1 Type: Letter/Methods

3 selscan 2.0: scanning for sweeps in unphased data

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12 Abstract

13 Haplotype-based scans to identify recent and ongoing positive selection have become 14 commonplace in evolutionary genomics studies of numerous species across the tree of life. 15 However, the most widely adopted approaches require phased haplotypes to compute the key 16 statistics. Here we release a major update to the selscan software that re-defines popular 17 haplotype-based statistics for use with unphased "multi-locus genotype" data. We provide 18 unphased implementations of iHS, nSL, XP-EHH, and XP-nSL and evaluate their performance 19 across a range of important parameters in a generic demographic history. Source code and 20 executables are available at https://www.github.com/szpiech/selscan.

21 **1 Introduction**

22 Haplotype-based summary statistics—such as iHS (Voight, et al. 2006), nSL (Ferrer-23 Admetlla, et al. 2014), XP-EHH (Sabeti, et al. 2007), and XP-nSL (Szpiech, et al. 2021)—have 24 become commonplace in evolutionary genomics studies to identify recent and ongoing positive 25 selection in populations (e.g., Colonna, et al. 2014; Zoledziewska, et al. 2015; Nedelec, et al. 26 2016; Crawford, et al. 2017; Meier, et al. 2018; Lu, et al. 2019; Zhang, et al. 2020; Salmon, et al. 27 2021). When an adaptive allele sweeps through a population, it leaves a characteristic pattern 28 of long high-frequency haplotypes and low genetic diversity in the vicinity of the allele. These 29 statistics aim to capture these signals by summarizing the decay of haplotype homozygosity as 30 a function of distance from a putatively selected region, either within a single population (iHS

and nSL) or between two populations (XP-EHH and XP-nSL). However, each of these statistics
 presumes that haplotype phase is known.

Recent work has shown that converting haplotype data into multi-locus genotype data is an effective approach for using haplotype-based selection statistics such as G12, LASSI, and saltiLASSI (Harris, et al. 2018; Harris and DeGiorgio 2020; DeGiorgio and Szpiech 2021) in unphased data. Recognizing this, we have reformulated the iHS, nSL, XP-EHH, and XP-nSL statistics to use multi-locus genotypes and provided an easy-to-use implementation in selscan 2.0 (Szpiech and Hernandez 2014). We also evaluate the performance of these unphased statistics under various generic demographic models.

40 **2 New Approaches**

When the --unphased flag is set in selscan v2.0+, biallelic genotype data is collapsed into multi-locus genotype data by representing the genotype as either 0, 1, or 2—the number of derived alleles observed. In this case, selscan v2.0+ will then compute iHS, nSL, XP-EHH, and XP-nSL as described below. We follow the notation conventions of Szpiech and Hernandez (2014).

46 **2.1 Extended Haplotype Homozygosity**

In a sample of *n* diploid individuals, let *C* denote the set of all possible genotypes at locus x_0 . For multi-locus genotypes, $C := \{0,1,2\}$, representing the total counts of a derived allele. Let $C(x_i)$ be the set of all unique haplotypes extending from site x_0 to site x_i either upstream or downstream of x_0 . If x_1 is a site immediately adjacent to x_0 , then $C(x_1) :=$ $\{00,01,02,10,11,12,20,21,22\}$, representing all possible two-site multi-locus genotypes. We can then compute the extended haplotype homozygosity (EHH) of a set of multi-locus genotypes as

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$$EHH(x_i) \sum_{h \in \mathcal{C}(x_i)} \frac{\binom{n_h}{2}}{\binom{n}{2}},$$

54 where n_h is the number of observed haplotypes of type *h*.

If we wish to compute the EHH of a subset of observed haplotypes that all contain the same 'core' multi-locus genotype, let $\mathcal{H}_c(x_i)$ be the partition of $\mathcal{C}(x_i)$ containing genotype $c \in \mathcal{C}$ at x_0 . For example, choosing a homozygous derived genotype (c = 2) as the core, $\mathcal{H}_2 :=$ {20,21,22}. Thus, we can compute the EHH of all individuals carrying a given genotype at site x_0 extending out to site x_i as

$$EHH_c(x_i) = \sum_{h \in \mathcal{H}_c(x_i)} \frac{\binom{n_h}{2}}{\binom{n_c}{2}},$$

61 where n_h is the number of observed haplotypes of type *h* and n_c is the number of observed

62 multi-locus genotypes with core genotype of *c*. Finally, we can compute the complement EHH of

63 a sample of multi-locus genotypes as

$$64 \qquad \qquad cEHH_c(x_i) = \sum_{h \in \mathcal{C}(x_i) \setminus \mathcal{H}_c(x_i)} \frac{\binom{n_h}{2}}{\binom{n_{c'}}{2}},$$

65 where $n_{c'}$ is the number of observed multi-locus genotypes with a core genotype of not *c*.

66 2.2 iHS and nSL

67 Unphased iHS and nSL are calculated using the equations above. First, we compute the 68 integrated haplotype homozygosity (iHH) for the homozygous ancestral (c = 0) and derived (c =69 2) core genotypes as

70
$$iHH_{c} = \sum_{i=1}^{|\mathcal{D}|} \frac{1}{2} (EHH_{c}(x_{i-1}) + EHH_{c}(x_{i})) d(x_{i-1}, x_{i}) + \sum_{i=1}^{|\mathcal{U}|} \frac{1}{2} (EHH_{c}(x_{i-1}) + EHH_{c}(x_{i})) d(x_{i-1}, x_{i}),$$

where \mathcal{D} is the set of downstream sites from the core locus and \mathcal{U} is the set of upstream sites. $d(x_{i-1}, x_i)$ is a measure of genomic distance between to markers and is the genetic distance in centimorgans or physical distance in basepairs for iHS (Voight, et al. 2006) or the number of sites observed for nSL (Ferrer-Admetlla, et al. 2014). We similarly compute the complement integrated haplotype homozygosity (ciHH) for both homozygous core genotypes as

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$$ciHH_{c} = \sum_{i=1}^{|\mathcal{D}|} \frac{1}{2} (cEHH_{c}(x_{i-1}) + cEHH_{c}(x_{i})) d(x_{i-1}, x_{i})$$

77
$$+ \sum_{i=1}^{|u|} \frac{1}{2} (cEHH_c(x_{i-1}) + cEHH_c(x_i)) d(x_{i-1}, x_i).$$

78 The (unstandardized) unphased iHS is then calculated as

79
$$iHS = \begin{cases} iHS_2, & \text{if } iHS_2 > iHS_0 \\ -iHS_0, & \text{otherwise'} \end{cases}$$

80 where
$$iHS_2 = \log_{10}\left(\frac{iHH_2}{ciHH_2}\right)$$
 and $iHS_0 = \log_{10}\left(\frac{iHH_0}{ciHH_0}\right)$. Unstandardized iHS scores are then

normalized in frequency bins, as previously described (Voight, et al. 2006; Ferrer-Admetlla, et

82 al. 2014). Unstandardized unphased nSL is computed similarly with the appropriate distance

83 measure. Large positive scores indicate long high-frequency haplotypes with a homozygous

84 derived core genotype, and large negative scores indicate long high-frequency haplotypes with

a homozygous ancestral core genotype. Clusters of extreme scores in both directions indicate

86 evidence for a sweep.

87 2.3 XP-EHH and XP-nSL

Unphased XP-EHH and XP-nSL are calculated by comparing the iHH between populations *A* and *B*, using the entire sample in each population. iHH in a population P is computed as

91
$$iHH_{P} = \sum_{i=1}^{|\mathcal{D}|} \frac{1}{2} (EHH(x_{i-1}) + EHH(x_{i})) d(x_{i-1}, x_{i}) + \sum_{i=1}^{|\mathcal{U}|} \frac{1}{2} (EHH(x_{i-1}) + EHH(x_{i})) d(x_{i-1}, x_{i}),$$

where the distance measure is given as centimorgans or basepairs for XP-EHH (Sabeti, et al. 2007) and number of sites observed for XP-nSL (Szpiech, et al. 2021). The XP statistics between population *A* and *B* are then computed as $XP = \log_{10} \left(\frac{iHH_A}{iHH_B}\right)$ and are normalized genome wide in a single bin. Large positive scores indicate long high-frequency haplotypes in 96 population *A*, and large negative scores indicate long high-frequency haplotypes in population

97 *B*. Clusters of extreme scores in one direction indicate evidence for a sweep in that population.

98 **3 Methods**

99 3.1 Simulations

100 We evaluate the performance of the unphased versions of iHS, nSL, XP-EHH, and XP-101 nSL under a generic two-population divergence model using the coalescent simulation program 102 discoal (Kern and Schrider 2016). We explore five versions of this generic model and name 103 them Demo 1 through Demo 5 (Table 1). Let N_0 and N_1 be the effective population sizes of 104 Population 0 and Population 1 after the split from their ancestral population (of size N_A). For 105 Demo 1, we keep a constant population size post-split and let $N_0 = N_1 = 10,000$. For Demo 2, 106 we keep a constant population size post-split and let $N_0 = 2N_1 = 10,000$. For Demo 3, we keep 107 a constant population size post-split and let $2N_0 = N_1 = 10,000$. For Demo 4, we initially set 108 $N_0 = N_1 = 10,000$ and let N_0 grow stepwise exponentially every 50 generations starting at 2,000 109 generations ago until $N_0 = 5N_1 = 50,000$. For Demo 5, we initially set $N_0 = N_1 = 10,000$ and let 110 N_1 grow stepwise exponentially every 50 generations starting at 2,000 generations ago until 111 $5N_0 = N_1 = 50,000.$

112 For each demographic history we vary the population divergence time $t_d \in$ 113 {2000, 4000, 8000} generations ago. For non-neutral simulations, we simulate a sweep in 114 Population 0 in the middle of the simulated region across a range of selection coefficients $s \in$ 115 $\{0.005, 0.01, 0.02\}$. We vary the frequency at which the adaptive allele starts sweeping as $e \in$ 116 $\{0, 0.01, 0.02, 0.05, 0.10\}$, where e = 0 indicates a hard sweep and e > 0 indicates a soft sweep, 117 and we also vary the frequency of the selected allele at time of sampling $f \in \{0.7, 0.8, 0.9, 1.0\}$ 118 as well as $g \in \{50, 100\}$ representing fixation of the sweeping allele g generations ago. For all 119 simulations we set the genome length to be L = 500,000 basepairs, the ancestral effective population size to be $N_A = 10,000$, the per site per generation mutation rate at $\mu = 2.35 \times 10^{-8}$, 120

121 and the per site per generation recombination rate at $r = 1.2 \times 10^{-8}$. For neutral simulations, we 122 simulate 1,000 replicates for each parameter set, and for non-neutral simulations we simulate 123 100 replicates for each parameter set. As iHS and nSL are single population statistics, we only 124 analyze Demo 1. Demo 3. and Demo 4 with these statistics, as Demo 2 and Demo 5 have a 125 constant size history identical to Demo 1 for Population 0, where the sweeps are simulated. 126 For all simulations, we compute the relevant statistics (--ihs, --nsl, --xpehh, or --xpnsl) 127 with selscan v2.0, using the --unphased and --trunc-ok flags. For iHS and XP-EHH, we also use 128 the --pmap flag in order to use physical distance instead of a recombination map.

129 **3.2 Power and False Positive Rate**

130 To compute power for iHS and nSL, we follow the approach of Voight et al. (2006). For 131 these statistics, each non-neutral replicate is individually normalized jointly with all matching 132 neutral replicates in 1% allele frequency bins. Because extreme values of the statistic are likely 133 to be clustered along the genome (Voight, et al. 2006), we then compute the proportion of 134 extreme scores (|iHS| > 2 or |nSL| > 2) within 100kbp non-overlapping windows. We then bin 135 these windows into 10 quantile bins based on the number of scores observed in each window 136 and call the top 1% of these windows as putatively under selection. We calculate the proportion 137 of non-neutral replicates that fall in this top 1% as the power. To compute the false positive rate, 138 we compute the proportion of neutral simulations that fall within the top 1%.

To compute power for XP-EHH and XP-nSL, we follow the approach of Szpiech et al. (2021). For these statistics, each non-neutral replicate is individually normalized jointly with all matching neutral replicates. Because extreme values of the statistic are likely to be clustered along the genome (Szpiech, et al. 2021), we then compute the proportion of extreme scores (XP-EHH > 2 or XP-nSL > 2) within 100kbp non-overlapping windows. We then bin these windows into 10 quantile bins based on the number of scores observed in each window and call the top 1% of these windows as putatively under selection. We calculate the proportion of non-

neutral replicates that fall in this top 1% as the power. To compute the false positive rate, we

147 compute the proportion of neutral simulations that fall within the top 1%.

148 **4 Results**

We find that the unphased versions of iHS and nSL have good power (Figures 1, S1-S4,
S13-16, and S21-24) to detect selection prior to fixation of the allele, with nSL generally
outperforming iHS. In smaller populations (Figure 1C and 1D), power does suffer relative to
larger populations (Figure 1A, 1B, 1E, 1F). Each of these statistics also have low false positive
rates hovering around 1% (Table S1).

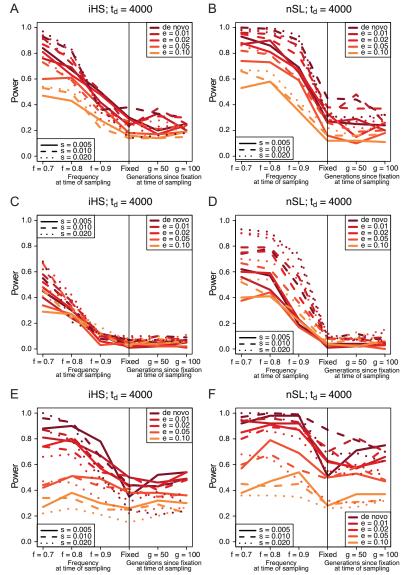
154 Similarly, we find that the unphased versions of XP-EHH and XP-nSL have good power 155 as well (Figures 2, 3, S5-S12, S17-S20, and S25-S32). When the sweep takes place in the 156 smaller of the two populations (Figure 2C and 2D), we see a similar decrease in power. When 157 one population is undergoing exponential growth (Figure 3) performance is generally guite 158 good, likely the result of a larger effective selection coefficient in large populations. These two-159 population statistics generally outperform their single-population counterparts, especially for 160 sweeps that have reached fixation recently. Each of these statistics also have low false positive 161 rates hovering around 1% (Table S1).

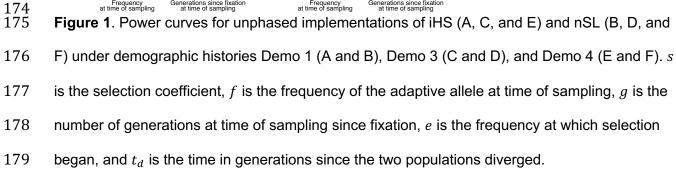
162 **5 Discussion**

We introduce multi-locus genotype versions of four popular haplotype-based selection statistics—iHS (Voight, et al. 2006), nSL (Ferrer-Admetlla, et al. 2014), XP-EHH (Sabeti, et al. 2007), and XP-nSL (Szpiech, et al. 2021)—that can be used when the phase of genotypes is unknown. We implement these updates in the latest v2.0 update of the program selscan (Szpiech and Hernandez 2014), with source code and pre-compiled binaries available at https://www.github.com/szpiech/selscan.

169 **6 Acknowledgements**

- 170 This work was supported by start-up funds from the Pennsylvania State University's
- 171 Department of Biology. Computations for this research were performed using the Pennsylvania
- 172 State University's Institute for Computational Data Sciences' Roar supercomputer.
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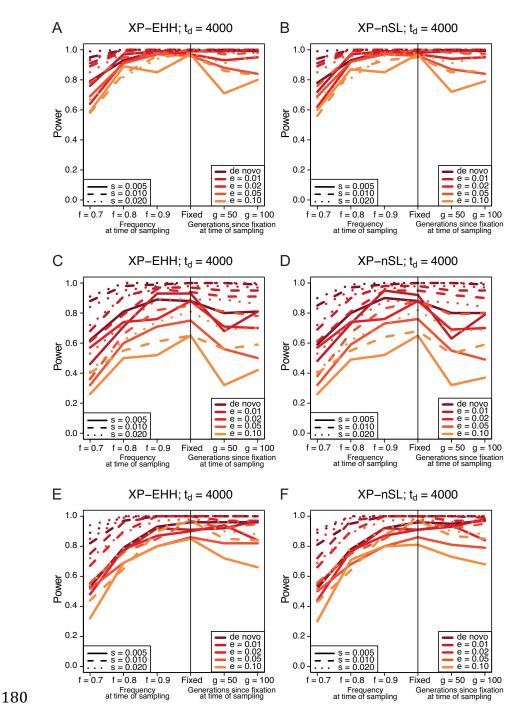


Figure 2. Power curves for unphased implementations of XP-EHH (A, C, and E) and XP-nSL (B, D, and F) under demographic histories Demo 1 (A and B), Demo 2 (C and D), and Demo 3 (E and F). *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

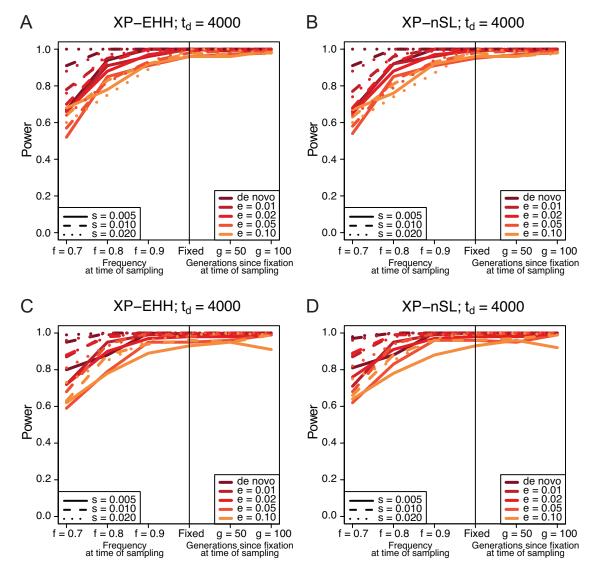
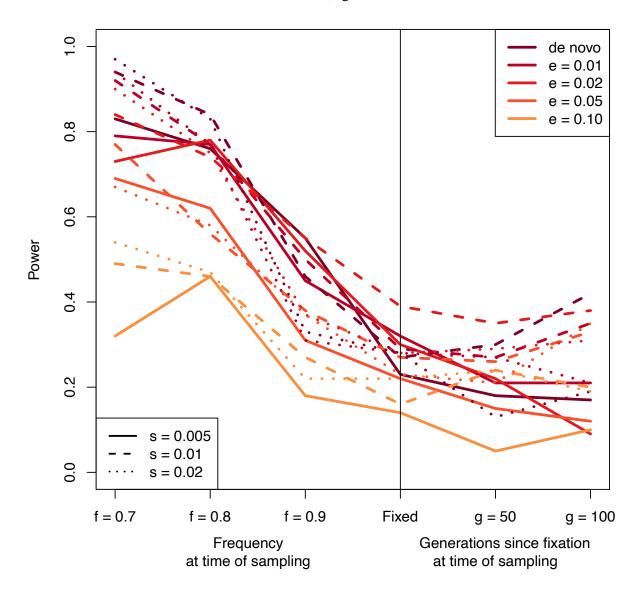


Figure 3. Power curves for unphased implementations of XP-EHH (A and C) and XP-nSL (B and D) under demographic histories Demo 4 (A and B), and Demo 5 (C and D). *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

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iHS; t_d = 2000



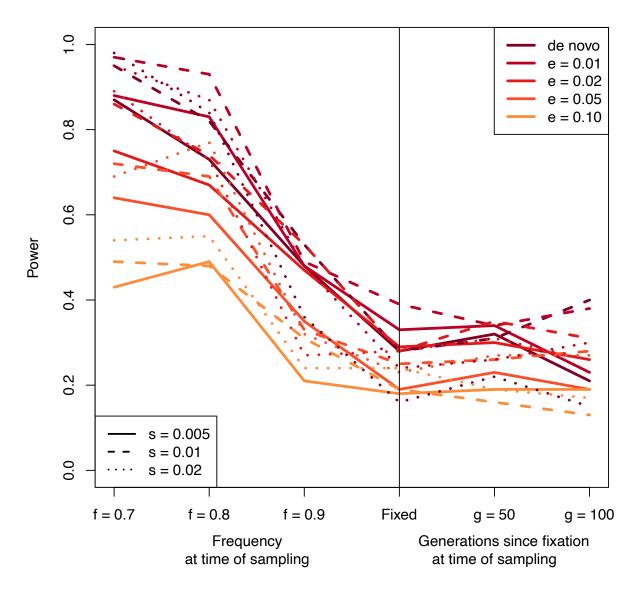


197 **Figure S1**. Demo 1 iHS $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the

198 frequency of the adaptive allele at time of sampling, g is the number of generations at time of

sampling since fixation, e is the frequency at which selection began, and t_d is the time in

iHS; t_d = 8000



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Figure S2. Demo 1 iHS $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the

frequency of the adaptive allele at time of sampling, g is the number of generations at time of sampling since fixation, e is the frequency at which selection began, and t_d is the time in

 $nSL; t_d = 2000$

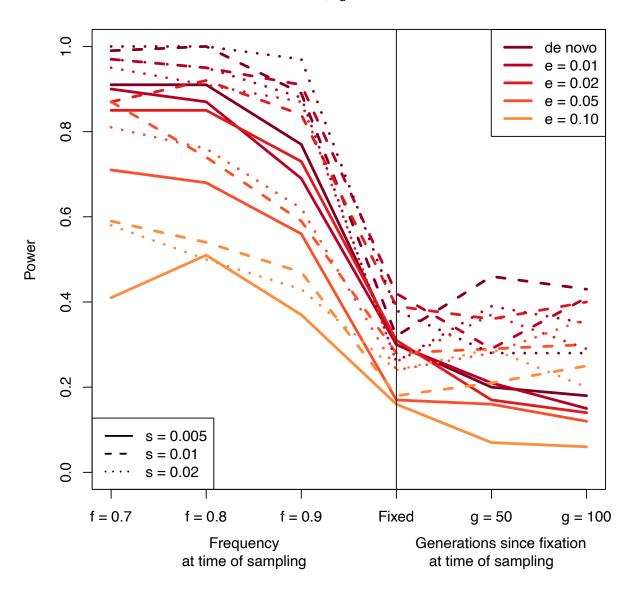


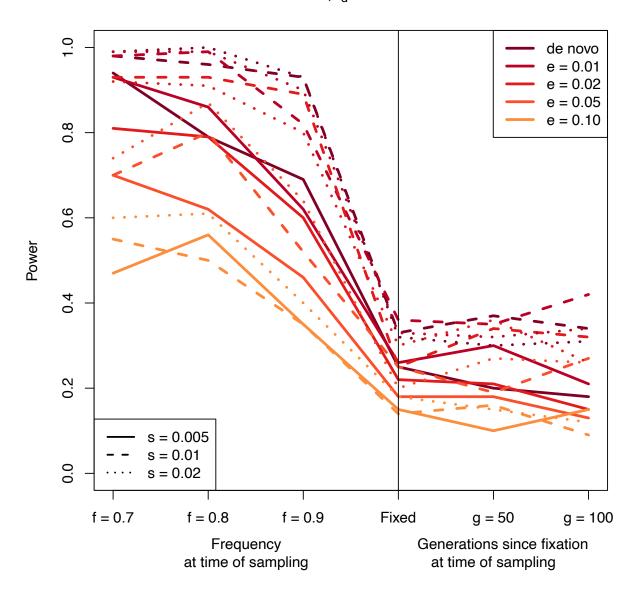


Figure S3. Demo 1 nSL $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the

208 frequency of the adaptive allele at time of sampling, g is the number of generations at time of

sampling since fixation, e is the frequency at which selection began, and t_d is the time in

nSL; t_d = 8000



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Figure S4. Demo 1 nSL $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the

213 frequency of the adaptive allele at time of sampling, g is the number of generations at time of

sampling since fixation, e is the frequency at which selection began, and t_d is the time in

 $XP-EHH; t_d = 2000$

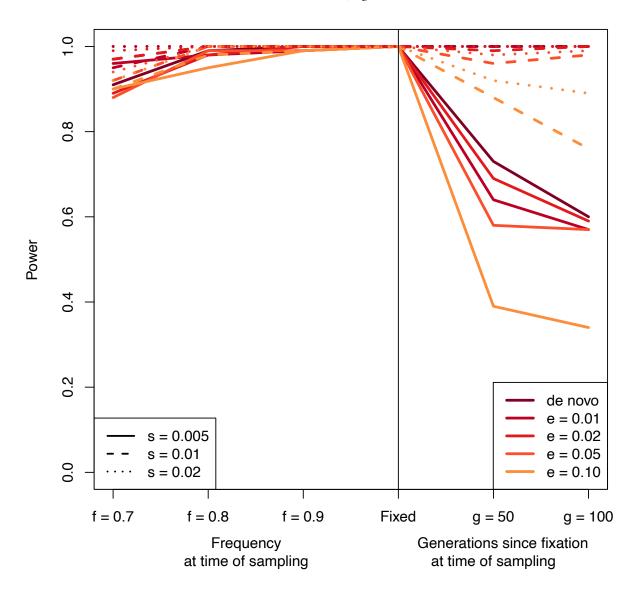


Figure S5. Demo 1 XP-EHH $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

 $XP-EHH; t_d = 8000$

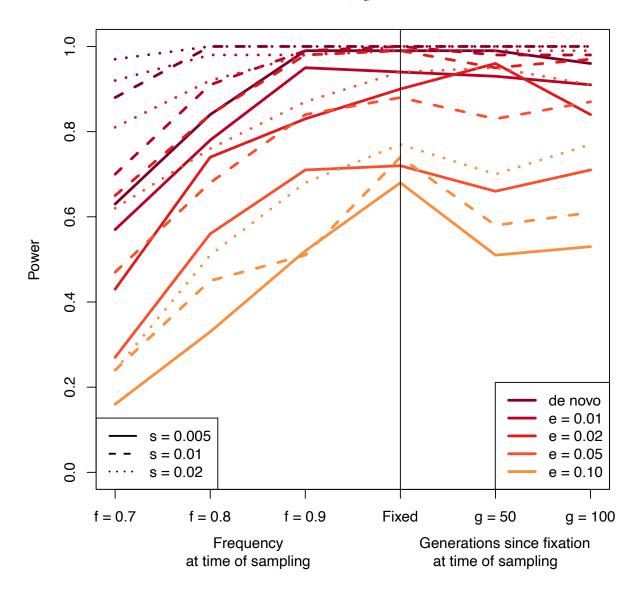


Figure S6. Demo 1 XP-EHH t_d = 8000 power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

XP–nSL; $t_d = 2000$

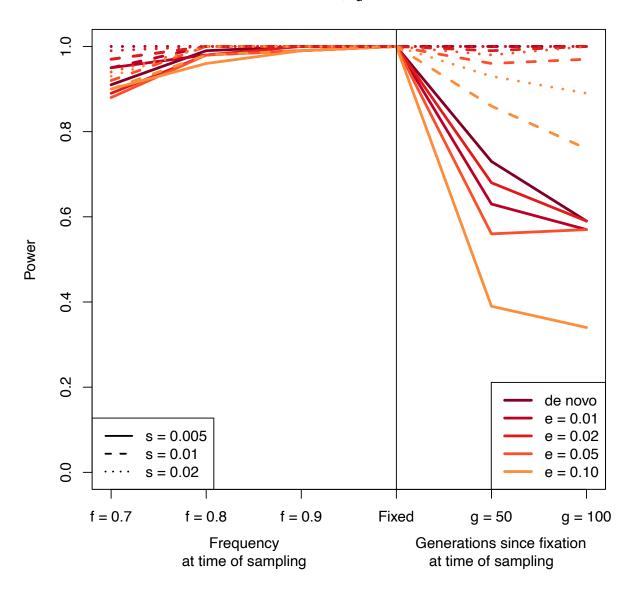


Figure S7. Demo 1 XP-nSL $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

 $XP-nSL; t_d = 8000$

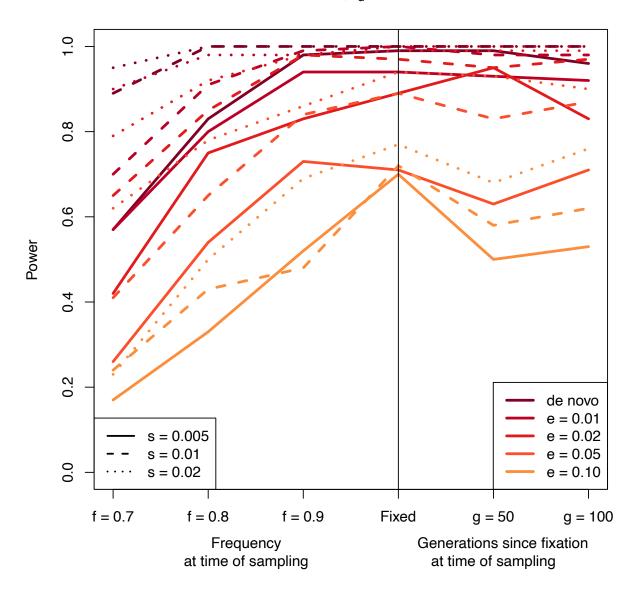


Figure S8. Demo 1 XP-nSL $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

 $XP-EHH; t_d = 2000$

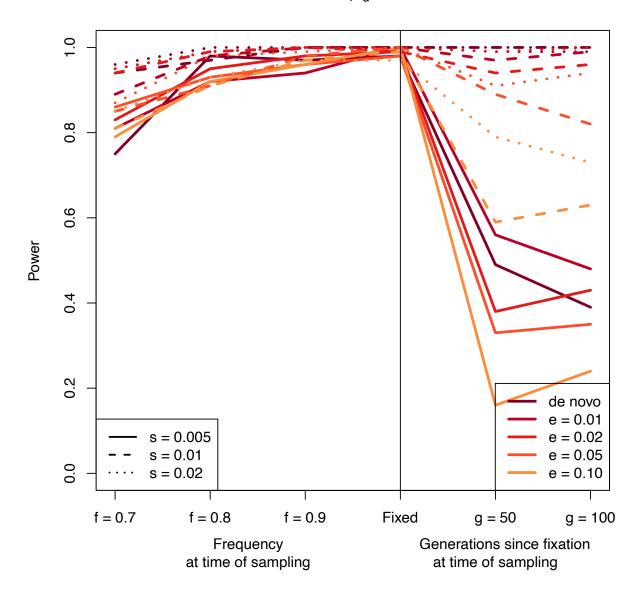


Figure S9. Demo 2 XP-EHH $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

 $XP-EHH; t_d = 8000$

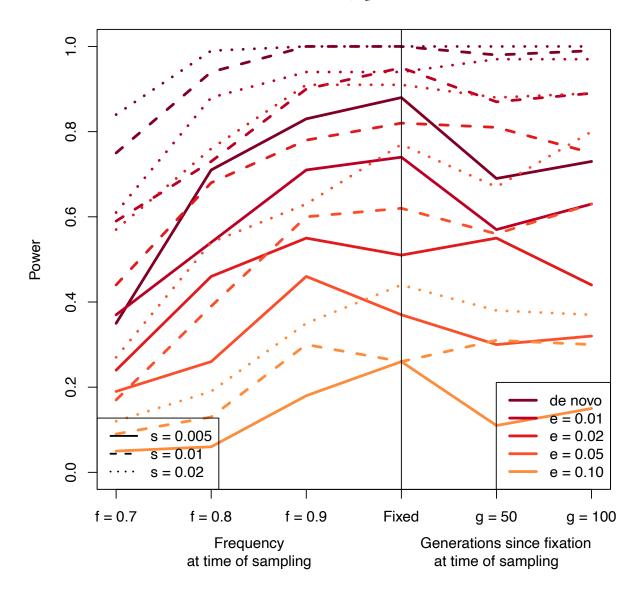




Figure S10. Demo 2 XP-EHH $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

XP–nSL; $t_d = 2000$

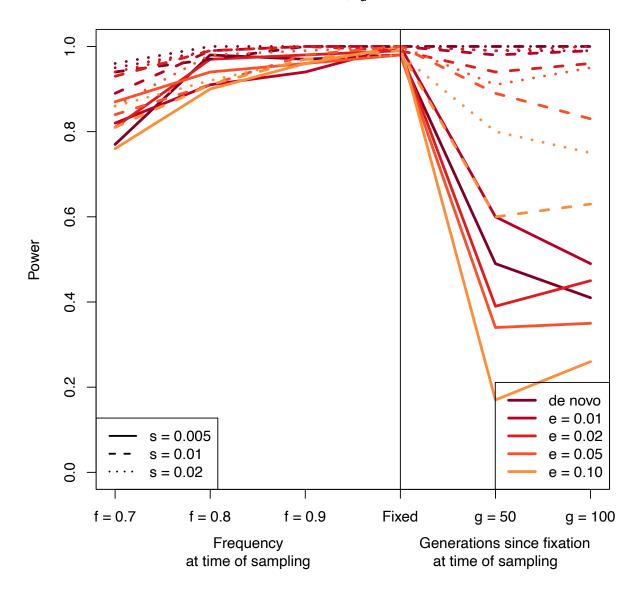


Figure S11. Demo 2 XP-nSL $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

 $XP-nSL; t_d = 8000$

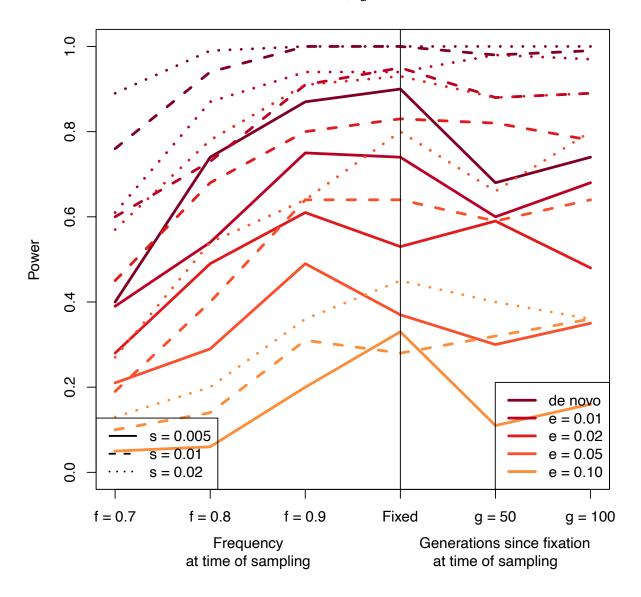
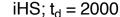


Figure S12. Demo 2 XP-nSL $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.



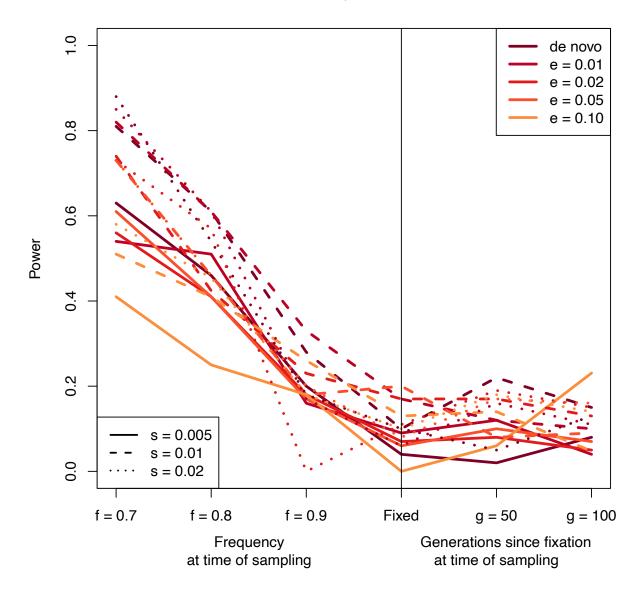
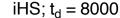




Figure S13. Demo 3 iHS $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.



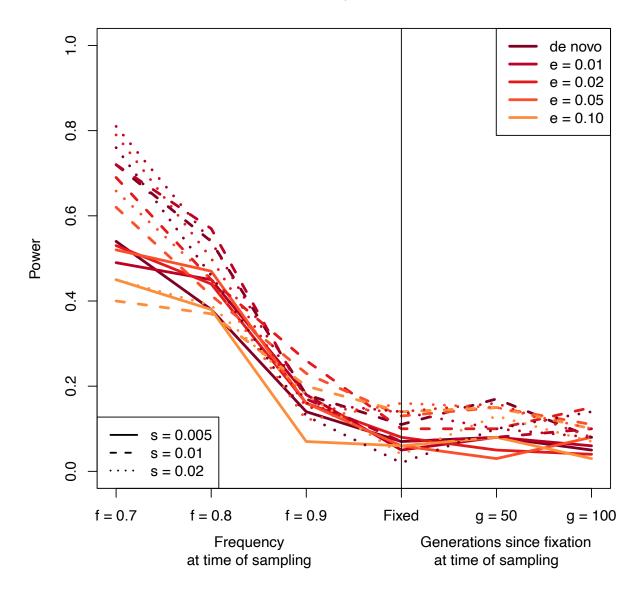
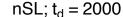
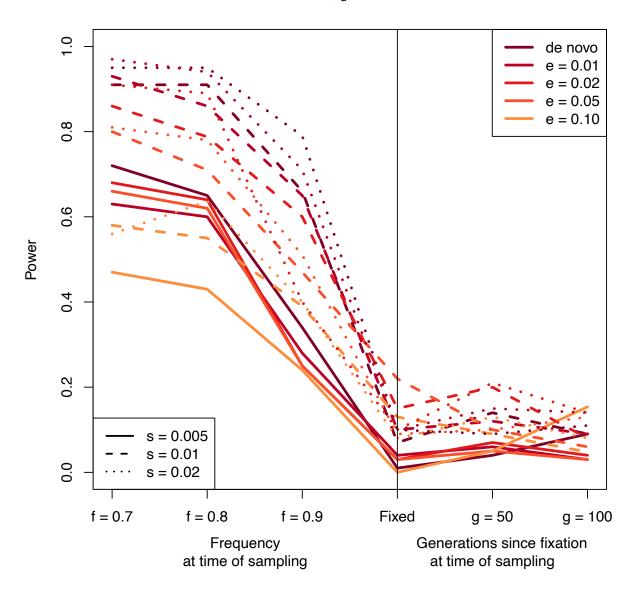


Figure S14. Demo 3 iHS $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.





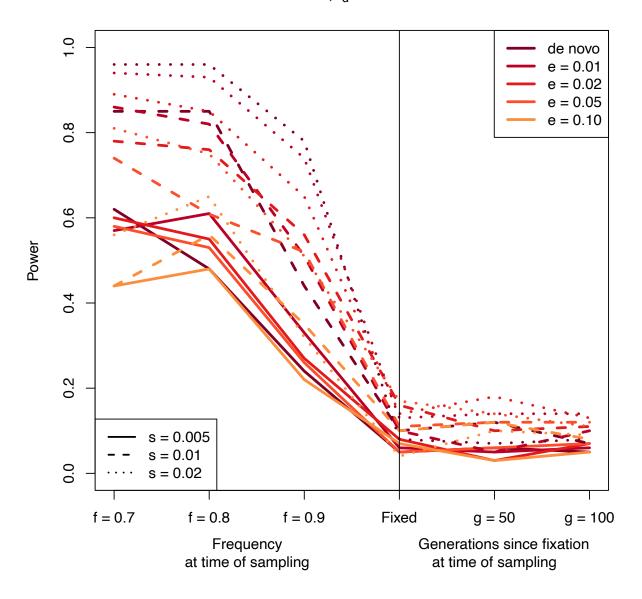
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Figure S15. Demo 3 nSL $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the

frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of

sampling since fixation, e is the frequency at which selection began, and t_d is the time in

nSL; t_d = 8000



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Figure S16. Demo 3 nSL $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the

273 frequency of the adaptive allele at time of sampling, g is the number of generations at time of

sampling since fixation, e is the frequency at which selection began, and t_d is the time in

 $XP-EHH; t_d = 2000$

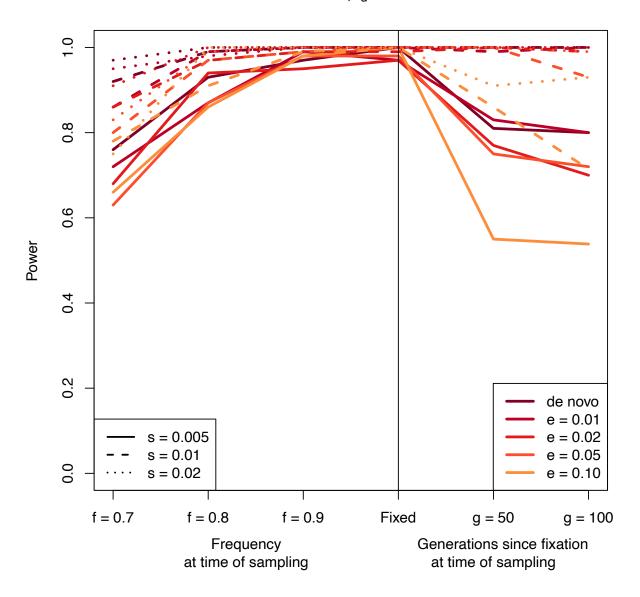


Figure S17. Demo 3 XP-EHH $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

 $XP-EHH; t_d = 8000$

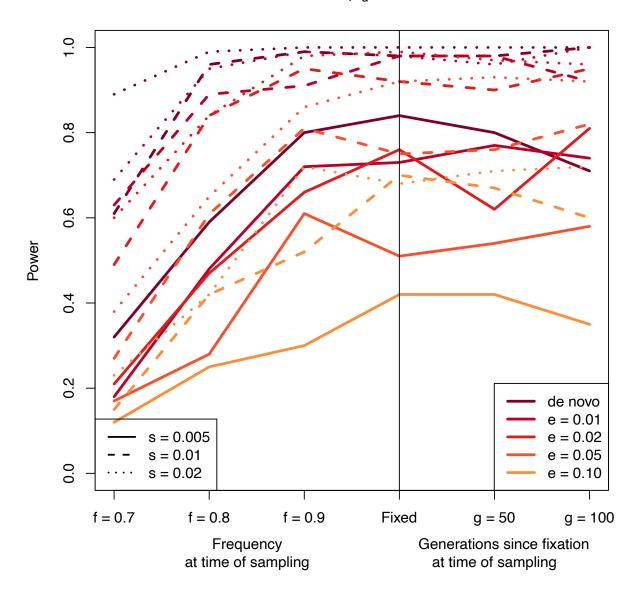


Figure S18. Demo 3 XP-EHH $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

XP–nSL; $t_d = 2000$

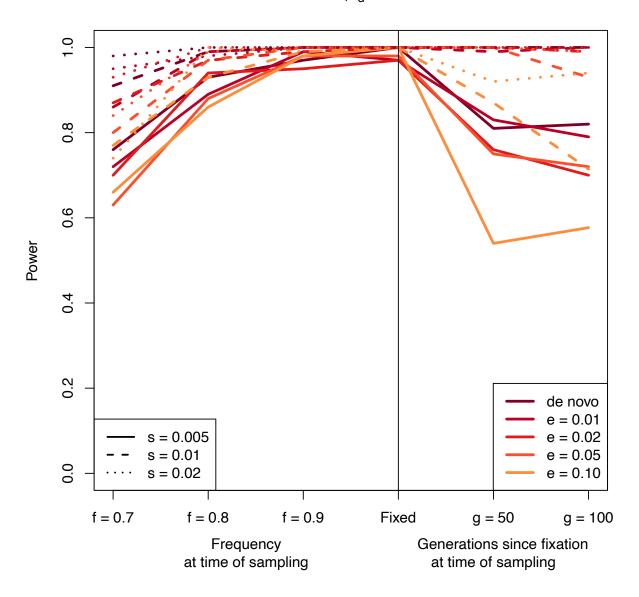


Figure S19. Demo 3 XP-nSL $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

XP–nSL; $t_d = 8000$

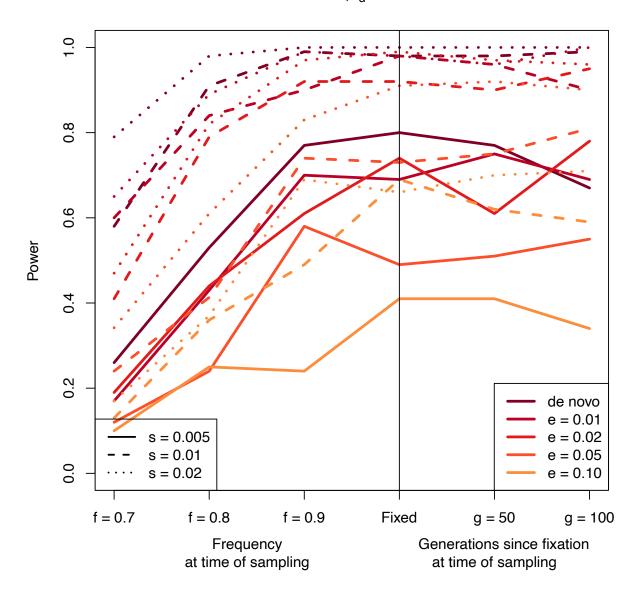
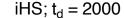


Figure S20. Demo 3 XP-nSL $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.



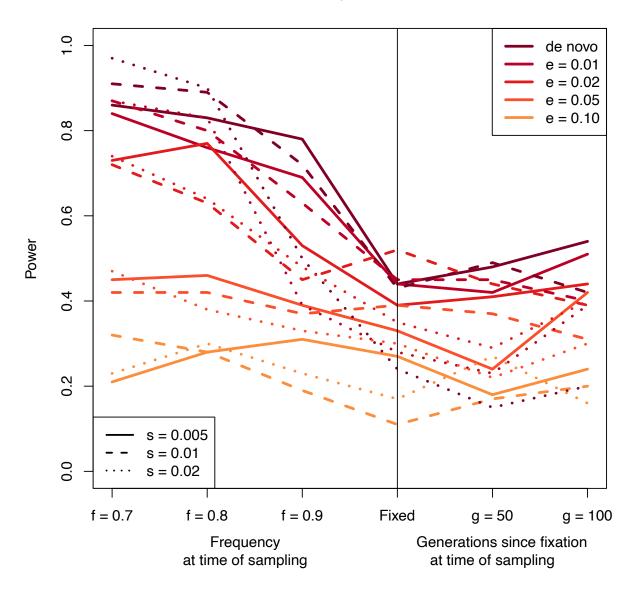




Figure S21. Demo 4 iHS $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

iHS; t_d = 8000

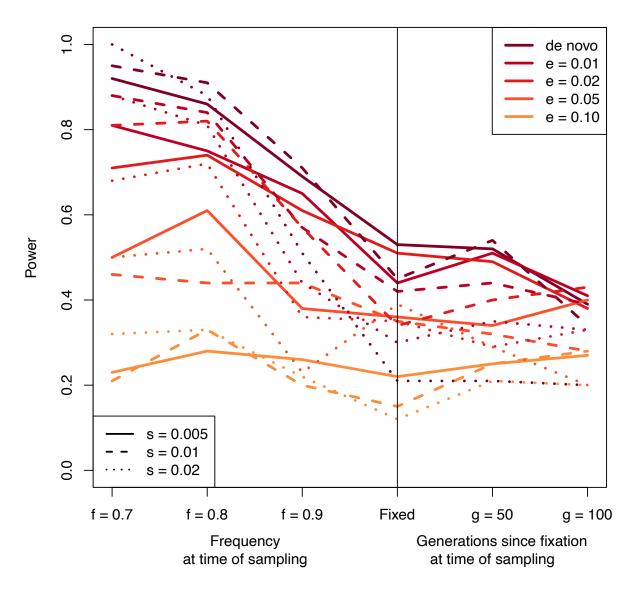
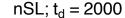
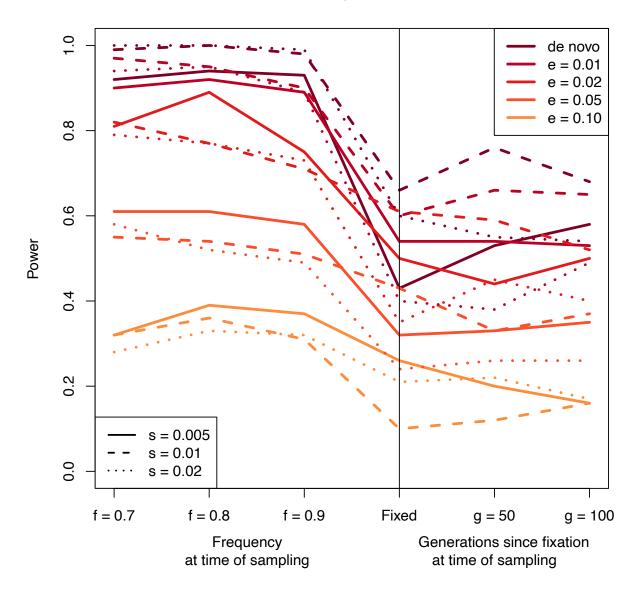




Figure S22. Demo 4 iHS $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.





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Figure S23. Demo 4 nSL $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the

308 frequency of the adaptive allele at time of sampling, g is the number of generations at time of

309 sampling since fixation, e is the frequency at which selection began, and t_d is the time in

nSL; t_d = 8000

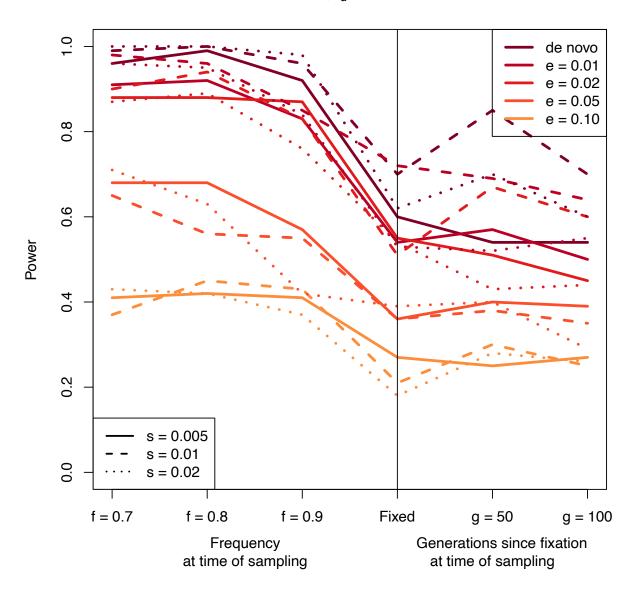




Figure S24. Demo 4 nSL $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the

313 frequency of the adaptive allele at time of sampling, g is the number of generations at time of

314 sampling since fixation, e is the frequency at which selection began, and t_d is the time in

 $XP-EHH; t_d = 2000$

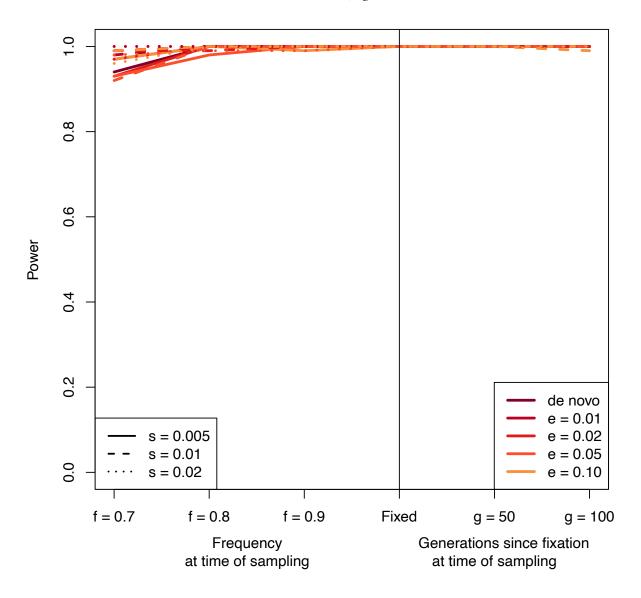


Figure S25. Demo 4 XP-EHH $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

XP-EHH; t_d = 8000

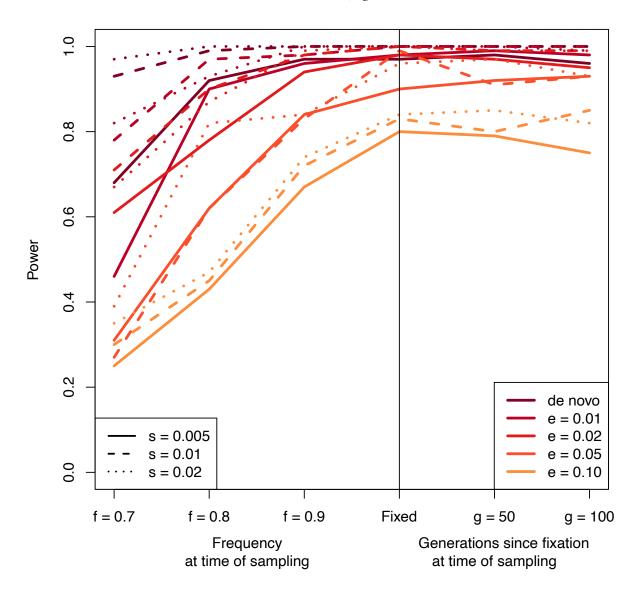
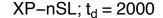


Figure S26. Demo 4 XP-EHH $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.



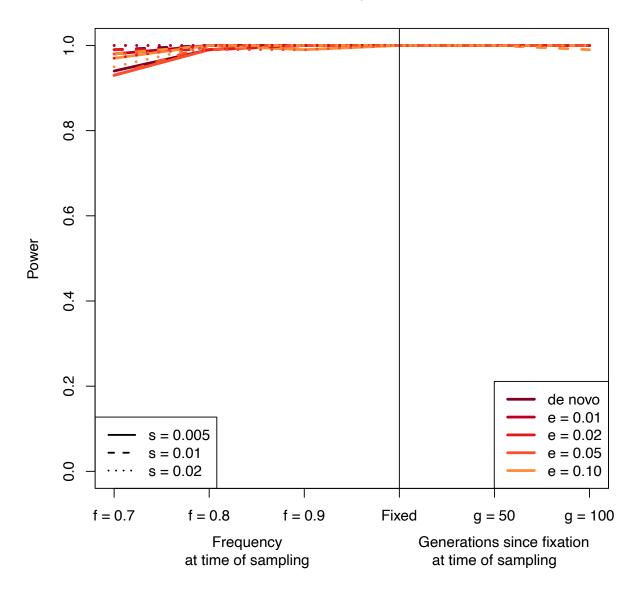


Figure S27. Demo 4 XP-nSL $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

XP–nSL; $t_d = 8000$

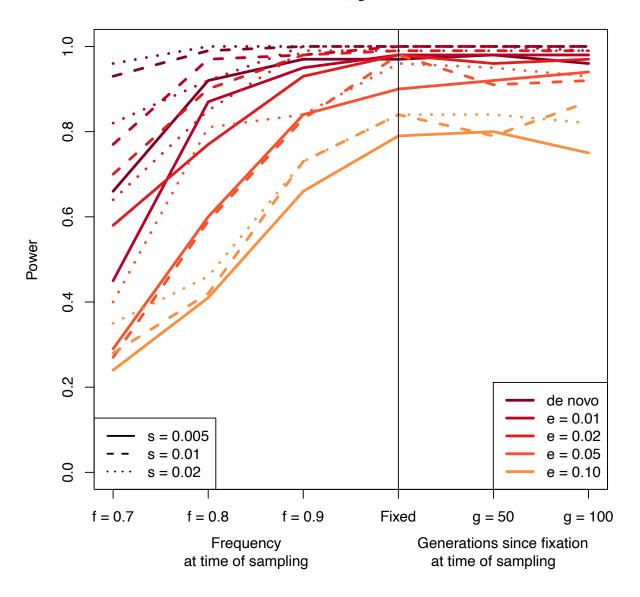


Figure S28. Demo 4 XP-nSL $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

 $XP-EHH; t_d = 2000$

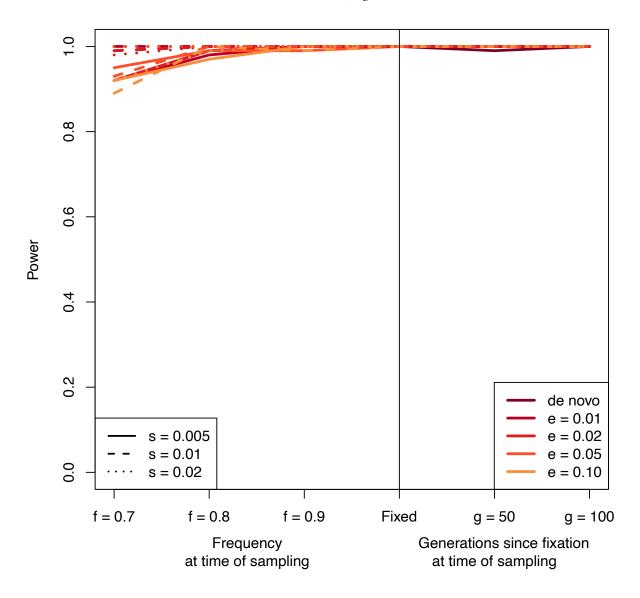


Figure S29. Demo 5 XP-EHH $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

 $XP-EHH; t_d = 8000$

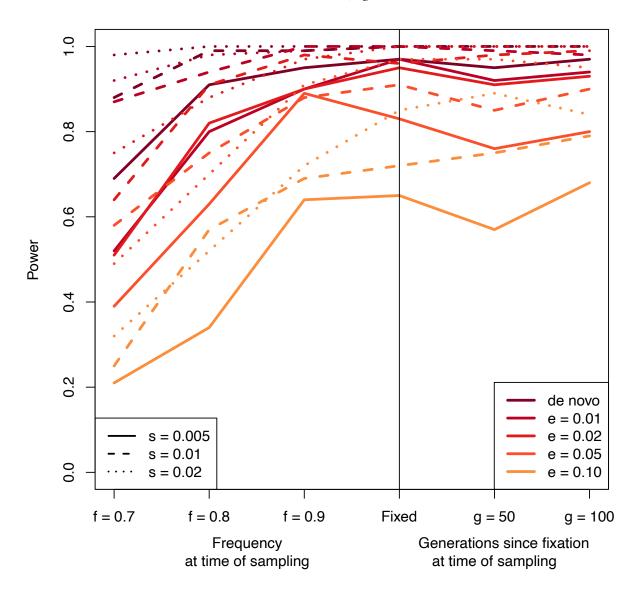


Figure S30. Demo 5 XP-EHH $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

XP–nSL; $t_d = 2000$

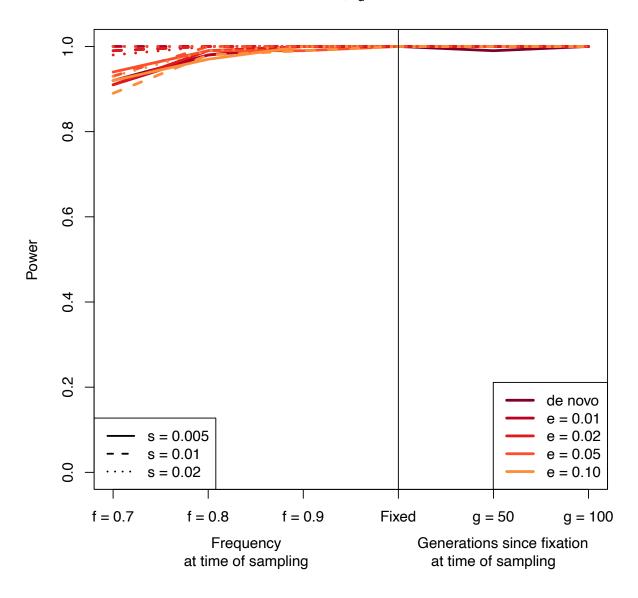


Figure S31. Demo 5 XP-nSL $t_d = 2000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

 $XP-nSL; t_d = 8000$

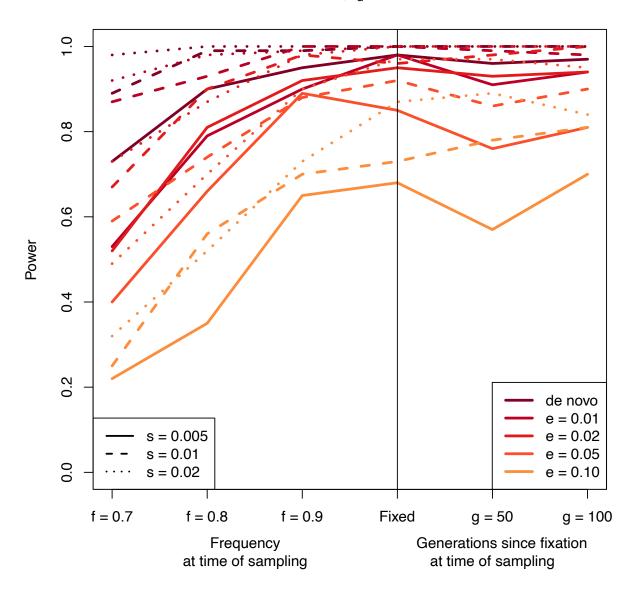




Figure S32. Demo 5 XP-nSL $t_d = 8000$ power curves. *s* is the selection coefficient, *f* is the frequency of the adaptive allele at time of sampling, *g* is the number of generations at time of sampling since fixation, *e* is the frequency at which selection began, and t_d is the time in generations since the two populations diverged.

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 Table 1. Demographic history parameters for simulations.

	N _A	N ₀ at split	N ₀ at present	N ₁ at split	N ₁ at present	t_d				
Demo 1	10,000	10,000	10,000	10,000	10,000	2,000/4,000/8,000				
Demo 2	10,000	10,000	10,000	5,000	5,000	2,000/4,000/8,000				
Demo 3	10,000	5,000	5,000	10,000	10,000	2,000/4,000/8,000				
Demo 4	10,000	10,000	50,000†	10,000	10,000	2,000/4,000/8,000				
Demo 5	10,000	10,000	10,000	10,000	50,000†	2,000/4,000/8,000				

362 [†]The reached via exponential growth starting 2,000 generations ago.

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364 **Table S1**. False positive rate computed from neutral simulations for varying t_d and demographic history.

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listory.							
		$t_d = 2000$	$t_d = 4000$	$t_d = 8000$			
	Demo 1	0.013	0.1	0.009			
iHS	Demo 3	0.007	0.013	0.007			
	Demo 4	0.015	0.018	0.008			
	Demo 1	0.01	0.015	0.008			
nSL	Demo 3	0.008	0.011	0.007			
	Demo 4	0.014	0.021	0.014			
	Demo 1	0.013	0.013	0.016			
	Demo 2	0.017	0.009	0.015			
XP-EHH	Demo 3	0.01	0.011	0.012			
	Demo 4	0.012	0.014	0.014			
	Demo 5	0.011	0.012	0.013			
	Demo 1	0.014	0.011	0.013			
	Demo 2	0.019	0.011	0.012			
XP-nSL	Demo 3	0.011	0.011	0.012			
	Demo 4	0.012	0.012	0.014			
	Demo 5	0.011	0.012	0.014			

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367 References 368

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370 Colonna V, Ayub Q, Chen Y, Pagani L, Luisi P, Pybus M, Garrison E, Xue Y, Tyler-Smith C,

- Genomes Project C, et al. 2014. Human genomic regions with exceptionally high levels 371 372 of population differentiation identified from 911 whole-genome sequences. Genome 373 Biol 15:R88.
- 374 Crawford NG, Kelly DE, Hansen MEB, Beltrame MH, Fan S, Bowman SL, Jewett E, Ranciaro A, 375 Thompson S, Lo Y, et al. 2017. Loci associated with skin pigmentation identified in
- 376 African populations. Science 358.
- 377 DeGiorgio M, Szpiech ZA. 2021. A spatially aware likelihood test to detect sweeps from 378 haplotype distributions. bioRxiv:2021.2005.2012.443825.
- 379 Ferrer-Admetlla A, Liang M, Korneliussen T, Nielsen R. 2014. On detecting incomplete soft 380 or hard selective sweeps using haplotype structure. Mol Biol Evol 31:1275-1291.
- 381 Harris AM, DeGiorgio M. 2020. A likelihood approach for uncovering selective sweep 382 signatures from haplotype data. Mol Biol Evol.

Harris AM, Garud NR, DeGiorgio M. 2018. Detection and Classification of Hard and Soft
 Sweeps from Unphased Genotypes by Multilocus Genotype Identity. Genetics 210:1429 1452.

- Kern AD, Schrider DR. 2016. Discoal: flexible coalescent simulations with selection.
 Bioinformatics 32:3839-3841.
- Lu K, Wei L, Li X, Wang Y, Wu J, Liu M, Zhang C, Chen Z, Xiao Z, Jian H, et al. 2019. Wholegenome resequencing reveals Brassica napus origin and genetic loci involved in its
 improvement. Nat Commun 10:1154.
- Meier JI, Marques DA, Wagner CE, Excoffier L, Seehausen O. 2018. Genomics of Parallel
 Ecological Speciation in Lake Victoria Cichlids. Mol Biol Evol 35:1489-1506.
- Nedelec Y, Sanz J, Baharian G, Szpiech ZA, Pacis A, Dumaine A, Grenier JC, Freiman A, Sams
 AJ, Hebert S, et al. 2016. Genetic Ancestry and Natural Selection Drive Population
 Differences in Immune Responses to Pathogens. Cell 167:657-669 e621.
- Sabeti PC, Varilly P, Fry B, Lohmueller J, Hostetter E, Cotsapas C, Xie X, Byrne EH, McCarroll
 SA, Gaudet R, et al. 2007. Genome-wide detection and characterization of positive
- 398 selection in human populations. Nature 449:913-918.
- Salmon P, Jacobs A, Ahren D, Biard C, Dingemanse NJ, Dominoni DM, Helm B, Lundberg M,
 Senar JC, Sprau P, et al. 2021. Continent-wide genomic signatures of adaptation to
 urbanisation in a songbird across Europe. Nat Commun 12:2983.
- 402 Szpiech ZA, Hernandez RD. 2014. selscan: an efficient multithreaded program to perform
 403 EHH-based scans for positive selection. Mol Biol Evol 31:2824-2827.
- 404 Szpiech ZA, Novak TE, Bailey NP, Stevison LS. 2021. Application of a novel haplotype-based
 405 scan for local adaptation to study high-altitude adaptation in rhesus macaques. Evol
 406 Lett 5:408-421.
- 407 Voight BF, Kudaravalli S, Wen X, Pritchard JK. 2006. A map of recent positive selection in
 408 the human genome. Plos Biology 4:e72.
- 409 Zhang SJ, Wang GD, Ma P, Zhang LL, Yin TT, Liu YH, Otecko NO, Wang M, Ma YP, Wang L, et
- 410 al. 2020. Genomic regions under selection in the feralization of the dingoes. Nat411 Commun 11:671.
- 412 Zoledziewska M, Sidore C, Chiang CWK, Sanna S, Mulas A, Steri M, Busonero F, Marcus JH,
- 413 Marongiu M, Maschio A, et al. 2015. Height-reducing variants and selection for short
 414 stature in Sardinia. Nat Genet 47:1352-1356.
- 415