2 surviving Aedes albopictus under optimal nutritional availability

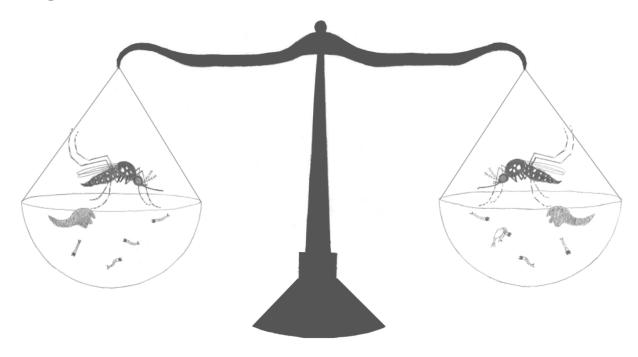
- 3 Marie C. Russell¹* and Lauren J. Cator¹
- ¹Department of Life Sciences, Imperial College London, Silwood Park Campus, Ascot, UK
- 6 *Correspondence: marie.clare.russell@gmail.com

8 Graphical abstract

1

4

7



Abstract

9

10

11

12

13

14

15

16

Background: Cyclopoid copepods have been applied successfully to limit populations of highly invasive *Aedes albopictus* mosquitoes that can vector diseases, including chikungunya, dengue, yellow fever, and Zika, to humans. However, there is concern that changes in certain vector traits, induced by exposure to copepod predation, might increase the risk of disease

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

transmission. In this study, we tested whether cyclopoid copepod predation has sublethal effects on the development time or adult size of Ae. albopictus under the scenario of an invasion in southeast England, which lies at the northern edge of the vector's expanding global range. **Methods:** Third instar Ae. albopictus larvae, hereafter referred to as "focal individuals", were placed in individual Petri dishes, each containing four newly-hatched Ae. albopictus larvae, which were counted, removed, and replaced daily. All focal individuals were provided with fish food ad libitum, and half were exposed to Megacyclops viridis copepod predators. The day of pupation was recorded for each focal individual, and the wing length of each focal adult was also measured. **Results:** Exposing late instar Ae. albopictus to predation decreased their chances of surviving to adulthood. Mortality of the focal individuals was 12.9% in the predator treatment, and 2.9% in the controls. Three focal larvae that died in the predator treatment showed signs of melanization, indicative of wounding. Among surviving Ae. albopictus, no significant difference in either pupation day or wing length was observed due to copepod predation. **Conclusions:** We found that *M. viridis* predation on smaller *Ae. albopictus* larvae does not significantly affect the pupation day or adult size of surrounding larger larvae that are less vulnerable to copepod attacks. This study controlled for density effects on size by maintaining a constant number of newly-hatched prey larvae surrounding each focal larva. Those working to control Ae. albopictus populations in the field should be made aware that increased adult body size can occur if copepod biocontrol agents are applied at lower than necessary levels. The absence of a significant sublethal impact from M. viridis copepod predation on surviving later-stage larvae in this analysis supports the use of M. viridis as a biocontrol agent.

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

Keywords predator-prey interactions, sublethal effects, vector traits **Background** Aedes albopictus is an important vector of dengue, chikungunya, yellow fever, and Zika [1]. This species is highly invasive due, in part, to its ability to lay desiccation-resistant eggs that can be transported across long distances, often by the shipment of used tires [2-6]. The northern limits of the global range of Ae. albopictus include North American populations in New York and Connecticut, USA [7], as well as European populations throughout Italy and France [8]. Several different methods have been proposed for controlling aedine mosquitoes in Europe, including chemical, genetic, and biological techniques [9]. Although space spraying with pyrethroid insecticide has been found to be effective against Ae, albopictus in Catalonia, this control strategy would require regular monitoring for insecticide resistance [10]. The Sterile Insect Technique (SIT) has previously been tested on Ae. albopictus mosquitoes from Italy, but it was not found to significantly reduce their population due to the reduced mating competitiveness of irradiated males [11]. Cyclopoid copepods have been used successfully as biocontrol agents against mosquito larvae in the US [12], Australia [13], Vietnam [14], and Italy [15]. Copepods are an especially convenient type of biocontrol because they are small enough to be distributed through a backpack sprayer [16]. When local predators are used to limit populations of a disease vector, there is a risk that these predation interactions could have unintended effects on the ability of surviving prey to reproduce and transmit disease.

68

69

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

heartworm, *Dirofilaria immitis* in Australia [26]. However, in comparison to how other

93

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

5

when controlling for differences due to sex and diet [37]. However, smaller Ae. albopictus

females are more likely to become infected with dengue and to disseminate the virus than larger females [38]. Furthermore, small females have been shown to be more likely than large females to take multiple bloodmeals within a single gonotrophic cycle [39], and the lifetime cumulative number of bloodmeals is also higher among smaller females [40]. This higher contact frequency with hosts increases the risk for disease transmission by small-sized *Ae*. *albopictus* vectors.

Previous studies have shown an increase in *Ae. albopictus* adult size after exposure to predation, but these studies were not designed to control for the greater per capita nutrition or decreased intraspecific competition that often occur when the population density has been significantly lowered due to successful predation [27, 41-43]. The phenomenon of increasing animal body size with decreasing population density has been documented across taxa [44]. This study is designed to test for sublethal effects of copepod predation on the development and adult size of *Ae. albopictus* that have been exposed to the act of predation during the later larval stages, while controlling for density effects. Due to the suitability of some areas in southeast England for *Ae. albopictus* populations [45], we used *Megacyclops viridis*, a likely copepod species for future biocontrol applications [46], collected from Longside Lake in Egham, UK, and *Ae. albopictus* that were originally collected in Montpellier, France. Our results show that while cyclopoid copepod predation by *M. viridis* significantly increases mortality of late-instar *Ae. albopictus*, the related predation cues do not significantly change development time or adult size of those late-instar *Ae. albopictus* that survive, assuming optimal nutritional availability.

Methods

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

7

ground fish food (Cichlid Gold Hikari®, Japan) was added ad libitum. After three hours, 500

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

dishes (Fig 1).

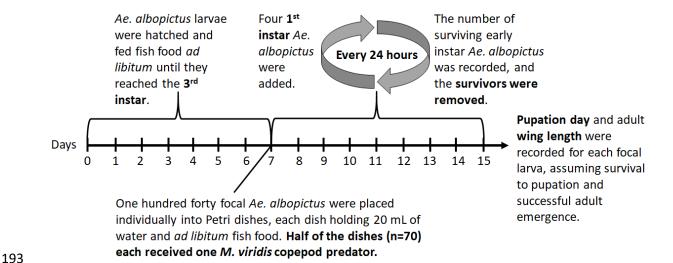


Fig 1. Experiment schedule

Each day, the number of surviving first or second instar larvae in each of the 140 Petri dishes was recorded, surviving first or second instars were removed, and four new first instars from the 18-hour hatch started on the previous day were added to each replicate (Fig 1). The status (dead or alive) of each focal larva was recorded daily, and in predator treatment replicates, the status of the copepod was also recorded. In the case of a focal larva death, the larva was preserved in 80% ethanol, and that replicate was removed from further observation. In the case of a copepod death, the copepod was preserved in 80% ethanol, and a new adult female *M. viridis* copepod from the September field collections was randomly chosen to replace it.

Pupation among focal individuals was recorded each day at 18:00 hrs. Pupae were transferred to 10 mL of spring water in a graduated cylinder with a mesh cover for emergence. Emerged adults were frozen at -20°C. Wings were removed and measured as a proxy for body size [52].

Data analysis:

All analyses were completed in R version 3.4.2 [53]. Welch two sample t-tests for samples of unequal variance were used to compare percent survival of first or second instar larvae between copepod absent and copepod present treatments. The possibility of a difference in the proportion of focal larvae emerging as adults based on copepod presence was examined using a Pearson's chi-squared test without Yates' continuity correction. Two Kruskal-Wallis tests, one for males and one for females, were used to compare adult wing lengths between copepod absent and copepod present treatments. A Kruskal-Wallis test was also used to compare pupation day distributions between predator present and predator absent treatments. A non-parametric local regression method ("loess", "ggplot2" package) was used to present the cumulative proportion of focal larvae pupated over time, by predator presence.

Results

Head capsule width measurements (mean = 0.59 mm, sd = 0.062 mm) of a subset of the focal mosquito larvae (n = 30) confirmed that they were third instars on the first day of exposure to copepod predation [51]. One hundred fifteen copepods (mean length = 1.75 mm, sd = 0.16 mm) were used throughout the experiment across the 70 predator treatment replicates.

Out of the 70 focal larvae that were not exposed to copepod predators, 68, or 97.1%, emerged successfully as adults; one died in the larval stage, and one died in the pupal stage. Out of the 70 focal larvae that were exposed to copepod predators, 61, or 87.1%, emerged successfully as adults; five died in the larval stage, and four died in the pupal stage. Results of a Pearson's chi-squared test showed that the probability of successful adult emergence was higher in the

absence of copepod predators (p-value = 0.0279). Three of the five focal larvae that died in

the presence of a predator showed clear signs of melanization in the abdominal region, most likely due to wounding from copepod attacks (Fig 2).

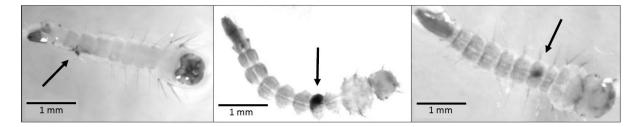


Fig 2. Evidence of melanization in focal larvae that died in the presence of a copepod predator on the eighth, ninth, and tenth days of observation, respectively

The percentage of first instar *Ae. albopictus* surviving each day was significantly lower in the presence of a copepod predator throughout the six days immediately following predator introduction (Table 1). No significant difference in first instar survival was observed on the last two days, when the number of remaining replicates was very low (Table 1).

Table 1. First instar percent survival by day and predator presence

Day of	Predator	Number of	Predator	Number of	P-value ^a
Experiment	Absent	Predator Absent	Present	Predator Present	
	$(mean \pm se)$	Replicates	$(mean \pm se)$	Replicates	
8	99.3 ± 0.5	70	6.8 ± 2.0	70	< 0.0001
9	99.3 ± 0.5	70	30.1 ± 4.1	69	< 0.0001
10	99.3 ± 0.5	70	47.8 ± 3.9	68	< 0.0001
11	100 ± 0.0	70	42.9 ± 4.3	67	< 0.0001
12	100 ± 0.0	42	55.9 ± 6.6	34	< 0.0001
13	100 ± 0.0	12	58.3 ± 8.9	12	0.0007
14	100 ± 0.0	5	33.3 ± 33.3	3	0.1835
15	100	1	25 ± 14.4	3	NA

^aP-value corresponds to a Welch two sample t-test for samples of unequal variance used to determine if there was a difference in first instar survival based on predator presence.

Ae. albopictus adult wing length data were left-skewed among both males and females. Female wing lengths (median = 2.87 mm) were significantly larger than male wing lengths (median = 2.33 mm, Kruskal-Wallis rank sum test, p-value < 0.0001). However, there were no significant differences in male wing length (Kruskal-Wallis rank sum test, p-value =

0.6387) or in female wing length (Kruskal-Wallis rank sum test, p-value = 0.1769), due to copepod presence (Fig 3a).

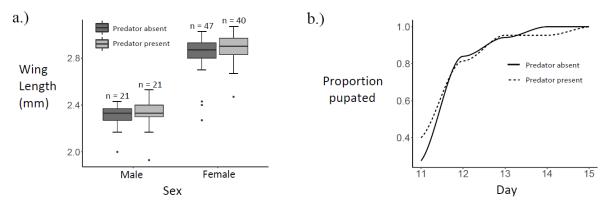


Fig 3. Size and development: a.) Boxplot of wing lengths by sex and predator presence, b.) Cumulative proportion pupated by predator presence

Pupation day data were right-skewed among both males and females. Sex did not affect the day of pupation (Kruskal-Wallis rank sum test, geometric mean \pm sd: males = 11.8 ± 1.1 , females = 11.9 ± 1.1 ; p-value = 0.2607). In addition, neither male pupation day (geometric mean \pm sd: copepod present = 11.8 ± 1.1 , copepod absent = 11.7 ± 1.1 ; Kruskal-Wallis rank sum test, p-value = 0.7819), nor female pupation day (geometric mean \pm sd: copepod present = 11.8 ± 1.1 , copepod absent = 12.0 ± 1.1 ; Kruskal-Wallis rank sum test, p-value = 0.1580), differed with copepod presence (Fig 3b).

Discussion

Although third and fourth instar *Ae. albopictus* larvae are generally less vulnerable to copepod predators than first and second instars [17], some of the third instar, fourth instar, and pupal stage deaths observed in this study were likely due to *M. viridis* copepod attacks. Three focal individuals that died in the larval stage showed signs of melanization (Fig. 2), a response triggered locally by cuticular wounding that results in the accumulation of melanin,

277

278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

297

298

299

300

a brown-black pigment, at the wound site [54-56]. Melanization is an energetically costly response that is likely to be influenced by nutritional status in mosquitoes [54, 57, 58]. Since the focal larvae in this study were provided with fish food ad libitum, it is unlikely that their immune responses were limited due to poor nutritional status. The location of the three melanization sites in the abdominal region (Fig. 2) is consistent with a previous study, which found that the larval abdomen was attacked more frequently by cyclopoid copepods than either the head/thorax region or the last body segment, containing the siphon [59]. While successfully emerging males were spread evenly between the predator and control treatments, less females emerged successfully from the predator treatment. These observations are consistent with those of a previous study showing that Ae. albopictus survivorship is skewed towards males in response to predation by *Toxorhynchites rutilus* [41]. A longer (five-week) semi-field study found that the Ae. albopictus sex ratio was skewed towards females after extended exposure to cyclopoid copepod predation [42]. However, lower larval densities have been shown to produce lower proportions of males in Ae. albopictus rearing [60]. One possible explanation for this is that the increased nutrient availability for each larva at lower densities might better support the larger body size of females. Thus, lower larval density resulting from predation, is likely to be the main cause of the female-dominated sex ratio that was previously observed [42]. In order for cyclopoid copepods to be effective biocontrols, enough adult female copepods need to be applied in order to quickly eliminate first and second instar mosquito larvae, which are the most vulnerable stages to copepod predation [17]. In some cases of incomplete control, Ae. albopictus adults that developed in the presence of predators emerged larger than adults that developed in control conditions because the lower larval density produced by

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

320

321

322

323

324

325

predators resulted in less intraspecific competition and greater per capita nutrition [41-43]. It has also been suggested that first instar larvae benefit nutritionally from decomposing dead conspecifics [27]. First and second instar larvae are likely to benefit the most from increased nutrition because most larval growth occurs between the first and third instar stages. Under the scenario of incomplete biocontrol leading to the emergence of larger Ae. albopictus adults, it is important to consider that Ae. albopictus females do not avoid copepods during oviposition [17, 42]. Therefore, the higher fecundity that is associated with larger female size [33, 36] is likely to be strongly counteracted by copepod predation against newly-hatched Ae. albopictus larvae of the next generation. Among Ae. albopictus adults that emerged successfully in this experiment, there was a significant difference in wing length due to sex (Fig. 3a), but there was not a significant difference in pupation day between sexes. Ae. albopictus males have previously been observed to be 17-20% smaller than females [61]. Accordingly, the male median wing length in this study is 18.8% smaller than the female median wing length, and wing length is known to correlate positively with mass in Ae. albopictus [52]. Previous work has shown that while mass clearly differs due to sex, development time in Ae. albopictus is less sexually dimorphic [62]. There was no significant difference in wing length or pupation day due to *M. viridis* predation cues (Fig. 3). Therefore, neither the greater reproductive success observed among larger Ae. albopictus [33-36], nor the higher risk for disease transmission observed among smaller female Ae. albopictus [38-40], is likely to result directly from M. viridis predation. A similar lack of predator impact on mosquito size and development time was observed when Toxorhynchites amboinensis was tested against newly-hatched Ae. polynesiensis in coconuts

327

328

329

330

331

332

333

334

335

336

337

338

339

340

341

342

343

344

345

346

347

348

349

350

Availability of data and materials:

References

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

373

374

- 1. Centers for Disease C, Prevention. Surveillance and Control of Aedes aegypti and
- Aedes albopictus in the United States. 2017. Contract No.: Report.
- 2. Eritja R, Escosa R, Lucientes J, Marques E, Molina R, Roiz D, et al. Worldwide
- invasion of vector mosquitoes: present European distribution and challenges for Spain.
- 381 Biological Invasions. 2005;7(1):87-97.
- 382 3. Juliano SA, Lounibos LP. Ecology of invasive mosquitoes: effects on resident species
- and on human health. Ecology Letters. 2005;8(5):558-74.
- 4. Lounibos LP. Invasions by insect vectors of human disease. Annual Review of
- 385 Entomology. 2002;47:233-66.
- 386 5. Medlock JM, Avenell D, Barrass I, Leach S. Analysis of the potential for survival and
- seasonal activity of Aedes albopictus (Diptera : Culicidae) in the United Kingdom. Journal of
- 388 Vector Ecology. 2006;31(2):292-304.
- 389 6. Benedict MQ, Levine RS, Hawley WA, Lounibos LP. Spread of the tiger: Global risk
- of invasion by the mosquito Aedes albopictus. Vector-Borne and Zoonotic Diseases.
- 391 2007;7(1):76-85.

- 392 7. Kache PA, Eastwood G, Collins-Palmer K, Katz M, Falco RC, Bajwa WI, et al.
- 393 Environmental Determinants of Aedes albopictus Abundance at a Northern Limit of Its
- Range in the United States. American Journal of Tropical Medicine and Hygiene.
- 395 2020;102(2):436-47.
- 8. European Centre for Disease P, Control, cartographers. Aedes albopictus current
- 397 known distribution: May 20202020.
- 398 9. Baldacchino F, Caputo B, Chandre F, Drago A, della Torre A, Montarsi F, et al.
- Control methods against invasive Aedes mosquitoes in Europe: a review. Pest management
- 400 science. 2015;71(11):1471-85.
- 401 10. Paaijmans K, Brustollin M, Aranda C, Eritja R, Talavera S, Pages N, et al. Phenotypic
- 402 insecticide resistance in arbovirus mosquito vectors in Catalonia and its capital Barcelona
- 403 (Spain). Plos One. 2019;14(7).
- 404 11. Bellini R, Balestrino F, Medici A, Gentile G, Veronesi R, Carrieri M. Mating
- 405 Competitiveness of Aedes albopictus Radio-Sterilized Males in Large Enclosures Exposed to
- Natural Conditions. Journal of Medical Entomology. 2013;50(1):94-102.
- 407 12. Marten GG. Elimination of Aedes-Albopictus from Tire Piles by Introducing
- 408 Macrocyclops-Albidus (Copepoda, Cyclopidae). Journal of the American Mosquito Control
- 409 Association. 1990;6(4):689-93.
- 410 13. Russell BM, Muir LE, Weinstein P, Kay BH. Surveillance of the mosquito Aedes
- aegypti and its biocontrol with the copepod Mesocyclops aspericornis in Australian wells and
- gold mines. Medical and Veterinary Entomology. 1996;10(2):155-60.
- 413 14. Kay BH, Nam VS, Van Tien T, Yen NT, Phong TV, Diep VTB, et al. Control of
- 414 Aedes vectors of dengue in three provinces of Vietnam by use of Mesocyclops (Copepoda)
- and community-based methods validated by entomologic, clinical, and serological
- surveillance. American Journal of Tropical Medicine and Hygiene. 2002;66(1):40-8.
- 417 15. Veronesi R, Carrieri M, MacCagnani B, Maini S, Bellini R. Macrocyclops Albidus
- 418 (Copepoda: Cyclopidae) for the Biocontrol of Aedes Albopictus and Culex Pipiens in Italy.
- Journal of the American Mosquito Control Association. 2015;31(1):32-43.
- 420 16. Marten GG. Issues in the development of cyclops for mosquito control. Arbovirus
- 421 Research in Australia. 1990;5:159-64.
- 422 17. Dieng H, Boots M, Tuno N, Tsuda Y, Takagi M. A laboratory and field evaluation of
- 423 Macrocyclops distinctus, Megacyclops viridis and Mesocyclops pehpeiensis as control agents

- of the dengue vector Aedes albopictus in a peridomestic area in Nagasaki, Japan. Medical and
- 425 Veterinary Entomology. 2002;16(3):285-91.
- 426 18. Preisser EL, Bolnick DI, Benard MF. Scared to death? The effects of intimidation and
- consumption in predator-prey interactions. Ecology. 2005;86(2):501-9.
- 428 19. Werner EE, Peacor SD. A review of trait-mediated indirect interactions in ecological
- 429 communities. Ecology. 2003;84(5):1083-100.
- 430 20. Ferrari MCO, Wisenden BD, Chivers DP. Chemical ecology of predator-prey
- interactions in aquatic ecosystems: a review and prospectus. Canadian Journal of Zoology.
- 432 2010;88(7):698-724.
- 433 21. Sih A. ANTIPREDATOR RESPONSES AND THE PERCEPTION OF DANGER
- 434 BY MOSQUITO LARVAE. Ecology. 1986;67(2):434-41.
- 435 22. Ferrari MCO, Messier F, Chivers DP. Variable predation risk and the dynamic nature
- of mosquito antipredator responses to chemical alarm cues. Chemoecology. 2007;17(4):223-
- 437 9.
- 438 23. Ferrari MCO, Messier F, Chivers DP. Threat-sensitive learning of predators by larval
- mosquitoes Culex restuans. Behavioral Ecology and Sociobiology. 2008;62(7):1079-83.
- 440 24. Ower GD, Juliano SA. The demographic and life-history costs of fear: Trait-mediated
- effects of threat of predation on Aedes triseriatus. Ecology and Evolution. 2019;9(7):3794-
- 442 806.
- 443 25. Roux O, Vantaux A, Roche B, Yameogo KB, Dabire KR, Diabate A, et al. Evidence
- 444 for carry-over effects of predator exposure on pathogen transmission potential. Proceedings
- of the Royal Society B-Biological Sciences. 2015;282(1821):20152430.
- van Uitregt VO, Hurst TP, Wilson RS. Reduced size and starvation resistance in adult
- 447 mosquitoes, Aedes notoscriptus, exposed to predation cues as larvae. Journal of Animal
- 448 Ecology. 2012;81(1):108-15.
- 449 27. Costanzo KS, Muturi EJ, Alto BW. Trait-mediated effects of predation across life-
- 450 history stages in container mosquitoes. Ecological Entomology. 2011;36(5):605-15.
- 451 28. Ohba S-Y, Ohtsuka M, Sunahara T, Sonoda Y, Kawashima E, Takagi M. Differential
- responses to predator cues between two mosquito species breeding in different habitats.
- 453 Ecological Entomology. 2012;37(5):410-8.
- 454 29. Buchanan AL, Hermann SL, Lund M, Szendrei Z. A meta-analysis of non-
- consumptive predator effects in arthropods: the influence of organismal and environmental
- 456 characteristics. Oikos. 2017;126(9):1233-40.
- 457 30. Kesavaraju B, Juliano SA. Nature of Predation Risk Cues in Container Systems:
- 458 Mosquito Responses to Solid Residues From Predation. Ann Entomol Soc Am.
- 459 2010;103(6):1038-45.
- 460 31. Kesavaraju B, Juliano SA. Behavioral Responses of Aedes albopictus to a Predator
- 461 Are Correlated with Size-Dependent Risk of Predation. Annals of the Entomological Society
- 462 of America. 2008;101(6):1150-3.
- 463 32. van Uitregt VO, Hurst TP, Wilson RS. Greater costs of inducible behavioural
- defences at cooler temperatures in larvae of the mosquito, Aedes notoscriptus. Evolutionary
- 465 Ecology. 2013;27(1):13-26.
- 466 33. Blackmore MS, Lord CC. The relationship between size and fecundity in Aedes
- albopictus. Journal of Vector Ecology. 2000;25(2):212-7.
- 468 34. Hatala AJ, Harrington LC, Degner EC. Age and Body Size Influence Sperm Quantity
- in Male Aedes albopictus (Diptera: Culicidae) Mosquitoes. Journal of Medical Entomology.
- 470 2018;55(4):1051-4.
- 471 35. Carrasquilla MC, Lounibos LP, Honorio NA, Murr S. Spermathecal Filling in Aedes
- aegypti and Aedes albopictus: Effects of Female and Male Body Sizes and Species. Journal
- 473 of Medical Entomology. 2019;56(2):334-40.

- 474 36. Armbruster P, Hutchinson RA. Pupal mass and wing length as indicators of fecundity
- in Aedes albopictus and Aedes geniculatus (Diptera : culicidae). Journal of Medical
- 476 Entomology. 2002;39(4):699-704.
- 477 37. Xue RD, Barnard DR, Muller GC. Effects of Body Size and Nutritional Regimen on
- 478 Survival in Adult Aedes albopictus (Diptera: Culicidae). Journal of Medical Entomology.
- 479 2010;47(5):778-82.
- 480 38. Alto BW, Reiskind MH, Lounibos LP. Size Alters Susceptibility of Vectors to
- 481 Dengue Virus Infection and Dissemination. American Journal of Tropical Medicine and
- 482 Hygiene. 2008;79(5):688-95.
- 483 39. Farjana T, Tuno N. Multiple Blood Feeding and Host-Seeking Behavior in Aedes
- aegypti and Aedes albopictus (Diptera: Culicidae). Journal of Medical Entomology.
- 485 2013;50(4):838-46.
- 486 40. Leisnham PT, Sala LM, Juliano SA. Geographic variation in adult survival and
- reproductive tactics of the mosquito Aedes albopictus. Journal of Medical Entomology.
- 488 2008;45(2):210-21.
- 489 41. Alto BW, Griswold MW, Lounibos LP. Habitat complexity and sex-dependent
- 490 predation of mosquito larvae in containers. Oecologia. 2005;146(2):300-10.
- 491 42. Cardoso HCB, da Silva BQ, de Assis TB, Lopez LCS. Effects of predation by the
- 492 copepod Mesocyclops ogunnus on the sex ratios of mosquito Aedes albopictus.
- 493 Hydrobiologia. 2013;705(1):55-61.
- 494 43. Dieng H, Boots M, Mwandawiro C, Satho T, Hasegawa M, Nyambura GJ, et al.
- 495 Effects of a copepod predator on the survivorship and development of Aedes albopictus
- 496 (Diptera: Culicidae). Medical Entomology and Zoology. 2003;54(2):187-92.
- 497 44. Peters RH. Animal abundance. In: Beck E, Birks HJB, Connor EF, editors. The
- ecological implications of body size. Cambridge: Cambridge University Press; 1983. p. 164 83.
- 500 45. Metelmann S, Caminade C, Jones AE, Medlock JM, Baylis M, Morse AP. The UK's
- suitability for Aedes albopictus in current and future climates. Journal of the Royal Society
- 502 Interface. 2019;16(152).
- 503 46. Russell MC, Qureshi A, Wilson CG, Cator LJ. Size, not temperature, drives cyclopoid
- 504 copepod predation of invasive mosquito larvae. PLOS ONE. 2021;16(2):e0246178.
- 505 47. Suarez MF, Marten GG, Clark GG. A Simple Method for Cultivating Fresh-Water
- 506 Copepods used in Biological-Control of Aedes-Aegypti. Journal of the American Mosquito
- 507 Control Association. 1992;8(4):409-12.
- 508 48. Donaldson GC, Keatinge WR, Nayha S. Changes in summer temperature and heat-
- related mortality since 1971 in North Carolina, South Finland, and Southeast England.
- 510 Environmental Research. 2003;91(1):1-7.
- 511 49. Bastin JF, Clark E, Elliott T, Hart S, van den Hoogen J, Hordijk I, et al.
- 512 Understanding climate change from a global analysis of city analogues. Plos One.
- 513 2019;14(7).
- 50. Gosling SN, McGregor GR, Paldy A. Climate change and heat-related mortality in six
- 515 cities Part 1: model construction and validation. International Journal of Biometeorology.
- 516 2007;51(6):525-40.
- 517 51. Teng HJ, Apperson CS. Identification of larval instars of Aedes albopictus (Skuse)
- and Aedes triseriatus (Say) (Diptera: culicidae) based on head capsule size. Journal of Vector
- 519 Ecology. 1996;21(2):186-91.
- 520 52. Siegel JP, Novak RJ, Ruesink WG. RELATIONSHIP BETWEEN WING LENGTH
- 521 AND DRY-WEIGHT OF MOSQUITOS. Journal of the American Mosquito Control
- 522 Association. 1994;10(2):186-96.

- 523 53. R Core Team. R: A language and environment for statistical computing. Vienna,
- Austria: R Foundation for Statistical Computing; 2020.
- 525 54. Christensen BM, Li JY, Chen CC, Nappi AJ. Melanization immune responses in
- mosquito vectors. Trends Parasitol. 2005;21(4):192-9.
- 527 55. Lai SC, Chen CC, Hou RF. Immunolocalization of prophenoloxidase in the process of
- wound healing in the mosquito Armigeres subalbatus (Diptera: Culicidae). Journal of
- 529 Medical Entomology. 2002;39(2):266-74.
- 530 56. Nakhleh J, El Moussawi L, Osta MA. The Melanization Response in Insect Immunity.
- In: Ligoxygakis P, editor. Insect Immunity. Advances in Insect Physiology. 522017. p. 83-
- 532 109.
- 533 57. Chapman RF, editor. The insects: structure and function: Cambridge University Press;
- 534 1998.
- 535 58. Lee KP, Simpson SJ, Wilson K. Dietary protein-quality influences melanization and
- immune function in an insect. Functional Ecology. 2008;22(6):1052-61.
- 537 59. Awasthi AK, Wu CH, Tsai KH, King CC, Hwang JS. How Does the Ambush
- Predatory Copepod Megacyclops formosanus (Harada, 1931) Capture Mosquito Larvae of
- 539 Aedes aegypti? Zoological Studies. 2012;51(7):927-36.
- 540 60. Puggioli A, Carrieri M, Dindo ML, Medici A, Lees RS, Gilles JRL, et al.
- Development of Aedes albopictus (Diptera: Culicidae) Larvae Under Different Laboratory
- 542 Conditions. Journal of Medical Entomology. 2017;54(1):142-9.
- 543 61. Briegel H, Timmermann SE. Aedes albopictus (Diptera: Culicidae): Physiological
- aspects of development and reproduction. Journal of Medical Entomology. 2001;38(4):566-
- 545 71.

- 546 62. Wormington JD, Juliano SA. Sexually dimorphic body size and development time
- 547 plasticity in Aedes mosquitoes (Diptera: Culicidae). Evolutionary Ecology Research.
- 548 2014;16(3):223-34.
- 63. Mercer DR, Wettach GR, Smith JL. Effects of larval density and predation by
- Toxorhynchites amboinensis on Aedes polynesiensis (Diptera : culicidae) developing in
- coconuts. Journal of the American Mosquito Control Association. 2005;21(4):425-31.
- 552 64. Kesavaraju B, Juliano SA. No Evolutionary Response to Four Generations of
- Laboratory Selection on Antipredator Behavior of Aedes albopictus: Potential Implications
- for Biotic Resistance to Invasion. Journal of Medical Entomology. 2009;46(4):772-81.