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KoT: an automatic implementation of the K/θ method for species delimitation

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Abstract

K/θ is a method to delineate species that rests on the calculation of the ratio between the
average distance K separating two putative species-level clades and the genetic diversity
θ of these clades. Although this method is explicitly rooted in population genetic theory,
it was never benchmarked due to the absence of a program allowing automated analyses.
For the same reason, its application by hand was limited to small datasets of a few tens of
sequences.
We present an automatic implementation of the K/θ method, dubbed KoT (short for "K
over Theta"), that takes as input a FASTA file, builds a neighbour-joining tree, and returns

⁹ putative species boundaries based on a user-specified K/θ threshold. This automatic imple-¹⁰ mentation avoids errors and makes it possible to apply the method to datasets comprising ¹¹ many sequences, as well as to test easily the impact of choosing different K/θ threshold ¹² ratios. KoT is implemented in Haxe, with a javascript webserver interface freely available at

https://eeg-ebe.github.io/KoT/

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¹⁴ Key words: 4X rule, K/θ , species delimitation, molecular systematics, DNA taxonomy

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INTRODUCTION

Methods to delineate species from sets of DNA sequences have been an intense field of 17 research for the last 20 years (Sites and Marshall, 2003; Flot, 2015). Some methods delimit 18 species based on phylogenetic trees, other on genetic distances, and yet others on allele 19 sharing (Fontaneto et al., 2015). Among these methods, one called K/θ (Birky et al., 2010; 20 Birky, 2013; Birky and Maughan, 2020) stands out be resting on the genealogical species 21 concept, in which closely related populations are considered as distinct species when their 22 lineages for a given locus (or set of loci) are reciprocally monophyletic, that is, "if their 23 loci coalesce more recently within the group than between any member of the group and 24 any organisms outside the group" (Baum and Shaw, 1995). Of course, sampling all lineages 25 from a population is usually impossible, but population genetic theory provides ways to 26 calculate the probability that the lineages of two populations are monophyletic given the 27 observation that the sequences samples from these populations form clades in a phylogenetic 28 tree (Hudson and Coyne, 2002). 29

In particular, Rosenberg (2003) provides a formula that uses Watterson's estimator 30 of genetic diversity $\theta = 4N_e\mu$ (Watterson, 1975) of each of two clades of sequences, the 31 number of sequences in each of them, and the mean pairwise sequence difference between 32 the two clades K to calculate the probability that the corresponding two populations are 33 reciprocally monophyletic, i.e. distinct species according to the genealogical species concept. 34 This formula is complex, but when the two θ values are similar and the number of sequences 35 in each clade is higher than three, a useful rule of thumb is that pairs of clades with a K/θ ratio higher than 4 have a probability of at least 0.95 of belonging to different species. This 37 forms the basis of the so-called "4X rule" (Birky and Barraclough, 2009), which has been 38

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widely used to delineate species in a variety of organisms. However, one may wish to choose a more stringent threshold: for instance, a K/θ ratio higher than 6 entails a probability of

 $_{41}$ monophyly higher than 0.99 (according to equation 9 in Rosenberg, 2003).

⁴² Despite the theoretical appeal of this method based on an explicit criterion inspired ⁴³ by population genetic, its practical application has been hampered by the lack of a program ⁴⁴ performing the needed calculations automatically. To fill this gap, we introduce here KoT ⁴⁵ (short for "K over Theta"), an automatic implementation of the K/θ method using the ⁴⁶ programming language Haxe (Dasnois, 2011).

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DESCRIPTION

⁴³ KoT takes as input an alignment of DNA sequences in the FASTA file format. Users ⁴⁹ can specify the K/θ threshold they wish to use to delineate species (by default, 4).

KoT starts by calculating all the pairwise nucleotide distances among the sequences 50 in the dataset. In case the input contains indels and/or missing data (encoded respectively as 51 "-" and as "N" or "?"), users can either ask KoT to filter out completely the corresponding 52 positions in the alignment ("complete deletion" mode) or to retain them ("pairwise deletion" 53 mode, in which case positions with missing or ambiguous data are ignored during pairwise 54 comparisons). From this set of pairwise distances, KoT then computes a neighbor-joining 55 (NJ) tree and the K/θ ratios of each pair of sister clades using the procedure outlined in 56 Birky and Maughan (2020). 57

To compute the genetic diversity θ of each clade, KoT starts by calculating the nucleotide diversity π (Nei and Li, 1979) as the mean of all nucleotide-level differences π_{ij} (number of nucleotide differences per nucleotide site between sequences *i* and *j*) among the $\frac{n(n-1)}{2}$ pairs of sequences in a clade of *n* sequences, i.e. $\pi = \frac{2}{n(n-1)} \sum_{i=1}^{n} \sum_{j=1}^{i-1} \pi_{ij}$ (Equation 10.6 in Nei, 1987). An equivalent way to calculate π found in the literature is to compare all pairs of haplotypes instead of all pairs of sequences: in that case, the formula above becomes $\pi = \frac{1}{n(n-1)} \sum_{ij} (n \times x_i)(n \times x_j) \pi_{ij}$ (where x_i is the frequency of haplotype *i* in the clade),

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which simplifies into $\pi = \frac{n}{n-1} \sum_{ij} x_i x_j \pi_{ij}$ (Equation 10.6 in Nei, 1987), i.e. the average heterozygosity multiplied by a sample size correction n/(n-1) (Nei and Tajima, 1981; Korunes and Samuk, 2021). As KoT uses the direct formula comparing pairs of sequences, however, no sample size correction is needed.

For clades made up of identical sequences (in which case $\pi = 0$), an upper bound of π 69 is sought by assuming that one sequence differs from the others by a single mutation, i.e. by 70 replacing one of the $\frac{n(n-1)}{2}$ pairwise distances with 1/L, where L is the sequence length; in 71 such case π become $\frac{2}{Ln(n-1)}$ (Birky et al., 2010). As this ratio is not defined for n = 1, KoT 72 uses n = 2 (i.e. $\pi = 1/L$) for clades comprising a single sequence. To estimate the genetic 73 diversity θ associated with a specific clade, KoT uses the formula $\theta = \frac{1}{\frac{1}{\pi} - \frac{4}{3}}$ (Equation 9 in 74 Tajima, 1996), which corrects for multiple hits based on the Jukes-Cantor model of sequence 75 evolution (Jukes and Cantor, 1969). 76

To compute the genetic divergence K between sister clades A and B, KoT computes the mean pairwise nucleotide distance $p = \sum_{i=1}^{n_A} \sum_{j=1}^{n_B} \pi_{ij}$ between the sequences in the two clades, where n_A stands for the number of individuals in clade A and n_B stands for the number of individuals in clade B, then corrects it for multiple substitutions using the Jukes-Cantor formula $K = -\frac{3}{4}ln(1-\frac{4}{3}p)$ (Jukes and Cantor, 1969). Using the Jukes-Cantor correction for calculating K is important to ensure that both terms of the ratio K/θ are computed using the same evolutionary model.

Finally, KoT computes the K/θ ratios of clades A and B and compares the smallest 84 of the two with the threshold chosen by the user to delineate species. These calculations 85 are performed iteratively from the leaves of the tree all the way to its root. When a ladder 86 structure ((A,B)C) or a polytomy (A,B,C) is encountered, KoT starts by comparing the two 87 clades separated by the smallest distance K, i.e. A and B: if the result of the calculation 88 does not support the hypothesis that A and B are different species, the A+B clade is then 89 compared to C to find out whether they are conspecific or heterospecific; whereas if the result 90 of the calculation indicates that A and B are likely two distinct species, C is compared with 91

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whichever of A and B has the smaller average distance K to C (Birky et al., 2010). If C is 92 then deemed to be distinct as well, the final result returned is three species A, B and C. 93 On the other hand, if the result of the calculation suggests that C is conspecific with e.g. B, 94 an additional comparison of C with A is warranted to ensure the transitivity of the result 95 (Dellicour and Flot, 2018). As the extra calculations this entails can take lots of time in 96 complex cases, this comparison is only performed if the box "transitivity" is checked by the 97 user prior to running the analysis. If the "transitivity" box is not checked (as by default), 98 KoT simply returns in such cases two species A and B+C; when the "transitivity" box is 99 checked, by contrast, KoT checks whether the K/θ ratio for the C vs. A comparison is also 100 above the user-selected threshold: if so, the final delimitation returned is a pair of species A 101 and B+C; whereas if the calculation does not support the monophyly of A vs. C, the final 102 result returned is a single species A+B+C. 103

¹⁰⁴ KoT outputs a tree in which the θ values of each pair of clades being compared are ¹⁰⁵ displayed on the tree next to the node uniting them, together with their K distance and ¹⁰⁶ the K/θ ratio (obtained using the larger of the two θ values). Colors are applied to the tree ¹⁰⁷ in order to visually delineate the different species. A partition list, i.e. a two-column table ¹⁰⁸ indicating, for each sequence in the input FASTA, the species to which it was attributed ¹⁰⁹ (Spöri and Flot, 2020), is also outputted below the tree where is can be easily copied/pasted ¹¹⁰ into other applications.

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BIOLOGICAL EXAMPLE

To investigate the behavior of KoT, we reanalyzed the COI dataset from one recent article (Stoch et al., 2020). In this article, a dataset of 34 COI sequences of specimens of the *Niphargus tatrensis* species complex was analyzed using a diversity of approaches: mPTP Kapli et al. (2017) delimited seven putative species, ABGD (Puillandre et al., 2012) returned ten of them, and bPTP (Zhang et al., 2013) and ST-GMYC (Pons et al., 2006) delineated eleven species-level units. The methods chiefly differed in their delimitation of

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species among the non-Austrian specimens included in the study but were largely congruent
in their treatment of the Austrian specimens, with mPTP, bPTP and ST-GMYC finding
four species and ABGD delimiting five species in Austria.

When run with a K/θ threshold ratio of 4 (Figure 1), KoT returned twelve species, 121 including five for Austria (separated by K/θ ratios of 21.39, 17.50, 4.52 and 5.98); with 122 a K/θ threshold ratio of 5 (Figure 2), the method returned eleven species, with precisely 123 the same putative boundaries as those obtained using bPTP and ST-GMYC (including 124 four species for Austria separated by K/θ ratios of 21.35, 17.44 and 5.94); finally, with a 125 K/θ threshold ratio of 6 (Figure 3) KoT returned eight species-level units, notably lumping 126 together all Austrian specimens into a single putative species. This highlights the sensitivity 127 of this method to the K/θ threshold parameter. 128

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AVAILABILITY

KoT is written in Haxe. Its source code is available at https://github.com/eeg-ebe/
 KoT, and a javascript webserver is freely accessible at https://eeg-ebe.github.io/KoT/.

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References

Baum, D. A. and K. L. Shaw. 1995. Genealogical perspectives on the species problem. *in* Experimental and molecular approaches to plant systematics (P. C. Hoch and A. G.
Stevenson, eds.) no. 53 in Monographs in systematics. Missouri Botanical Garden, St.
Louis.

Birky, C. W. 2013. Species detection and identification in sexual organisms using population
 genetic theory and DNA sequences. PLoS ONE 8:e52544.

¹³⁹ Birky, C. W., J. Adams, M. Gemmel, and J. Perry. 2010. Using population genetic theory
¹⁴⁰ and DNA sequences for species detection and identification in asexual organisms. PLoS
¹⁴¹ ONE 5:e10609.

REFERENCES

- ¹⁴² Birky, C. W. and H. Maughan. 2020. Evolutionary genetic species detected in prokaryotes ¹⁴³ by applying the K/ϑ ratio to DNA sequences. preprint bioRxiv.
- ¹⁴⁴ Birky, W. C. and T. G. Barraclough. 2009. Asexual speciation. Pages 201–216 in Lost Sex
- ¹⁴⁵ (I. Schön, K. Martens, and P. Dijk, eds.). Springer Netherlands, Dordrecht.
- ¹⁴⁶ Dasnois, B. 2011. HaXe 2 Beginner's Guide. Packt Publishing Ltd.
- Dellicour, S. and J.-F. Flot. 2018. The hitchhiker's guide to single-locus species delimitation.
 Molecular Ecology Resources 18:1234–1246.
- ¹⁴⁹ Flot, J.-F. 2015. Species delimitation's coming of age. Systematic Biology 64:897–899.
- Fontaneto, D., J.-F. Flot, and C. Q. Tang. 2015. Guidelines for DNA taxonomy, with a focus
 on the meiofauna. Marine Biodiversity 45:433–451.
- ¹⁵² Hudson, R. R. and J. A. Coyne. 2002. Mathematical consequences of the genealogical species
 ¹⁵³ concept. Evolution 56:1557–1565.
- Jukes, T. H. and C. R. Cantor. 1969. Evolution of protein molecules. Pages 21-132 in
- ¹⁵⁵ Mammalian protein metabolism (H. N. Munro, ed.) vol. 3. Academic Press, New York.
- Kapli, P., S. Lutteropp, J. Zhang, K. Kobert, P. Pavlidis, A. Stamatakis, and T. Flouri. 2017.
 Multi-rate Poisson tree processes for single-locus species delimitation under maximum
 likelihood and Markov chain Monte Carlo. Bioinformatics 33:1630–1638.
- ¹⁵⁹ Korunes, K. L. and K. Samuk. 2021. pixy: Unbiased estimation of nucleotide diversity and
 ¹⁶⁰ divergence in the presence of missing data. Molecular Ecology Resources 21:1359–1368.
- ¹⁶¹ Nei, M. 1987. Molecular evolutionary genetics. Columbia University Press, New York.
- ¹⁶² Nei, M. and W. H. Li. 1979. Mathematical model for studying genetic variation in terms of
- restriction endonucleases. Proceedings of the National Academy of Sciences 76:5269–5273.
- ¹⁶⁴ Nei, M. and F. Tajima. 1981. DNA polymorphism detectable by restriction endonucleases.
- 165 Genetics 97:145–163.

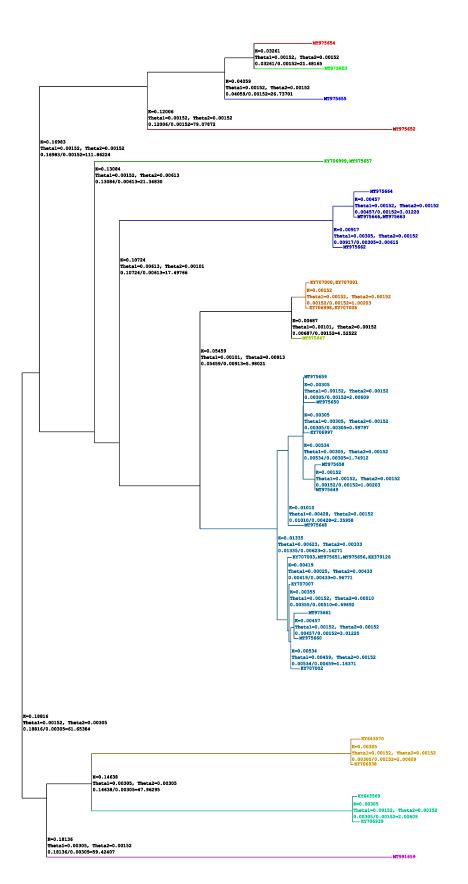
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REFERENCES

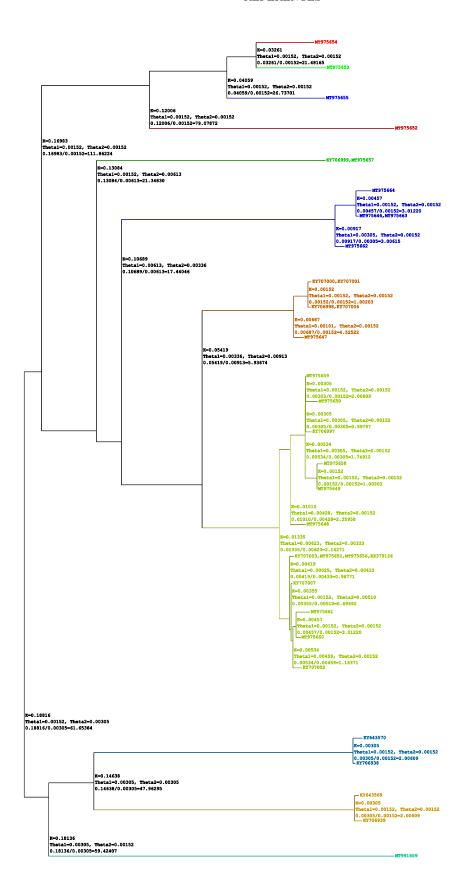
166	Pons, J., T. G. Barraclough, J. Gomez-Zurita, A. Cardoso, D. P. Duran, S. Hazell,
167	S. Kamoun, W. D. Sumlin, and A. Vogler. 2006. Sequence-based species delimitation
168	for the DNA taxonomy of undescribed insects. Systematic Biology 55:595–609.
169	Puillandre, N., A. Lambert, S. Brouillet, and G. Achaz. 2012. ABGD, Automatic Barcode

- Gap Discovery for primary species delimitation. Molecular Ecology 21:1864–1877.
- ¹⁷¹ Rosenberg, N. A. 2003. The shapes of neutral gene genealogies in two species: probabilities ¹⁷² of monophyly, paraphyly, and polyphyly in a coalescent model. Evolution 57:1465–1477.
- Sites, J. W. and J. C. Marshall. 2003. Delimiting species: a Renaissance issue in systematic
 biology. Trends in Ecology & Evolution 18:462–470.
- Spöri, Y. and J.-F. Flot. 2020. HaplowebMaker and CoMa: two web tools to delimit species
 using haplowebs and conspecificity matrices. Methods in Ecology and Evolution 11:1434–
 1438.
- Stoch, F., E. Christian, and J.-F. Flot. 2020. Molecular taxonomy, phylogeny and biogeography of the *Niphargus tatrensis* species complex (Amphipoda, Niphargidae) in Austria.
 Organisms Diversity & Evolution 20:701–722.
- Tajima, F. 1996. The amount of DNA polymorphism maintained in a finite population when
 the neutral mutation rate varies among sites. Genetics 143:1457–1465.
- Watterson, G. A. 1975. On the number of segregating sites in genetical models without
 recombination. Theoretical Population Biology 7:256–276.
- ¹⁸⁵ Zhang, J., P. Kapli, P. Pavlidis, and A. Stamatakis. 2013. A general species delimitation
 ¹⁸⁶ method with applications to phylogenetic placements. Bioinformatics 29:2869–2876.

REFERENCES



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REFERENCES
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REFERENCES

