| 1 | Microbiome assembly predictably shapes diversity across a range of disturbance |
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16 Abstract

17 Diversity is frequently linked to the functional stability of ecological communities. However, its 18 association with assembly mechanisms remains largely unknown, particularly under fluctuating 19 disturbances. Here, we subjected complex bacterial communities in bioreactor microcosms to 20 different frequencies of organic loading shocks, tracking temporal dynamics in their assembly, 21 structure and function. Null modelling revealed a stronger role of stochasticity at intermediate 22 disturbance frequencies, preceding the formation of a peak in α -diversity. Communities at extreme 23 ends of the disturbance range had the lowest α -diversity and highest within-treatment similarity in 24 terms of β -diversity, with stronger deterministic assembly. Stochasticity prevailed during the initial 25 successional stages, coinciding with better specialized function (nitrogen removal). In contrast, 26 general functions (carbon removal and microbial aggregate settleability) benefited from stronger 27 deterministic processes. We showed that changes in assembly processes predictably precede changes 28 in diversity under a gradient of disturbance frequencies, advancing our understanding of the 29 mechanisms behind disturbance-diversity-function relationships.

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Key words: intermediate stochasticity hypothesis, ISH, intermediate disturbance hypothesis, IDH,
 succession, diversity, disturbance, community structure, community function, stochastic assembly,
 deterministic assembly.

34 Introduction

Microbes exist typically as diverse, complex and dynamic communities¹, responsible for all 35 biogeochemical cycles worldwide². These microbial communities or microbiomes provide crucial 36 37 functions for global climate regulation, human health, biotechnology and bioremediation³. Microbial diversity is often related to community function⁴ and the ability to withstand environmental 38 fluctuations that typically occur as disturbances⁵. Given the growing human population and its impact 39 on natural and engineered ecosystems⁶, management and conservation practices are faced with 40 41 increasing frequencies and magnitudes of various disturbances that occur on different scales. A 42 concept of ecology that can be used to explore possible outcomes is the intermediate disturbance 43 hypothesis (IDH), which predicts a diversity peak at intermediate levels of disturbance due to 44 competition-colonization trade-offs faced by organisms⁷. Although the IDH has been influential in $ecology^8$ and ecosystem conservation^{9,10}, it is not a coexistence mechanism as initially thought¹¹. 45 Further, many studies have not found the diversity pattern predicted by the IDH^{12,13} and its relevance 46 as a prediction tool is up for debate^{14,15}. Therefore, studies are needed to address the mechanisms 47 48 behind the observed disturbance-diversity relationships¹⁶.

49 Intermediate frequencies of exposure to a xenobiotic pollutant in our recent replicated sludge 50 bioreactor study demonstrated higher α -diversity and relative influence of stochastic assembly compared to other exposure levels, after a short succession period of 35 days¹⁷. We hypothesized that 51 52 when intermediate disturbance levels result in unpredictable environments where specialized traits are 53 less advantageous to taxa, the stochastic equalization of competitive advantages would lead to a 54 higher α -diversity, a causal relationship we named the intermediate stochasticity hypothesis (ISH)¹⁷. 55 In contrast, either no disturbance or press disturbance conditions at the extreme ends of a disturbance 56 range would allow fewer adapted organisms to dominate, thus lowering the α -diversity. Unlike the 57 IDH, the ISH incorporates the role of assembly mechanisms as shapers of community structure (α -58 and β - diversity) across a disturbance gradient. Further, it predicts patterns not only in species 59 richness but also in higher-order α -diversity indices, since variations in the underlying assembly 60 mechanisms would affect abundance distributions of taxa. The ISH also considers that the output of a

61 stochastic process is affected by some uncertainty, which means that there are several possible paths 62 for the evolution of the structure and function of a community. In this regard, stochasticity operating 63 at intermediate levels of disturbance in replicated systems could lead to similar high α -diversity 64 (local, e.g., within a reactor), but not necessarily to similar β -diversity (compositional variation across sites, e.g., between reactors) and community function¹⁷. The idea of community assembly processes 65 66 underlying the observed patterns of diversity is reasonable, as such processes are believed to shape community structure¹⁸, which also links them to ecosystem function. These processes, either 67 deterministic or stochastic, are known to act in combination to form community assembly¹⁹⁻²² and can 68 cause replicate communities to reach a similar or variable structure and function^{17,23}. Further, while 69 recent studies have reported positive correlations of strength of stochasticity with α -diversity in 70 bacterial²⁴ and fungal²⁵ communities, the role of assembly processes behind diversity patterns under 71 72 fluctuating disturbances is still unclear.

73 The objective of this work was to test the central tenet of the ISH that intermediate 74 disturbance frequencies promote stochastic assembly processes, resulting in increased α -diversity and variable β -diversity¹⁷. We used an experimental system comprised of activated sludge sequencing 75 76 batch reactors harboring complex microbial communities collected from a full-scale wastewater 77 treatment plant. These were subjected to different frequencies of organic loading shocks, tracking temporal dynamics in their overall assembly, structure and function, without focusing on any 78 particular taxa. The reactors had a working volume of 25 mL, representing a microcosm scale²⁶. 79 Replicates (n = 5) received double organic loading either never (L0, undisturbed), every 8, 6, 4 or 2 80 81 days (L1-4, intermediately-disturbed), or every day (L5, press-disturbed), for 42 days. Samples were 82 analyzed using 16S rRNA gene metabarcoding and effluent chemical characterization. Patterns of α -83 and β -diversity were employed to assess temporal dynamics of bacterial community structure. 84 Assembly mechanisms were quantified via null model analysis of phylogenetic turnover for each 85 bioreactor.

86 Results

87 Intermediate disturbance frequencies exhibit higher taxonomic and phylogenetic α-diversity

Taxonomic α -diversity was evaluated using Hill diversity indices²⁷ of orders zero (⁰D, taxa 88 richness), one (¹D) and two (²D), the latter being a robust estimate of microbial diversity¹⁷. 89 Phylogenetic α -diversity was also considered through Faith's phylogenetic distance²⁸ unweighted 90 91 (PD) and abundance-weighted (PD_w). There was a temporal decrease in α -diversity for all 92 disturbance frequency levels compared to the sludge inoculum for both taxonomic and phylogenetic 93 α -diversity indices (Fig. 1A, Fig. S1). This drop was more pronounced, particularly within the first 14 94 days, when variability across same-level replicates was also highest. From d14 onwards, disturbance frequency had a significant effect on α -diversity (²D, Welch's ANOVA P_{adj} = <0.001-0.015) (Fig. 95 96 1A). A peak in α -diversity at intermediate frequencies of disturbance was observed for all unweighted 97 (⁰D, PD) and abundance-weighted (¹D, ²D, PD_w) indices evaluated in this study (Fig. 1A, Fig. S2, Fig. S3). Such parabolic pattern was significant from d21 onwards for ²D (Welch's ANOVA $P_{adj} \le 0.003$), 98 99 from d28 onwards for ¹D (Welch's ANOVA $P_{adj} = 0.002-0.01$), PD (Welch's ANOVA $P_{adj} = 0.003-0.01$) 0.037) and PD_W (Welch's ANOVA P_{adj} = 0.005-0.013), and from d35 onwards for ^{0}D (Welch's 100 101 ANOVA $P_{adj} = 0.03 - 0.035$).

102 Community assembly temporal dynamics precede α -diversity patterns across disturbance frequencies

103 Assembly processes were first evaluated by modelling the phylogenetic dispersion of a given community against the null expectation, through the nearest taxon index (NTI)²⁹. We observed higher 104 105 stochasticity at the initial stages of the experiment (d0-14), which decreased in relative intensity over 106 time across disturbance levels for both unweighted (NTI) and abundance-weighted (NTI_w) indices 107 (Fig. 1A, Fig. S2). There was a stronger role of stochastic assembly processes at intermediate 108 disturbance frequencies as shown by NTI values closer to zero (i.e., lower |NTI| values); this was significant from d14 onward (NTI Welch's ANOVA $P_{adj} = \langle 0.001 - 0.037 \rangle$ but was reduced towards 109 110 the end of the study becoming non-significant on d42. Games-Howell post-hoc grouping indicated 111 that the parabolic pattern of NTI across disturbance frequency levels preceded (d14-35) the formation

112 of a peak in α -diversity (d21-42) at intermediate levels of disturbance, with two to three groups 113 significantly differentiated (Fig. 1A). Stochastic assembly processes were less prevalent when 114 abundance weighing was included in the calculation of the NTI index (NTI_w) , meaning that 115 phylogenetic dispersion compared to the null expectation was higher among individual organisms 116 than it was among taxa. Nonetheless, there was a significant peak in NTI_w values at intermediate 117 frequencies of disturbance on d7 and d14 (NTI_W Welch's ANOVA $P_{adj} = 0.001$). This parabolic 118 pattern of NTI_w was evident on d7, preceding that of NTI, but disappeared on d21 and reverted from 119 d28 onwards. Also, significant phylogenetic signals were observed via mantel correlogram analysis 120 (Fig. S5) mostly across relatively short phylogenetic distances, justifying the use of phylogenetic null 121 modelling to evaluate community assembly processes in this study.

122 Stochastic assembly was more important when α -diversity was higher, particularly for phylogenetic diversity. This was shown by significant Kendall correlation τ values (0.24-0.46, P_{adj} < 123 124 0.001) between NTI and α -diversity indices (Figs. 1B-C, Fig. S4). Kendall correlation τ values were 125 also positive (0.20-0.26) and significant ($P_{adj} < 0.001$) between NTI_W and phylogenetic α -diversity 126 indices (PD, PD_w) and unweighted taxonomic α -diversity (⁰D), but not between NTI_w and 127 abundance-weighted taxonomic α -diversity (¹D, ²D) (Fig. S4). The estimation of all the 128 aforementioned indices over time using rarefied ASV sequencing data yielded the same significant 129 patterns via Welch's ANOVA, with the exception of NTI_w on d21 and d42 (see supplementary file).

β-diversity patterns display similarity at low and high disturbance frequencies and higher variability
at intermediate ones

132 Community structure in terms of β -diversity showed temporal changes, which varied across 133 disturbance levels for both Unifrac phylogenetic distances (Fig. 2A) and Bray-Curtis taxonomic 134 distances (Fig. 2B). Unconstrained ordination displayed a dispersion effect in overall community 135 structure over time, particularly after 7 days, with communities in each reactor diverting from each 136 other (Fig. 2A). To disentangle the effect of disturbance from temporal dynamics, constrained 137 ordination via canonical analysis of principal coordinates (CAP) was used at each time point (Fig.

138 2B). Group-average cluster similarity (60%) was included to detect formations of clusters of 139 community structure. Differences in β -diversity across disturbance levels were statistically significant 140 at all time points evaluated (PERMANOVA $P_{adj} < 0.001$), without significant effects of 141 heteroscedasticity (PERMDISP $P_{adj} > 0.14$) (Table S1). Replicate reactors at the undisturbed (L0) and 142 press-disturbed level (L5) clustered separately from intermediate disturbance levels on all sampling 143 days (except on d7 and d21 for L0) (Fig. 2B), both levels having 0% misclassification error at all time 144 points assessed (Fig. 2C). Comparatively, reactors at intermediate disturbance frequencies (L1-4) 145 clustered together and showed higher dispersion across replicates within the same level, with CAP 146 misclassification errors above zero (Fig. 2B-C). Thus, replicate reactors were less similar to each 147 other at intermediate levels of disturbance, while replicates at low (undisturbed) and high (press-148 disturbed) disturbance frequencies were more similar. Likewise, community assembly assessed via the beta nearest taxon index $(\beta NTI)^{30}$ showed a higher relative contribution of stochasticity at 149 150 intermediate levels of disturbance (Fig. 2D), with β NTI values closer to zero, indicating that 151 phylogenetic turnover across within-treatment replicates was closer to the null expectation. Similarly 152 to what we observed through the NTI, the relative importance of stochasticity decreased with time in 153 the experiment (*i.e.*, higher $|\beta NTI|$ values) and when abundance weighing was included in the 154 calculation of the β NTI values (β NTI_w) (Fig. S6). The observed temporal changes in bacterial 155 community structure across disturbance frequencies were consistent with phylum-level dynamics in 156 relative abundances (Fig. S7), although the focus of this study was on overall community dynamics 157 and not on any particular group of taxa.

158 Community function dynamics and correlations with community structure and assembly

Bacterial community function was assessed over time via influent chemical oxygen demand (COD) removal, sludge volume index (SVI), and influent total Kjeldahl nitrogen (TKN) removal, as measure of carbon removal, sludge settleability and nitrogen removal, respectively (Fig. 3A). Carbon removal and sludge settleability, which are functions associated with a broad range of taxa (*i.e.*, general functions), improved over time during the experiment. High carbon removal (> 0.97) was achieved at all disturbance frequency levels from d21 onwards, with no significant differences on

165 days 35 and 42, after a period of high variability for same-level replicates during the first 14 days. 166 Sludge settleability increased with disturbance frequency, with undisturbed (L0) reactors showing the 167 lowest settleability from d21 onwards and intermediately disturbed levels reaching the highest 168 settleability on d42 (SVI Welch's ANOVA $P_{adj} = 0.018$). The nitrogen removal function (TKN 169 removal), which is related to specialized bacteria (ammonia oxidizers), significantly differed across 170 disturbance frequencies (TKN removal Welch's ANOVA P_{adj} < 0.001) with the highest removal at 171 intermediately disturbed levels during the first 21 days. From d28 onwards, L0 to L4 reactors had 172 similarly high average nitrogen removal (> 0.9), and only the press disturbed reactors (L5) continued 173 to have lower nitrogen removal (< 0.7) than that of the initial sludge inoculum (0.8). Effluent values 174 of TKN, ammonia, nitrite and nitrate showed that TKN removal occurred via nitrification (Fig. S8).

175 Carbon removal had an overall significant negative Kendall correlation with α -diversity 176 indices (τ < -0.21, P_{adj} < 0.001), whereas sludge settleability and nitrogen removal showed non-177 significant correlations with α -diversity across the study (Fig. S9). Correlations between general 178 functions of carbon removal and sludge settleability and both NTI and NTI_w were negative and 179 significant across all time points and disturbance frequencies of the study (Fig. 3B-C, Fig. S9), 180 indicating improved performance of these functions under stronger deterministic assembly 181 mechanisms. Nitrogen removal had a non-significant overall correlation with NTI and NTI_{W} (Fig. 3D, 182 Fig. S9), which became positive and significant when only the first 21 days of the study were 183 considered (NTI $\tau_{d0-21} = 0.39$, $P_{adj} < 0.001$, Fig. 3D; NTI_W $\tau_{d0-21} = 0.46$, $P_{adj} < 0.001$, Fig. S10), 184 suggesting better performance of this function under higher stochastic assembly throughout the initial 185 weeks of the study.

186 Discussion

In this study we found stochastic assembly processes to be more important at intermediate disturbance frequencies where the highest α -diversity was also observed, together with high β diversity dispersion across within-treatment replicates as predicted by the ISH¹⁷. Furthermore, we showed that a peak in the relative contribution of stochasticity preceded a peak in α -diversity across a disturbance frequency range. Also, we observed that higher stochasticity during initial successional

192 stages correlated with better nitrogen removal (specialized function) at intermediate disturbance 193 frequencies, while carbon removal and microbial aggregate settleability (general functions) improved 194 in step with more deterministic forces. These findings highlight the utility of the ISH for a 195 mechanistic understanding of disturbance-diversity-function relationships.

196 We expanded our earlier work¹⁷ by using a different type of disturbance and microbial community inoculum, a relevant scenario given the multidimensional nature of disturbance³¹. 197 198 Employing taxonomic and phylogenetic diversity metrics, in both unweighted and abundance-199 weighted forms, allowed us to cover a broader aspect of α -diversity. Taxonomic resolution was also 200 improved by the use of amplicon sequence variants (ASVs) compared to operational taxonomic unit (OTU) clustering³² with about one to two orders of magnitude fewer spurious units³³, allowing for a 201 202 better estimation of unweighted α -diversity (*i.e.*, taxa richness). We further verified that the observed patterns occurred independently of data rarefaction, given the lack of consensus about this practice³⁴ 203 and the fact that it is known to affect (mainly unweighted) estimations of α -diversity³⁵. Assembly 204 205 processes were tracked over time using a phylogenetic null modelling methodology, which has been tested and recommended in microbial ecology^{20,30,36}. Additionally, general and specific functions were 206 207 evaluated against structure and assembly. All the aforementioned enhancements allowed us to test the 208 ISH, while also gaining new insights into the role of assembly processes behind disturbance-induced 209 changes in community structure and function over time.

210 Our experimental system produced a succession scenario in which bacterial communities had to 211 adapt to change from a full-scale system to a bioreactor microcosm setup along a disturbance frequency gradient, similarly to what we described in a prior study³⁷. With regards to community 212 213 structure, succession led to a significant hump-backed pattern of α -diversity for all composition- and abundance-based indices employed in the study, which occurred after 21 days for ²D, 28 days for ¹D, 214 215 PD and PD_w, and 35 days for ⁰D. Thus, the observed dynamics in community structure were stronger in terms of relative abundances than richness (²D, ¹D vs. ⁰D), as well as at the phylogenetic versus 216 217 taxonomic level (PD vs. ⁰D). The appearance of higher phylogenetic α -diversity at intermediate levels 218 of disturbance for both unweighted (PD) and abundance-weighed (PD_w) indices suggests that

considering evolutionary relationships among organisms³⁸ could also aid in assessing the effect of 219 220 varying disturbances on community structure under succession. In our study, disturbance promoted 221 the co-occurrence of phylogenetically distinct organisms, suggesting that additional niches were 222 created at intermediate disturbance frequencies that were occupied by ecologically different species, 223 thus reducing competitive exclusion. Conversely, phylogenetic clustering at undisturbed and press-224 disturbed levels can be interpreted as communities structured by environmental filtering²⁹. 225 Additionally, temporal analysis of community structure in terms of β -diversity revealed three different 226 clusters for undisturbed, press-disturbed and intermediately disturbed reactors. Further comparison of 227 replicates within the same disturbance frequency level showed higher β -diversity variability at 228 intermediate disturbance levels, which was coherent with prior observations in freshwater ponds³⁹ and 229 sludge bioreactors¹⁷ where β -diversity increased with stochastic assembly. Our findings are relevant 230 for understanding disturbance-diversity relationships, since few studies have reported parabolic α -231 diversity patterns using abundance-based indices⁸. Furthermore, variations in β -diversity among 232 ecological communities that are subject to large and fluctuating disturbances are believed to provide 233 insights about the mechanisms driving changes in α -diversity and function⁴⁰.

234 We observed similar trends of phylogenetic dispersion within a single community (NTI) and 235 the phylogenetic turnover between communities of the same treatment level (β NTI), compared to the 236 null expectation. Stochasticity was more important during initial successional stages of the study, with 237 initial NTI and β NTI values closer to zero (*i.e.*, closer to the null expectation of the model). 238 Relatively, the overall strength of deterministic processes increased with time, with higher |NTI| and 239 $|\beta$ NTI| values. Similarly, late succession stages were shown to be governed by deterministic processes in plant forest⁴¹ and microbial groundwater communities⁴². Furthermore, α -diversity-based temporal 240 241 assembly dynamics revealed a parabolic pattern in NTI and NTI_w, through the disturbance frequency 242 gradient, which was evident after 14 and 7 days of the study, respectively, before the appearance of 243 similar parabolic patterns across various α -diversity indices. This preceding pattern is considered here 244 as a strong indicator of assembly mechanisms operating to shape community structure. It is, therefore, 245 plausible that stochastic assembly mechanisms were first favored at intermediate disturbance

246 frequencies, prompting subsequent changes of community structure that resulted in the observed higher α -diversity as the ISH proposes¹⁷. These observations are also coherent with the idea that 247 248 secondary succession is community assembly in $action^{43}$. The disturbance range in this study 249 produced different secondary succession scenarios, with communities in the sludge of each bioreactor 250 likely experiencing different re-colonization processes from their bacterial seed-bank (i.e, lowabundance or rare taxa), via stochastic processes such as priority effects⁴⁴ followed by historical 251 contingency⁴⁵ and legacy effects³. Importantly, external dispersal processes⁴⁶ (*i.e.*, bacterial 252 253 immigration) could not influence community assembly since bioreactors within this study were 254 operated as closed systems. Indeed, microbial seed-banks are thought to contribute to the maintenance 255 of microbial diversity⁴⁷ and have been described as essential for understanding temporal community 256 changes⁴⁸. Further, stochastic assembly processes were shown to be more preponderant within the rare fraction of the microbial community²². Nonetheless, other processes might also be promoting 257 stochastic assembly at intermediate disturbance frequencies, like ecological drift³⁶ and feedback 258 mechanisms linked to density dependence and species interactions⁴⁹. Hence, a disturbance frequency 259 260 gradient can not only result in nonlinearities for growth rates that would affect the outcome of competition^{14,31}, it could also alter the relative contribution of stochastic and deterministic 261 262 mechanisms of community assembly that underlie changes in community structure¹⁷. Furthermore, 263 our results showed that, over a range of disturbance frequencies, assessing temporal community 264 assembly patterns during succession can act as a sentinel of upcoming patterns of diversity.

265 Stochasticity was positively correlated with better nitrogen (as TKN) removal via nitrification 266 at intermediate disturbance frequencies during the initial successional stages where stochastic 267 processes were also generally prevalent. Nitrification functions are carried out by specific taxa (*i.e.*, 268 nitrifiers), which are slow growers, nutritionally inflexible, sensitive to inhibitors and less phylogenetically diverse than many other key functional guilds⁵⁰. Yet, the recruitment of nitrifying 269 270 organisms from the microbial seed-bank was important for the recovery of nitrification, following the 271 removal of a long-term disturbance of altering food-to-biomass and carbon-to-nitrogen ratios in 272 sludge bioreactors, although resilience varied across identically treated replicates⁵¹. Also, partial 273 recovery of nitrification in sludge bioreactors was observed at intermediate frequencies of 3-

274 chloroaniline disturbance, where stochastic assembly processes and within-treatment variability were 275 also higher¹⁷. Conversely, general functions of carbon removal and settleability performed better 276 when deterministic processes were stronger (higher |NTI| values). Carbon removal was better when α -277 diversity was lower, similarly to what was reported previously using a different xenobiotic 278 disturbance in bioreactors¹⁷. Hence, a more diverse community does not necessarily translate into better ecosystem functions^{17,52}. Our data suggest that general functions thrive during stronger 279 280 deterministic processes, while specialized functions might be favored by stochasticity at initial 281 successional stages. Future studies assessing the effect of fluctuating disturbances on community 282 diversity and function should also consider the type of function (e.g., specific or general), the stage of 283 succession after the disturbance, and the underlying assembly mechanisms.

284 The observed patterns in community assembly, structure and function were time-dependent. 285 The ISH successional pattern appears to be transient, as assembly mechanisms across disturbance 286 frequency levels were not significantly different towards the end of the study on d42, while α -287 diversity continued to display a significant parabolic pattern. If the gradient of disturbance frequencies 288 is maintained over time, then the peak in α -diversity at intermediate levels might continue during the 289 late successional stages, but this remains to be investigated. Nonetheless, most relevant bacteria in 290 activated sludge have generation times of less than 24 h. Hence, the 42-day length of this study 291 represented around tens to hundredths of generations of many different taxa, allowing the detection of 292 significant patterns in assembly and structure. Further research in a variety of ecosystems is needed to 293 validate the broad applicability of the ISH, particularly considering that disturbance can vary in type, frequency, intensity, driver and impact^{31,53}. Studies at different scales are also necessary since 294 295 ecological patterns can vary across spatial, temporal and phylogenetic scales³, while the effect of 296 dispersal processes could also be evaluated within open systems.

Although a similar study on communities of larger organisms would require considerably larger scales of space and time, some modelling approaches suggest that ISH-like patterns (Fig. 4) could emerge in community assembly and structure under varying disturbances. For example, forest fire modelling showed that intermediate lightning strike frequency values yielded higher diversity with a

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301 close balance between stochastic and deterministic forces, which were highly sensitive to probabilistic events leading the system to diverse trajectories⁵⁴. Likewise, a conceptual model developed for plants 302 303 and animals suggested that high variation in resource abundance and location in space and time, 304 which could be caused by disturbance, would favor diversity via adaptation through novelty and 305 innovation (*i.e.*, stochasticity) generation⁵⁵. The predictions of the ISH could help to identify cases 306 when disturbance-induced stochastic assembly promotes alternative states of community structure that 307 compromise or enhance ecosystem function, so as to design mitigation or intensification strategies. 308 Furthermore, it could be used to promote community resistance and resilience to future disturbances 309 via increased α -diversity and functional-gene diversity. Alternatively, this theoretical framework 310 could help develop functionally resilient communities that do not occur naturally, through the 311 stochastic mechanisms that are initially elicited at intermediate frequencies of disturbance. Therefore, 312 we propose that the ISH has potential for a general understanding of disturbance-induced changes in 313 community structure and function during succession, by integrating the influence of the underlying 314 assembly processes over time.

315 Materials and Methods

316 Experimental design and function analyses

317 We employed 30 sequencing batch bioreactors at a microcosm scale (25-mL working volume), 318 inoculated with activated sludge from a full-scale wastewater treatment plant in Singapore and 319 operated for 42 days at 30°C in an incubator shaker. The daily complex synthetic feeding regime (adapted from Santillan et al.⁵¹) included double organic loading at varying disturbance frequencies. 320 321 Six levels of disturbance were set in quintuplicate independent reactors (n = 5), which received double 322 organic loading either never (undisturbed), every eight, six, four, or two days (intermediately-323 disturbed), or every day (press-disturbed). Level numbers were assigned from 0 to 5 (0 for no 324 disturbance, 1 to 5 for low to high disturbance frequency). Disturbance frequency was further calculated from the rate of high organic loading at each disturbance level resulting in values of 0, $\frac{1}{8}$, 325 $\frac{1}{6}$, $\frac{1}{4}$, $\frac{1}{2}$, and 1. Ecosystem function, in the form of process performance parameters at the end of a 326 cycle, was measured weekly in accordance with Standard Methods⁵⁶ where appropriate, and targeted 327

328 the removal of soluble COD and TKN from the mixed liquor after feeding. Sludge settling capacity 329 was measured via the SVI (mL/g), considering 30 minutes of settling time. Concentrations in the 330 mixed liquor of the bioreactors after feeding (*i.e.*, beginning of a new cycle) were regularly 305.8 331 (±7.4) mg COD/L and 45.6 (±0.8) mg TKN/L, or 594.7 (±18.6) mg COD/L and 46.1 (±0.2) mg 332 TKN/L when double organic loading occurred. A food-to-biomass ratio (F:M) control approach was used as previously described⁵¹, for which biomass was measured weekly as total suspended solids 333 334 (TSS) after which sludge wastage was done to target a TSS of 1,500 mg/L. The latter resulted in 335 average solids residence time (SRT) values of 30, 26, 23, 22, 19 and 15 days, for disturbance levels 336 from 0 to 5, respectively. Note that these SRT values are well above the doubling times of relevant bacteria in activated sludge⁵⁷. Sludge samples of 2 mL (m = 184) were collected on the initial day of 337 338 the study (four samples, taken at random from the inoculum mix) and weekly from each reactor 339 afterwards (180 samples), for DNA extraction as previously described³⁷.

340 16S rRNA gene metabarcoding and reads processing

Bacterial 16S rRNA metabarcoding was done in two steps as described in Santillan et al.⁵¹. Primer set 341 342 341f/785r targeted the V3-V4 variable regions of the 16S rRNA gene⁵⁸. The libraries were sequenced 343 in-house at SCELSE on an Illumina MiSeq (v.3) with 20% PhiX spike-in, at 300 bp paired-end read-344 length. Sequenced sample libraries were processed with the dada2 (v.1.3.3) R-package³³, allowing 345 inference of ASVs³². Illumina adaptors and PCR primers were trimmed prior to quality filtering. 346 Sequences were truncated after 280 and 255 nucleotides for forward and reverse reads, respectively. 347 After truncation, reads with expected error rates higher than 3 and 5 for forward and reverse reads, 348 respectively, were removed. After filtering, error rate learning, ASV inference and denoising, reads 349 were merged with a minimum overlap of 20 bp. Chimeric sequences (0.17% on average) were 350 identified and removed. For a total of 184 samples, an average of 18,086 reads were kept per sample 351 after processing, representing 47% of the average forward input reads. Taxonomy was assigned using the SILVA database $(v.132)^{59}$. Diversity and assembly analyses were carried on both unrarefied and 352 353 rarefied datasets. To generate the rarefied dataset, samples were rarefied to the lowest number of reads 354 (5,089) in a sample after processing (Fig. S11).

355 Bacterial community structure analyses and statistics

356 All reported P-values for statistical tests in this study were corrected for multiple comparisons using a false discovery rate (FDR) of 5%. Hill diversity indices²⁷ were used to quantify taxonomic α -diversity 357 as described elsewhere¹⁷. Phylogenetic α -diversity was assessed through Faith's phylogenetic 358 distance²⁸ (PD) including its abundance-weighted version (PD_w). Community structure in terms of 359 360 taxonomic β -diversity was evaluated through: i) canonical analysis of principal coordinates (CAP) 361 ordination including ellipses of 60% group-average cluster similarity; ii) misclassification error 362 analysis for each disturbance frequency level over the six time points sampled, via the leave-one-out 363 allocation of observations to groups from CAP; and iii) multivariate tests of permutational analysis of 364 variance (PERMANOVA) and permutational analysis of dispersion (PERMDISP); all from Bray-365 Curtis dissimilarity matrixes at each time point sampled (30 bioreactors, n = 5), constructed from square-root transformed abundance data using PRIMER $(v.7)^{60}$. Phylogenetic β -diversity was 366 367 assessed via non-metric multidimensional (NMDS) ordination of a weighted Unifrac dissimilarity 368 matrix, constructed from Hellinger transformed abundance data of all 184 samples using the phyloseq⁶¹ R-package (v.1.30.0) in R. The ggplot2 package (v.3.3.2) in R⁶² was used for local 369 370 polynomial regression fitting via the *loess* function (including 95% confidence intervals) and box 371 plots construction (using Tukey style whiskers). The ggdist R-package (v.2.4.1) was used to make the 372 BNTI raincloud plot. Univariate testing through Welch's analysis of variance (ANOVA) with Games-Howell post-hoc grouping was done using the $rstatix^{63}$ (v.0.6.0) R-package. Kendall correlations were 373 374 done using the $ggpubr^{64}$ package (v.0.4.0) in R. Heat maps for bacterial phyla relative abundances were constructed using the *ampvis2*⁶⁵ package (v.2.6.2) in R. 375

376 Bacterial community assembly analyses and statistics

The effect of underlying assembly mechanisms was assessed using phylogenetic-based null modelling approaches on both α - and β -diversity. First, the nearest taxon index (NTI)²⁹ was calculated for each community to assess whether α -diversity was more or less structured than would be expected by random chance. The model uses the mean nearest taxon distance (MNTD)²⁹, which quantifies the phylogenetic distance between each ASV in one community, as a measure of the clustering of closely

382 related ASVs. Phylogenetic relatedness of ASVs was characterized by multiple-alignment of ASV sequences using *decipher* (v.2.14.0) R-package⁶⁶. The phylogenetic tree was then constructed and a 383 384 GTR+G+I maximum likelihood tree was fitted using the *phangorn* (v.2.5.5) R-package⁶⁷. To quantify 385 the degree to which MNTD deviates from a null model expectation, ASVs and abundances were 386 shuffled across the tips of the phylogenetic tree. After shuffling, MNTD was recalculated to obtain a 387 null value, and repeating the shuffling 1,000 times provided a null distribution. Then, NTI was 388 calculated as the difference between the mean of the null distribution and the observed MNTD in 389 units of standard deviation²⁹. The closer to zero a NTI value is, the closer to the null expectation (*i.e.*, 390 higher stochasticity) is the phylogenetic dispersion of a given community. Positive NTI values 391 suggest phylogenetic clustering while negative values indicate phylogenetic overdispersion. Second, 392 β -diversity null modelling via the β -nearest taxon index (β NTI) was done to investigate if the 393 phylogenetic turnover across two samples was significantly more or less similar than would be expected by just random chance³⁰. The model uses the β -mean nearest taxon distance (β MNTD), 394 395 which quantifies the phylogenetic distance between pairs of ASVs drawn from two distinct 396 communities. To quantify the degree to which β MNTD deviates from a null model expectation, ASVs 397 and abundances were shuffled across the tips of the phylogenetic tree. After shuffling, β MNTD was 398 recalculated to obtain a null value, and repeating the shuffling 1,000 times provided a null 399 distribution. Then, β NTI was calculated as the difference between the mean of the null distribution 400 and the observed β MNTD in units of standard deviation³⁰. The closer to zero a β NTI value is, the 401 closer to the null expectation (*i.e.*, higher stochasticity) is the phylogenetic turnover between two 402 communities. By convention, a value of $|\beta NTI| > 2$ indicates that the observed turnover is 403 significantly deterministic, while $|\beta NTI| < 2$ indicates dominance of stochastic assembly processes²⁰. 404 Similarly, here we consider that |NTI| < 2 indicates dominance of stochastic phylogenetic clustering. 405 Both unweighted and abundance-weighted NTI and β NTI values were calculated. These analyses were done using the $metagMisc^{68}$ (v.0.0.4) and $picante^{69}$ (v.1.8.2) R-packages. To test for a 406 407 phylogenetic signal across phylogenetic distances, Mantel correlograms were constructed using the vegan⁷⁰ (v.2.5.6) R-package, relating between-ASV niche differences to between-ASV phylogenetic 408

409 distances across a given phylogenetic distance, following the previously described methodology^{20,30}.

410 Environmental niches were constructed from bioreactor effluent process data (COD removal, TKN

411 removal and SVI). Phylogenetic distances were quantified for 50 phylogenetic distance bins, while the

412 significance of Pearson correlations was assessed using 1,000 permutations and FDR (5%) correction.

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420 Author Contributions

421 Both authors conceived the idea. ES designed and performed the experiment, as well as data 422 processing and analyses. SW obtained the funding for the study. ES wrote the manuscript draft. Both 423 authors contributed to manuscript editing.

424 **Data availability**

DNA sequencing data are available at NCBI BioProjects with accession number PRJNA723443. See supplementary information for details about the sludge inoculum collection, synthetic feed preparation, and additional figures of diversity and community assembly metrics, correlations, heat maps and data rarefaction. Diversity analyses on rarefied data and all other relevant data to reproduce the results of this study are available as supplementary files in the online version of this manuscript.

430 **Competing interests**

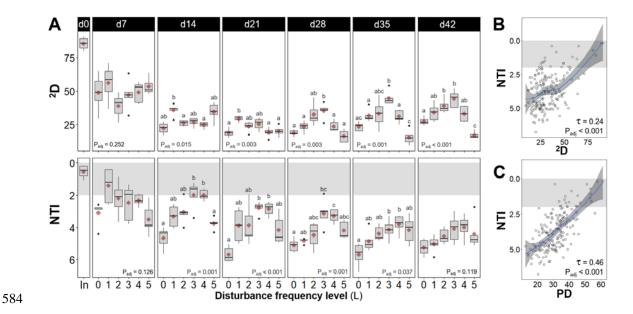
431 The authors declare no competing interests.

| 432 433 | | | | | |
|------------|----|--|--|--|--|
| 434 | 1 | Widder, S. et al. Challenges in microbial ecology: building predictive understanding of | | | |
| 435 | | community function and dynamics. <i>ISME J.</i> 10 , 2557-2568 (2016). | | | |
| 436 | 2 | Flemming, HC. & Wuertz, S. Bacteria and archaea on Earth and their abundance in | | | |
| 437 | - | biofilms. <i>Nat. Rev. Microbiol.</i> 17 , 247–260 (2019). | | | |
| 438 | 3 | Ladau, J. & Eloe-Fadrosh, E. A. Spatial, Temporal, and Phylogenetic Scales of Microbial | | | |
| 439 | U | Ecology. Trends Microbiol. (2019). | | | |
| 440 | 4 | Wagg, C., Bender, S. F., Widmer, F. & van der Heijden, M. G. A. Soil biodiversity and soil | | | |
| 441 442 | · | community composition determine ecosystem multifunctionality. <i>Proceedings of the National Academy of Sciences</i> 111 , 5266-5270 (2014). | | | |
| 443 | 5 | Shade, A. <i>et al.</i> Fundamentals of microbial community resistance and resilience. <i>Front.</i> | | | |
| 444 | 5 | Microbiol. 3, 1-19 (2012). | | | |
| 445 | 6 | Zalasiewicz, J., Williams, M., Steffen, W. & Crutzen, P. The New World of the | | | |
| 446 | 0 | Anthropocene. <i>Environ. Sci. Technol.</i> 44, 2228-2231 (2010). | | | |
| 440 447 | 7 | Connell, J. H. Diversity in tropical rain forests and coral reefs. <i>Science</i> 199 , 1302-1310 | | | |
| 448 | / | (1978). | | | |
| 448 449 | 8 | Svensson, J. R., Lindegarth, M., Jonsson, P. R. & Pavia, H. Disturbance–diversity models: | | | |
| | 0 | what do they really predict and how are they tested? <i>Proceedings of the Royal Society B:</i> | | | |
| 450 | | | | | |
| 451 452 | 9 | Biological Sciences 279, 2163-2170 (2012). | | | |
| 452 453 | 9 | Yuan, Z. Y., Jiao, F., Li, Y. H. & Kallenbach, R. L. Anthropogenic disturbances are key to maintaining the biodiversity of grasslands. <i>Sci. Rep.</i> 6 , 22132-22132 (2016). | | | |
| 455 454 | 10 | Sasaki, T. <i>et al.</i> Management applicability of the intermediate disturbance hypothesis across | | | |
| 454 | 10 | Mongolian rangeland ecosystems. <i>Ecol. Appl.</i> 19 , 423-432 (2009). | | | |
| 455 456 | 11 | | | | |
| 450 457 | 11 | Roxburgh, S. H., Shea, K. & Wilson, J. B. The intermediate disturbance hypothesis: Patch dynamics and mechanisms of species coexistence. <i>Ecology</i> 85 , 359-371 (2004). | | | |
| 457 | 12 | Mackey, R. L. & Currie, D. J. The diversity-disturbance relationship: Is it generally strong | | | |
| 458 459 | 12 | and peaked? <i>Ecology</i> 82 , 3479-3492 (2001). | | | |
| 460 | 13 | Kershaw, H. M. & Mallik, A. U. Predicting Plant Diversity Response to Disturbance: | | | |
| 460 | 15 | Applicability of the Intermediate Disturbance Hypothesis and Mass Ratio Hypothesis. <i>Crit.</i> | | | |
| 462 | | <i>Rev. Plant Sci.</i> 32 , 383-395 (2013). | | | |
| 463 | 14 | Fox, J. W. The intermediate disturbance hypothesis should be abandoned. <i>Trends Ecol. Evol.</i> | | | |
| 464 | 14 | 28 , 86-92 (2013). | | | |
| 465 | 15 | Sheil, D. & Burslem, D. Defining and defending Connell's intermediate disturbance | | | |
| 466 | 15 | hypothesis: a response to Fox. <i>Trends Ecol. Evol.</i> 28 , 571-572 (2013). | | | |
| 467 | 16 | Shea, K., Roxburgh, S. H. & Rauschert, E. S. J. Moving from pattern to process: coexistence | | | |
| 468 | 10 | mechanisms under intermediate disturbance regimes. <i>Ecol. Lett.</i> 7 , 491-508 (2004). | | | |
| 469 | 17 | Santillan, E., Seshan, H., Constancias, F., Drautz-Moses, D. I. & Wuertz, S. Frequency of | | | |
| 470 | 17 | disturbance alters diversity, function, and underlying assembly mechanisms of complex | | | |
| 471 | | bacterial communities. <i>NPJ Biofilms Microbiomes</i> 5 , 1-8 (2019). | | | |
| 472 | 18 | Leibold, M. A., Chase, J. M. & Ernest, S. K. M. Community assembly and the functioning of | | | |
| 473 | 10 | ecosystems: how metacommunity processes alter ecosystems attributes. <i>Ecology</i> 98 , 909-919 | | | |
| 474 | | (2017). | | | |
| 475 | 19 | Chase, J. M. & Myers, J. A. Disentangling the importance of ecological niches from | | | |
| 476 | 17 | stochastic processes across scales. <i>Philosophical Transactions of the Royal Society B</i> - | | | |
| 477 | | Biological Sciences 366 , 2351-2363 (2011). | | | |
| 478 | 20 | Dini-Andreote, F., Stegen, J. C., van Elsas, J. D. & Salles, J. F. Disentangling mechanisms | | | |
| 479 | 20 | that mediate the balance between stochastic and deterministic processes in microbial | | | |
| 480 | | succession. <i>Proc. Natl. Acad. Sci. USA</i> 112 , E1326-E1332 (2015). | | | |
| 481 | 21 | Santillan, E., Seshan, H. & Wuertz, S. Press xenobiotic 3-chloroaniline disturbance favors | | | |
| 482 | | deterministic assembly with a shift in function and structure of bacterial communities in | | | |
| 483 | | sludge bioreactors. ACS ES&T Water 1, 1429-1437 (2021). | | | |
| 484 | 22 | Santillan, E., Constancias, F. & Wuertz, S. Press disturbance alters community structure and | | | |
| 485 | | assembly mechanisms of bacterial taxa and functional genes in mesocosm-scale bioreactors. | | | |
| 486 | | mSystems 5, e00471-00420 (2020). | | | |

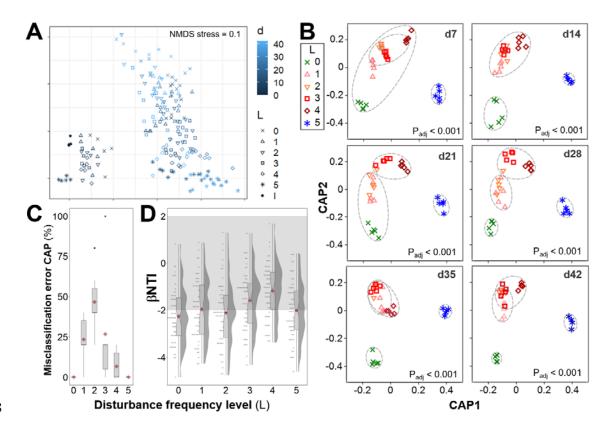
| 487 | 23 | Zhou, J. Z. et al. Stochastic assembly leads to alternative communities with distinct functions |
|------------|-----|---|
| 488 | | in a bioreactor microbial community. mBio 4, 1-8 (2013). |
| 489 | 24 | Xun, W. et al. Diversity-triggered deterministic bacterial assembly constrains community |
| 490 | | functions. Nat. Commun. 10, 3833 (2019). |
| 491 | 25 | Gao, C. et al. Fungal community assembly in drought-stressed sorghum shows stochasticity, |
| 492 | | selection, and universal ecological dynamics. Nat. Commun. 11, 34 (2020). |
| 493 | 26 | Drake, J. M. & Kramer, A. M. Mechanistic analogy: how microcosms explain nature. Theor. |
| 494 | | <i>Ecol.</i> 5 , 433-444 (2012). |
| 495 | 27 | Hill, M. O. Diversity and evenness: a unifiying notation and its consequences. <i>Ecology</i> 54, |
| 496 | • • | 427-432 (1973). |
| 497 | 28 | Faith, D. P. Conservation evaluation and phylogenetic diversity. <i>Biol. Conserv.</i> 61 , 1-10 |
| 498 | • | (1992). |
| 499 | 29 | Webb, C. O., Ackerly, D. D., McPeek, M. A. & Donoghue, M. J. Phylogenies and |
| 500 | • | Community Ecology. Annu. Rev. Ecol. Syst. 33, 475-505 (2002). |
| 501 | 30 | Stegen, J. C. <i>et al.</i> Quantifying community assembly processes and identifying features that |
| 502 | 0.1 | impose them. <i>ISME J.</i> 7 , 2069-2079 (2013). |
| 503 | 31 | Miller, A. D., Roxburgh, S. H. & Shea, K. How frequency and intensity shape diversity- |
| 504 | 22 | disturbance relationships. Proc. Natl. Acad. Sci. USA 108, 5643-5648 (2011). |
| 505 | 32 | Callahan, B. J., McMurdie, P. J. & Holmes, S. P. Exact sequence variants should replace |
| 506 | 22 | operational taxonomic units in marker-gene data analysis. <i>ISME J.</i> 11 , 2639–2643 (2017). |
| 507 | 33 | Callahan, B. J. <i>et al.</i> DADA2: High resolution sample inference from Illumina amplicon data. |
| 508 | 24 | Nat. Methods 13, 581-583 (2016). |
| 509 | 34 | Weiss, S. <i>et al.</i> Normalization and microbial differential abundance strategies depend upon |
| 510 | 25 | data characteristics. <i>Microbiome</i> 5 , 27 (2017). |
| 511 512 | 35 | McMurdie, P. J. & Holmes, S. Waste not, want not: why rarefying microbiome data is inadmissible. <i>BL</i> of <i>Comm. Biol.</i> 10 , 21002521 (2014) |
| 512 | 36 | inadmissible. <i>PLoS Comp. Biol.</i> 10 , e1003531 (2014). Zhou, J. & Ning, D. Stochastic community assembly: does it matter in microbial ecology? |
| 515 514 | 50 | Microbiol. Mol. Biol. Rev. 81, 1-32 (2017). |
| 514 | 37 | Santillan, E., Seshan, H., Constancias, F. & Wuertz, S. Trait-based life-history strategies |
| 516 | 57 | explain succession scenario for complex bacterial communities under varying disturbance. |
| 517 | | <i>Environ. Microbiol.</i> 21 , 3751-3764 (2019). |
| 518 | 38 | Martiny, A. C., Treseder, K. & Pusch, G. Phylogenetic conservatism of functional traits in |
| 519 | 50 | microorganisms. ISME J. 7, 830-838 (2013). |
| 520 | 39 | Chase, J. M. Stochastic Community Assembly Causes Higher Biodiversity in More |
| 521 | 57 | Productive Environments. Science 328 , 1388-1391 (2010). |
| 522 | 40 | Mori, A. S., Isbell, F. & Seidl, R. Beta-diversity, community assembly, and ecosystem |
| 522 | 10 | functioning. Trends Ecol. Evol. 33 , 549-564 (2018). |
| 524 | 41 | Chai, Y. <i>et al.</i> Patterns of taxonomic, phylogenetic diversity during a long-term succession of |
| 525 | | forest on the Loess Plateau, China: insights into assembly process. <i>Sci. Rep.</i> 6 , 27087 (2016). |
| 526 | 42 | Zhou, J. Z. et al. Stochasticity, succession, and environmental perturbations in a fluidic |
| 527 | | ecosystem. Proc. Natl. Acad. Sci. USA 111, E836-E845 (2014). |
| 528 | 43 | Lebrija-Trejos, E., Pérez-García, E. A., Meave, J. A., Bongers, F. & Poorter, L. Functional |
| 529 | | traits and environmental filtering drive community assembly in a species-rich tropical system. |
| 530 | | <i>Ecology</i> 91 , 386-398 (2010). |
| 531 | 44 | Nemergut, D. R. et al. Patterns and processes of microbial community assembly. Microbiol. |
| 532 | | Mol. Biol. Rev. 77, 342-356 (2013). |
| 533 | 45 | Fukami, T. Historical Contingency in Community Assembly: Integrating Niches, Species |
| 534 | | Pools, and Priority Effects. Annu. Rev. Ecol. Evol. Syst. 46, 1-23 (2015). |
| 535 | 46 | Hanson, C. A., Fuhrman, J. A., Horner-Devine, M. C. & Martiny, J. B. H. Beyond |
| 536 | | biogeographic patterns: processes shaping the microbial landscape. Nat. Rev. Microbiol. 10, |
| 537 | | 497-506 (2012). |
| 538 | 47 | Lennon, J. T. & Jones, S. E. Microbial seed banks: the ecological and evolutionary |
| 539 | | implications of dormancy. Nat. Rev. Microbiol. 9, 119 (2011). |
| 540 | 48 | Shade, A. et al. Conditionally rare taxa disproportionately contribute to temporal changes in |
| 541 | | microbial diversity. <i>mBio</i> 5, 1-9 (2014). |
| | | |

| 542 | 49 | Holyoak, M. & Loreau, M. Reconciling empirical ecology with neutral community models. |
|-----|----|---|
| 543 | | <i>Ecology</i> 87 , 1370-1377 (2006). |
| 544 | 50 | Wagner, M. et al. Microbial community composition and function in wastewater treatment |
| 545 | | plants. Antonie Van Leeuwenhoek International Journal of General and Molecular |
| 546 | | Microbiology 81 , 665-680 (2002). |
| 547 | 51 | Santillan, E., Phua, W. X., Constancias, F. & Wuertz, S. Sustained organic loading |
| 548 | | disturbance favors nitrite accumulation in bioreactors with variable resistance, recovery and |
| 549 | | resilience of nitrification and nitrifiers. Sci. Rep. 10, 21388 (2020). |
| 550 | 52 | Shade, A. Diversity is the question, not the answer. <i>ISME J.</i> 11 , 1-6 (2017). |
| 551 | 53 | Graham, E. B. et al. Toward a generalizable framework of disturbance ecology through |
| 552 | | crowdsourced science. Frontiers in Ecology and Evolution 9 (2021). |
| 553 | 54 | Savage, M., Sawhill, B. & Askenazi, M. Community Dynamics: What Happens When We |
| 554 | | Rerun the Tape? J. Theor. Biol. 205, 515-526 (2000). |
| 555 | 55 | Allen, C. R. & Holling, C. Novelty, adaptive capacity, and resilience. Ecol. Soc. 15 (2010). |
| 556 | 56 | Rice, E. W., Baird, R. B. & Eaton, A. D. Standard Methods for the Examination of Water and |
| 557 | | Wastewater. 23 edn, (APHA-AWWA-WEF, 2017). |
| 558 | 57 | Tchobanoglous, G., Stensel, H. D., Tsuchihashi, R. & Burton, F. L. Wastewater engineering: |
| 559 | | treatment and resource recovery. 5th edn, (McGraw Hill Education 2013). |
| 560 | 58 | Thijs, S. et al. Comparative evaluation of four bacteria-specific primer pairs for 16S rRNA |
| 561 | | gene surveys. Front. Microbiol. 8, 1-15 (2017). |
| 562 | 59 | Glöckner, F. O. et al. 25 years of serving the community with ribosomal RNA gene reference |
| 563 | | databases and tools. J. Biotechnol. 261, 169-176 (2017). |
| 564 | 60 | Clarke, K. R. & Gorley, R. N. PRIMER v7: User Manual/Tutorial. (PRIMER-E, 2015). |
| 565 | 61 | McMurdie, P. J. & Holmes, S. phyloseq: an R package for reproducible interactive analysis |
| 566 | | and graphics of microbiome census data. PLoS One 8, e61217 (2013). |
| 567 | 62 | Wickham, H. ggplot2: elegant graphics for data analysis. 2nd edn, (Springer-Verlag, 2016). |
| 568 | 63 | Kassambara, A. rstatix: pipe-friendly framework for basic statistical tests. <i>R-package</i> (v.0.6.0) |
| 569 | | (2020). |
| 570 | 64 | Kassambara, A. ggpubr: "ggplot2" based publication ready plots. <i>R-package</i> (v.0.1.6) (2017). |
| 571 | 65 | Albertsen, M., Karst, S. M., Ziegler, A. S., Kirkegaard, R. H. & Nielsen, P. H. Back to basics |
| 572 | | - the influence of DNA extraction and primer choice on phylogenetic analysis of activated |
| 573 | | sludge communities. PLoS One 10, 15 (2015). |
| 574 | 66 | Wright, E. S. Using decipher v2.0 to analyze big biological sequence data in R. <i>R Journal</i> 8, |
| 575 | | 352-359 (2016). |
| 576 | 67 | Schliep, K. P. phangorn: phylogenetic analysis in R. Bioinformatics 27, 592-593 (2010). |
| 577 | 68 | Mikryukov, V. metagMisc: Miscellaneous functions for metagenomic analysis. <i>R-package</i> |
| 578 | | (v.0.0.4) (2020). |
| 579 | 69 | Kembel, S. W. et al. Picante: R tools for integrating phylogenies and ecology. Bioinformatics |
| 580 | | 26 , 1463-1464 (2010). |
| 581 | 70 | Oksanen, F. J. et al. vegan: community ecology package. R-package (v.2.5.6) (2019). |
| 582 | | |
| 583 | | |

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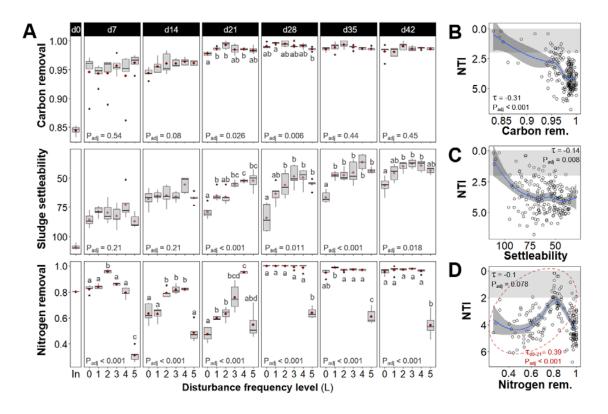
585 Fig. 1 – Community dynamics in α -diversity. (A) Community structure assessed via 2nd order true α -586 diversity (²D, upper panels) and community assembly evaluated via the nearest taxon index (NTI, 587 lower panels), from bacterial ASV data for different frequencies of organic loading disturbance (n =588 5). Disturbance frequency levels (L): 0 (undisturbed), 1-4 (intermediately disturbed), 5 (press-589 disturbed). In: sludge inoculum (day 0, n = 4). Each panel represents a sampling day, red diamonds display mean values. Characters above boxes display Games-Howell post-hoc grouping ($P_{adj} < 0.05$). 590 591 Welch's ANOVA P-values adjusted at 5% FDR shown within panels. Correlations of (B) ^{2}D and (C) 592 phylogenetic diversity (PD) versus NTI from bacterial ASV data across all frequency levels and time 593 points evaluated in this study (m = 184). Kendall correlation τ - and adjusted P-values are indicated 594 within the panel. Blue line represents locally estimated scatterplot smoothing regression (loess) with 595 confidence interval in dark-grey shading. Note the inverted y-axis for NTI, as values closer to zero 596 indicate a higher relative contribution of stochastic assembly. Shaded in grey is the zone of significant 597 stochastic phylogenetic dispersion, |NTI| < 2.



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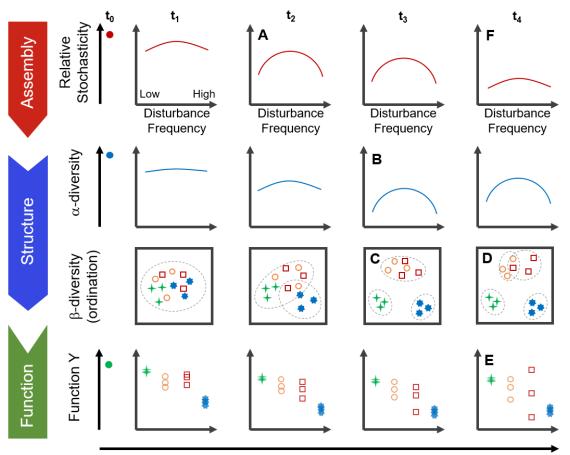
Fig. 2 – Temporal dynamics of β -diversity community structure and assembly for bacterial ASV data 599 600 across different frequencies of organic loading disturbance (n = 5 bioreactors). (A) Unconstrained 601 NMDS ordination (weighed Unifrac β -diversity, Hellinger transformed data) for all 184 samples 602 collected. Disturbance frequency levels (L): 0 (undisturbed), 1-4 (intermediately disturbed), 5 (press-603 disturbed). I: Sludge inoculum (day 0, n = 4). (B) Constrained canonical analysis of principal 604 coordinates (CAP) ordinations (Bray-Curtis β -diversity, squared root transformed data) on different 605 sampling days, including ellipses of 60% group-average cluster similarity and PERMANOVA 606 adjusted P-values. (C) Misclassification errors at each disturbance frequency level, via the leave-one-607 out allocation of observations to groups from CAP at each time point after d0 (n = 6 sampling days). 608 Bray-Curtis β -diversity, squared root transformed data. Red diamonds display mean values. (**D**) Beta 609 nearest taxon index (β NTI) at each disturbance frequency level, from pairwise comparisons across 610 within-treatment replicates at each time point after d0 (n = 60 comparisons). Red diamonds display 611 mean values. Notches show the 95% confidence interval for the median. When notches do not 612 overlap, the medians can be judged to differ significantly. Shaded in grey is the zone where stochastic 613 processes significantly dominate, $|\beta NTI| < 2$. βNTI values closer to zero indicate a higher relative 614 contribution of stochastic assembly.

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616 Fig. 3 – Community function assessed via influent chemical oxygen demand removal (carbon 617 removal, upper panels), sludge volume index (sludge settleability, middle panels), and influent total 618 Kjeldahl nitrogen removal (nitrogen removal, lower panels) for different frequencies of organic 619 loading disturbance (n = 5). Disturbance frequency levels (L): 0 (undisturbed), 1-4 (intermediately 620 disturbed), 5 (press-disturbed). In: sludge inoculum (day 0, n = 4). Each panel represents a sampling 621 day, red diamonds display mean values. Characters above boxes display Games-Howell post-hoc 622 grouping (P_{adj} < 0.05). Welch's ANOVA P-values adjusted at 5% FDR shown within panels. 623 Correlations of (B) carbon removal, (C) sludge settleability, and (D) nitrogen removal, versus NTI 624 from bacterial ASV data across all frequency levels and time points evaluated in this study (m = 184). 625 Kendall correlation τ - and adjusted P-values are indicated within the panels. Blue line represents 626 locally estimated scatterplot smoothing regression (loess) with confidence interval in dark-grey 627 shading. Shaded in grey is the zone of significant stochastic phylogenetic dispersion, |NTI| < 2. Red 628 ellipse and τ - and P-value in panel (**D**) indicate data at initial stages of succession (d0 to d21). Note 629 the inverted axis for sludge settleability, as it increases with decreasing SVI values, and for NTI, since 630 values closer to zero indicate a higher relative contribution of stochastic assembly.

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Time since start of secondary succession

632 Fig. 4 - Conceptual representation of the intermediate stochasticity hypothesis (ISH) to describe 633 patterns of assembly and structure along a disturbance frequency gradient, for communities in 634 secondary succession (starting at time point t_0). (A) Initially, stochastic assembly mechanisms (e.g., 635 priority effects, historical contingency and legacy effects) are favored at intermediate disturbance 636 frequencies, promoting re-colonization processes from the low-abundance fraction of the community 637 or seed-bank. (B) Subsequently, these are followed by changes in the community structure that 638 manifest as a peak of α -diversity at intermediate levels of disturbance. (C) At least three separated 639 clusters of β -diversity ordination would form over time across the disturbance range. However, 640 stochasticity operating at intermediate disturbance levels may lead to variable within treatment (**D**) β -641 diversity and (E) community function. (F) The overall relative contribution of stochasticity decreases 642 with succession time. The observed patterns of diversity are stronger in terms of relative abundances 643 than richness, as well as at the phylogenetic versus the taxonomic level.