapworthella – Tannuolina – Sinosachites* zone; refs. Qian, 1977; 1999) indicates that these rocks correspond to the Meishucunian Stage that is the equivalent of the lowermost Cambrian Terreneuvian Stage. Radiochronology (U-Pb method; refs. Sawaki et al., 2008; Peng et al., 2012) confirms that the Kuanchuanpu Formation is approximately 535 Ma. Secondarily phosphatized fossils were extracted from rocks via a standard acid digestion in 7% acetic acid. Dried residues with a grain-size >60 µm were sorted and picked under a binocular microscope. Twelve specimens of Olivooidae (Cnidaria) bearing well-preserved muscle fibers were selected for the present study and mounted for SEM (FEI Quanta 400 FEG scanning electron microscope at Northwest University, China; Au-coating, high-vacuum). They belong to *Sinaster petalon* Wang et al. (2017) (ELISN115-39), *Hanagyroia orientalis* Wang et al. (2020) (ELISN107-470) and Olivooidae sp. (ELISN150-278, ELISN111-54, ELISN052-33, ELISN045-143, ELISN012-16, ELISN061-19, ELISN087-64, ELISN088-48, ELISN087-33 and ELISN098-19). All specimens are deposited in the collections of the Shaanxi Key Laboratory of Early Life & Environments and the Department of Geology, Northwest University, China. Data concerning these specimens are available on request from JH and XW.

Two-week-old *Clytia hemisphaerica* medusae, newly released *Eirene sp.* medusae, one-month-old *Chrysaora colorata* and *Pelagia noctiluca* metaephyrae were raised in the laboratory (Villeranche-sur-mer) following Lechable et al. (2020) and Ramondenc et al. (2017) culture protocols. Fixation followed by Phalloidin (actin) and Hoechst (nuclei) staining were performed on the four species as described for *Clytia hemisphaerica* in Sinigaglia et al. (2020). Samples were mounted in 50% citifluor AF1 antifadent mountant and imaged using Leica SP8 confocal and Zeiss Axio-Observer microscopes.

Extant priapulid worms (*Priapulus caudatus*) (see figure supplement 7) were collected (JV) from near the Kristineberg Marine Station (Sweden), fixed with glutaraldehyde and dried (Critical Point) for SEM observations (Univ. Lyon).

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and the Paris Aquarium for providing the Eirene polyp stains. We thank the Marine Resources Centre (CRBM and PIV imaging platform) of Institut de la Mer de Villefranche (IMEV), supported by EMBRC-France. The CRBM is supported by EMBRC-France, whose French state funds are managed by the ANR within the Investments of the Future program under reference ANR-10-INBS-02.

Author contributions

X. W., J. V. and J. H. designed the research. X. W., X.-G. Y and J. H. collected rocks (Shizhonggou section, Shaanxi Province, China) and analyzed fossils. X. W. wrote the manuscript with input from all other co-authors. L. L. provided comparative data and images of modern cnidarians and Q. O. made insightful remarks on the MS.

Supplements

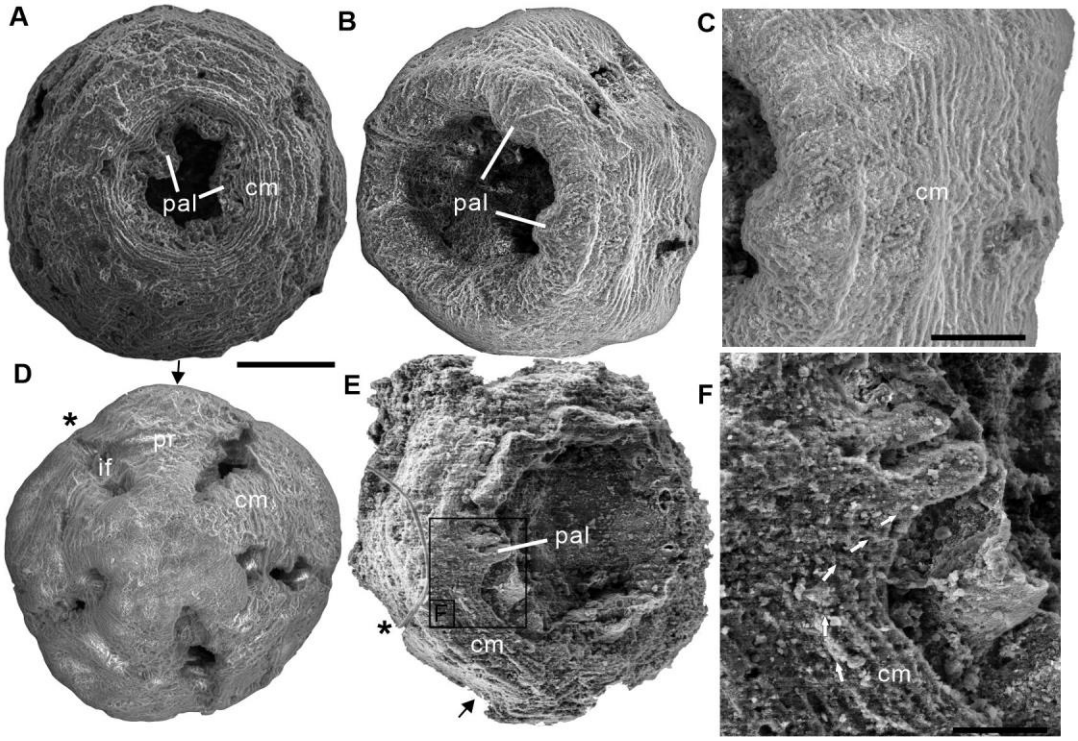

figure supplement 1. The three scanning electron micrographs of the *Olivooides* specimens ELISN012-16, ELISN045-143 and ELISN111-54 from the Kuanchuanpu Formation, south China. (**A**) Oral view showing the external morphology of ELISN045-143 without periderm. (**A**), (**D**) ELISN012-16. General view of oral side (**A**), details of muscles fibers in perradial apertural lobes and general view of aboral side (**D**). (**B**) Oral view showing the external morphology of ELISN045-143 without periderm. (**C**) Close-up view of a showing the circular muscles. (**E**) Oral view showing the external morphology of ELISN111-54. (**F**) The enlarged view of c showing that the circular muscles extending into the triangular perradial apertural lobes marked by the giant white arrows. Abbreviations: cm, circular muscle; pal, perradial apertural lobe; *, perradii; →, interradii. Scale bars represent: 200 μm in (**A**), (**B**), (**D**) and (**E**); 50 μm in (**C**); 25 μm in (**F**).

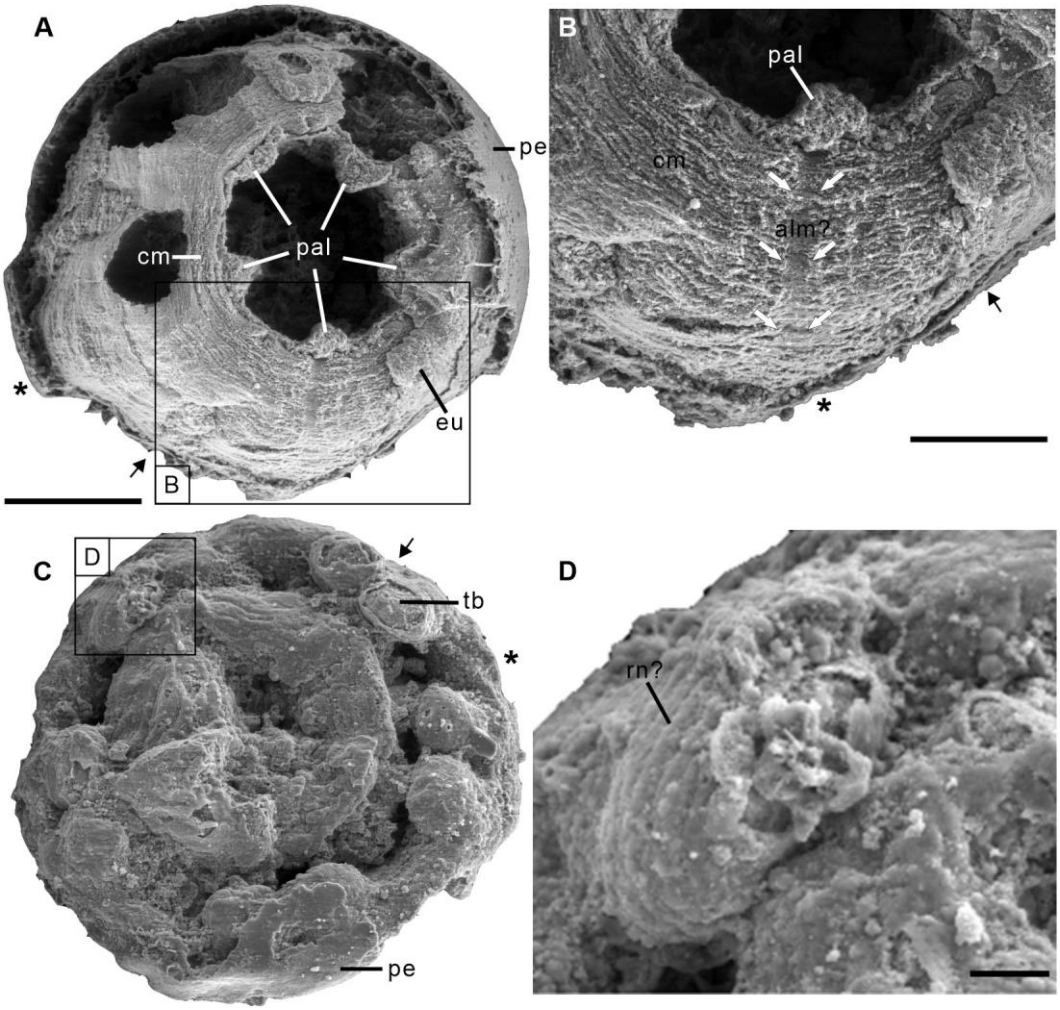

figure supplement 2. The two scanning electron micrographs of the *Olivooides* specimens ELISN115-39 (*Sinaster petalon*; Wang et al. 2017) and ELISN107-470 (*Hanagyroia orientalis*; Wang et al. 2020) from the Kuanchuanpu Formation, south China. (**A**) Oral view showing the external morphology of ELISN115-39. (**B**) Close-up view of (**A**) showing the possible longitudinal muscle bands marked by the two rows of giant white arrows. (**C**) Oral view showing the external morphology of ELISN107-470. (**D**) The enlarged view of (**C**) showing the possible rings of nematocysts on the tentacles. Abbreviations: alm?, the possible longitudinal muscle bands at adradii; cm, circular muscle; eu, exumbrella; pal, perradial apertural lobe; pe, periderm; *, perradii; →, interradii. Scale bars represent: 200 µm in (**A**), (**C**); 50 µm in (**B**); 25 µm in (**D**).

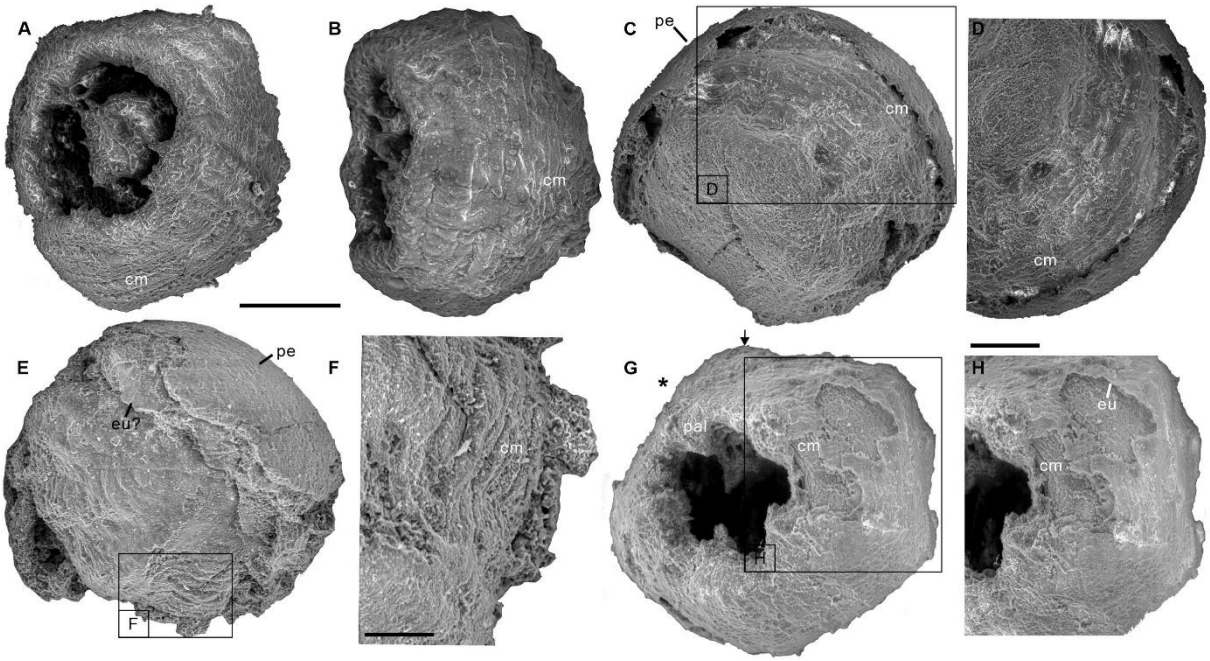

figure supplement 3. The four scanning electron micrographs of the *Olivooides* specimens ELISN087-64 (**A, B**), ELISN088-48 (**C, D**), ELISN087-33 (**E, F**) and ELISN98-19 (**G, H**) from the Kuanchuanpu Formation, south China. (**A**) General view of oral side. (**B**) Lateral view. (**C**) General view of aboral side. (**D**) The enlarged view of (**C**) showing the circular muscles. (**E**) Lateral view. (**F**) The enlarged view of (**E**) showing that the circular muscles. (**G**) General view of oral side. (**H**) The enlarged view of (**G**) showing the circular muscles. Abbreviations: cm, circular muscle; eu, exumbrella; pal, perradial apertural lobe; *, perradii; →, interradii. Scale bars represent: 200 μm in (**A-C**), (**E**) and (**G**); 50 μm in (**F**), (**H**); 25 μm in (**D**).

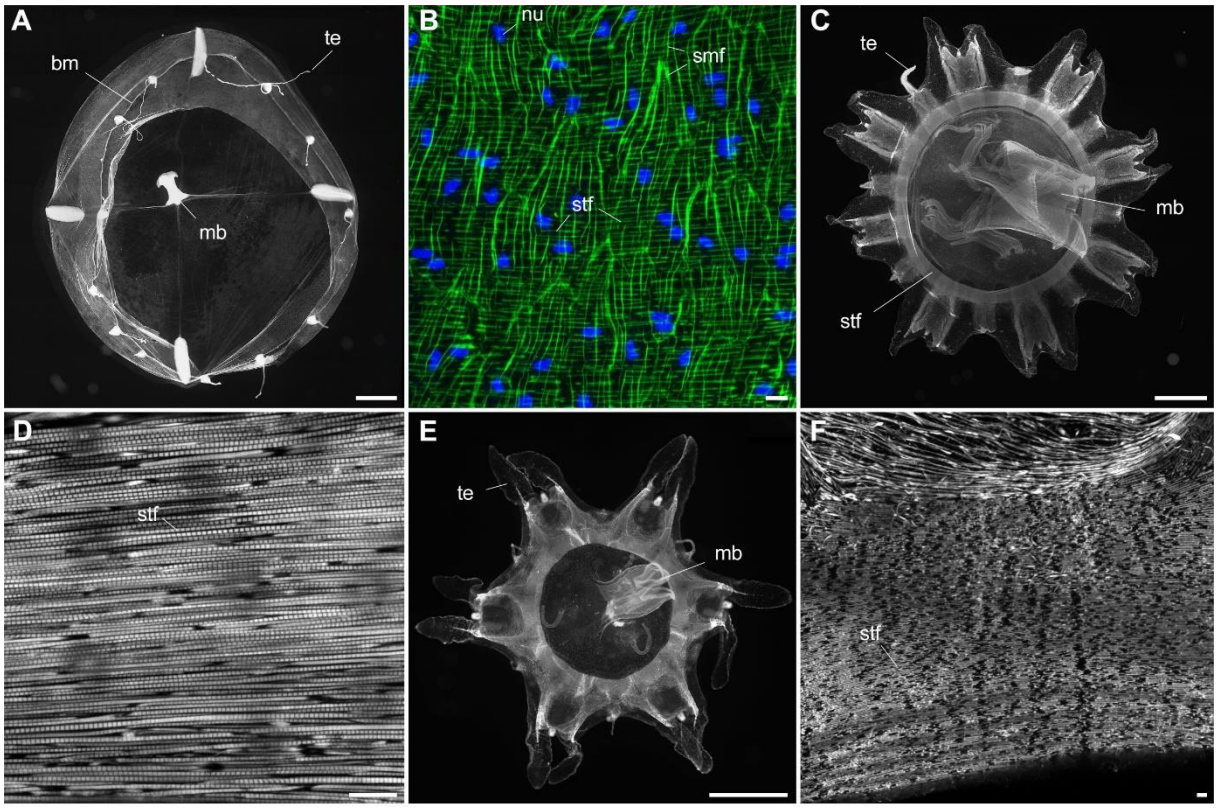

figure supplement 4. Myoepithelial muscle network shown in extant medusozoan cnidarians. (**A**), (**B**) *Clytia hemisphaerica* (Hydrozoa) mature medusa, general oral view and details of fiber network (subumbrella). (**C**), (**D**) *Pelagia noctiluca* (Scyphozoa) metaephyra, general oral view and details of closely-packed circular striated muscle fibers (subumbrella). (**E**), (**F**) *Chrysaora colorata* (Scyphozoa) metaephyra, general oral view and details of circular striated muscle fibers. Green and blue colours (**B**) correspond to actin (phalloidin) and DNA (Hoechst) staining. Specimens represented by black and white images (**A**) and (**C-F**) were stained for actin (phalloidin). Abbreviations are as follows: bm, bell margin; mb, manubrium; nu, nucleus; smf, smooth (radial) muscle fiber; stf, striated (circular) muscle fiber; te, tentacle. Scale bars: 1000 µm in (**A**), (**C**) and (**E**); 10 µm in (**B**), (**D**) and (**F**).

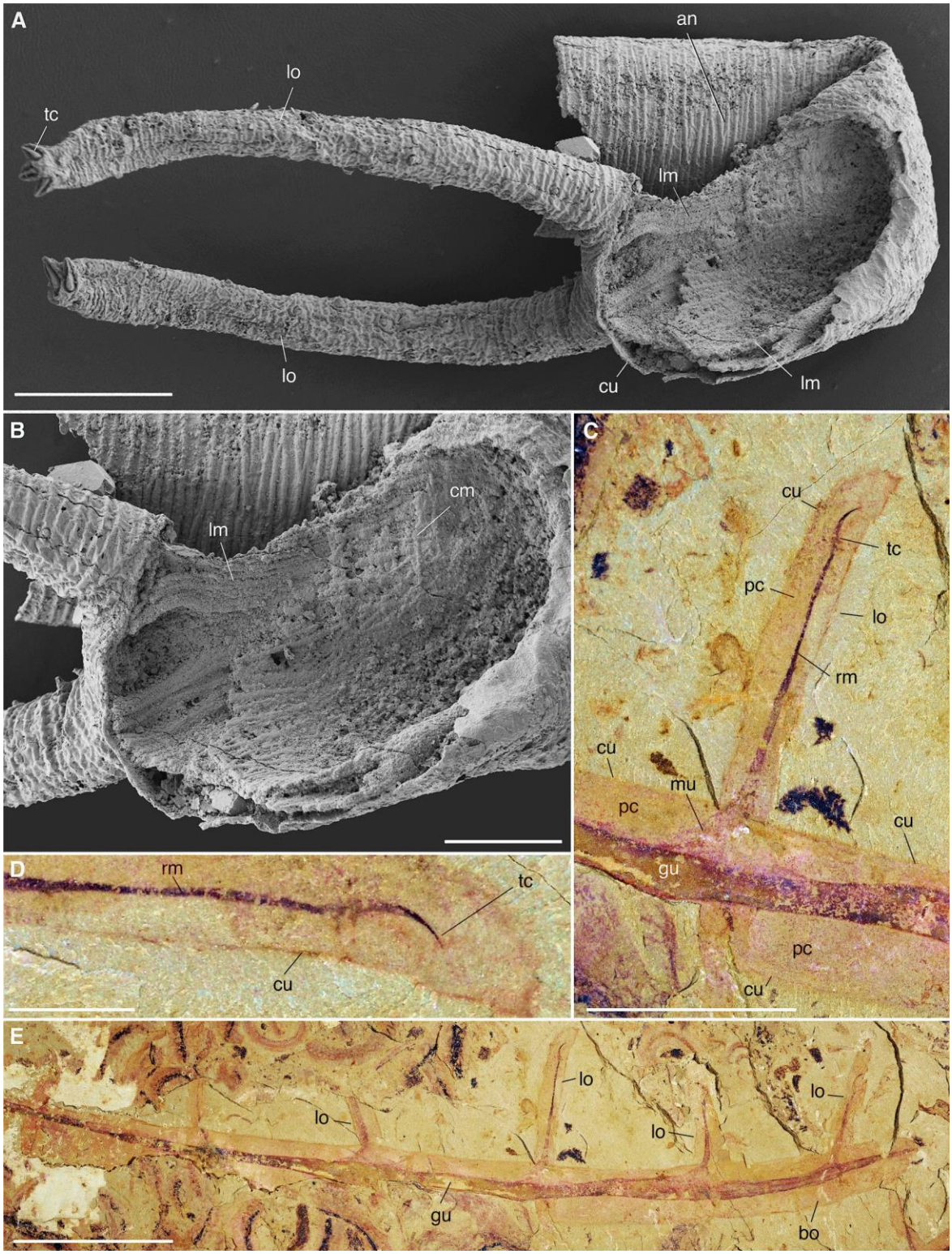

figure supplement 5. Muscle system in Cambrian lobopodians (Ecdysozoa, Panarthropoda). (**A**), (**B**) *Tritonychus phanerosarkus* from the Yu'anshan Formation (Xiaotan section, Cambrian Series 2, Stage 3), Yongshan, Yunnan Province, China; general view and details of a secondarily phosphatized specimen showing a pair of walking appendages (lobopods) and the inner body wall lined with muscle fibers (see Zhang et al., 2016). (**C-E**) *Paucipodia haikouensis* from the Chengjiang Lagerstätte, ELI-JS0001a (see Vannier and Martin, 2017); close-up of lobopod and terminal claw, and general view;

457 note muscle bridging claw to gut area. (**A**), (**B**) are SEM images (Courtesy Prof. Xiguang Zhang; see
458 Zhang et al., 2016). Abbreviations: an, annuli; cm, circular muscle; cu, cuticle; gu, gut; lm, longitudinal
459 muscle; lo, lobopod; mf, muscle fiber; mu, muscle; pc, primary cavity; rm, retractor muscle; tc,
460 terminal claw. Scale bars: 10 mm in (**E**), 5 mm in (**C**), 1 mm in (**D**), 200 µm in (**A**) and 100 µm in (**B**).
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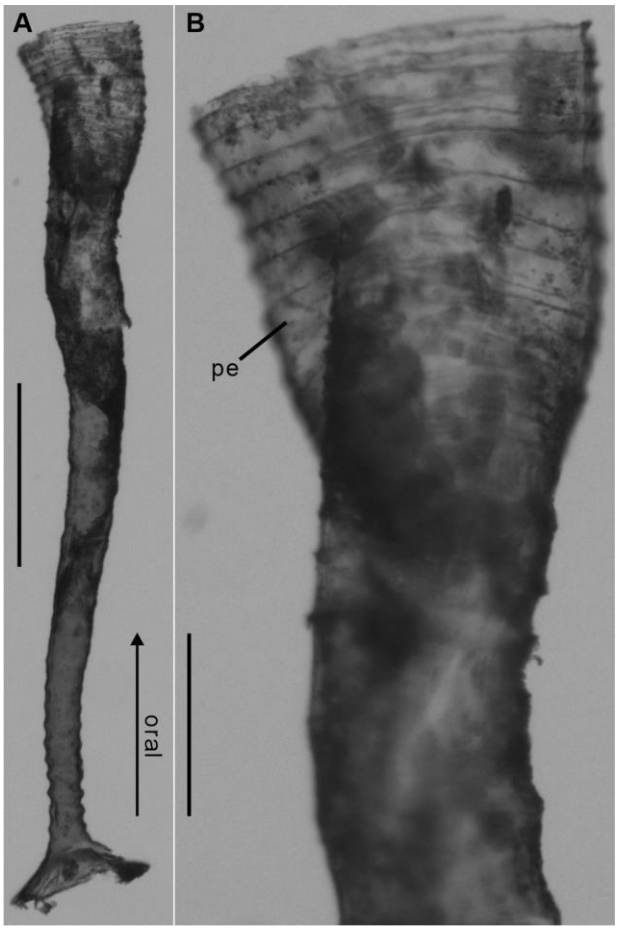

462

463 **figure supplement 6. Polyp of Coronatae sp. from South China Sea** showing the periderm. (**A**), (**B**)
464 Lateral view and details showing corrugations. (Courtesy Dr. Xikun Song). Abbreviations are as
465 follows: pe, periderm. Scale bars: 400 µm in (**A**); 100 µm in (**B**).
466

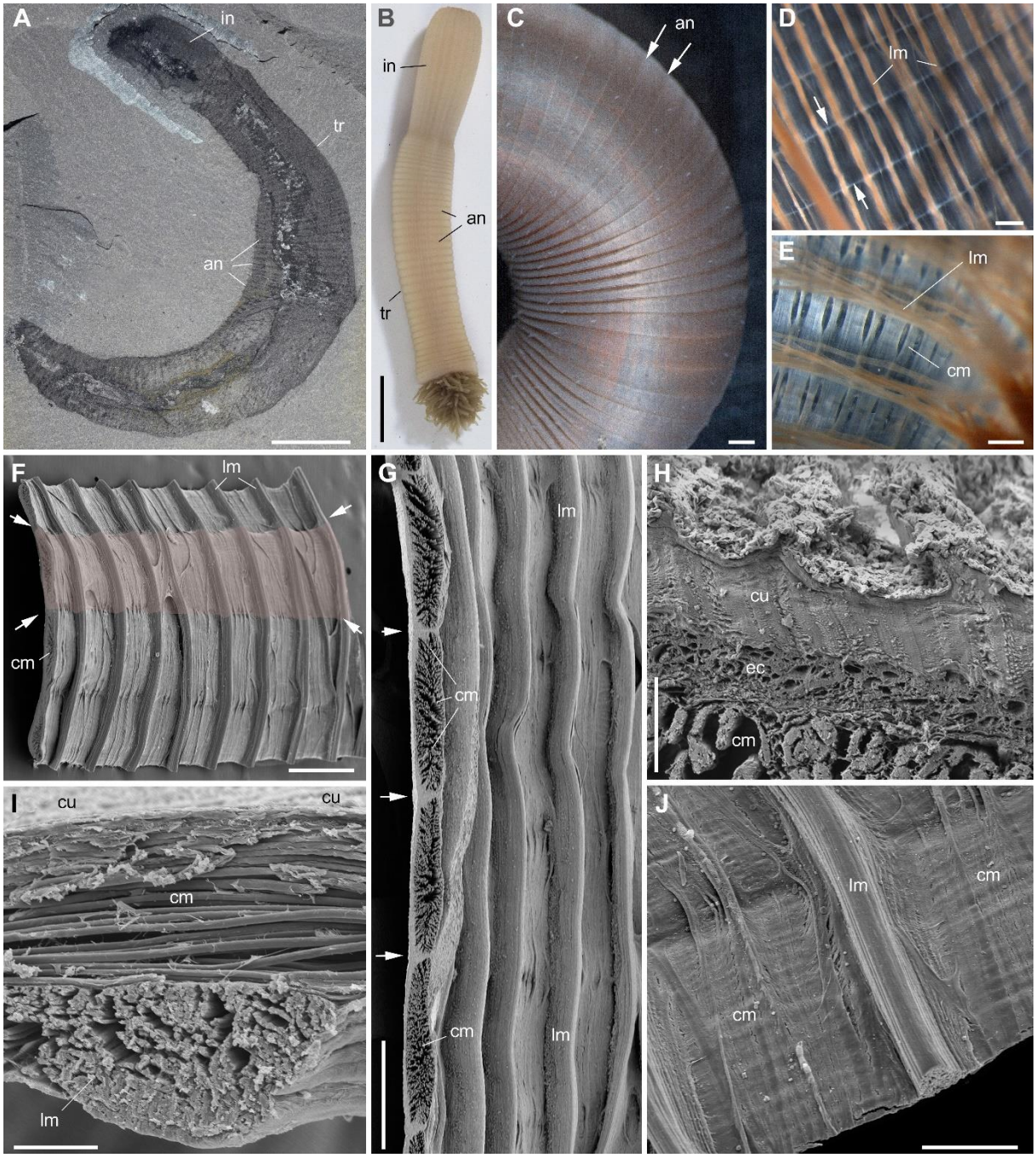

figure supplement 7. Muscle system in priapulid worms (Ecdysozoa). (**A**) *Ottoia prolifica* from the mid-Cambrian Burgess Shale, general view showing annulated body. (**B-J**) *Priapulus caudatus* from Sweden. (**B**) general view of live specimen in sea water. (**C**) annulated trunk. (**D**) muscles (trunk). (**E**) muscles (introvert). (**F**) fragment of trunk showing the internal body wall with strong longitudinal muscles (pink area corresponds to one annulation). (**G**) transverse section through circular muscles (one bundle per annulation). (**H**) transverse section through body wall across cuticle, epithelial cells and circular muscles. (**I**) transverse section through body wall showing circular muscles overlying longitudinal ones. (**J**) inner wall of body showing both circular and longitudinal muscles. Small white arrows indicate annulation boundaries. (**A-E**) are light photographs. (**F-J**) are SEM images. Abbreviations are as follows: an, annulation; cm, circular muscle; cu, cuticle; ec, epithelial cells;

introvert; lm, longitudinal muscle; tr, trunk. Scale bars: 1 cm in (**A**), (**B**); 1 mm in (**C**); 500 µm in (**D-G**); 200 µm in (**J**); 50 µm in (**I**) and 10 µm in (**H**).

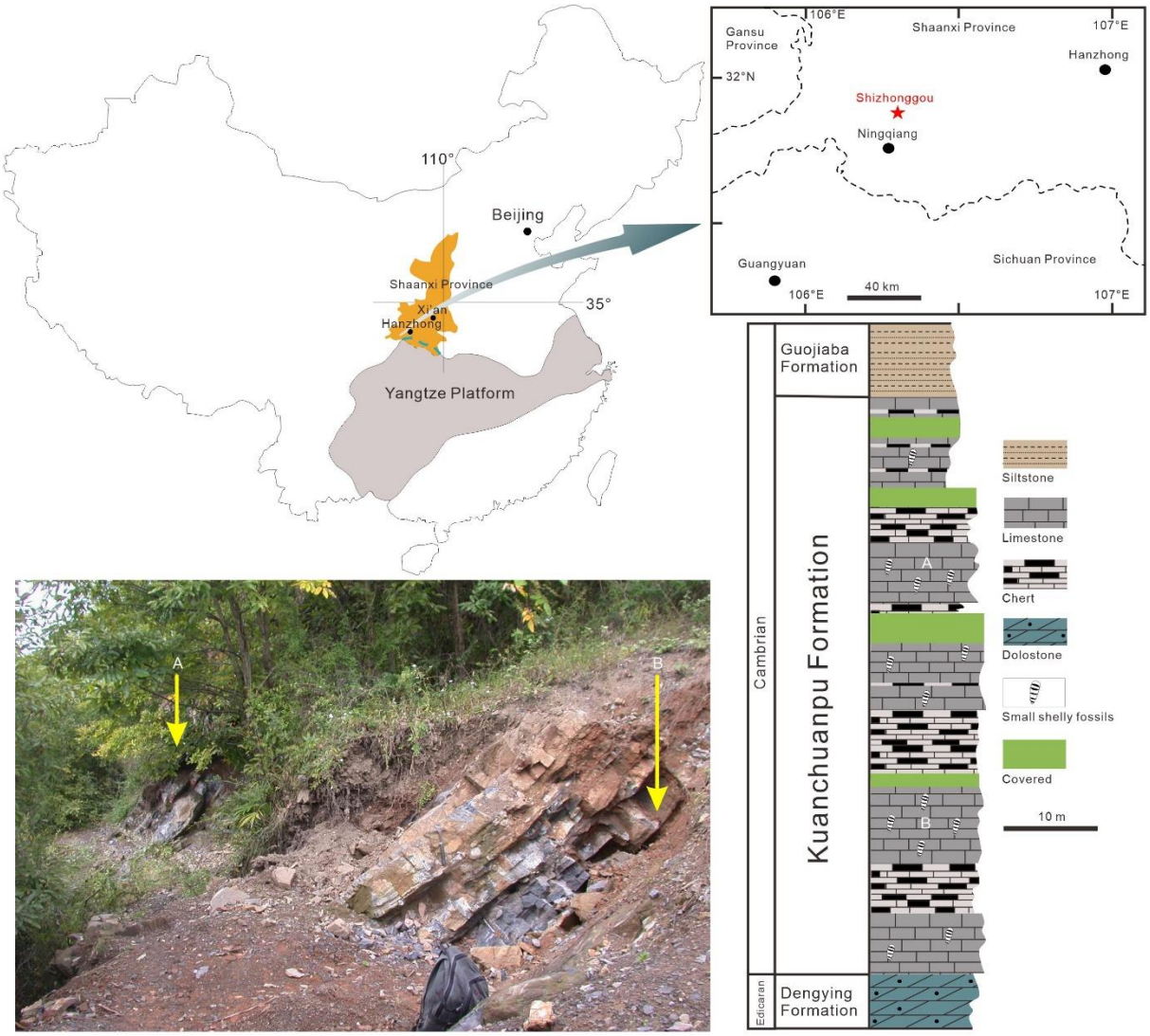

figure supplement 8. Origin of the fossil material. Location map (Ningqiang area, Southern Shaanxi Province, China), photograph of outcrop (Shizhonggou Section; see red star) and stratigraphic position of fossil-bearing rock samples (**A**), (**B**) within the Kuanchuanpu Formation. Modified from Steiner et al., 2014.

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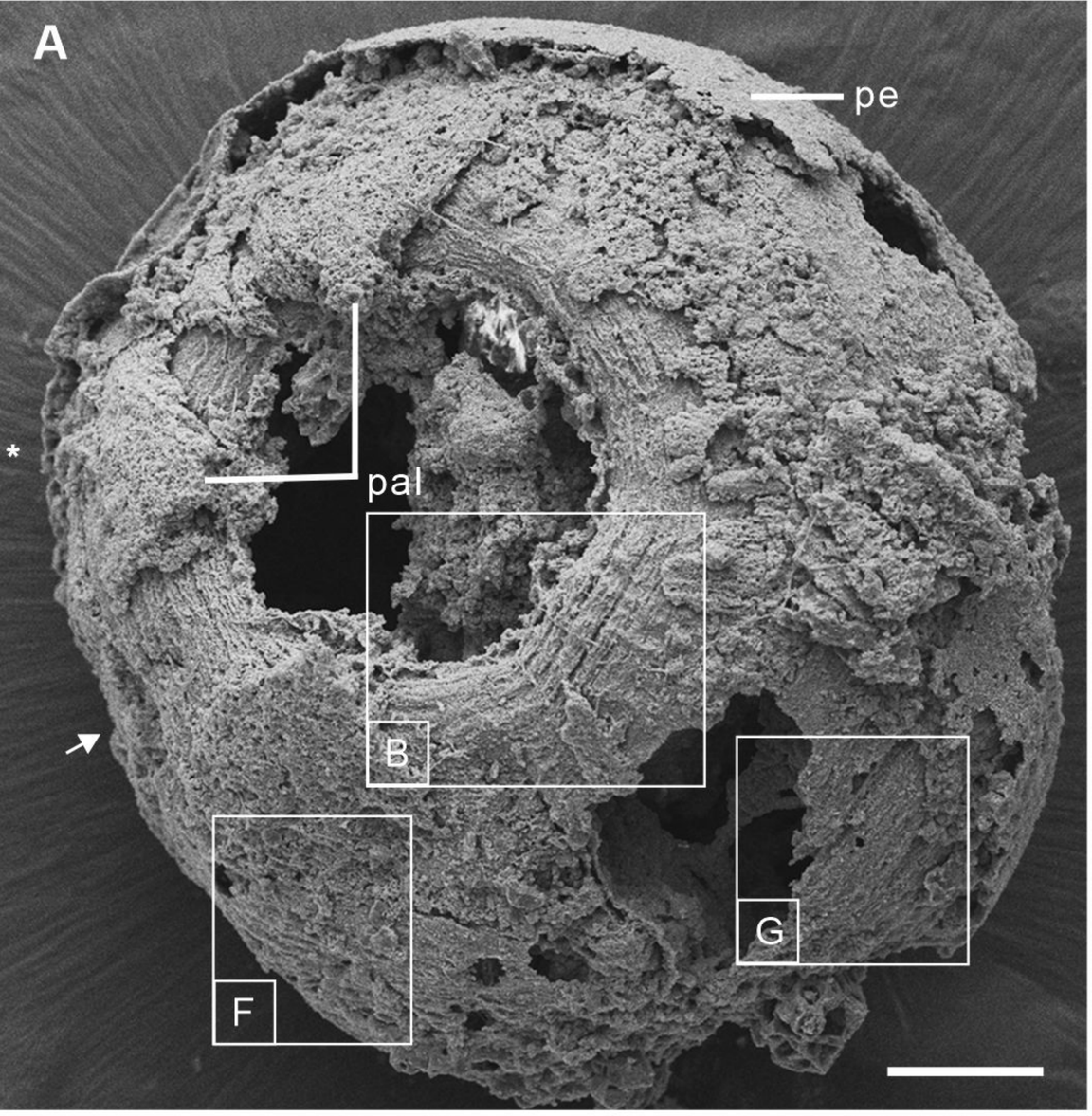

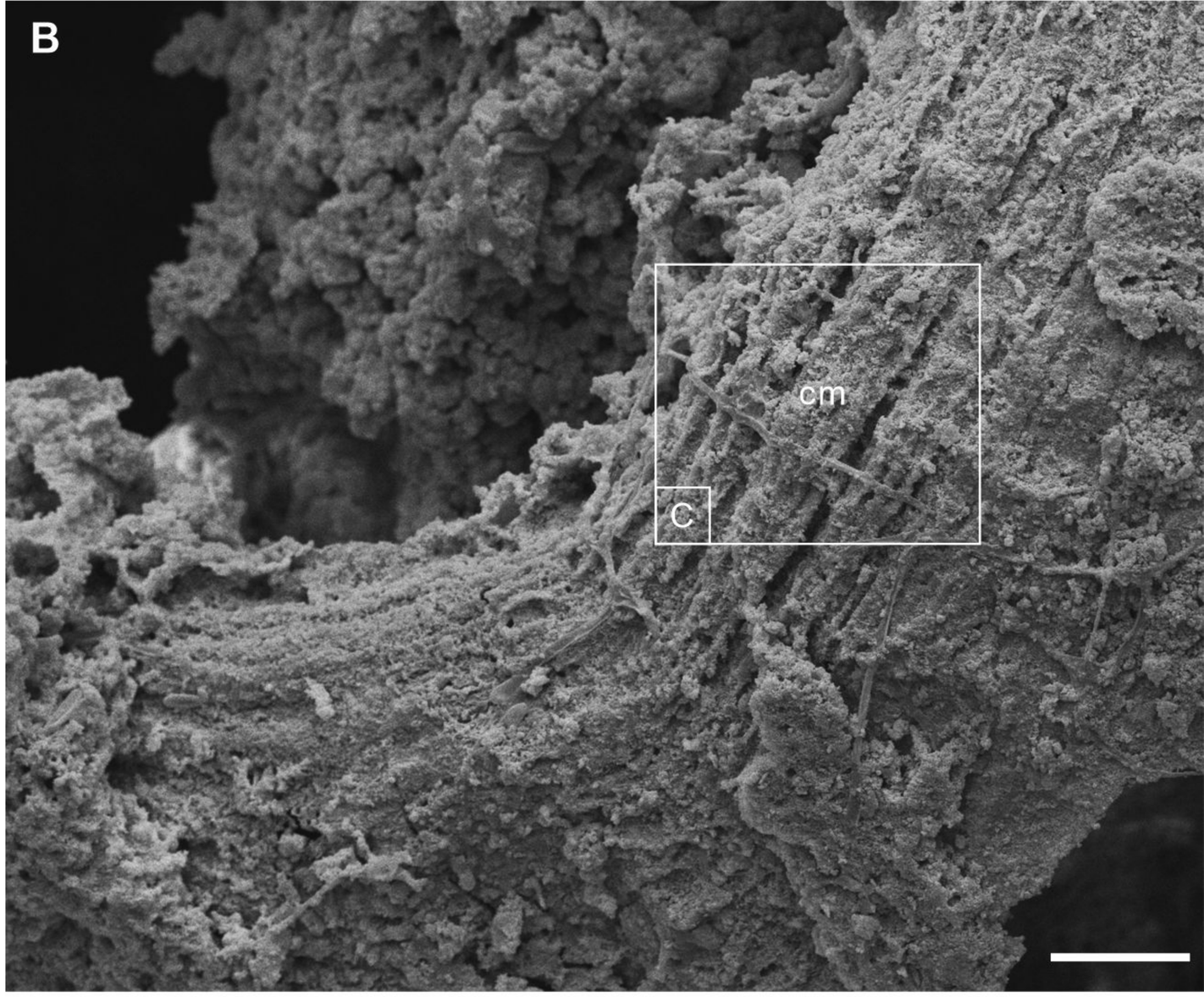

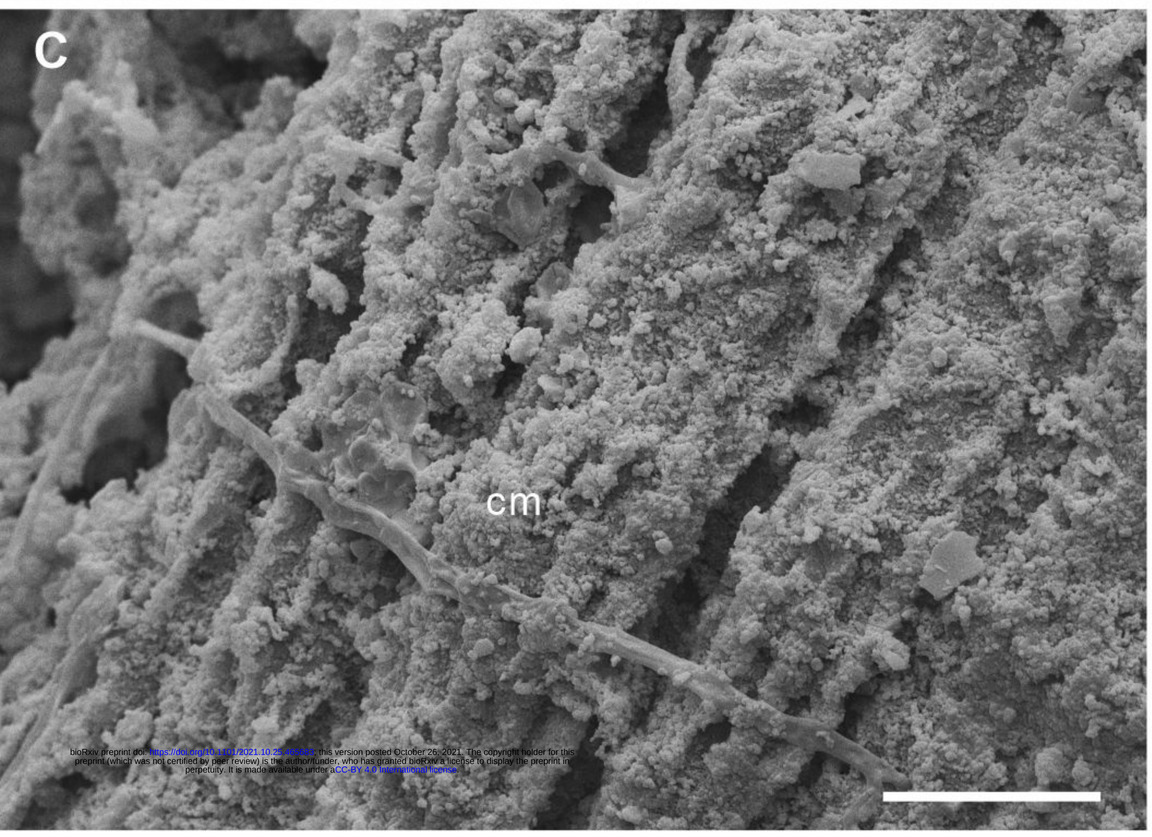

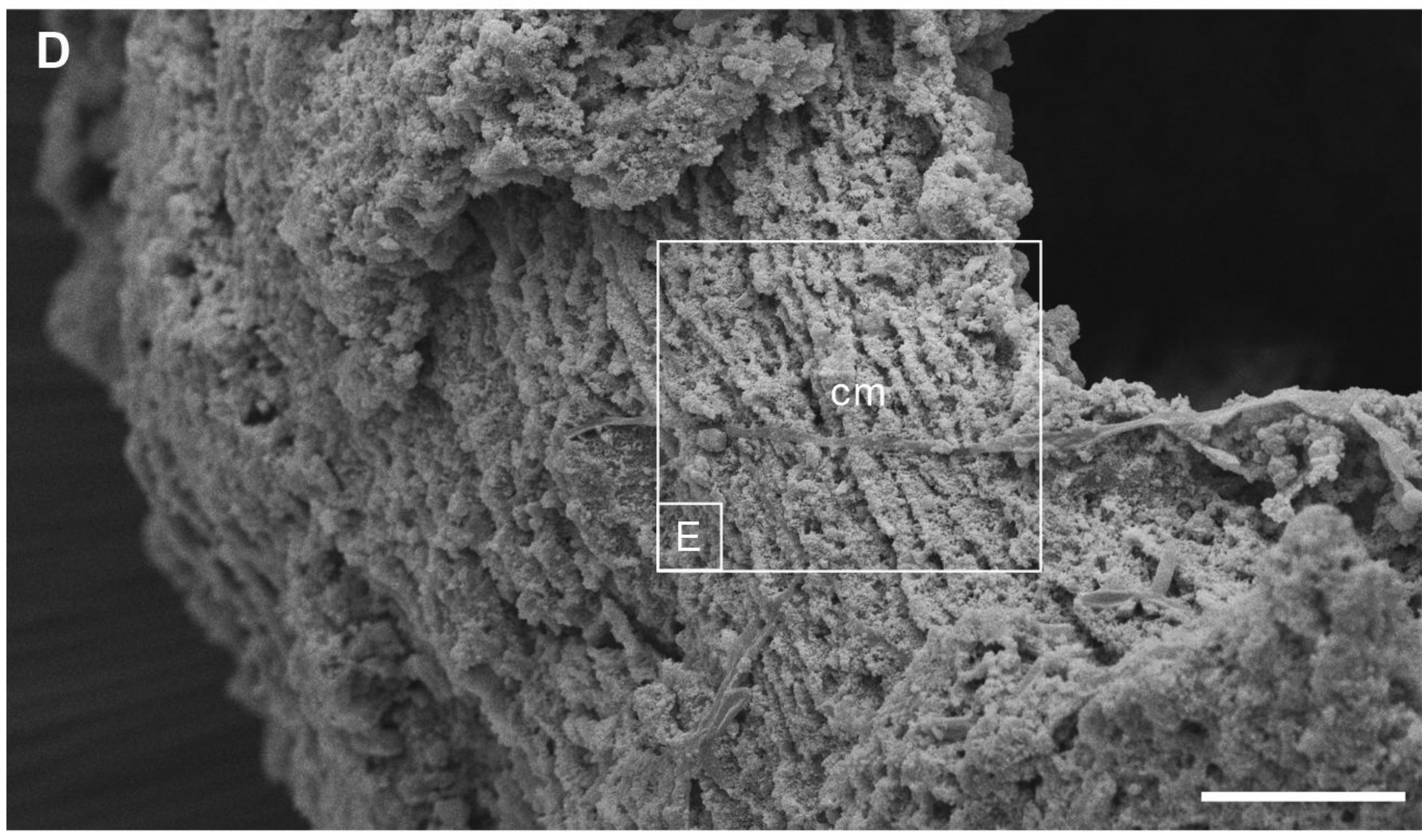

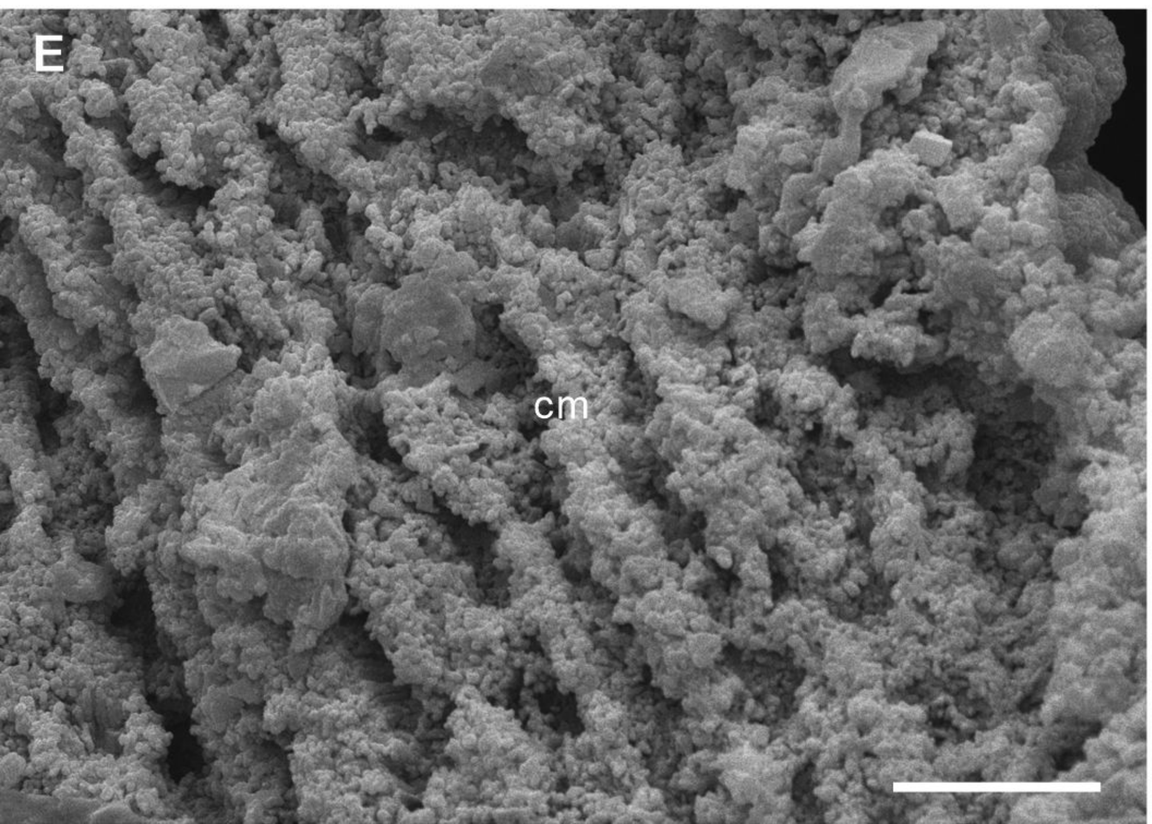

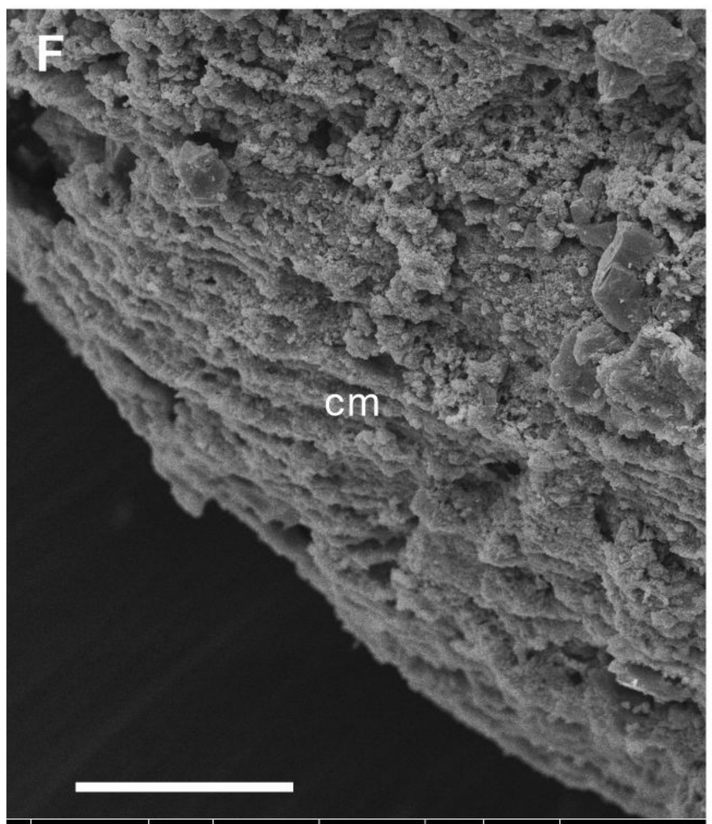

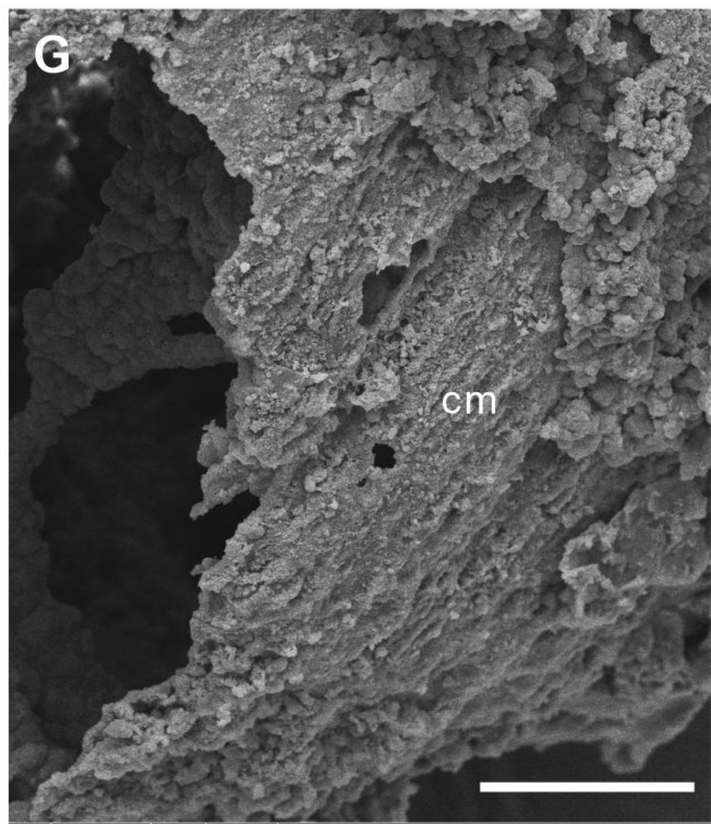

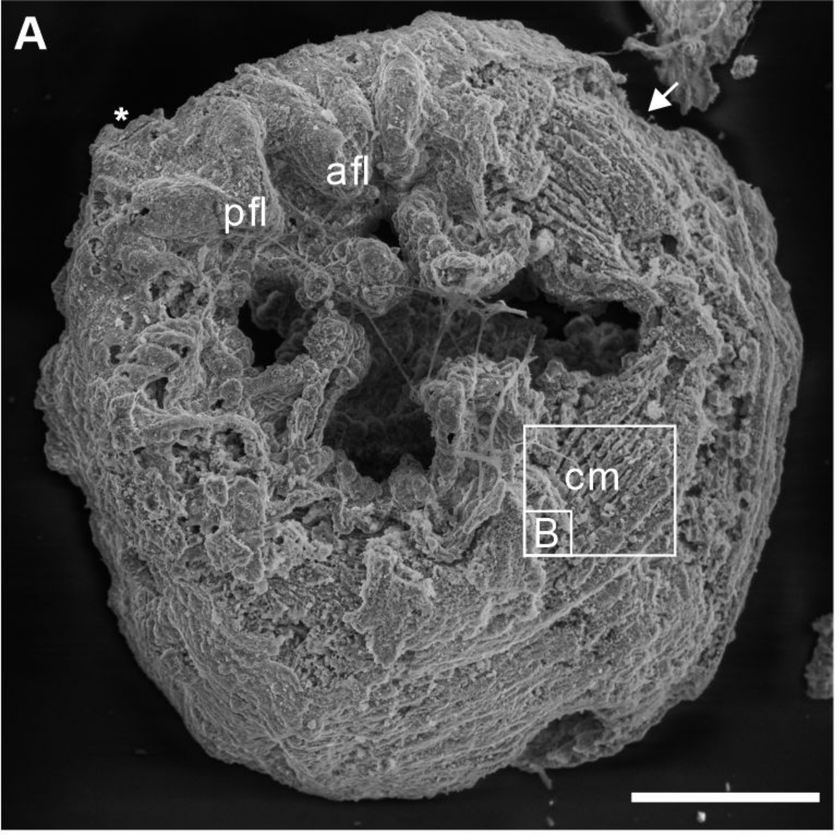
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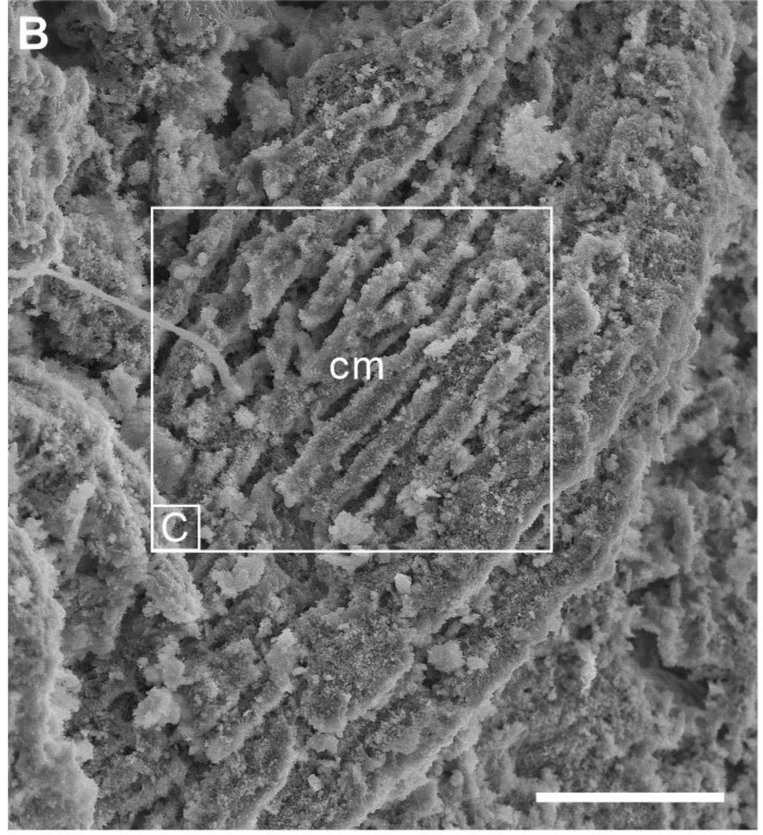
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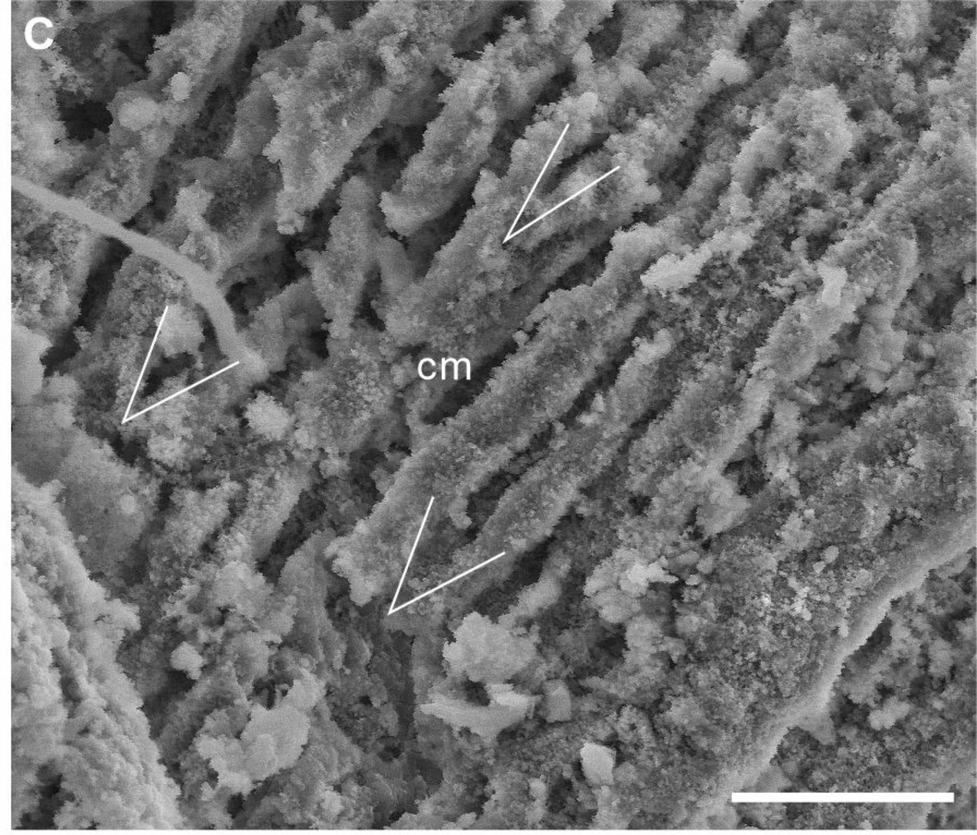
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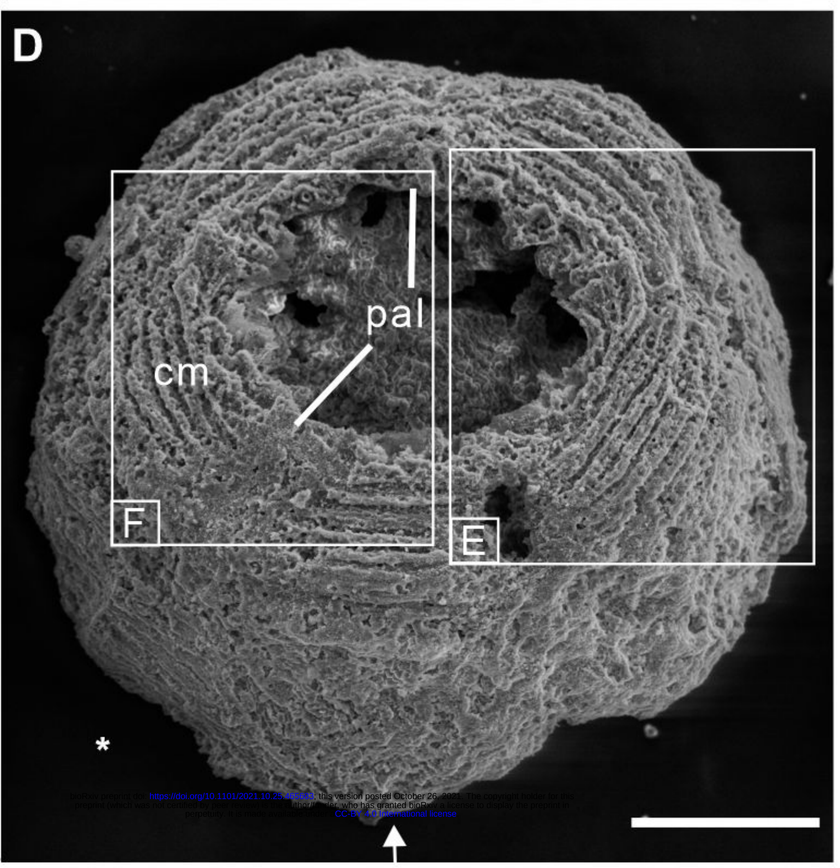
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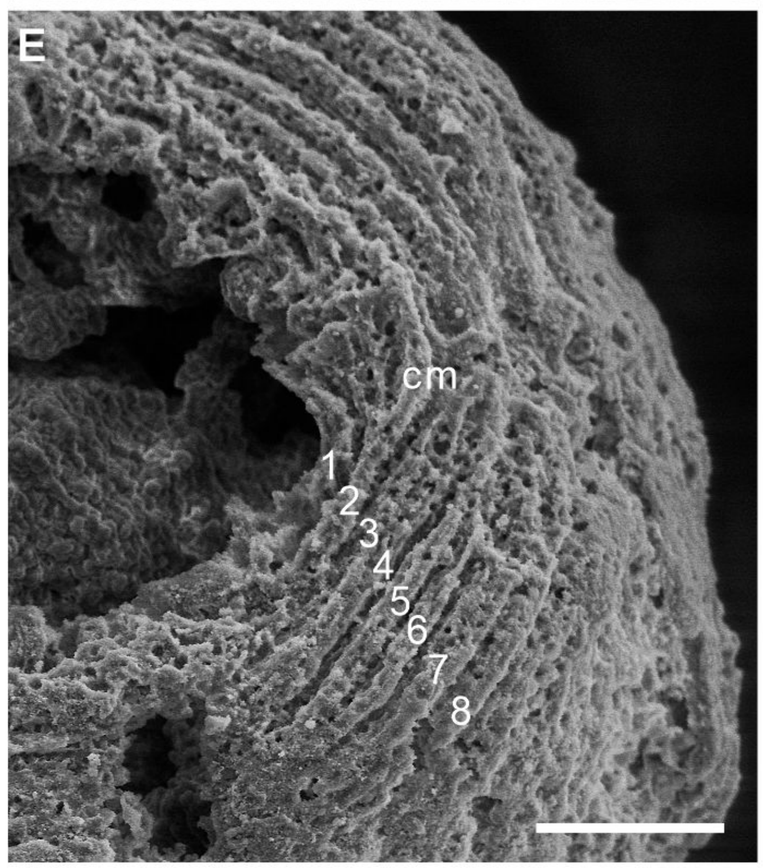
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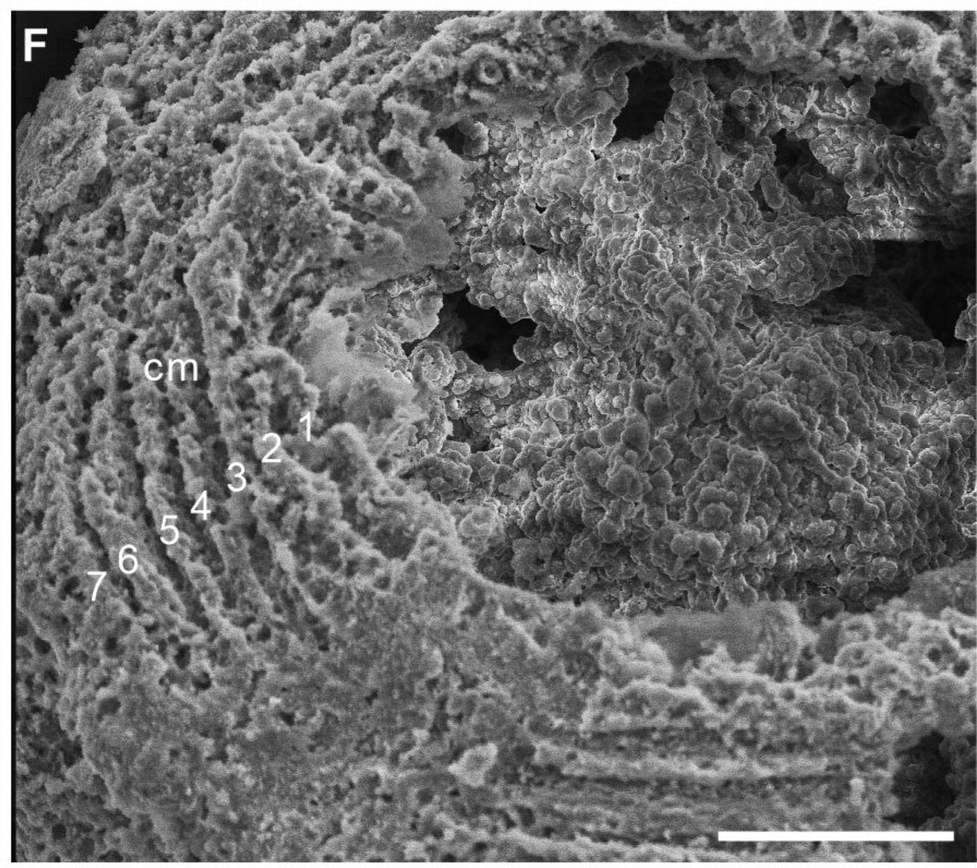
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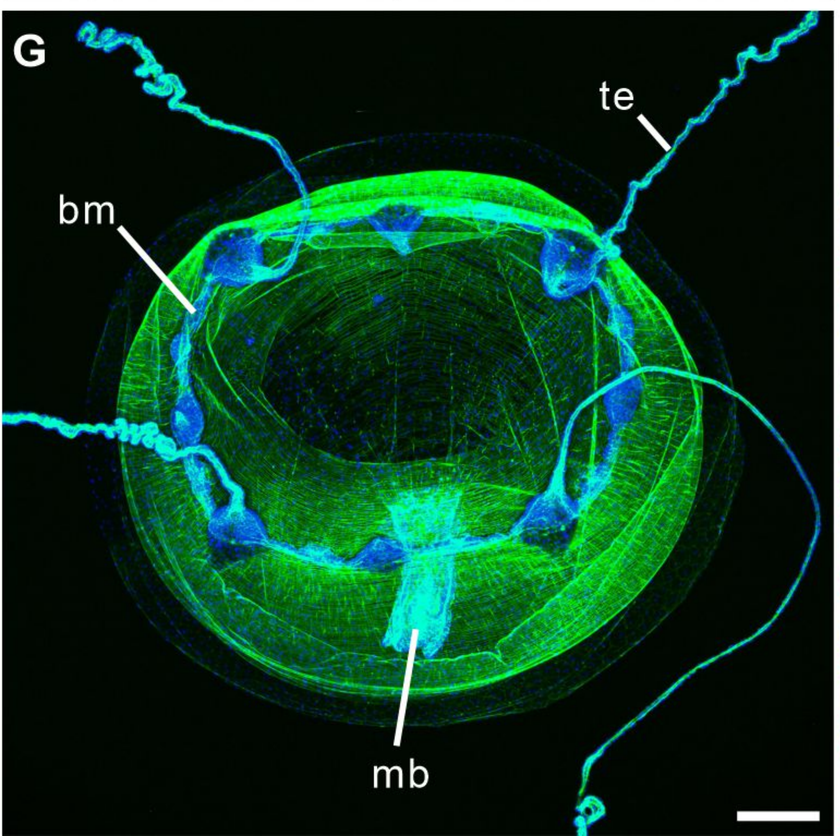
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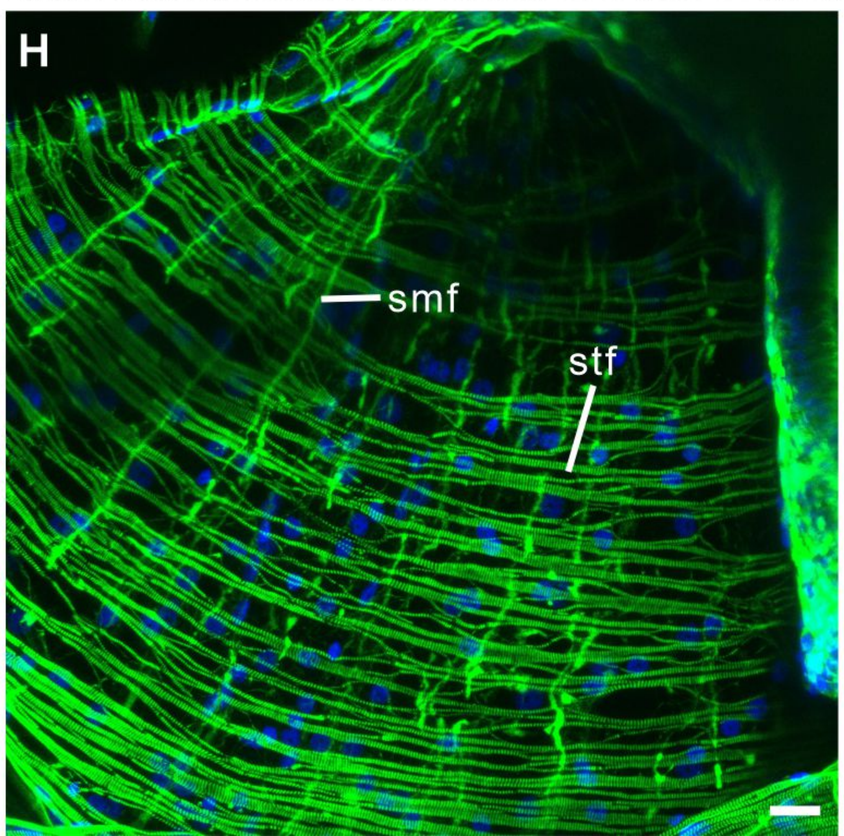
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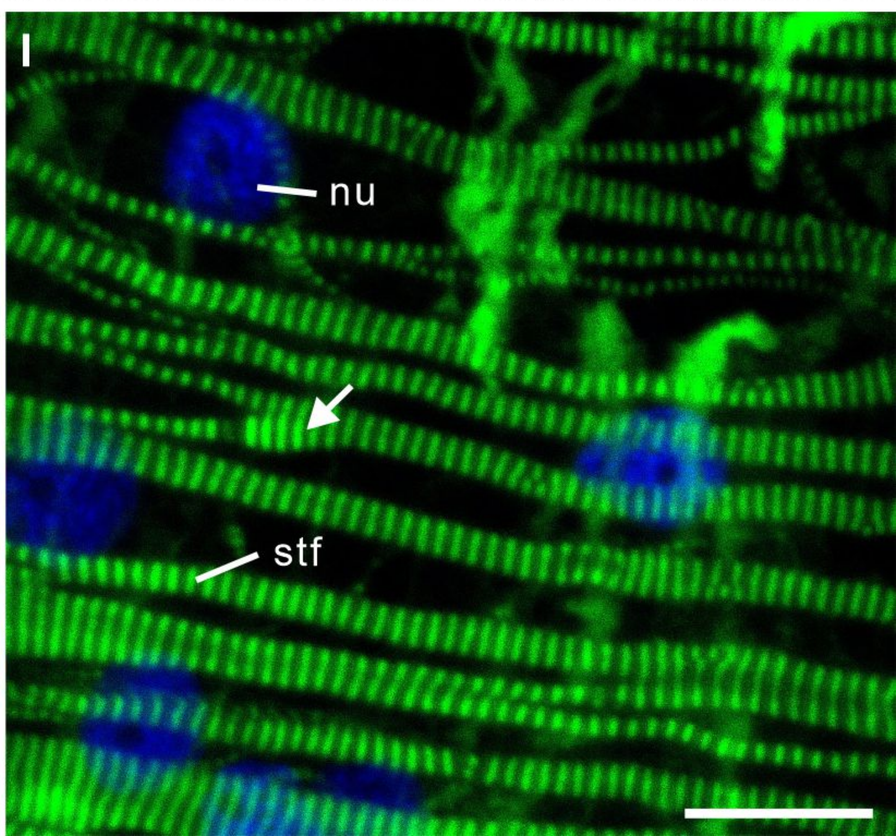
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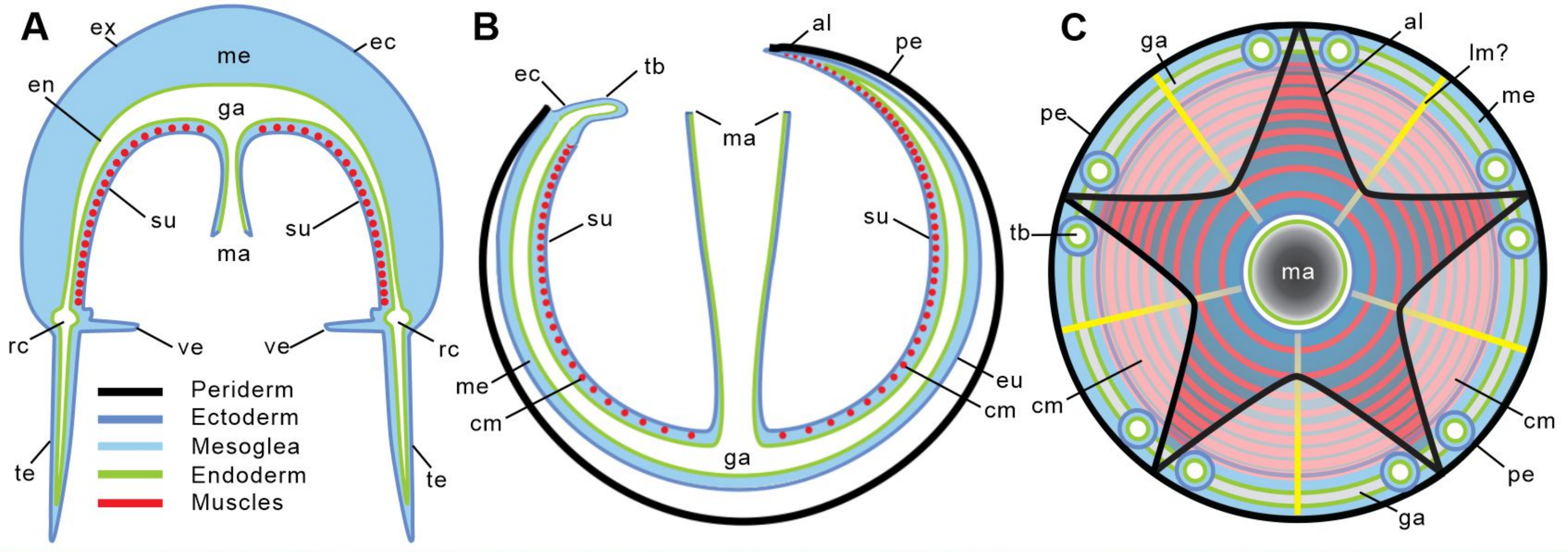

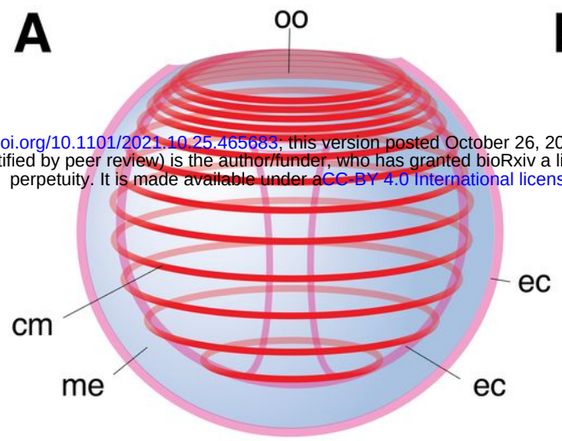

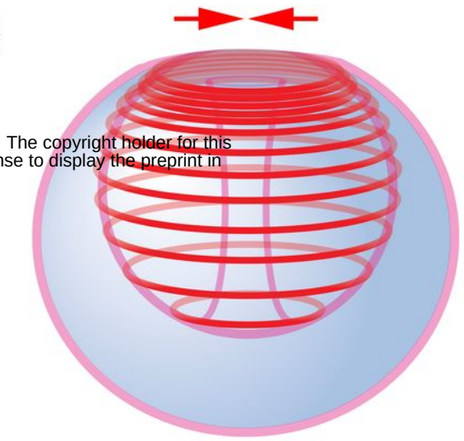

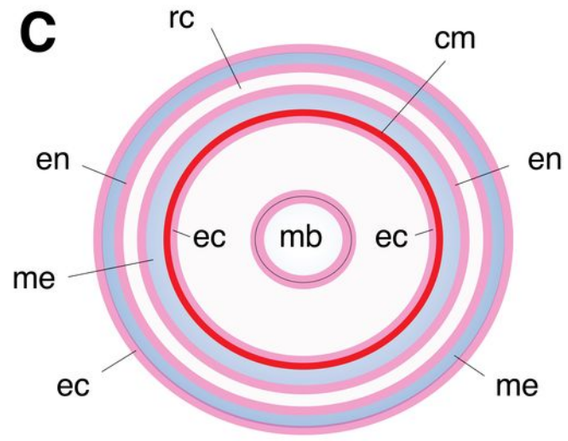

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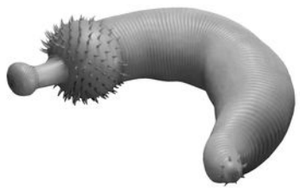

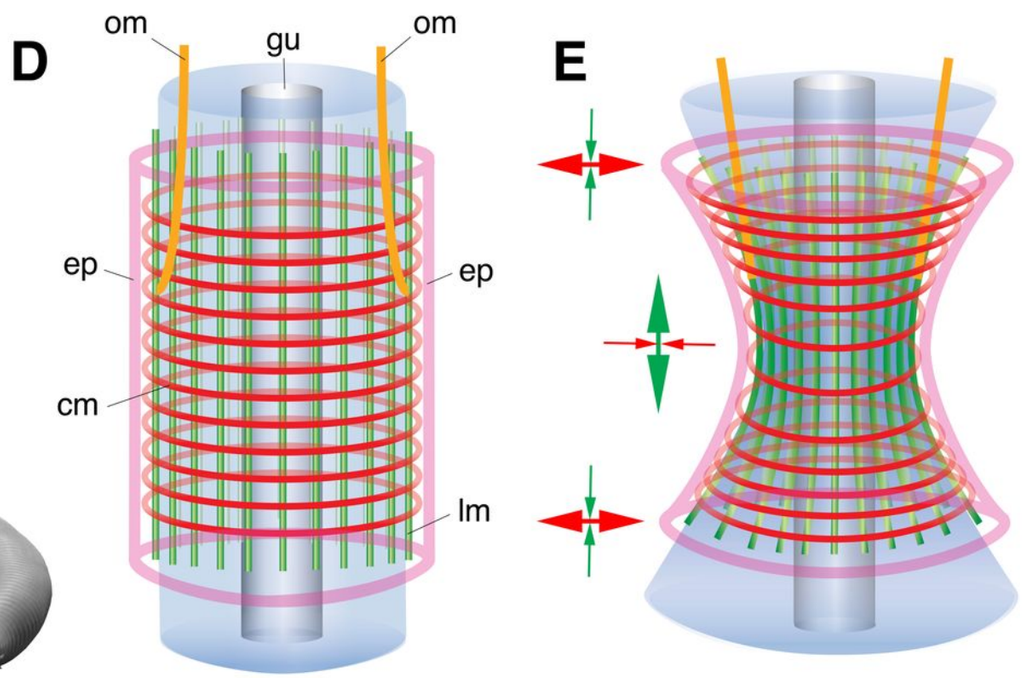

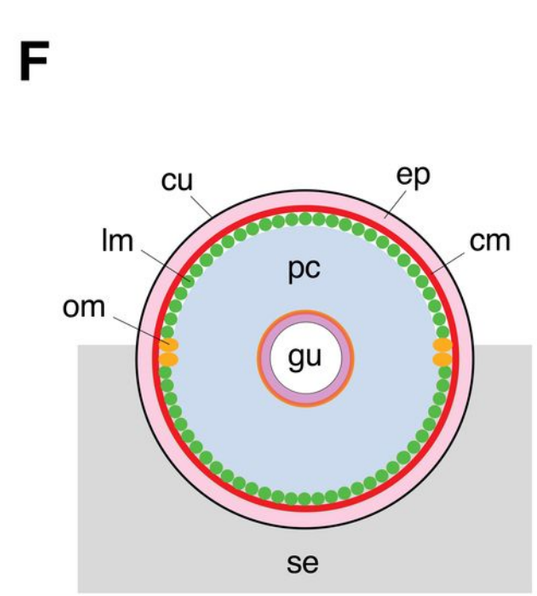

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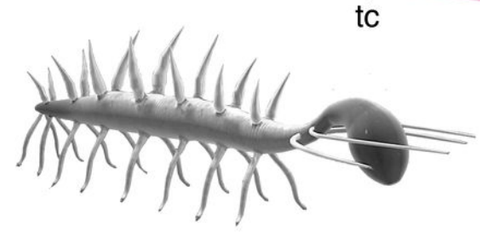

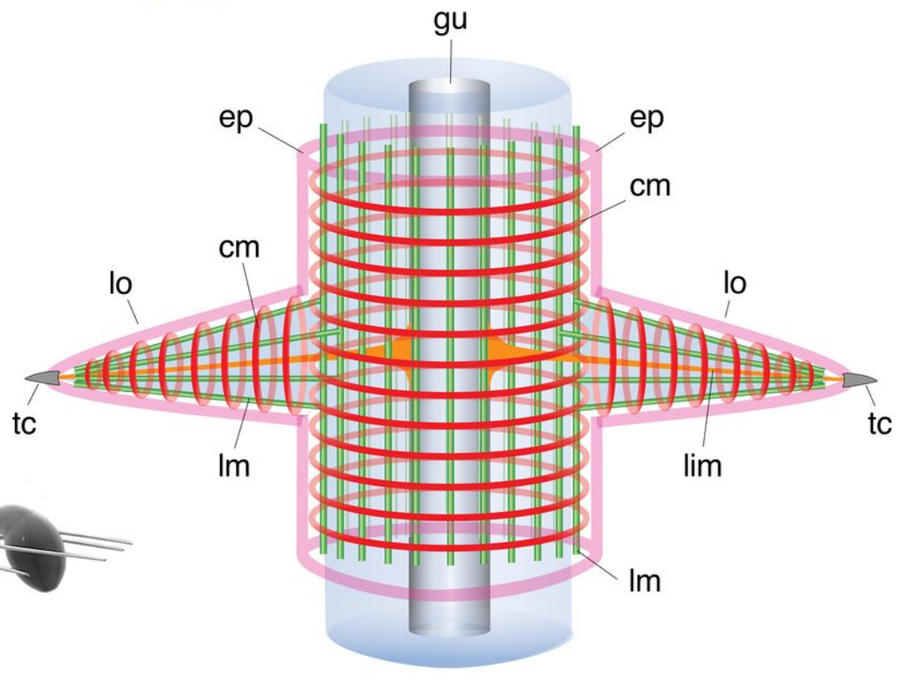

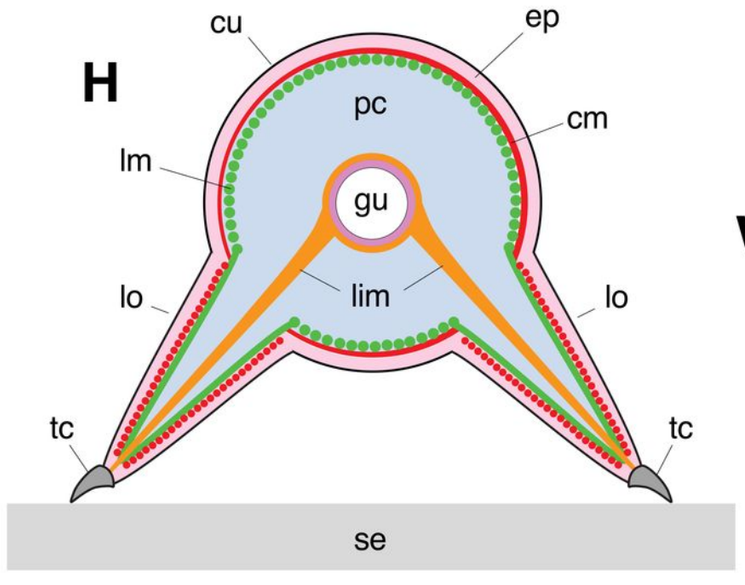

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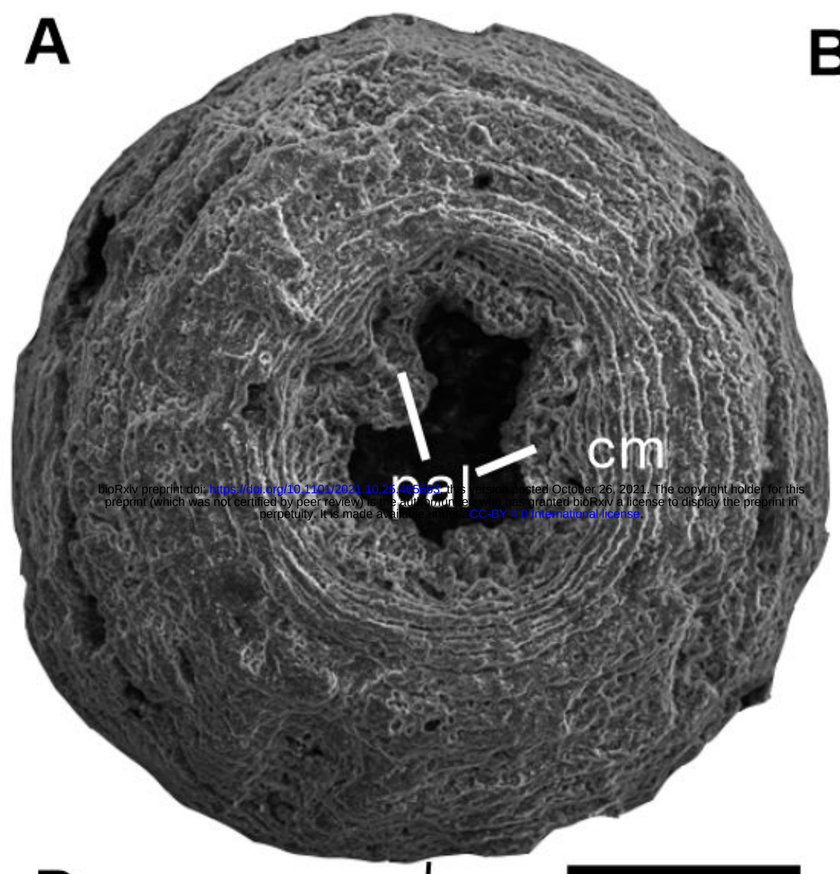

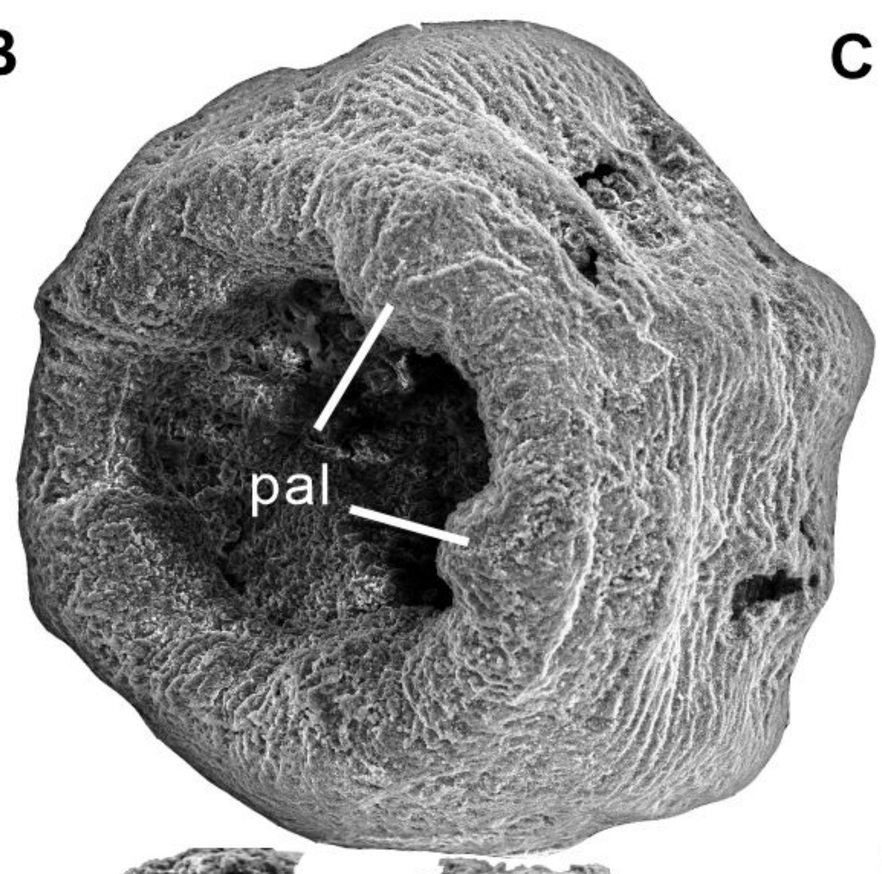

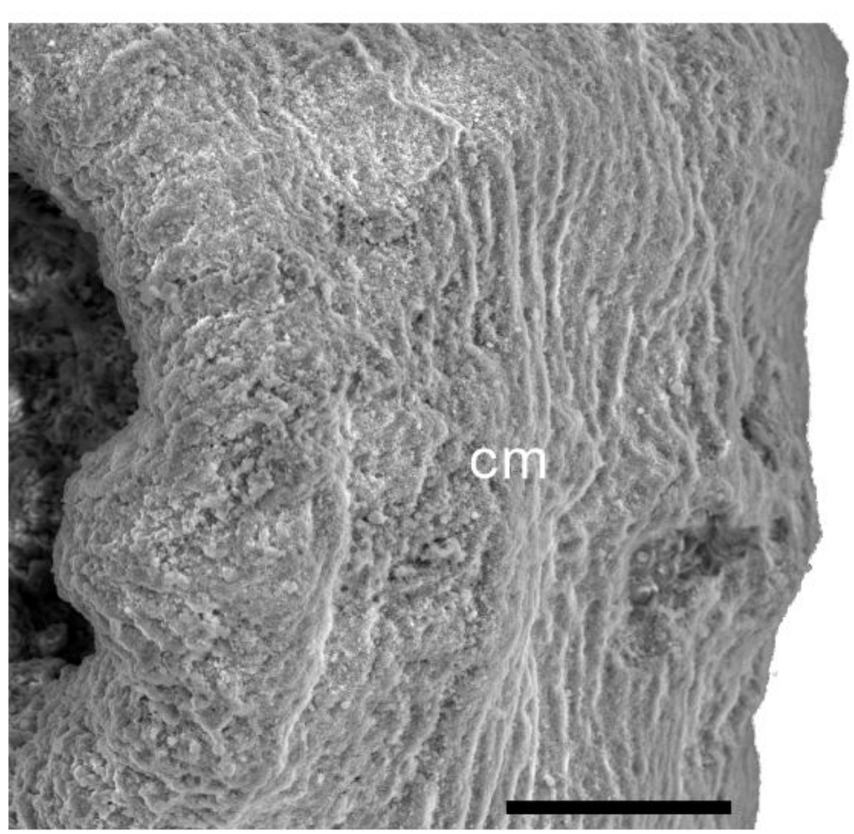

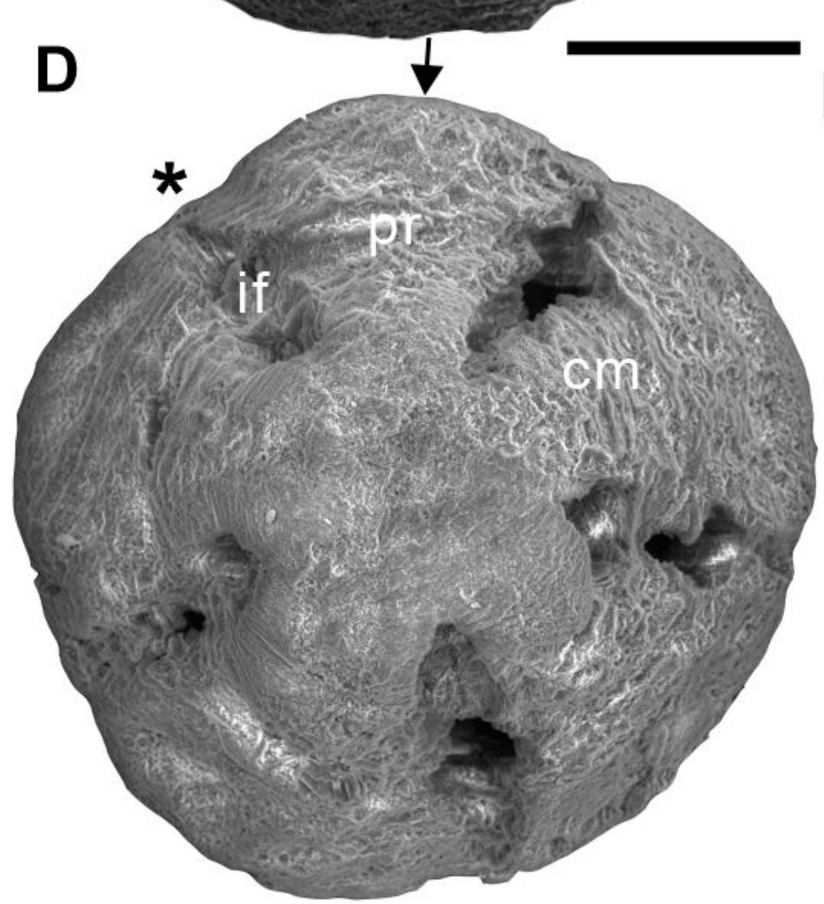

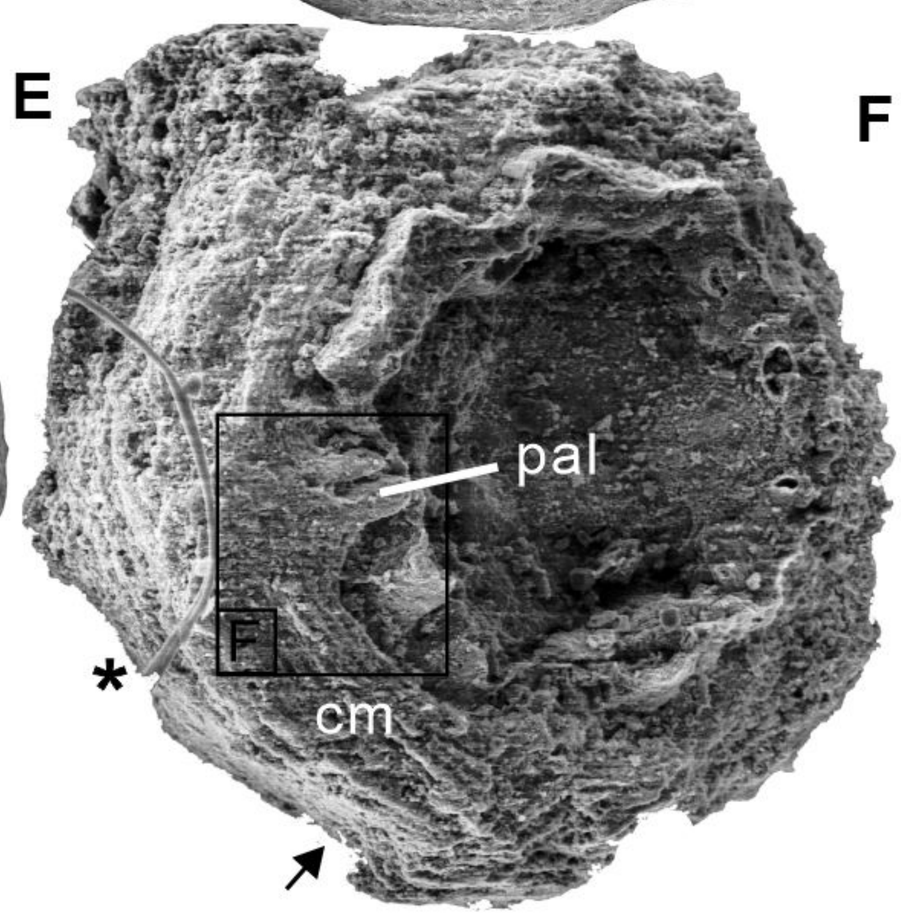

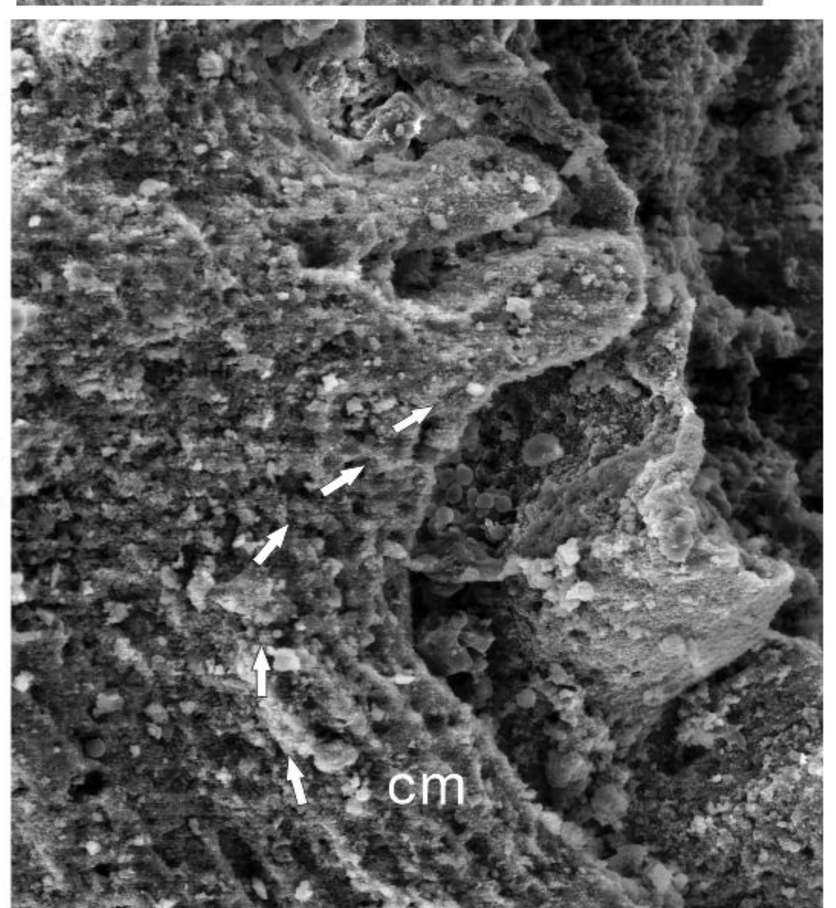

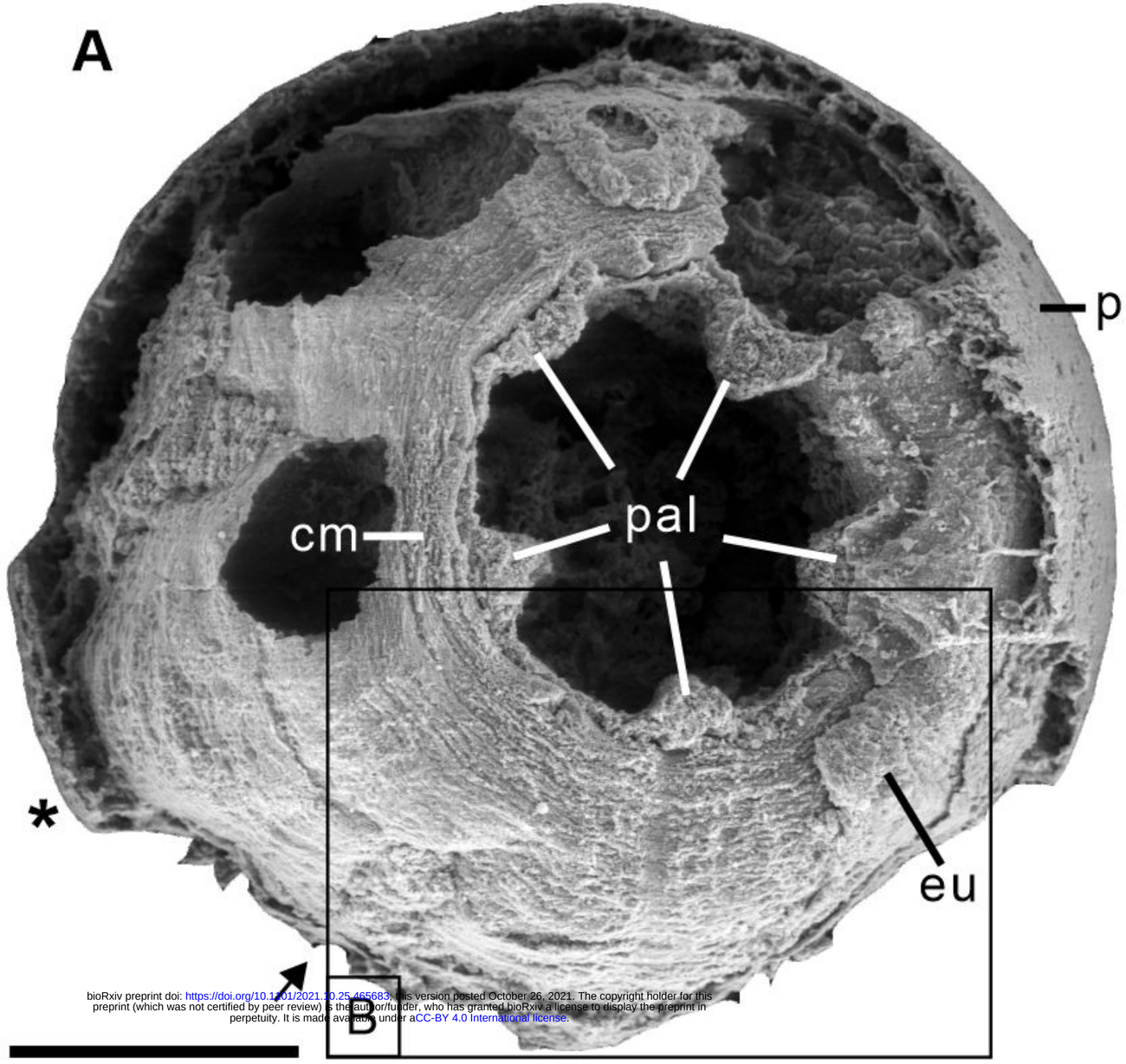

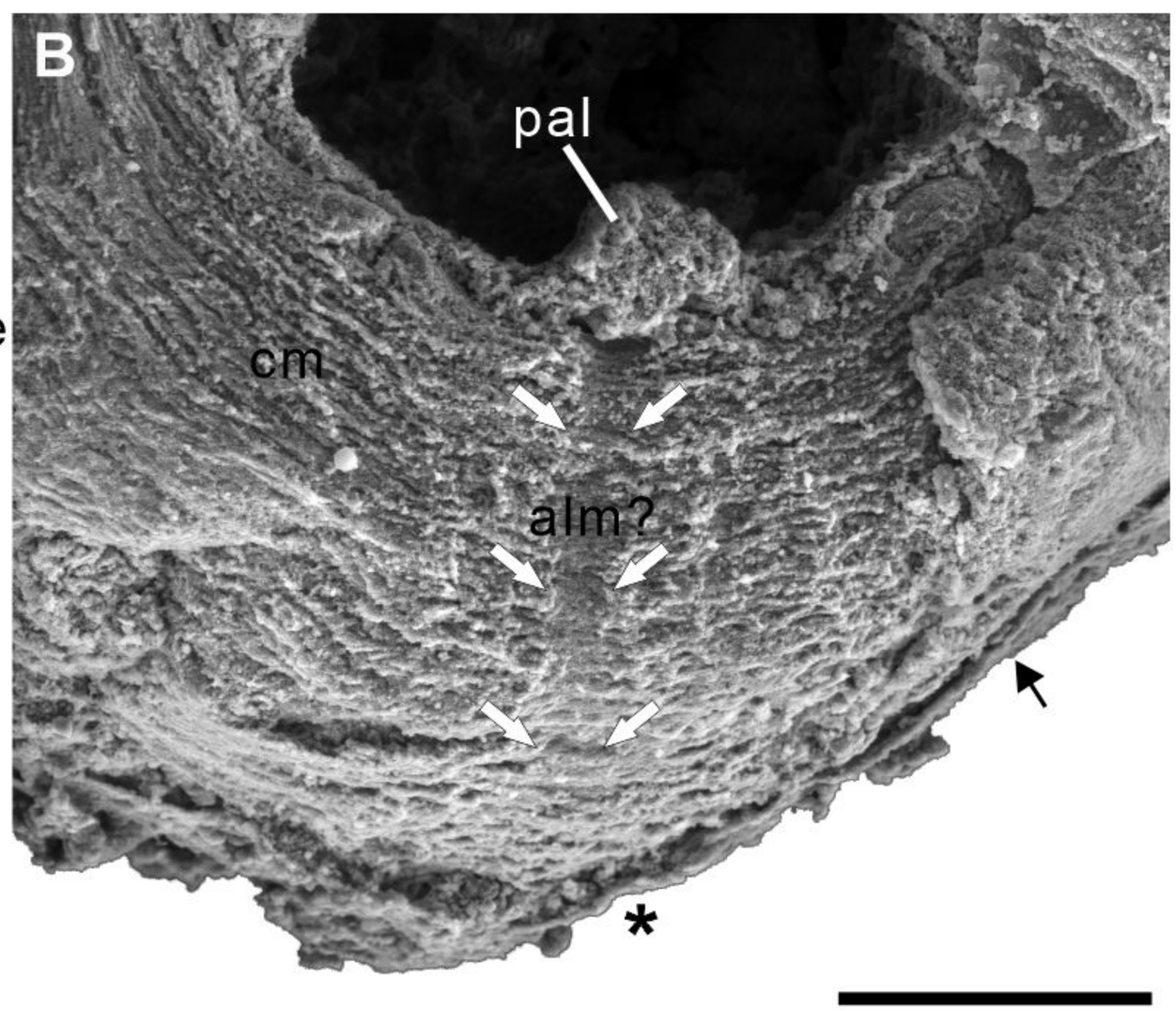

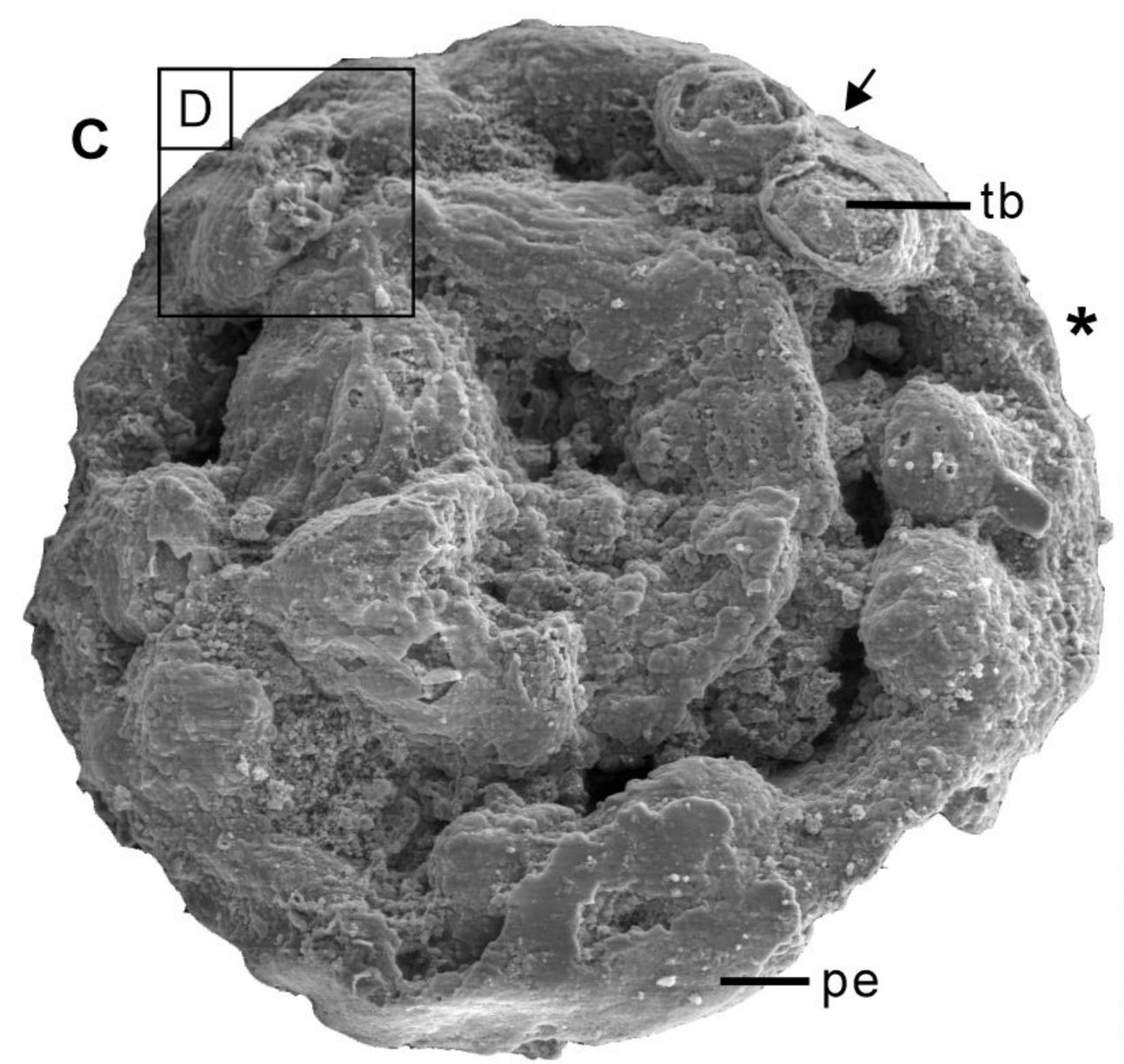

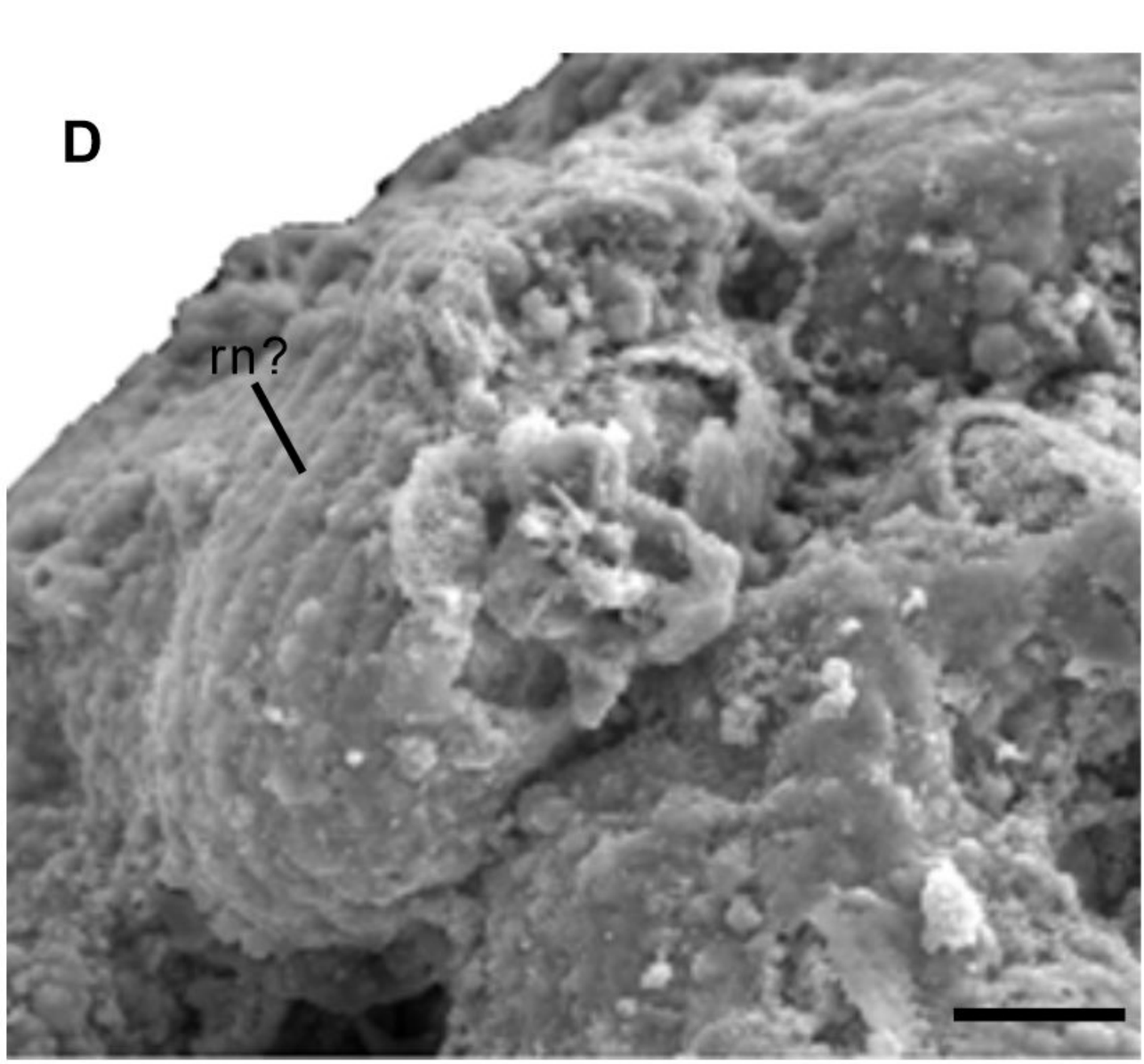

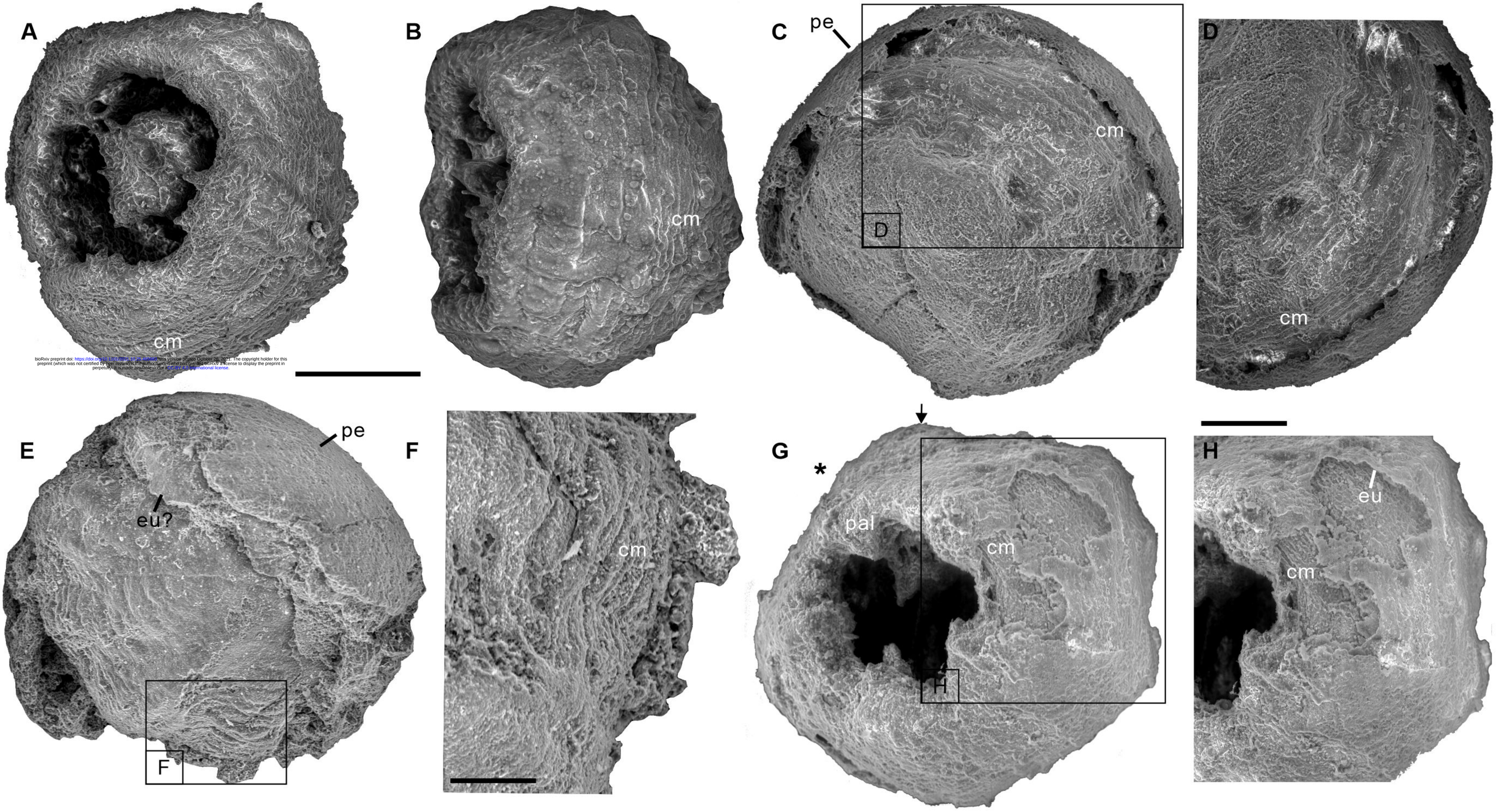

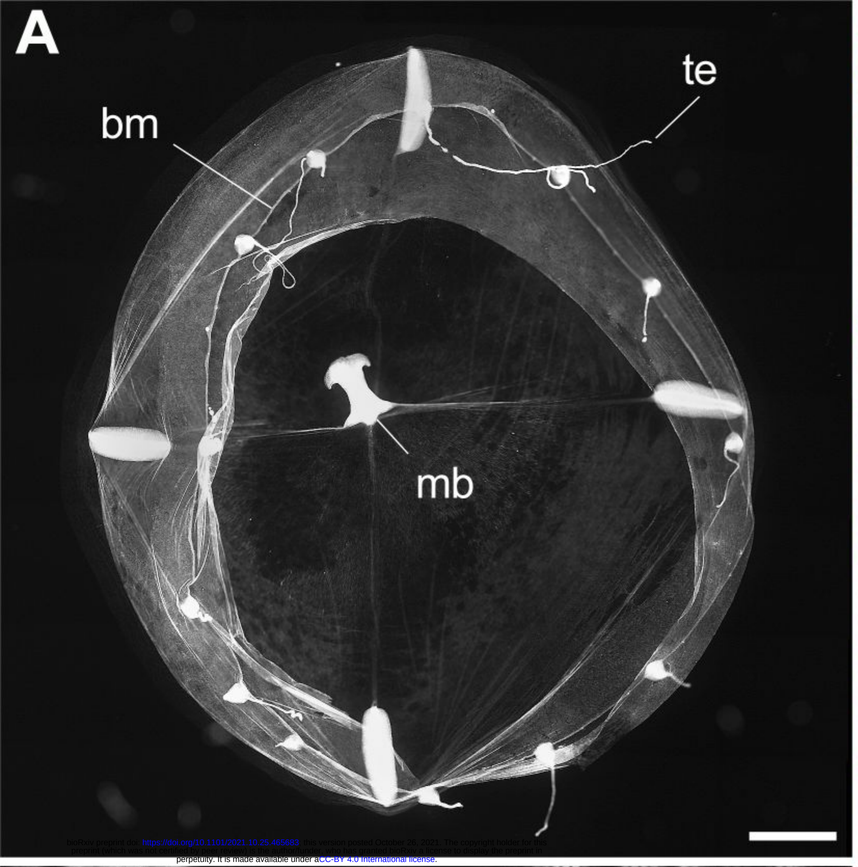

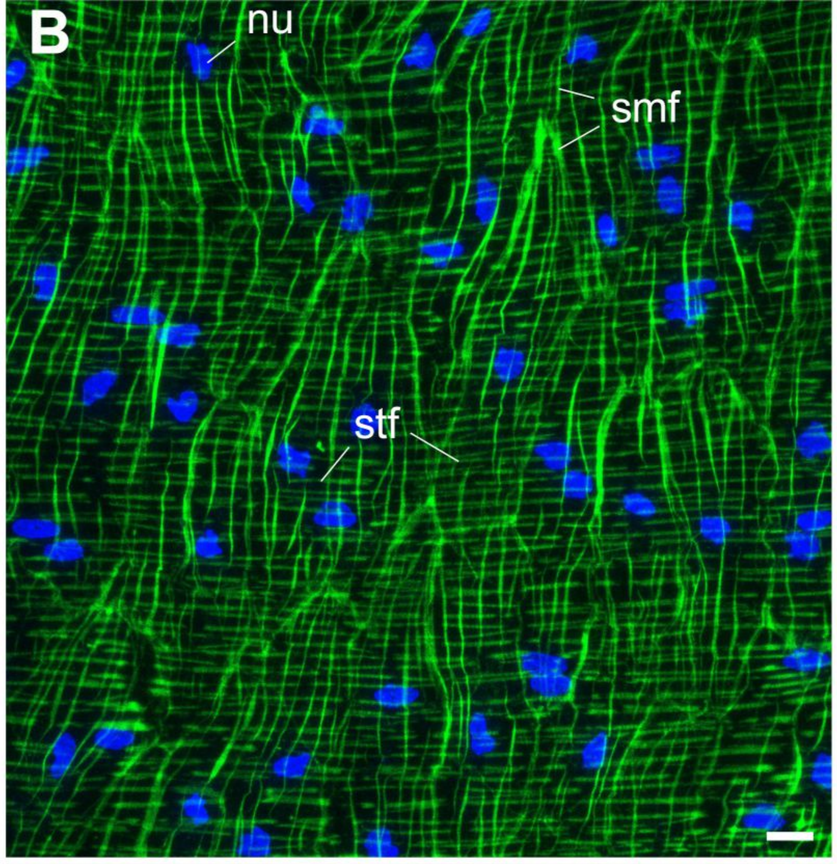

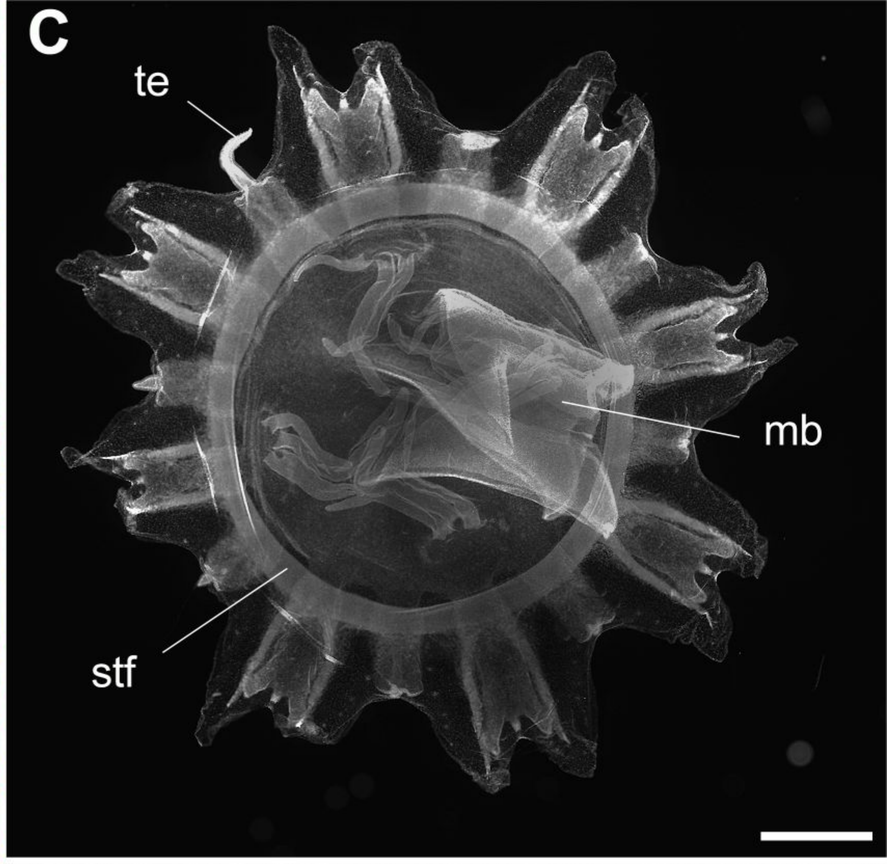

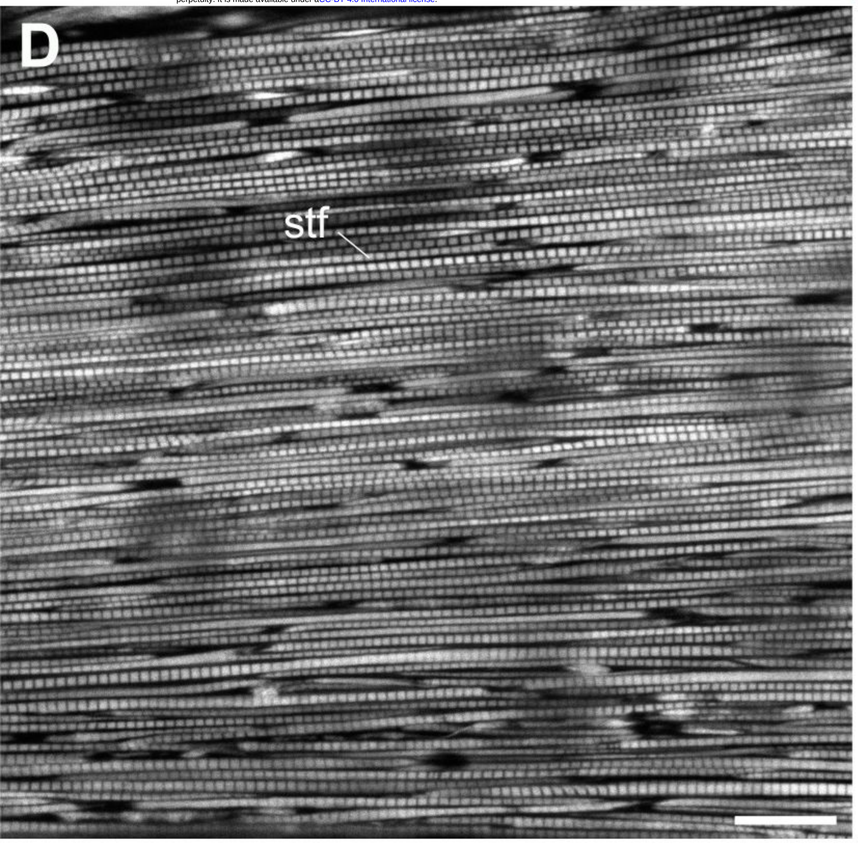

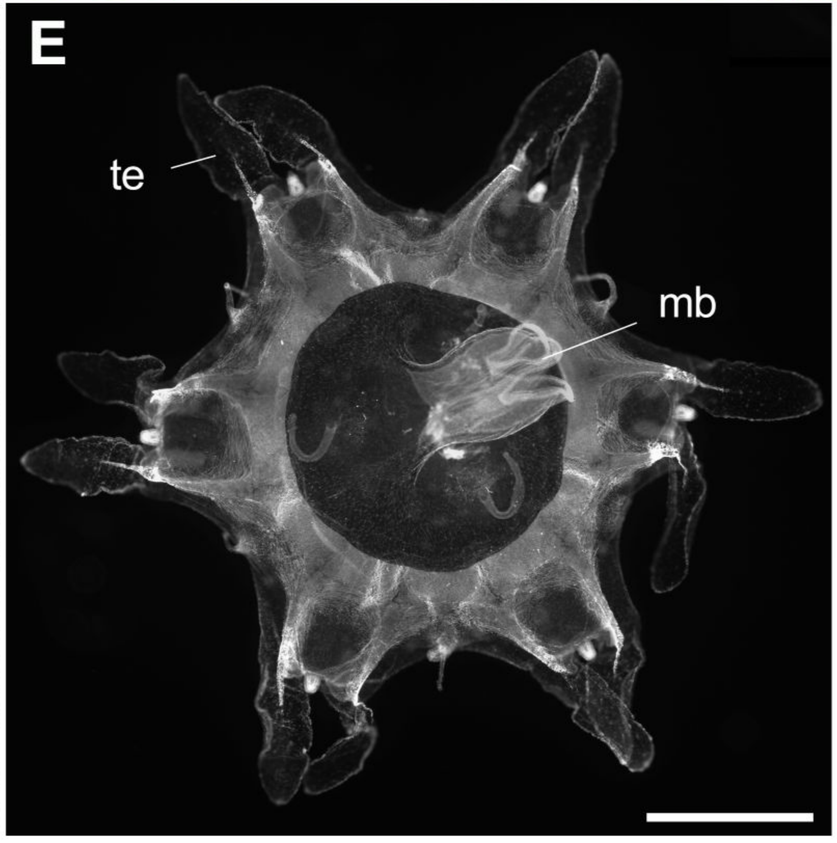

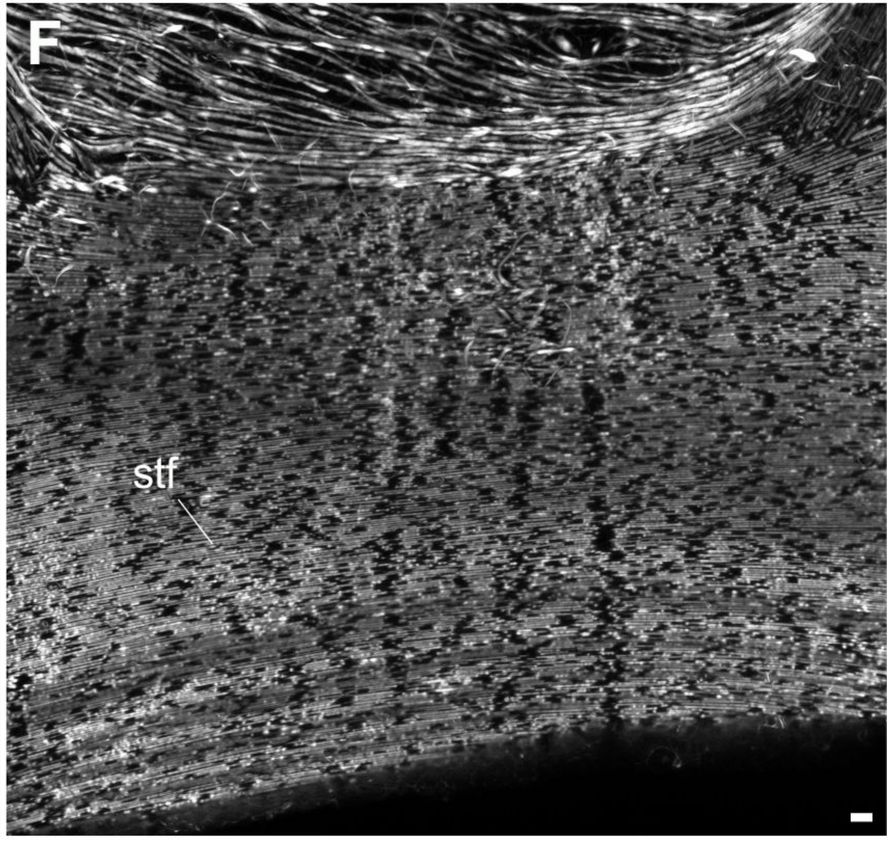

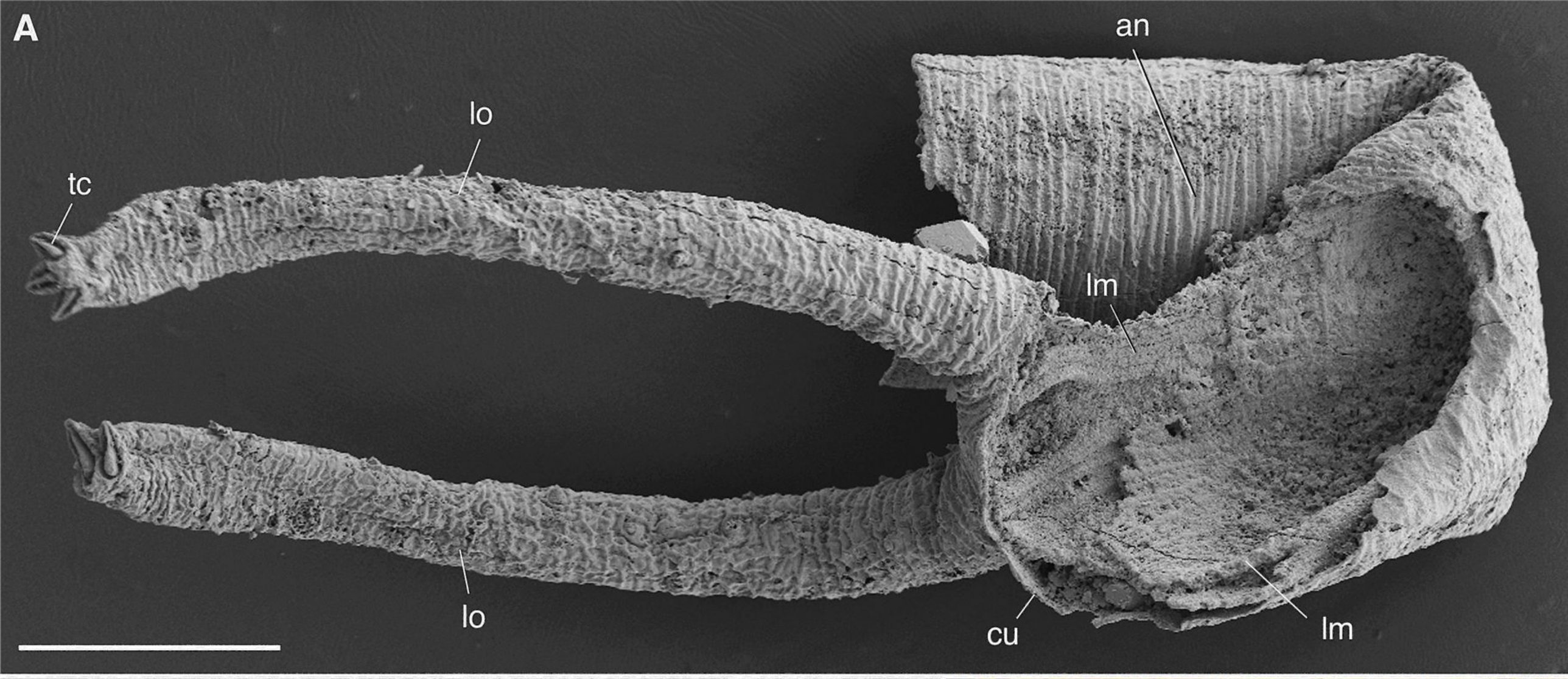

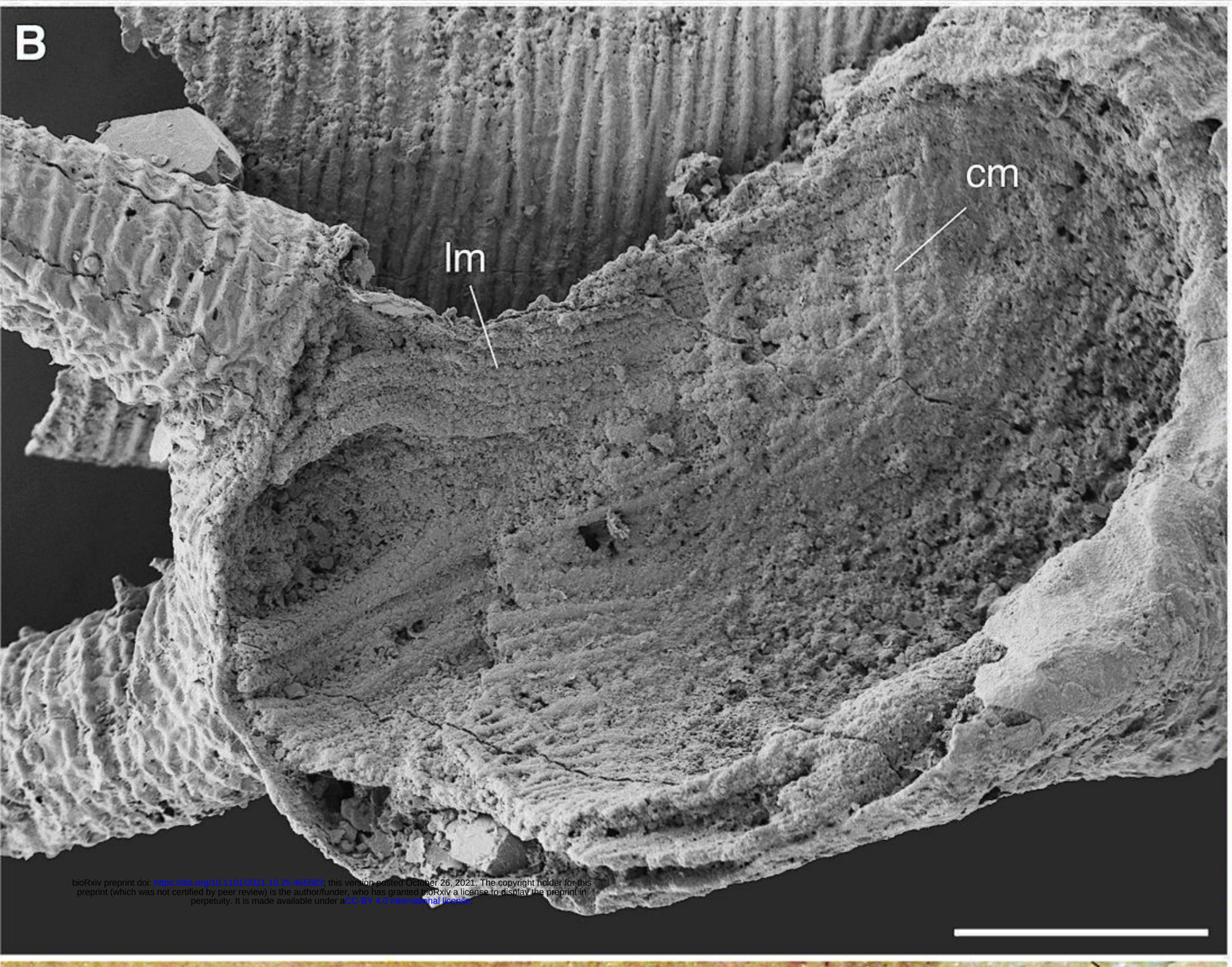

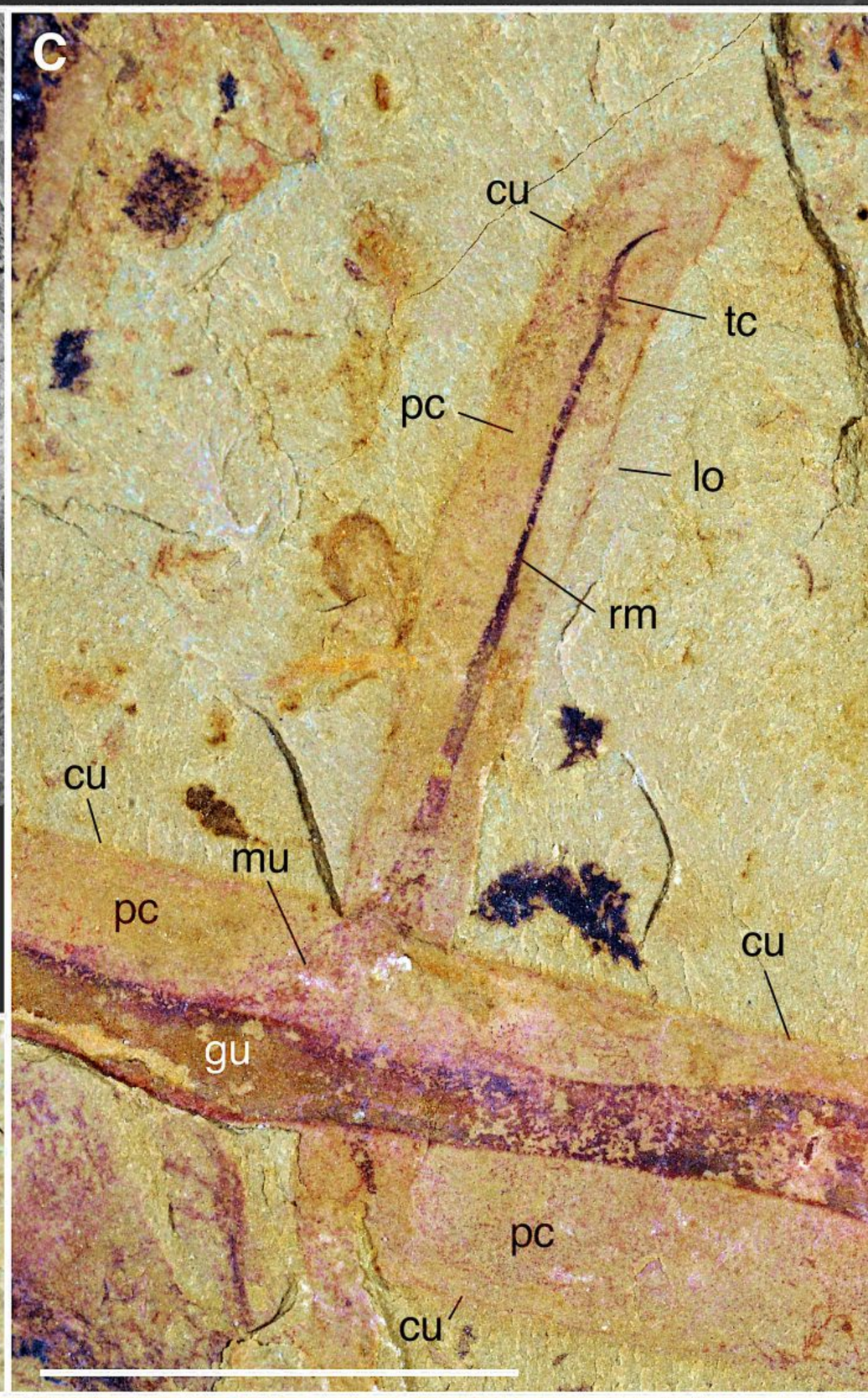

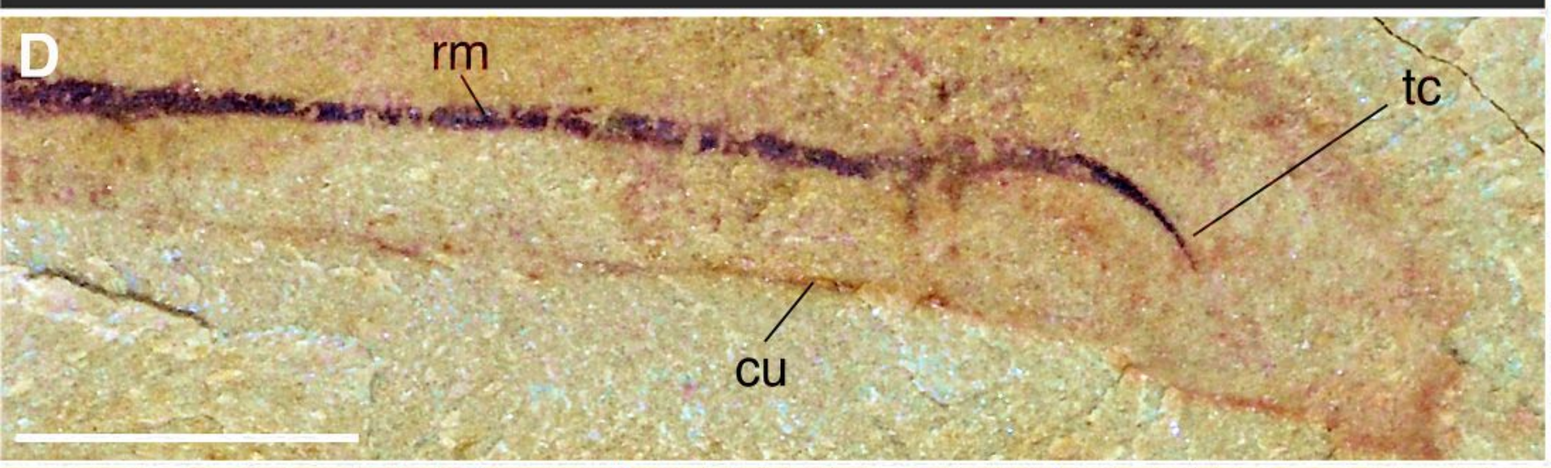

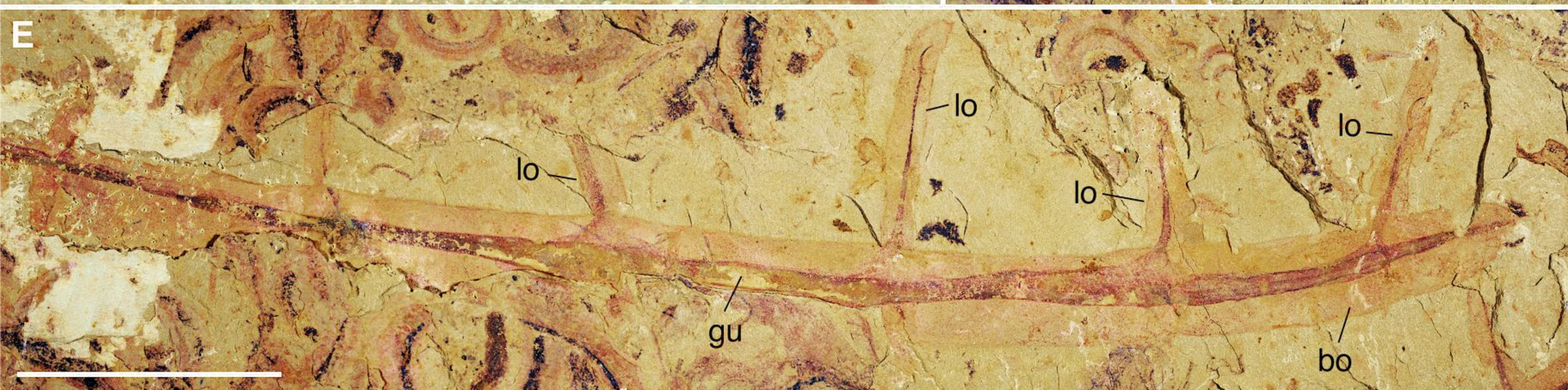

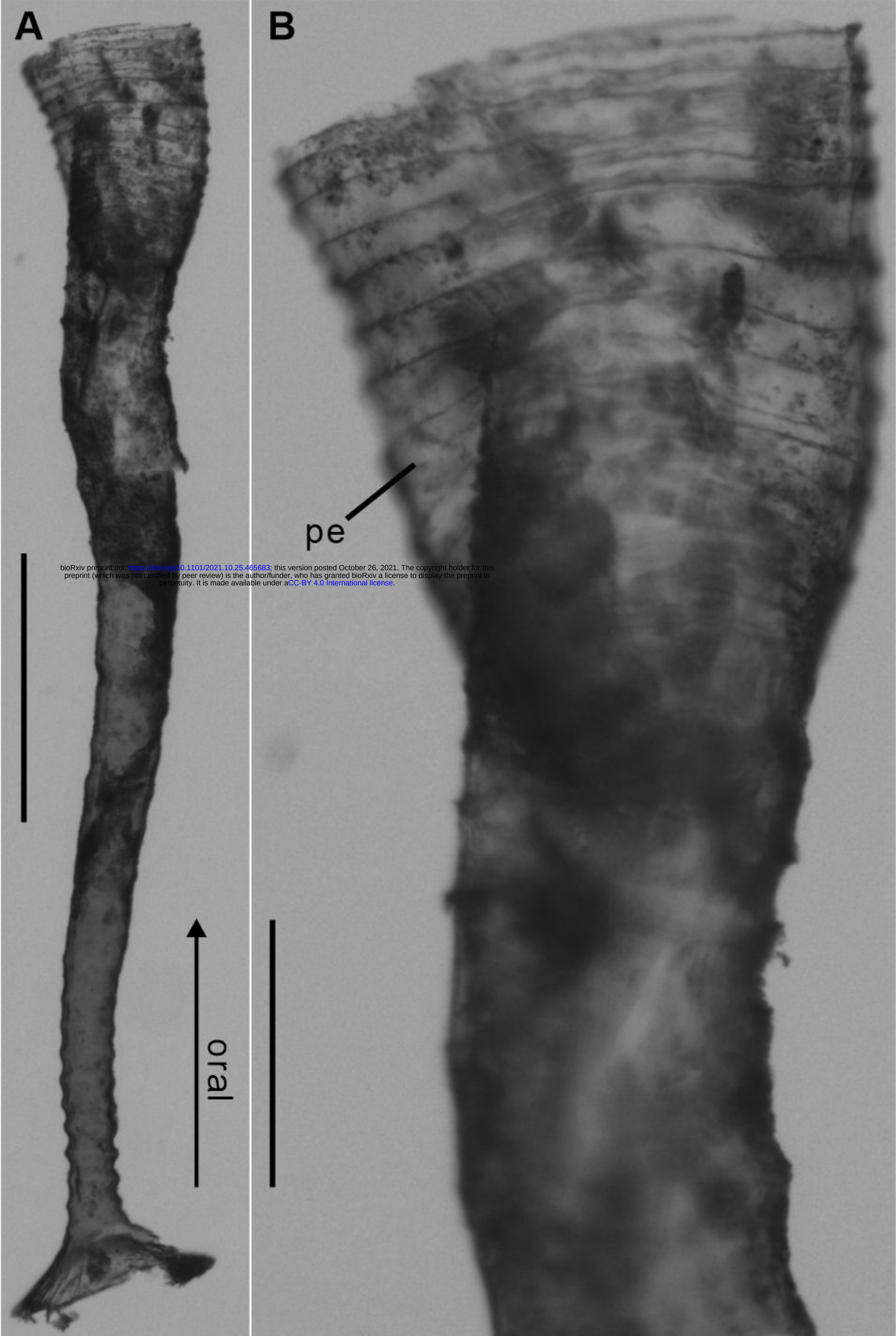

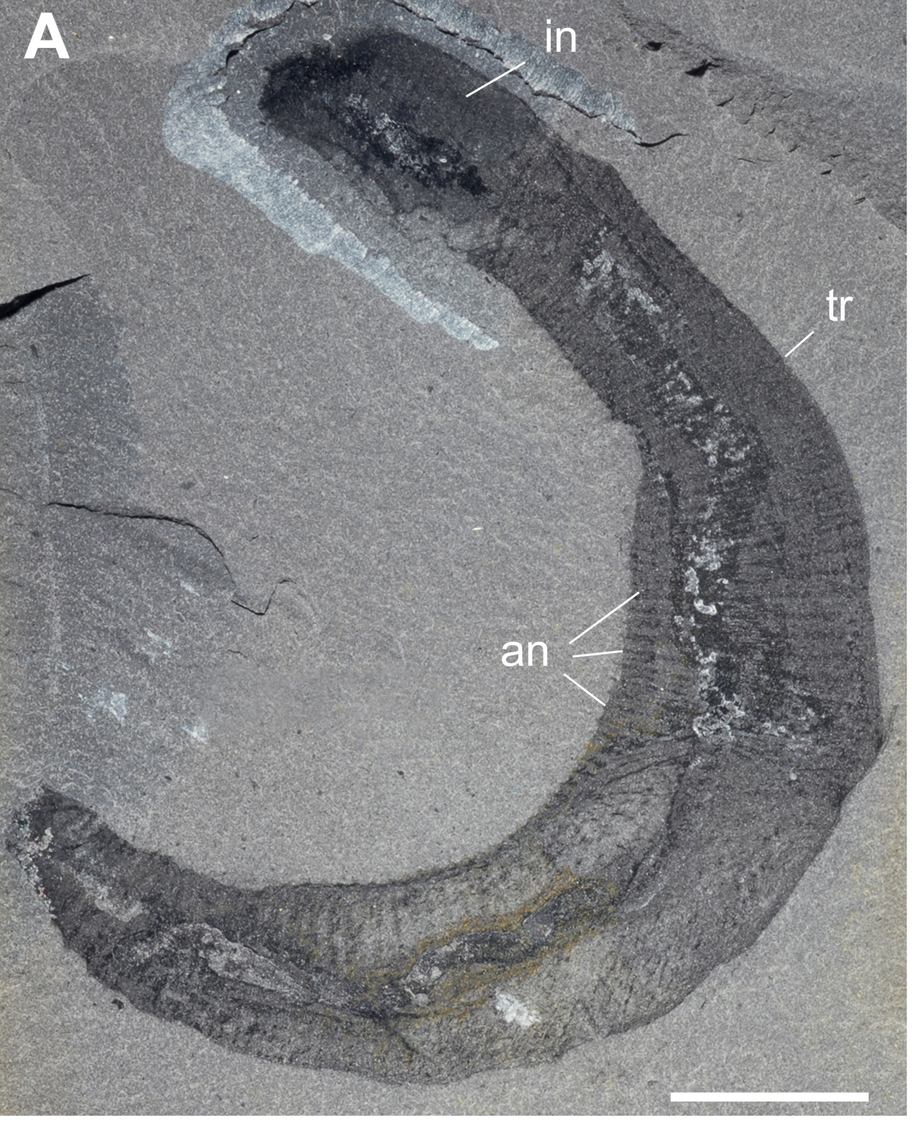
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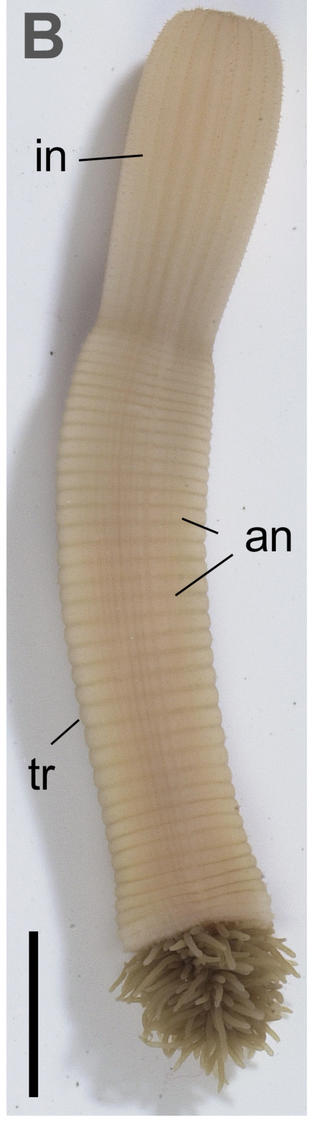
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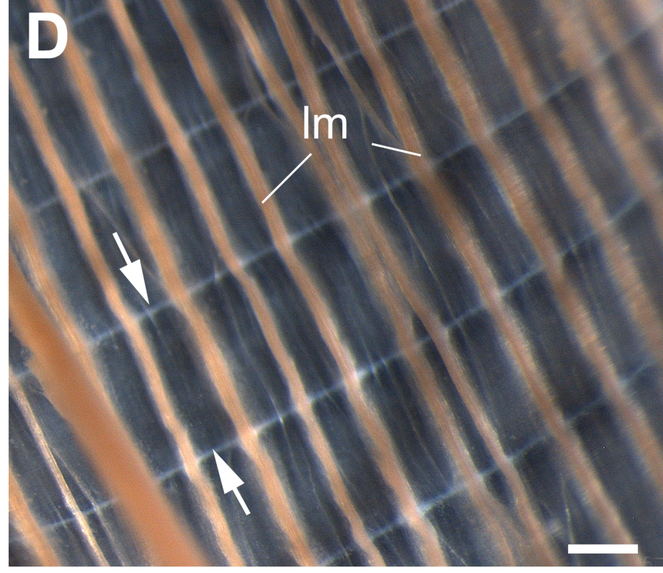
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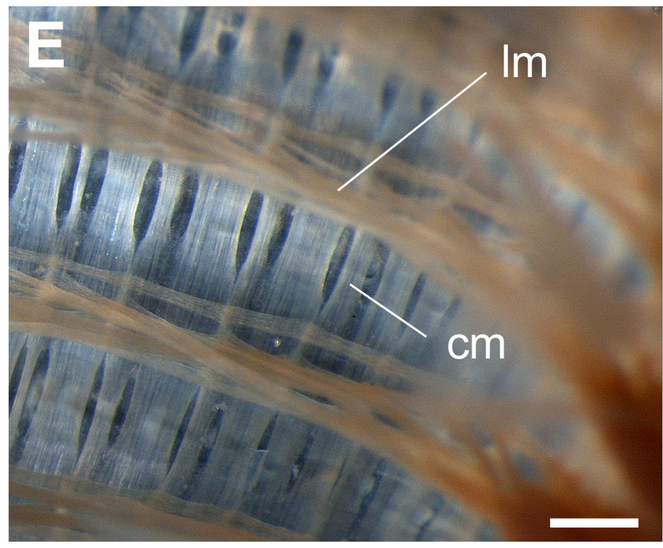
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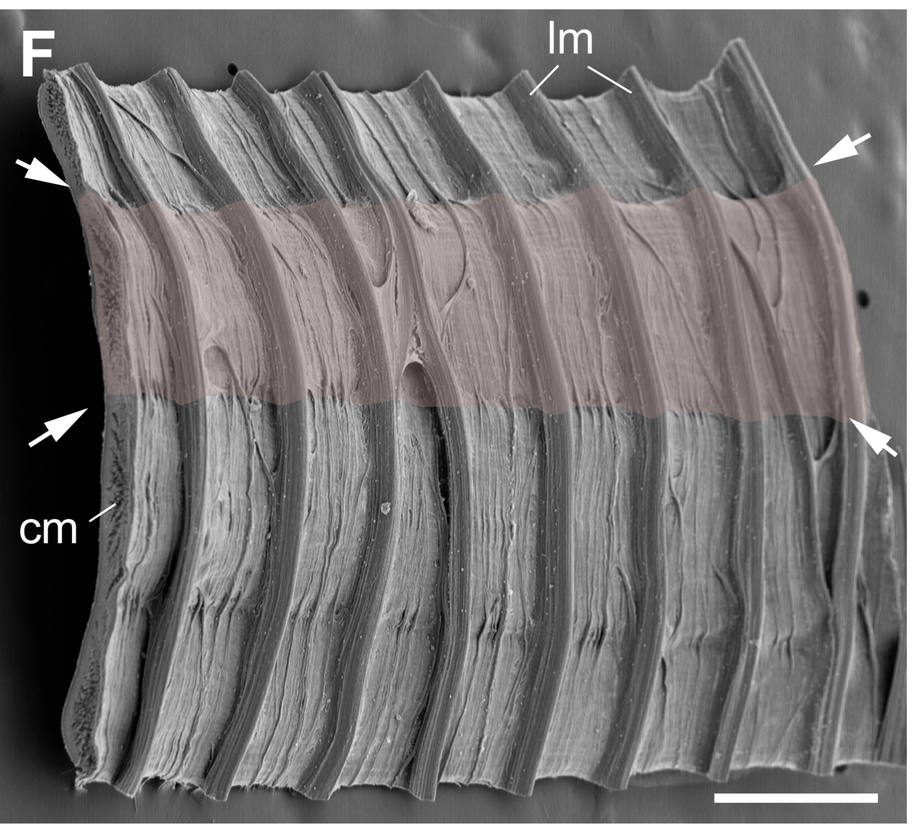
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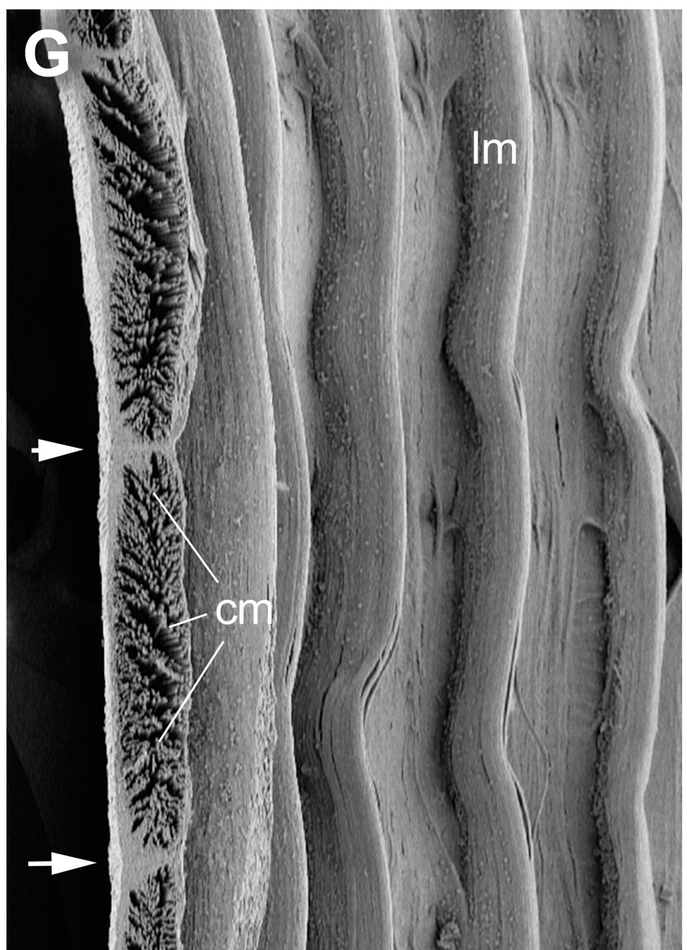
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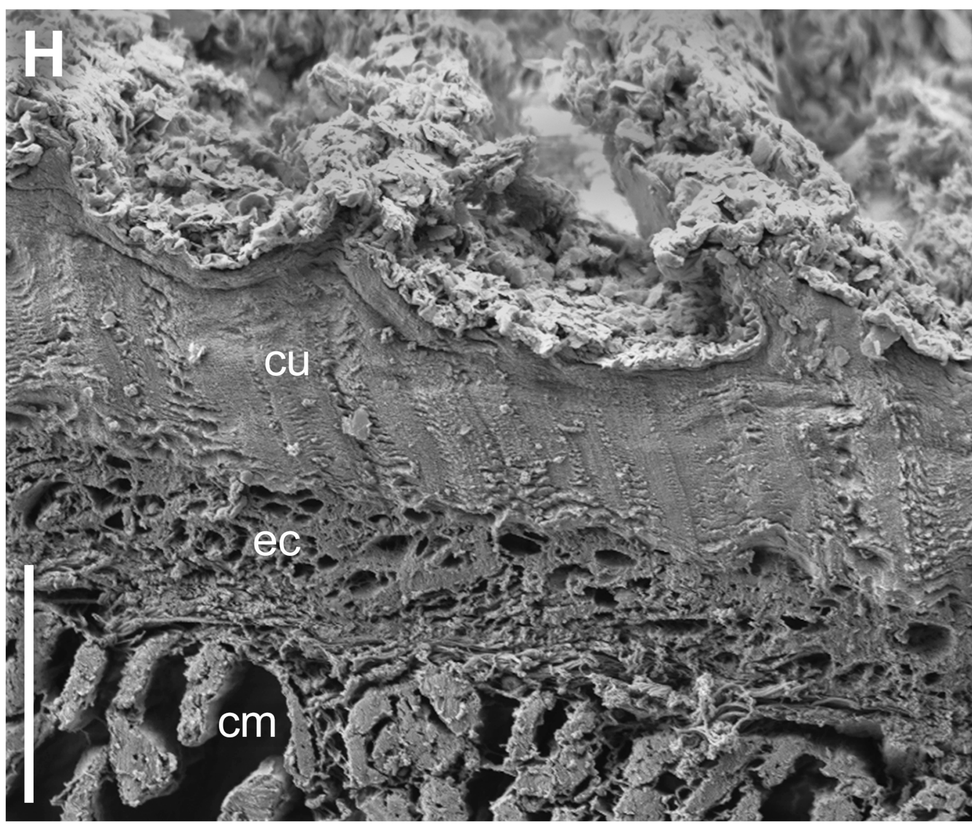
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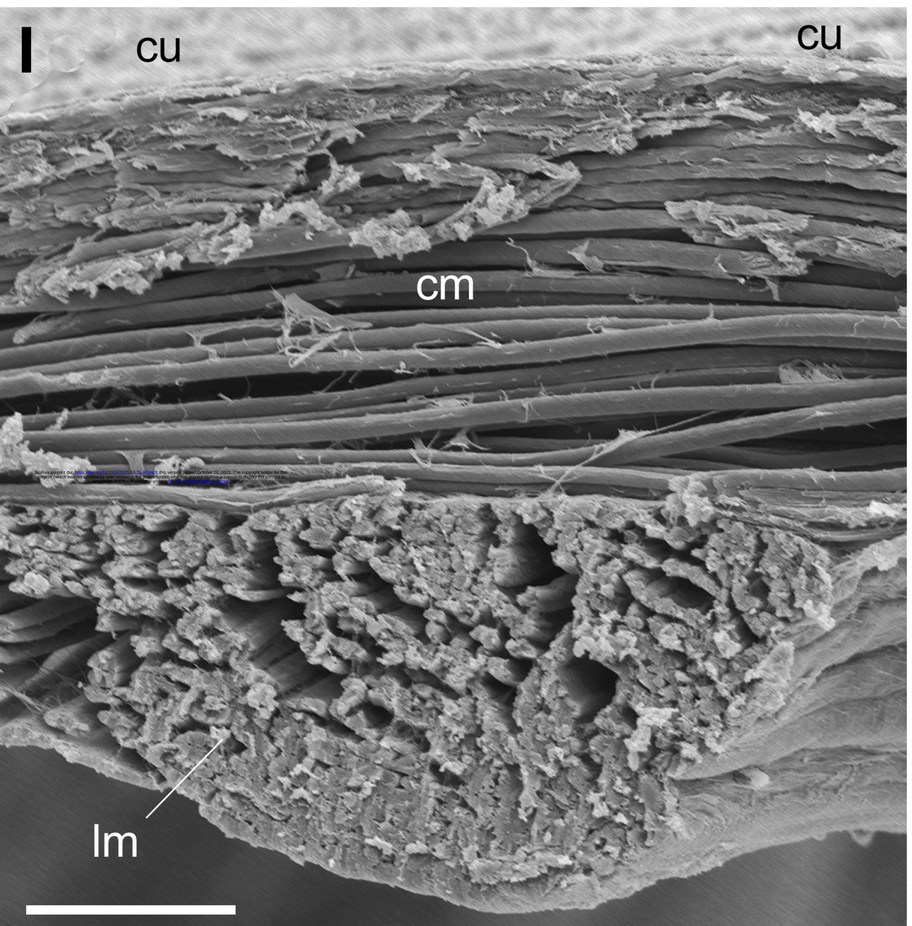
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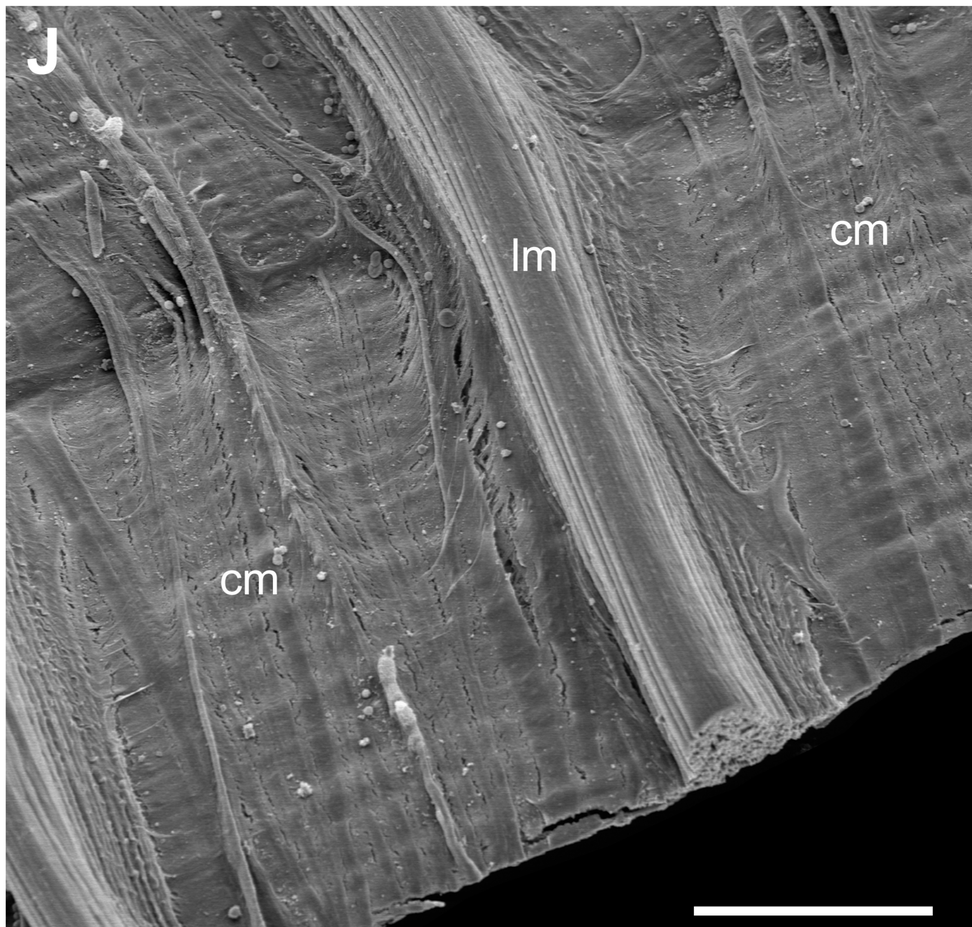
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