

1 **Host genomic influence on bacterial composition in the switchgrass rhizosphere**

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29

30 **Abstract**

31 Host genetic variation can shape the diversity and composition of associated microbiomes, which may reciprocally
32 influence host traits and performance. While the genetic basis of phenotypic diversity of plant populations in nature
33 has been studied, comparatively little research has investigated the genetics of host effects on their associated
34 microbiomes. Switchgrass (*Panicum virgatum*) is a highly outcrossing, perennial, grass species with substantial
35 locally adaptive diversity across its native North American range. Here, we compared 383 switchgrass accessions in
36 a common garden to determine the host genotypic influence on rhizosphere bacterial composition. We hypothesized
37 that the composition and diversity of rhizosphere bacterial assemblages would differentiate due to genotypic
38 differences between hosts (potentially due to root phenotypes and associated life history variation). We observed
39 higher alpha diversity of bacteria associated with upland ecotypes and tetraploids, compared to lowland ecotypes
40 and octoploids, respectively. Alpha diversity correlated negatively with flowering time and plant height, indicating
41 that bacterial composition varies along switchgrass life history axes. Narrow-sense heritability (h^2) of the relative
42 abundance of twenty-one core bacterial families was observed. Overall compositional differences among tetraploids,
43 due to genetic variation, supports wide-spread genotypic influence on the rhizosphere microbiome. Lastly, a
44 genome-wide association study identified 1,861 single-nucleotide polymorphisms associated with 110 families and
45 genes containing them related to potential regulatory functions. Our findings suggest that switchgrass genomic and
46 life-history variation influences bacterial composition in the rhizosphere, potentially due to host adaptation to local
47 environments.

48

49 **Introduction**

50 Macroscopic organisms host diverse microbiomes, which often differ substantially among individuals
51 (Smith et al., 2015; Trivedi et al., 2020; Wilson et al., 2020). Some of the proximate causes of this variation in
52 microbiome composition arise from differences between host environments and traits, the latter of which can be
53 divided into genetic and plastic components. Host genetic differences can affect a wide range of traits that influence
54 associated microbiome composition, including traits that determine the supply of resources used by microbes or
55 traits that control environmental variables affecting microbial fitness in other ways (e.g. pH) (Lebeis et al., 2015;
56 Sasse et al., 2018). These host genetic influences on microbiomes may arise from different evolutionary processes.

57 For example, host traits that affect microbiome composition may evolve neutrally or in response to non-microbial
58 components of the local environment (Zeng et al., 2015). Alternatively, host trait variation could have evolved as a
59 direct response to beneficial interactions with microbes in the environment (Gould et al., 2018). In the latter case,
60 hosts could either influence the abundance of microbes that are simply present in the environment, and functionally
61 important to the host, or hosts could form species or strain-specific symbiotic relationships where the host depends
62 on and influences the abundance of those taxa for reproductive success (Parker et al., 2006).

63 Because of environmental and dispersal limitations, the same microbes may not be present across the entire
64 range of a host, although functionally similar ones might be. It has been shown that reciprocal interactions between
65 host immune systems and host-associated microorganisms can lead to preferential recruitment of beneficial
66 microbial types and repel pathogenic types, to varying degrees (Hacquard et al., 2017; Jones et al., 2019). This
67 suggests that hosts might (directly or indirectly) select for functional microbial traits from certain lineages. Several
68 studies have observed the presence of ‘core microbiota’ – persistent members of the microbiome that appear across a
69 large portion of a host’s population or species (Risely, 2020), but it is often unclear whether these microorganisms
70 are functionally important or commensal to the host. Yet, because microbial traits are often phylogenetically
71 conserved and host selection of microbes can occur in related microbial lineages (Lemanceau et al., 2017; Martiny et
72 al., 2015), a core microbiome might be preserved for higher taxonomic levels but undetectable for lower ones, at the
73 same occupancy frequency threshold. The study of diverse host genotypes and phenotypes in common environments
74 could reveal the genetic and host physiological drivers of variation in associated microbiomes and, potentially, types
75 of microbes that are functionally important to the host.

76 The demonstrated influence of host trait variation on microbial assemblages (Hestrin et al., 2021; Jones et
77 al., 2019; Wagner et al., 2016) suggests that microbiome composition can be treated as an extension of host
78 phenotype, one that may be under selective pressure by both the host and the environment (Bordenstein & Theis,
79 2015; Hunter, 2018; Moran & Sloan, 2015; Wagner et al., 2016; Whitham et al., 2003). Genome-wide association
80 studies (GWAS) have shown that the relationship between hosts and specific microbial taxa can be linked to single-
81 nucleotide polymorphisms (SNPs) (Beilsmith et al., 2019). However, at the microbiome level, it becomes
82 challenging to directly link host genotype and microbial composition, due to the high diversity and dimensionality of
83 microbiomes (Wray et al., 2013). Common garden experiments and genetic mapping offer a window into host

84 genetic impacts on host-associated microbial composition. The resolution of association mapping can be improved
85 by exploiting high levels of genomic diversity within and between populations (Holland, 2007).

86 Switchgrass (*Panicum virgatum*) is a C4 perennial grass species that has been extensively studied as a
87 bioenergy crop since 2005 (Bouton, 2007; McLaughlin & Kszos, 2005; Parrish & Fike, 2005; Sanderson et al.,
88 2006). Unlike first generation biofuels, switchgrass can be grown on marginal land with minimal inputs while also
89 providing ecological services (e.g. wildlife habitat and soil erosion mitigation) (Werling et al., 2014). In addition to
90 its ecological and agricultural value, switchgrass provides an ideal system for studying the influence of host
91 genotypic diversity on rhizosphere bacterial composition (i.e., bacteria living on or in close proximity to the root
92 system of the plant) because of the high degree of genotypic diversity in the species.

93 Switchgrass is primarily an outcrossing species with high genetic diversity within populations, showing
94 evidence of inbreeding depression, which increases heterozygosity in the genome (Sharma et al., 2012). The species
95 is comprised of two locally-adapted ecotypes, upland and lowland, based on differences in phenotype, physiology,
96 and habitat (Das et al., 2004). Separate migration events fostered by interglacial periods, in addition to gene
97 duplication through polyploidization, have also played a major role in shaping the genomic diversity of switchgrass
98 (Casler et al., 2015). Lowland switchgrass is primarily allotetraploid (4X: n = 36) and exhibits disomic inheritance
99 – chromosome inheritance that follows diploid association patterns. Upland accessions tend to vary in their ploidy
100 level (4X: n = 36, 6X: n = 54, 8X: n = 72) and exhibit widespread aneuploidy between and within localized
101 populations (Costich et al., 2010). Additionally, populations of switchgrass are adapted to local climate conditions
102 and linkage mapping suggests a polygenic basis to this local adaptation (Lowry et al., 2019). Here, we studied
103 genotypes largely from the geographically Northeastern, upper Southern, and Midwestern United States that can be
104 genetically clustered into five groups (roughly North, South, East, West, and Northeast, with some geographic
105 overlap) (Supplemental Figure 1) (Lu et al., 2013). Genetic variation in switchgrass has been linked to measures of
106 plant fitness and performance, including biomass production (Lowry et al., 2019) and soil carbon inputs (Adkins et
107 al., 2016; Stewart et al., 2017). While substantial research has explored the switchgrass microbiome, key gaps
108 remain, including the functional role of non-fungal microorganisms and host mechanisms that underly switchgrass–
109 microbiome interactions (Hestrin et al., 2021).

110 Vascular plants influence microbes in the rhizosphere through rhizodeposits (Dennis et al., 2010) and
111 carbon-based root exudates (Compant et al., 2010; Hu et al., 2018; Yu & Hochholdinger, 2018). Root exudates

112 provide signaling and substrate compounds for beneficial microbial recruitment (Carvalhais et al., 2013;
113 Lakshmanan et al., 2012), and antibiotic compounds that defensively modulate the microbiome composition against
114 pathogenic taxa (Baetz & Martinolia, 2014; Lebeis et al., 2015). When available, root exudates potentially represent
115 a larger portion of carbon in the soil compared to other carbon inputs from the environment (e.g. leaf litter)
116 (Heijboer et al., 2018). Switchgrass can benefit from a range of microbes that are influenced by root exudates
117 (Hestrin et al., 2021).

118 In this study, we analyzed bacterial 16S rRNA gene sequences from total DNA extracted from rhizosphere
119 soil samples for 383 switchgrass accessions from 63 local populations planted within a common garden in Ithaca,
120 NY, USA. Our aim was to better understand the influence of switchgrass genotype on the composition of bacteria in
121 the rhizosphere. We hypothesized that overall bacterial composition and diversity within the growing site would
122 differentiate due to host genetic variation along the three dimensions of switchgrass genetic diversity (ecotype,
123 ploidy level, and genetic cluster), as is true for multiple fitness-related traits (Lovell et al., 2021). Separately,
124 bacteria experience higher rates of gene loss, horizontal gene transfer, and shorter generation times compared to
125 their hosts. So, microbial specialization on a given host can be evolutionarily fast compared to changes in the plant
126 community (Bever et al., 2012). Given that the evolutionary response rate is mismatched between switchgrass
127 genotypes and their microbiomes, we sought to determine the lowest taxonomic level for which a potential host
128 influence could be detected. We show that switchgrass genotype influences bacteria in the rhizosphere at, at least,
129 the family level. Our findings demonstrate how plant host genotype can influence host-associated microbiomes, the
130 taxonomic level at which the microbiome is influenced, and the host genetic architecture involved.

131

132 Materials and Methods

133 Switchgrass Germplasm Data

134 Previously, exome capture sequencing was used to characterize variation in the genic regions of the
135 genome from the Northern Switchgrass Association Panel used in this study ($n = 537$) (Evans et al., 2018). For this
136 study, the HapMapv2 matrix was obtained from the Dryad Digital Repository (10.5061/dryad.mp6cp) in 2019.
137 Several modifications were made to the original dataset. The dataset was modified in *python* (G. Van Rossum &
138 Drake, 2009) to reflect heterozygote positions more conservatively than the original, unfiltered, dataset, such that
139 heterozygote positions with read count ratios greater than 0.75 or less than 0.25 were called homozygous relative to

140 the appropriate allele (e.g. A(20)/T(1) = A). SNPs with a minor-allele frequency (MAF) below 0.05 were excluded,
141 resulting in 103,776 SNPs for all genotypes studied here ($n = 362$) spanning eighteen chromosomes: Chr01- 09
142 (subgenomes a and b) of the v1.1 *P. virgatum* reference genome (www.phytozome.net) and fifteen unanchored
143 scaffolds (Evans et al., 2018). The unanchored scaffolds of Evans et al. (2018) were renamed to the corresponding
144 contig names within the v1.1 reference genome using *python*. Phylogenetic tree generated in Tassel v5 (Bradbury et
145 al., 2007).

146 We compared bacterial composition to published data on several important switchgrass functional and life
147 history traits, using data from 2009, 2010, and 2011 field seasons at the common garden in Ithaca, NY (genetic
148 variation in these traits is largely consistent from year to year) (Lipka et al., 2014). The 2011 measurements for
149 anthesis date and full plant height were used to identify correlations with rhizosphere alpha diversity present in
150 2016.

151

152 **Rhizosphere sampling of switchgrass accessions**

153 The original switchgrass association panel consisted of 3-10 clonally propagated individuals each from 66
154 discrete switchgrass local populations grown from seed in the greenhouse of the USDA-ARS Dairy Forage Research
155 Center in Madison WI in 2007 (Lu et al., 2013). The accessions were sourced from much of the natural range of
156 switchgrass in the eastern and midwestern United States. Young plants were deposited in the common garden in
157 Ithaca, NY, USA in 2008. In 2016, plugs were collected for transplanting from 525 cloned accessions, representing
158 all but two (perished) original populations. Prior to replanting the plugs, soil samples were collected into a
159 polythene bag from each of the switchgrass accessions by vigorously shaking off soil attached to the roots. The bags
160 with soil and fine roots were initially stored at 4°C while being processed - each soil sample was thoroughly mixed
161 into a uniform distribution and then transferred to 50 ml conical cryogenic tubes and stored at -80°C prior to soil
162 DNA extraction.

163

164 **Soil DNA extractions**

165 To determine the composition of bacteria established in the switchgrass rhizosphere, we selected soil
166 samples from 383 of the 525 cloned switchgrass accessions, based on a phylogenetically determined set of clones
167 from 63 of the local populations in the association panel. Total DNA was extracted from ~300 mg of soil using the

168 NucleoSpin Soil 96 kit (Macherey-Nagel, Düren, Germany). Lysis was performed using the Buffer SL with
169 Enhancer SX, and was performed on the FastPrep 24 homogenizer (MP Biomedicals, Santa Ana, CA, USA) at 4.0
170 m/s for 30 s.

171

172 Amplicon Sequencing

173 Briefly, amplicons targeting the V3-V4 region of the 16S rRNA gene were generated from all 383 samples
174 using universal bacterial primers 515F (5'-GTGYCAGCMGCCGCGTAA-3') and 806R (5'-
175 GGAATACNVGGGTWTCTAAT-3') (Apprill, McNally, Parsons, & Weber, 2015; Parada, Needham, & Fuhrman,
176 2016) with overhangs for attaching barcodes and standard Illumina overhang adaptors in a second PCR step (full
177 protocol outlined in the supplemental methods of Trexler & Bell (2019)). Sequencing was performed on an Illumina
178 MiSeq using the 2 × 250 cycle v2 kit. Raw reads are available through NCBI SRA under project number
179 PRJNA689762.

180

181 Amplicon sequence analysis pipeline

182 The raw 16S rRNA gene sequences were analyzed using an adapted version of the *dada2* pipeline
183 (Callahan et al., 2016). *dada2* was used to process the raw sequences into Amplicon Sequence Variants (ASVs).
184 Reads were truncated above 240 bp and below 160 bp. Sequences with any missing reads after truncation were
185 discarded. Reads were then truncated at the first instance of a quality score less than 2. After truncation, reads that
186 matched against the phiX Genome were discarded. Reads with higher than two "expected errors" were also
187 discarded. Expected errors are calculated from the nominal definition of the quality score: $EE = \text{sum}(10^{(Q/10)})$.
188 Filtered sequences were used to determine the error rate using the *dada2* function: *learnErrors()*. The filtered
189 sequences were then processed with the core sample inference algorithm, *dada()*, incorporating the learned error
190 rates. Forward and reverse sequence reads were then merged. A sequence table was made and chimeras were
191 removed using the "consensus" method. Taxonomic assignments for the ASVs were defined against the SILVA 138
192 ribosomal RNA gene database (Quast et al., n.d.) using DECIPHER v2.14.0 (Wright, n.d.). The sequence variant
193 table and taxonomy table were exported for downstream processing in *phyloseq* (McMurdie & Holmes, 2013).

194 In *phyloseq*, ASVs designated "NA" at the phylum level, non-Bacterial entries at the domain level, and
195 those associated to chloroplasts and mitochondria were removed from the taxonomy file before further processing.

196 Finally, we also removed samples with no ASVs remaining after pruning. Rarefying samples has been shown to
197 effectively reduce false discovery rates when there are large differences between the average sample library size
198 (Weiss et al., 2017). Our data was characteristic of this scenario. Therefore, we rarefied samples to 1000 sequences,
199 to retain as many samples as possible while also maintaining a sufficient sampling depth to detect the most prevalent
200 taxa comprising the switchgrass core microbiome (See below).

201

202 **Alpha diversity analysis**

203 Using the rarefied ASV dataset, we generated alpha diversity metrics using *phyloseq* (McMurdie &
204 Holmes, 2013). Statistical significance was calculated using the vegan 2.5.6 package (Oksanen et al., 2019), and
205 other core functions in R version 3.6.3 (Ihaka & Gentleman, 1996). The Chao1 and Shannon diversity indices were
206 calculated using the estimate_richness() function in *phyloseq* and then used to test for differences in alpha diversity
207 between genotypes differing in ecotype, ploidy level, and genetic cluster groups using an analysis of variance
208 (ANOVA) (R version 3.6.3). Correlations between diversity metrics, anthesis date and full plant height were
209 generated using the Pearson correlation test.

210

211 **Beta diversity and core microbiome analysis**

212 Bacterial diversity data rarely conform to the assumptions of MANOVA-like procedures, largely due to
213 inflated zero values among rare taxa that skew the distribution, thus non-parametric methods based on permutation
214 tests are preferred (M. J. Anderson, 2001). A non-parametric, one-way analysis of variance (NPMANOVA) is also
215 tolerant of non-independent observations (e.g. microbe-microbe interactions). We therefore calculated an
216 NPMANOVA using the *adonis2* function in vegan to test for significant differences between the ecotype, ploidy
217 level, and genetic cluster groups on the overall rhizosphere microbiome composition using pairwise Bray-Curtis
218 distances at the ASV level.

219 The ‘core microbiome’ can be defined as microbial taxa that associate with a host above a particular
220 occupancy frequency threshold (i.e. their prevalence). Biological justifications for such thresholds tend to be
221 subjective, often ranging between 30% and 95% (Risely, 2020). However, core taxa prevalence within a population
222 could be a reflection of an environmental response by the host due to local adaptation, resulting in higher intergroup
223 prevalence of certain ‘core taxa’ relative to the whole population. Thus, we determined two core microbiomes: A

224 high-fidelity core (prevalence $\geq 90\%$), to determine taxa that reliably associate with the vast majority of our panel,
225 and a low-fidelity core (prevalence $\geq 10\%$), to study host genetic variation effects on bacterial composition. Taxa
226 present within our rarefied dataset were initially agglomerated using the `tax_grom()` function at all taxonomic levels
227 in *phyloseq*. Sample counts were then transformed to relative abundances. A filter was applied to subset
228 phylogenetic groups in abundance greater than 2%. Each core microbiome was determined using the *microbiome*
229 package in R (Lahti, Shetty, & Blake, 2017). The `core()` function in the *microbiome* package (detection = 0,
230 prevalence = 0.9 and 0.1, respectively) was applied at each taxonomic level to determine the core taxa among all
231 genotypes.

232

233 Heritability Estimates

234 Heritability is an estimate that describes the proportion of phenotypic variance that is due to genetic
235 variance. Narrow-sense SNP-based heritability (h^2) was estimated using the *sommer* package in R as: σ_A^2 / σ_p^2 , where
236 σ_A^2 is the additive variance and σ_p^2 is the total phenotypic variance (additive + dominance) (Covarrubias-Pazaran,
237 2016). The core function of the *sommer* package is the ‘`mmer`’ function that fits multivariate linear mixed models.
238 The modified exome capture SNP dataset, described above, was used to generate a kinship matrix used in ‘`mmer`’ to
239 estimate narrow-sense heritability for each of bacterial families’ abundance. The *sommer* package includes a
240 tolerance parameter for the matrix inverse when singularities are encountered in the estimation procedure. Here,
241 inversion tolerance parameters were adjusted to 10 to avoid model singularity.

242

243 Redundancy Analysis (RDA) and variance partitioning

244 To determine how whole microbiome variation changed across host genomic variation, much of which is
245 due to population structure, we implemented redundancy analysis (RDA). RDA models variation in a set of
246 response variables as a function of a set of explanatory variables. We used the first 10 principal components of
247 switchgrass SNPs (Evans et al. 2014) as explanatory variables that describe population structure in switchgrass
248 (Brown, Bray, & Pachter, 2018) and log-scaled ASV counts for each switchgrass genotype as the response variables.
249 We performed the RDA to explain the relative proportion of total genotypic variance on the overall microbiome
250 composition. The `rda()` function in *vegan* (Oksanen et al., 2019) was used to perform the RDA for the tetraploid
251 accessions. For host genomic analyses like this RDA, we focused on tetraploids; since they (1) represent both upland

252 and lowland ecotypes, (2) exhibit disomic inheritance unlike octoploids, and (3) exhibit more inter-group alpha
253 diversity variation compared to octoploids.

254 We identified outlier ASVs with the strongest loadings, based on inter-quartile range criteria, on the first
255 two RDA canonical axes; these ASVs are those most representative of microbiome-wide turnover across
256 switchgrass hosts of different genomic background. ASVs that were labeled as outliers were then assigned
257 taxonomic rank using the *phyloseq* method described above and compared to families in the original ASV dataset.

258 To dissect how genomic variation combined with ecotype, ploidy, and phenotypes to explain bacterial
259 compositional turnover we also implemented variance partitioning of the RDA. Variance partitioning allows one to
260 estimate the portions of compositional turnover explained by multiple sets of factors (Peres-Neto et al., 2006), which
261 here were the first 10 principal components of SNPs, ecotype, geography-of-origin (latitude, longitude, and their
262 squared values), and our two focal phenotypes (anthesis date and height). We implemented two versions of variance
263 partitioning with RDA: one on ASV counts and the other on family abundances, to test whether family-level
264 composition was differently associated with these factors than was ASV level composition.

265

266 **Genome-wide association study (GWAS)**

267 To identify specific loci in the switchgrass genome linked to variation in rhizosphere bacterial composition,
268 a genome-wide association study (GWAS) was conducted using the *statgenGWAS* package in R (Rossum et al.,
269 2020). This fast single trait GWAS method was developed by Biometris, following the method described in Kang et.
270 al (2010). The modified SNP dataset, described above, was once again used for this analysis.

271 Family-level abundance counts data (1) were imported from *phyloseq* using the rarified *phyloseq* object
272 and a chromosome positional map (2) was generated from the SNP matrix (3). These three components were used to
273 construct the R object used by *statgenGWAS*. Initial results from the heritability estimates and RDA indicated that
274 certain core families of bacteria were influenced by switchgrass genotype. Therefore, to determine associated genes,
275 a single trait GWAS, where the trait was the abundance of each of the families, was completed using a Generalized
276 Least Squares (GLS) method for estimating the marker effects and corresponding p-values. Our GWAS included
277 random effects correlated according to the kinship matrix, which was calculated with the VanRaden method
278 (VanRaden, 2008). A p-value threshold is required for this method to minimize the False Discovery Rate (FDR)
279 following the algorithm proposed by Brzyski et al. (2017). The method limits the number of SNPs in statistical

280 linkage with each other passing FDR control. Significant SNPs were identified using a 0.01 threshold for the false-
281 discovery rate (FDR). Linkage disequilibrium (LD) in switchgrass decays over kilobase length. Following
282 Grabowski et al. (2017), we used a window of 25kb to identify genes potentially linked to significant SNPs. Thus, in
283 this study, we defined SNPs within 25 kb as being “within LD of significant SNPs”. For each trait, *statgenGWAS*
284 outputs significant SNPs and SNPs within the defined cutoff. A full description of the method can be found in
285 Rossum et al. (2020).

286

287 **GO Enrichment Analysis**

288 Significant SNPs associated with bacterial family abundance and positions within 25 kb of significant
289 SNPs were then used to identify associated genes and gene ontologies. Here, we focused on three families:
290 Xanthobacteraceae, Sphingomonadaceae, and Micromonosporaceae. The Xanthobacteraceae and
291 Sphingomonadaceae families were chosen because they represent the high-fidelity core microbiome.
292 Micromonosporaceae was chosen due to its high relative heritability within our panel and its importance in
293 agriculture applications (Trujillo, Hong, & Genilloud, 2014). *Phytozome* was used to identify Gene IDs for each
294 family using the *P. virgatum v1.1* reference genome at www.phytozome.net. Gene IDs for corresponding SNPs were
295 then used to perform a Singular Enrichment Analysis (SEA) against the gene ID (ver. 4) background for *P. virgatum*
296 using the agrigo v2 web program (Du, Zhou, Ling, Zhang, & Su, 2010). Default parameters were used.

297

298 **Results**

299 **Amplicon Sequence Analysis**

300 For 383 initial rhizosphere soil samples, 96,902 amplicon sequences variants (ASVs) were obtained
301 following initial quality filtering and sequence processing in *dada2* (Callahan et al., n.d.). After sample pruning
302 (removing non-bacteria from the taxonomy table) and rarefaction (1,000 sequences/sample) in *phyloseq*, 365
303 samples containing 31,181 ASVs remained. Counts for unique phylogenetic classifications are as follows: Domain:
304 1, Phylum: 26, Class: 59, Order: 142, Family: 268, Genus: 510.

305

306 **Alpha diversity analysis**

307 We detected significant differences in alpha diversity (Chao1 and Shannon Diversity) between groups.
308 Upland ecotypes, which are adapted to regions at higher latitudes and those geographically closer to our growing
309 site, exhibited higher rhizosphere diversity compared to lowland ecotypes (Figure 1A, Upland-Lowland: $p.adj$
310 (Shannon) = $1.58e^{-2}$). Tetraploids, which are present in both upland and lowland ecotype groups, also exhibited
311 higher diversity overall, compared to octoploids, which are exclusively upland (Figure 1B, 4X-8X: $p.adj$ (Shannon)
312 = <0.001). Finally, the North genetic cluster, primarily comprised of upland tetraploids, exhibited the highest
313 rhizosphere diversity among all of the genetic clusters and there was a significant difference between the East and
314 West genetic clusters (Figure 1C, North-East: $p.adj$ (Shannon) = <0.001 , North-West: $p.adj$ (Shannon) = <0.001 ,
315 North-Northeast: $p.adj$ (Shannon) = <0.001 , North-South: $p.adj$ (Shannon) = <0.001 , East-West: $p.adj$ (Shannon) =
316 $5.92e^{-2}$). All diversity measurements for accessions can be found in Supplemental Table 1.

317 We found that Chao1 and Shannon diversity were negatively correlated with anthesis date (Chao1: $r = -$
318 0.17 , p-value = 0.0014; Shannon: $r = -0.18$, p-value = <0.001) and full plant height (Chao1: $r = -0.18$, p-value =
319 0.0012 ; Shannon: $r = -0.13$, p-value = 0.017). We also considered these trait-diversity associations stratified by
320 ecotype, ploidy, and genetic cluster (Figure 2). In general, the negative diversity association with anthesis date and
321 height was broadly consistent, with Shannon diversity negatively correlated with plant height for Upland ecotypes (r
322 = -0.25 , p-value = <0.001), octoploids ($r = -0.23$, p-value = 0.0045), hybrids ($r = -0.73$, p-value = 0.016), the North
323 genetic cluster ($r = -0.27$, p-value = 0.011), the Northeast genetic cluster ($r = -0.22$, p-value = 0.061), and the West
324 genetic cluster ($r = -0.24$, p-value = 0.032) (Figure 2A,B,C). Likewise, Shannon diversity negatively correlated with
325 anthesis date for Upland ecotypes ($r = -0.19$, p-value = 0.002), tetraploids ($r = -0.23$, p-value = 0.0025), hybrids ($r =$
326 -0.94 , p-value = <0.001), and the Northeast genetic cluster ($r = -0.45$, p-value = <0.001) (Figure 2D,E,F). A subset
327 ($n = 7$) of ecotype/ploidy/genetic cluster combinations showed positive trait correlations with Shannon diversity, but
328 these were not significant.

329

330 Beta diversity analysis

331 We performed separate multivariate analyses of variance on a Bray-Curtis distance matrix using the
332 NPMANOVA test. There were significant differences in overall bacterial composition between ecotypes ($p <$
333 0.001), ploidy levels ($p < 0.001$), and genetic clusters ($p < 0.001$), indicating differences in overall composition
334 associated with switchgrass genetics and life-history.

335 Although all taxonomic levels were tested, the family level was identified as the lowest taxonomic rank
336 with the ability to retain a high-fidelity core (Supplemental Figure 2). The high-fidelity core microbiome (>=90% of
337 samples) contained two families: Xanthobacteraceae and Sphingomonadaceae. The low-fidelity core microbiome
338 (>=10% of samples) contained 110 families (Supplemental Table 2).

339

340 **Host Genomic Influence**

341 Of the 110 families in the core microbiome, twenty-one showed narrow-sense SNP-based heritability (h^2)
342 significantly greater than zero (Figure 3). The standard errors of narrow-sense heritability estimates are very large
343 and there is considerable uncertainty in the estimate overall (Furlotte, Heckerman, & Lippert, 2014). Therefore, we
344 consider estimates with standard errors that do not intersect zero as showing evidence of heritability (i.e. heritable).
345 Heritability estimates for low-fidelity core taxa in the switchgrass rhizosphere ranged from 0.106 to 0.539, with a
346 mean of 0.241. Heritability estimates presented here for bacterial lineages have a range similar to that of corn and
347 sorghum (Deng et al., 2021). The heritability estimates indicate that, in part, switchgrass genotype influences the
348 variability in the relative abundance of certain families within our growing site. Heritability estimates for all
349 bacterial families can be found in Supplemental Table 2.

350 Next, we performed a redundancy analysis (RDA) to characterize how the whole rhizosphere bacterial
351 assemblage changes in association with host genomic background (Figure 4, Supplemental Figure 3). The RDA
352 results indicate that genotypic variability among tetraploids (i.e. first 10 PCs of SNPs) had modest influences on
353 bacterial ASV composition ($R^2 = 0.063$, $R^2_{\text{adjusted}} = 0.007$). Switchgrass SNP PC1, which is associated with
354 separation between the North and the Northeast and South genetic clusters, loaded most strongly to the first primary
355 canonical axis (RDA1, PC1 = 0.99, the next strongest loaded PC was PC10, with a 0.097 loading). Switchgrass SNP
356 PC7, which is associated the variance in the North genetic cluster, loaded most strongly to the second canonical axis
357 (RDA2, PC7 = 0.899). The proportion of explained variance in bacterial ASV composition was fairly low along
358 these individual canonical axes (RDA1 = 0.9%, RDA2 = 0.7%), which was unsurprising given the large diversity in
359 the bacterial composition. Still, the RDA identified bacterial ASV compositional turnover in association with
360 switchgrass population structure, as shown by the turnover between the North and the Northeast and South
361 switchgrass genetic clusters (Figure 4).

362 Outlier ASVs were determined for each of the first two canonical RDA axes (n = 9,215). These outlier
363 ASVs are most representative of the variation in bacterial composition among the tetraploid genotypes. We then
364 identified the taxonomic ranks for the outlier ASVs. Our results show that Xanthobacteraceae and
365 Sphingomonadaceae represent the high-fidelity core for both the outlier ASV dataset and the original ASV dataset,
366 signifying host genotypic influence might be strongest on these families. When we consider the low-fidelity core (n
367 = 110 families), 70.9% of families in the RDA outlier ASV core microbiome are in original core microbiome
368 (Supplemental Table 2). When we consider just the heritable families, 80.95% of heritable families in the outlier
369 ASV core microbiome are in the original core microbiome (Supplemental Table 2).

370 When we applied variance partitioning to estimate the portion of ASV-level compositional turnover
371 explained by SNP PCs, ecotype, geographic origin (i.e. latitude and longitude coordinates), and the two phenotypes,
372 we found that the total R^2_{adjusted} was low, approximately 1% of ASV composition. However, variance partitioning of
373 family-level turnover found these 4 factors explained a larger portion, with $R^2_{\text{adjusted}} = 0.096$ (Figure 4). This variance
374 partitioning found that the largest portion of bacterial compositional variation was explained by the collinear portion
375 of SNP PCs, ecotype, and geographic origin ($R^2_{\text{adjusted}} = 0.0866$), suggesting that geographic population structure
376 associated with ecotypic variation explains a large portion of turnover in bacterial family composition.

377

378 **Genome-wide association study**

379 A primary limitation of heritability estimates and the RDA was that we could not account for which SNPs
380 have the greatest effect on compositional variation. To overcome this, a genome-wide association study (GWAS)
381 was conducted to identify specific loci associated with the relative abundance of families in the core microbiome. In
382 total, 1,861 SNPs were found to be associated with the abundance of the 110 tested core families. 878 SNPs were
383 found to be associated with the abundance of the 21 heritable core families. Furthermore, 36 SNPs were found to be
384 associated with the abundance of the two families (Xanthobacteraceae and Sphingomonadaceae) that represent the
385 high-fidelity core microbiome (Figure 6, Supplemental Table 2).

386

387 **GO Enrichment Analysis**

388 Despite the general difficulty identifying specific molecular functions or pathways in non-model plant
389 species, resources for gene ontology (GO) still remain useful for biological inference (Tian et al., 2017). Therefore,

390 we completed a GO enrichment analysis for genes harboring significant SNPs and positions within 25kb of
391 significant SNPs to determine possible cellular processes (P), cellular components (C), and molecular functions (F)
392 associated with three core family abundances (Supplemental Table 3). Primarily, we identified two types of cellular
393 processes: metabolic and stimuli response processes. Broadly, we identified several molecular functions relating to
394 ATPase activity, hydrolase activity, and pyrophosphatase activity (Supplemental Figure 4, Supplemental Table 3).
395 Functions involving ATP hydrolysis as an energy source either typically catalyze a reaction or drive membrane
396 transport against a concentration gradient (Berg, Tymoczko, & Stryer, 2002). In the context of our GWAS, this
397 could suggest possible root exudate activity whereby switchgrass genotypes differentially transport exudates against
398 a lower soil compound gradient influencing core family abundances. Perturbing these regions in future studies could
399 reveal the underlying pathways and mechanisms. Lastly, genes harboring SNPs associated with Sphingomonadaceae
400 abundance, in particular, were enriched for GO terms associated with cellular membrane components,
401 metallopeptidase and metalloendopeptidase activity, and cellular processes that relate to metabolism and response to
402 stimuli (Supplemental Table 3). Again, given the context of our GWAS, we suspect that the genes associated with
403 these GO terms could interact with Sphingomonadaceae populations in the soil. More work involving these genes
404 could expose perturbations that more predictably affect Sphingomonadaceae abundance in switchgrass.

405

406 Discussion

407 Macroscopic organisms often exhibit striking variation among individuals in the microbial assemblages
408 they host, but the genetic basis and ecological relevance of this variation is often unclear. Common garden
409 experiments with diverse genotypes offer a powerful window into dissecting the ecological genomics of host
410 influences on their microbiomes. Previous studies have demonstrated that genomic differences among switchgrass
411 genotypes and populations may, in part, have led to much of the adaptive phenotypic differences observed between
412 switchgrass populations (Adkins et al., 2016; Lowry et al., 2019). We add to these observations by showing a similar
413 phenomenon within a less visible portion of the switchgrass phenotype. We found that switchgrass genotype
414 influences the rhizosphere microbiome in a variety of ways relative to bacterial alpha and beta diversity as a result of
415 variability within the switchgrass genome.

416 We demonstrated that the alpha diversity (Chao1 and Shannon diversity) of rhizosphere bacteria within a
417 common garden was significantly differentiated between switchgrass ecotypes, ploidy levels, and genetic clusters.

418 These groupings with distinct bacterial assemblages are associated with life history variation and locally-adapted
419 traits in this species. Furthermore, alpha diversity within each group typically had no correlation with, or was
420 negatively correlated with, anthesis date and plant height (Figure 2). Flowering time is an important adaptation to
421 local growing conditions and has major fitness consequences for the plant (Grabowski et al., 2017b). In general, in
422 common garden experiments, upland ecotypes have an anthesis date 2–4 weeks earlier than lowland ecotypes
423 (Schwartz & Amasino, 2013). As plants move through different developmental stages the diversity and availability
424 of metabolites influencing the rhizosphere microbiome shifts (Wagner, 2021). Rhizosphere sampling in this study
425 occurred in May, 2016. Therefore, it's reasonable to expect genotypes at different stages of development at the time
426 of sampling to exert different selective pressure on the microbiome composition. Future research examining
427 temporal exudate availability in the rhizosphere of switchgrass could expand on this hypothesis.

428 It is already widely hypothesized that hosts mediate their microbiomes relative to environmental
429 conditions, resource availability, and microbial function, not necessarily always a specific species or strain
430 (Lemanceau et al., 2017; Martiny et al., 2015). Thus, some basis for comparing taxa between host groupings
431 becomes important when determining the taxonomic level at which host influence is detectable. A “core”
432 microbiome can be defined in a variety of ways, and may not represent microbes that have the most important
433 functional impacts on a system (Bell & Bell, 2021). For example, commensal microorganisms remain widespread
434 throughout host populations (Zeng et al., 2015). Still, the consistent occurrence of microbial taxa provides an
435 opportunity to assess relationships between host genetics and microbial recruitment. Here, we applied the “common
436 core” concept (Risely, 2020) to identify bacterial taxa that were consistently associated with diverse switchgrass
437 genotypes in a common garden. Given the disparity in evolutionary pace (i.e. there are many bacterial generations
438 during the lifecycle of a switchgrass plant), it is unlikely that switchgrass has mechanisms to distinguish between
439 closely related bacterial taxa unless doing so provided a substantial fitness advantage (Yin et al., 2021). Therefore,
440 the core microbiome provides both a basis for measuring bacterial abundance across multiple switchgrass genotypes
441 as well as a means to detect host genomic influence at the lowest possible taxonomic rank.

442 For example, Singer et al. (2019) observed a large core endophytic switchgrass microbiome dominated by
443 root-colonizing bacterial genera such as *Streptomyces*, *Pseudomonas*, and *Bradyrhizobium*, while rhizosphere
444 diversity was more variable between their two sampling sites. Likewise, Singer et al. (2019b) observed different
445 core bacterial classes associated with upland and lowland switchgrass ecotypes, with each ecotype preferentially

446 enriched for Alphaproteobacteria and Actinobacteria, respectively. For our study, out of 268 families, only two
447 associated with the majority ($\geq 90\%$) of our rhizosphere samples (Xanthobacteraceae and Sphingomonadaceae) and
448 less than half ($n = 110$) associated with more than 10% of our samples. Xanthobacteraceae and Sphingomonadaceae
449 are families in the Alphaproteobacteria class. We were unable to detect any genera or species associated with our
450 genotypes at the 90% occupancy threshold. This indicates that a large swath of the switchgrass rhizosphere
451 microbiome assembly is stochastic and only certain lineages were able to consistently associate with our genotypes
452 in the common garden.

453 The abundance of twenty-one of the 110 families were determined to show evidence of heritability. After
454 reasonably establishing heritability of select core families, the RDA allowed us to identify how genome-wide
455 variation in switchgrass influenced the rhizosphere microbiome composition. RDA methods used in microbial
456 ecology typically explain the variance in microbiome composition due to environmental variables, such as
457 temperature or pH (Paliy & Shankar, 2016). However, here, we modeled microbial compositional data as a function
458 of host genomic data. As such, the RDA in this study represents the compositional variance of the microbiome due
459 to genetic variation in the hosts. We then extended the analysis to explore outlier ASVs along each of the first two
460 RDA axes, because those ASVs represent taxa associated with host genomic influence on overall turnover in
461 bacterial composition. Once clustered at the family level, we determined that seventeen out of the 21 heritable
462 families were in the outlier ASV dataset. Additionally, the results of the variance partitioning suggest that
463 geographic population structure associated with ecotypic variation explains a larger portion of turnover in bacterial
464 family composition compared to ASV composition. Thus, we determined that genotypic variability in switchgrass
465 sampled from across its natural range influences the composition of the rhizosphere bacterial microbiome,
466 particularly for tetraploids.

467 The ecophysiology of host-microbe interactions in our study is unclear, requiring detailed investigation, but
468 there may be hints in the function of switchgrass genes that are strongly associated with core family abundances. We
469 conducted a GWAS to identify potential loci that strongly associate with core family abundances. The outcrossing
470 nature of switchgrass and its rapid decay in LD can help pinpoint causal loci with GWAS. Several considerations
471 had to be made though, in order to reliably trust the significance of the resulting QTLs. First, we considered only
472 tetraploids for the same reasons as the RDA, that they (1) represent both upland and lowland ecotypes, (2) exhibit
473 disomic inheritance unlike octoploids, increasing the effect size of additive effects, and (3) exhibit the most inter-

474 group alpha diversity variation compared to octoploids or hybrids. We used exome capture SNPs (Evans et. al.
475 (2018), which are more effective than genotype-by-sequencing (GBS) in their ability to tag causal polymorphisms,
476 compared to GBS SNPs that are often far from any genes (Kaur et al., 2014). The identification of genes containing
477 significant SNPs could provide insights into potential host-microbe interaction pathways. To explore this
478 relationship further, we chose to focus on three core families: Xanthobacteraceae, Sphingomonadaceae, and
479 Micromonosporaceae.

480 We identified hundreds of switchgrass genes containing SNPs (including those within 25 kb of significant
481 SNPs) associated with Micromonosporaceae ($n = 315$), Xanthobacteraceae ($n = 578$), and Sphingomonadaceae ($n =$
482 718) abundance in the rhizosphere. Many species within the Micromonosporaceae family degrade chitin, cellulose,
483 lignin, and pectin, and play an important role in the turnover of organic plant material. Moreover, many strains
484 produce useful secondary metabolites and enzymes used by plants (Trujillo et al., 2014). Members of
485 Xanthobacteraceae have been experimentally identified as nitrate reducers (C. R. Anderson et al., 2011) and
486 therefore their regulation could be important to host metabolic activity. Our GO enrichment analysis indicates that
487 genes associated with Micromonosporaceae and Xanthobacteraceae abundance are involved in hydrolase,
488 pyrophosphatase, and ATPase activity suggesting that the enrichment for this set of genes plays an important role in
489 overall plant fitness. Members of the Sphingomonadaceae family are known to be antagonistic against plant
490 pathogens and induce plant growth promotion (Glaeser & Kämpfer, 2014). GO terms for genes associated with this
491 family are enriched for metallopeptidase and metalloendopeptidase activity, membrane cellular components, and
492 processes involved in stimuli response. Altogether, this suggests that the activity of these genes is linked to
493 environmental stimuli, possibly initiated by Sphingomonadaceae, and interacting with plant cell membranes. Future
494 work examining these families could make these connections to plant function more robust.

495 In conclusion, bacterial composition in the switchgrass rhizosphere is influenced by host genotypic
496 variability. Rhizosphere diversity shows major differences among switchgrass ecotypes, ploidy levels, genetic
497 clusters, and life-history axes. In particular, the North switchgrass genetic cluster was most distinct at our common
498 garden site. Two high-fidelity core families, Xanthobacteraceae and Sphingomonadaceae, were observed across the
499 majority of our switchgrass genotypes. Twenty-one bacterial families were found to exhibit evidence for heritability.
500 Though the explained variance was low, a redundancy analysis revealed widespread genotypic influence on the
501 composition of the rhizosphere bacterial microbiome and outlier ASVs along each primary axis belonged largely to

502 our defined core microbiome. Finally, we identified 1,861 SNPs associated with the abundance of core microbiome
503 families and discussed potential pathway mechanisms that could influence a subset of those families that we
504 expected could influence host fitness. A GO enrichment analysis determined several functional and cellular
505 processes for which host genotypic influence could be expressed, notably by hydrolase activity. This study should
506 function as a foundation on which further exploration can be achieved investigating the relationship between
507 switchgrass genotype and its associated rhizosphere microbiome in different environmental contexts or under
508 artificial genetic perturbation.

509

510

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524

525 References

- 526 Adkins, J., Jastrow, J. D., Morris, G. P., Six, J., & de Graaff, M. A. (2016). Effects of switchgrass cultivars and
527 intraspecific differences in root structure on soil carbon inputs and accumulation. *Geoderma*, 262, 147–154.
528 doi: 10.1016/j.geoderma.2015.08.019
- 529 Anderson, C. R., Condron, L. M., Clough, T. J., Fiers, M., Stewart, A., Hill, R. A., & Sherlock, R. R. (2011).
530 Biochar induced soil microbial community change: Implications for biogeochemical cycling of carbon,
531 nitrogen and phosphorus. *Pedobiologia*, 54(5–6), 309–320. doi: 10.1016/j.pedobi.2011.07.005
- 532 Anderson, M. J. (2001). A new method for non-parametric multivariate analysis of variance. *Austral Ecology*, 26(1),
533 32–46. doi: 10.1111/j.1442-9993.2001.01070.pp.x

- 534 Apprill, A., McNally, S., Parsons, R., & Weber, L. (2015). Minor revision to V4 region SSU rRNA 806R gene
535 primer greatly increases detection of SAR11 bacterioplankton. *Aquatic Microbial Ecology*. doi:
536 10.3354/ame01753
- 537 Baetz, U., & Martinoia, E. (2014, February 1). Root exudates: The hidden part of plant defense. *Trends in Plant
538 Science*, Vol. 19, pp. 90–98. doi: 10.1016/j.tplants.2013.11.006
- 539 Beilsmith, K., Thoen, M. P. M., Brachi, B., Gloss, A. D., Khan, M. H., & Bergelson, J. (2019). Genome-wide
540 association studies on the phyllosphere microbiome: Embracing complexity in host-microbe interactions. *The
541 Plant Journal*, 97(1), 164–181. doi: 10.1111/tpj.14170
- 542 Bell, T. H., & Bell, T. (2021). Many roads to bacterial generalism. *FEMS Microbiology Ecology*. doi:
543 10.1093/femsec/fiaa240
- 544 Berg, J. M., Tymoczko, J. L., & Stryer, L. (2002). *A Family of Membrane Proteins Uses ATP Hydrolysis to Pump
545 Ions Across Membranes*. Retrieved from <https://www.ncbi.nlm.nih.gov/books/NBK22464/>
- 546 Bever, J. D., Platt, T. G., & Morton, E. R. (2012). Microbial population and community dynamics on plant roots and
547 their feedbacks on plant communities. *Annual Review of Microbiology*, 66, 265–283. doi: 10.1146/annurev-
548 micro-092611-150107
- 549 Bordenstein, S. R., & Theis, K. R. (2015). Host biology in light of the microbiome: Ten principles of holobionts and
550 hologenomes. *PLoS Biology*, 13(8). doi: 10.1371/journal.pbio.1002226
- 551 Bouton, J. H. (2007, December 1). Molecular breeding of switchgrass for use as a biofuel crop. *Current Opinion in
552 Genetics and Development*, Vol. 17, pp. 553–558. doi: 10.1016/j.gde.2007.08.012
- 553 Bradbury, P. J., Zhang, Z., Kroon, D. E., Casstevens, T. M., Ramdoss, Y., & Buckler, E. S. (2007). TASSEL:
554 software for association mapping of complex traits in diverse samples. *Bioinformatics*, 23(19), 2633–2635.
555 doi: 10.1093/BIOINFORMATICS/BTM308
- 556 Brown, B. C., Bray, N. L., & Pachter, L. (2018). Expression reflects population structure. *PLoS Genetics*, 14(12),
557 e1007841. doi: 10.1371/journal.pgen.1007841
- 558 Brzyski, D., Peterson, C. B., Sobczyk, P., Candès, E. J., Bogdan, M., & Sabatti, C. (2017). Controlling the rate of
559 GWAS false discoveries. *Genetics*. doi: 10.1534/genetics.116.193987
- 560 Callahan, B. J., McMurdie, P. J., Rosen, M. J., Han, A. W., Johnson, A. J. A., & Holmes, S. P. (n.d.). *DADA2: High
561 resolution sample inference from Illumina amplicon data*. doi: 10.1038/nmeth.3869
- 562 Callahan, B. J., McMurdie, P. J., Rosen, M. J., Han, A. W., Johnson, A. J. A., & Holmes, S. P. (2016). DADA2:
563 High-resolution sample inference from Illumina amplicon data. *Nature Methods*, 13(7), 581–583. doi:
564 10.1038/nmeth.3869
- 565 Carvalhais, L. C., Dennis, P. G., Fan, B., Fedoseyenko, D., Kierul, K., Becker, A., ... Borris, R. (2013). Linking
566 Plant Nutritional Status to Plant-Microbe Interactions. *PLoS ONE*, 8(7). doi: 10.1371/journal.pone.0068555
- 567 Casler, M. D., Vogel, K. P., & Harrison, M. (2015). Switchgrass Germplasm Resources. *Crop Science*, 55(6), 2463–
568 2478. doi: 10.2135/cropsci2015.02.0076
- 569 Comptant, S., Clément, C., & Sessitsch, A. (2010, May 1). Plant growth-promoting bacteria in the rhizo- and
570 endosphere of plants: Their role, colonization, mechanisms involved and prospects for utilization. *Soil Biology
571 and Biochemistry*, Vol. 42, pp. 669–678. doi: 10.1016/j.soilbio.2009.11.024
- 572 Costich, D. E., Friebel, B., Sheehan, M. J., Casler, M. D., & Buckler, E. S. (2010). Genome-size Variation in
573 Switchgrass (*Panicum virgatum*): Flow Cytometry and Cytology Reveal Rampant Aneuploidy. *The Plant
574 Genome*, 3(3), 130–141. doi: 10.3835/plantgenome2010.04.0010
- 575 Covarrubias-Pazaran, G. (2016). Genome-Assisted prediction of quantitative traits using the r package sommer.
576 *PLoS ONE*. doi: 10.1371/journal.pone.0156744
- 577 Das, M. K., Fuentes, R. G., & Taliaferro, C. M. (2004). Genetic Variability and Trait Relationships in Switchgrass.
578 *Crop Science*, 44(2), 443–448. doi: 10.2135/cropsci2004.4430
- 579 Deng, S., Caddell, D. F., Xu, G., Dahlen, L., Washington, L., Yang, J., & Coleman-Derr, D. (2021). Genome wide
580 association study reveals plant loci controlling heritability of the rhizosphere microbiome. *ISME Journal*, 1–
581 14. doi: 10.1038/s41396-021-00993-z
- 582 Dennis, P. G., Miller, A. J., & Hirsch, P. R. (2010). Are root exudates more important than other sources of
583 rhizodeposits in structuring rhizosphere bacterial communities? *FEMS Microbiology Ecology*. doi:
584 10.1111/j.1574-6941.2010.00860.x
- 585 Du, Z., Zhou, X., Ling, Y., Zhang, Z., & Su, Z. (2010). agriGO: A GO analysis toolkit for the agricultural
586 community. *Nucleic Acids Research*. doi: 10.1093/nar/gkq310
- 587 Evans, J., Sanciangco, M. D., Lau, K. H., Crisovan, E., Barry, K., Daum, C., ... Buell, C. R. (2018). Extensive
588 Genetic Diversity is Present within North American Switchgrass Germplasm. *The Plant Genome*, 11(1), 1–16.
589 doi: 10.3835/plantgenome2017.06.0055

- 590 Furlotte, N. A., Heckerman, D., & Lippert, C. (2014). Quantifying the uncertainty in heritability. *Journal of Human*
591 *Genetics*. doi: 10.1038/jhg.2014.15
- 592 Glaeser, S. P., & Kämpfer, P. (2014). The family Sphingomonadaceae. In *The Prokaryotes: Alphaproteobacteria*
593 and *Betaproteobacteria* (Vol. 9783642301971, pp. 641–707). doi: 10.1007/978-3-642-30197-1_302
- 594 Gould, A. L., Zhang, V., Lamberti, L., Jones, E. W., Obadia, B., Korasidis, N., ... Ludington, W. B. (2018).
595 Microbiome interactions shape host fitness. *Proceedings of the National Academy of Sciences of the United*
596 *States of America*, 115(51), E11951–E11960. doi: 10.1073/pnas.1809349115
- 597 Grabowski, P. P., Evans, J., Daum, C., Deshpande, S., Barry, K. W., Kennedy, M., ... Casler, M. D. (2017a).
598 Genome-wide associations with flowering time in switchgrass using exome-capture sequencing data. *New*
599 *Phytologist*, 213(1), 154–169. doi: 10.1111/nph.14101
- 600 Grabowski, P. P., Evans, J., Daum, C., Deshpande, S., Barry, K. W., Kennedy, M., ... Casler, M. D. (2017b).
601 Genome-wide associations with flowering time in switchgrass using exome-capture sequencing data. *New*
602 *Phytologist*. doi: 10.1111/nph.14101
- 603 Hacquard, S., Spaepen, S., Garrido-Oter, R., & Schulze-Lefert, P. (2017). Interplay between Innate Immunity and
604 the Plant Microbiota. *Annual Review of Phytopathology*, 55, 565–589. doi: 10.1146/annurev-phyto-080516-
605 035623
- 606 Heijboer, A., de Ruiter, P. C., Bodelier, P. L. E., & Kowalchuk, G. A. (2018). Modulation of Litter Decomposition
607 by the Soil Microbial Food Web Under Influence of Land Use Change. *Frontiers in Microbiology*, 9(NOV),
608 2860. doi: 10.3389/fmicb.2018.02860
- 609 Hestrin, R., Lee, M. R., Whitaker, B. K., & Pett-Ridge, J. (2021). The switchgrass microbiome: A review of
610 structure, function, and taxonomic distribution. *Phytobiomes Journal*. doi: 10.1094/pbiomes-04-20-0029-fi
- 611 Holland, J. B. (2007, April 1). Genetic architecture of complex traits in plants. *Current Opinion in Plant Biology*,
612 Vol. 10, pp. 156–161. doi: 10.1016/j.pbi.2007.01.003
- 613 Hu, L., Robert, C. A. M., Cadot, S., Zhang, X., Ye, M., Li, B., ... Erb, M. (2018). Root exudate metabolites drive
614 plant-soil feedbacks on growth and defense by shaping the rhizosphere microbiota. *Nature Communications*,
615 9(1), 1–13. doi: 10.1038/s41467-018-05122-7
- 616 Hunter, P. (2018). The revival of the extended phenotype. *EMBO Reports*, 19(7). doi: 10.15252/embr.201846477
- 617 Ihaka, R., & Gentleman, R. (1996). R: A Language for Data Analysis and Graphics. *Journal of Computational and*
618 *Graphical Statistics*, 5(3), 299–314. doi: 10.1080/10618600.1996.10474713
- 619 Jones, P., Garcia, B. J., Furches, A., Tuskan, G. A., & Jacobson, D. (2019, May 31). Plant host-associated
620 mechanisms for microbial selection. *Frontiers in Plant Science*, Vol. 10, p. 862. doi: 10.3389/fpls.2019.00862
- 621 Kang, H. M., Sul, J. H., Service, S. K., Zaitlen, N. A., Kong, S. Y., Freimer, N. B., ... Eskin, E. (2010). Variance
622 component model to account for sample structure in genome-wide association studies. *Nature Genetics*, 42(4),
623 348–354. doi: 10.1038/ng.548
- 624 Kaur, P., & Gaikwad, K. (2017). From genomes to gene-omes: Exome sequencing concept and applications in crop
625 improvement. *Frontiers in Plant Science*. doi: 10.3389/fpls.2017.02164
- 626 Lahti, L., Shetty, S., & Blake, T. (2017). Tools for microbiome analysis in R. *Microbiome Package Version 0.99*.
- 627 Lakshmanan, V., Kitto, S. L., Caplan, J. L., Hsueh, Y. H., Kearns, D. B., Wu, Y. S., & Bais, H. P. (2012). Microbe-
628 associated molecular patterns-triggered root responses mediate beneficial Rhizobacterial recruitment in
629 Arabidopsis. *Plant Physiology*, 160(3), 1642–1661. doi: 10.1104/pp.112.200386
- 630 Lebeis, S. L., Paredes, S. H., Lundberg, D. S., Breakfield, N., Gehring, J., McDonald, M., ... Dangl, J. L. (2015).
631 Salicylic acid modulates colonization of the root microbiome by specific bacterial taxa. *Science*, 349(6250),
632 860–864. doi: 10.1126/science.aaa8764
- 633 Lemanceau, P., Blouin, M., Muller, D., & Moënne-Loccoz, Y. (2017). Let the Core Microbiota Be Functional.
634 *Trends in Plant Science*. doi: 10.1016/j.tplants.2017.04.008
- 635 Lipka, A. E., Lu, F., Cherney, J. H., Buckler, E. S., Casler, M. D., & Costich, D. E. (2014). Accelerating the
636 Switchgrass (*Panicum virgatum L.*) Breeding Cycle Using Genomic Selection Approaches. *PLoS ONE*, 9(11),
637 e112227. doi: 10.1371/journal.pone.0112227
- 638 Lovell, J. T., MacQueen, A. H., Mamidi, S., Bonnette, J., Jenkins, J., Napier, J. D., ... Schmutz, J. (2021). Genomic
639 mechanisms of climate adaptation in polyploid bioenergy switchgrass. *Nature*, 590(7846), 438–444. doi:
640 10.1038/s41586-020-03127-1
- 641 Lowry, D. B., Lovell, J. T., Zhang, L., Bonnette, J., Fay, P. A., Mitchell, R. B., ... Juenger, T. E. (2019). QTL ×
642 environment interactions underlie adaptive divergence in switchgrass across a large latitudinal gradient.
643 *Proceedings of the National Academy of Sciences of the United States of America*. doi:
644 10.1073/pnas.1821543116
- 645 Lu, F., Lipka, A. E., Glaubitz, J., Elshire, R., Cherney, J. H., Casler, M. D., ... Costich, D. E. (2013). Switchgrass

- 646 Genomic Diversity, Ploidy, and Evolution: Novel Insights from a Network-Based SNP Discovery Protocol.
647 *PLoS Genetics*. doi: 10.1371/journal.pgen.1003215
648 Martiny, J. B. H., Jones, S. E., Lennon, J. T., & Martiny, A. C. (2015, November 6). Microbiomes in light of traits:
649 A phylogenetic perspective. *Science*, Vol. 350. doi: 10.1126/science.aac9323
650 McLaughlin, S. B., & Kszos, L. A. (2005). Development of switchgrass (*Panicum virgatum*) as a bioenergy
651 feedstock in the United States. *Biomass and Bioenergy*, 28(6), 515–535. doi: 10.1016/j.biombioe.2004.05.006
652 McMurdie, P. J., & Holmes, S. (2013). phyloseq: An R Package for Reproducible Interactive Analysis and Graphics
653 of Microbiome Census Data. *PLoS ONE*, 8(4), e61217. doi: 10.1371/journal.pone.0061217
654 Moran, N. A., & Sloan, D. B. (2015). The Hologenome Concept: Helpful or Hollow? *PLoS Biology*, 13(12). doi:
655 10.1371/journal.pbio.1002311
656 Oksanen, J., Blanchet, F. G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., ... Maintainer, H. W. (2019).
657 vegan: Community Ecology Package. R package version 2.5-5. <https://CRAN.R-project.org/package=vegan>.
658 *Community Ecology Package*.
659 Palazzo, A. F., & Gregory, T. R. (2014). The Case for Junk DNA. *PLoS Genetics*. doi:
660 10.1371/journal.pgen.1004351
661 Paliy, O., & Shankar, V. (2016). Application of multivariate statistical techniques in microbial ecology. *Molecular
662 Ecology*. doi: 10.1111/mec.13536
663 Parada, A. E., Needham, D. M., & Fuhrman, J. A. (2016). Every base matters: Assessing small subunit rRNA
664 primers for marine microbiomes with mock communities, time series and global field samples. *Environmental
665 Microbiology*. doi: 10.1111/1462-2920.13023
666 Parker, M. A., Malek, W., & Parker, I. M. (2006). Growth of an invasive legume is symbiont limited in newly
667 occupied habitats. *Diversity & Distributions*, 12(5), 563–571.
668 doi: 10.1111/j.1366-9516.2006.00255.x
669 Parrish, D. J., & Fike, J. H. (2005, September). The biology and agronomy of switchgrass for biofuels. *Critical
670 Reviews in Plant Sciences*, Vol. 24, pp. 423–459. doi: 10.1080/07352680500316433
671 Peres-Neto, P. R., Legendre, P., Dray, S., & Borcard, D. (2006). VARIATION PARTITIONING OF SPECIES
672 DATA MATRICES: ESTIMATION AND COMPARISON OF FRACTIONS. *Ecology*, 87(10), 2614–2625.
673 doi: 10.1890/0012-9658
674 Quast, C., Pruesse, E., Yilmaz, P., Gerken, J., Schweer, T., Yarza, P., ... Glöckner, F. O. (n.d.). *The SILVA
675 ribosomal RNA gene database project: improved data processing and web-based tools*. doi:
676 10.1093/nar/gks1219
677 Risely, A. (2020). Applying the core microbiome to understand host–microbe systems. *Journal of Animal Ecology*.
678 doi: 10.1111/1365-2656.13229
679 Rossum, B. van, Kruijer, W., Eeuwijk, F. van, & Boer, M. (2020). Package “statgenGWAS” Title Genome Wide
680 Association Studies. doi: 10.1038/ng.548
681 Sanderson, M. A., Adler, P. R., Boateng, A. A., Casler, M. D., & Sarath, G. (2006). Switchgrass as a biofuels
682 feedstock in the USA. *Canadian Journal of Plant Science*, 86(5), 1315–1325. doi: 10.4141/p06-136
683 Sasse, J., Martinoia, E., & Northen, T. (2018, January 1). Feed Your Friends: Do Plant Exudates Shape the Root
684 Microbiome? *Trends in Plant Science*, Vol. 23, pp. 25–41. doi: 10.1016/j.tplants.2017.09.003
685 Schwartz, C., & Amasino, R. (2013). Nitrogen Recycling and Flowering Time in Perennial Bioenergy Crops.
686 *Frontiers in Plant Science*, 0(APR), 76. doi: 10.3389/FPLS.2013.00076
687 Sharma, M. K., Sharma, R., Cao, P., Jenkins, J., Bartley, L. E., Qualls, M., ... Ronald, P. C. (2012). A Genome-
688 Wide Survey of Switchgrass Genome Structure and Organization. *PLoS ONE*, 7(4), e33892. doi:
689 10.1371/journal.pone.0033892
690 Singer, E., Bonnette, J., Kenaley, S. C., Woyke, T., & Juenger, T. E. (2019). Plant compartment and genetic
691 variation drive microbiome composition in switchgrass roots. *Environmental Microbiology Reports*, 11(2),
692 185. doi: 10.1111/1758-2229.12727
693 Singer, E., Bonnette, J., Woyke, T., & Juenger, T. E. (2019). Conservation of Endophyte Bacterial Community
694 Structure Across Two *Panicum* Grass Species. *Frontiers in Microbiology*, 10(SEP), 2181. doi:
695 10.3389/FMICB.2019.02181
696 Smith, C. C. R., Snowberg, L. K., Gregory Caporaso, J., Knight, R., & Bolnick, D. I. (2015). Dietary input of
697 microbes and host genetic variation shape among-population differences in stickleback gut microbiota. *ISME
698 Journal*, 9(11), 2515–2526. doi: 10.1038/ismej.2015.64
699 Stewart, C. E., Roosendaal, D., Denef, K., Pruessner, E., Comas, L. H., Sarath, G., ... Soundararajan, M. (2017).
700 Seasonal switchgrass ecotype contributions to soil organic carbon, deep soil microbial community
701 composition and rhizodeposit uptake during an extreme drought. *Soil Biology and Biochemistry*, 112, 191–

- 702 203. doi: 10.1016/j.soilbio.2017.04.021
703 Tian, T., Liu, Y., Yan, H., You, Q., Yi, X., Du, Z., ... Su, Z. (2017). AgriGO v2.0: A GO analysis toolkit for the
704 agricultural community, 2017 update. *Nucleic Acids Research*. doi: 10.1093/nar/gkx382
705 Trexler, R. V., & Bell, T. H. (2019). Testing sustained soil-to-soil contact as an approach for limiting the abiotic
706 influence of source soils during experimental microbiome transfer. *FEMS Microbiology Letters*. doi:
707 10.1093/femsle/fnz228
708 Trivedi, P., Leach, J. E., Tringe, S. G., Sa, T., & Singh, B. K. (2020, November 1). Plant–microbiome interactions:
709 from community assembly to plant health. *Nature Reviews Microbiology*, Vol. 18, pp. 607–621. doi:
710 10.1038/s41579-020-0412-1
711 Trujillo, M. E., Hong, K., & Genilloud, O. (2014). The family Micromonosporaceae. In *The Prokaryotes:*
712 *Actinobacteria* (pp. 499–569). doi: 10.1007/978-3-642-30138-4_196
713 Van Rossum, G., & Drake, F. L. (2009). *Python 3 Reference Manual*. Scotts Valley, CA: CreateSpace.
714 VanRaden, P. M. (2008). Efficient methods to compute genomic predictions. *Journal of Dairy Science*, 91(11),
715 4414–4423. doi: 10.3168/jds.2007-0980
716 Wagner, M. R. (2021). Prioritizing host phenotype to understand microbiome heritability in plants. *New Phytologist*.
717 doi: 10.1111/NPH.17622
718 Wagner, M. R., Lundberg, D. S., Del Rio, T. G., Tringe, S. G., Dangl, J. L., & Mitchell-Olds, T. (2016). Host
719 genotype and age shape the leaf and root microbiomes of a wild perennial plant. *Nature Communications*, 7.
720 doi: 10.1038/ncomms12151
721 Weiss, S., Xu, Z. Z., Peddada, S., Amir, A., Bittinger, K., Gonzalez, A., ... Knight, R. (2017). Normalization and
722 microbial differential abundance strategies depend upon data characteristics. *Microbiome*, 5(1), 27. doi:
723 10.1186/s40168-017-0237-y
724 Werling, B. P., Dickson, T. L., Isaacs, R., Gaines, H., Gratton, C., Gross, K. L., ... Landis, D. A. (2014). Perennial
725 grasslands enhance biodiversity and multiple ecosystem services in bioenergy landscapes. *Proceedings of the*
726 *National Academy of Sciences of the United States of America*, 111(4), 1652–1657. doi:
727 10.1073/pnas.1309492111
728 Whitham, T. G., Young, W. P., Martinsen, G. D., Gehring, C. A., Schweitzer, J. A., Shuster, S. M., ... Kuske, C. R.
729 (2003, March 1). Community and ecosystem genetics: A consequence of the extended phenotype. *Ecology*,
730 Vol. 84, pp. 559–573. doi: 10.1890/0012-9658(2003)084[0559:CAEGAC]2.0.CO;2
731 Wilson, A. S., Koller, K. R., Ramaboli, M. C., Nesengani, L. T., Ocvirk, S., Chen, C., ... O’Keefe, S. J. D. (2020,
732 March 1). Diet and the Human Gut Microbiome: An International Review. *Digestive Diseases and Sciences*,
733 Vol. 65, pp. 723–740. doi: 10.1007/s10620-020-06112-w
734 Wray, N. R., Yang, J., Hayes, B. J., Price, A. L., Goddard, M. E., & Visscher, P. M. (2013, July). Pitfalls of
735 predicting complex traits from SNPs. *Nature Reviews Genetics*, Vol. 14, pp. 507–515. doi: 10.1038/nrg3457
736 Wright, E. S. (n.d.). *DECIPHER v2.0 to Analyze Big Biological Sequence Data in R*.
737 Yin, C., Casa Vargas, J. M., Schlatter, D. C., Hagerty, C. H., Hulbert, S. H., & Paulitz, T. C. (2021). Rhizosphere
738 community selection reveals bacteria associated with reduced root disease. *Microbiome 2021* 9:1, 9(1), 1–18.
739 doi: 10.1186/S40168-020-00997-5
740 Yu, P., & Hochholdinger, F. (2018). The role of host genetic signatures on root–microbe interactions in the
741 rhizosphere and endosphere. *Frontiers in Plant Science*, Vol. 871. doi: 10.3389/fpls.2018.01896
742 Zeng, Q., Sukumaran, J., Wu, S., & Rodrigo, A. (2015). Neutral Models of Microbiome Evolution. *PLOS*
743 *Computational Biology*, 11(7), e1004365. doi: 10.1371/journal.pcbi.1004365
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745 **Data Availability**

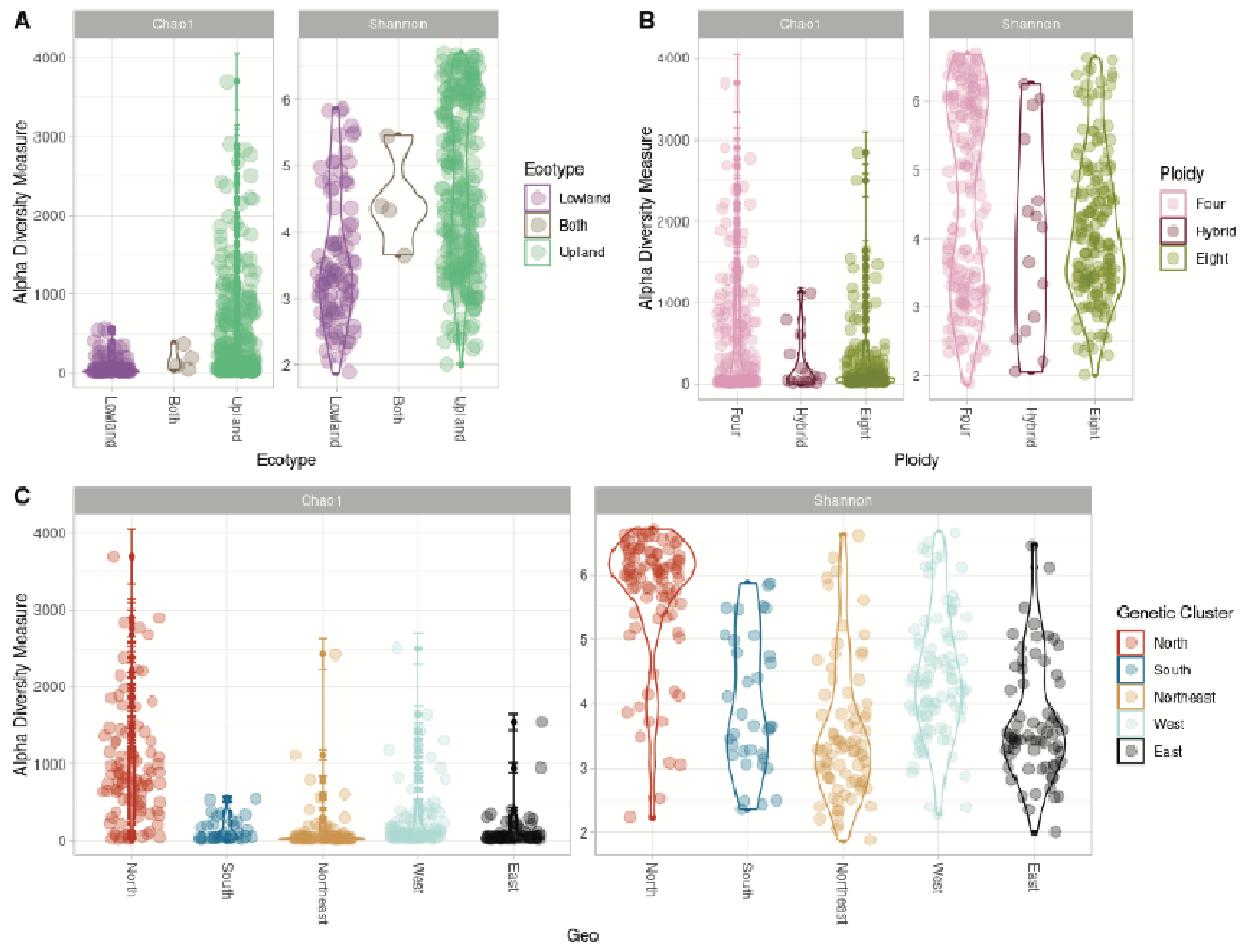
746 Raw amplicon sequence reads are deposited in the SRA (BioProject PRJNA689762).

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748 **Figure and Table Captions**

749 Figures:

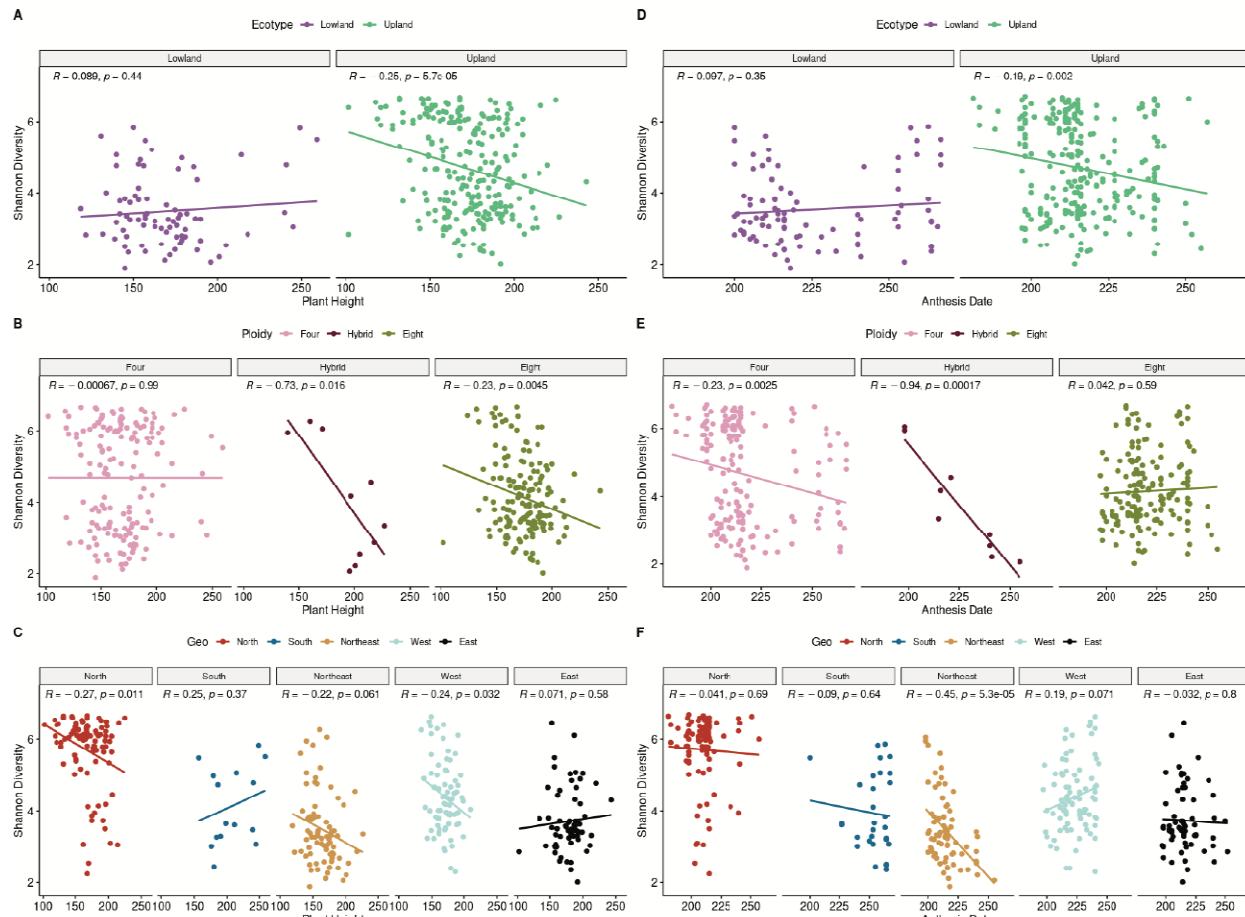
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Figure 1:

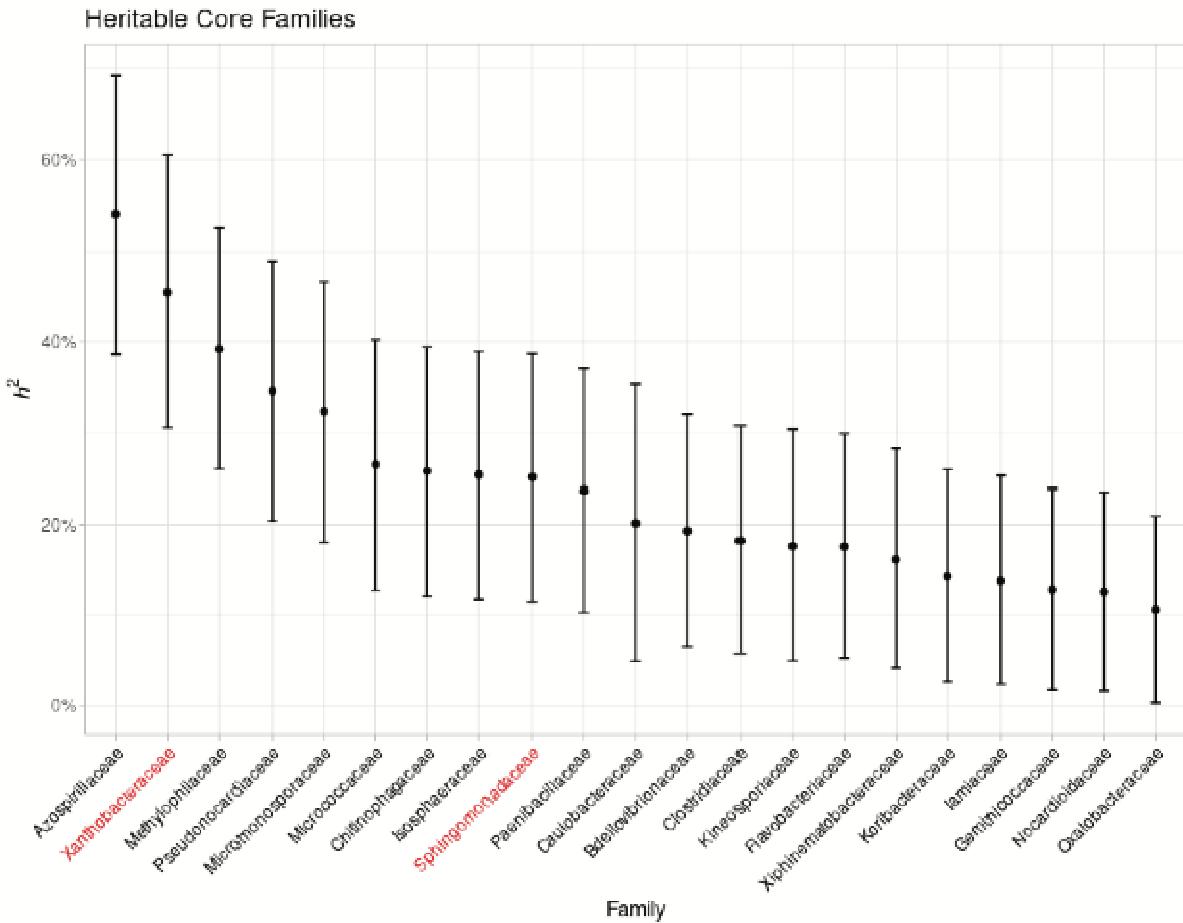
A: Alpha diversity by Ecotype. Alpha-diversity, measured by Chao1 and Shannon diversity Index, plotted for switchgrass ecotypes with Lowland (purple), Both (brown), and Upland (green). **B: Alpha diversity by Ploidy.** Alpha-diversity, measured by Chao1 and Shannon diversity Index, plotted for switchgrass ploidy levels with Tetraploid - Four (pink), Hybrid (maroon), and Octoploid - Eight (olive). **C: Alpha diversity by Genetic cluster.** Alpha-diversity, measured by Chao1 and Shannon diversity Index, plotted for switchgrass genetic clusters with North (red), South (dark blue), Northeast (beige), West (light blue), and East (black).



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Figure 2:

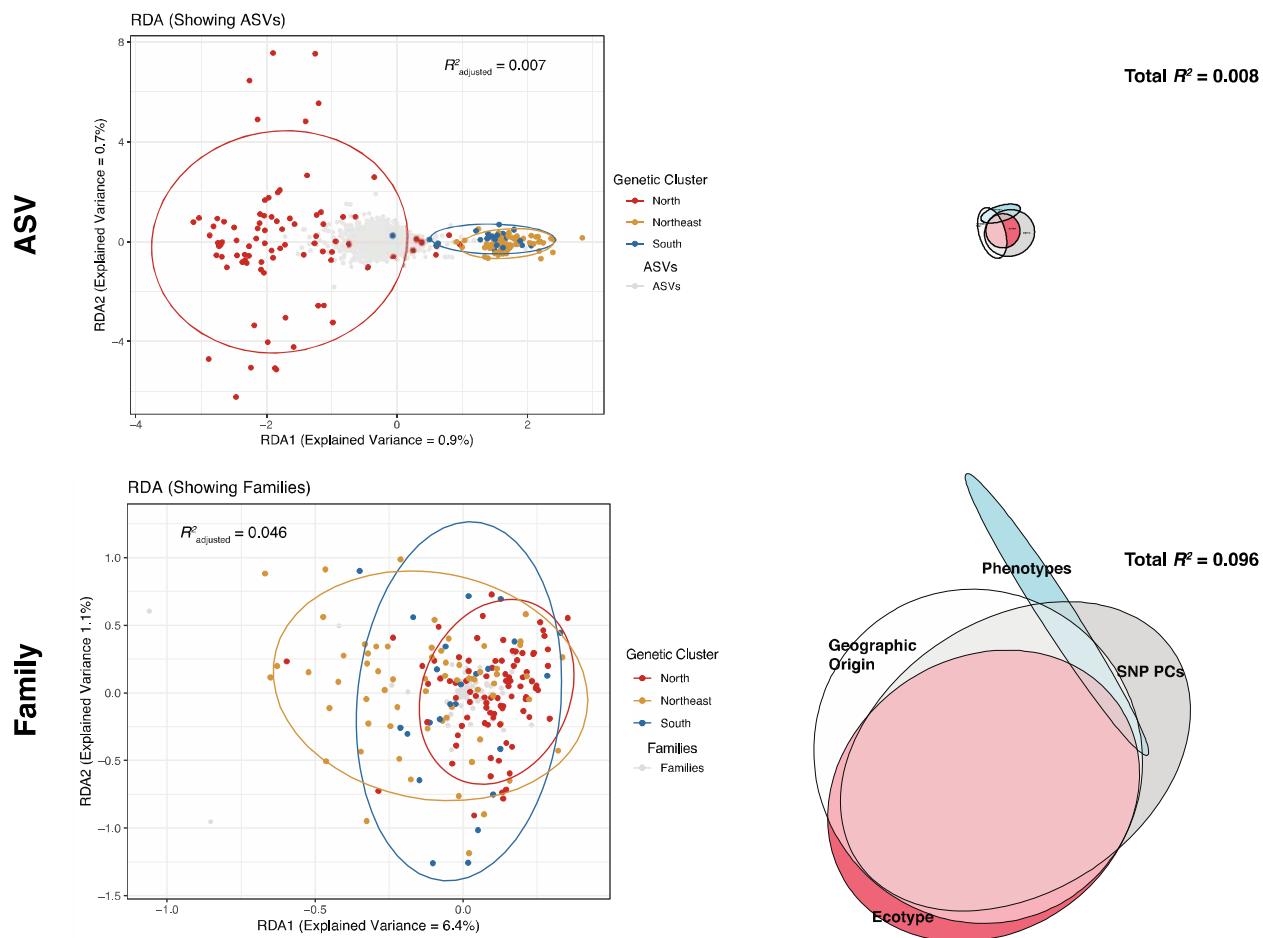
Relationships between Shannon diversity and plant height. A: Ecotype: plotted for switchgrass ecotypes with Lowland (purple), Both (brown), and Upland (green). B: Ploidy: plotted for switchgrass ploidy levels with Tetraploid - Four (pink), Hybrid (maroon), and Octoploid - Eight (olive). C: Genetic cluster: plotted for switchgrass genetic clusters with North (red), South (dark blue), Northeast (beige), West (light blue), and East (black). **Relationships between Shannon diversity and anthesis date.** D: Ecotype: plotted for switchgrass ecotypes with Lowland (purple), Both (brown), and Upland (green). E: Ploidy: plotted for switchgrass ploidy levels with Tetraploid - Four (pink), Hybrid (maroon), and Octoploid - Eight (olive). F: Genetic cluster: plotted for switchgrass genetic clusters with North (red), South (dark blue), Northeast (beige), West (light blue), and East (black).



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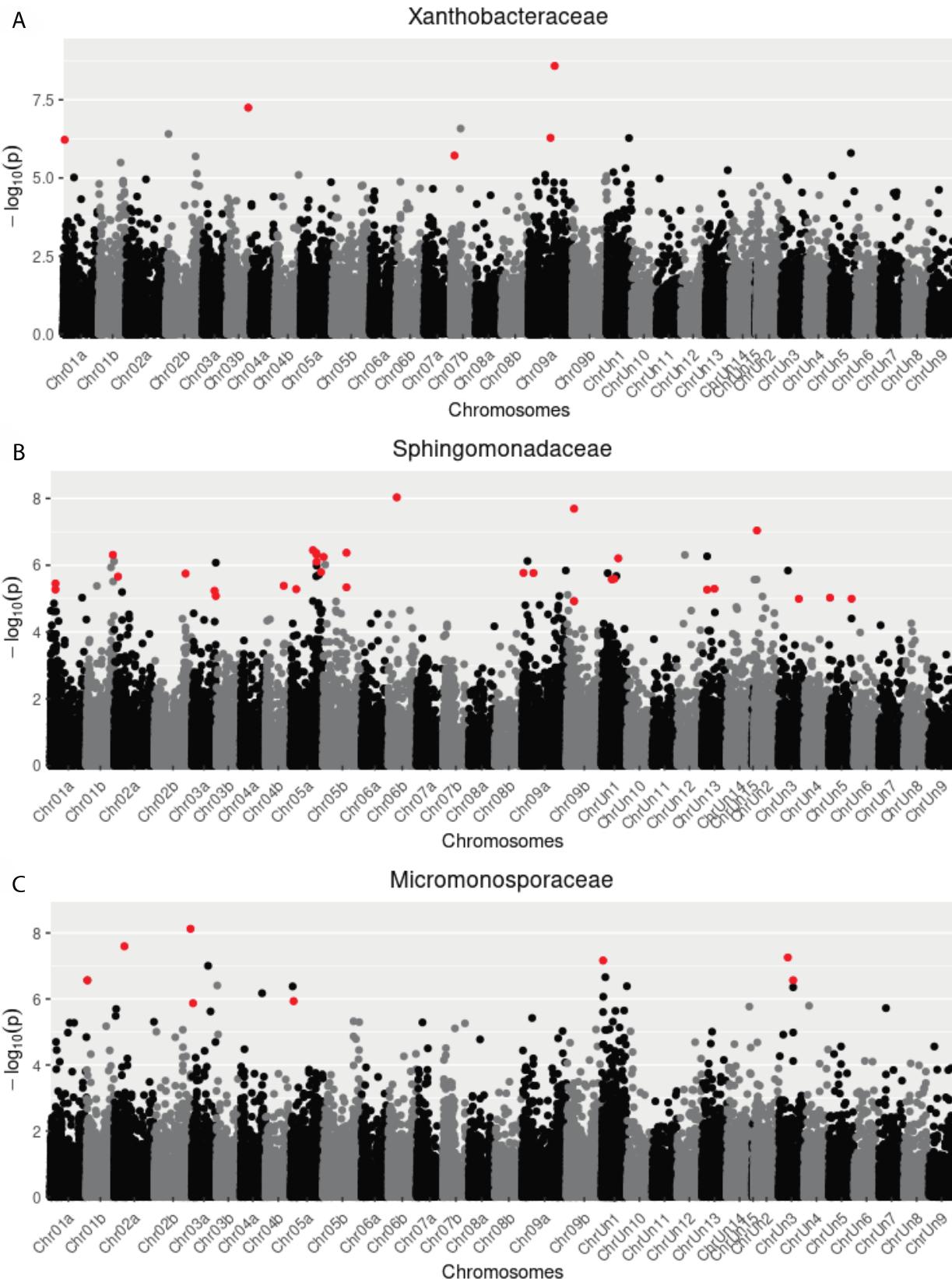
Figure 3:

Heritability estimates for bacterial families in the switchgrass rhizosphere. For 110 bacterial families that were observed in more than 10% of switchgrass samples, narrow-sense heritability (h^2) was calculated using a mixed linear model. 21 families had heritability estimates with standard error greater than zero. Families observed in more than 90% of our samples are represented in red.



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Figure 4:
Redundancy Analyses and Variance Partitioning. **A:** Results of RDA analysis presenting amplicon sequence variants (Grey) and the North (Red), Northeast (Beige), and South (Blue) genetic clusters. **B:** Results of variance partitioning analysis for ASVs presenting Ecotype (Red), SNP PCs (Grey), Phenotypes (Blue), and Geographic Origin (White). **C:** Results of RDA analysis presenting rhizosphere bacterial families (Grey) and the North (Red), Northeast (Beige), and South (Blue) genetic clusters. **D:** Results of variance partitioning analysis for rhizosphere bacterial families presenting Ecotype (Red), SNP PCs (Grey), Phenotypes (Blue), and Geographic Origin (White).

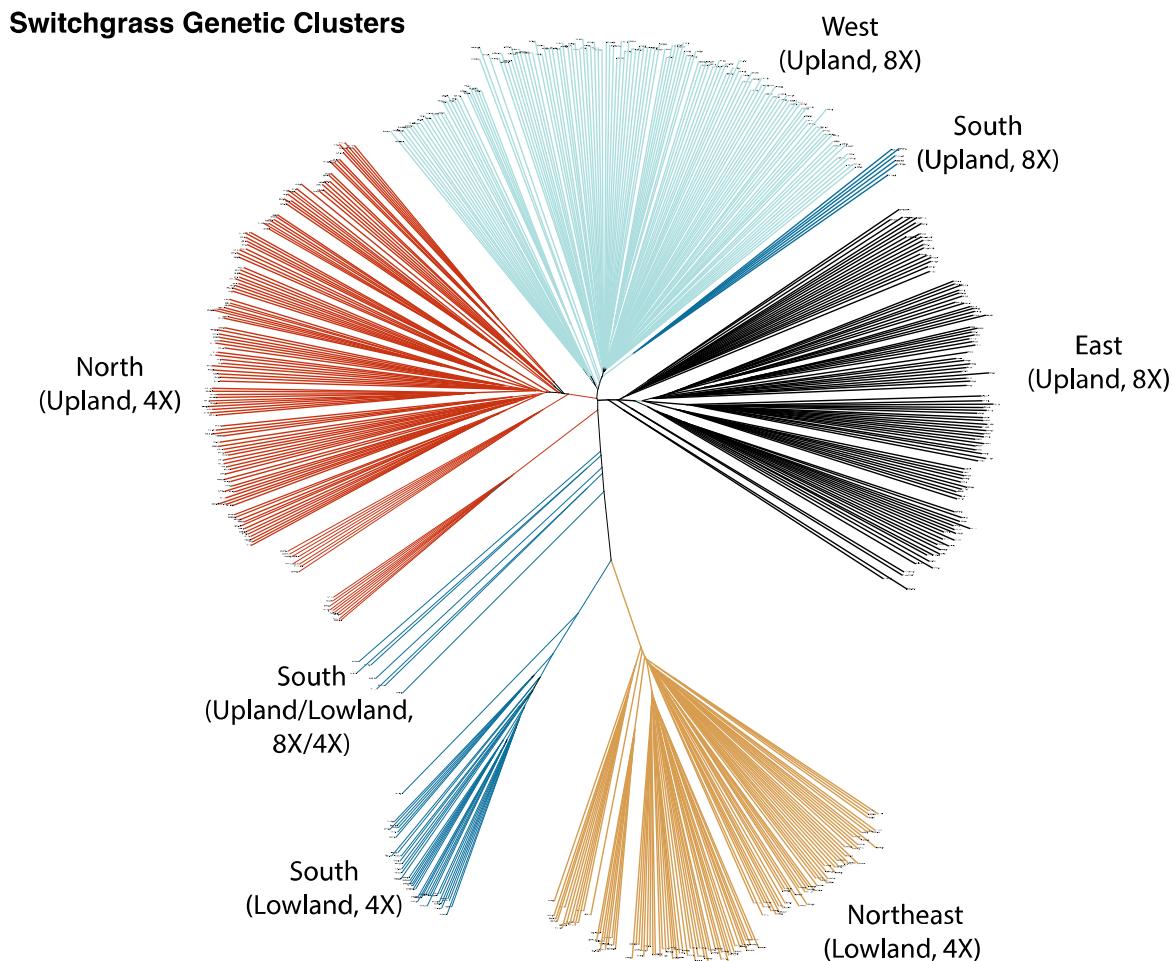


790 **Genome-wide association study (GWAS): A:** Manhattan plot presenting significant SNPs (Red) associated with
791 Xanthobacteraceae rhizosphere abundance. **B:** Manhattan plot presenting significant SNPs (Red) associated with
792 Sphingomonadaceae rhizosphere abundance. **C:** Manhattan plot presenting significant SNPs (Red) associated with
793 Micromonosporaceae rhizosphere abundance.

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795 Supplemental Figures:

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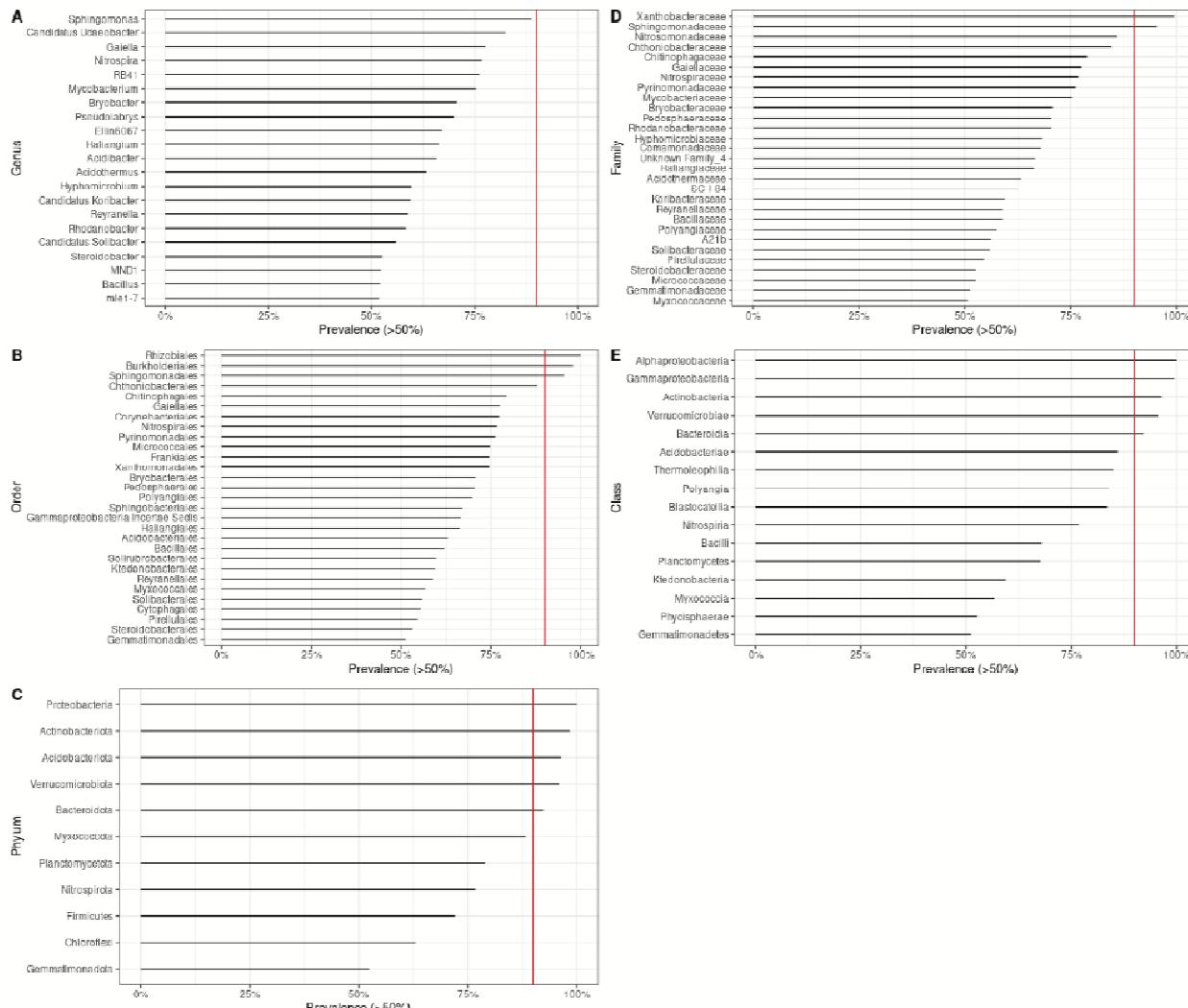


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798 **Supplemental Figure 1:**

799 **Phylogenetic analysis:** Results of phylogenetic analysis based on exome capture dataset from Evans et al. 2018 with
800 switchgrass genetic clusters colored as North (red), South (dark blue), Northeast (beige), West (light blue), and East
801 (black).

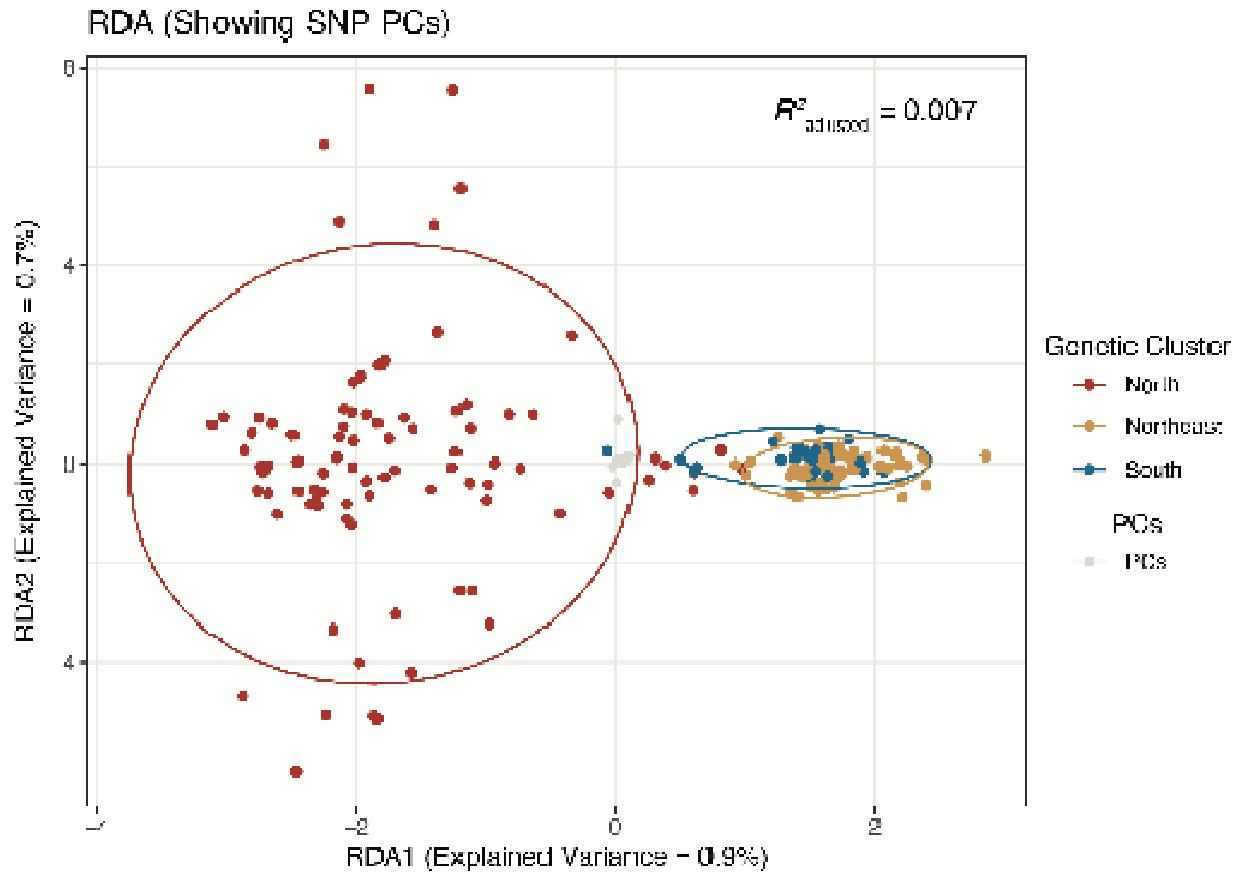
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Supplemental Figure 2:

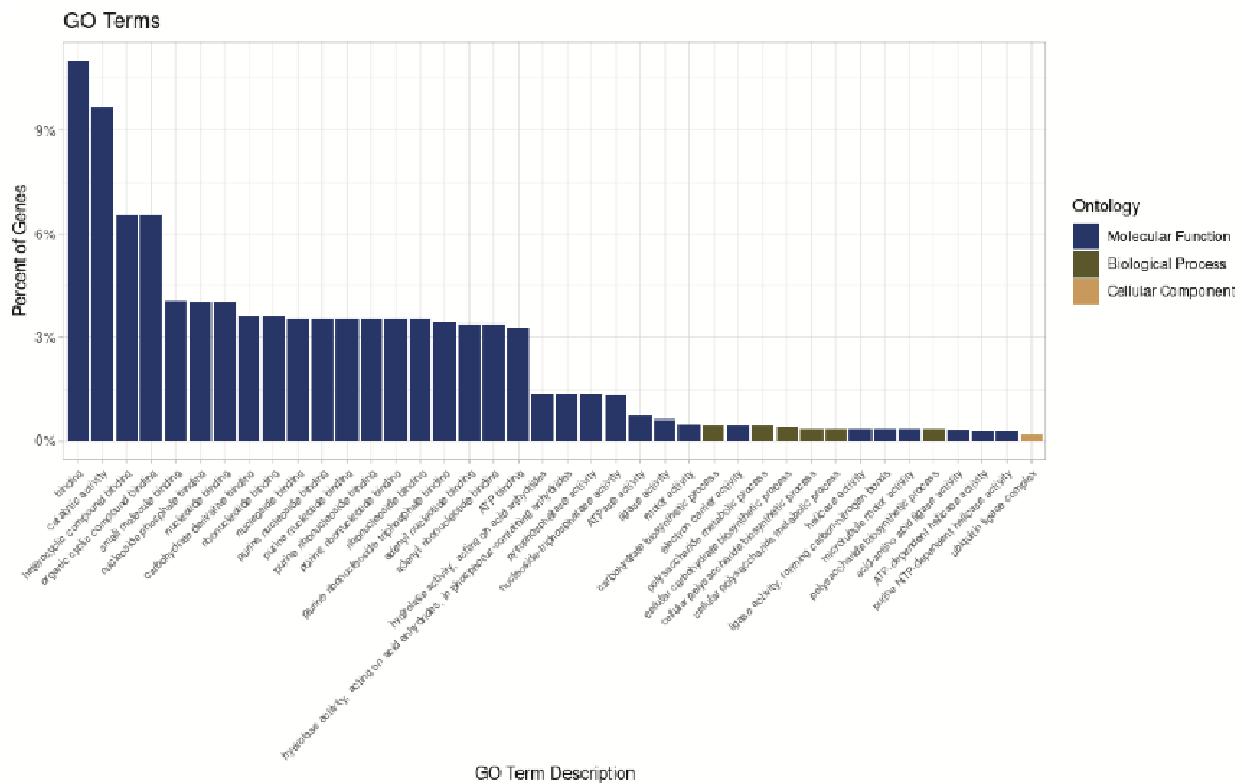
Prevalence of rhizosphere bacterial taxa: The prevalence of rhizosphere bacterial taxa in greater than 50% of switchgrass samples shown for **A: Genus**, **B: Family**, **C: Order**, **D: Class**, and **E: Phylum**. The 90% occupancy threshold for each group is indicated by a vertical red line.



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Supplemental Figure 3:

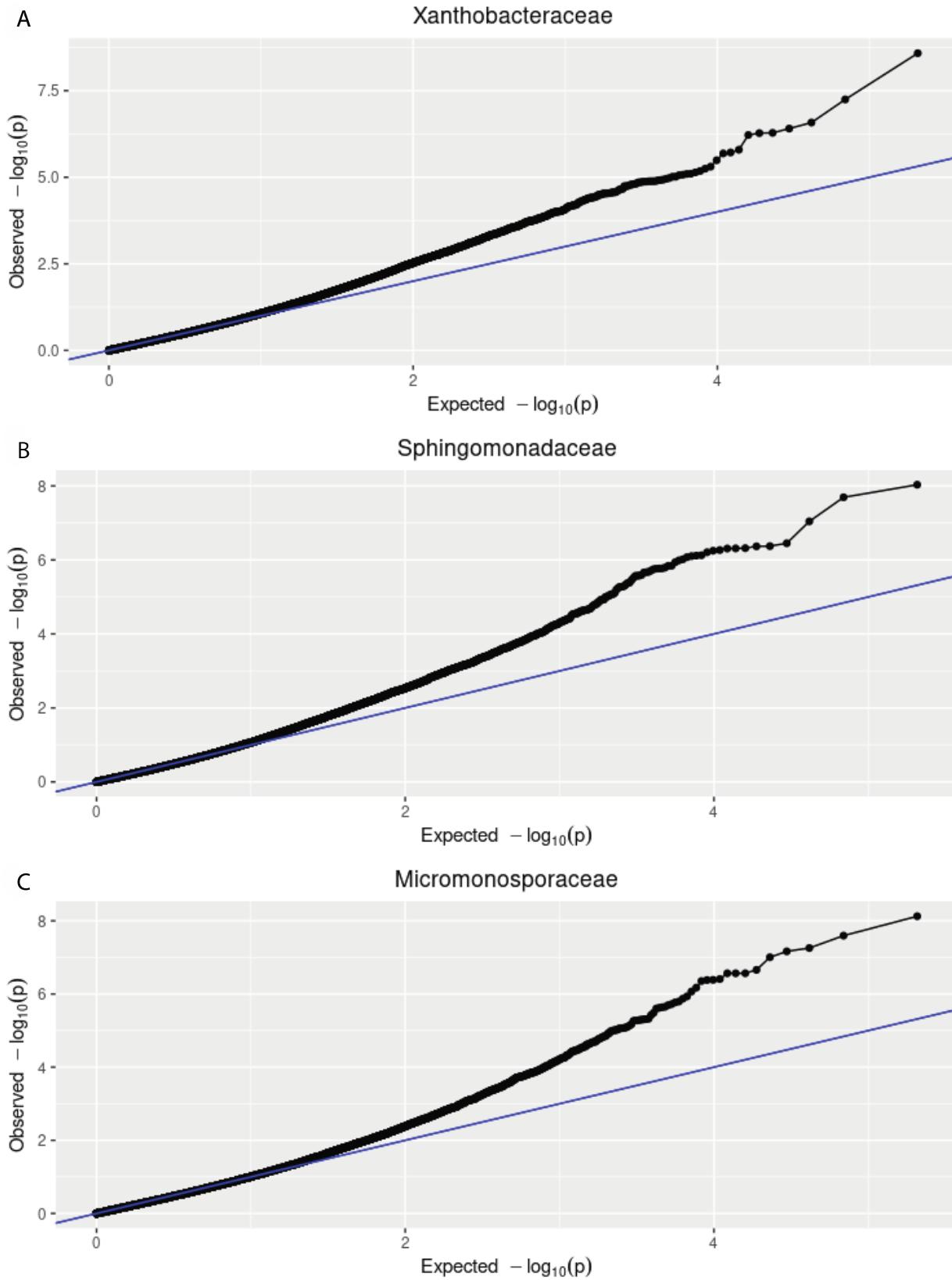
Redundancy Analyses (PCs): Results of RDA analysis presenting SNP PCs (grey) and the North (red), Northeast (beige), and South (dark blue) genetic clusters.



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Supplemental Figure 4:

GO Terms: Percent of enriched GO terms related to Molecular Function (dark blue), Biological Process (olive), and Cellular Component (beige).



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Supplemental Figure 5:

821 ***Genome-wide association study (GWAS): A:*** QQ plot presenting significant SNPs (red) associated with
 822 Xanthobacteraceae rhizosphere abundance. ***B:*** QQ plot presenting significant SNPs (red) associated with
 823 Sphingomonadaceae rhizosphere abundance. ***C:*** QQ plot presenting significant SNPs (red) associated with
 824 Micromonosporaceae rhizosphere abundance.

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 826 Supplemental Tables:
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Accession	Observ ed	Chao1	se.chao1	ACE	se.ACE	Shannon	Simpson	InvSimpson	Fisher
Blackwell.01 .2	284	369.56	23.394779	354.272	8.002603	5.3424889	2	0.993834	132.26882
			1.4225048	918	93			162.179695	2
			157.090	7	641	56	0.983254	21.150173	
Blackwell.01	82	82.75		145.800	5.671623	4.215349	0.986778	59.715753	3
			13.659610	4	369	98	7	75.6315232	1
Blackwell.02	132	909		125.653	4.950851	4.5327523			40.732534
			128.111	9	226	1	3		36.441366
Blackwell.03	122	111		194.578	5.869440	4.8451448	0.986856	76.0803408	1
			9.7584752	9	467	07	3	102.438025	61.407364
Blackwell.04	175	923			0.960768	2.3923152			2.1091676
			232.689	0	13	92	4	0.899676	9.96770464
Blackwell.05	13	13		226.089	5.819401	5.0021228			76.920499
			12.043015	2	895	91	9	0.991456	117.041199
Blackwell.06	203	655			2.229670	3.3590414			7.0549535
Carthage.01 .2	35	35		34.6522	2.343733	3.1504598			6.5554321
			33.3333	0	35	06	2	0.960402	25.2538007
Carthage.01	33	333		0.9230919	33.4661	1.792922	0.94669	18.7582067	3
			0.4923659	6	222	59	5		6.5554321
Carthage.02	33	33		0.4946522	103	95	8	0.95452	21.9876869
			0.4946522	6	516	26	5		10.234894
Carthage.03	47	47		0.4898979		1.147710	0.969314	32.588151	7
			0.4898979	5		2.9982538			4.6508688
Carthage.04	25	25		26.12		0.966091	0.941572	17.1150818	2
				5	2	2.4329510			2.5030219
Carthage.05	15	15		0	15	78	3	0.900104	10.0104108
			0.2472676		46.3328	2.872233			9.9583175
Carthage.06	46	46		8	042	76	1	0.967658	30.9195473
CaveinRock. 01	241.217	22.700631		225.941	7.275428	4.7746719			63.520861
	179	391		8	994	77	3	0.988734	88.7626487
CaveinRock. 03	34	34		0	34	72	5	0.957934	23.7721676
CaveinRock. 04	11	11		0	NA	NA	4	0.816972	5.46364491
			1.4216995		52.5269	3.487480			1.7290385
CaveinRock. 06	51	51.75		5	868	35	7	0.964654	28.2917445
CaveinRock. 07	19	19		0	19	83	6	0.906036	11.361576
			2.026144		9	9	9	0.872682	3.3283031
CaveinRock. 08	19	19		0	19	83	6	0.872682	3.3283031
			756.309		725.718	14.55862			342.72404
Dacotah.02	468	278		9	703	1	5.9079517	0.996468	4
			50.450369		1308.02	17.00212		283.125708	
Dacotah.03	676	424		8	044	02	8	0.998092	916.05373
			77.663680					524.109015	4

Dacotah.04	423	624.741	38.926336	620.717	13.96987	5.7900730			276.56328
		573	4	968	77	9	0.99604	252.525253	7
		674.666	37.119183	708.293	15.52590	5.9068336			337.92165
Dacotah.05	465	667	8	812	06	3	0.996498	285.551114	5
		1104.38	65.879614	1139.99	17.53679	6.2936985			713.14069
Dacotah.07	625	71	1	626	3	4	0.997828	460.405157	8
		221.95544	2679.47	13.78880	6.6552991				2260.3525
Dacotah.08	828	2680.32	6	419	54	8	0.9986	714.285714	9
				2.304418	3.0658291				5.5839911
ECS.1.05	29	29	0	29	54	5	0.942692	17.4495707	2
		2.2998855	20.5890	1.683910					3.1175139
ECS.1.06	18	19	8	909	07	2.5683886	0.914612	11.7112475	4
				2.788866	3.6244770				9.6838002
ECS.12.01	45	45	0	45	76	8	0.969748	33.0556657	4
		183.769	17.238614	165.971	5.763388	4.6236123			
ECS.12.02	148	231	5	715	15	2	0.987436	79.5924865	47.997644
				92.3956	4.726708				
ECS.12.03	92	92.2	0.6211081	644	63	4.2780631	0.983272	59.7800096	24.694105
		135.312	1.6025110	136.017	4.915418	4.6706732			41.613637
ECS.12.05	134	5	7	432	19	1	0.988754	88.9205051	9
		0.6211025	91.4664	4.756108	4.3463872				24.331359
ECS.12.07	91	91.2	9	354	06	3	0.985614	69.5120256	3
		126.583	5.4482389	125.221	5.179940	4.4153944			35.190936
ECS.2.02	119	333	4	544	28	3	0.984792	65.7548659	6
		290.454		272.205	6.573931	5.1440032			
ECS.2.03	235	546	18.730274	486	01	3	0.992416	131.85654	96.81155
		0.7360481	81.5651	4.487695	4.1634070				20.806009
ECS.2.04	81	81.25	2	007	33	4	0.981912	55.285272	8
		0.3817603	76.5797	4.342524					19.113207
ECS.2.06	76	76.1	9	785	45	4.0168183	0.977958	45.3679339	7
		2.3326329	67.2962	3.995960	4.0140341				16.184541
ECS.2.07	67	68	5	963	89	7	0.979062	47.7600535	4
		0.2472066	45.3072	2.933789	3.5627293				9.6838002
ECS.10.01	45	45	2	902	86	5	0.967504	30.7730182	4
				1.624807	2.9845207				4.6508688
ECS.10.03	25	25	0	25	68	5	0.940842	16.9038845	2
		4.5993925	32.8037	2.514591	3.0079953				
ECS.10.07	31	34	4	5	17	2	0.937756	16.0658055	6.0650427
		0.4909902	28.2733	2.356810	3.0237944				5.3470387
ECS.10.08	28	28	5	113	57	5	0.943754	17.7790421	8
				3.089571	3.5389183				9.4113548
ECS.10.09	44	44	0	44	9	4	0.966258	29.6366546	7
		2.3232382	39.4975	2.415816	3.3910601				7.8209646
ECS.10.10	38	39	4	37	29	4	0.961654	26.0783393	9
		117.833	1.2604964	118.395	4.950770	4.5256214			34.366727
ECS.11.01	117	333	7	017	42	5	0.987064	77.3036487	8
		0.0988571	44.4333	2.851068	3.5181090				9.4113548
ECS.11.02	44	44	1	048	81	1	0.96517	28.7108814	7
				2.053959	3.2729350				6.3090836
ECS.11.04	32	32	0	32	59	9	0.957414	23.4818955	3
				1.337712	2.6862938				3.3283031
ECS.11.05	19	19	0	19	11	4	0.92127	12.7016385	1

ECS.11.07	157	218.875	25.182649	196.417	6.860286	4.5762160			52.304464
			8	512	26	5	0.986162	72.2647782	6
					2.2086196				
HighTide.02	11	11	0	NA	NA	1	0.86129	7.20928556	1.7290385
					1.345185	2.8546305			3.7583769
HighTide.03	21	21	0	21	42	9	0.9339	15.128593	1
					1.788854	2.6534529			3.5419457
HighTide.05	20	20	0	20	38	5	0.913368	11.5430788	8
					0.953462	2.0604604			
HighTide.09	11	11	0	11	59	1	0.848558	6.60318802	1.7290385
					0.968245	2.5352006			2.7047898
HighTide.10	16	16	0	16	84	8	0.909588	11.0604787	6
			5.7942620	136.481	5.060855	4.6272811			40.732534
Kanlow.01	132	139.8	5	022	93	3	0.988286	85.3679358	1
				59.2239	3.562309	3.8346694			13.710755
Kanlow.02	59	59	0.1652482	335	36	8	0.974376	39.0259132	8
			2.3277386	48.5768	3.106327	3.6482240			10.513519
Kanlow.03	48	49	8	655	2	8	0.970032	33.3689269	9
					2.034190	3.0615609			5.5839911
Kanlow.04	29	29	0	29	51	8	0.943474	17.6909741	2
			197.676	8.8938170	202.486	6.234049	4.7330135		62.459927
Kanlow.05	177	471	5	714	7	2	0.987688	81.2215725	9
					2.304418	3.0572103			5.5839911
Kanlow.06	29	29	0	29	54	4	0.944458	18.0043931	2
					3.016620	3.2779203			8.3425609
Kanlow.08	40	40	0	40	63	6	0.950638	20.2584984	6
					1.951800	2.4982881			3.7583769
Kanlow.10	21	21	0	21	15	9	0.87997	8.33125052	1
					2.236067	3.3007340			7.3080809
KY1625.01	36	36	0	36	98	1	0.955154	22.2985328	3
					1.609630	2.8223172			
KY1625.02	22	22	0	22	11	1	0.930418	14.3715329	3.9775371
			0.4885042	22.5871	1.832393				
KY1625.03	22	22	1	936	2	2.8498394	0.93646	15.7381177	3.9775371
			2.3145502	28.1911	2.095781	3.0126158			5.1125109
KY1625.04	27	28	5	385	78	9	0.944648	18.0661945	2
			0.4952605	53.1403	3.309600	3.7910374			11.936998
KY1625.06	53	53	6	509	16	2	0.974256	38.8440025	1
			0.2465033	36.3734	2.316959				7.3080809
KY1625.07	36	36	2	074	53	3.323476	0.957368	23.4565585	3
Pathfinder.0			0.4894725	25.1111	1.066883				4.4238312
1	24	24	1	111	2	2.8738709	0.931104	14.5146308	1
Pathfinder.0			0.4918693	31.3805	1.810100	3.2172318			
2	31	31	8	41	82	1	0.952968	21.2621194	6.0650427
Pathfinder.0			0.4960158	63.1347	3.615392	4.0034066			14.932265
3	63	63	7	518	81	9	0.979342	48.4073967	5
Pathfinder.0			0.1653594	64.1680	3.910412	3.9565316			15.242462
4	64	64	6	336	95	2	0.97814	45.7456542	8
Pathfinder.0			497.149	32.223917	481.942	10.68062	5.5926925		195.49644
5	354	254	2	64	65	1	0.995268	211.327134	5
Pathfinder.0			0.0992619	68.3272	4.026175	3.9687013			16.502381
6	68	68	8	39	65	1	0.977834	45.1141388	4

		266.675		262.922	6.869075				88.440004
Shelter.01	222	676	15.401994	522	03	5.0768225	0.992116	126.839168	9
		148.157	4.7464328	147.604	5.125929	4.6565174			44.758093
Shelter.04	141	895	2	547	49	1	0.988222	84.9040584	5
		107.111	4.9755659	105.959	5.042556	4.3239777			28.042367
Shelter.05	101	111	3	064	18	8	0.984116	62.9564342	7
				2.620767	3.4252775				7.8209646
Shelter.06	38	38	0	38	26	8	0.962992	27.0211846	9
				41.3370	2.594245	3.4853419			8.6065827
Shelter.07	41	41	0.1646216	185	3	4	0.964894	28.4851592	4
				2.247805	3.4858016				7.8209646
Shelter.08	38	38	0	38	95	3	0.965726	29.1766354	9
				0.980580	3.1696484				4.8804419
Sunburst.01	26	26	0	26	68	6	0.955562	22.503263	6
				1.615145	2.9328576				4.1993718
Sunburst.02	23	23	0	23	71	3	0.94052	16.8123739	9
				0.4890096	24.0864	1.483973	3.0044659		4.1993718
Sunburst.04	23	23	5	198	93	3	0.946512	18.6957822	9
		61.3333	0.9245561	61.5347	3.627977	3.9264577			14.317643
Sunburst.05	61	333	5	21	36	1	0.977988	45.4297656	8
		231.823		231.577	6.244902	4.9978561			77.506253
Sunburst.06	204	529	11.04251	917	37	5	0.99154	118.20331	5
			0.4923659	33.3279	2.307691	3.2662388			6.5554321
Sunburst.07	33	33	6	766	93	2	0.956992	23.2514881	1
		1300.36	84.349896	1313.73	17.21792				849.79393
SW102.01	661	913	8	961	41	6.3621531	0.997996	499.001996	8
		2380.54	191.69492	2405.20	15.69208	6.6029326			
SW102.02	798	688	1	226	59	9	0.998492	663.129974	1832.3246
		758.794	47.138486	755.958	15.62673	5.9716331			369.39039
SW102.03	484	393	3	608	43	7	0.99687	319.488818	3
		793.045	43.863284	800.693	15.20757				
SW102.04	522	455	9	061	04	6.0783177	0.997264	365.497076	440.66336
		1037.39	72.873095	1011.43	17.55745	6.1260082			518.14414
SW102.05	557	823	3	725	61	5	0.997286	368.459838	6
		545.011	31.403449	565.668	12.82308	5.6877861			236.27209
SW102.06	391	111	6	334	41	8	0.995596	227.066303	9
			2.2303578	77.2480	4.314111	4.0450301			19.113207
SW109.01	76	77.5	1	166	34	7	0.97858	46.6853408	7
			41.2737	2.709759					8.6065827
SW109.02	41	41	0.2469324	255	07	3.4328486	0.961836	26.2027041	4
			60.039323	973.887	18.10678	6.1203808			503.93733
SW109.03	551	946	6	63	14	5	0.997308	371.471025	8
		107.857	6.3350402	104.429	4.952243	4.3188250			27.662922
SW109.04	100	143	3	098	42	3	0.984176	63.1951466	1
		144.807	1.1265561	146.463	5.203400				46.134760
SW109.05	144	692	4	757	52	4.7150251	0.989222	92.7815921	6
			2.3261795	44.7873	2.485914	3.5940011			9.4113548
SW109.06	44	45	1	431	29	9	0.968868	32.12129	7
		458.222	33.945054	435.872	9.938721	5.4450033			160.03554
SW109.07	317	222	8	385	04	3	0.994486	181.356547	6
		82.8571	1.3946665	83.3088	4.499746	4.1874380			21.150173
SW110.01	82	429	3	896	92	5	0.982186	56.1356237	3

SW110.02	527	825.411	48.780317	772.572	14.60961	6.1071805			450.97506
		765	8	416	38	1	0.997406	385.505012	8
		798.962	39.216606	830.401	15.60662	6.1457769			496.98551
SW110.03	548	264	9	116	49	6	0.997474	395.882819	9
					3.603239				14.013230
SW110.04	60	60	0	60	28	3.8856459	0.97624	42.0875421	2
		102.333	0.7585402	102.712	4.970449	4.3883292			28.423670
SW110.05	102	333	2	117	37	4	0.98537	68.3526999	5
			18.055641	293.765	7.543141	5.1113165			99.470471
SW110.06	239	296.5	1	997	39	5	0.992004	125.062531	7
					2.247805	3.4529103			7.8209646
SW112.01	38	38	0	38	95	5	0.963954	27.7423293	9
			0.4933771	38.2777	1.942205	3.4570502			7.8209646
SW112.03	38	38	9	778	17	9	0.964716	28.3414579	9
			1.2952673	57.4978	3.514503	3.7856122			13.111649
SW112.04	57	57.5	1	983	42	5	0.972816	36.7863449	3
		350.258		346.502	7.517912	5.4187040			139.50889
SW112.05	293	621	16.574448	027	2	1	0.994522	182.548375	4
		96.6666		96.1614	4.781793	4.3380314			25.793473
SW112.06	95	667	2.2045363	415	98	2	0.9846	64.9350649	8
					2.065116	3.3471525			6.8040632
SW112.07	34	34	0	34	43	7	0.960178	25.1117473	3
		998.937	55.867297	1045.23	17.58806	6.2536799			645.37092
SW114.01	604	5	2	435	44	5	0.997728	440.140845	7
		386.571	17.559381	375.829	7.422961	5.5696491			165.45875
SW114.02	323	429	7	321	37	5	0.99548	221.238938	9
		785.269	38.483632	812.478	15.36338	6.1335031			
SW114.03	542	231	2	316	57	9	0.997442	390.930414	483.37524
		2838.97	254.66957	2870.12	16.33949	6.6092716			1946.6333
SW114.04	807	222	4	007	29	9	0.998482	658.761528	9
		495.142	29.713622	473.017	10.17081	5.6495506			205.95653
SW114.05	364	857	5	313	75	8	0.995624	228.519196	8
		88.6666	4.4908603	87.4215	4.615603	4.1227654			21.844089
SW114.07	84	667	8	719	31	6	0.98023	50.5816894	3
		0.0992509		67.2028	4.068502	3.9451636			16.184541
SW114.08	67	67	3	536	12	1	0.976966	43.4140835	4
		9.6567964		311.412	5.471092	5.4390300			132.26882
SW115.01	284	311.55	4	824	91	6	0.994722	189.465707	2
		2898.46		2884.04	13.35152	6.6685517			2419.7627
SW115.02	837	61	249.31555	551	03	5	0.998622	725.689405	8
		1466.80		1517.40	18.08361	6.4207845			1004.4280
SW115.03	694	667	98.459852	599	24	8	0.998112	529.661017	4
		822.007	44.036882	824.114	15.88440	6.1353856			485.61692
SW115.04	543	092	6	014	11	3	0.997444	391.236307	7
		20.348782		328.695	8.397857	5.1688365			
SW115.06	262	333.4	6	018	74	4	0.992		125
		126.47829		1753.29	17.03139	6.4819939			115.55145
SW116.02	728	1740.7	5	48	67	9	0.99825	571.428571	1204.2124
		2207.68	167.39303	2281.26	15.23266	6.6061864			
SW116.03	798	085	3	827	02	1	0.998506	669.344043	1832.3246
		1813.15	134.05897	1866.32	17.50813	6.4907478			1251.8622
SW116.05	735	217	7	534	57	8	0.998256	573.394495	8

SW116.07	542	930.128	59.339305	999.748	19.75033	6.0799997				
		1447.09	92.913821	1475.47	16.84388	6.4515826				
SW116.08	706	317	4	377	07	4	0.998206	557.413601	7	
		713.439	38.111760	787.171	16.89353	5.9544505				374.59615
SW116.09	487	394	3	713	21	1	0.996664	299.760192	7	
		366.428	21.837920	368.981	8.768288	5.3205579				133.06144
SW122.01	285	571	2	817	86	9	0.99364	157.232704	7	
		2.5842331	69.2796	4.071990						16.184541
SW122.02	67	69	3	504	03	3.8993284	0.975504	40.8229915	4	
		413.838	15.307925	421.873	8.208279					195.49644
SW122.03	354	71	4	189	85	5.6665673	0.995926	245.459008	5	
		131.666	8.0006549	126.221	5.170465	4.4950221				35.605862
SW122.05	120	667	2	269	05	9	0.986472	73.920757	2	
		105.333	4.1251151	103.015	4.694652	4.4101508				28.423670
SW122.06	102	333	3	706	7	5	0.985426	68.6153424	5	
		0.6206871	50.8760	3.353897	3.6167772					11.076870
SW123.01	50	50.2	9	602	4	8	0.968186	31.4327026	8	
		0.4890096	23.5247		2.8611842					4.1993718
SW123.02	23	23	5	303	1.7591	2	0.933426	15.020879	9	
		74.1111	75.6248	4.313761	3.8457485					17.467286
SW123.04	71	111	3.1006765	977	74	6	0.972622	36.5256776	6	
		347.837	30.158536	331.011	8.837619	5.0507301				101.49086
SW123.05	242	838	5	052	16	8	0.991038	111.582236	4	
				1.371988	3.3944481					6.8040632
SW123.06	34	34	0	34	68	7	0.96294	26.9832704	3	
		4.3392619	100.045	4.810379	4.3102378					26.909539
SW123.08	98	102.2	8	873	46	8	0.98365	61.1620795	2	
				2.701351	3.3406406					7.5634245
SW123.09	37	37	0	37	01	8	0.958218	23.9337514	5	
		41.605091	761.943	15.29225	5.9929967					385.21207
SW124.01	493	740	3	89	28	9	0.996924	325.097529	7	
		1292.58	81.263119	1335.18	17.82383	6.3785069				884.35986
SW124.02	669	599	2	265	77	6	0.998028	507.099392	2	
		32.108803	498.286	10.69483	5.6405265					209.17261
SW124.03	367	514	5	973	89	3	0.99553	223.713647	1	
		896.641	927.474	18.04594	6.0501102					448.89384
SW124.04	526	026	58.341897	031	61	2	0.99703	336.700337	2	
		864.282	53.513328	924.436	18.47727	6.0417601				444.75983
SW124.05	524	258	7	656	51	1	0.996972	330.250991	4	
		894.063	47.921104	960.870	18.78322	6.1707030				552.94865
SW124.06	571	694	1	215	19	1	0.9974	384.615385	2	
		667.657	30.555908	697.271	14.53763	6.0207062				385.21207
SW127.01	493	343	3	655	25	1	0.99709	343.642612	7	
		78.6666	2.2042773	78.9334	4.343880	4.0427247				19.448027
SW127.03	77	667	9	648	99	9	0.978958	47.5239996	4	
				2.641022	3.5083058					8.3425609
SW127.04	40	40	0	40	53	8	0.965962	29.3789294	6	
		196.111	7.8214233	194.488	5.495054	4.9072919				64.054480
SW127.05	180	111	9	214	92	7	0.990942	110.399647	1	
		1.0591134	100.137	4.847002	4.3011674					27.285333
SW127.06	99	99.6	4	629	36	3	0.982466	57.032052	1	

SW127.07	43	43	0	43	1	3	0.968068	31.3165477	9.1409943	4	
		392.484	20.896337	394.986	8.967902	5.4558879				155.60752	
SW128.01	312	375	4	534	23	1	0.99458	184.501845	3		
		150.076		154.143	5.565713	4.3789688				45.673925	
SW128.02	143	923	4.1280238	344	67	5	0.982148	56.0161327	4		
		0.4866642	19.3706	2.048086	2.3949373				3.3283031		
SW128.03	19	19	6	19	57	6	0.881198	8.41736671	1		
				3.903123					15.242462		
SW128.04	64	64	0	64	75	3.9237705	0.975576	40.9433344	8		
				2.215646	3.2258587				6.5554321		
SW128.05	33	33	0	33	84	1	0.952642	21.1157566	1		
				1.309307	2.2966286				2.3044443		
SW128.06	14	14	0	14	34	3	0.875366	8.02349279	1		
		861.389	53.433810	830.197	15.52612	6.0907763			453.06585		
SW129.01	528	831	7	982	43	3	0.997306	371.195249	4		
		538.780	29.045800	537.935	11.50814	5.7503007					
SW129.02	399	22	4	175	23	1	0.996034	252.143217	245.87671		
		828.436	49.952721	802.482	15.18090				438.62905		
SW129.03	521	975	2	816	18	6.0772823	0.997272	366.568915	3		
		911.059	54.945604	970.392	18.66920	6.1061963					
SW129.04	549	702	2	393	72	6	0.997182	354.861604	499.29176		
		42.080110	641.626	14.17244	5.7986501				279.25447		
SW129.05	425	646.625	5	598	85	5	0.996154	260.0104	9		
		1142.28	78.015661	1180.96	19.09270	6.2106376			621.49858		
SW129.06	596	347	3	226	1	2	0.997542	406.834825	1		
				2.190890					5.8233357		
SW31.01	30	30	0	30	23	3.0852854	0.945866	18.4726789	9		
				0.963624	2.2390348				2.3044443		
SW31.03	14	14	0	14	11	1	0.858202	7.05228565	1		
				1.839732					4.8804419		
SW31.04	26	26	0	26	42	3.0493745	0.946344	18.6372447	6		
		79.3333	0.9249512	79.4706	4.431733	4.1405708			20.123282		
SW31.05	79	333	5	263	96	2	0.98132	53.5331906	8		
				1.322875	2.5340035				2.7047898		
SW31.06	16	16	0	16	66	6	0.911206	11.2620222	6		
		0.4937104	40.1868	2.684726	3.4847579				8.3425609		
SW31.07	40	40	4	67	94	6	0.96575	29.1970803	6		
		1083.81		1081.42	16.80824	6.2911341			696.29238		
SW33.01	620	579	64.532596	018	14	2	0.997848	464.684015	7		
		362.205	12.554748	362.792	6.639766				162.73204		
SW33.02	320	48	6	142	2	5.5708467	0.995528	223.613596	9		
		1428.79	98.343364	1366.97	16.58372	6.3986478			925.37053		
SW33.03	678	286	1	366	52	9	0.998096	525.210084	6		
		1639.92	112.71225	1628.25	16.21770	6.4920891					
SW33.04	729	715	2	185	83	6	0.998294	586.166471	1210.8657		
		2501.04	199.10825	2617.03		6.6425969			2147.5297		
SW33.05	821	511	3	967	14.67729	6	0.99857	699.300699	2		
		1303.60	84.013939	1349.35	18.81321	6.3653057			862.55656		
SW33.06	664	265	2	139	05	1	0.997982	495.540139	8		
		1177.84	79.315215	1203.65	18.52369	6.2509562			670.28051		
SW38.03	612	091	4	042	44	9	0.997674	429.922614	5		

SW38.05	124	128.230 769 183.230	3.5599718 1 10.632199	127.799 801 186.067	4.976695 05 6.446704	4.5357387 8 4.6670977			37.284421 1 53.283425
SW38.08	159	769	11.643834	433 86.0334	68 4.512650	2 3.8596990			2 19.784719
SW40.01	78	93.6	3 1.5802298	614 87.1796	94 4.619396	8 4.1893661			8 22.545448
SW40.03	86	87	7 3.4443596	919 116.662	96 5.085906	4 4.4602852			7 33.144492
SW40.05	114	117.5	9 0.4947642	351 48.1610	97 2.873662	1 3.7165490			1 10.794182
SW40.07	49	49	0 5	49 024	79 47	6 3.7158136			1 10.513519
SW40.08	48	48	664.853	34.724433	702.893	14.94659			9 344.33832
SW40.09	469	659	1 172.75986	196 2170.37	72 17.03852	2 6.5359544			1 1462.4471
SW43.04	762	2152.04	4 446.181	26.003820	263 434.897	82 9.737036			6 179.55857
SW43.05	338	818	6 209.972	751 5.5984958	3 212.743	6 5.474219			6 74.599851
SW43.06	199	973	7 763.266	621 41.327118	28 817.411	1 16.79084			3 416.84738
SW43.07	510	667	6 1757.07	937 125.32180	3 1871.02	3 18.99345			1 1273.0791
SW43.08	738	534	9 633.294	408 38.729967	74 644.634	6.4967984 13.91327			4 283.33186
SW43.09	428	737	4 1410.72	653 94.611359	25 1451.75	5 18.50389			7 263.019463
SW46.01	680	109	1 715.722	494 38.512163	5 733.864	6.3937451 14.59347			934.8075 379.86965
SW46.02	490	222	3 382.166	85 21.991383	81 382.836	8 8.730470			9 143.63640
SW46.04	298	667	6 1852.18	675 131.96638	75 1875.62	6 16.01166			8 1445.1947
SW46.05	760	121	9 1363.33	984 93.472807	99 1397.52	8 18.65401			1 621.890547
SW46.06	659	813	9 1147.76	822 69.956191	17 1162.16	6 17.26003			2 841.41469
SW49.01	634	471	6 959.281	993 57.225214	7 1041.20	6 19.01996			1 550.38240
SW49.03	570	69	4 225.028	129 8.3254188	96 225.001	8 5.352650			8 383.729854
SW49.04	206	571	3 9.1880682	183 215.042	5 4.734906	5 5.1035270			2 78.684440
SW49.05	203	222.375	9 3696.56	156 356.66937	71 3821.17	6 13.28291			8 141.964793
SW49.07	861	122	6 160.687	219 6.5400071	27 157.389	2 4.955924			9 2946.5009
SW50.01	150	5	8 0.4930066	41 36.3805	38 1.828162	5 3.3943770			4 93.405567
SW50.02	36	36	5 41	46 41	2 46				3 7.3080809

SW50.04	77	77	0.4967426	77.1556	4.371533	4.1016497			19.448027
		270.634	14.994995	273.208	7.329285	5.0486945	0.978962	47.5330355	4
SW50.05	226	146	6	032	65	4	0.99152	117.924528	5
			14.347272	274.058	7.230033	5.0337627			90.972487
SW50.06	226	269.4	6	509	75	1	0.991252	114.311843	5
		421.245	32.003419	422.220	10.58460	5.2786440			137.87913
SW51.04	291	283	8	328	53	6	0.993042	143.71946	6
			0.0990867	55.3411	3.295062	3.7914025			12.520377
SW51.05	55	55	4	99	38	6	0.97375	38.0952381	3
			5.3706097	78.9042	4.324772	4.0147670			19.113207
SW51.06	76	81.25	9	093	9	9	0.97785	45.1467269	7
		150.769	7.3624733	146.966	5.283612	4.6034867			43.850023
SW51.08	139	231	9	957	92	6	0.987286	78.6534529	6
			0.1234276	40.3710	2.809075	3.4342469			8.3425609
SW51.09	40	40	1	482	33	6	0.963066	27.0753236	6
		162.437	8.1461593	159.136	5.737042	4.6733323			
SW58.01	148	5	8	152	13	3	0.98829	85.3970965	47.997644
			27.463211	173.401	6.877900	4.3815745			39.425196
SW58.02	129	190.75	4	199	91	9	0.984006	62.5234463	8
			3.9161574	81.2211	4.368121				19.113207
SW58.03	76	80.5	1	229	71	3.8240742	0.971712	35.3506787	7
			1.2954545	60.5070	3.621399	3.9010984			14.013230
SW58.06	60	60.5	5	73	39	5	0.97694	43.3651344	2
					3.198557	3.7778140			11.648288
SW58.07	52	52	0	52	37	1	0.973852	38.2438427	1
					3.135249	3.6251846			10.234894
SW58.09	47	47	0	47	15	6	0.9697	33.0033003	7
		2218.63	168.30984	2254.01	14.65760	6.6111904			1856.8282
SW63.02	800	83	8	11	05	3	0.998522	676.589987	8
			55.708170	1015.40	18.56500	6.2140248			595.80752
SW63.03	587	971.2	2	894	61	8	0.997602	417.014179	1
		1402.29		1394.20	18.30181	6.3834456			902.29902
SW63.04	673	787	95.766148	466	41	8	0.998032	508.130081	9
		1151.76	70.944254	1149.66	17.10747	6.3137867			741.09718
SW63.05	633	667	4	334	1	6	0.997894	474.833808	4
		1122.24	65.186885	1154.21	17.85312				766.62362
SW63.06	640	54	4	806	42	6.329736	0.997926	482.160077	5
		516.944	23.458376	539.202	11.40023	5.7776168			253.27906
SW63.07	405	444	7	793	07	9	0.996194	262.743037	7
			0.4947642	48.1944	2.971564	3.7002456			10.513519
SW64.01	48	48	5	849	78	5	0.972234	36.0152705	9
			0.2462960	34.4562	2.009731	3.2804634			6.8040632
SW64.02	34	34	9	359	91	4	0.955036	22.2400142	3
		52.3333	0.9242565	52.7819	3.208699	3.6578331			11.648288
SW64.03	52	333	8	731	85	7	0.968262	31.5079715	1
			0.4435179	63.7052	3.955878				14.932265
SW64.04	63	63.125	9	147	82	3.6988139	0.968618	31.8654006	5
				41.2576	2.803450	3.4153709			8.6065827
SW64.06	41	41	0.2469324	295	83	2	0.95907	24.431957	4
		0.4918693	31.3759	2.176306	3.1894652				
SW64.07	31	31	8	414	75	3	0.953168	21.3529211	6.0650427

SW65.03	26	26	0	26	97	8	0.942742	17.4648084	4.8804419	6	
			1.8138697	45.4161	3.142597	3.4204977			9.4113548		
SW65.05	44	45	5	932	25	4	0.960964	25.6173788	7		
					2.978181	3.5906407			9.9583175		
SW65.06	46	46	0	46	53	7	0.966146	29.538607	9		
			0.2456518	29.4444	1.666988	3.0944588			5.5839911		
SW65.07	29	29	4	444	54	3	0.947566	19.0715948	2		
					1.358732	2.9796503			4.8804419		
SW65.08	26	26	0	26	44	4	0.936222	15.6793879	6		
	1541.64	103.54737		1673.12	20.12552	6.4499819			1104.1769		
SW65.09	712	286	9	092	32	9	0.99815	540.540541	9		
	947.271	62.304263		958.196	18.50230	6.1037410			487.86919		
SW65.10	544	186	9	283	51	2	0.997246	363.108206	8		
					2.215646	3.2698721			6.5554321		
SW781.03	33	33	0	33	84	7	0.956548	23.0139004	1		
					2.504050	3.3988180			7.5634245		
SW781.05	37	37	0	37	77	8	0.962778	26.865832	5		
					1.788854				3.5419457		
SW781.06	20	20	0	20	38	2.6724301	0.916226	11.9368778	8		
					1.809068	2.7808027					
SW781.07	22	22	0	22	07	3	0.925568	13.4350817	3.9775371		
					2.018433	2.9845488			5.1125109		
SW781.08	27	27	0	27	57	4	0.936848	15.8348113	2		
					1.658312	3.4308854			7.3080809		
SW782.01	36	36	0	36	4	7	0.96345	27.3597811	3		
			2.2305169	83.0866	4.500469				21.150173		
SW782.02	82	83.5	4	84	05	4.2050299	0.9829	58.4795322	3		
	292.108	13.706247		292.026	6.788258	5.2337778			107.68916		
SW782.03	251	696	1	065	42	4	0.99333	149.925038	4		
	280.576	22.286481		254.600	6.628178	5.0358842			85.327629		
SW782.09	217	923	4	492	14	1	0.991516	117.86893	4		
	723.038	38.663407		766.500	15.83018				385.21207		
SW786.01	493	168	6	847	16	5.9810855	0.996788	311.332503	7		
	735.857	35.791741		756.524	14.82240				434.58801		
SW786.02	519	143	1	259	52	6.0826	0.9973	370.37037	5		
	741.791	43.869913		743.850	15.16405				369.39039		
SW786.03	484	304	6	712	72	5.9462728	0.996526	287.85262	3		
			25.764350	467.603	10.14834	5.6409390			201.72496		
SW786.06	360	471.8	9	276	56	1	0.995624	228.519196	6		
				376.528	8.822307	5.3767999			143.63640		
SW786.07	298	369.25	19.054615	849	66	8	0.993932	164.798945	8		
	1012.78			1072.72	19.09012				612.80147		
SW786.09	593	378	60.040602	665	33	6.2193803	0.997598	416.319734	3		
	1499.92	99.820617		1534.39	17.08353	6.4470481			1069.5348		
SW787.01	706	857	3	205	78	5	0.998186	551.267916	7		
	1056.46			1114.73	18.86348	6.2232462			624.42799		
SW787.02	597	154	65.253601	09	22	9	0.997602	417.014179	6		
	2767.94	236.81439		2755.97	14.27855	6.6440253			2178.8469		
SW787.03	823	068	3	481	99	7	0.998572	700.280112	4		
	1982.06	141.77635		2026.35	15.14634	6.5868475			1673.6418		
SW787.04	784	579	9	704	98	1	0.99848	657.894737	1		

SW787.05	482	646.802 92 297.396	29.564399 1 11.386718	665.689 818 297.021	13.94684 58 6.246260	5.9994619 6 5.3102846	0.997048	338.753388	365.95694 7 117.01568
SW787.06	264	226	2	915	36	6	0.993794	161.134386	1 14.932265
SW788.01	63	63	0	63	01	7	0.979754	49.3924726	5 2.5030219
SW788.02	15	15	0	15	18	8	0.879288	8.28418053	3 2.7047898
SW788.04	16	16	2	16.584	71	5	0.876364	8.08825908	6
SW788.05	11	11	0	11	59	7	0.844528	6.43202635	1.7290385 5.3470387
SW788.06	28	28	5	479	8	2	0.94003	16.6750042	8 8.8727322
SW788.07	42	42	8	794	97	3	0.96585	29.2825769	1 4.4238312
SW788.10	24	24	1	198	51	8	0.948544	19.4340796	1 40.732534
SW789.01	132	192.2	3	52	8	6	0.982354	56.6700669	1 27.662922
SW789.06	100	100	3	378	97	8	0.985366	68.3340167	1 10.513519
SW789.07	48	48	6	802	7	2	0.970726	34.1600055	9 147.84082
SW789.08	303	148	8	768	9	5.4550575	0.994744	190.258752	4 141.15073
SW790.01	295	337.6	3	637	78	7	0.995088	203.583062	4 6.5554321
SW790.03	33	33	8	898	31	8	0.951062	20.4340186	1 9.9583175
SW790.07	46	47	6	713	18	9	0.962304	26.5280136	9 2.9096498
SW793.02	17	17	0	17	33	2.4908301	0.902418	10.2477916	3 7.0549535
SW793.03	35	35	0	35	91	8	0.954492	21.9741584	3 3.1175139
SW793.06	18	18	0	18	83	8	0.917158	12.0711716	4 3.5419457
SW793.09	20	20	0	20	43	4	0.930386	14.3649266	8 1.5445238
SW793.10	10	10	0	10	3	9	0.810434	5.27520758	2
SW795.04	22	22	0	22	11	9	0.930508	14.3901456	3.9775371 7.5634245
SW795.07	37	37	6	31	65	1	0.96396	27.7469478	5 2.1091676
SW795.09	13	13	0	13	92	2.3976174	0.898864	9.887676	9 2.7047898
SW795.10	16	16	0	16	5	2.4102199	0.896332	9.64617818	6 2.9096498
SW796.03	17	17	3	148	65	2	0.909696	11.0737066	3

SW796.04	26	26	0	26	1.358732	3.1038568			4.8804419
		149.555	4.9829943	149.631	5.390597	4.6730675	0.95008	20.0320513	6
SW796.05	142	556	8	339	72	8	0.988782	89.1424496	5
					2.070196				7.0549535
SW796.06	35	35	0	35	68	3.3809776	0.96205	26.3504611	3
					3.125290	3.4371735			9.1409943
SW796.10	43	43	0	43	68	4	0.959992	24.995001	4
					1.620185	2.9123211			4.4238312
SW797.01	24	24	0	24	18	6	0.93673	15.805279	1
					3.603117	3.8380698			13.111649
SW797.02	57	57	0	57	56	1	0.974412	39.0808191	3
					1.596871	2.6838457			3.5419457
SW797.04	20	20	0	20	94	1	0.919916	12.4868888	8
					0.963624	2.2614517			2.3044443
SW797.06	14	14	0	14	11	6	0.870718	7.73502885	1
		563.082	27.427164	578.552	12.38594	5.8351822			279.25447
SW797.09	425	569	4	045	97	9	0.996424	279.642058	9
					3.195741	3.5668079			10.234894
SW798.05	47	47	0	47	85	6	0.965328	28.8417167	7
					0.982607	3.2202277			5.5839911
SW798.06	29	29	0	29	37	5	0.955364	22.4034412	2
		110.571	2.5150130	112.088	5.114972	4.3651484			30.750531
SW798.09	108	429	3	235	26	8	0.984544	64.699793	5
			0.2457980	30.4786	2.053989	3.2099464			5.8233357
SW798.10	30	30	2	994	87	8	0.955386	22.4144887	9
					1.333333	2.7259285			3.1175139
SW799.03	18	18	0	18	33	1	0.928784	14.0417884	4
					1.866513				
SW799.05	31	31	0	31	05	3.2578298	0.957222	23.3765019	6.0650427
		55.3333	0.9243672	55.5916	3.449167	3.7821489			12.520377
SW799.07	55	333	7	843	61	8	0.974084	38.5862016	3
			16.813203	209.034	6.798887	4.7630813			60.363131
SW799.09	173	216.125	6	524	43	8	0.989052	91.3408842	1
		195.791	11.029161	192.486	6.073507				59.327189
SW799.10	171	667	9	382	98	4.7611125	0.988738	88.7941751	1
			0.1647618	44.3347	2.854134	3.5303744			9.4113548
SW802.04	44	44	4	037	42	6	0.96553	29.010734	7
					3.174901	3.7404316			11.076870
SW803.03	50	50	0	50	57	7	0.972994	37.0288084	8
			0.4909902	28.4176	1.767283	3.1044229			5.3470387
SW803.04	28	28	5	955	31	3	0.948632	19.4673727	8
					1.341640	2.7913704			3.5419457
SW803.05	20	20	0	20	79	2	0.931386	14.5742851	8
		51.3333		51.7428	3.216859	3.6694280			
SW803.06	51	333	0.9242168	336	98	1	0.969834	33.1499039	11.361576
			12.108790	186.530	5.539810	4.8045816			58.812315
SW803.07	170	196.4	1	556	24	8	0.989402	94.3574259	3
					1.648863	3.3003673			6.3090836
SW803.08	32	32	0	32	25	9	0.958406	24.0419291	3
					1.888800	3.4276346			7.5634245
SW803.09	37	37	0	37	56	8	0.96211	26.3921879	5

SW805.01	31	31	0	31	78	4	0.95692	23.2126277	6.0650427	
			0.2480694	65.2219	3.869117	3.9193552			15.554577	
SW805.02	65	65	7	077	21	5	0.976268	42.1371987	7	
			0.2477168	55.2919	3.052005	3.8404970			12.520377	
SW805.03	55	55	5	282	22	2	0.975504	40.8229915	3	
	407.118	17.846479	404.106	7.986731	5.5998246				179.55857	
SW805.04	338	421	7	58	12	1	0.99555	224.719101	6	
				0.974679	2.8595438				3.5419457	
SW805.05	20	20	0	20	43	6	0.937284	15.9448944	8	
	195.387	2.6507250	197.021	4.033269	5.0639293				70.623828	
SW805.06	192	097	9	517	06	9	0.99266	136.239782	2	
				1.653872	3.3595380				6.8040632	
SW805.07	34	34	0	34	46	4	0.9603	25.1889169	3	
			1.8401708	97.7915	4.795703	4.3725341			26.535662	
SW805.08	97	98.2	9	191	85	2	0.984734	65.5050439	1	
	295.489	14.789478	296.018	6.956748					106.29379	
SW806.01	249	796	3	045	49	5.2054738	0.99304	143.678161	2	
			4.6273642	58.0724	3.499970	3.8171965			13.111649	
SW806.02	57	60	6	869	53	7	0.97427	38.865138	3	
					1.833030	2.9655511			4.6508688	
SW806.03	25	25	0	25	28	9	0.94062	16.8406871	2	
			0.4873397	21.1111	1.212469	2.8358486			3.5419457	
SW806.04	20	20	2	111	61	1	0.93648	15.7430731	8	
			2.3175348		1.928104	3.1881191			5.8233357	
SW806.05	30	31	4	32.12	02	5	0.953784	21.6375281	9	
			0.4923659	33.2941	1.999130	3.3099720			6.5554321	
SW806.06	33	33	6	9	2	1	0.959222	24.5230271	1	
			5.8565645	175.825	4.707394	4.9275278			58.812315	
SW806.08	170	179.5	5	824	14	9	0.991544	118.259224	3	
			0.6202686	86.0507	4.598278	4.1318451			22.193839	
SW806.09	85	85.25	3	291	97	9	0.980432	51.103843	2	
			11.998471	178.978	6.737979				45.215037	
SW808.01	142	170.12	5	261	83	4.4453889	0.984218	63.3633253	5	
			2.8816147	59.6076	3.647304	3.7775234			13.410226	
SW808.02	58	60	7	403	05	4	0.973232	37.3580395	7	
			0.1237684	51.3347	3.156760	3.7051349				
SW808.03	51	51	4	152	34	5	0.972308	36.1115124	11.361576	
				1.866513	3.2341032					
SW808.04	31	31	0	31	05	9	0.95598	22.7169468	6.0650427	
			0.2429563		1.362106				3.1175139	
SW808.05	18	18	3	18.6	71	2.570515	0.912676	11.4516055	4	
	216.562	11.718162	217.109	6.422373	4.8525849				67.849120	
SW808.06	187	5	8	214	53	3	0.989662	96.7305088	5	
	407.246	22.131121	408.480	9.021575	5.4834924				162.73204	
SW808.07	320	154	7	747	09	8	0.99469	188.323917	9	
			194.946	5.639601	4.8970522					
SW809.01	178	204.4	12.108828	914	04	1	0.99097	110.741971	62.989346	
			0.4935481	39.3469	2.351994	3.4342916				
SW809.02	39	39	2	388	6	7	0.962318	26.5378695	8.0806827	
			1.2941636	45.1479	2.681644	3.5009398			9.4113548	
SW809.03	44	44.5	5	562	41	2	0.963256	27.2153277	7	

SW809.04	167	227.714	22.863570	213.805	7.162040	4.6610082			57.279981
		286	5	3	01	8	0.98754	80.2568218	7
		304.615	28.805505	272.532	7.484375	5.0031086			85.327629
SW809.05	217	385	6	498	64	9	0.991174	113.301609	4
				41.3100	2.484734	3.4734062			8.6065827
SW809.06	41	41	0.4938648	306	86	4	0.96344	27.3522976	4
		0.4850712	18.1111	1.201914	2.5543183				2.9096498
SW809.07	17	17	5	111	86	6	0.908938	10.9815291	3
				41.2538	2.373659	3.5205353			8.6065827
SWG32.02	41	41	0.4938648	84	21	5	0.965334	28.8467086	4
		101.333	15.071557	90.1604	4.694679	4.1069418			21.496200
SWG32.03	83	333	8	386	9	9	0.980706	51.8295843	3
				9.2962342	163.648	5.377482			50.850983
SWG32.04	154	170.5	9	136	33	4.7879969	0.990256	102.627258	5
		317.285	8.8111908	322.004	5.593233	5.4785320			
SWG32.05	292	714	6	987	27	2	0.995124	205.086136	138.69251
		353.137	14.792722	345.239	6.557215	5.5101147			148.69107
SWG32.08	304	931	3	587	13	1	0.995228	209.555742	3
					0.966091	2.4866551			2.5030219
SWG39.01	15	15	0	15	78	2	0.904668	10.4896572	3
		528.135	22.387572	526.681	10.84031	5.8614962			275.22571
SWG39.02	422	135	9	273	27	3	0.996624	296.208531	7
		249.807	15.468139	235.861	6.028905	5.0531145			81.068072
SWG39.05	210	692	5	084	78	8	0.992116	126.839168	8
		546.722	26.836037	545.455	11.21525	5.8283957			267.31096
SWG39.06	416	772	2	834	23	3	0.99649	284.900285	5
		13.789171	246.842	6.033289					
SWG39.07	215	252.5	4	409	66	5.0622552	0.992004	125.062531	84.09905
					1.870828	3.2234867			6.3090836
Timber.04	32	32	0	32	69	1	0.951664	20.6885137	3
					0.963624	2.3617307			2.3044443
Timber.06	14	14	0	14	11	5	0.89116	9.1877986	1
		10.091014	206.560	5.448567	4.9804185				69.507464
Timber.07	190	212	5	338	22	1	0.991758	121.329774	7
		891.149	54.734200	954.593	19.35277	6.0832006			474.51298
WS4U.02	538	606	7	369	81	9	0.997136	349.162011	6
		19.937958	341.111	7.778163	5.3394405				129.90842
WS4U.03	281	351	3	39	45	1	0.99381	161.550889	7
		613.032	605.653	12.90464	5.8068552				277.90619
WS4U.05	424	609	36.62875	172	5	4	0.996194	262.743037	9
		138.55759	1951.51	16.95901	6.5255707				1387.0462
WS4U.06	753	1887	6	607	04	8	0.998336	600.961539	8
		619.205	35.894854	641.760	13.53377	5.7890665			283.33186
WS4U.07	428	882	9	819	11	3	0.995836	240.153698	7
		1088.04	66.696762	1137.36	18.48859				667.10792
WS4U.08	611	082	7	841	39	6.2573398	0.997706	435.919791	1
					2.738612	3.0383803			6.3090836
WS98SB.01	32	32	0	32	79	7	0.938828	16.3473485	3
		2426.93	197.63015	2484.73	15.80698	6.6025057			1844.5148
WS98SB.02	799	651	8	584	47	6	0.998486	660.501982	3
		1114.49	71.024387	1119.28	17.63597	6.2590029			663.95241
WS98SB.04	610	64	9	408	84	8	0.997732	440.917108	1

		793.105	45.941701	795.717	15.66741	6.0524142			422.68149
WS98SB.05	513	691	3	803	56	2	0.997164	352.609309	6
		603.319	25.719927	643.275	13.57448	5.9470239			337.92165
WS98SB.06	465	149	7	296	65	6	0.996834	315.85597	5
			86.3404	4.601967	4.1681999				22.193839
WS98SB.07	85	86	1.5802193	574	23	5	0.981536	54.1594454	2
				2.070196	3.3413749				7.0549535
WS98SB.09	35	35	0	35	68	1	0.959136	24.4714174	3
		148.687	6.5399460	147.014	5.396665				43.398891
WS98SB.10	138	5	5	608	83	4.5484329	0.985916	71.0025561	2

828

829 **Supplemental Table 1:**

830 **Alpha Diversity Metrics:** Results from the Alpha Diversity analysis for 365 switchgrass genotypes.

831

family	snp	h2	se	>0.9	>0.1	>0.1 (outliers)
Xanthobacteraceae	5	0.45489741	0.15006082	TRUE	TRUE	TRUE
Sphingomonadaceae	31	0.25080413	0.13608337	TRUE	TRUE	TRUE
Micrococcaceae	89	0.26493375	0.1376574	FALSE	TRUE	TRUE
Chitinophagaceae	NA	0.25752125	0.13692033	FALSE	TRUE	TRUE
Koribacteraceae	6	0.14298423	0.11619551	FALSE	TRUE	TRUE
Mycobacteriaceae	15	0.06959644	0.08384037	FALSE	TRUE	TRUE
Pyrinomonadaceae	8	0.0688696	0.08341623	FALSE	TRUE	TRUE
Pirellulaceae	NA	0.00794786	0.13035936	FALSE	TRUE	TRUE
Myxococcaceae	NA	0	0.17460424	FALSE	TRUE	TRUE
Polyangiaceae	NA	0	0.1601444	FALSE	TRUE	TRUE
Reyranellaceae	18	0	0.13129499	FALSE	TRUE	TRUE
Bacillaceae	12	0	0.11080426	FALSE	TRUE	TRUE
Chthoniobacteraceae	7	0	0.08612535	FALSE	TRUE	TRUE
Nitrosomonadaceae	1	0	0.0821527	FALSE	TRUE	TRUE
A21b	2	0	0.06449511	FALSE	TRUE	TRUE
Haliangiaceae	80	0	0.05771382	FALSE	TRUE	TRUE
Rhodanobacteraceae	11	0	0.05728122	FALSE	TRUE	TRUE
Nitrospiraceae	NA	0	0.05377501	FALSE	TRUE	TRUE
Solibacteraceae	NA	0	0.04764666	FALSE	TRUE	TRUE
Comamonadaceae	5	0	0.04073533	FALSE	TRUE	TRUE
Pedosphaeraceae	NA	0	0.04072739	FALSE	TRUE	TRUE
Gaiellaceae	1	0	0.03478031	FALSE	TRUE	TRUE
Bryobacteraceae	4	0	0.03154768	FALSE	TRUE	TRUE
Gemmamimonadaceae	3	0	0.0254416	FALSE	TRUE	TRUE
Steroidobacteraceae	NA	0	0.01622218	FALSE	TRUE	TRUE
Acidothermaceae	3	-0.0067312	0.00802897	FALSE	TRUE	TRUE
SC-I-84	NA	NA	NA	FALSE	TRUE	TRUE
Unknown Family_4	NA	NA	NA	FALSE	TRUE	TRUE

Hyphomicrobiaceae	NA	NA	NA	FALSE	TRUE	TRUE
Micromonosporaceae	8	0.32336921	0.14287441	FALSE	TRUE	TRUE
Caulobacteraceae	2	0.2011677	0.15233479	FALSE	TRUE	TRUE
Bdellovibrionaceae	12	0.19268472	0.12781354	FALSE	TRUE	FALSE
Xiphinematobacteraceae	4	0.16183436	0.12038463	FALSE	TRUE	TRUE
Nocardioidaceae	1	0.12583433	0.1099005	FALSE	TRUE	TRUE
Oxalobacteraceae	22	0.10620934	0.10302864	FALSE	TRUE	TRUE
Xanthomonadaceae	16	0.08703035	0.09431779	FALSE	TRUE	TRUE
Rhizobiaceae	1	0.08088047	0.16039007	FALSE	TRUE	TRUE
Intrasporangiaceae	91	0.07681519	0.08921872	FALSE	TRUE	TRUE
Sphingobacteriaceae	NA	0.06792159	0.08320344	FALSE	TRUE	TRUE
Thermoanaerobaculaceae	11	0.05084146	0.10051362	FALSE	TRUE	TRUE
Opitutaceae	NA	0.01157036	0.0346631	FALSE	TRUE	TRUE
Diplorickettsiaceae	2	0.00896882	0.03285347	FALSE	TRUE	TRUE
Blastocatellaceae	NA	0	0.63004088	FALSE	TRUE	TRUE
Microcillaceae	NA	0	0.35296219	FALSE	TRUE	TRUE
Acetobacteraceae	NA	0	0.22778964	FALSE	TRUE	TRUE
B1rii41	NA	0	0.08574337	FALSE	TRUE	TRUE
Vermiphilaceae	NA	0	0.08323864	FALSE	TRUE	TRUE
Burkholderiaceae	31	0	0.06809019	FALSE	TRUE	TRUE
Vicinamibacteraceae	20	0	0.0513239	FALSE	TRUE	TRUE
Frankiaceae	2	0	0.03337879	FALSE	TRUE	TRUE
Microbacteriaceae	42	0	0.02255735	FALSE	TRUE	TRUE
Solirubrobacteraceae	2	0	0.02232189	FALSE	TRUE	TRUE
Deulosiaceae	3	0	0.02229932	FALSE	TRUE	TRUE
Ktedonobacteraceae	20	0	0.01184748	FALSE	TRUE	TRUE
Rhizobiales Incertae Sedis	NA	NA	NA	FALSE	TRUE	TRUE
X67-14	NA	NA	NA	FALSE	TRUE	FALSE
TRA3-20	NA	NA	NA	FALSE	TRUE	TRUE
WD2101 soil group	NA	NA	NA	FALSE	TRUE	TRUE
JG30-KF-AS9	NA	NA	NA	FALSE	TRUE	TRUE
Beijerinckiaceae	NA	NA	NA	FALSE	TRUE	TRUE
env.OPS 17	NA	NA	NA	FALSE	TRUE	FALSE
Azospirillaceae	1	0.53923556	0.15319359	FALSE	TRUE	TRUE
Methylophilaceae	NA	0.39228257	0.13234329	FALSE	TRUE	TRUE
Pseudonocardiaceae	3	0.34591637	0.14266118	FALSE	TRUE	TRUE
Isosphaeraceae	24	0.2535165	0.13616372	FALSE	TRUE	TRUE
Paenibacillaceae	11	0.23699145	0.13424919	FALSE	TRUE	FALSE
Clostridiaceae	309	0.18235269	0.12480892	FALSE	TRUE	TRUE
Kineosporiaceae	64	0.17626353	0.12688325	FALSE	TRUE	TRUE

Flavobacteriaceae	211	0.1756527	0.12278504	FALSE	TRUE	FALSE
Iamiaceae	25	0.138249	0.11405764	FALSE	TRUE	FALSE
Geminicoccaceae	50	0.12800102	0.11102073	FALSE	TRUE	TRUE
Labraceae	7	0.11543509	0.13795443	FALSE	TRUE	TRUE
Chthonomonadaceae	33	0.06042788	0.08631568	FALSE	TRUE	FALSE
Phaselicystidaceae	NA	0.04328935	0.04395835	FALSE	TRUE	TRUE
cvE6	14	0.01969061	0.06606464	FALSE	TRUE	FALSE
Anaeromyxobacteraceae	NA	0.01845726	0.02758436	FALSE	TRUE	TRUE
Solimonadaceae	25	0.01797846	0.04217796	FALSE	TRUE	FALSE
Phycisphaeraceae	11	0.01003618	0.07399336	FALSE	TRUE	FALSE
Nakamurellaceae	56	0.00733123	0.0313196	FALSE	TRUE	TRUE
Rubritaleaceae	2	0.00299102	0.02880787	FALSE	TRUE	FALSE
Spirochaetaceae	3	0.00111513	0.0238385	FALSE	TRUE	FALSE
Alicyclobacillaceae	8	5.04E-05	0.03636558	FALSE	TRUE	TRUE
SM2D12	60	0	0.21567637	FALSE	TRUE	FALSE
Moraxellaceae	2	0	0.07885071	FALSE	TRUE	TRUE
Rhodomicrobiaceae	11	0	0.07551665	FALSE	TRUE	FALSE
A0839	1	0	0.05401663	FALSE	TRUE	FALSE
Planococcaceae	1	0	0.04551408	FALSE	TRUE	FALSE
Geodermatophilaceae	NA	0	0.03953639	FALSE	TRUE	TRUE
Hyphomonadaceae	6	0	0.02803625	FALSE	TRUE	FALSE
Streptomycetaceae	11	0	0.0244267	FALSE	TRUE	TRUE
Geobacteraceae	28	0	0.02304121	FALSE	TRUE	TRUE
Pseudomonadaceae	24	0	0.02231061	FALSE	TRUE	TRUE
URHD0088	81	-0.0028494	0.01910465	FALSE	TRUE	FALSE
Abditibacteriaceae	87	-0.005207	0.01375589	FALSE	TRUE	FALSE
Illumatobacteraceae	3	-0.0054262	0.02290175	FALSE	TRUE	FALSE
Verrucomicrobiaceae	1	-1.8867917	79.8301507	FALSE	TRUE	FALSE
Cellulomonadaceae	15	-1.9632452	67.3985434	FALSE	TRUE	TRUE
Fibrobacteraceae	NA	-2.4936366	137.92784	FALSE	TRUE	FALSE
Thermoactinomycetaceae	23	-2.5751422	150.57728	FALSE	TRUE	FALSE
Peptostreptococcaceae	45	-3.8013187	81.2594218	FALSE	TRUE	FALSE
Parachlamydiaceae	14	-4.6415211	71.2198938	FALSE	TRUE	FALSE
Gemmataceae	NA	NA	NA	FALSE	TRUE	FALSE
Dongiaceae	NA	NA	NA	FALSE	TRUE	FALSE
CPla-3 termite group	NA	NA	NA	FALSE	TRUE	FALSE
Acidobacteriaceae (Subgroup 1)	NA	NA	NA	FALSE	TRUE	TRUE
KF-JG30-B3	NA	NA	NA	FALSE	TRUE	FALSE
X37-13	NA	NA	NA	FALSE	TRUE	FALSE
NS11-12 marine group	NA	NA	NA	FALSE	TRUE	FALSE

AKYH767	NA	NA	NA	FALSE	TRUE	TRUE
Nostocaceae	NA	NA	NA	FALSE	TRUE	FALSE
Caldilineaceae	88	0.76067873	0.15879238	FALSE	FALSE	FALSE
Holophagaceae	271	0.65671928	0.15669245	FALSE	FALSE	FALSE
Gallionellaceae	339	0.65416872	0.15683858	FALSE	FALSE	FALSE
Dethiobacteraceae	NA	0.62208092	0.1131101	FALSE	FALSE	FALSE
Amoebophilaceae	243	0.55321342	0.15365691	FALSE	FALSE	FALSE
Coriobacteriaceae	483	0.48429688	0.15094295	FALSE	FALSE	FALSE
Proteiniboraceae	209	0.43875513	0.1493015	FALSE	FALSE	FALSE
Aerococcaceae	231	0.42740944	0.1488965	FALSE	FALSE	FALSE
Sanguibacteraceae	470	0.37437605	0.14575336	FALSE	FALSE	FALSE
Wohlfahrtiimonadaceae	397	0.35941743	0.14514329	FALSE	FALSE	FALSE
Prevotellaceae	69	0.32828338	0.14307546	FALSE	FALSE	FALSE
Sporichthyaceae	197	0.28849102	0.14004792	FALSE	FALSE	FALSE
Alcaligenaceae	368	0.28154792	0.13934373	FALSE	FALSE	FALSE
Defluviicoccaceae	315	0.2798717	0.13911257	FALSE	FALSE	FALSE
Leptospiraceae	229	0.21808269	0.13091858	FALSE	FALSE	FALSE
Cyclobacteriaceae	521	0.20954394	0.13018992	FALSE	FALSE	FALSE
Obscuribacteraceae	219	0.18240802	0.12479559	FALSE	FALSE	FALSE
Spirosomaceae	411	0.17824146	0.12530481	FALSE	FALSE	FALSE
Erysipelotrichaceae	218	0.17748616	0.1232979	FALSE	FALSE	FALSE
Brevibacillaceae	162	0.17234433	0.12269324	FALSE	FALSE	FALSE
Omnitrophaceae	1	0.17212481	0.12300393	FALSE	FALSE	FALSE
Paludibacteraceae	221	0.16980054	0.1239147	FALSE	FALSE	FALSE
Akkermansiaceae	NA	0.16815901	0.12097812	FALSE	FALSE	FALSE
Cryomorphaceae	NA	0.16815901	0.12097812	FALSE	FALSE	FALSE
Bacteroidaceae	95	0.15706174	0.11871881	FALSE	FALSE	FALSE
Leptotrichiaceae	411	0.14468155	0.11591839	FALSE	FALSE	FALSE
Cellvibrionaceae	23	0.13897261	0.11354993	FALSE	FALSE	FALSE
Morganellaceae	NA	0.13096179	0.11149072	FALSE	FALSE	FALSE
Hydrogenophilaceae	418	0.12965953	0.11115586	FALSE	FALSE	FALSE
Terrimicrobiaceae	220	0.12007945	0.10722102	FALSE	FALSE	FALSE
WX65	60	0.11502143	0.10310093	FALSE	FALSE	FALSE
Legionellaceae	154	0.11389058	0.10533955	FALSE	FALSE	FALSE
Sumerlaeaceae	158	0.10742225	0.10392821	FALSE	FALSE	FALSE
Sandaracinaceae	243	0.10576886	0.10177004	FALSE	FALSE	FALSE
Nocardiaceae	35	0.10035008	0.09940176	FALSE	FALSE	FALSE
Halomonadaceae	200	0.08853067	0.15415457	FALSE	FALSE	FALSE
Anaerolineaceae	360	0.08144329	0.09026023	FALSE	FALSE	FALSE
Thermaceae	205	0.07620147	0.14974382	FALSE	FALSE	FALSE

Cytophagaceae	19	0.07041701	0.08371033	FALSE	FALSE	FALSE
Magnetospirillaceae	308	0.06858003	0.09539432	FALSE	FALSE	FALSE
Arcobacteraceae	217	0.06132036	0.07754565	FALSE	FALSE	FALSE
Ruminococcaceae	4	0.05736914	0.07498634	FALSE	FALSE	FALSE
Methyloligellaceae	99	0.05513853	0.07494552	FALSE	FALSE	FALSE
Bacteriovoracaceae	58	0.04963317	0.07027572	FALSE	FALSE	FALSE
Neisseriaceae	117	0.04484694	0.06957396	FALSE	FALSE	FALSE
AKIW781	65	0.04173726	0.06354817	FALSE	FALSE	FALSE
Rubinispphaeraceae	20	0.04112421	0.063642	FALSE	FALSE	FALSE
Gemellaceae	250	0.03846134	0.06204274	FALSE	FALSE	FALSE
Trueperaceae	250	0.03685005	0.06030392	FALSE	FALSE	FALSE
Saccharimonadaceae	250	0.03685005	0.06030392	FALSE	FALSE	FALSE
Oxobacteraceae	250	0.03685005	0.06030392	FALSE	FALSE	FALSE
Oligosphaeraceae	284	0.03302006	0.05765196	FALSE	FALSE	FALSE
Thermomonosporaceae	119	0.0249567	0.0495929	FALSE	FALSE	FALSE
Rhodobacteraceae	22	0.02005565	0.04473901	FALSE	FALSE	FALSE
Pasteurellaceae	302	0.01828478	0.04338533	FALSE	FALSE	FALSE
Lachnospiraceae	104	0.01781246	0.05359081	FALSE	FALSE	FALSE
Selenomonadaceae	302	0.01553585	0.04045114	FALSE	FALSE	FALSE
DEV007	369	0.01132861	0.0358079	FALSE	FALSE	FALSE
Alteromonadaceae	64	0.0105716	0.04212393	FALSE	FALSE	FALSE
Schlesneriaceae	26	0.01026488	0.03455747	FALSE	FALSE	FALSE
Saprospiraceae	199	0.00747033	0.03160117	FALSE	FALSE	FALSE
Actinospicaceae	327	0.00683266	0.03874494	FALSE	FALSE	FALSE
UBA12409	56	0.00375163	0.02858096	FALSE	FALSE	FALSE
Bifidobacteriaceae	267	0.00373363	0.02719774	FALSE	FALSE	FALSE
Sulfurimonadaceae	222	0.00269238	0.02610891	FALSE	FALSE	FALSE
Tannerellaceae	268	0.0022849	0.02572081	FALSE	FALSE	FALSE
Thermomicrobiaceae	221	0.00193853	0.02508234	FALSE	FALSE	FALSE
Vampirovibrionaceae	299	0.00166096	0.02470817	FALSE	FALSE	FALSE
Sulfurospirillaceae	512	0	1.41266348	FALSE	FALSE	FALSE
Sporomusaceae	55	0	0.53576159	FALSE	FALSE	FALSE
Rhodocyclaceae	339	0	0.46270774	FALSE	FALSE	FALSE
Veillonellaceae	NA	0	0.28029337	FALSE	FALSE	FALSE
BSV26	26	0	0.1916218	FALSE	FALSE	FALSE
Micropepsaceae	22	0	0.17047885	FALSE	FALSE	FALSE
Dysgonomonadaceae	320	0	0.10911275	FALSE	FALSE	FALSE
Eggerthellaceae	NA	0	0.10837229	FALSE	FALSE	FALSE
Leuconostocaceae	NA	0	0.10837229	FALSE	FALSE	FALSE
Streptosporangiaceae	267	0	0.09141077	FALSE	FALSE	FALSE

Crocinitomicaceae	7	0	0.08330471	FALSE	FALSE	FALSE
Chloroflexaceae	73	0	0.06733627	FALSE	FALSE	FALSE
Rhodothermaceae	NA	0	0.05466097	FALSE	FALSE	FALSE
LWQ8	240	0	0.03622526	FALSE	FALSE	FALSE
Fimbriimonadaceae	415	0	0.03347359	FALSE	FALSE	FALSE
Acidiferrobacteraceae	67	0	0.02598065	FALSE	FALSE	FALSE
Silvanigrellaceae	5	0	0.02305175	FALSE	FALSE	FALSE
Caloramatoraceae	416	0	0.02255735	FALSE	FALSE	FALSE
Chromobacteriaceae	92	0	0.02255735	FALSE	FALSE	FALSE
Halieaceae	272	0	0.02226287	FALSE	FALSE	FALSE
Weeksellaceae	26	0	0.02212612	FALSE	FALSE	FALSE
Longimicrobiaceae	57	0	0.0220318	FALSE	FALSE	FALSE
Simkaniaceae	5	0	0.02027306	FALSE	FALSE	FALSE
Carnobacteriaceae	538	0	0.0185994	FALSE	FALSE	FALSE
Blattabacteriaceae	NA	0	0.0148498	FALSE	FALSE	FALSE
Nannocystaceae	3	0	0.01281651	FALSE	FALSE	FALSE
Hymenobacteraceae	96	0	0.01103008	FALSE	FALSE	FALSE
Enterobacteriaceae	281	-0.001057	0.02115837	FALSE	FALSE	FALSE
Desulfovibrionaceae	419	-0.0016626	0.02045536	FALSE	FALSE	FALSE
Staphylococcaceae	22	-0.0459734	0.15984776	FALSE	FALSE	FALSE
S32	418	-0.17952	4.0598948	FALSE	FALSE	FALSE
Desulfobulbaceae	408	-0.9083787	105.776702	FALSE	FALSE	FALSE
Barnesiellaceae	408	-0.9083787	105.776702	FALSE	FALSE	FALSE
Psychromonadaceae	278	-1.7288246	91.5697065	FALSE	FALSE	FALSE
Vulgatibacteriaceae	84	-1.7783036	1.20791225	FALSE	FALSE	FALSE
Kaстиaceae	324	-1.8310767	83.8592642	FALSE	FALSE	FALSE
Bogoriellaceae	280	-1.8538133	80.2846575	FALSE	FALSE	FALSE
Idiomarinaceae	169	-2.4796445	131.65033	FALSE	FALSE	FALSE
Microtrichaceae	85	-2.9002719	609.137226	FALSE	FALSE	FALSE
Paracaedibacteriaceae	13	-2.9232174	367.275503	FALSE	FALSE	FALSE
Prolixibacteriaceae	13	-3.0826234	175.050572	FALSE	FALSE	FALSE
Woeseiaceae	73	-3.1008482	168.423304	FALSE	FALSE	FALSE
Demequinaceae	NA	-3.2695624	121.800573	FALSE	FALSE	FALSE
Calalkalibacillaceae	20	-3.3221406	114.828941	FALSE	FALSE	FALSE
Halanaerobiaceae	75	-3.5777428	90.7245235	FALSE	FALSE	FALSE
Achopleplasmataceae	111	-3.6316076	87.8364168	FALSE	FALSE	FALSE
Erysipelatoclostridiaceae	96	-3.7160219	84.1578126	FALSE	FALSE	FALSE
Alcanivoracaceae1	70	-3.7730001	82.1393022	FALSE	FALSE	FALSE
Aneurinibacillaceae	84	-3.8207342	80.6500404	FALSE	FALSE	FALSE
Streptococcaceae	56	-4.0198736	76.1528722	FALSE	FALSE	FALSE

Deinococcaceae	60	-4.0612542	75.4414778	FALSE	FALSE	FALSE
Porphyromonadaceae	23	-7.0024208	86.3138376	FALSE	FALSE	FALSE
Corynebacteriaceae	42	-14.232355	203.724539	FALSE	FALSE	FALSE
KD3-93	NA	NA	NA	FALSE	FALSE	FALSE
Lactobacillaceae	NA	NA	NA	FALSE	FALSE	FALSE
CWT CU03-E12	NA	NA	NA	FALSE	FALSE	FALSE
Desulfitobacteriaceae	NA	NA	NA	FALSE	FALSE	FALSE
JG30-KF-CM45	NA	NA	NA	FALSE	FALSE	FALSE
Christensenellaceae	NA	NA	NA	FALSE	FALSE	FALSE
Unknown Family	NA	NA	NA	FALSE	FALSE	FALSE
Actinomycetaceae	NA	NA	NA	FALSE	FALSE	FALSE
Amb-16S-1034	NA	NA	NA	FALSE	FALSE	FALSE
bac2nit3	NA	NA	NA	FALSE	FALSE	FALSE
27F-1492R	NA	NA	NA	FALSE	FALSE	FALSE
Caedibacteraceae	NA	NA	NA	FALSE	FALSE	FALSE
type III	NA	NA	NA	FALSE	FALSE	FALSE
Symbiobacteriaceae	NA	NA	NA	FALSE	FALSE	FALSE
Bacteroidetes vad in HA17	NA	NA	NA	FALSE	FALSE	FALSE
Shewanellaceae	NA	NA	NA	FALSE	FALSE	FALSE
Parvibaculaceae	NA	NA	NA	FALSE	FALSE	FALSE
LiUU-11-161	NA	NA	NA	FALSE	FALSE	FALSE
Coxiellaceae	NA	NA	NA	FALSE	FALSE	FALSE
Catenulisporaceae	NA	NA	NA	FALSE	FALSE	FALSE
Inquilinaceae	NA	NA	NA	FALSE	FALSE	FALSE
UCG-010	NA	NA	NA	FALSE	FALSE	FALSE
Rubrobacteriaceae	NA	NA	NA	FALSE	FALSE	FALSE
[Eubacterium] coprostanoligenes group	NA	NA	NA	FALSE	FALSE	FALSE
Phormidiaceae	NA	NA	NA	FALSE	FALSE	FALSE
AB1	NA	NA	NA	FALSE	FALSE	FALSE
Streptosporangiales Incertae Sedis	NA	NA	NA	FALSE	FALSE	FALSE
Pseudanabaenaceae	NA	NA	NA	FALSE	FALSE	FALSE
Dermabacteraceae	NA	NA	NA	FALSE	FALSE	FALSE
Caldicoprobacteraceae	NA	NA	NA	FALSE	FALSE	FALSE
NS9 marine group	NA	NA	NA	FALSE	FALSE	FALSE
Puniceicoccaceae	NA	NA	NA	FALSE	FALSE	FALSE
Rickettsiaceae	NA	NA	NA	FALSE	FALSE	FALSE
Desulfitibacteraceae	NA	NA	NA	FALSE	FALSE	FALSE
Chitinibacteraceae	NA	NA	NA	FALSE	FALSE	FALSE
Desulfallas-Sporotomaculum	NA	NA	NA	FALSE	FALSE	FALSE
Syntrophomonadaceae	NA	NA	NA	FALSE	FALSE	FALSE

832 **Supplemental Table 2:**

833 **Bacterial Family Association Data:** Data represents the number of significant SNPs associated with each family,
 834 the heritability estimate and standard error, and an indicator for their presence above the occupancy thresholds 0.9
 835 and 0.1 for the full ASV dataset and 0.1 for the outlier ASV dataset.
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GO term	Ontology	Description	Number in input list	Number in BG/Ref	p-value	FDR	Family
GO:0016887	F	ATPase activity	15	698	0.0044	0.04	Xanthobacte raceae
GO:0016817	F	hydrolase activity, acting on acid anhydrides	26	1415	0.0019	0.02	Xanthobacte raceae
GO:0016818	F	hydrolase activity, acting on acid anhydrides, in phosphorus-containing anhydrides	26	1391	0.0015 0.0003	0.02	Xanthobacte raceae
GO:0016874	F	ligase activity	16	600	8	0.04	Xanthobacte raceae
GO:0003774	F	motor activity	6	152	0.0045 0.0007	0.02	Xanthobacte raceae
GO:0017111	F	nucleoside-triphosphatase activity	26	1324	5	0.02	Xanthobacte raceae
GO:0016462	F	pyrophosphatase activity	26	1357	0.0011	0.02	Xanthobacte raceae
GO:0019787	F	small conjugating protein ligase activity	6	130	0.0021	0.02	Xanthobacte raceae
GO:0004842	F	ubiquitin-protein ligase activity	6	130	0.0021	0.03	Sphingomon adaceae
GO:0005623	C	cell	103	8213	0.0085	0.03	Sphingomon adaceae
GO:0044464	C	cell part	103	8213	0.0085	0.03	Sphingomon adaceae
GO:0016021	C	integral to membrane	27	1616	0.0083	0.03	Sphingomon adaceae
GO:0031224	C	intrinsic to membrane	27	1651	0.011	0.03	Sphingomon adaceae
GO:0016020	C	membrane	58	4123	0.0063 0.0005	0.01	Sphingomon adaceae
GO:0044425	C	membrane part	37	2042	2	0.02	Sphingomon adaceae
GO:0004222	F	metalloendopeptidase activity	6	100	0.0007 0.0000	0.00	Sphingomon adaceae
GO:0008237	F	metallopeptidase activity	8	119	44	0.02	Sphingomon adaceae
GO:0070727	P	cellular macromolecule localization	10	311	0.0016 0.0006	0.01	Sphingomon adaceae
GO:0044237	P	cellular metabolic process	142	11155	9	0.00	Sphingomon adaceae
GO:0009987	P	cellular process	190	14248	0.0079	0.00	Sphingomon adaceae
GO:0034613	P	cellular protein localization	10	311	0.0016 0.0001	0.00	Sphingomon adaceae
GO:0009719	P	response to endogenous stimulus	5	48	9	0.00	Sphingomon adaceae
GO:0009725	P	response to hormone stimulus	5	48	9	0.00	Sphingomon adaceae
GO:0010033	P	response to organic substance	5	48	9	0.00	Sphingomon adaceae
GO:0008026	F	ATP-dependent helicase activity	5	232	0.0084	0.04	Micromonos poraceae
GO:0016887	F	ATPase activity	12	698	0.0004	0.00	Micromonos poraceae
GO:0042623	F	ATPase activity, coupled	9	422	8	0.03	Micromonos poraceae
GO:0004386	F	helicase activity	8	477	0.0042	0.04	Micromonos poraceae
GO:0016787	F	hydrolase activity	43	5652	0.0075 0.0002	0.00	Micromonos poraceae
GO:0016817	F	hydrolase activity, acting on acid anhydrides	19	1415	1	0.00	Micromonos poraceae
GO:0016818	F	hydrolase activity, acting on acid anhydrides, in phosphorus-containing anhydrides	19	1391	7	0.00	Micromonos poraceae

GO:00									
17111	F	nucleoside-triphosphatase activity	19	1324	9	0.0000	0.00	Micromonos	
GO:00								poraceae	
70035	F	purine NTP-dependent helicase activity	5	232	0.0084	0.04	0.04	Micromonos	
GO:00								poraceae	
16462	F	pyrophosphatase activity	19	1357	2	0.0001	0.00	Micromonos	
GO:00								poraceae	
07242	P	intracellular signaling cascade	7	291	0.001	0.04	0.04	Micromonos	
GO:00								poraceae	
07264	P	small GTPase mediated signal transduction	7	289	0.001	0.04	0.04	Micromonos	
								poraceae	

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Supplemental Table 3:

GO Term Data: Data represents the Ontologies, Descriptions, Number of genes in the input list and reference genome, p-value, and FDR for GO terms associated with Xanthobacteraceae, Sphingomonadaceae, and Micromonosporaceae abundances.