

1 **Mycorrhizal inoculation and treatment with biochar and compost from pruning waste**  
2 **improve the qualitative properties of a calcareous soil under wheat cultivation**

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9

10 **Abstract**

11 Most calcareous soils have relatively low levels of organic matter. To address this issue and improve  
12 the qualitative properties of calcareous soils, soils can be treated with mycorrhizal fungi and/or  
13 exogenous organic material such as biochar or compost derived from tree pruning waste. To evaluate  
14 the effect of pruning waste biochar (PWB) and pruning waste compost (PWC) derived from apple and  
15 grape trees combined with arbuscular mycorrhizal fungi (AMF) on the biological indices of calcareous  
16 soils, a rhizobox study on wheat plants using a completely randomized design was conducted under  
17 greenhouse conditions. The studied factors included the source of the type of organic material applied  
18 (PWB, PWC, and control), the nature of the microbial inoculation (inoculation with AMF or no  
19 inoculation), and the zone to which the treatments were applied (rhizosphere and non-rhizosphere soil).  
20 At the end of the plant growth period, organic carbon (OC), microbial biomass carbon (MBC), microbial  
21 biomass phosphorous (MBP), microbial respiration (BR), substrate-induced respiration (SIR), alkaline  
22 (ALP), acid (ACP) phosphatase enzyme activities in the rhizosphere and non-rhizosphere soils, and root  
23 mycorrhizal colonization were determined. Simultaneous application of a source of organic matter and  
24 AMF inoculation significantly increased the OC and biological indices of soil relative to those observed  
25 when applying organic matter without AMF inoculation. Additionally, MBC, MBP, ACP and ALP -  
26 enzymes activities in the rhizosphere zone were significantly higher than in the non-rhizosphere. AMF  
27 increased BR and SIR levels in the rhizosphere by 13.06% and 7.95% compared to non-rhizosphere,  
28 respectively. It can be concluded that in calcareous soils with low organic carbon contents, organic

29 amendments such as PWC and PWB can improve soil biological properties by increasing microbial  
30 activity and changing the properties of the rhizosphere.

31

32 **Keywords:** pruning waste, biological Indices, rhizosphere, rhizobox

33

## 34 1. Introduction

35

36 The arid and semi-arid regions have alkaline calcareous soils with low contents of organic matter,  
37 which adversely affects crop yields in these regions [1]. One of the most important ways to improve the  
38 organic matter content is to properly manage the use of plant residues such as waste material from the  
39 pruning of fruit trees. Tree pruning waste can be converted into biochar or compost, which can then be  
40 applied to soils to replenish or increase their nutrient content. This improves the soil's physical and  
41 chemical characteristics and is important for supporting the life of soil microorganisms and maintaining  
42 a sustainable dynamic equilibrium between the biotic and abiotic components of the soils [2, 3]. Biochar  
43 is a carbon-enriched solid generated by the pyrolysis of biomass under conditions with little or no  
44 oxygen [4]. The composition and properties of biochar depend on the pyrolysis conditions including the  
45 temperature, heating rate, duration, pressure, and input material [5]. Conversely, compost is formed by  
46 the decomposition of organic matter mediated by microorganisms under hot, humid, aerobic conditions  
47 [6]. Compost is rich in mineral nutrients, some of which are released and made available to plants in  
48 soil gradually and continuously [7]. In addition to its physical and chemical properties, soil quality is  
49 closely related to its biological characteristics [8]. Biological indicators are thus key descriptors of soil  
50 quality, and variation in soil performance should be measured using biochemical parameters and indices  
51 (e.g., BR, SIR, MBC, MBP, ACP, and ALP) that represent the diversity of soil microorganisms as well  
52 as their distribution and metabolic activity. Soil microbial respiration is essentially a cellular process  
53 that involves several biochemical reactions; the rate of microbial respiration is an indicator of both the  
54 status and activity of soil microbes but also reflects the rate of organic matter decomposition and cycling  
55 of certain nutrients within the soil [9].

56 Enzymatic activity and microbial biomass are the main biological indicators of soil. It seems that  
57 among the biochemical parameters, MBC (41%) and the activity of phosphatase (ACP or ALP) enzymes  
58 (28%) have been widely used [10].

59 There are contradictory reports about the impact of biochar on soil quality. Liu et al. reported that  
60 biochar did not significantly influence soil respiration [11]. However, other studies concluded that the  
61 pores of biochar provide protective habitats for microorganisms, leading to increases in the soil's  
62 content of various mineral nutrients, energy, and carbon; as a result, biochar administration was found  
63 to increase soil biological activity and help maintain soil quality [12, 13]. Another study found that  
64 biochar increased microbial biomass and activity and also, it can enhance enzymatic activities in soil;  
65 the increase in the activity of enzymes like phosphatases and suggested that the increased activity of  
66 microorganisms following biochar administration might improve the availability of nutrients to other  
67 microorganisms as a result of increased root exudation [14].

68 Studies on compost have shown that it contains a large population of microorganisms. Therefore,  
69 in addition to increasing the organic matter and nutrient content of the soil, compost administration  
70 increases and changes its microbial population [15]. Some rhizosphere microorganisms such as AMF  
71 can stimulate soil microbial activity and promote increases in activity of phosphatase and soil microbial  
72 biomass [16]. Accordingly, multiple studies have found that treatment with biochar and compost  
73 promote mycorrhizal root colonization, spore production, and hyphal expansion, thereby improving soil  
74 quality [17, 18]. Active root systems continuously release organic compounds into the rhizosphere that  
75 promote the growth and activity of the microbial community in the soil as well as the overall health of  
76 the soil system [19, 20].

77 The effect of compost and biochar is not confined to the placement of organic matter in and outside  
78 the rhizosphere; it also strongly affects microbial activity in the soil outside the rhizosphere [21]. When  
79 studying rhizosphere processes, it is helpful to confine root growth to a fixed volume of soil in order to  
80 increase root density and accelerate the sampling of the rhizosphere soil. To this end, the concept of the  
81 rhizobox was developed; a rhizobox is like a pot except that it hinders direct contact between the roots  
82 and the soil without disrupting the mobilization of the soil solution [22, 23].

83 This work investigates the effects of applying biochar and pruning waste compost derived from  
84 apple trees and grapevines on some biological characteristics of calcareous soils in the presence of  
85 AMF. The combined effects of treatment with biochar and AMF on soil biological processes have not  
86 been studied extensively, and most biochar administration studies have focused on acidic soils in  
87 tropical and humid climates; consequently, there is little data on their effects in warm and arid regions  
88 with non-acidic soils.

89

## 90 2. Materials and Methods

91

### 92.1. Soil sampling and preparation

93

94 This study used a randomized factorial complete block design with three replicates. The  
95 experiments were conducted under greenhouse condition in rhizoboxes. The factors were the organic  
96 matter source (pruning waste biochar [PWB], pruning waste compost [PWC], and control [no added  
97 organic matter]), microbial inoculation (AMF and non-inoculation [-AMF]), and the zone to which the  
98 treatment was applied (rhizosphere or non-rhizosphere soil). Soil samples were collected from the  
99 surface layer (0–30 cm) of a region of non-arable land in Salmas, West Azerbaijan Province, Iran. The  
100 samples were air-dried and sieved to pass through a 2 mm mesh, then sterilized in an autoclave at 121°C  
101 at 1.5 atm for 2 hours. Before sterilization, some of the soil's physicochemical properties were  
102 determined, as shown in Table 1 [24].

103

104

**Table 1.** Physicochemical properties of the studied soil

soil texture	pH	EC (dS m <sup>-1</sup> )	O.C (%)	CaCO <sub>3</sub> (%)	N (%)	P (mg kg <sup>-1</sup> )	K (mg kg <sup>-1</sup> )
Loamy sand	7.53	0.47	0.25	14.25	0.08	7.64	98

105

### 106.2. Preparation of biochar and compost

107

108 To prepare biochar, fruit tree pruning waste was collected from orchards in Urmia County in West  
109 Azerbaijan Province. The waste was then oven-dried at 65 °C for 48 hours. The dried samples were  
110 placed in a reactor (a steel cylinder with a diameter of 7 cm and a height of 31 cm) and then heated to  
111 350 °C in an electric furnace to form biochar. The PWC was taken from the research greenhouse of the  
112 Soil Science Department in Urmia University. Finally, the PWB and PWC were ground and screened  
113 with a 0.5-mm mesh. No ash was observed on the biochar surface, implying that oxygen had been  
114 removed and it had been produced correctly. Table 2 summarizes the characteristics of the biochar and  
115 compost, which were determined using the analytical methods described by Rajkovich et al. and Alfano  
116 et al. [25, 26].

117

118

**Table 2.** Characteristics of the tested PWC and PWB

Characteristics	unit	Pruning waste biochar (PWB)	Pruning waste compost (PWC)
pH	-	7.29	7.05
EC	dS m <sup>-1</sup>	0.08	17.87
N	%	0.54	3.72
C	%	67.53	30.02
P	%	0.27	7.54

119

### 120.3. Greenhouse Experiment

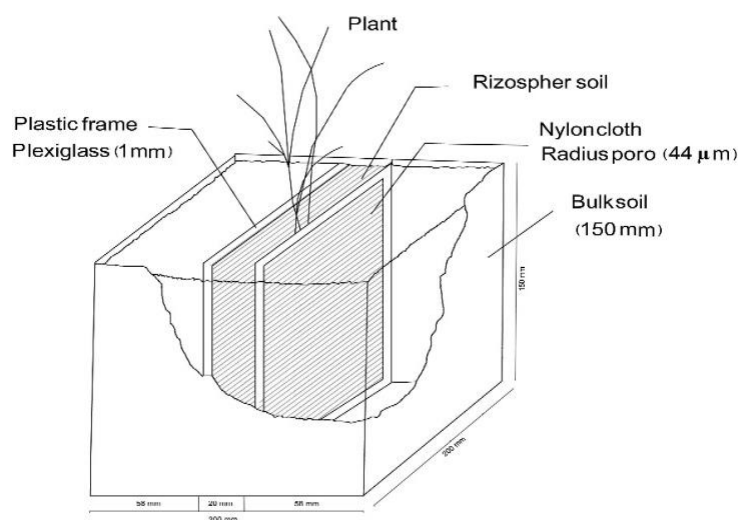
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#### 122.3.1. Rhizobox experiment

123

124 Lab-built rhizoboxes with dimensions of 20 × 15 × 20 cm (length × width × height) were used in  
125 the greenhouse experiments. The inner space of the rhizoboxes was divided into two zones using  
126 325 mesh nylon cloth: (i) a 2 cm thick rhizosphere zone, and (ii) a 5.8 cm thick non-rhizosphere zone  
127 on either side of the rhizosphere zone (Fig. 1). PWB and PWC derived from apple trees and grape vines  
128 was incorporated into the soil so as to provide 1.5% net organic C; each box thus contained 5.80 kg soil  
129 complemented with 41.19 g compost kg<sup>-1</sup> soil (PWC), 22.21 g biochar kg<sup>-1</sup> soil (PWB), or sterilized  
130 inoculated soil (control). In addition, a rock phosphate source providing 80 mg P kg<sup>-1</sup> soil was placed 5  
131 cm below the seeds to act as an insoluble P source.

132 Microbial inoculation was performed using microbial strains from the microbial bank of the Soil  
133 Science Department of Urmia University and included the mycorrhizal fungus *Glomus fasciulatum*.  
134 The amount of mycorrhiza inoculum was 70 g per box and it was uniformly dispersed under the seeds  
135 with a 0.5-cm spacing [27]. After adding the inoculum, seeds of the wheat cultivar 'Pishtaz' (*Triticum*  
136 *aestivum* L.) were disinfected with 0.5% sodium hypochlorite and sown in the rhizosphere zones of the  
137 rhizoboxes. After the seeds germinated, the seedlings were thinned to provide four plants per rhizobox.  
138 During their growth periods, the plants were irrigated with distilled water and fed with P-free Rorison  
139 nutrient solution to satisfy their nutrient requirements [28].  
140



141

142

**Figure 1.** A schematic of rhizobox

143

### 144 2.3.2. Soil analysis

145

146 At the end of the 65-day wheat growth period, two soil samples were collected from the non-  
147 rhizosphere zone (specifically, from the 2 cm furthest from the rhizosphere zone) and combined.  
148 Rhizosphere soil was then sampled by removing the plexiglass framework of the box and then removing  
149 the soil from the box. Main and secondary roots were then carefully removed from the rhizosphere soil  
150 and two samples were collected and combined. Two final samples were thus obtained from each  
151 rhizobox, one representing the rhizosphere zone and the other representing the non-rhizosphere zone.

152 The OC [24], MBC [29], MBP [30], BR [31], SIR [32], ACP and ALP [33] and arbuscular mycorrhizal  
153 fungi colonization percentage [34] were then determined for each sample.

154

### 155 2.3.3. Statistical Analysis

156

157 Statistical analyses including analysis of variance and comparison of data means was performed  
158 using Duncan's Multiple Range Test with a significance threshold of  $p < 0.05$  using the SAS software  
159 package (Version 9.2).

160

## 161 3. Results

162

163 Comparisons of means for the interaction between organic sources and AMF inoculation  
164 ( $p < 0.001$ ) revealed that treatments involving AMF inoculation increased organic C and soil biological  
165 parameters to a much greater degree than treatments without inoculation. Even without the addition of  
166 exogenous organic material, the soil biological parameters after inoculation were substantially higher  
167 than under non-inoculated control conditions. However, in the absence of added organic matter, AMF  
168 inoculation did not significantly increase the soil's organic C content (Table 3). The highest soil organic  
169 C content and biological parameter values were observed for the treatment combining AMF inoculation  
170 with compost addition. However, the organic C content did not differ significantly between the compost  
171 and biochar treatments under non-inoculation conditions. Biochar treatments yielded higher organic C  
172 levels than control treatments under both inoculation and non-inoculation conditions, but the increase  
173 in organic C was lower than that caused by adding compost. A comparison of means for the interactive  
174 effect of organic source and microbial inoculation on MBC and MBP showed that the addition of  
175 organic matter and AMF inoculation had a significant effect ( $p < 0.001$ ) when compared to the control  
176 treatment without AMF inoculation. In the compost treatments, the presence of AMF increased MBC  
177 and MBP levels 1.66- and 5.24-fold, respectively. Similarly, in the biochar treatments, AMF inoculation  
178 increased MBC and MBP levels 1.37- and 1.34-fold, respectively. Under non-inoculation conditions,  
179 adding compost increased MBC and MBP to a greater degree than adding biochar (Table 3). Compost

180 treatment combined with AMF inoculation also increased BR and SIR, and this increase was 1.28 times  
 181 greater in the presence of AMF than without inoculation. Similar trends were observed for biochar  
 182 treatment.

183

184 **Table 3.** Effects of the organic amendment source and microbial inoculation on OC, MBC, MBP, BR  
 185 and SIR

Microbial inoculation	Organic sources	OC (%)	MBC (mg kg <sup>-1</sup> )	MBP (mg kg <sup>-1</sup> )	BR (mg CO <sub>2</sub> d <sup>-1</sup> kg <sup>-1</sup> )	SIR (mg CO <sub>2</sub> d <sup>-1</sup> kg <sup>-1</sup> )
+AMF	PWB	1.64 <sup>b</sup>	789.3 <sup>b</sup>	20.20 <sup>b</sup>	53.15 <sup>b</sup>	100.5 <sup>b</sup>
	PWC	2.0 <sup>a</sup>	957.1 <sup>a</sup>	85.72 <sup>a</sup>	56.17 <sup>a</sup>	112.3 <sup>a</sup>
	Cont.	0.44 <sup>d</sup>	429.0 <sup>d</sup>	7.43 <sup>d</sup>	25.97 <sup>e</sup>	51.99 <sup>e</sup>
-AMF	PWB	0.87 <sup>c</sup>	574.0 <sup>c</sup>	15.01 <sup>c</sup>	34.86 <sup>d</sup>	78.21 <sup>d</sup>
	PWC	0.97 <sup>c</sup>	573.7 <sup>c</sup>	16.33 <sup>c</sup>	43.63 <sup>c</sup>	87.48 <sup>c</sup>
	Cont.	0.32 <sup>d</sup>	264.5 <sup>e</sup>	3.52 <sup>e</sup>	8.61 <sup>f</sup>	15.46 <sup>f</sup>
LSD <sub>0.05</sub>		0.18	7.31	2.41	2.88	4.34

186 Means followed by the same letters are not significantly different according to Duncan's multiple range test at the  
 187  $p < 0.05$  level. +AMF, -AMF, PWB, PWC, and Cont denote inoculation with arbuscular mycorrhizal fungi, without  
 188 micorrhizal inoculation, pruning waste biochar, pruning waste compost, and control (no added organic matter),  
 189 respectively.

190

191 Adding organic matter (especially compost) to the soil caused the organic matter content of both  
 192 the rhizosphere zone and the non-rhizosphere zone to differ significantly from that in control treatments  
 193 ( $p < 0.05$ ; Table 4). The soil organic carbon content was higher in the rhizosphere than in non-  
 194 rhizosphere in all treatments, but the difference was not significant in all cases. The addition of organic  
 195 matter to the rhizosphere and non-rhizosphere zones significantly increased their MBC and MBP  
 196 contents ( $p < 0.01$ ) when compared to controls (Table 4). The highest MBC content was observed when  
 197 rhizosphere soil was amended with compost; under these conditions, the MBC content of rhizosphere  
 198 soil was 1.30% higher than that of non-rhizosphere soil. The MBC of the rhizosphere soil was lowest  
 199 under organic matter-free control conditions; even under these conditions, the MBC of the rhizosphere  
 200 soil was 3.49% higher than that of the non-rhizosphere soil. The highest MBP was observed in the  
 201 rhizosphere zone after treatment with compost, which was 5.55% higher than the MBP in the non-  
 202 rhizosphere. Biochar also improved these biological parameters when compared to the control, albeit  
 203 to a lesser degree than compost; it increased both BR and SIR in the rhizosphere significantly more  
 204 ( $p < 0.001$ ) than in the non-rhizosphere zone (Table 4). The BR and SIR in both the rhizosphere and the  
 205 non-rhizosphere were higher under the organic amendment treatments than under control conditions;



206 the highest BR (5263 mg CO<sub>2</sub> kg<sup>-1</sup> soil) and SIR (103.3 mg CO<sub>2</sub> kg<sup>-1</sup> soil) were observed in rhizosphere  
207 zone after compost treatment.

208

209 **Table 4.** OC, MBC, MBP, BR and SIR in different soil zones with different organic amendments

Soil	Organic sources	OC (%)	MBC (mg kg <sup>-1</sup> )	MBP (mg kg <sup>-1</sup> )	BR (mg CO <sub>2</sub> d <sup>-1</sup> kg <sup>-1</sup> )	SIR (mg CO <sub>2</sub> d <sup>-1</sup> kg <sup>-1</sup> )
Rhizosphere	PWB	1.29 <sup>bc</sup>	687 <sup>c</sup>	18.98 <sup>c</sup>	46.99 <sup>b</sup>	92.61 <sup>b</sup>
	PWC	1.52 <sup>a</sup>	770.3 <sup>a</sup>	52.41 <sup>a</sup>	52.63 <sup>a</sup>	103.3 <sup>a</sup>
	Cont.	0.42 <sup>d</sup>	352.7 <sup>e</sup>	6.71 <sup>e</sup>	20.27 <sup>d</sup>	37.13 <sup>d</sup>
Non-rhizosphere	PWB	1.22 <sup>c</sup>	676.3 <sup>d</sup>	16.23 <sup>d</sup>	41.03 <sup>c</sup>	86.06 <sup>c</sup>
	PWC	1.45 <sup>ab</sup>	760.4 <sup>b</sup>	49.65 <sup>b</sup>	47.16 <sup>b</sup>	96.51 <sup>b</sup>
	Cont.	0.35 <sup>d</sup>	340.8 <sup>f</sup>	4.24 <sup>f</sup>	14.31 <sup>e</sup>	30.33 <sup>e</sup>
LSD <sub>0.05</sub>		0.18	7.31	2.41	2.88	4.34

210 Means followed by the same letters are not significantly different according to Duncan's multiple range test at  
211 the p<0.05 level. PWB, PWC, and Cont. denote pruning waste biochar, pruning waste compost and Control  
212 (without organic matter), respectively.

213

214 AMF inoculation increased organic C in both the rhizosphere and non-rhizosphere zones when  
215 compared to the non-inoculated control treatment (Table 5). However, the organic C contents of the  
216 rhizosphere and non-rhizosphere did not differ significantly ( $p < 0.05$ ) under either inoculation or non-  
217 inoculation conditions. Additionally, the microbial biomass C and P levels in the +AMF treatments  
218 were 1.22 and 3.25 times higher, respectively, than in the -AMF treatments (Table 5). On the other  
219 hand, MBC and MBP were much higher in the rhizosphere than in the non-rhizosphere ( $p < 0.001$ ).  
220 AMF inoculation significantly ( $p < 0.001$ ) increased BR and SIR in both the rhizosphere and non-  
221 rhizosphere zones when compared to controls (Table 5). Additionally, AMF inoculation increased BR  
222 and SIR in the rhizosphere by 13.06 and 7.95%, respectively, relative to the non-rhizosphere.

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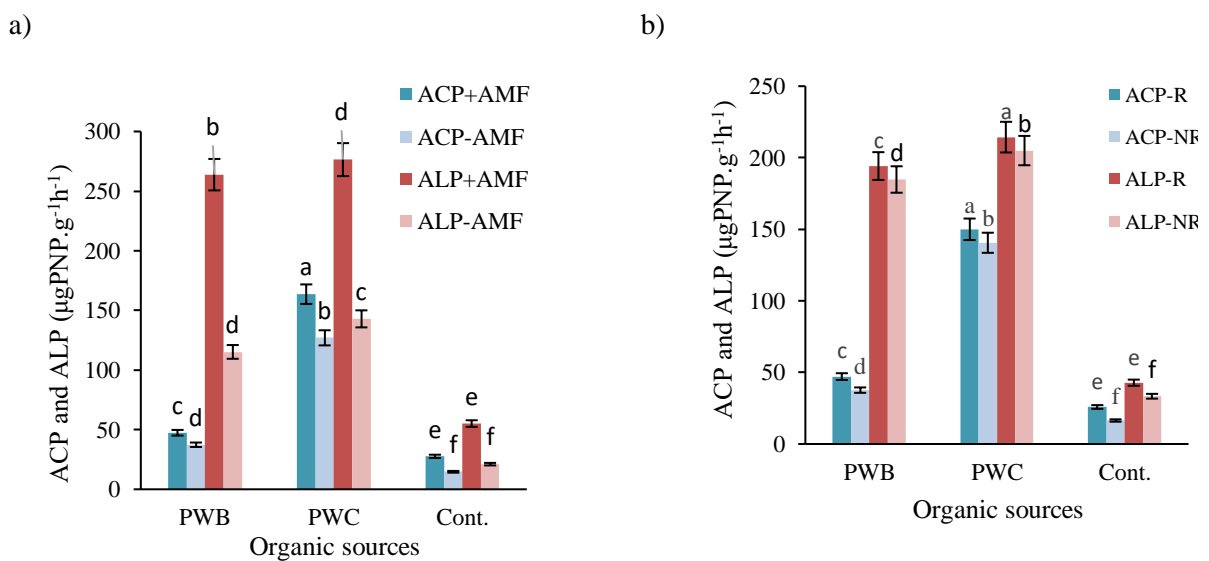
224 **Table 5.** OC, MBC, MBP, BR and SIR in rhizosphere and non-rhizosphere soil with and without microbial  
225 inoculation

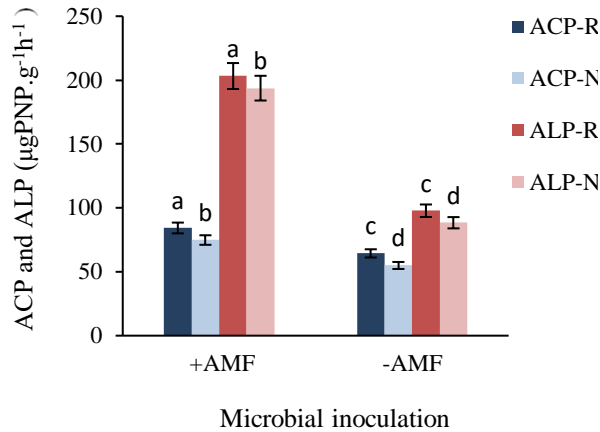
Microbial inoculation	Soil	OC (%)	MBC (mg kg <sup>-1</sup> )	MBP (mg kg <sup>-1</sup> )	BR (mg CO <sub>2</sub> d <sup>-1</sup> kg <sup>-1</sup> )	SIR (mg CO <sub>2</sub> d <sup>-1</sup> kg <sup>-1</sup> )
+AMF	Rhizosphere	1.39 <sup>a</sup>	730.9 <sup>a</sup>	39.08 <sup>a</sup>	47.86 <sup>a</sup>	91.64 <sup>a</sup>
	Non-rhizosphere	1.33 <sup>a</sup>	719.4 <sup>b</sup>	36.49 <sup>b</sup>	42.33 <sup>b</sup>	84.89 <sup>b</sup>
-AMF	Rhizosphere	0.75 <sup>b</sup>	475.7 <sup>c</sup>	12.99 <sup>c</sup>	32.06 <sup>c</sup>	63.73 <sup>c</sup>
	Non-rhizosphere	0.69 <sup>b</sup>	465.6 <sup>d</sup>	10.26 <sup>d</sup>	26.01 <sup>d</sup>	57.04 <sup>d</sup>
LSD <sub>0.05</sub>		0.18	7.31	2.41	2.88	4.34

226 Means followed by the same letters are not significantly different at the  $p < 0.05$  level according to Duncan's test.  
227 +AMF and -AMF denote inoculation with arbuscular mycorrhizal fungi and without mycorrhiza, respectively.

228

229 The highest activity of phosphatase enzymes was measured in compost applied inoculated with AMF.  
230 AMF inoculation increased ACP and ALP enzymes by 5.92 and 5.01 times compared to non-inoculated  
231 control, respectively. The lowest activity of phosphatase enzymes was observed in non-inoculated  
232 control that was not amended with organic matter. Among non-inoculated organic treatments, compost  
233 showed higher activity of these enzymes than biochar, although biochar enhanced these enzymes'  
234 activity compared to control. Also, its effect under AMF inoculation on the increased activity of the  
235 enzymes was in the second rank of magnitude after compost (Figure 2a). Organic treatments increased  
236 the activity of phosphatase enzymes in both rhizosphere and non-rhizosphere when compared to control  
237 (Figure 2b). According to the results, when PWC was applied, the activity of ACP and ALP enzyme  
238 was 1.06 and 1.04 times as great in the rhizosphere as in the non-rhizosphere region, and both regions  
239 showed significant differences when they were treated with compost than treated with biochar and  
240 control (Figure 2b). According to Figure 2b, it is observed that in all treatments, the activity of the ALP  
241 enzyme is greater than that of ACP. In control treatments, too, compost was more effective than biochar  
242 in prospering enzymatic activity. AMF inoculation increased the activity of phosphatase enzymes so  
243 that the activity of ACP and ALP enzymes under inoculation conditions was 1.30 and 2.07 times as  
244 great in the rhizosphere compared to non-rhizosphere region (Figure 2c).  
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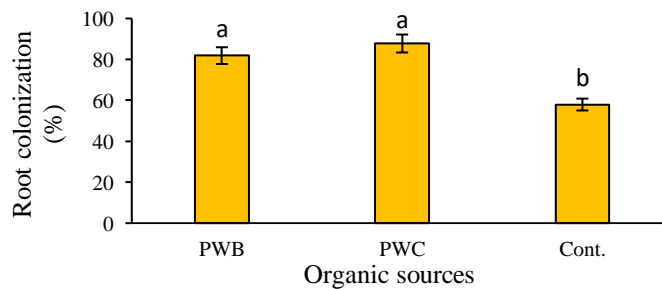


**Figure 2.** Means comparison of a) organic sources and microbial inoculation (+AMF, -AMF) on the soil ACP and ALP b) organic sources on the soil ACP and ALP, and c) microbial inoculation on the soil ACP and ALP. PWB, PWC, Cont., ACP, ALP, +AMF, -AMF, R and NR are pruning waste biochar, pruning waste compost, control (no organic mater), acid and alkaline phosphatase enzymes, inoculation with arbuscular mycorrhizal fungi, without mycorrhiza, rhizosphere and non-rhizosphere regions, respectively.

246

247 Finally, analysis of variance revealed that organic amendment and AMF significantly increased root  
248 mycorrhizal colonization ( $p < 0.001$ ) when compared to controls (Fig. 3). Colonization was highest  
249 under the PWC and PWB treatments (51.45 and 41.24% higher than controls, respectively).

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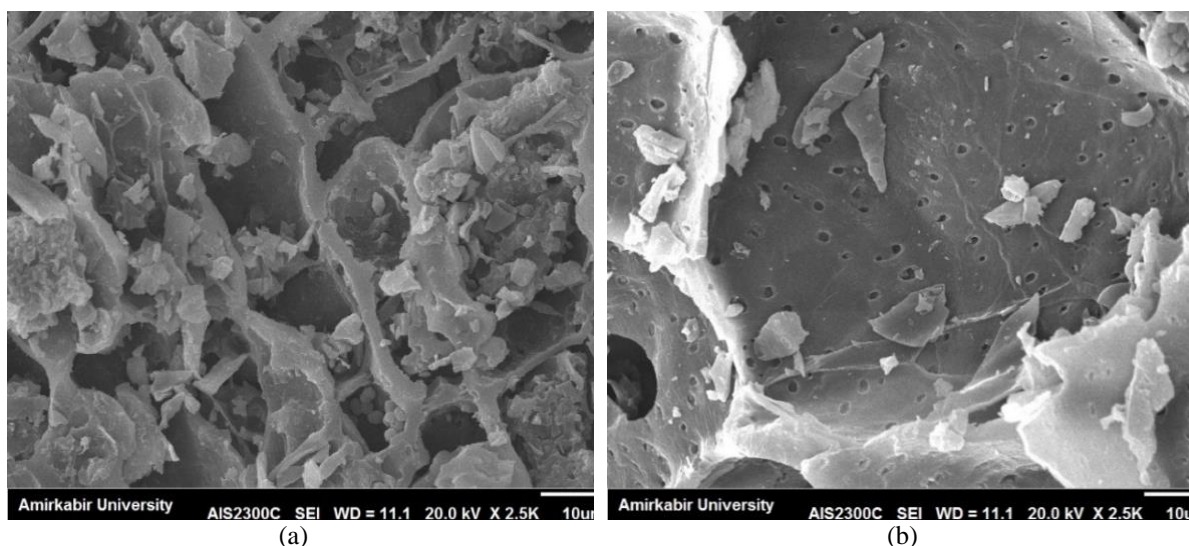
252 **Figure 3.** The effect of the organic source and AMF on root colonization (%) PWB, PWC and Cont. denote  
253 pruning waste biochar, pruning waste compost, and control (no organic mater), respectively.

254

#### 255 4. Discussion

256 Biochar and compost amendment both increased the soil's organic C content, but when organic  
257 amendment was combined with AMF inoculation, amendment with compost increased organic C more  
258 than amendment with biochar. This may be due to the stable carbon skeleton of biochar, which is likely  
259 to resist microbial decomposition and may thus enable a slow gradual release of organic C into the soil.

260 For example, there is evidence that the lifespan of biochar carbon in soil is between 100 and 1000 years,  
261 which is 10-1000 times longer than that of soil organic carbon, making it a valuable long-term carbon  
262 source [35]. The increase in MBC following organic amendment may be due to the introduction of  
263 beneficial substrates for microorganisms that stimulate their activity and increase soil biological activity  
264 [36]. Accordingly, Babalol et al. reported that applying plant residue compost to soil increased its  
265 population of microorganisms, leading to an increase in MBP [37]. Amendment with wood biochar can  
266 also increase soil microbial activity both by providing a favorable habitat for microorganisms and by  
267 serving as a source of moisture, carbon, labile resources, and nutrients. Fig. 4 presents scanning electron  
268 microscopy (SEM) images of the apple and grape biochars used in this work, revealing that they consist  
269 of irregularly shaped porous particles. The porosity of biochar gives it a high specific area, enabling the  
270 adsorption of large quantities of dissolved organic matter, gases, and minerals. Additionally, the  
271 diameter of the pores in wood-derived biochars ranges from 2 to 80  $\mu\text{m}$ ; this size range is adequate to  
272 support mycorrhizal fungi [38]. Consequently, biochar provides a very favorable habitat for  
273 microorganisms, especially AMF; its pores provide shelter against predators and protection from  
274 drought while also hosting resources that help satisfy their carbon, energy, and nutrient requirements.  
275 Consequently, biochar amendment increases the biological activity of mycorrhizal fungi [13]. Jin  
276 reported that the interaction of biochar with AMF increased the population of mycorrhizal fungi and  
277 the MBC content of rhizospheric soil to a greater degree than in non-rhizosphere soil [39]. The increase  
278 in MBC was more pronounced among mycorrhizal fungi than bacteria, which was attributed to more  
279 efficient use of resources by the fungi and more effective carbon assimilation due to the extensive  
280 hyphal network of the fungi. Importantly, fungi can establish hyphal bridges between biochar and plant  
281 roots.  
282



283 **Figure 4.** Scanning electron microscope (SEM) imagery of biochar of (a) apple pruning waste and (b) grape  
284 pruning waste at pyrolysis temperature of 350°C  
285

286 Amendment with compost increases the organic C content of the soil as well as both plant growth  
287 and microbial activity. This normally leads to increased respiration (both BR and SIR). Compost  
288 amendment induced a greater increase in both rates of respiration than amendment with biochar, which  
289 may be related to its greater ease of decomposition. Obviously, the availability of adequate nutrients is  
290 a key determinant of the activity and growth of microorganisms. The effect of biochar on respiration  
291 differs from that of compost due to its structure and long half-life; Liu et al. found that biochars did not  
292 significantly influence soil respiration [11], whereas Steiner et al. observed increased microbial activity  
293 and growth following biochar amendment [40]. In this work, despite an increase in the microbial  
294 population following the addition of glucose to the biochar-treated soil, the rate of soil respiration (SIR)  
295 was unchanged. These results imply that the application of nutrient-rich but slowly decomposing  
296 biochars can support microbial population growth. Biochars also had some positive impacts on soil  
297 biological characteristics; the formation of biochars from wood at low pyrolysis temperatures leads to  
298 the retention of diverse compounds that promote microbial growth (e.g., sugars and aldehydes) on the  
299 surface of the biochar [41]. The short-term increase in microbial activity and population as well as soil  
300 respiration following biochar amendment is probably related to these labile constituents of freshly  
301 prepared biochars [42].

302 In general, the rhizosphere is the main zone in which organic compounds accumulate and  
303 decompose. It is thus unsurprising that it contains more organic matter than the non-rhizosphere. It

304 seems that the active carbon of compost is partially decomposed after the compost is applied and that  
305 some of this carbon is added to the soil's carbon reserve, increasing the organic carbon content of the  
306 soil. Biochar amendment also increased the soil's organic carbon content, but to a lesser degree than  
307 treatment with compost. Zaman et al. reported that the application of urban waste compost increased  
308 soil MBC by enhancing the soil's content of N, C, and soluble organic C [43]. This is consistent with  
309 our finding that the soil's organic carbon content (Table 4) and total absorbed nitrogen (Table 2) were  
310 higher after compost amendment than after treatment with other organic materials. Additionally, De  
311 Neergaard and Magid found that the application of organic matter improved the MBC content in the  
312 rhizosphere of rye to a greater degree than that of the non-rhizosphere zone [44], and a study on the  
313 response of the soil microbial community to biochar derived from wheat and willow trees, Watzinger  
314 et al. indicated that biochar application increased microbial biomass and changed the microbial  
315 community structure [45]. MBP can be either a vital sink for soluble P when microbes compete with  
316 plants for P or a crucial source of P that helps partially meet plants' P requirements. Soil microbes can  
317 thus help maintain stable levels of labile P in soils with low P availability. Redel et al. incorporated  
318 compost into soil and observed a positive association between MBP content and Olsen P (determined  
319 by extraction with sodium bicarbonate) [46]. Similarly, in this work, compost amendment increased  
320 available P in the soil more than any other treatment (Table 2). Biochar amendment has also been  
321 reported to increase MBP, possibly due to its high C content and positive effect on nutrient availability.  
322 Compost amendment strongly increased the microbial respiration rate because compost is rich in  
323 oxygenated organic compounds such as carbohydrates that are more readily degraded by microbes than  
324 aromatic carbon and alkyl carbon chains [47]. However, an earlier study on the effect of biochar on BR  
325 under rhizobox conditions [48] found that biochar increased BR, leading to a significantly higher rate  
326 of microbial respiration in the rhizosphere than in the non-rhizosphere zone.

327 Microorganisms can be regarded as active organic particles in the soil with charged and catalytically  
328 active surfaces and the ability to produce and exude a wide range of organic compounds including  
329 carbohydrates, and enzymes [49]. A soil's MBC content is widely interpreted as a measure of its content  
330 of soil microbes and their activity. However, it does not distinguish between living and dead microbes;  
331 it is likely that AMF cells are included in the soil microbial biomass when their life cycle ends. The

332 increase in MBC following AMF inoculation may thus be partly due to the addition of dead cells to the  
333 rhizosphere soil, as suggested in earlier studies on the maize rhizosphere [50]. The beneficial effects of  
334 the rhizosphere on the soil biological characteristics have also been highlighted by Zhao et al. [51]. In  
335 addition to the increases in rhizospheric MBC due to organic matter amendment and microbial  
336 inoculation, plants release organic compounds including sugars, amino acids, and vitamins into the  
337 rhizosphere, further reinforcing its differences from the non-rhizosphere zone. These root exudations  
338 also increase microbial biomass by making the rhizosphere more hospitable to microbes, which in turn  
339 increases plant growth and the production of microbial metabolites [52]. Consequently, the rhizosphere  
340 is characterized by elevated microbial activity and has a higher content of organic matter and nutrients  
341 than the rest of the soil, allowing it to play a key role in meeting plants' nutrient requirements to support  
342 photosynthesis while also supporting a diverse microbial community leading to high levels of soil  
343 biological activity (and thus high levels of BR, SIR, MBC, and MBP) [53]. Organic matter application  
344 can boost microbial activity to improve enzymatic activities via three ways: (i) the application of organic  
345 matter to soil enhances soil microbial activity by microorganism-based energy pathway, resulting in  
346 enzyme synthesis; (ii) the enzymes accompanying organic matter reduce energy for the activity of  
347 microorganisms of soil for the synthesis of the enzymes, thereby increasing their activity and the rate  
348 of organic matter decomposition in soil; (iii) organic matter is partially decomposed in soil and then, it  
349 protects the enzymes against enzymatic hydrolysis by adsorbing and physically grasping the enzymes  
350 [54]. The enhancement of the activity of soil phosphatase enzymes under the treatments with PWC and  
351 PWB may be associated with the increased level of microbial biomass in response to the applied organic  
352 matter, soil nutrients, and the improvement of soil (physical, chemical and biological) characteristics,  
353 like nutrient and water holding capacity. Also, the higher performance of PWC in enhancing the activity  
354 of phosphatase enzymes as compared to PWB is likely to reflect its higher rate of decomposition and  
355 the availability of its N content, which stimulates microbial activity. The decline of ACP enzyme  
356 activity versus ALP under the application of organic matter, especially biochar, was not surprising  
357 because the studied soil, which had alkaline pH and calcareous conditions (Table 1), and the applied  
358 biochar, which had alkaline pH (Table 2), were not suitable for the activity of this enzyme. Jin et al.  
359 reported that the activity of ACP enzyme was decreased after the application of manure biochar,

360 whereas the activity of ALP enzyme was increased [55]. The loss of ACP enzyme activity was attributed  
361 to the increase in soil pH by biochar application. Fungi need extra-cellular enzymes, such as  
362 phosphatase, to decompose the substrate of their surrounding medium into smaller molecules and to  
363 mobilize these molecules into the cells and also to use them in their metabolic activities. Thus, the  
364 medium around a fungus is studied with the exudation of extra-cellular enzymes. Since extra-cellular  
365 enzymes decompose promptly, therefore they should be adsorbed to soil particles to maintain their  
366 activities. In this respect, specific areas are of particular importance. These areas can be the surface of  
367 soil particulates, plant root, or organic matter like compost or biochar. The activity of extra-cellular  
368 enzymes depends on the site of the enzyme that is interacting with the surface of biochar particles [56].  
369 If the enzyme's active site is not covered and is operational and free to react with environment, its  
370 activity will be augmented. Still, if it is blocked, its activity will decline [57]. Therefore, some groups  
371 of enzymes become more active by the application of biochar and the other groups lose their activity  
372 depending on molecular composition, folding properties, and their adsorption potential [58]. The  
373 augmented activity of phosphatase in the rhizosphere versus non-rhizosphere can be associated with the  
374 microbial and root activity of the plant. Similarly, in a study on the effect of organic fertilizer on the  
375 activities of phosphatase enzymes in the rhizosphere of plants using rhizobox, Balik et al. reported that  
376 the action of ACP and ALP enzymes were significantly increased in bulk soil [59].

377 The rate by which microorganisms can synthesize and release phosphatase varies with soil pH. As  
378 pH is increased, ALP becomes more stable and active so that its activity is maximized at pH = 11. In  
379 addition, ACP is mostly exuded by fungi and the optimal pH for it is acidic to neutral conditions. Still,  
380 bacteria predominantly exude ALP and are more active at pH > [60]. Furthermore, ACP is produced  
381 mainly by plants and microorganisms, but microorganisms predominantly exude ALP.

382 Organic matter amendment also improves the physical properties of the soil, making it better able  
383 to support the development and activity of microorganisms and various bioprocesses [61]. The  
384 improvements in soil biochemical properties and nutrient availability induced by PWC or PWB  
385 amendment were reinforced by inoculation with AMF, and also enhanced root mycorrhizal  
386 colonization. Similarly, Mäder et al. reported that the application of compost to soil increased  
387 mycorrhizal colonization by 30-60% in the wheat rhizosphere [62]. Amendment with hardwood biochar



388 (2% w/w) similarly increased root colonization, which was attributed to increased growth of the external  
389 hyphae of arbuscular mycorrhiza [17]. Biochar can influence the symbiosis of arbuscular mycorrhiza  
390 and plant roots by (i) changing the availability of nutrients affecting the relationships between plants  
391 and AMF and/or their physicochemical properties, (ii) stimulating microbial activity and population  
392 growth by strengthening mycorrhizal symbiosis, (iii) disturbing chemical signals or detoxifying  
393 chemicals that inhibit the activity of mycorrhiza fungi, and (iv) providing physical shelter for  
394 mycorrhiza fungi against adverse conditions.

395

## 396 **5. Conclusion**

397

398 The results presented herein show that combining organic matter amendment with microbial  
399 inoculation can have significant positive effects on soil quality parameters. Adding organic matter such  
400 as compost or biochar to the soil and inoculation with AMF in the root zone strongly increased soil  
401 biological activity, with particularly strong increases in soil quality parameters (BR, SIR, MBC, and  
402 MBP) occurring in the rhizosphere. Combined compost amendment and AMF inoculation led to the  
403 highest OC, MBC, and MBP values, but the effect of biochar amendment on root mycorrhizal  
404 colonization was similar to that of adding compost. In general, amendment with compost had a stronger  
405 positive effect on the studied soil biological parameters than treatment with biochar, which may be due  
406 to the structure and greater stability of biochar. It should be noted that not all soil quality parameters  
407 respond in the same way to microbial inoculation and organic matter amendment, and that the efficiency  
408 of plant responses to these treatments will depend on the experimental conditions including the plant  
409 type and species, the experiment type (pot or field study), the choice of organic matter, and the nutrient  
410 ratios. Consequently, it will be necessary to confirm the results presented herein by studying the  
411 combined effects of organic amendment and microbial inoculation in greenhouse and field studies on  
412 various plant species and to assess the economic viability of organic amendment, especially with  
413 biochar. Nevertheless, the results presented herein show that combining organic matter amendment with  
414 AMF inoculation can strongly increase the quality, biological activity, and biodiversity of calcareous  
415 alkaline and may represent a very promising alternative to the costly application of chemical fertilizers.

416

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