

1 The natural history of the black soldier fly,
2 *Hermetia illucens*: insights from complete
3 mitochondrial genome sequences.

4 Authors : Guilliet J*, Baudouin G, Pollet N, Filée J*

5 address : guilliet@egce.cnrs-gif.fr jonathan.filee@egce.cnrs-gif.fr- Laboratoire Evolution, Génomes

6 Comportement, Ecologie CNRS Université Paris Sud UMR 9191, IRD UMR 247 CNRS Avenue de la Terrasse, Bâtiment 13, Boite Postale 1

7 91198 Gif sur Yvette France

8 **Abstract**

9 **The Black Soldier Fly (BSF) *Hermetia illucens* is a cosmopolitan fly heavily used by industrial**
10 **companies to reduce biowaste and produce protein and fat for poultry and aquaculture feed.**
11 **However, the natural history and the genetic diversity of the BSF is poorly known. In this study, we**
12 **analysed 677 CO1 sequences derived from samples found all over the five continents leading us to**
13 **the discovery of 52 haplotypes including 10 major haplotypes. We refined the definition of these**
14 **haplotypes by sequencing 59 mitochondrial genomes. We could derive an estimate of the separation**
15 **events of the different haplotypes at more than two million years for the oldest branches. This**
16 **worldwide cryptic genetic and genomic diversity is mirrored at local scale in France in which we**
17 **found five major haplotypes sometimes in sympatry. Our data resolve the phylogenetic relationships**
18 **between the major lineages and give insights into the dispersal and the numbers of BSF neo-**
19 **introduction at global and local scales. Our results indicate that the genetic and genomic diversity of**
20 **commercial BSF stock is very low and these brood stock participated in the dissemination of the BSF**
21 **in the wild. Taken together these results call for a better understanding of the genomic diversity of**

22 **the BSF to unravel possible specific adaptations of the different lineages for industrial needs and to**
23 **initiate the selection process.**

24

25 Keywords : *Hermetia illucens* ; phylogeny ; mitochondrial genome ; phylogeography

26 Introduction

27 The black soldier fly (BSF) *Hermetia illucens* (Linnaeus, 1758) is a Diptera of the Stratiomyidae family.
28 It probably originated from Mexico, then subsequently spread to the south of the USA and across Latin
29 America over the last thousand years (Hauser et al. 2015). These geographical areas are now
30 considered as its natural area (Rozkosný, 1983, Hauser et al. 2015). The BSF is now a cosmopolitan fly
31 that can be found in tropical, subtropical, and tempered regions, from the 40th parallel North and the
32 45th South (Rozkošný 1983 ; Mason et al, 2009) where it is considered as an exotic, non-invasive
33 species (<https://tinyurl.com/5353hvjv>, Wang and Shelomi, 2017). This fly likely spread through
34 shipping routes (Picker et al, 2002). While studies based on naturalistic observations show that it was
35 first discovered in Malta in 1926 (Lindner, 1936), it has been suggested that it was introduced in Italy
36 about 500 years ago (Benelli et al, 2014). The BSF appeared in the early 1950s in the South-East of
37 France Barbier 1952 before spreading through the South-West, (Dauphin 2003) along the Rhône valley
38 (Richoux 2009) and finally along the Atlantic coast (Maquart et al, 2020). Currently, the BSF can be
39 found on the whole European territory with few exceptions. It is reasonable to assume that the rise of
40 international traffic and the industrial use of the BSF may have led to a steady flow of introductions in
41 France and all over the world (Leclerq 1969, Sheppard et al, 1994).

42 The BSF was initially used in forensic entomology to date the post-mortem time (Lord et al, 1994), now
43 it is heavily used by industrial companies to reduce biowastes (Gold et al, 2020 ; Hoc et al, 2020), to
44 produce protein (Hoc et al, 2020 ; Ewald et al, 2020 ;Giannetto et al, 2019) and fat (Hoc et al, 2020 ;
45 Ewald et al, 2020 ; Spranghers et al, 2017). These products are respectively used to make animal food
46 (Huis 2013) or biodiesel (Leong et al, 2016 ; Surendra et al, 2020).

47 BSF industries face similar issues as other farms: knowing how to master the biological and genetic
48 aspects of their model to get the best out of it. To this end, it is important to know the genetic diversity
49 of BSF existing internationally, nationally, and locally. A lot of articles exist about the nutritional aspect
50 of the BSF (Spranghers et al, 2017 ; Heuel et al, 2021 ; Schiavone et al, 2019 ; Neumann et al, 2018 ;
51 Biasato et al, 2019) or its microbiota (Khamis et al, 2020 ; Erickson et al, 2004 ; Liu et al, 2008), few
52 have focused on the genetic diversity of the BSF. We have some evidence on the extent of BSF genetic
53 diversity from previous studies. 10 haplotypes based on the CO1 gene were found in South Korea with
54 245 individuals (Park et al, 2017). Still on the CO1 gene, and on a worldwide sampling, 56 haplotypes
55 were found with a divergence rate up to 4.9% and it has been shown that individuals belonging to
56 divergent lineages are still interfertile (Ståhls et al 2020). Finally, based on 15 microsatellite markers,
57 16 genetic clusters were found with hot spots in South America (Kaya et al, 2021). Other works focused
58 on more fundamental aspects of BSF and led to the assembly of a reference mitochondrial genome as
59 well as a chromosomal-scale nuclear genome (Qi et al, 2017 ; Generalovic et al, 2020 ; Zhan et al, 2020).

60 Within the framework of the BSF industry, phylogeography can allow on the one side to account for
61 genetic diversity at different scales (global, national, and local) and on the other side, to have keys on
62 the geographical origin of the species present in a territory or used by companies. This study may
63 suggest local adaptations or some phenotypic plasticity, due to habituation to different biomes.

64 In this study, we quantified this diversity at different scales: worldwide by studying the variation within
65 the CO1 gene, and more locally at the level of France. We also analysed the role of livestock BSF
66 farming in the diversity present on the French territory. Our report is based on data obtained by
67 sequencing 113 individuals for the CO1 gene and also the mining of 564 CO1 sequences. We also
68 sequenced and assembled 59 mitochondrial genomes. We also show the great and hidden genetic
69 diversity within the species *Hermetia illucens* at all observation scales with a specific focus on France
70 (90 French individuals were analysed, allowing us to find 9 different haplotypes). In addition, we are

71 also providing here a date of the separation events of the different haplotypes found. This separation
72 turns out to be more than two million years old for the most distant haplotypes.

73 Materials and methods

74 Sampling

75 We decided to carry out a global sampling followed by a specific focus on France. A detailed table of
76 samples and associated metadata is provided in (SUPP_TABLE 001 (SUPP_MATERIAL)). The origin of all
77 CO1 sequences used in our study is summarized in Table 1 and a summary of sample numbers by
78 continent is provided in Table 2. Samples were collected during summer 2020 and stored in 100%
79 ethanol at -20°C prior to DNA extraction. In France, the collection was done with the help of the
80 compost citizen network (<https://reseaucompost.org/>). The larvae came exclusively from compost and
81 the adults collected were close to the composting area. Between one and 10 individuals (larvae or
82 adults) were collected per area. In some French localities, collections in the wild have been made near
83 BSF breeding areas (about five kilometres). For the other parts of the world, the individuals came either
84 from compost or from breeding farms.

85 We gathered all the CO1 sequences available on NCBI and Bold, as well as whole genome sequencing
86 data in SRA, and reconstructed the mitochondrial genome. In France, sampling was carried out during
87 the summer of 2020 in shared and personal compost sets. We obtained larvae in the areas of Blois,
88 Bordeaux, Toulouse, Montpellier, Lyon, Paris. In Africa, individuals from compost or livestock farms
89 were collected during 2019. In Latin America, larvae from French Guiana compost were collected
90 during the summer of 2019. An adult found in 2018 in Mexico. In Asia, individuals collected from
91 Taiwan either from a breeding farm or compost. These were mainly L5 larvae and pupae; some adults
92 were also recovered. We had the surprise in the Bayonne area to have BSF-like larvae, which turned
93 out to be *Exaireta spinigera* larvae.

94 DNA extraction

95 We extracted DNA from larvae, pupae and imago under the same conditions by grinding individuals in
96 liquid nitrogen using a mortar and pestle. The grinded samples were then processed according to the
97 protocol of the Macherey-Nagel AXG 100 (<https://tinyurl.com/46a7xejr>) kit for the isolation of
98 genomic DNA from tissue with some modification : we added a second purification step with 3.5 ml of
99 freshly prepared ethanol and centrifugation at 15,000 rpm at 4 ° C for 15 min just before the final re-
100 suspension. The DNA pellet was finally re-suspended in 100 µL of Tris 10 mM EDTA 1 mM pH 8.0. The
101 purity was evaluated by spectrophotometry using a Nanodrop (Desjardins and Conklin 2010), and the
102 concentration was quantified using the Qubit kit DNA Broad range (<https://tinyurl.com/evaah44c>).
103 DNA integrity was monitored using routine agarose gel electrophoresis.

104 CO1 Sequencing

105 We designed specific CO1 primers from the reference mitochondrial DNA (Qi et al, 2017) (SUPP_TABLE
106 002 (SUPP_MATERIAL)), then a mix was realized using NEB 5X standard buffer One Taq. The mix is then
107 amplified in ONETAQ HOT 45 PCR. We purified the PCR products using the Illustra Exostar 1-step GE
108 Healthcare kit before Sanger cycle sequencing. Cycle sequencing reactions were performed using the
109 BigDye 3.1 chemistry, purified by precipitation and sequenced using an ABI 3130 sequencer (Applied
110 Biosystems). Base calling was performed using the Sequencing analysis software version 5.3 (Applied
111 Biosystems). We assembled forward and reverse reads using the Geneious
112 (<https://www.geneious.com>) assembler with the sensitivity set at medium/fast. The regions
113 containing the primers were excised and we kept only the sequences having more than 75% of high
114 quality basecalls.

115

116 Multiple sequence alignment and haplotype network reconstruction

117 We aligned CO1 sequences obtained in the laboratory as well as those recovered from the databases
118 using the MAFFT (Katoh and Standley 2013) aligner included as a plug-in in the Geneious
119 (<https://www.geneious.com>) software. We used the algorithm G-INS-i with a score matrix 200 PAM /
120 k=2, a gap open penalty at 1.53 and offset value at 0.123. We exported the multiple sequence

121 alignment in nexus format ([Maddison et al, 1997](#)) for haplotype network reconstruction and
122 visualization using POPART ([Leigh and Bryant 2015](#)). We modified the output file to add geographical
123 origins to the sequences.

124 Shotgun sequencing and mitochondrial DNA assembly

125 A total of 56 BSF individuals were used for whole-genome shotgun sequencing using an Illumina
126 method by Novogene UK ([Cock et al, 2010](#) ; [Hansen et al, 2010](#) ; [Erlich et al, 2008](#) ; [Jiang et al, 2011](#) ;
127 [Yan et al, 2013](#)). The technique used is pair-end with a coverage rate of 4,68X (SD 0,50). The resulting
128 sequences were checked for quality and counted for read parity. We take advantage of the fact that
129 mtDNA is highly repeated in insect DNA. In insects, mtDNA represents on average 0.42% of the total
130 DNA in the genome sequence project ([Meng et al, 2019](#)). As stipulated, we used 5000 kb of
131 data/sample (4-5x coverage) for the mitochondrial assembly, which in average contains
132 $(5000 * 0,42) / 100 = 21000\text{kb}$ of mtDNA. As the mitochondrial genome is about 15 kb, we obtain a
133 coverage of 1400x for this genome which is largely sufficient to obtain a complete genome in one
134 contig. The sequences were assembled de novo with the MEGAHit ([Dinghua et al, 2015](#)) software
135 (based options); the mitochondrial genomes were thus recovered from the assemblies by blast
136 ([Altschul et al, 1990](#)) using the reference BSF mitochondrial genome sequence (NC_035232). Using
137 MITOS online software ([Bernt et al, 2013](#)) the mitochondrial DNAs were annotated, and the D-Loop
138 zone removed. To these sequences were added 4 complete mitochondrial sequences from databases
139 or reconstructed via read sequences available on the SRA ([Leinonen et al, 2011](#)) using the same
140 assembly procedure and then aligned under the same conditions as previously mentioned.

141 CO1 and whole mitochondrial genome phylogeny

142 We use the MEGAX ([Sudhir et al, 2018](#)) software to determine the best model and then compute a ML
143 Tree with 500 Bootstraps using a complete deletion parametre. We visualized the data with the online
144 tool ITOL ([Letunic and Bork, 2021](#)), and we rooted the tree with the Stratiomyidae *Exaireta spinigera*.
145 We chose *Exaireta spinigera* because it was the closest species for which we have a complete

146 mitochondrial genome (the blastn result of the complete mitochondrial genome of *Hermetia illucens*
147 vs *Exaireta spinigera* gives us 81.28% of similarity). To simplify the interpretation of the phylogeny, we
148 have kept only a single identical sequence per haplotype.

149 Time tree

150 We used the BEAST software (Suchard et al, 2018) to estimate the divergence time of the different BSF
151 groups. The analysis was performed with an expansion growth model and an uncorrelated lognormal
152 relaxed clock with the proposed insect molecular clock. 10 million iterations were performed and then
153 with the TreeAnnotator (Helfrich et al, 2018) the obtained trees were clustered with a burn of 25% of
154 the trees. The visualization of the final tree was done with the online software ITOL¹³.

155 Results

156 Haplotype Network

157 We reconstructed an haplotype network from the analysis of 677 sequences spanning 658 nucleotides
158 of the CO1 mitochondrial gene. A total of 52 distinct haplotypes were delineated from the analysis of
159 this CO1 fragment that were made of one to 230 fly samples (Figure 1).

160 We found 73 segregating sites with a Tajima-D of 45.1413 suggesting a lack of rare alleles, which could
161 come from a contraction of the population.

162 There were 31 haplotypes made of a single sequence. We observed 11 haplotypes made of between
163 two and four sequences derived from the same continent; that total of 42 haplotypes will be referred
164 to as minor haplotypes. We observed 10 haplotypes made of between five and 230 sequences
165 including 6 multi continental haplotypes that we will refer to as major haplotypes. In conclusion, we
166 found an important diversity both within and between continents using the CO1 phylogenetic
167 mitochondrial gene. Minor haplotypes represent 8.5% of all sequences (58 sequences) and 80.8% of
168 haplotypes. Major haplotypes represent 91.5% of the sequences (619 sequences) and 19.2% of the
169 haplotypes.

170 In Europe, we found 10 distinct haplotypes from 118 sequences, nine for Asia from 277 sequences,
171 five for Oceania from 106 sequences, 30 for South America from 55 sequences, four in Africa from 78
172 sequences and seven in North America from 43 sequences. Some major haplotypes are represented in
173 a more restricted area, for example haplotype B is mainly present in Asia (figure 1). We observed a
174 high haplotype diversity in South America with 28 minor haplotypes that contain between one and
175 four sequences. The haplotype C contains all the sequences derived from farmed strains of the industry
176 working on BSF.

177

178 Global distribution analysis

179 We visualized the actual distribution of the different haplotypes found previously on a distribution
180 map (figure 2).

181 Each of the haplotypes from A to J is now represented with a color; haplotypes that are numbered in
182 the haplotype network (figure 1) are now represented in grey.

183 We observed a great variability in South America highlighted on the map by the grey pie charts
184 representing mostly minor haplotypes. We observe 22 singletons in South America out of the 52
185 haplotypes found in total, which is more than 40% of the total diversity. On the other hand, we found
186 six out of ten major haplotypes in Europe.

187 Haplotype A and C were the most represented haplotypes in our sampling: Haplotype A was present
188 on all continents, and represented 34% of all sequences, and haplotype C was only absent from South
189 America, and represented 18% of all sequences. Some haplotypes seemed to be restricted to certain
190 regions of the world, such as haplotype G which was present in all the localities in West Africa and
191 surprisingly in a Portuguese sample, or the haplotype H in South America. Haplotype F was present in
192 Europe, Uganda, and South Korea. Haplotype E was more present in Asia and Oceania as well as in
193 North America. Finally, some haplotypes such as B, I and J present rare sequences found in more
194 restricted areas such as B in Korea or H and J in France (figure 2).

195 CO1 Phylogeny

196 During our CO1 gene analyses, some BSF-like larvae were identified as *Exaireta spinigera* larvae. The
197 sequences were amplified and sequenced with the same primers as for BSF.

198 By taking only one sequence for each haplotype we have reconstructed a phylogeny of the CO1 gene
199 sequence. The CO1 phylogeny was rooted with the *Exaireta spinigera* (figure 3). A sample from
200 Cameroon was revealed to belong to a closely unidentified species of BSF and was termed *Hermetia*
201 *sp.* Cameroon in our subsequent analysis (blastn score between *Hermetia* sp. from Cameroon and BSF
202 gives us 85.01% identity with an E Value of 2e-174).

203 The CO1 gene phylogeny supports the analyses performed with the haplotype network (Figure 1),
204 showing remarkable sequence diversity among the CO1 gene. Interestingly, all the commercial strains
205 are found in the C haplotype (a detailed phylogenetic tree is given as a supplementary with an asterisk
206 placed on individuals from farms - (SUPP_Figure 003 (SUPP_MATERIAL))).

207 A group containing haplotypes 35 to 42 and haplotypes B and C forms a divergent group from the
208 others, with strong statistical support (bootstrap value of 0.99). Some sequences of individuals
209 captured in the wild have been found near BSF commercial breeding sites, for example, we have
210 captured C haplotype individuals in Avignon close by companies that are working with the BSF.

211 Although the global resolution of the deep branches of the tree is low, it's also possible to better
212 understand the relationship between the haplotypes: haplotype B and C are close to each other and
213 well separated from other haplotypes. The other haplotypes are not very well resolved with the CO1
214 sequences. Finally, the samples coming from Latin America are scattered all over the tree, showing
215 their great diversity and supporting the hypothesis of the geographical origin of the BSF.

216 The global BSF diversity is mirrored at the local level in France

217 Among the 10 major haplotypes, our sampling effort in France showed that five are present in France
218 in addition to a unique haplotype only found in France (Figure 4) in La Rochelle. Haplotype A remains
219 the dominant haplotype in France (71.1% of all sequences) in seven of the eight localities (excluding

220 Beaufort en Vallée where the flies come from a company). Regarding the haplotype C, BSF belonging
221 to this group have been found in four locations in the regions of Poitiers, Bordeaux, Beaufort-En-Vallée
222 and Montpellier. Except for Beaufort-En-Vallée, where the flies come from a company, the other BSF
223 come from natural environments. BSF industries exist in all these localities. More surprisingly, different
224 haplotypes coexist in some identical localities. For example we found three different haplotypes in the
225 region of Paris and four in the region of Montpellier. Finally, we found a singular haplotype in Colomiers
226 (J) and La Rochelle (1) .

227 Complete mitochondrial genome analyses

228 To better understand BSF phylogeny and to estimate the timing of haplotype divergence, we
229 reconstructed and analysed the mitochondrial genomes of 60 individuals including 3 *Exaireta spinigera*
230 (used as an outgroup). The 60 reconstructed mitochondrial genomes have the typical structure of
231 insects mtDNA : 13 protein-coding genes, 22 tRNA, 2 rRNA-coding genes as well as non-coding regions
232 and control regions (an example is given as a supplementary (SUPP_Figure 004 (SUPP_MATERIAL)).
233 The genetic differences are homogeneously distributed on the whole mtDNA and do not seem to be
234 localized on precise genes (Figure 5A). A great genetic diversity at the mitochondrial genome level is
235 observed. Two very homogeneous groups A and C are well represented, and sequences of the C
236 haplotype are very divergent all along the genomes from the others. The two sequences with the most
237 differences are 55-F-Angouleme and 21-Ghana with a similarity percentage of 96.373% (all results are
238 in supplementary (SUPP_Figure 005 (SUPP_MATERIAL)).

239 We also wanted to know if the mutations are equally (or not) distributed between the different genes.
240 Therefore, we compared the level of similarity of the 13 mitochondrial genes of the 57 BSFs (fig 5B) to
241 those of *Exaireta spinigera*. The results are presented as percent sequence similarity.

242 We grouped the individuals into 3 different categories:

243 - Haplotype A (Figure 5 B1), here all genes show the same differences with those of *Exaireta spinigera*
244 only the NAD6 gene shows slight differences.

245 - Haplotype C (Figure 5 B2), we can see that most of the mitochondrial genes are identical except for
246 the ATP8 gene which is clearly different for four individuals (11 Angouleme; 14 Kenya; 19 Kenya and
247 15 Kenya - Table available in supplementary 4 material). Slight differences are also present in the NAD3
248 and NAD6 genes.

249 -Haplotype D-F-G-H-I (Figure 5 B3), the profiles are all slightly different, with a stronger disparity for
250 the ATP8, NAD1 and NAD5 genes, especially the Mexican one (57). The profiles show the same pattern
251 except for the ATP8 gene of the individual from Mexico (57) which is more conserved.

252 Complete mitochondrial genome phylogeny

253 The complete mitochondrial genome phylogeny demonstrates the very high level of genetic
254 divergence among BSF populations that were found with the CO1 gene analysis (Fig 6A). The increased
255 sensitivity of the analysis, due to the size of the sequences (13.96 kb, 13 genes) robustly resolves the
256 relationships between different haplotypes identified previously (nodes >95%). In addition, it allows
257 us to distinguish subgroups within the previously determined haplotypes. Two major divisions seem to
258 emerge within the tree: group C (which includes all commercial strains) and all others. Within the C
259 group two subgroups emerge (11 – F – Angouleme; 14 Kenya; 19 Kenya; 15 Kenya and the others).

260 The other large group contains the sequences of the A D I G and H haplotypes. The I G and H haplotypes
261 are related and close to each other. The very cosmopolitan A haplotype is well conserved with a limited
262 level of mitochondrial sequence divergences.

263 Moreover, the I G and H haplotypes, although phylogenetically close, are found in very disparate areas
264 (the G haplotype is mostly in Africa, the I in Europe, and the H in Latin America) supporting a multiple
265 worldwide introduction.

266 Strikingly the high level of the conservation of the ATP8 sequence described above in the Mexican
267 sample (haplotype D) does not explain the basal position of the sequence in the phylogeny; indeed,
268 we tested the phylogeny by removing the ATP8 gene and no change was found (SUPP_Figure 006
269 (SUPP_MATERIAL)).

270 The ATP8 gene alone is not sufficient to explain the appearance of the observed subgroups. However,
271 unlike the CO1 gene which varies only slightly within this haplotype, the NAD2, NAD3, NAD5 and NAD6
272 genes are more variable.

273 The average percentage of identity between haplotypes A and C is about 96.39%. Between haplotype
274 C and D 96,54%. Between haplotype C and H 96.40% and 96.43% between C and I. Between A and D
275 97.82% (all results are in SUPP_Figure 005 (SUPP_MATERIAL)).

276 Divergence time of the different groups

277 The time tree (fig 6B) gives us information about the divergence time of the different mitochondrial
278 genomes. We obtained identical Bayesian and ML trees. We noted a relatively long divergence time
279 between haplotype C and the others (2.2 million years). The others have diverged more recently (about
280 1.2 million years). Within the C haplotype, a divergence has been about 300.000 years while in the
281 other groups the divergence times are much lower, about 20.000 years for the A haplotype.

282 The time of divergence with *Exaireta spinigera* has been estimated at 110 million years, which is
283 confirmed by the literature that gives us 112 million years with the TIMETREE tool (Hedges et al, 2015).

284 Discussion

285 *Hermetia illucens* is now considered as a species of major interest for the food industry (Tomberlin and
286 Huis 2020). Thus, it became urgent to study the genetic diversity within the different populations at
287 different scales. The analysis of the different haplotypes in the same geographical area is a powerful
288 tool to study the recent history of BSF. An in-depth study of the genetic variations between the
289 different haplotypes may allow us to better resolve the scenario of the arrival of BSF in a territory.

290 In this study, we uncovered a high genetic variability of mitogenomes within the species *Hermetia*
291 *illucens*. We found that the haplotype C is the most divergent. The dating of the separation events with
292 the other haplotypes goes back more than 2 million years. We found a well resolved separation within
293 haplotype C (>95%); the other haplotypes are also well resolved (>95%).

294 Our study confirms the genetic diversity that was found in previous papers ([Kaya et al, 2021](#) ; [Park et](#)
295 [al, 2017](#) ; [Ståhls et al 2020](#)). We found a comparable number of genetic clusters although we add many
296 more sequences of diverse origin. For example, Ståhls et al, found 56 distinct haplotypes with 418 CO1
297 sequences and we found a comparable number of haplotypes based on the analysis of 677 CO1
298 sequences. These results suggest that we have now captured a major part of the extant BSF worldwide
299 diversity. However in South America we found 30 haplotypes among the 55 sequences sampled, that
300 open-up the possibility of the existence of a still unknown genetic diversity in this part of the world.

301 With the greatest genetic diversity being found in South America, the hypothesis of the geographical
302 origin of BSF in Latin America is further confirmed. Apart from this continent of origin, there seem to
303 be a huge number of neo-introductions on all continents with divergent mitochondrial lineages. The
304 analysis of complete mitogenomes supports the hypothesis of several distinct introductions in Europe
305 as well as on other continents. Our results contradict the scenario of Kaya et al based on
306 microsatellites for a single BSF introduction in Asia and Australia. Indeed, our data support the
307 presence of highly divergent major haplotypes A, C and E in Australia and major haplotypes A, B, C in
308 Asia in addition to several orphan haplotypes. As the divergence times between these major
309 haplotypes is estimated around two million years, this pattern is incompatible with a single
310 introduction by man in the recent period but supports multiple and independent introductions. In the
311 same way, Kaya et al. proposes at least three distinct introductions in Europe. Our data considerably
312 increases this number: at least seven different haplotypes are found (A, I, J, G, F, C and E) supporting
313 at least seven distinct introductions. Similarly, at least four introductions in Africa arose. By increasing
314 the sample size and using more resolute markers, we obtained a more precise description of BSF
315 genetic diversity and a better understanding of its natural history. However, due to the frequent,
316 multiple, and probably ongoing introductions worldwide of the BSF, the understanding of the exact
317 introduction history might be very challenging.

318 Even at a national scale, the BSF genetic diversity is huge. By taking the example of France, we were
319 able to show that the diversity observed at the global scale is also found at the national scale (five of
320 the 10 major haplotypes found worldwide). Moreover, at the local level, we have haplotype mixtures,
321 confirming the observations made on microsatellite data (Kaya et al, 2021). For example, in the region
322 of Paris we have a mix of three haplotypes (A, F and I), and in the region of Montpellier, a mix of four
323 haplotypes (A, B, C and E) – Figure 4.

324 France contains on its territory a great diversity of haplotypes, whose origin can be explained either by
325 the historical introduction of the year 1950 (Barbier 1952), in addition to more recent neo-
326 introductions. The haplotype A, which is by far the most common haplotype in wild BSF found in France
327 (71.1%) and in Europe, may correspond to the initial introduction. However, it is difficult to understand
328 the exact scenario of the introduction of BSF in France: for example, the divergence times within
329 French flies belonging to haplotype A (20.000 years) tend to prove the recent but multiple
330 introductions in France of flies belonging to this group rather than a single introduction by men
331 followed by a diversification on the territory. Our data supports a complex scenario involving multiple
332 and repetitive introduction of the BSF from closely related (same phylogenetic groups) and distantly
333 related (divergent phylogenetic groups) individuals in France. A possible scenario involves an initial
334 introduction of the haplotype A in the 1950s (Barbier 1952) which spread gradually on the territory.
335 More recent introductions arose explaining the occurrence of the haplotypes J, G and I. Finally,
336 introductions related to the industrial activity of the BSF of the haplotype C arose.

337 Jefferey Tomberlin, a pioneer in the field, also indicates in a recent article on the structure and
338 demography of the BSF based on microsatellite data (Kaya et al, 2021), that the captive populations
339 used for breeding in Europe and North America derive from a common, genetically related strain. Our
340 analysis based on CO1 and complete mitochondrial genome sequences confirm this result: the
341 haplotype that we have named C (for commercial) contains all the sequences of industrial origin with
342 very few genetic diversity. Interestingly, this commercial group does not have any South American

343 relatives but [Kayla et al.](#) reports wild north American populations that are related (but not similar) to
344 the haplotype C. This result suggests a possible North American origin for the BSF commercial stock.
345 Our data supports a possible alternative scenario because both network haplotypes and CO1
346 phylogeny indicate that the C haplotype is closely related to the B haplotypes, as well as 8 other
347 haplotypes distributed in Asia and Europe. However, the exact origin of the commercial stocks might
348 be difficult to decipher due to the naturalisation of the C haplotypes in many locations.

349 Indeed, certain individuals collected in the wild and belonging to this group were collected in the
350 vicinity (around 5 km) of industries breeding BSF. For example, if we focus on the commercial
351 haplotype analysis in France (the C haplotype), it is possible that the wild BSF belonging to the
352 haplotype C have escaped from industries in the regions of Poitiers, Bordeaux, Avignon, and
353 Montpellier. This last point raises the question of the biosecurity of BSF farms around the world.
354 Indeed, if the industries carrying out breeding of BSF do not opt for an optimal safety, the escape of
355 BSF of industrial origin can lead either to an establishment of the BSF in territories where it was not
356 yet present, or to a replacement-hybridization of the local populations by the industrial strain. This
357 would lead inexorably to a loss of the local genetic diversity thus potentially erasing some local
358 adaptation to specific environmental conditions.

359 However, it is interesting to note that in [Ståhls et al 2020](#), in contrast to [Kaya et al 2021](#) and our study,
360 some genetic diversity in the breeding strains in Europe and North America have been evidenced (10
361 haplotypes). This may be due to a broader sampling effort of commercial strains. Indeed, STAHLs had
362 the opportunity to sample many small farms leading to 292 BSF sequences from rearing cultures. This
363 allowed them to access a broader genetic diversity not found in our study and in Kaya et al. In addition,
364 it is also possible that smaller farms did not obtain BSF from the dealers who constituted most of the
365 larger industries, but directly from *in Natura* acquisitions. Nevertheless, our results confirm that the
366 industrial actors work with the same and unique strain.

367 We also evidence through this study that the mitochondrial genome is perfectly suited to differentiate
368 haplotypes and to resolve the phylogenetic relationships between them. It allows us to date the
369 natural history of the BSF, bringing a sum of details that were impossible to distinguish based only on
370 the CO1. For example, we report puzzling variations in the level of conservation of the ATP8 genes for
371 some samples (haplotypes C and D). Interestingly, these variations have already been found in other
372 insect species (for example in coleoptera see [Zhang et al 2019](#), and in grasshoppers see [Li et al 2018](#)).
373 The ATP8 gene is a small gene (160 bp on average in insects) that encodes a subunit of the complex V
374 of the ATP synthase. In *Hermetia illucens*, the initiator codon of the ATP8 gene is ATT, is 167 bp long
375 and it overlaps with the ATP6 gene. It has been assumed in other species that the modification of the
376 sequence of the ATP8 gene can affect the structure of mitochondrial complex V and thus its function.
377 Furthermore, it has been shown that changes within ATP synthase can be explained by adaptation to
378 different ecological environments ([Peng et al, 2019](#) ; [Gu et al, 2016](#) ; [Zhang et al, 2017](#)). This indicates
379 that some local adaptation driven by the ATP8 genes may exist in BSF, as it has already been shown in
380 other insects. This can lead to an adaptation to survive in high altitude environments, i.e., at a lower
381 level of oxygen and low temperature ([Luo et al, 2013](#)). Even if these variations did not change the
382 phylogeny of the BSF it can be a clue to some local adaptation signal.

383 Complete mitochondrial genome data allows us to make a robust and well resolved time tree of the
384 species. The divergence times between the different haplotypes are surprisingly long (more than two
385 million years for the most divergent lineages) and the differences observed between the sequences
386 tend to prove that there is still unsuspected and hidden genomic diversity for the species. If we
387 compare it to *Drosophila*, two million years correspond to many speciation events leading to different
388 reproductively isolated species ([Russo et al, 2013](#)). By contrast, [Stahls et al](#) indicated that BSF belonging
389 to phylogenetically divergent haplotypes are still able to cross and generate offspring, indicating that
390 there is no apparent reproductive isolation in laboratory conditions. Interestingly, our data evidence
391 the cohabitation of different haplotypes at the same locality that could lead to crosses between
392 haplotypes. Further investigations are thus needed to understand the possible genetic barriers

393 between highly divergent BSF groups but also the possible patterns of introgression in natural BSF
394 populations. This phenomenon might considerably confuse the elucidation of the population structure
395 and the natural history of these species.

396 Finally, it is interesting to recall that, if indeed, most of the industrial actors use the same haplotype
397 for the industry, the other haplotypes present in France are very divergent from this one. Strikingly,
398 we found in one compost bin in the region of Paris flies belonging to two different haplotypes (A and
399 F). Thus, crossbreeding between these two haplotypes would bring a strong genetic mix that could be
400 beneficial for breeding. Associated with this significant genetic diversity, BSF may also have some
401 significant phenotypic diversity that might be useful for specific industrial usage and selection
402 processes to increase performances or to ameliorate peculiar traits or behaviours (Zhou et al, 2013).

403 Conclusion

404 During this study, through the analysis of 60 mitochondrial genomes, we were able to propose a robust
405 and well resolved time tree of the BSF that confirms the huge and hidden genetic diversity within the
406 species *Hermetia illucens*. Our result elucidates the phylogenetic relationship and time divergence
407 between the different BSF lineages and provides a plausible scenario for BSF expansion in the world
408 via multiple and independent introduction at the global and local scale. With the use of 677 COI gene
409 sequences coming from a broad geographical sample, we were able to highlight the genetic diversity
410 at the level of the different continents evidencing recurrent and multiple introductions outside the
411 geographical area of origin in Latin America. However, the exact scenario of introductions, even at a
412 national scale in France, are still difficult to understand due to probable multiple and independent
413 introductions in addition to possible interbreeding between different strains that coexist in the same
414 geographical area. We have also shown that a large part of the BSF industrial companies work with
415 flies that derive from a unique strain. We would also like to remind that the apparent biosecurity
416 deficiency within some BSF industrial companies probably led to escapement and possible local
417 introduction/replacement of natural populations of BSF. Finally, our data indicated that it becomes
418 extremely interesting to take a closer look at the nuclear genomes' evolution of the BSF to better

419 understand the natural history of the BSF as well as specific adaptations of the different lineages that
420 might be useful for industrial needs and to initiate the selection process for peculiar traits.

421 **Availability of data and materials**

422 The dataset generated during the study has been deposited in the European Nucleotide Archive
423 database under Accession Numbers PRJEB48031 Bioprotect. Alignment and raw data can be
424 downloaded following this link <http://gofile.me/2ppPR/QWjM33T7y> the supplementaries can be
425 downloaded following this link : <http://gofile.me/2ppPR/rwaGSXDQU>

426

427 **Contributions**

428 JG carried out the sampling, performed data curation, and wrote the original draft. NP performed data
429 curation and reviewed the manuscript. GB helped with the data curation, and reviewed the
430 manuscript. JF conceptualised and supervised the study, performed data curation, reviewed, and
431 finalized the manuscript.

432 **Funding**

433 This work was funded in the framework of a partnership between the CNRS and the CycleFarms
434 company, JG is supported by a CIFRE PhD Grant from the Agence Nationale pour la Recherche et la
435 Technologie (ANRT)

436 **Competing interests**

437 This project was partially granted by the private company CycleFarms but at any moment, the company
438 has influenced the design and the analysis of the results nor the writing of the manuscript and its
439 conclusions. No commercial products have been developed based on the outputs of this article. None
440 of the authors have any competing interests.

441 **Acknowledgements**

442 We would like to thank Florian Laville and Frederic Marion-Poll for his continuous support and help
443 during this project. We warmly thank the members of the Réseau Compost Citoyen for their precious

444 help during the sampling in France. We also thank Matan Shelomi (National Taiwan University), Meng-
445 Kun Wu, Shu-Min Chen, Jing-Jiun Huang (Yi Mi Community College, Taiwan) for flies from Taiwan. We
446 thank the international Centre of Insect Physiology and Ecology (ICIPE) from Nairobi City, Kenya. We
447 thank Mathieu Chouteau from CNRS LEESIA, in French Guiana. We finally thank Benoit Gilles for his
448 help at the beginning of this project.

449 References

- 450 1 - Rozkosný, R. A Biosystematic Study of the European Stratiomyidae (Diptera): Volume 2 - Clitellariinae, Hermediinae, Pachygasterinae and
451 Bibliography. Series Entomologica. Springer Netherlands, 1983. <https://www.springer.com/gp/book/9789061931355>.
- 452 2 - Hauser, Martin, et Marshall Woodley. « The historical spread of the Black Soldier Fly, *Hermetia illucens* (L.) (Diptera, Stratiomyidae,
453 Hermetiinae), and its establishment in Canada. » *Journal of the Kansas Entomological Society* 146 (1 décembre 2015): 51-54.
- 454 3 - https://ec.europa.eu/environment/nature/invasivealien/list/index_en.html
- 455 4 - Wang, Yu-Shiang, et Matan Shelomi. « Review of Black Soldier Fly (*Hermetia Illucens*) as Animal Feed and Human Food ». *Foods* (Basel,
456 Switzerland) 6, n° 10 (18 octobre 2017). <https://doi.org/10.3390/foods6100091>.
- 457 5 - Mason, Franco, Rudolf Rozkošný, et Martin Hauser. « A Review of the Soldier Flies (Diptera: Stratiomyidae) of Sardinia ». *Zootaxa* 2318,
458 n° 1 (22 décembre 2009): 507-30. <https://doi.org/10.11646/zootaxa.2318.1.20>.
- 459 6 - Picker, Mike, Charles Griffiths, et Alan Weaving. *Field Guide to Insects of South Africa*. Penguin Random House South Africa, 2002.
- 460 7 - Lindner, Erwin. *Die amerikanische Hermetia illucens L. im Mittelmeergebiet: (Stratiomyidae, Dipt.)*. Akadem. Verlagsges., 1936.
- 461 8 - Benelli, Giovanni, Angelo Canale, Alfio Raspi, et Gino Fornaciari. « The Death Scenario of an Italian Renaissance Princess Can Shed Light
462 on a Zoological Dilemma: Did the Black Soldier Fly Reach Europe with Columbus? » *Journal of Archaeological Science* 49 (1 septembre 2014)
463 : 203-5. <https://doi.org/10.1016/j.jas.2014.05.015>.
- 464 9 - Barbier, Jean. « Introduction en France d'un Diptère Stratiomyide américain ». *Bulletin de la Société entomologique de France* 57, n° 7
465 (1952) : 108-108.
- 466 10 - Dauphin, Patrick. « Présence de *Hermetia illucens* (LINNÉ, 1758) dans le sud-ouest de la France (Diptera Stratiomyidae) », s. d., 3.
- 467 11 - Richoux, Philippe. « Sur la présence d'*Hermetia illucens* (Linnaeus, 1758) (Diptère Stratiomyidae) dans la région lyonnaise/Presence of
468 *Hermetia illucens* (Linnaeus, 1758) (Diptera Stratiomyidae) in the region of Lyons ». *Publications de la Société Linnéenne de Lyon* 78, n° 5
469 (2009) : 137-38. <https://doi.org/10.3406/linly.2009.13722>.
- 470 12 - Pierre-Olivier, Maquart, Richard Denis, et Jesse Willems. « First record of the Black Soldier Fly, *Hermetia illucens*, in the Western regions
471 of France (Vendée, Loire-Atlantique, Ille-et-Vilaine) with notes on its worldwide repartition (Diptera, Stratiomyidae) ». *Bulletin de la Société*
472 *entomologique de France* 125 (20 mars 2020) : 13-18. https://doi.org/10.32475/bsef_2104.
- 473 13 - *Leclercq, M. (1969). Dispersion et transport des insectes nuisibles : Hermetia illucens L. (Diptera Stratiomyidae). Bull. Rech. Agron.*
474 *Gembloux N.S., 4, 139-43*
- 475 14 - Craig Sheppard, D., G. Larry Newton, Sidney A. Thompson, et Stan Savage. « A Value-Added Manure Management System Using the
476 Black Soldier Fly ». *Bioresource Technology* 50, n° 3 (1 janvier 1994): 275-79. [https://doi.org/10.1016/0960-8524\(94\)90102-3](https://doi.org/10.1016/0960-8524(94)90102-3).

- 477 15 - Lord, W. D., M. L. Goff, T. R. Adkins, et N. H. Haskell. « The Black Soldier Fly *Hermetia Illucens* (Diptera: Stratiomyidae) As a Potential
478 Measure of Human Postmortem Interval: Observations and Case Histories ». *Journal of Forensic Science* 39, n° 1 (1 janvier 1994): 215-22.
479 <https://doi.org/10.1520/JFS13587J>.
- 480 16 - Gold, Moritz, Jeffery K. Tomberlin, Stefan Diener, Christian Zurbrügg, et Alexander Mathys. « Decomposition of Biowaste Macronutrients,
481 Microbes, and Chemicals in Black Soldier Fly Larval Treatment: A Review ». *Waste Management (New York, N.Y.)* 82 (décembre 2018): 302-
482 18. <https://doi.org/10.1016/j.wasman.2018.10.022>.
- 483 17 - Gold, Moritz, Melanie Binggeli, Fabienne Kurt, Tomas de Wouters, Markus Reichlin, Christian Zurbrügg, Alexander Mathys, et Michael
484 Kreuzer. « Novel Experimental Methods for the Investigation of *Hermetia Illucens* (Diptera: Stratiomyidae) Larvae ». *Journal of Insect Science*
485 *(Online)* 20, n° 3 (1 mai 2020): 21. <https://doi.org/10.1093/jisesa/ieaa057>.
- 486 18 - Hoc, B., M. Genva, M.-L. Fauconnier, G. Lognay, F. Francis, et R. Caparros Megido. « About Lipid Metabolism in *Hermetia Illucens* (L.
487 1758): On the Origin of Fatty Acids in Prepupae ». *Scientific Reports* 10, n° 1 (17 juillet 2020): 11916. [https://doi.org/10.1038/s41598-020-
488 68784-8](https://doi.org/10.1038/s41598-020-68784-8).
- 489 19 - Ewald, Nils, Aleksandar Vidakovic, Markus Langeland, Anders Kiessling, Sabine Sampels, et Cecilia Lalander. « Fatty Acid Composition of
490 Black Soldier Fly Larvae (*Hermetia Illucens*) - Possibilities and Limitations for Modification through Diet ». *Waste Management (New York,*
491 *N.Y.)* 102 (1 février 2020): 40-47. <https://doi.org/10.1016/j.wasman.2019.10.014>.
- 492 20 - Giannetto, Alessia, Sabrina Oliva, Carlos Frederico Ceccon Lanes, Fabio de Araújo Pedron, Domenico Savastano, Cosimo Baviera, Vincenzo
493 Parrino, et al. « *Hermetia Illucens* (Diptera: Stratiomyidae) Larvae and Prepupae: Biomass Production, Fatty Acid Profile and Expression of Key
494 Genes Involved in Lipid Metabolism ». *Journal of Biotechnology*, 31 octobre 2019. <https://doi.org/10.1016/j.jbiotec.2019.10.015>.
- 495 21 - Sprangers, Thomas, Matteo Ottoboni, Cindy Klootwijk, Anneke Obyn, Stefaan Deboosere, Bruno De Meulenaer, Joris Michiels, Mia
496 Eeckhout, Patrick De Clercq, et Stefaan De Smet. « Nutritional Composition of Black Soldier Fly (*Hermetia Illucens*) Prepupae Reared on
497 Different Organic Waste Substrates ». *Journal of the Science of Food and Agriculture* 97, n° 8 (juin 2017): 2594-2600.
498 <https://doi.org/10.1002/jsfa.8081>.
- 499 22 - Huis, Arnold van. *Edible Insects: Future Prospects for Food and Feed Security*. FAO Forestry Paper 171. Rome: Food and Agriculture
500 Organization of the United Nations, 2013.
- 501 23 - Leong, Siew Yoong, Shamsul Rahman Mohamed Kutty, Amirhossein Malakahmad, et Chew Khun Tan. « Feasibility Study of Biodiesel
502 Production Using Lipids of *Hermetia Illucens* Larva Fed with Organic Waste ». *Waste Management (New York, N.Y.)* 47, n° Pt A (janvier 2016):
503 84-90. <https://doi.org/10.1016/j.wasman.2015.03.030>.
- 504 24 - Surendra, K. C., Jeffery K. Tomberlin, Arnold van Huis, Jonathan A. Cammack, Lars-Henrik L. Heckmann, et Samir Kumar Khanal. «
505 Rethinking Organic Wastes Bioconversion: Evaluating the Potential of the Black Soldier Fly (*Hermetia Illucens* (L.)) (Diptera: Stratiomyidae)
506 (BSF) ». *Waste Management (New York, N.Y.)* 117 (novembre 2020): 58-80. <https://doi.org/10.1016/j.wasman.2020.07.050>.
- 507 25 - Heuel, M., C. Sandrock, F. Leiber, A. Mathys, M. Gold, C. Zurbrügg, I. D. M. Gangnat, M. Kreuzer, et M. Terranova. « Black Soldier Fly
508 Larvae Meal and Fat Can Completely Replace Soybean Cake and Oil in Diets for Laying Hens ». *Poultry Science* 100, n° 4 (avril 2021): 101034.
509 <https://doi.org/10.1016/j.psj.2021.101034>.
- 510 26 - Schiavone, A., S. Dabbou, M. Petracci, M. Zampiga, F. Sirri, I. Biasato, F. Gai, et L. Gasco. « Black Soldier Fly Defatted Meal as a Dietary
511 Protein Source for Broiler Chickens: Effects on Carcass Traits, Breast Meat Quality and Safety ». *Animal: An International Journal of Animal*
512 *Bioscience* 13, n° 10 (octobre 2019): 2397-2405. <https://doi.org/10.1017/S1751731119000685>.

- 513 27 - Neumann, Carmen, Susanne Velten, et Frank Liebert. « N Balance Studies Emphasize the Superior Protein Quality of Pig Diets at High
514 Inclusion Level of Algae Meal (*Spirulina Platensis*) or Insect Meal (*Hermetia Illucens*) When Adequate Amino Acid Supplementation Is Ensured
515 ». *Animals: An Open Access Journal from MDPI* 8, n° 10 (3 octobre 2018): E172. <https://doi.org/10.3390/ani8100172>.
- 516 28 - Biasato, Ilaria, Manuela Renna, Francesco Gai, Sihem Dabbou, Marco Meneguz, Giovanni Perona, Silvia Martinez, et al. « Partially
517 Defatted Black Soldier Fly Larva Meal Inclusion in Piglet Diets: Effects on the Growth Performance, Nutrient Digestibility, Blood Profile, Gut
518 Morphology and Histological Features ». *Journal of Animal Science and Biotechnology* 10 (2019): 12. <https://doi.org/10.1186/s40104-019-0325-x>.
- 519
- 520 29 - Khamis, Fathiya M., Fidelis L. O. Ombura, Komivi S. Akutse, Sevgan Subramanian, Samira A. Mohamed, Komi K. M. Fiaboe, Weerachai
521 Saijuntha, et al. « Insights in the Global Genetics and Gut Microbiome of Black Soldier Fly, *Hermetia Illucens*: Implications for Animal Feed
522 Safety Control ». *Frontiers in Microbiology* 11 (2020): 1538. <https://doi.org/10.3389/fmicb.2020.01538>.
- 523 30 - Erickson, Marilyn C., Mahbub Islam, Craig Sheppard, Jean Liao, et Michael P. Doyle. « Reduction of *Escherichia Coli* O157:H7 and
524 *Salmonella Enterica* Serovar Enteritidis in Chicken Manure by Larvae of the Black Soldier Fly ». *Journal of Food Protection* 67, n. 4 (avril 2004):
525 685-90. <https://doi.org/10.4315/0362-028x-67.4.685>.
- 526 31 - Liu, Qiaolin, Jeffery K. Tomberlin, Jeff A. Brady, Michelle R. Sanford, et Ziniu Yu. « Black Soldier Fly (Diptera: Stratiomyidae) Larvae Reduce
527 *Escherichia coli* in Dairy Manure ». *Environmental Entomology* 37, n° 6 (1 décembre 2008) : 1525-30. [https://doi.org/10.1603/0046-225X-](https://doi.org/10.1603/0046-225X-37.6.1525)
528 [37.6.1525](https://doi.org/10.1603/0046-225X-37.6.1525).
- 529 32 - Park Soyeon, Hansu Choi, Ji-young Choi, et Gilsang Jeong. « Population Structure of the Exotic Black Soldier Fly, *Hermetia illucens* (Diptera:
530 Stratiomyidae) in Korea ». Consulté le 1 avril 2021. <https://doi.org/10.13047/KJEE.2017.31.6.520>.
- 531 33 - Ståhls, Gunilla, Rudolf Meier, Christoph Sandrock, Martin Hauser, Ljiljana Šašić Zorić, Elina Laiho, Andrea Aracil, et al. « The Puzzling
532 Mitochondrial Phylogeography of the Black Soldier Fly (*Hermetia Illucens*), the Commercially Most Important Insect Protein Species ». *BMC*
533 *Evolutionary Biology* 20, n° 1 (24 mai 2020): 60. <https://doi.org/10.1186/s12862-020-01627-2>.
- 534 34 - Kaya, Cengiz, Tomas N. Generalovic, Gunilla Ståhls, Martin Hauser, Ana C. Samayoa, Carlos G. Nunes-Silva, Heather Roxburgh, et al. «
535 Global population genetic structure and demographic trajectories of the black soldier fly, *Hermetia illucens* ». *BMC Biology* 19, n° 1 (5 mai
536 2021): 94. <https://doi.org/10.1186/s12915-021-01029-w>.
- 537 35 - Qi, Yingju, Jingyang Xu, Xiaoxuan Tian, Yichuan Bai, et Xishu Gu. « The complete mitochondrial genome of *Hermetia illucens* (Diptera:
538 Stratiomyidae) ». *Mitochondrial DNA Part B* 2, n° 1 (1 janvier 2017): 189-90. <https://doi.org/10.1080/23802359.2017.1307708>.
- 539 36 - Generalovic, Tomas N., Shane A. McCarthy, Ian A. Warren, Jonathan M. D. Wood, James Torrance, Ying Sims, Michael Quail, et al. « A
540 High-Quality, Chromosome-Level Genome Assembly of the Black Soldier Fly (*Hermetia Illucens* L.) ». *BioRxiv*, 15 novembre 2020,
541 2020.11.13.381889. <https://doi.org/10.1101/2020.11.13.381889>.
- 542 37 - Zhan, Shuai, Gangqi Fang, Minmin Cai, Zongqing Kou, Jun Xu, Yanghui Cao, Liang Bai, et al. « Genomic Landscape and Genetic
543 Manipulation of the Black Soldier Fly *Hermetia Illucens*, a Natural Waste Recycler ». *Cell Research* 30, n° 1 (janvier 2020): 50-60.
544 <https://doi.org/10.1038/s41422-019-0252-6>.
- 545 41 - Leinonen R, Sugawara H, Shumway M; International Nucleotide Sequence Database Collaboration. The sequence read archive. *Nucleic*
546 *Acids Res.* 2011 Jan;39(Database issue): D19-21. doi: 10.1093/nar/gkq1019. Epub 2010 Nov 9. PMID: 21062823; PMCID: PMC3013647.
- 547 43 – <https://www.mn-net.com/fr/nucleobond-axg-100-midi-columns-for-high-integrity-dna-740545>

- 548 44 - Desjardins P, Conklin D. NanoDrop microvolume quantitation of nucleic acids. *J Vis Exp*. 2010 Nov 22;(45) :2565. doi : 10.3791/2565.
549 PMID: 21189466; PMCID: PMC3346308.
- 550 47 - Geneious R11 (<https://www.geneious.com>)
- 551 48 - Katoh K, Standley DM. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol Biol*
552 *Evol*. 2013 Apr;30(4):772-80. doi: 10.1093/molbev/mst010. Epub 2013 Jan 16. PMID: 23329690; PMCID: PMC3603318.
- 553 49 - Maddison DR, Swofford DL, Maddison WP. NEXUS: an extensible file format for systematic information. *Syst Biol*. 1997 Dec;46(4):590-
554 621. doi: 10.1093/sysbio/46.4.590. PMID: 11975335.
- 555 50 - Leigh, JW, Bryant D (2015). PopART: Full-feature software for haplotype network construction. *Methods Ecol Evol* 6(9):1110–1116.
- 556 51 - Cock P.J.A. et al (2010). The Sanger FASTQ file format for sequences with quality scores, and the Solexa/Illumina FASTQ variants. *Nucleic*
557 *acids research* 38, 1767-1771.
- 558 52 - Hansen K.D. et al (2010). Biases in Illumina transcriptome sequencing caused by random hexamer priming. *Nucleic acids research* 38,
559 e131-e131.
- 560 53 - Erlich Y. et al (2008). Alta-Cyclic: a self-optimizing base caller for next-generation sequencing. *Nature Methods*,5,679-682.
- 561 54 - Jiang L.C. et al (2011). Synthetic spike-in standards for RNA-seq experiments. *Genome research* 21, 1543-1551.
- 562 55 - Yan L.Y. et al (2013). Single-cell RNA-Seq profiling of human preimplantation embryos and embryonic stem cells. *Nat Struct Mol Biol*.
- 563 56 - Dinghua Li, Chi-Man Liu, Ruibang Luo, Kunihiko Sadakane, Tak-Wah Lam, MEGAHIT: an ultra-fast single-node solution for large and
564 complex metagenomics assembly via succinct de Bruijn graph, *Bioinformatics*, Volume 31, Issue 10, 15 May 2015, Pages 1674–1676,
565 <https://doi.org/10.1093/bioinformatics/btv033>
- 566 57 - Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. Basic local alignment search tool. *J Mol Biol*. 1990 Oct 5;215(3):403-10. doi:
567 10.1016/S0022-2836(05)80360-2. PMID: 2231712.
- 568 58 - Bernt M, Donath A, Jühling F, Externbrink F, Florentz C, Fritzsch G, Pütz J, Middendorf M, Stadler PF. MITOS: improved de novo metazoan
569 mitochondrial genome annotation. *Mol Phylogenet Evol*. 2013 Nov;69(2):313-9. doi: 10.1016/j.ympev.2012.08.023. Epub 2012 Sep 7. PMID:
570 22982435.
- 571 59 - Sudhir Kumar, Glen Stecher, Michael Li, Christina Knyaz, and Koichiro Tamura (2018) MEGA X: Molecular Evolutionary Genetics Analysis
572 across computing platforms. *Molecular Biology and Evolution* 35:1547-1549
- 573 60 - Letunic I and Bork P (2021) *Nucleic Acids Res* doi: 10.1093/nar/gkab301 Interactive Tree Of Life (iTOL) v5: an online tool for phylogenetic
574 tree display and annotation
- 575 61 - Marc A Suchard, Philippe Lemey, Guy Baele, Daniel L Ayres, Alexei J Drummond, Andrew Rambaut, Bayesian phylogenetic and
576 phylodynamic data integration using BEAST 1.10, *Virus Evolution*, Volume 4, Issue 1, January 2018, vey016,
577 <https://doi.org/10.1093/ve/vey016>
- 578 62 - Helfrich, Philipp, Elias Rieb, Giuseppe Abrami, Andy Lücking and A. Mehler. "TreeAnnotator: Versatile Visual Annotation of Hierarchical
579 Text Relations." *LREC* (2018).
- 580 63 – <https://www.thermofisher.com/order/catalog/product/Q32850#Q32850>
- 581 65 – Hedges, S. Blair, Julie Marin, Michael Suleski, Madeline Paymer, et Sudhir Kumar. « Tree of Life Reveals Clock-Like Speciation and
582 Diversification ». *Molecular Biology and Evolution* 32, n. 4 (1 avril 2015): 835-45. <https://doi.org/10.1093/molbev/msv037>.

- 583 66 - Tomberlin, J.k., et A. van Huis. « Black soldier fly from pest to ‘crown jewel’ of the insects as feed industry: an historical perspective ».
584 *Journal of Insects as Food and Feed* 6, n. 1 (6 février 2020): 1-4. <https://doi.org/10.3920/JIFF2020.0003>.
- 585 67 - Zhang, Shou-ke, Jin-ping Shu, Yang-dong Wang, Ya-ning Liu, Han Peng, Wei Zhang, et Hao-jie Wang. « The Complete Mitochondrial
586 Genomes of Two Sibling Species of Camellia Weevils (Coleoptera: Curculionidae) and Patterns of Curculionini Speciation ». *Scientific Reports*
587 9, n. 1 (4 mars 2019): 3412. <https://doi.org/10.1038/s41598-019-39895-8>.
- 588 68 - Li, Xiao-Dong, Guo-Fang Jiang, Li-Yun Yan, Ran Li, Yuan Mu, et Wei-An Deng. « Positive Selection Drove the Adaptation of Mitochondrial
589 Genes to the Demands of Flight and High-Altitude Environments in Grasshoppers ». *Frontiers in Genetics* 9 (2018).
590 <https://doi.org/10.3389/fgene.2018.00605>.
- 591 69 - Peng, Quekun, Lei Tang, Shuai Tan, Zhigang Li, Jifei Wang, et Fangdong Zou. « Mitogenomic Analysis of the Genus Pseudois: Evidence of
592 Adaptive Evolution of Morphological Variation in the ATP Synthase Genes ». *Mitochondrion* 12, n. 5 (1 septembre 2012): 500-505.
593 <https://doi.org/10.1016/j.mito.2012.07.107>.
- 594 70 - Gu, Peng, Wei Liu, Yong-fang Yao, Qing-yong Ni, Ming-wang Zhang, Di-yan Li, et Huai-liang Xu. « Evidence of adaptive evolution of alpine
595 pheasants to high-altitude environment from mitogenomic perspective ». *Mitochondrial DNA Part A* 27, n. 1 (2 janvier 2016): 455-62.
596 <https://doi.org/10.3109/19401736.2014.900667>.
- 597 71 - Zhang, Qi-Lin, Li Zhang, Tian-Xuan Zhao, Juan Wang, Qian-Hua Zhu, Jun-Yuan Chen, et Ming-Long Yuan. « Gene Sequence Variations and
598 Expression Patterns of Mitochondrial Genes Are Associated with the Adaptive Evolution of Two Gynaephora Species (Lepidoptera:
599 Lymantriinae) Living in Different High-Elevation Environments ». *Gene* 610 (30 avril 2017) : 148-55.
600 <https://doi.org/10.1016/j.gene.2017.02.014>.
- 601 72 - Luo, Yongjun, Xiaohong Yang, et Yuqi Gao. « Mitochondrial DNA response to high altitude: A new perspective on high-altitude
602 adaptation ». *Mitochondrial DNA* 24, n. 4 (1 août 2013) : 313-19. <https://doi.org/10.3109/19401736.2012.760558>.
- 603 73 - Russo, Claudia A. M., Beatriz Mello, Annelise Frazão, et Carolina M. Voloch. « Phylogenetic Analysis and a Time Tree for a Large
604 Drosophilid Data Set (Diptera: Drosophilidae) ». *Zoological Journal of the Linnean Society* 169, n. 4 (2013). <https://doi.org/10.1111/zoi.12062>.
- 605 74 - Zhou, Fen, Jeffery K. Tomberlin, Longyu Zheng, Ziniu Yu, et Jibin Zhang. « Developmental and Waste Reduction Plasticity of Three Black
606 Soldier Fly Strains (Diptera: Stratiomyidae) Raised on Different Livestock Manures ». *Journal of Medical Entomology* 50, n. 6 (novembre 2013):
607 1224-30. <https://doi.org/10.1603/me13021>.
- 608 75 - <https://reseaucompost.org/>
- 609 76 - Meng, Guanliang, Yiyuan Li, Chentao Yang, et Shanlin Liu. « MitoZ: a toolkit for animal mitochondrial genome assembly, annotation and
610 visualization ». *Nucleic Acids Research* 47, n. 11 (20 juin 2019): e63. <https://doi.org/10.1093/nar/gkz173>.

ORIGIN	NUMBER OF INDIVIDUALS (CO1)	NUMBER OF INDIVIDUALS (MITOCHONDRIA)
COLLECTION FROM THE WILD	91	43
INDUSTRIAL ORIGIN	23	13
MINED FROM DATABASES (GENBANK ³⁹ & BOLD ⁴⁰)	560	1
ASSEMBLED FROM SRA ⁴¹ (RAW SEQUENCES)	3	3
TOTAL	677	60

Table 1: Origin of CO1 sequences and mitochondrial genomes

GEOGRAPHIC ORIGIN	NUMBER OF INDIVIDUALS – CO1	NUMBER OF INDIVIDUALS – MITOCHONDRIA
SOUTH AMERICA	55	4
NORTH AMERICA	43	2
ASIA	277	4
OCEANIA	106	0
AFRICA	78	11
EUROPE (INCLUDING FRANCE)	118 (90)	39 (38)
TOTAL	677	60

Table 2: Geographic origin of CO1 sequences and mitochondrial genomes

FIG 1

Haplotype network generated by PopArt after multiple alignment, based on the CO1 gene of *Hermetia illucens*. Analysis of the differences between the CO1 gene sequences of 677 individuals. The pie charts represent the same haplotype, and their relative size gives an indication of the number of individuals sharing this haplotype. The number of bars between each segment indicates the number of differences between the two sequences. Within a pie chart the colors represent the geographical origin of the individuals. The major haplotypes are those with more than five sequences (A B C E F G H 30 38 & 41). The minor haplotypes are the others. The list of sequences and their exact origin is available as an additional document.

Fig 2

Global distribution of the different *Hermetia illucens* haplotypes based on the CO1 gene. The pie charts represent the proportion of each haplotype within a country. The name of the country is followed by the number of individuals analysed in that country. A colored band representing the continents is next to each pie chart. The colors represent the haplotypes determined in figure 1. The grey haplotypes are those that were represented with a number in figure 1. A separation is made in the pie chart when the haplotypes change for the same country. For example, Venezuela contains three unique haplotypes, meaning that these three individuals are different from each other. A summary table can be found in the supplementary table 3.

FIG 3

ML phylogenetic tree. Rooted phylogeny with *Exaireta spinigera* (Stratiomyidae). Bootstrap values are materialized with colored circles: green for values > 0.85 and red for values between 0.7 and 0.85. We only kept one sequence per haplotype to visualise the relationship between them. Next to each haplotype, colored circles represent the geographical origin of the species present in this haplotype. The order indicates the relative abundance of the individuals of a

container in each haplotype. The statistical support was estimated with 500 Bootstraps with MegaX software using the Tamura 92 G+I model.

FIG 4

Distribution of the different haplotypes of *Hermetia illucens* in France. The dots represent the precise area of collect. The pie charts represent the distribution of haplotypes in the corresponding area, the names of the main cities in each area are indicated above as well as the number of sequenced individuals.

Paris includes Vincennes and Nogent Sur Marnes. Blois includes Valloire Sur Cisse. Lyon includes Saint Marcellin En Forez, Corbas, Champagne Au Mont d'Or and Dardilly. Montpellier includes Avignon, Trescléoux and Dieulefit. Bordeaux includes Lormont La Force and Périgueux. Poitiers includes Saint Georges Lès Baillargeaux.

Fig 5A

Multiple alignment of mitochondrial DNA sequences from *Hermetia illucens*. Alignment performed with the MAFFT aligner. The colors of the names come from their haplotypes determined with the CO1 sequences. Each black mark represents a difference with the reference (1 – China). Above the alignment there is a representation of the mitochondrial genes.

FIG 5B

Sequence conformity of mitochondrial genes of *Hermetia illucens* to *Exaireta spinigera*. The sequences are grouped according to haplotypes. The A haplotype (B1), the C haplotype (B2) and the other haplotypes (B3). The sequences inside each graph are those represented in the alignment (5A). Sequences from 23 to 46 plus sequences 51, 52, and 55 for haplotype A (5-B1); from 1 to 22 for haplotype C (5-B2); and from 47 to 57 (except sequences 51, 52 and 55) for the other haplotypes (5-B3).

FIG 6A

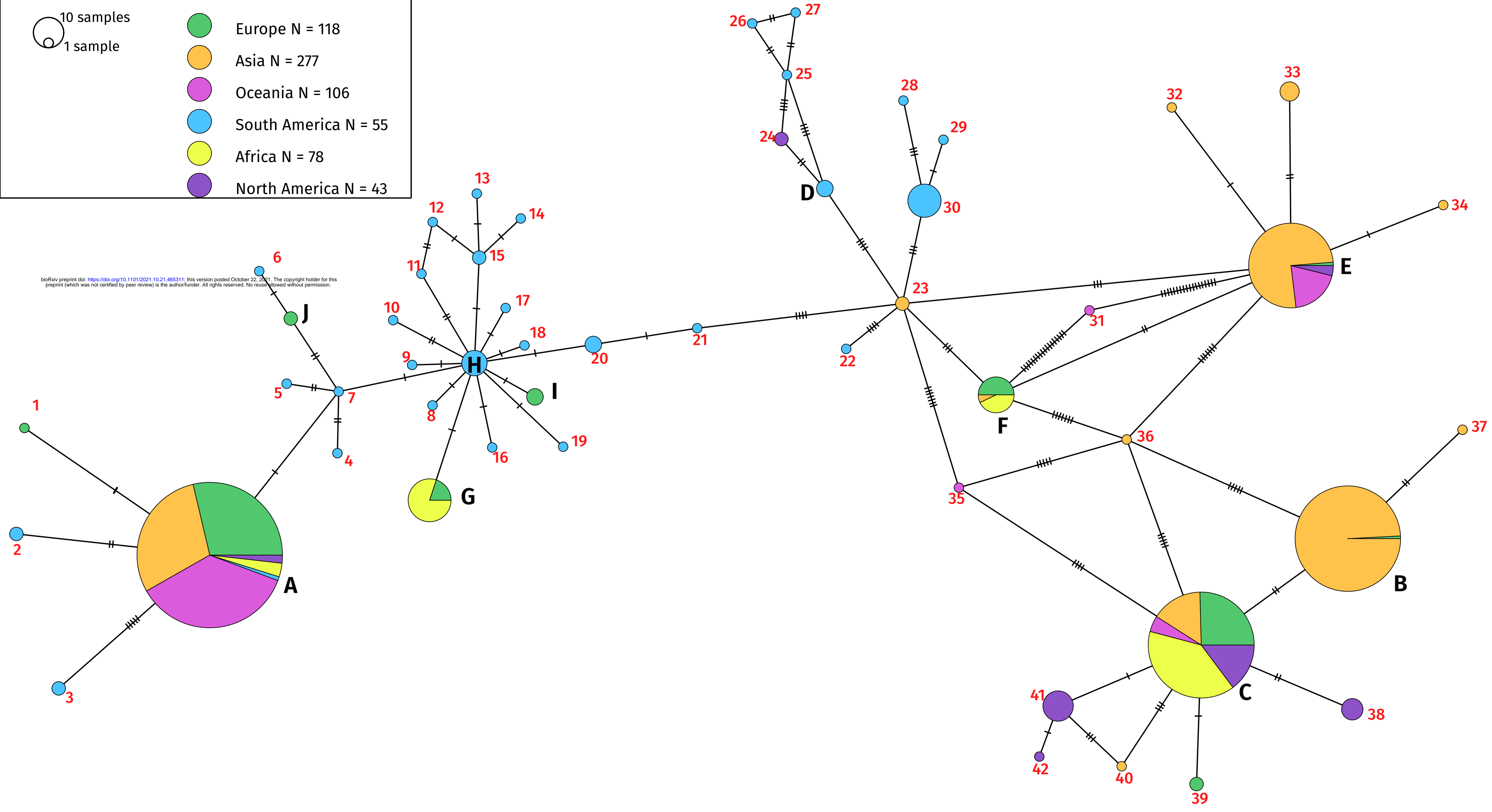
ML phylogenetic tree, of 57 mitochondrial DNA sequences of *Hermetia illucens*, deprived of D-Loop regions. MAFFT was used to align the sequences, 500 Bootstrap replicas were performed. The outgroup used is *Exaireta spinigera* (Stratiomyidae). The colors of the names correspond to the haplotypes determined with the CO1 gene sequences (Figure 1). Bootstrap values are indicated by a green circle when they are > 0.95. Commercial individuals are marked with an asterisk.

FIG 6B

Bayesian time tree obtained with BEAST of 60 mitochondrial genomes of *Hermetia illucens* rooted with *Exaireta spinigera*. The scale is in millions of years. A break in scale is made at the root level. The colors of the names correspond to the haplotypes determined with the CO1 sequences (figure 1). Commercial individuals are marked with an asterisk.

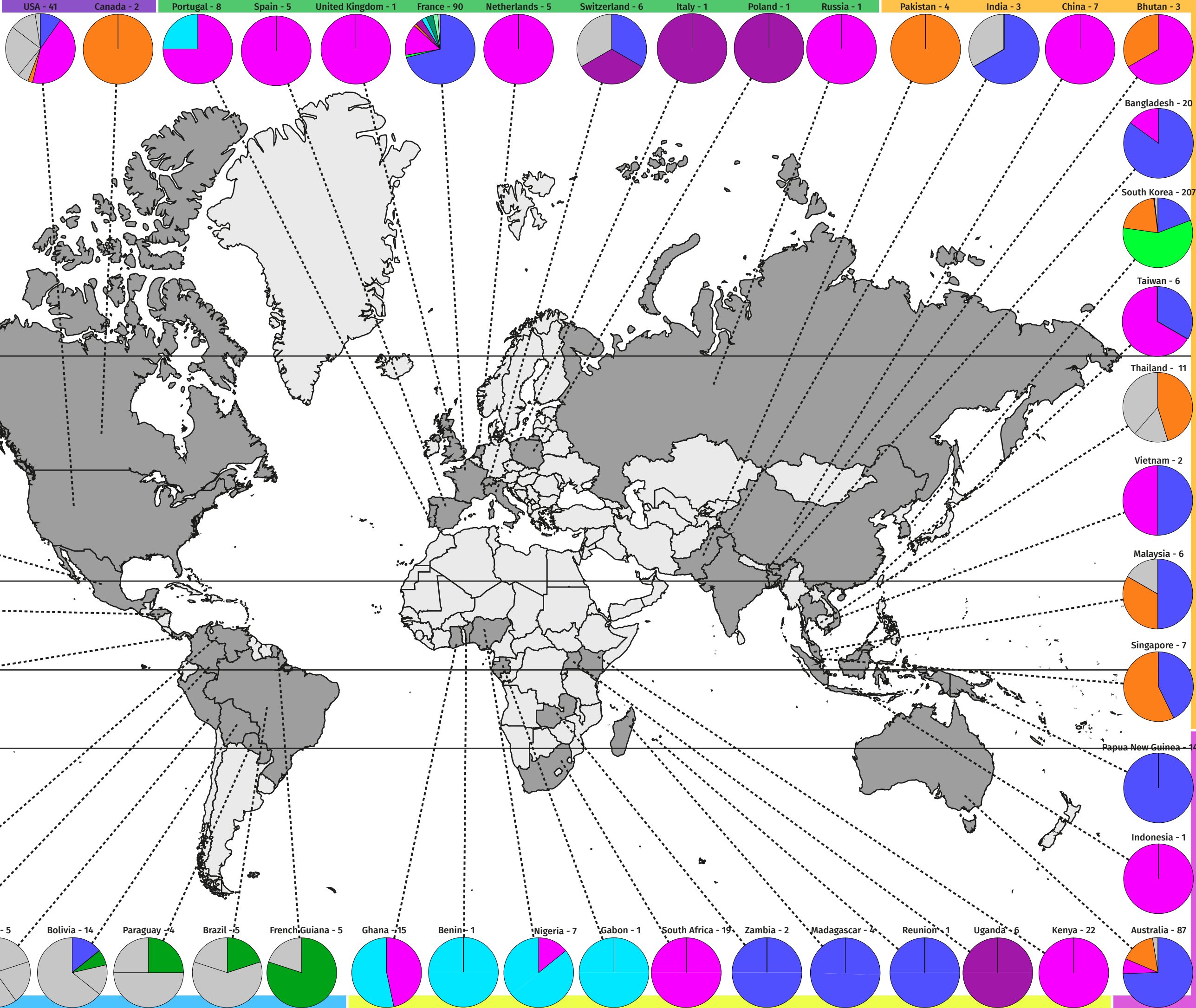
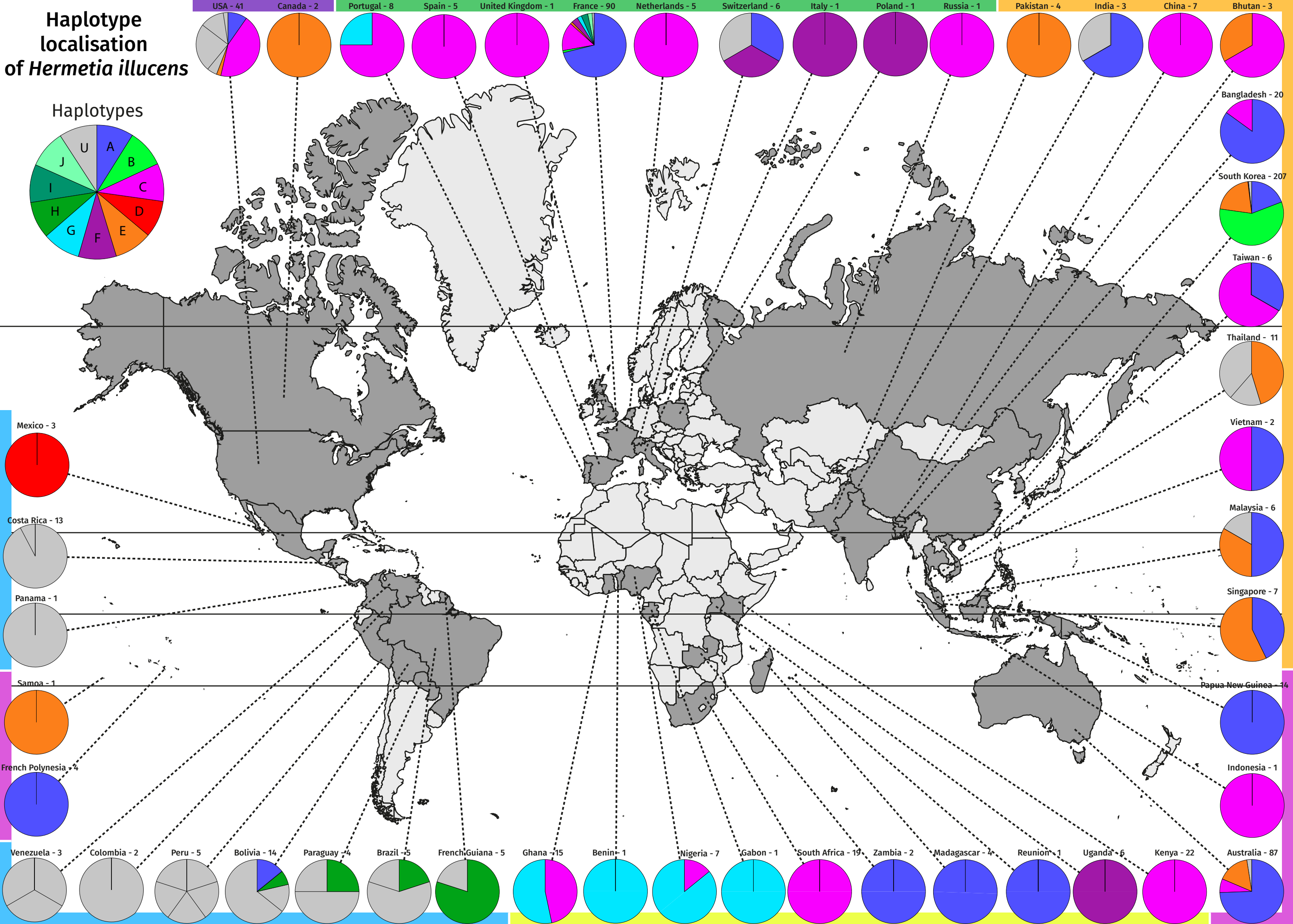
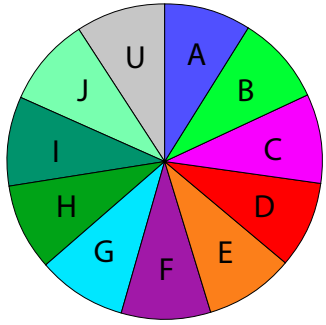


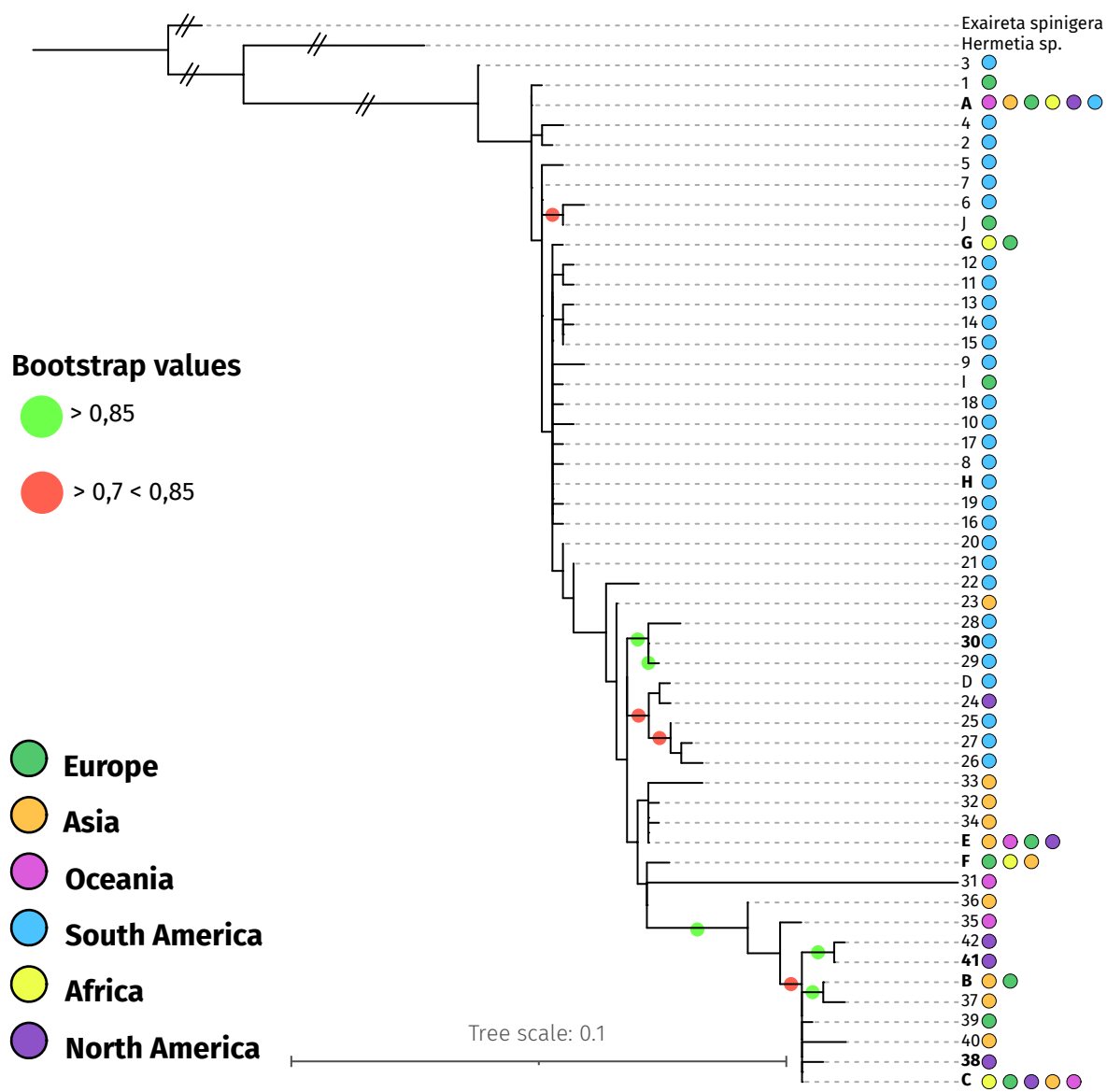
bioRxiv preprint doi: <https://doi.org/10.1101/2021.10.21.465311>; this version posted October 22, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

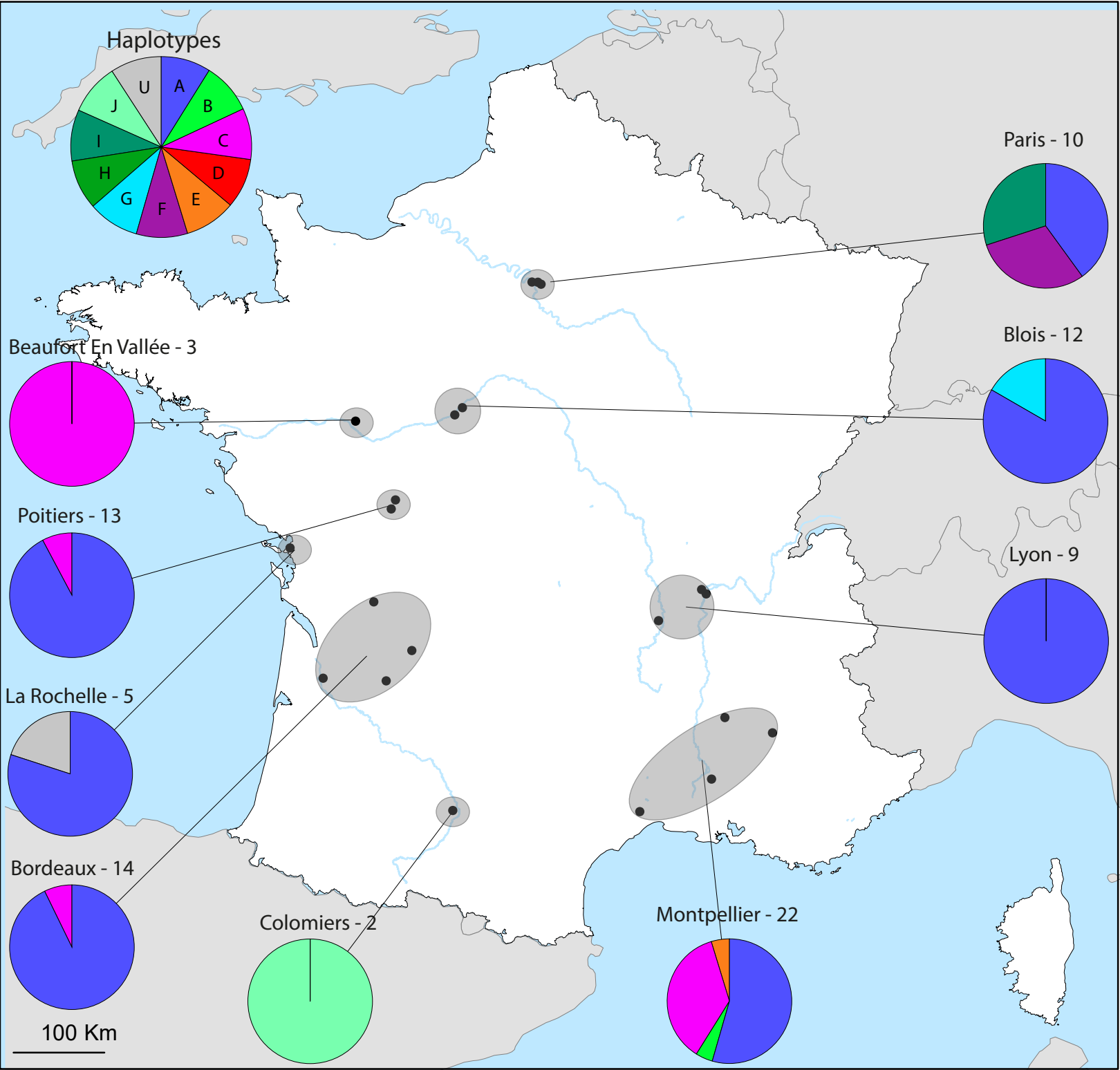


Haplotype localisation of *Hermetia illucens*

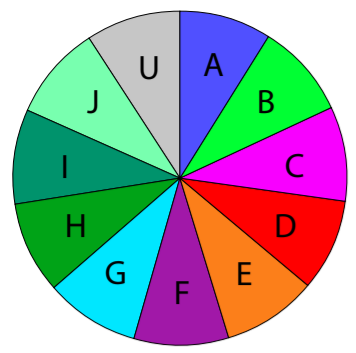
Haplotypes



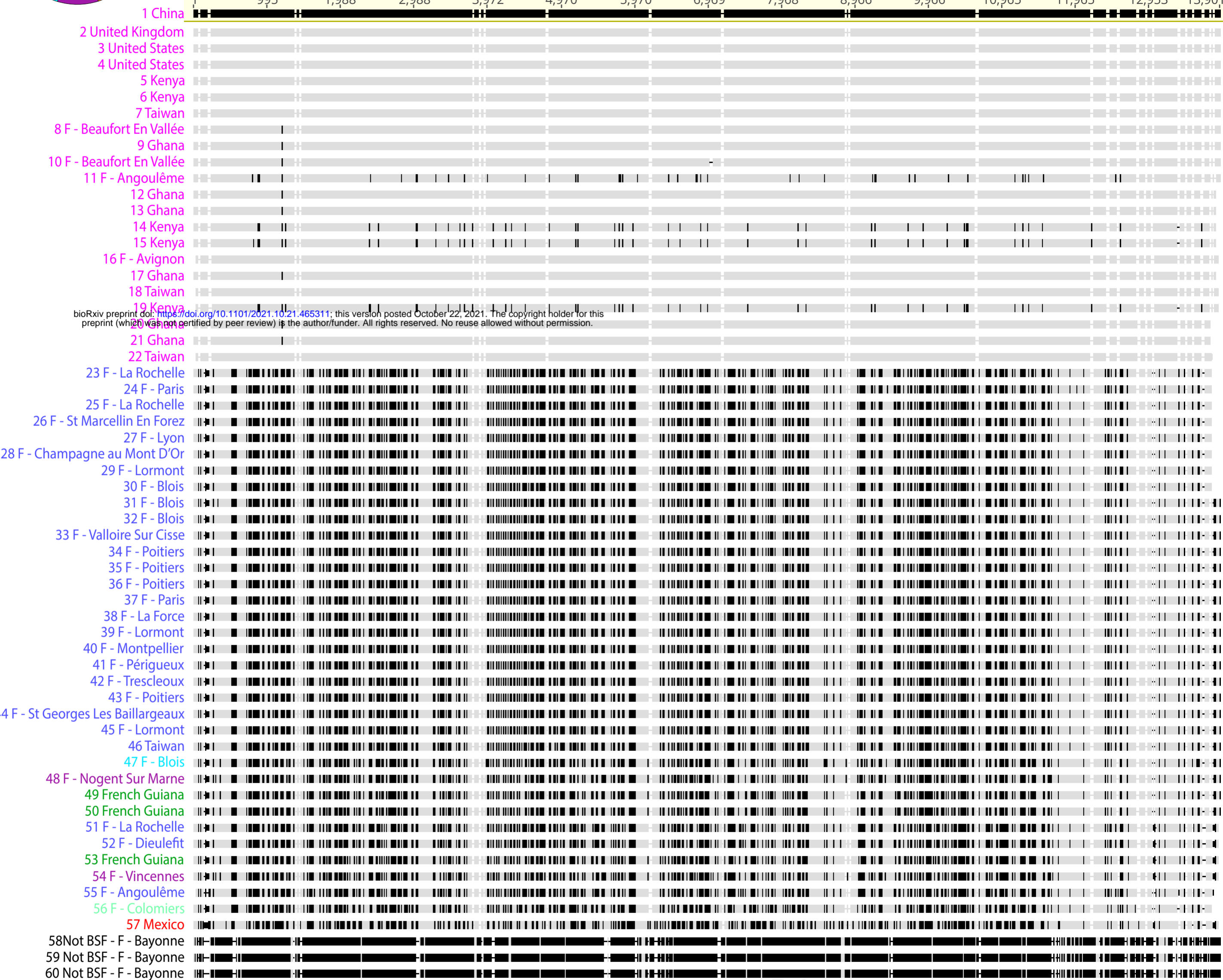
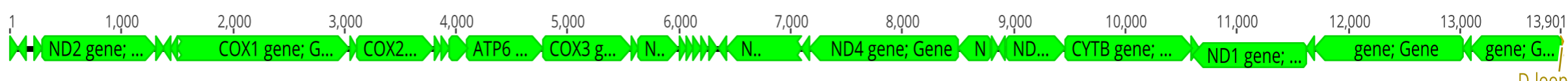




Haplotypes



A

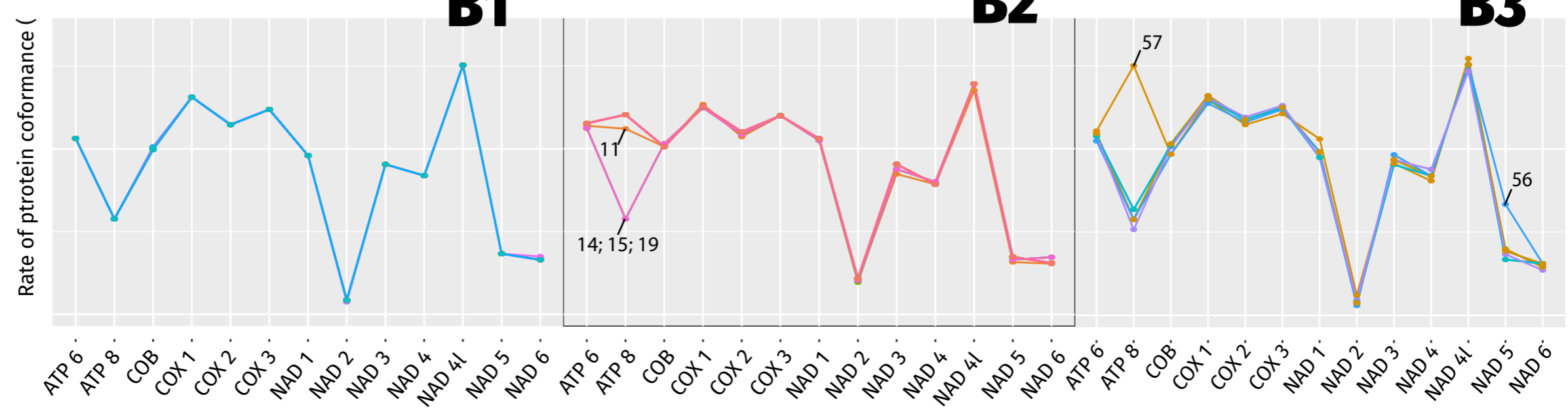


bioRxiv preprint doi: <https://doi.org/10.1101/2021.10.21.465311>; this version posted October 22, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

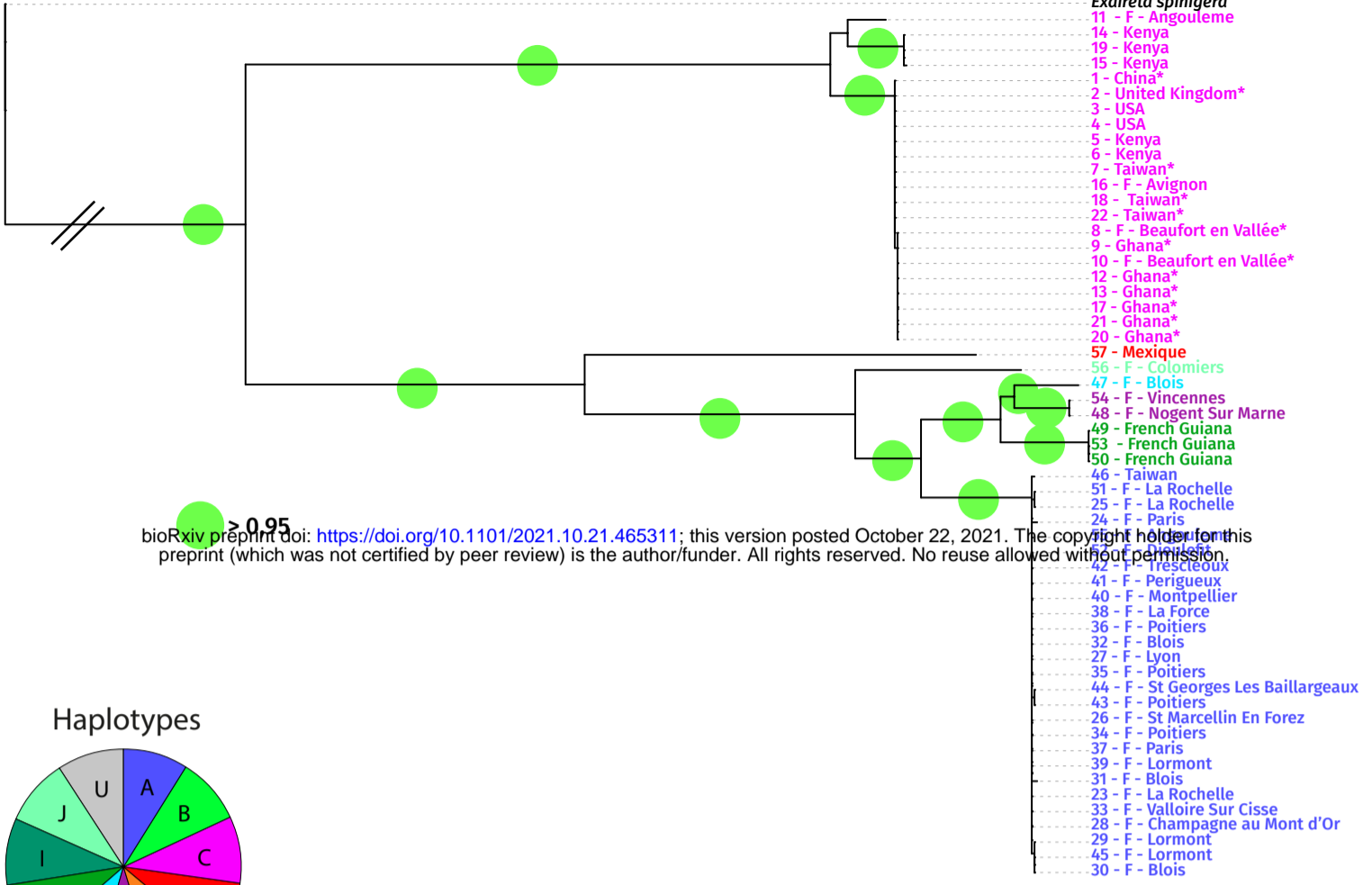
B1

B2

B3



Tree scale: 0.01



Tree scale : 1
(in million years)

