

1 **Intraspecific interaction of host plant influences local distribution of specialist**

2 **herbivores through metabolic alterations in leaves**

3

4 **Last revision: July 30, 2021**

5

6 Haruna Ohsaki^{1*}, Atsuko Miyagi², Maki Kawai-Yamada², Akira Yamawo^{1*}

7 **Author information:** ¹Department of Biological Sciences, Faculty of Agriculture and

8 Life Science, Hirosaki University, 3 Bunkyo-cho, Hirosaki, Aomori 036-8561, Japan;

9 ²Graduate School of Science and Engineering, Saitama University, 225 Shimo-Okubo,

10 Sakura-ku, Saitama-city, Saitama 338-8570, Japan

11

12 *Corresponding authors: Haruna Ohsaki and Akira Yamawo, E-Mail:

13 mric3706hrn@gmail.com, yamawo.aki@gmail.com, Phone: +81 172 39 3822

14

15 **Author contributions:** H. O. & A. Y. developed the core idea, designed the

16 experiments, carried out field, cultivation and mesocosm experiments, and analysed

17 data. A. M. & M. K.-Y. analysed the primary metabolites. H. O., A. M., M. K. & A. Y.

18 wrote the article.

19

20 **COMPETING INTERESTS**

21 The authors declare that they have no competing interests.

22

23 **Abstract**

24 1. Recent studies suggest that changes in leaf traits due to interactions between plants
25 affect the resource utilisation and distribution of herbivores. However, this has not
26 yet been confirmed experimentally. Here, we investigated the effects of phenotypic
27 plasticity in leaf traits of *Rumex obtusifolius* (host plant) in response to the intra-
28 and interspecific interaction on distribution of two leaf beetles, *Gastrophysa*
29 *atrocyanea* (specialist herbivore) and *Galerucella grisescens* (generalist herbivore).

30 2. We investigated the local population density of *R. obtusifolius* plants and the
31 presence of leaf beetles on the plants at five study sites. Leaf chemicals (condensed
32 tannins and total phenolics) were compared between aggregated and solitary *R.*
33 *obtusifolius* plants. To clarify the effects of the interaction environment of *R.*
34 *obtusifolius* plants on their leaf traits and resource utilisation by leaf beetles, we
35 conducted cultivation and preference experiments. Leaf chemicals (chlorophylls,
36 organic acids, primary metabolites, condensed tannins and total phenolics) and
37 preferences of adult leaf beetles were compared between intraspecific, interspecific
38 plant interaction, or no-interaction treatments. Finally, we evaluated the effects of
39 interaction between *R. obtusifolius* on leaf beetle distribution in mesocosm
40 experiments.

41 3. In the field, the presence of the specialist leaf beetle, *G. atrocyanea*, was positively
42 correlated with the local population density (rosette overlap ratio) of *R. obtusifolius*
43 plants; however, no correlation was observed in the case of the generalist leaf
44 beetle, *G. grisescens*. In the cultivation experiment, plants in the intraspecific
45 interaction treatment increased their leaf contents of condensed tannins and total
46 phenolics, and *G. atrocyanea* consumed more of these leaves than leaves in other
47 treatments. Similar results were observed in the field. In the mesocosm experiment,
48 larger numbers of *G. atrocyanea* were distributed on *R. obtusifolius* plants exposed
49 to below-ground intraspecific interaction than on plants not exposed to intraspecific
50 interaction.

51 4. Our results provide experimental evidence that leaf-trait changes in response to
52 intraspecific interaction between host plants influence specialist herbivore
53 distribution. This highlights the need to integrate plant–plant interactions into our
54 understanding of plant–animal interactions.

55 **Keywords:** herbivory, host selection, phenotypic plasticity, plant–plant interactions,
56 plant–herbivore interactions, resource concentration hypothesis, *Rumex*, secondary
57 metabolites

58 INTRODUCTION

59 To improve our understanding of plant–animal interactions, numerous ecologists have
60 tried to predict herbivorous insect distribution by using the local population density of
61 host plants. Root (1973) predicted that herbivores would be concentrated on host plants
62 growing in high-density populations or monocultures (resource concentration
63 hypothesis). This prediction has been supported by several studies (e.g., Stephens &
64 Myers, 2012; Nerlekar, 2018). On the other hand, the possibility of an inverse
65 distribution pattern, in which herbivores are concentrated on low-density or solitary host
66 plants (Yamamura, 1999; Otway et al., 2005), has been proposed as the resource
67 dilution hypothesis, and these predictions have also been supported by several studies
68 (e.g., Fagundes et al., 2019; Coutinho et al., 2019). These conflicting patterns have been
69 reported for several herbivore species, and some species have even been found to be
70 unresponsive to resource distribution (Rhains and English-Loeb, 2003; Tuller et al.,
71 2013). Regardless, the mechanism that produces the uneven distribution of each
72 herbivore species has not been elucidated.

73 Differences in the local population density of host plants are likely to reflect
74 differences in the quality of the host plants, because the local population density of
75 plants is linked to their interaction environments; host plants present at high density are

76 exposed to direct intraspecific interaction. In contrast, host plants present at low density
77 are exposed to direct interspecific interaction or no interaction. Many studies have
78 reported that plant–plant direct interactions influence herbivory (Hambäck &
79 Beckerman, 2003; Muiruri et al., 2019; Yamawo, 2021). For example, plant competition
80 for resources induces plastic changes in the plants’ resource allocation; these changes
81 can affect root or shoot growth. The changes in resource allocation can also influence
82 the expression of leaf thickness, leaf mass per area, and primary (essential nutrients) and
83 secondary (potentially plant-protective compounds) metabolites in the leaves (Bartron
84 & Bowers, 2006; Broz et al., 2010; Mraja et al., 2011; Takigahira & Yamawo, 2019;
85 Yamawo, 2021). Therefore, variations in the interaction environment induce changes in
86 expression of the chemical traits of host plant leaves (Barton & Bowers, 2006; Broz et
87 al., 2010; Mraja et al., 2011; Muiruri et al., 2019), so they are likely to influence leaf
88 herbivory and the distribution of herbivores. In fact, several studies have strongly
89 suggested that changes in leaf traits in response to differences in the interaction
90 environment of host plants influence herbivory or herbivore distribution (e.g., Broz et
91 al. 2010; Muiruri et al., 2019; Yamawo, 2021). However, to our knowledge, no
92 experimental evidence has yet been provided for this effect.

93 Specialist herbivores are often attracted by secondary metabolites in their host

94 plants; they adapt to these chemicals because they use them as cues to recognise the
95 host plants (e.g., Wheat et al., 2007; Goodey et al., 2015). Brassicaceae plants produce
96 glucosinolates to prevent herbivory by generalist herbivores; however, *Brevicoryne*
97 *brassicae* (cabbage aphid), which specialises in Brassicaceae plants, prefers these
98 glucosinolates (Titayavan & Altieri, 1990). Therefore, a high content of secondary
99 metabolites induced in host plants by intraspecific interaction may attract the plants'
100 specialist herbivores. In contrast, generalist herbivores avoid secondary metabolites
101 (e.g., alkaloids, phenolics and condensed tannins) in the leaves of host plants (e.g.,
102 Schoonhoven et al., 2005; Macel, 2011; Jeschke et al., 2017). Here, we hypothesised
103 that intraspecific interaction increases the content of secondary metabolites in plant
104 leaves, and that this increase would lead to the aggregation of specialist herbivores. In
105 contrast, generalist herbivores may gravitate towards low-density host plants to avoid
106 high contents of secondary metabolites. Therefore, differences in resource quality due to
107 variations in local population density within a plant population could induce either a
108 concentrated or a low-density distribution of herbivores, depending on the resource
109 concentration (Root, 1973) or resource dilution (Otway et al., 2005) hypothesis, when
110 compared with the distribution predicted on the basis of resource quantity alone
111 (Fretwell & Lucas, 1969).

112 Here, we focused on *Rumex obtusifolius* L. (broad-leaved dock; Polygonaceae)
113 as a host plant, and two leaf beetles, *Gastrophysa atrocyanea* Motschulsky
114 (Chrysomelidae), which is a specialist herbivore of *Rumex* plants, and *Galerucella*
115 *grisescens* (Joannis) (Chrysomelidae), which is a generalist herbivore of Polygonaceae
116 plants (see details in Supplemental methods). To test our hypothesis, we investigated the
117 relationships between the local population density of *R. obtusifolius* plants and the
118 herbivores' distributions in the field. Next, to clarify the effect of the interaction
119 environment on leaf traits of *R. obtusifolius* plants and resource utilisation by the two
120 leaf beetles, we conducted cultivation and preference experiments in adult leaf beetles.
121 Finally, we evaluated the effects of intraspecific interaction of *R. obtusifolius* plants on
122 the distribution of the leaf beetles by using a mesocosm experiment. On the basis of the
123 results, we discuss the effects of plant–plant interaction on herbivore distributions.

124

125

126 **MATERIALS AND METHODS**

127 **Field survey**

128 To reveal the relationships between the local population density of *Rumex obtusifolius*
129 and the distribution of leaf beetles, we conducted field surveys in April and May 2018,

130 at a time when the populations of both leaf beetles are large. Five grasslands were
131 selected as field-survey sites (5 April, Tomino-cho, Hirosaki City, Aomori Prefecture,
132 40°35'N 140°28'E; 13 April, Ozawa, Hirosaki City, Aomori Prefecture, 40°34'N
133 140°27'E; 28 April, Ohara, Hirosaki City, Aomori Prefecture, 40°34'N 140°26'E; 22
134 April, Nagoya City, Aichi Prefecture, 35°09'N 136°58'E; 5 May, Morioka City, Iwate
135 Prefecture, 39°42'N 141°08'E, Figure S1). These sites are all at least 2 km apart. At each
136 site we set up one square quadrat (Tomino-cho and Ohara, 10 × 10 m; Ozawa, 8 × 8 m;
137 Iwate, 4 × 6 m; Nagoya; 4 × 4 m) including varying local population densities and sizes
138 of *R. obtusifolius* plants. The maximum size of each quadrat was determined as 100 m²;
139 in the case of small *R. obtusifolius* populations we adjusted the size of the quadrat
140 downward to include all *R. obtusifolius* individuals.

141

142 ***Survey of local population densities of R. obtusifolius and herbivore distributions on***

143 ***R. obtusifolius***

144 In each quadrat, a corner was used as the origin of two axes, x and y, which we used to
145 plot coordinates. From the origin, we described the positions of all *R. obtusifolius*
146 individuals, except for first-year seedlings that had cotyledons, to a precision of 1 cm by
147 using a ruler. The longest rosette diameters of the described *R. obtusifolius* plants were

148 recorded as the plant size. We also recorded the presence or absence of each herbivore
149 species on each *R. obtusifolius* individual, regardless of the beetles' developmental
150 stages (egg, larva or adult).

151 By using these data, a bubble chart was created by converting the positions of
152 plants into distributions on a map and the rosette sizes into bubble sizes (Figure S2). As
153 an indicator of the local population density of *R. obtusifolius*, the area of one rosette
154 overlapping with the rosettes of neighbouring individuals was calculated by using image
155 analysis software (Adobe Photoshop Elements 2.0, Adobe Systems, San Jose, CA,
156 USA). The overlap ratio (overlapping area/total rosette area) was used to represent the
157 population density for analytical purposes. Because leaf beetles often retire into the soil
158 around the host plants, making it difficult to evaluate their numbers accurately, we used
159 binomial data (presence or absence) to analyse their distribution. The correlations
160 between the overlap ratio of the rosettes and the presence of *Gastrophysa atrocyanea* or
161 *Galerucella grisescens* were examined.

162 ***Measurement of leaf traits in field plants***

163 To reveal the effects of local population density on the secondary chemicals of *R.*
164 *obtusifolius* in the field, we measured the leaf secondary metabolites of *R. obtusifolius*
165 plants that grew alone or were aggregated. In April 2018, leaves of *R. obtusifolius* plants

166 were collected from three study sites in Aomori Prefecture, northern Japan (Hirosaki-
167 city: 40° 35'N 140° 28'E, Fujisaki-city: 40° 39' N 140° 29' E, Itayanagi-city: 40° 40' N
168 140°28' E). Each site was at least 10 km apart from the next site. To exclude the effects
169 of reproduction and leaf damage, we selected non-flowering individuals that had no
170 herbivores and no leaf damage. An *R. obtusifolius* plant was defined as “Solitary” when
171 there were no conspecific individuals within 30 cm from the edge of the widest rosette
172 ($N = 15$), and *R. obtusifolius* plants with five or more conspecific individuals within a
173 radius of 1 m from the centre of the plant were defined as “Aggregated” ($N = 25$). The
174 widest rosettes of these plants were about 30 cm in diameter. We selected the youngest,
175 fully expanded leaves. These leaves were analysed for secondary metabolites, namely
176 the contents of total phenolics and condensed tannin, which are well known as major
177 secondary metabolites in the *Rumex* genus (Feduraev et al., 2019). We measured the leaf
178 contents of total phenolics and condensed tannins in accordance with the methods of
179 Feeny (1970) and Dudt and Shure (1994).

180 ***Leaf beetle choice experiment using leaf sections from naturally growing plants***

181 In April 2018, Solitary and Aggregated *R. obtusifolius* plants (85 individuals each) with
182 rosette diameters of about 30 cm were selected at random in Hirosaki City. We collected
183 the youngest fully expanded leaves from the plants. We cut one 2-cm piece from the

184 base of each collected leaf. A wet filter paper (8 cm in diameter) was placed in a
185 covered Petri dish (8.5 cm in diameter), and a piece of leaf from a Solitary plant and a
186 piece of leaf from an Aggregated plant were placed on it with one adult of *G.*
187 *atrocyanea* or *G. grisescens*. The Petri dishes were kept in a growth chamber (25 °C,
188 12L, 12D). After 24 h, the damage to each leaf piece was estimated by image analysis.
189 More details of the methods are given in the Supplementary methods.

190

191

192 **Cultivation experiments**

193 ***Cultivation design***

194 To examine the effects of the interaction environment on leaf traits and leaf beetle
195 preferences, we conducted cultivation experiments. To prepare enough samples to
196 measure leaf traits and leaf beetle preferences, two experiments were conducted.
197 Experiment 1 was conducted in 2017 to estimate the effects of interaction environment
198 on leaf secondary metabolite contents and plant biomass. Experiment 2 was conducted
199 in 2019 to estimate the effects of interaction environment on leaf primary metabolite
200 and chlorophyll contents and leaf beetle preferences.

201 In September 2016, a total of more than 700 seeds of *R. obtusifolius* were

202 collected from four individual plants in the field in Hirosaki City. Each individual was
203 separated by at least 2 km. As interspecific competitors, we focused on *Plantago*
204 *asiatica* L., *Trifolium repens* L. and *Festuca ovina* L. These species are the dominant
205 competitors of *R. obtusifolius* in Japan (Ohsaki, 2020). A total of 100 seeds of *P.*
206 *asiatica* were collected from two individuals in the field in Aomori Prefecture. A total of
207 100 seeds of *T. repens* were collected from individuals in the field in Saga Prefecture.
208 For *F. ovina*, commercially available seeds (Kaneko Seeds Co., Gunma, Japan) were
209 used. The seeds were stored in a refrigerator at 4 °C until the experiments began. Seeds
210 from each mother plant were mixed and sown on the surface of wet sand (2 cm deep)
211 during March 2017 for Experiment 1 and during March 2019 for Experiment 2. The
212 containers were kept in a growth chamber (25 °C, 12L, 12D). All plants had developed
213 their first true leaves by the beginning of the experiment.

214 In April 2017 and 2019, to obtain the focal plants, we planted one *R.*
215 *obtusifolius* seedling in each pot (10.5 cm diameter × 9 cm high) containing seed-free
216 garden soil (Mori Sangyo Co., Hokkaido, Japan). These pots were assigned to three
217 interaction treatments: no-interaction treatment as a control (2017, $N = 49$; 2019, $N =$
218 35), intraspecific interaction treatment (2017, $N = 66$; 2019, $N = 66$) and interspecific
219 interaction treatment (2017, $N = 153$; 2019, $N = 98$). In the no-interaction treatment, to

220 provide a volume of soil similar to that used in the interaction treatment for each plant,
221 the pots were divided into halves with a plastic plate to block any below-ground
222 interaction, and one seedling of *R. obtusifolius* was planted in each half of the pot. In the
223 intraspecific interaction treatment, we planted another *R. obtusifolius* seedling beside
224 the focal plant as a competitor with no plastic plate. In the interspecific interaction
225 treatment, a seedling of another species (*P. asiatica*, 2017, $N = 58$; 2019, $N = 35$; *T.*
226 *repens*, 2017, $N = 41$; 2019, $N = 30$; *F. ovina*, 2017, $N = 54$; 2019, $N = 33$) was planted
227 next to the target *R. obtusifolius* seedling. In the interaction treatment, the distance
228 between seedlings was about 2 cm. All pots were placed randomly and maintained in
229 the growth chambers (25 °C, 12L, 12D) and watered once a day for 30 days.

230

231 ***Measurement of leaf traits in cultivated plants***

232 Experiment 1

233 After 30 days, we analysed total phenolics and condensed tannins. Plants were
234 harvested and dried at 50 °C for 3 days. The plants were then weighed on an electronic
235 balance to the nearest 0.1 mg. The leaves were used to analyse total phenolics and
236 condensed tannins by using the methods in field survey.

237

238 Experiment 2

239 After 30 days, we measured chlorophyll content and five organic acids as plant nutrients
240 (see details in Supplementary methods). The chlorophyll content reflects the plant's
241 nitrogen concentration and has been found to indirectly affect herbivore survival and
242 distribution (Scheirs & De Bruyn, 2004, Sousa-Souto et al., 2018). Also, organic acids
243 in the plant are necessary for the optimal development of phytophagous insects (Offor,
244 2010). Therefore, by measuring these, we examined changes in nutrient condition in
245 response to interactions between plants.

246

247 ***Leaf beetle choice experiment***

248 To reveal whether changes in leaf chemical contents induced in *R. obtusifolius* by
249 interaction influenced the preferences of leaf beetles, we conducted choice experiments
250 with the *R. obtusifolius* leaves used in cultivation experiment 2. The combinations of
251 leaf pairs were as follows: intraspecific interaction versus interspecific interaction;
252 interspecific interaction versus no-interaction treatment; and no-interaction treatment
253 versus intraspecific interaction. The experimental design and conditions were similar to
254 that described for the choice experiment using field leaves (see Supplementary
255 methods).

256

257 **Mesocosm experiments**

258 To determine the effects of the interaction environment of *R. obtusifolius* on the
259 distribution of *G. atrocyanea*, we conducted a mesocosm experiment in November 2019
260 and July 2020. In November 2019, we estimated the effects of intraspecific interaction
261 of *R. obtusifolius* plants on the distribution of leaf beetles by using plants of the same
262 patch size in a “one-to-one-pot experiment” (Figure 1). In July 2020, we conducted a
263 “one-to-three-pot experiment” to clarify the effects of intraspecific interaction of *R.*
264 *obtusifolius* on the distribution of leaf beetles by adding the effects of patch size (i.e.,
265 resource amount) of the host plants.

266 We used two types of pot, namely interaction-treatment pots and no-
267 interaction-treatment pots. In both types of treatment pot, two seedlings of *R.*
268 *obtusifolius* were planted. The no-interaction-treatment pot was divided in half by a
269 plastic plate to block below-ground interaction between *R. obtusifolius* plants. The
270 interaction-treatment pot allowed below-ground interaction between *R. obtusifolius*
271 plants, because this type of pot had no plastic plate (see Supplementary methods).

272 ***One-to-one-pot experiment***

273 We prepared 20 containers (911 mm × 602 mm × 207 mm). In each container, an

274 interaction-treatment pot and a no-interaction-treatment pot were placed 30 cm away
275 from the edge of the pot. The containers were surrounded by soil to a depth of 15 cm to
276 allow the beetles free access to the pods, as they would have in the field (Figure 1 a, b).
277 For data analysis, each container was allocated an ID. Five *G. atrocyanea* females were
278 released on the soil in the centre of each container, the top of which was then covered
279 with 1-mm-mesh white cloth. The containers were placed in a greenhouse (15 °C), and
280 the numbers of beetles on the plants were counted after 24 h.

281 ***One-to-three-pot experiment***

282 In this experiment, we set up two types of conditions, namely “quantity conditions” (25
283 containers) and “quantity + quality conditions” (24 containers). For the quantity
284 conditions, we set up patches of two sizes by using four no-interaction-treatment pots.
285 We placed three pots together to represent large patches and one pot by itself to
286 represent small patches (Figure 1c). For the quantity + quality conditions, we set up
287 patches of two sizes by using one no-interaction-treatment pot and three interaction-
288 treatment pots; the three interaction-treatment pots represented large patches and the
289 single no-interaction-treatment pot represented small patches. In all containers, pots
290 were set up as in the one-to-one-pot experiment and under the same controlled
291 conditions. For the quantity conditions 125 beetles were used, and 120 beetles were

292 used for the quantity + quality conditions.

293

294 **Statistical analysis**

295 All statistical analyses were performed by using R v.3.6.1 software (R Development

296 Core Team, 2019). All data met the statistical assumptions of normality and

297 homoscedasticity according to the Kolmogorov–Smirnov test and F-test, and statistical

298 analyses were performed appropriately depending on the data set structure. All tests

299 were two tailed. The significance level was set at 0.05.

300

301 **Field survey data analysis**

302 ***Survey of herbivore distribution on R. obtusifolius***

303 We analysed the effects of the local population density of *R. obtusifolius* on the

304 distribution of leaf beetles by using generalised linear mixed models (GLMMs) with a

305 binomial distribution and logit function, followed by the Chi-square test. The models

306 included presence or absence of leaf beetles as fixed terms and overlap ratio of *R.*

307 *obtusifolius* rosettes for each plant, species of leaf beetles, and their interaction as

308 explanatory variables. When the relationship between overlap ratio of rosette area and

309 presence or absence of leaf beetles differed between leaf beetle species, the relationship

310 between these was analysed for each beetle species. Site ID was included as a random
311 effect in these models. False discovery rate (FDR) correction for multiple comparisons
312 was then applied.

313 ***Measurement of leaf traits in field plants***

314 Leaf chemical traits (content of condensed tannin or total phenolics) were compared
315 between Solitary and Aggregate plants by using GLMMs with Gaussian distribution and
316 an identity link, followed by an F-test; the models included leaf chemical traits as fixed
317 terms and plant density (Solitary or Aggregated) as an explanatory variable. Site ID was
318 included as a random effect in the models. FDR correction for multiple comparisons
319 was then applied.

320

321 ***Leaf beetle choice experiment using leaf sections from naturally growing plants***

322 Consumed areas of leaves were compared between local *R. obtusifolius* population
323 densities for each leaf beetle species. We used GLMMs with Gaussian distribution and
324 an identity link, followed by an F-test; the models included area consumed by leaf
325 beetles as a fixed term and plant density (Solitary or Aggregated) as an explanatory
326 variable. Petri dish ID was included as a random effect in the models. FDR correction
327 for multiple comparisons was then applied.

328

329 **Cultivation experiments data analysis**

330 *Measurement of leaf traits in cultivated plants*

331 We used Gamma distributions for the dry weights of plants, Gaussian distributions for
332 the chlorophyll content of leaves, and Poisson distributions for the leaf contents of
333 condensed tannin and total phenolics. We compared plant dry weights and leaf traits
334 (condensed tannin, total phenolics and chlorophyll content) between the cultivation
335 treatments by using GLMMs. Gamma or Poisson distributions with a log link followed
336 by a Chi-square test were applied, and Gaussian distributions with an identifying link
337 followed by an F-test were applied. These models included each plant trait as fixed
338 terms and interaction treatment (no-, intraspecific or interspecific interaction treatment)
339 as an explanatory variable. Parent plant ID was included as a random effect in the
340 models. When there was an interaction effect between each plant trait and interaction
341 treatment, we conducted multiple comparisons by FDR correction.

342 Organic acids were analysed by using a principal component analysis (PCA)
343 based on the correlation matrix of variables. Scores on the first (PC1) and second (PC2)
344 axes of the PCA were compared between interaction treatments by using GLMMs with
345 a Gaussian distribution and an identity link, followed by an F-test. The models included

346 PC1 or PC2 as fixed terms, interaction treatment (no-, intraspecific or interspecific
347 interaction treatment) as an explanatory variable and parent plant ID as a random effect.
348 When there was an interaction between PC1 or PC2 and interaction treatments, we
349 conducted multiple comparison by using FDR correction.

350 *Leaf beetle choice experiment*

351 The leaf area consumed by the leaf beetles was compared between interaction
352 treatments (no-, intraspecific or interspecific interaction). Data sets for female beetles
353 were analysed by using GLMMs with a Gamma distribution and a log link, followed by
354 a Chi-square test, and data sets of male beetles were analysed by using the Wilcoxon
355 signed-rank test because the data sets contained some 0 values. The analysis was
356 conducted for each species of leaf beetle and for each sex of each species. FDR
357 correction for multiple comparisons was then applied to each data set.

358

359 **Mesocosm experiments data analysis**

360 *One-to-one-pot experiment*

361 The numbers of leaf beetles per patch were compared between cultivation treatments by
362 using GLMMs with Gaussian distributions and an identity link, followed by an F-test;
363 the models included number of leaf beetles on the patch as a fixed term and interaction

364 treatment (interaction or no- interaction) as an explanatory variable. Container ID was
365 included as a random effect in the models.

366 ***One-to-three-pot experiment***

367 Number of leaf beetles per patch or number of leaf beetles per pot (representing leaf
368 beetle density) was compared between patch sizes (quantity) and cultivation conditions
369 (quality) by using GLMMs with Poisson distributions and a log-link, followed by a Chi-
370 square test; the models included number of leaf beetles per patch or per pot as fixed
371 terms and patch size (small or large), cultivation conditions (quantity or quantity +
372 quality condition) and their interaction as an explanatory variable. Container ID was
373 included as a random effect in these models. When there was an interaction between
374 patch size and cultivation conditions, we conducted multiple comparisons by FDR
375 correction.

376

377

378 **RESULTS**

379 **Field survey**

380 More than 60 *R. obtusifolius* individuals were growing within each quadrat; the major
381 herbivores were *G. atrocyanea* and *G. grisescens* (Table 1). The relationship between

382 the overlap ratio of *R. obtusifolius* rosettes and the presence of leaf beetles differed
383 among leaf beetle species ($\chi^2 = 81.032$, $df = 2$, $P < 0.001$). There was a significant
384 positive correlation between the overlap ratio of *R. obtusifolius* rosettes and the
385 presence of *G. atrocyanea* (estimate coefficient = 0.786, $\chi^2 = 12.764$, $df = 1$, $P < 0.001$).
386 In contrast, the presence of *G. grisescens* was not significantly correlated with the
387 overlap ratio of *R. obtusifolius* rosettes (estimate coefficient = -0.456, $\chi^2 = 2.451$, $df = 1$,
388 $P = 0.117$).

389 Contents of total phenolics and condensed tannin tended to be higher in
390 Aggregated plants than in Solitary plants, but this relationship did not reach statistical
391 significance (total phenolics, $F = 3.910$, $P = 0.096$, Figure 2a; condensed tannin, F
392 $= 4.882$, $P = 0.067$, Figure 2b). Females of *G. atrocyanea* consumed significantly more
393 leaf tissue from Aggregated plants than from Solitary plants ($F = 7.837$, $P = 0.037$,
394 Figure 3a). In contrast, for males of *G. atrocyanea* ($F = 1.779$, $P = 0.323$, Figure 3b)
395 and for both sexes of *G. grisescens* (female, $F = 1.421$, $P = 0.323$, Figure 3c; male, $F =$
396 0.494 , $P = 0.490$, Figure 3d), there were no differences in the area of feeding damage
397 between Aggregated and Solitary leaves.

398

399

400 **Cultivation experiments**

401 The biomass and chlorophyll content of *R. obtusifolius* did not differ among interaction
402 treatments (biomass: $x^2 = 7.081$, $df = 4$, $P = 0.132$, Figure 4a; chlorophyll: $F = 1.444$, P
403 $= 0.239$, Figure 4b). The contents of total phenolics of *R. obtusifolius* differed
404 significantly among treatments; they were higher in the order of intraspecific, no-, and
405 interspecific interaction treatment (Figure 4c). Plants subjected to the intraspecific
406 interaction treatment had a significantly higher content of condensed tannins than those
407 undergoing the no- or interspecific interaction treatments (Figure 4d).

408 We found that PC1 and PC2 explained 61.0% and 20.7%, respectively, of the
409 total variance of the organic acid composition data. The PC1 value did not differ among
410 interaction treatments ($F = 2.068$, $P = 0.154$, Figure S3a). Plants subjected to the no-
411 interaction treatment had significantly lower PC2 values than those undergoing the
412 intraspecific or interspecific interaction treatments (Figure S3b).

413 Females of *G. atrocyanea* consumed significantly more leaf tissue from the
414 intraspecific interaction treatment plants than from the no-interaction plants ($x^2 = 5.470$,
415 $df = 1$, $P = 0.029$, Figure 5a) and from the interspecific interaction plants ($x^2 = 6.064$, df
416 $= 1$, $P = 0.029$, Figure 5b). There was no significant difference between the no
417 interaction and interspecific interaction treatments in terms of the area of leaf eaten by

418 females of *G. atrocyanea* ($x^2 = 1.832$, $df = 1$, $P = 0.176$, Figure 5c). For males of *G.*
419 *atrocyanea*, there were no differences between treatments in the area of leaf consumed
420 (no-interaction versus intraspecific interaction, $z = 0.329$, $P = 1$, Figure 5d; interspecific
421 interaction versus intraspecific interaction, $z = 2.139$, $P = 1$, Figure 5e; no-interaction
422 versus interspecific interaction, $z = 0$, $P = 0.097$, Figure 5f).

423

424 **Mesocosm experiments**

425 In the one-to-one-pot experiment, a significantly greater number of *G. atrocyanea* were
426 distributed on the *R. obtusifolius* plants in the interaction treatment than in the no-
427 interaction treatment ($F = 5.556$, $P = 0.030$, Figure 6a). In the one-to-three-pot
428 experiment, the effect of patch size on the distribution of *G. atrocyanea* differed
429 significantly with the cultivation conditions ($x^2 = 6.540$, $df = 2$, $P = 0.038$, Figure 6b).
430 Under both types of cultivation condition, large patches had significantly more beetles
431 than small patches (quantity conditions, $x^2 = 18.301$, $df = 1$, $P < 0.001$; quantity + quality
432 conditions, $x^2 = 55.474$, $df = 1$, $P < 0.001$, Figure 6b). This trend was more pronounced
433 under quantity + quality conditions. Moreover, the effect of patch size on the number of
434 *G. atrocyanea* per pot (i.e., the leaf beetle density) differed significantly between
435 cultivation conditions (interaction treatment \times patch size; $z = -2.067$, $P = 0.039$, Figure

436 6c). Although the densities of leaf beetles in small and large patches were similar under
437 quantity conditions ($z = -0.308$, $P = 0.758$), under quantity + quality conditions the
438 large patches had a greater density of leaf beetles than small patches ($z = 2.118$, $P =$
439 0.034 ; Figure 6c).

440

441 **DISCUSSION**

442 We found here that intraspecific interaction induced changes in the leaf metabolite
443 contents of *Rumex obtusifolius* and affected resource utilisation by the specialist leaf
444 beetle, *G. atrocyanea*, but not by the generalist leaf beetle, *G. grisea*. In addition,
445 we showed experimentally that this type of resource utilisation affected the distribution
446 of *G. atrocyanea*. These results support our hypothesis, providing experimental
447 evidence that differences in the local population density of the host plant led to plastic
448 changes in leaf metabolite contents, affecting the resource utilisation and distribution
449 patterns of specialist herbivores.

450 ***Variations in leaf traits***

451 In the field, Aggregated *R. obtusifolius* plants tended to have higher contents of total
452 phenolics and condensed tannin than Solitary plants (Figure 2). This result suggests that
453 aggregation of *R. obtusifolius* plants induced changes in leaf chemical traits. In fact, in

454 the cultivation experiments, the contents of total phenolics and condensed tannin in the
455 leaves of *R. obtusifolius* were significantly higher under intraspecific interaction
456 conditions than under interspecific interaction or no-interaction conditions (Figure 4c,
457 d). Increased contents of secondary metabolites in the presence of a conspecific
458 neighbour have been reported in several plant species, and it has been suggested that
459 metabolic alterations in leaves in response to intraspecific interaction are common in
460 plants (Barton & Bowers, 2006; Ormeño et al., 2007; Broz et al., 2010, but see Kigathi
461 et al., 2013). In many plant species intraspecific competition is more intense than
462 interspecific competition (Adler et al., 2018), and such intraspecific competition causes
463 limitation of soil nutrients and water (Craine & Dybzinski, 2013; Takigahira &
464 Yamawo, 2019). It is well known that limitation of soil nutrients and water for plants
465 induces the accumulation of secondary metabolites in the leaves (reviewed in Akula &
466 Ravishankar, 2011). Thus, aggregation of *R. obtusifolius* plants may increase the leaf
467 contents of secondary metabolites, such as total phenolics and condensed tannin,
468 through soil resource competition.

469 Another possible hypothesis is that Aggregated plants invest more in defence
470 than do Solitary plants through recognition of con-specific neighbours, because
471 aggregated plants often consumed by specialist leaf beetles. Leaf-trait alteration based

472 on neighbour recognition has also been reported in several plant species (Yamawo &
473 Mukai 2020; Yamawo, 2015, 2021). Neighbour recognition can therefore be a cause of
474 leaf-trait alteration in *R. obtusifolius* plants. However, the history of interaction between
475 specialist leaf beetles and *R. obtusifolius* plants is weak, because *R. obtusifolius* is an
476 exotic species in Japan. To understand the adaptive importance of leaf-trait alteration in
477 *R. obtusifolius* plants, we would need to perform an additional study in a region to
478 which *R. obtusifolius* is native.

479 The contents of primary metabolites are strongly affected by light conditions
480 (Kitazaki et al., 2018). For example, experiments with lettuce, *Lactuca sativa*, have
481 shown that the pattern of accumulation of primary metabolites, such as sugars and
482 amino acids, is affected by light quality, intensity and exposure time (Kitazaki et al.,
483 2018). In fact, our cultivation experiment found differences in the content of primary
484 metabolites between no-interaction and interaction treatments (Figure S3b). Changes in
485 the contents of primary metabolites in leaves may depends on presence of neighbour
486 plants, regardless of the identity of neighbour.

487 ***Preferences and distribution of leaf beetles***

488 The local *R. obtusifolius* population density affected the amounts of leaf consumed by
489 the specialist leaf beetle, *G. atrocyanea*. In the experiment using leaves from the field,

490 females of *G. atrocyanea* preferred to consume the leaves of aggregated *R. obtusifolius*
491 plants than of Solitary plants, despite similar quantities of leaves being provided for the
492 beetles (Figure 3a). In the experiment using the leaves of cultivated plants, females of
493 *G. atrocyanea* also preferred the leaves of *R. obtusifolius* plants exposed to intraspecific
494 interaction over those of plants exposed to no interaction (Figure 5a). These preference
495 pattern are consistent with the increases in the leaf contents of secondary metabolites
496 (total phenolics and condensed tannins) (Figures 2 and 4c, d) but not with the variations
497 in primary metabolites (Figures S3). Therefore, we concluded that females of *G.*
498 *atrocyanea* selected leaves on the basis of increases induced in the leaf secondary
499 metabolite content by the host plant's interactive environment. *G. atrocyanea* beetles are
500 specialist herbivores of *Rumex* plants (Suzuki, 1985). Many herbivore specialists use
501 host-specific secondary metabolites for host searching or detecting (Schoonhoven et al.,
502 2005; Ômura, 2018). This type of host searching may reflect the feeding preferences of
503 *G. atrocyanea*. Females of *G. atrocyanea* lay eggs on the plants on which they feed, and
504 the hatched larvae feed on the same plants. The larvae of *G. atrocyanea* require large
505 amounts of food, and plants are often completely consumed (Suzuki, 1985). For this
506 reason, the selection of aggregated plant leaves by *G. atrocyanea* females during the
507 reproductive season is linked to the securing of food resources for the next generation.

508 This may be associated with niche specialisation in coevolution among host plants and
509 specialist herbivores (Schoonhoven et al. 2005; Abrahamson, 2008). In contrast, no
510 preference was observed among males of *G. atrocyanea*, possibly because males use
511 fewer resources than females with egg masses.

512 Do the preferences of leaf beetles affect the beetles' distribution? Our
513 mesocosm experiment provided robust evidence that changes in leaf traits based on
514 intraspecific interaction can affect the distribution of the specialist leaf beetles, *G.*
515 *atrocyanea* (Figure 6). When the plant patch sizes were similar, approximately 1.7 times
516 more leaf beetles were distributed in the interaction treatment patch than in the no-
517 interaction treatment patch (Figure 6a). Effects of interaction between host plants were
518 also found in the one-to-three-pot experiment. Greater numbers of leaf beetles were
519 distributed on the large patches than on the small patches, and this trend was more
520 pronounced under quantity + quality conditions than under quantity conditions (Figure
521 6b). This finding is consistent with the distribution of *G. atrocyanea* in the field
522 (Suzuki, 1985).

523 Moreover, in the conditions under which both the patch size and the
524 competitive environment (and thereby leaf traits) differed, the leaf beetle density was
525 significantly higher on large patches than on small ones, but it did not differ in

526 conditions under which only the patch size differed (Figure 6c). Root (1973) predicted
527 that “Herbivores are more densely distributed as the patch size of the feeding site
528 increases (resource concentration hypothesis).” According to this prediction, the
529 response of *G. atrocyanea* to differences in leaf traits leads to the concentration of
530 beetles relative to the food resource. In other words, in this system, our results strongly
531 suggested that differences in metabolic alterations in leaves through intraspecific
532 interactions in plants induced the concentrated distribution of herbivores on resources.

533 In contrast, numbers of the generalist leaf beetle, *G. grisea*, were not
534 correlated with the local population density of *R. obtusifolius* in the field, and these
535 beetles did not select the leaves of *R. obtusifolius* on the basis of the interaction
536 environment. These results did not support our hypothesis that generalist herbivores
537 accumulate on low-density host plants to avoid high levels of secondary metabolites.
538 Generalist herbivores respond to a variety of chemicals besides those measured as leaf
539 traits in this study (Schoonhoven et al., 2005; War et al., 2012). Perhaps other leaf traits,
540 which could not be measured here, may have been involved in the preferences of *G.*
541 *grisea* and varied according to the interaction environment, thus masking the
542 avoidance effect of secondary metabolites on the generalist leaf beetles. Another
543 possible reason why the findings did not support our hypothesis is the effects of

544 resource competition among herbivores. In some cases, resource competition among
545 herbivores influences herbivore distribution (e.g., Suzuki 1986; Schoonhoven et al.,
546 2005; Godinho et al., 2020). A previous study pointed out that *G. grisea* is
547 vulnerable to resource competition from *G. atrocyanea* (Suzuki, 1986). It may therefore
548 prioritise the avoidance of competitors over plant availability when deciding where to
549 feed (Suzuki, 1985). Several studies, as well as the resource dilution hypothesis
550 proposed by Otway et al. (2005), have pointed out that herbivore density per plant may
551 be higher when the host density is low (e.g. Yamamura, 1999). Our results suggest that
552 these phenomena may be caused not only by differences in the local population density
553 of the host plants but also indirectly by interactions with other herbivorous insects. To
554 determine whether these results are general or specific to certain herbivores, several
555 species, including generalists, may need to be tested.

556

557

558 **CONCLUSIONS**

559 Our findings provide experimental evidence that intraspecific interaction between host
560 plants affects specialist herbivore distribution. Many researchers have worked to
561 unravel the relationship between the distribution of herbivores and the local population
562 density of host plants. Some herbivores have shown a positive response to resource

563 abundance, as in the resource concentration hypothesis proposed by Root (1973),
564 whereas others, as in the resource dilution hypothesis proposed by Otway et al. (2005),
565 have shown a negative response. These studies have focused on the amount of food
566 available and have assumed that leaf traits are always constant. Our results indicate that
567 herbivore responses to resource quantity and quality may interact with each other as
568 factors governing herbivore distribution. Therefore, herbivore responses to the local
569 population density of host plants can be understood from a plant–plant interaction
570 perspective, highlighting the need to integrate plant–plant interactions into our
571 understanding of plant–animal interactions in nature.

572

573 **ACKNOWLEDGEMENTS**

574 This work was supported by JSPS Grants-in-Aid for Scientists (grant no. 18K19353 and
575 19H03295 to AY) from the Japan Society for the Promotion of Science and a Sasakawa
576 Scientific Research Grant (Study Number: 2019 - 5023).

577

578 **REFERENCES**

579 Abrahamson, W. G. (2008). *Specialization, speciation, and radiation: the evolutionary*
580 *biology of herbivorous insects*. Univ of California Press, Berkeley, CA.

- 581 Adler, P. B., Smull, D., Beard, K. H., Choi, R. T., Furniss, T., Kulmatiski, A., Meiners, J.
582 M., Tredennick, A. T., & Veblen, K. E. (2018). Competition and coexistence
583 in plant communities: intraspecific competition is stronger than interspecific
584 competition. *Ecology letters*, *21*, 1319-1329. <https://doi.org/10.1111/ele.13098>
- 585 Akula, R., & Ravishankar, G. A. (2011). Influence of abiotic stress signals on secondary
586 metabolites in plants. *Plant Signaling & Behavior*, *6*, 1720-1731.
587 <https://doi.org/10.4161/psb.6.11.17613>
- 588 Barton, K. E., & Bowers, M. D. (2006). Neighbor species differentially alter resistance
589 phenotypes in *Plantago*. *Oecologia*, *150*, 442-452.
590 <https://doi.org/10.1007/s00442-006-0531-z>
- 591 Broz, A. K., Broeckling, C. D., De-la-Peña, C., Lewis, M. R., Greene, E., Callaway, R.
592 M., Sumner, L. W., & Vivanco, J. M. (2010). Plant neighbor identity
593 influences plant biochemistry and physiology related to defense. *BMC Plant*
594 *Biology*, *10*, 1-14. <https://doi.org/10.1186/1471-2229-10-115>
- 595 Coutinho, R. D., Cuevas-Reyes, P., Fernandes, G. W., & Fagundes, M. (2019).
596 Community structure of gall-inducing insects associated with a tropical shrub:
597 regional, local and individual patterns. *Tropical Ecology*, *60*, 74-82.
598 <https://doi.org/10.1007/s42965-019-00010-7>

- 599 Craine, J. M., & Dybzinski, R. (2013). Mechanisms of plant competition for nutrients,
600 water and light. *Functional Ecology*, 27, 833-840.
601 <https://doi.org/10.1111/1365-2435.12081>
- 602 Dudt, J. F., & Shure, D. J. (1994). The influence of light and nutrients on foliar
603 phenolics and insect herbivory. *Ecology*, 75, 86-98.
604 <https://doi.org/10.2307/1939385>
- 605 Fagundes, M., Barbosa, E. M., Oliveira, J. B., Brito, B. G., Freitas, K. T., Freitas, K. F.,
606 & Reis-Junior, R. (2019). Galling inducing Insects associated with a tropical
607 shrub: the role of resource concentration and species interactions. *Ecología*
608 *Austral*, 29, 012-019. <https://doi.org/10.25260/EA.19.29.1.0.751>
- 609 Feduraev, P., Chupakhina, G., Maslennikov, P., Tacenko, N., & Skrypnik, L. (2019).
610 Variation in phenolic compounds content and antioxidant activity of different
611 plant organs from *Rumex crispus* L. and *Rumex obtusifolius* L. at different
612 growth stages. *Antioxidants*, 8, 237. <https://doi.org/10.3390/antiox8070237>
- 613 Feeny, P. (1970). Seasonal changes in oak leaf tannins and nutrients as a cause of spring
614 feeding by winter moth caterpillars. *Ecology*, 51, 565-581.
615 <https://doi.org/10.2307/1934037>
- 616 Fretwell, D. S., & Lucas, H. L. J. (1969). On territorial behavior and other factors

- 617 influencing habitat distribution in birds. *Acta Biotheoretica*, *19*, 16–36.
- 618 <https://doi.org/10.1007/BF01601>
- 619 Godinho, D. P., Janssen, A., Li, D., Cruz, C., & Magalhães, S. (2020). The distribution
620 of herbivores between leaves matches their performance only in the absence
621 of competitors. *Ecology and Evolution*, *10*, 8405-8415.
- 622 <https://doi.org/10.1002/ece3.6547>
- 623 Goodey, N. A., Florance, H. V., Smirnoff, N., & Hodgson, D. J. (2015). Aphids pick
624 their poison: selective sequestration of plant chemicals affects host plant use
625 in a specialist herbivore. *Journal of Chemical Ecology*, *41*, 956-964.
- 626 <https://doi.org/10.1007/s10886-015-0634-2>
- 627 Hambäck, P. A., & Beckerman, A. P. (2003). Herbivory and plant resource competition:
628 a review of two interacting interactions. *Oikos*, *101*, 26-37.
- 629 <https://doi.org/10.1034/j.1600-0706.2003.12568.x>
- 630 Jeschke, V., Kearney, E. E., Schramm, K., Kunert, G., Shekhov, A., Gershenson, J., &
631 Vassão, D. G. (2017). How glucosinolates affect generalist lepidopteran
632 larvae: growth, development and glucosinolate metabolism. *Frontiers in Plant*
633 *Science*, *8*, 1995. <https://doi.org/10.3389/fpls.2017.01995>
- 634 Kigathi, R. N., Weisser, W. W., Veit, D., Gershenson, J., & Unsicker, S. B. (2013).

- 635 Plants suppress their emission of volatiles when growing with conspecifics.
636 *Journal of Chemical Ecology*, 39, 537-545. [https://doi.org/10.1007/s10886-](https://doi.org/10.1007/s10886-013-0275-2)
637 013-0275-2
- 638 Kitazaki, K., Fukushima, A., Nakabayashi, R., Okazaki, Y., Kobayashi, M., Mori, T.,
639 Nishizawa, T., Reyes-Chin-Wo, S., Michelmore, R. W., Shoji, K., & Kusano,
640 M. (2018). Metabolic reprogramming in leaf lettuce grown under different
641 light quality and intensity conditions using narrow-band LEDs. *Scientific*
642 *Reports*, 8, 1-12. <https://doi.org/10.1038/s41598-018-25686-0>
- 643 Macel, M. (2011). Attract and deter: a dual role for pyrrolizidine alkaloids in plant–
644 insect interactions. *Phytochemistry Reviews*, 10, 75-82.
645 <https://doi.org/10.1007/s11101-010-9181-1>
- 646 Makuchi, T., & Sakai, H. (1984). Seedling survival and flowering of *Rumex*
647 *obtusifolius* L. in various habitats. *Weed Research*, 29, 123–130.
648 <https://doi.org/10.3719/weed.29.123>
- 649 Mraja, A., Unsicker, S. B., Reichelt, M., Gershenson, J., & Roscher, C. (2011). Plant
650 community diversity influences allocation to direct chemical defence in
651 *Plantago lanceolata*. *PLoS One*, 6, e28055.
652 <https://doi.org/10.1371/journal.pone.0028055>

- 653 Muiruri, E. W., Barantal, S., Iason, G. R., Salminen, J. P., Perez - Fernandez, E., &
654 Koricheva, J. (2019). Forest diversity effects on insect herbivores: do leaf
655 traits matter?. *New Phytologist*, *221*, 2250-2260.
656 <https://doi.org/10.1111/nph.15558>
- 657 Nerlekar, A. N. (2018). Seasonally dependent relationship between insect herbivores
658 and host plant density in *Jatropha nana*, a tropical perennial herb. *Biology*
659 *Open* *7*, 1–7. <https://doi.org/10.1242/bio.035071>
- 660 Offor, E. (2010). The nutritional requirements of phytophagous insects: why do insects
661 feed on plants? *Available at SSRN*, 1535274.
- 662 Ohsaki, H., Mukai, H., & Yamowo, A. (2020). Biochemical recognition in seeds:
663 Germination of *Rumex obtusifolius* is promoted by leaves of facilitative adult
664 conspecifics. *Plant Species Biology*, *35*, 233-242.
665 <https://doi.org/10.1111/1442-1984.12275>
- 666 Ômura, H. (2018). Plant secondary metabolites in host selection of butterfly. *Chemical*
667 *Ecology of Insects* (ed. by J. Tabata), pp. 3–27. CRC Press, Boca Raton,
668 Florida.
- 669 Ormeño, E., Bousquet-Mélou, A., Mévy, J. P., Greff, S., Robles, C., Bonin, G., &
670 Fernandez, C. (2007). Effect of intraspecific competition and substrate type

- 671 on terpene emissions from some Mediterranean plant species. *Journal of*
672 *Chemical Ecology*, 33, 277-286. <https://doi.org/10.1007/s10886-006-9219-4>
- 673 Otway, S. J., Hector, A., & Lawton, J. H. (2005). Resource dilution effects on specialist
674 insect herbivores in a grassland biodiversity experiment. *Journal of Animal*
675 *Ecology*, 74, 234-240. <https://doi.org/10.1111/j.1365-2656.2005.00913.x>
- 676 R Development Core Team. (2019). R: A language and environment for statistical
677 computing. Vienna, Austria: R Foundation for Statistical Computing.
- 678 Rhainds, M., & English - Loeb, G. (2003). Testing the resource concentration
679 hypothesis with tarnished plant bug on strawberry: density of hosts and patch
680 size influence the interaction between abundance of nymphs and incidence of
681 damage. *Ecological Entomology*, 28, 348-358. [https://doi.org/10.1046/j.1365-](https://doi.org/10.1046/j.1365-2311.2003.00508.x)
682 [2311.2003.00508.x](https://doi.org/10.1046/j.1365-2311.2003.00508.x)
- 683 Root, R. B. (1973). Organization of a plant - arthropod association in simple and
684 diverse habitats: the fauna of collards (*Brassica oleracea*). *Ecological*
685 *Monographs*, 43, 95-124. <https://doi.org/10.2307/1942161>
- 686 Scheirs, J., & De Bruyn, L. (2004). Excess of nutrients results in plant stress and
687 decreased grass miner performance. *Entomologia experimentalis et applicata*,
688 113, 109-116. <https://doi.org/10.1111/j.0013-8703.2004.00215.x>

- 689 Scherber, C., Eisenhauer, N., Weisser, W.W., Schmid, B., Voigt, W., Fischer, M. *et al.*
690 (2010). Bottom-up effects of plant diversity on multitrophic interactions in a
691 biodiversity experiment. *Nature*, 468, 553-556.
692 <https://doi.org/10.1038/nature09492>
- 693 Schoonhoven, L.M., van Loon, J.J.A. & Dicke, M. (2005). *Insect–Plant Biology*, 2nd
694 edn. Oxford University Press, Oxford.
- 695 Sousa-Souto, L., Bocchiglieri, A., Dias, D. D. M., Ferreira, A. S., & José Filho, P. D. L.
696 (2018). Changes in leaf chlorophyll content associated with flowering and its
697 role in the diversity of phytophagous insects in a tree species from a semiarid
698 Caatinga. *PeerJ*, 6, e5059. <https://doi.org/10.7717/peerj.5059>
- 699 Stephens, A. E., & Myers, J. H. (2012). Resource concentration by insects and
700 implications for plant populations. *Journal of Ecology*, 100, 923-931.
701 <https://doi.org/10.1111/j.1365-2656.2005.00913.x>
- 702 Suzuki, N. (1985). Resource utilization of three chrysomelid beetles feeding on *Rumex*
703 plants with diverse vegetational background. *Japanese Journal of Ecology*,
704 35, 225-234. https://doi.org/10.18960/seitai.35.2_225
- 705 Suzuki, N. (1986). Interspecific competition and coexistence of the two chrysomelids,
706 *Gastrophysa atrocyanea* Motschulsky and *Galerucella vittaticollis* Baly

- 707 (Coleoptera: Chrysomelidae), under limited food resource conditions.
- 708 *Ecological Research*, 1, 259-268. <https://doi.org/10.1007/BF02348683>
- 709 Takigahira, H., & Yamawo, A. (2019). Competitive responses based on kin-
- 710 discrimination underlie variations in leaf functional traits in Japanese beech
- 711 (*Fagus crenata*) seedlings. *Evolutionary Ecology*, 33, 521-531.
- 712 <https://doi.org/10.1007/s10682-019-09990-3>
- 713 Titayavan, M., & Altieri, M. A. (1990). Synomone-mediated interactions between the
- 714 parasitoid *Diaeretiella rapae* and *Brevicoryne brassicae* under field
- 715 conditions. *Entomophaga*, 35, 499-507. <https://doi.org/10.1007/BF02375084>
- 716 Tuller, J., Queiroz, A. C. M., Luz, G. R., & Silva, J. O. (2013). Gall-forming insect
- 717 attack patterns: a test of the Plant Vigor and the Resource Concentration
- 718 Hypotheses. *Biotemas*, 26, 45-51. [https://doi.org/10.5007/2175-](https://doi.org/10.5007/2175-7925.2013v26n1p45)
- 719 [7925.2013v26n1p45](https://doi.org/10.5007/2175-7925.2013v26n1p45)
- 720 War, A. R., Paulraj, M. G., Ahmad, T., Buhroo, A. A., Hussain, B., Ignacimuthu, S., &
- 721 Sharma, H. C. (2012). Mechanisms of plant defense against insect herbivores.
- 722 *Plant Signaling & Behavior*, 7, 1306-1320. <https://doi.org/10.4161/psb.21663>
- 723 Wheat, C. W., Vogel, H., Wittstock, U., Braby, M. F., Underwood, D., & Mitchell-Olds,
- 724 T. (2007). The genetic basis of a plant–insect coevolutionary key innovation.

- 725 *Proceedings of the National Academy of Sciences*, 104, 20427-20431.
- 726 <https://doi.org/10.1073/pnas.0706229104>
- 727 Yamamura, K. (1999). Relation between plant density and arthropod density in cabbage
728 fields. *Population Ecology*, 41, 177-182.
- 729 <https://doi.org/10.1007/s101440050020>
- 730 Yamawo A. (2015) Relatedness of neighboring plants alters the expression of indirect
731 defense traits in an extrafloral nectary-bearing plant. *Evolutionary Biology*,
732 42, 12-19. <https://doi.org/10.1007/s11692-014-9295-2>
- 733 Yamawo A. (2021). Intraspecific competition favors ant-plant protective mutualism.
734 *Plant Species Biology*. <https://doi.org/10.1111/1442-1984.12331>
- 735 Yamawo, A., & Mukai, H. (2020). Outcome of interspecific competition depends on
736 genotype of conspecific neighbours. *Oecologia*, 193, 415-423.
- 737 <https://doi.org/10.1007/s00442-020-04694-w>
- 738

739 Table 1 Herbivores of *Rumex obtusifolius* and numbers and proportions of infested plants.

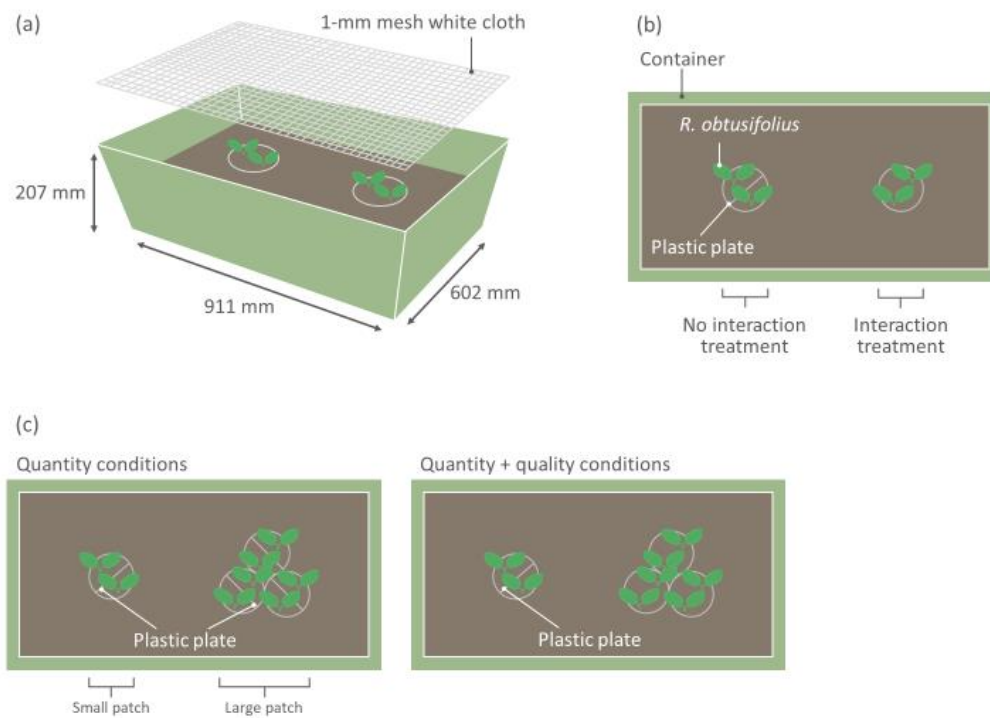
740

Taxon	Tomino-cho (<i>N</i> = 213)		Ozawa (<i>N</i> = 441)		Ohara (<i>N</i> = 112)		Iwate (<i>N</i> = 195)		Nagoya (<i>N</i> = 62)	
	No.	(%)	No.	(%)	No.	(%)	No.	(%)	No.	(%)
<i>Gastrophysa atrocyanea</i> Motschulsky (Chrysomelidae: Coleoptera)	14	6.57	195	44.22	84	75.00	49	25.13	44	22.56
<i>Galerucella grisescens</i> Joannis (Chrysomelidae: Coleoptera)	29	13.62	54	12.24	109	97.32	50	25.64		
<i>Bothrogonia ferruginea</i> Fabricius (Tettigellidae: Hemiptera)	1	0.47			1	0.89				
<i>Mantura clavareau</i> Heikertinger (Chrysomelidae: Coleoptera)	1	0.47					1	0.51		
<i>Aphis rumicis</i> Linnaeus (Aphididae: Hemiptera)							15	7.69		
Dermaptera (Insecta)					1	0.89				
Helicoidea (Pulmonata)							60	30.77		
Lepidoptera (Insecta)							4	2.05		
Unknown	1	0.47								

741

742

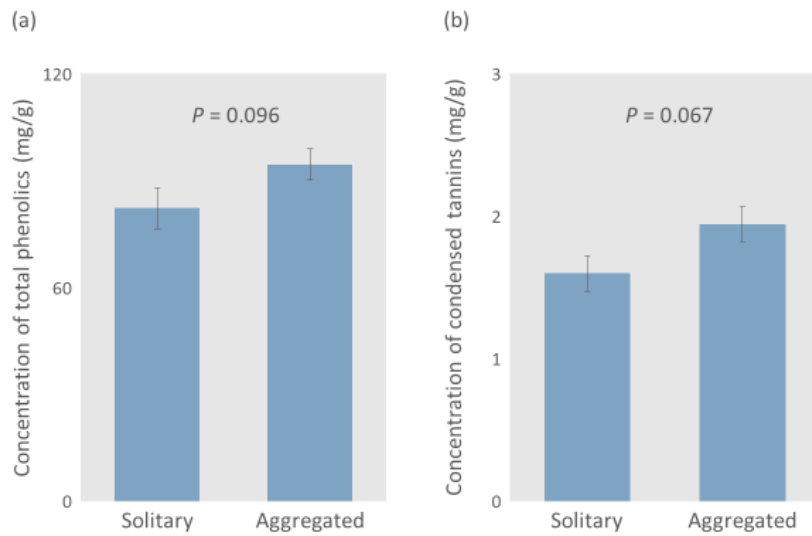
743 **FIGURES**



744

745 **Figure 1**

746 (a) Experimental setup in mesocosm experiment. In all containers, the area around the
747 pots was filled with soil to a depth of 15 cm to allow the beetles free access to the
748 plants, as in the field. (b) In the one-to-one-pot experiment, the interaction and no-
749 interaction treatment pots were placed 30 cm away from each other in the container. (c)
750 In the one-to-three-pot experiment, two sets of conditions were set up, namely “quantity
751 conditions” and “quantity + quality conditions.” Under quantity conditions, two patch
752 sizes were created by using four no-interaction pots. Under quantity + quality
753 conditions, two patch sizes were created by using one no-interaction pot and three
754 interaction pots. The distance between the large and small patches was 30 cm in each
755 container.



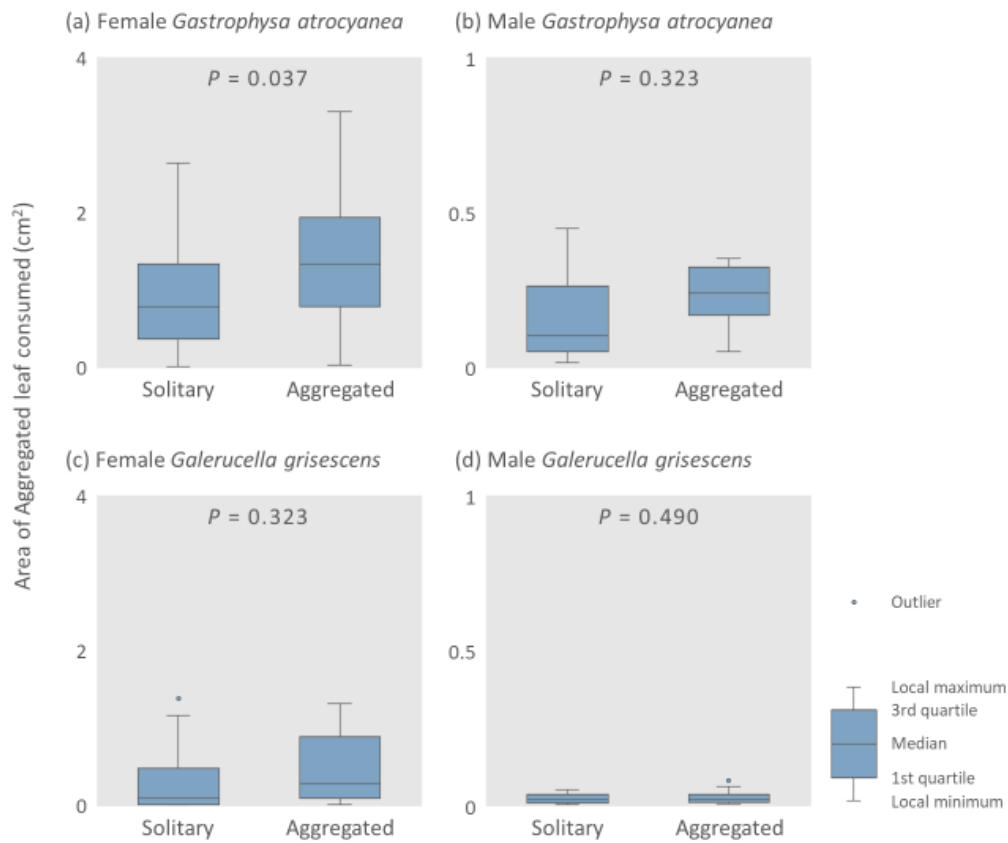
756

757 Figure 2

758 Contents of (a) total phenolics and (b) condensed tannin in Solitary ($N = 15$) and

759 Aggregated ($N = 25$) plants. Bars represent SE. P -values are for the results of GLMM

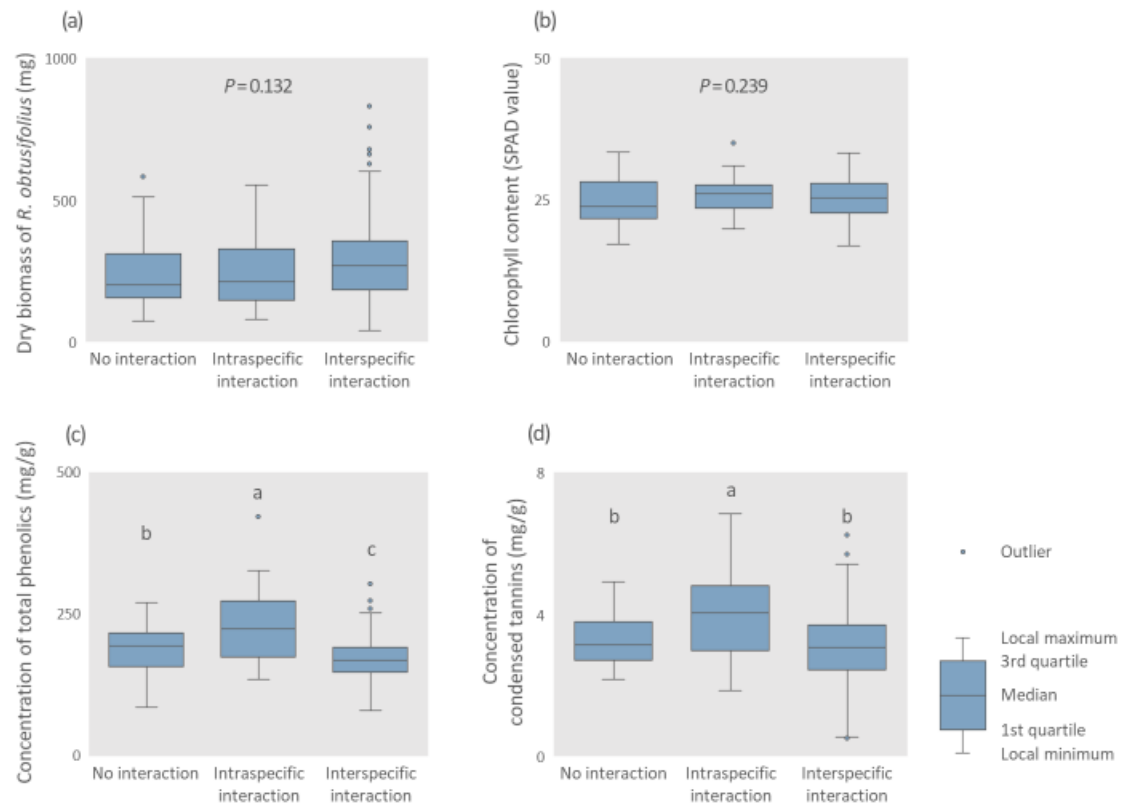
760 analysis.



761

762 Figure 3

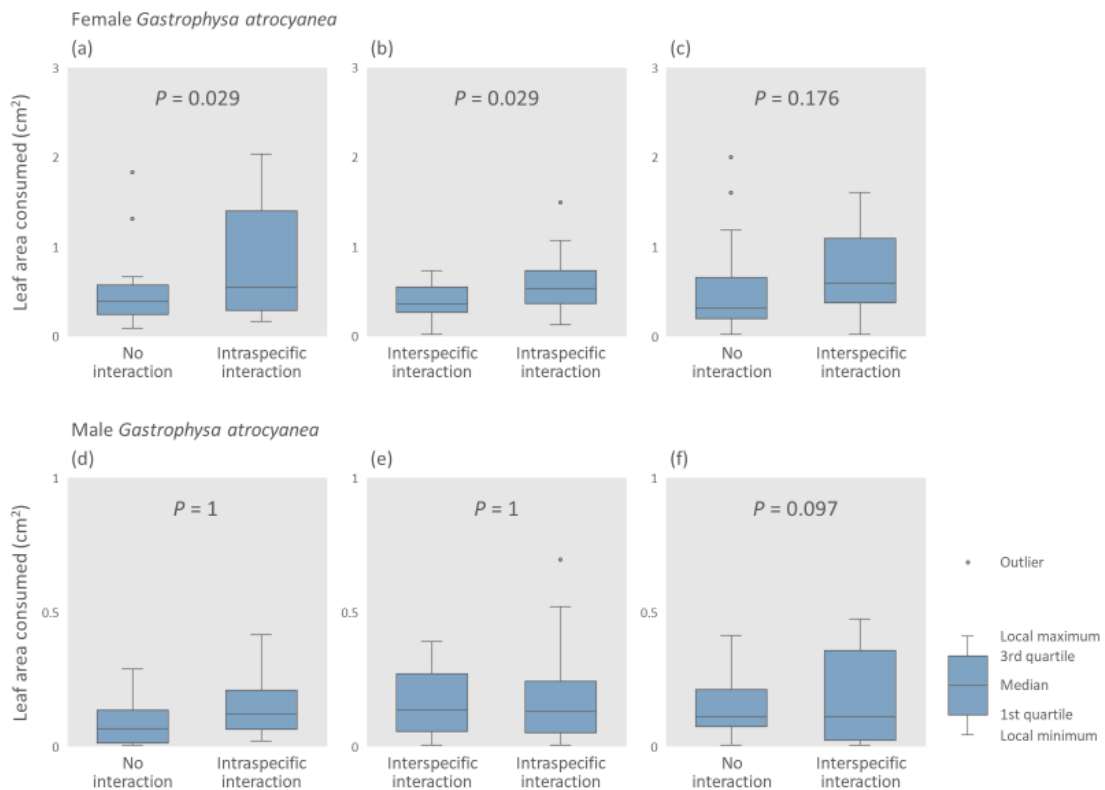
763 Areas of leaf consumed by (a) female and (b) male *Gastrophysa atrocyanea* and (c)
764 female and (d) male *Galerucella griseascens* in the choice experiment using leaves from
765 the field. *P*-values are for the results of GLMM analysis.



766

767 Figure 4

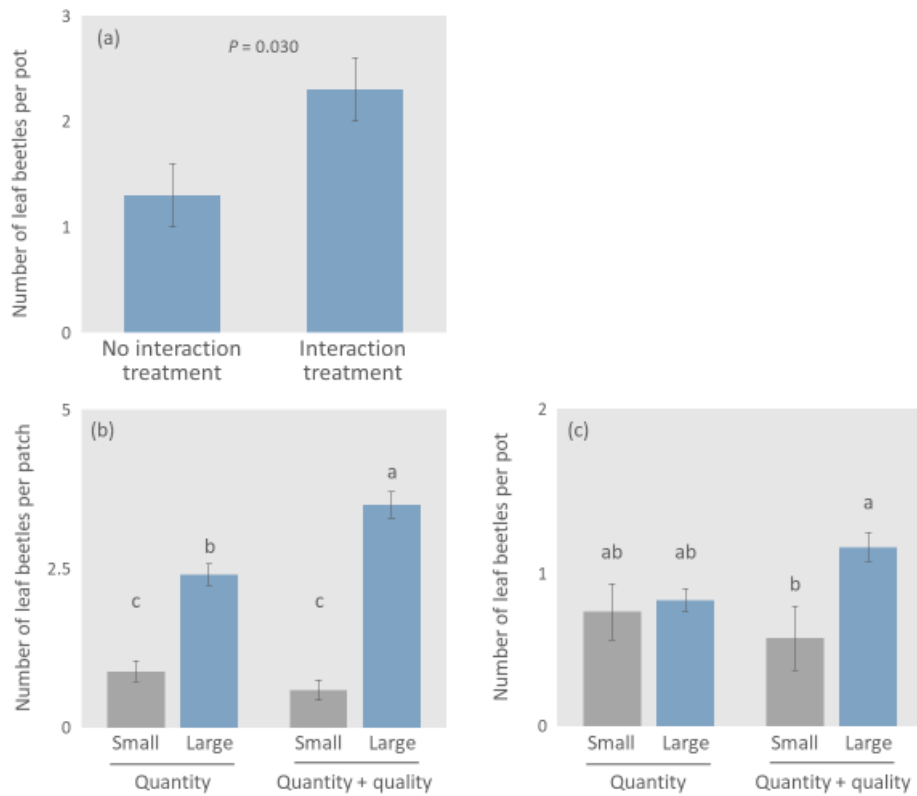
768 Dry biomass and leaf traits of *Rumex obtusifolius* in the cultivation experiment. (a) Dry
769 biomass of whole plant, (b) chlorophyll content, (c) content of total phenolics and (d)
770 content of condensed tannins in leaves. Different letters denote significant differences
771 (GLMM, $P < 0.05$).



772

773 Figure 5

774 Leaf areas consumed by (a to c) female and (d to f) male *Gastrophysa atrocyanea* in the
775 choice experiment using cultivated plant leaves. The combinations of leaf pairs of
776 treatments were as follows: (a, d) no interaction versus intraspecific interaction; (b, e)
777 interspecific interaction versus intraspecific interaction; (c, f) no interaction versus
778 interspecific interaction.



779

780 Figure 6

781 Numbers of leaf beetles in the mesocosm experiment. (a) Number of leaf beetles per pot

782 in each treatment (no interaction or interaction treatment) in the one-to-one-pot

783 experiment. (b) Number of leaf beetles per patch and (c) number of leaf beetles per pot

784 in each patch (small and large patches) under each set of conditions (quantity and

785 quantity + quality conditions) in the one-to-three-pot experiment. Bars represent SE.

786 Different letters indicate significant differences (GLMM, $P < 0.05$).

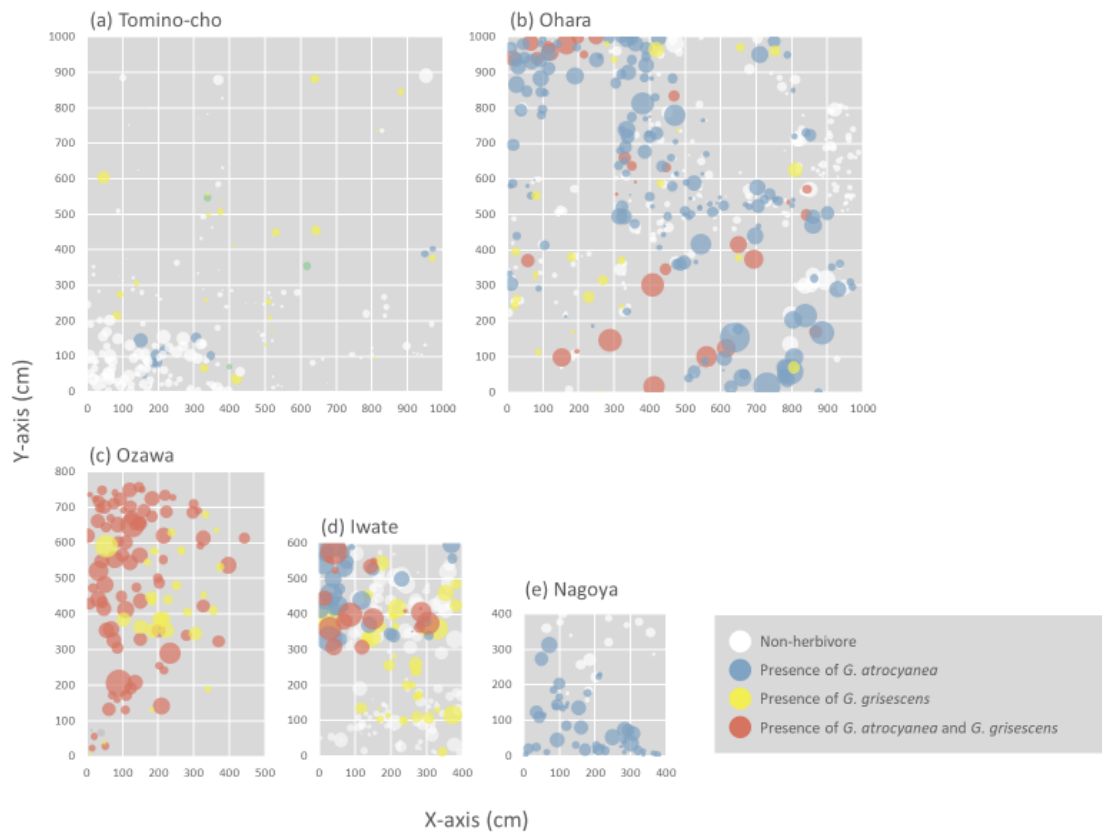


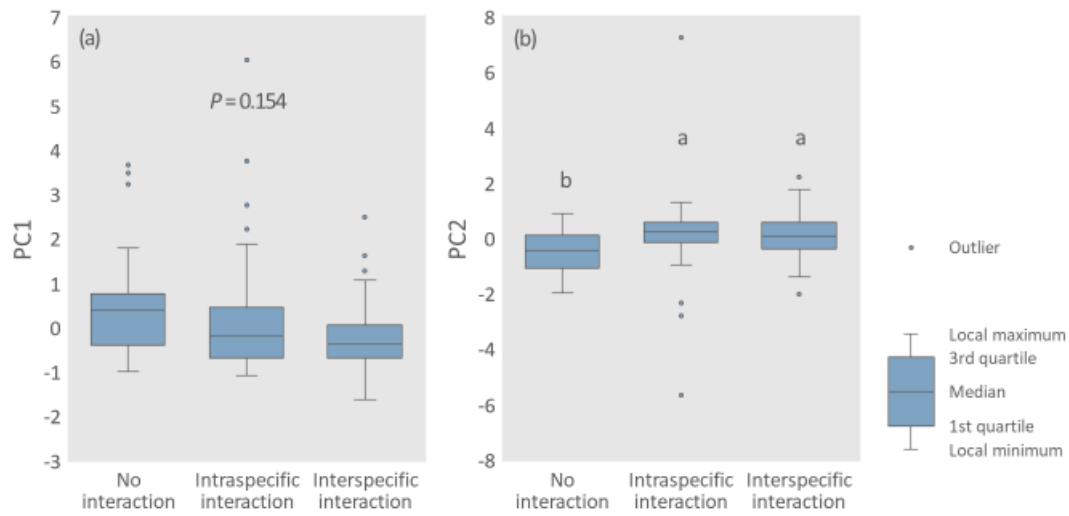
787

788 Figure S1

789 Geographic locations of the five study sites used in the field survey in Japan. These sites

790 were at least 2 km apart.





796

797 Figure S3

798 Boxplots of principal component values. (a) PC1 and (b) PC2 for organic acids in the
799 cultivation experiment. Different letters indicate significant differences (GLMM, $P <$
800 0.05).

801 **SUPPLEMENTARY METHODS**

802

803 **Study species**

804 *Rumex obtusifolius* is a perennial herb native to Europe. In the early 1900s, *R.*
805 *obtusifolius* plants were brought to Japan as contaminants of grasses imported from
806 Europe (Makuchi & Sakai, 1984), and they invaded all regions of Japan except
807 Okinawa. This species grows in wet grassland in Japan. The plants often aggregate with
808 conspecific individuals, and they form patches of various sizes within the population
809 (Van Evert et al., 2011) because their seed germination is promoted by the presence of
810 the leaves of conspecifics via water-soluble-chemical exposure (Ohsaki et al., 2020).
811 The species contains condensed tannins and phenolics as secondary metabolites in the
812 leaves (Feduraev et al., 2019).

813 *Rumex obtusifolius* leaves are consumed mainly by two leaf beetles (Table 1),
814 *Gastrophysa atrocyane* and *Galerucella grisescen*. *Gastrophysa atrocyanea* is a
815 univoltine species and specialist herbivore of *Rumex* plants. It occurs on *Rumex* plant
816 species from spring (March) to mid-summer (July) in Japan (Suzuki, 1985). *Galerucella*
817 *grisescens* is a multivoltine species and a generalist herbivore of plants of the
818 Polygonaceae family. It occurs on *Rumex* species from spring (March) to early winter
819 (December) in Japan (Suzuki, 1985). Previous studies have shown that larger patches of
820 *Rumex* plants are associated with *G. atrocyanea* and smaller patches with *G. grisescens*
821 (Suzuki, 1985). This is related to differences in resource utilisation by the two species
822 and to competition advantage: *G. atrocyanea* requires greater amounts of food resources
823 than *G. grisescens*, and *G. grisescens*, which is less competitive, avoids competition
824 with *G. atrocyanea* (Suzuki, 1985).

825

826 **Field survey**

827 ***Total phenolics and condensed tannins analysis***

828 We dried focal plants at 50 °C in an oven over 3 days. Dried leaf tissues were powdered
829 in a mill. Total phenolics were extracted from 20 mg of leaf powder in 10 mL of 50%
830 methanol for 1 h in an ultrasound bath at 40 °C. The content of total phenolics (mg/g)
831 was measured by using the Folin–Ciocalteu method (Julkunen-Tiitto, 1985). Condensed
832 tannins were extracted from 50 mg of dry leaves and were quantified by radial diffusion
833 assay with tannic acid as a standard (Hagerman, 1987).

834

835 ***Leaf beetle choice experiment using leaf sections from naturally growing plants***

836 In April 2018, Solitary and Aggregated *Rumex obtusifolius* plants (85 individuals each)
837 with rosette diameters of about 30 cm were selected at random in Hirosaki City. We
838 collected the youngest fully expanded leaves from the plants. These leaves had no
839 damage. We cut one 2-cm piece from the base of each collected leaf. A wet filter paper
840 (8 cm in diameter) was placed in a covered Petri dish (8.5 cm in diameter), and a piece
841 of leaf from a Solitary plant and a piece of leaf from an Aggregated plant were placed
842 on it. Adults of *G. atrocyanea* (36 females and 14 males) and *G. grisescens* (22 females
843 and 13 males) were also collected from the field in Hirosaki City. One beetle was placed
844 in the centre of the Petri dish, which was then kept in a growth chamber for 24 h at
845 25 °C, with a 12L 12D cycle. Each leaf piece was scanned by using an image scanner,
846 and the consumed area of the leaf was measured by using ImageJ bundled with 64-bit
847 Java 1.8.0_172 image analysis software (Abràmoff et al., 2004). This experimental
848 design is generally adopted for choice experiments with herbivores (e.g., Blüthgen,

849 2007; Sato et al., 2014; Shirahama et al., 2017; Lackner et al., 2019).

850

851

852 **Cultivation experiments**

853 ***Leaf beetle choice experiment***

854 To reveal whether changes in leaf chemical contents induced by interaction in *R.*

855 *obtusifolius* influenced the preferences of leaf beetles, we conducted choice experiments

856 with the *R. obtusifolius* leaves used in the cultivation experiment. The experimental

857 design was similar to that described for the choice experiment using field leaves. Leaf

858 pairs were cut into 1.5-cm pieces taken from the leaf base and placed on a Petri dish.

859 The combinations of leaf pairs were as follows: no-interaction treatment versus

860 intraspecific interaction; interspecific interaction versus intraspecific interaction; and

861 no-interaction versus interspecific treatment. In the interspecific interaction treatment,

862 treatments using three different species of plants (*P. asiatica*, *T. repens* and *F. ovina*)

863 were used equally. In this experiment, we used only *G. atrocyanea*, because in the

864 choice experiment using field leaves only *G. atrocyanea* had expressed a significant

865 preference for group leaves (see Results). Male ($N = 65$) and female ($N = 69$) beetles

866 were collected from Hirosaki City and assigned to the three different interaction

867 treatments (intraspecific interaction versus interspecific interaction, 25 males and 25

868 females; interspecific interaction versus no interaction, 15 males and 19 females; no

869 interaction versus intraspecific interaction, 25 males and 25 females). Experimental

870 conditions were the same as those in the choice experiment using field leaves.

871

872 ***Measurement of leaf traits in cultivated plants***

873 Chlorophyll content. First, a non-destructive chlorophyll meter (SPAD -502 Plus;
874 Konica Minolta, Tokyo, Japan) was used to measure the chlorophyll content in the most
875 recently fully expanded leaves of each cultivated *R. obtusifolius* on the final day of
876 cultivation. A chlorophyll meter is one of the most commonly used diagnostic tools for
877 rapid and non-destructive estimation of chlorophyll content in leaves; the resulting
878 SPAD values are positively correlated with chlorophyll content (Shibaeva et al., 2020).
879 Each leaf was measured twice—in the central part on both sides of the main vein—and
880 the average value was determined. Next, one longitudinal half of each leaf was used to
881 measure primary metabolites. The other halves of the leaves were used in the leaf beetle
882 choice experiment.

883 *Organic acids analysis*

884 Organic acids were extracted according to the method of Miyagi et al. (2010). The leaf
885 halves were frozen with liquid nitrogen and stored in a freezer at -80°C until
886 measurement. Organic acids were extracted according to the method of Miyagi et al.
887 (2010). Frozen leaves (about 50 mg) were milled, and 50% methanol containing 50 mM
888 PIPES (1,4-piperazineethanesulfonate) as an internal standard were added. After initial
889 centrifugation (22,000g, 5 min, 4°C), the supernatant was transferred to a 3-kDa cut-off
890 filter (Millipore, Billerica, MA, USA) and recentrifuged (14,000g, 30 min, 4°C). Five
891 primary metabolites—oxalate, isocitrate, citrate, glycolate and threonate—were selected
892 for analysis. These metabolites had been found in *R. obtusifolius* in our preliminary
893 survey. The resulting filtrate was quantified by capillary electrophoresis triple
894 quadrupole mass spectrometry (CE-QQQ-MS, CE; 7100, MS; 6420 Triple Quad
895 LC/MS, Agilent Technologies, Santa Clara, CA, USA) with multi-reaction monitoring
896 mode according to the method of Miyagi et al. (2019). Because of the small leaf size,

897 plants for which we did not have enough samples for analysis were excluded from the
898 analysis, and the final number of plants analysed for organic acids was slightly smaller
899 than the original collection (no interaction, $N = 35$; intraspecific interaction, $N = 63$;
900 interspecific interaction, $N = 97$).

901

902

903 **Mesocosm experiment**

904 *Preparation of plants and insects*

905 In August 2018, seeds of *R. obtusifolius* were collected from three individuals in
906 Hirosaki City. Individuals were separated by at least 2 km. In October 2019 and June
907 2020, seeds were germinated in the same way as described in the cultivation experiment
908 and used for the mesocosm experiments. Two weeks later, we planted two *R.*
909 *obtusifolius* seedlings in each pot (10.5 cm diameter \times 9 cm high) containing seed-free
910 garden soil (Mori Sangyo Co., Ltd, Japan). As a no-interaction treatment, pots were
911 divided in half by a plastic plate to block below-ground interaction of the plants in each
912 experiment (2019, $N = 35$; 2020, $N = 84$), because changes in leaf chemical content in
913 response to conspecific neighbours depend on direct interaction below the ground
914 (Ohsaki, 2020). The other pots were assigned to the interaction treatment, which
915 consisted of two *R. obtusifolius* seedlings with no plastic plate (2019, $N = 35$; 2020, $N =$
916 74). All pots were watered once a day and maintained at 25 °C, 12L 12D for 30 days in
917 growth chambers. *Gastrophysa atrocyanea* were collected from the field and kept in the
918 laboratory until required for the experiment.

919

920

921 **References For Supplementary Methods**

922 Abramoff, M. D., Magalhães, P. J., & Ram, S. J. (2004). Image processing with ImageJ.

923 *Biophotonics international*, 11, 36-42.

924 Blüthgen, N., & Metzner, A. (2007). Contrasting leaf age preferences of specialist and

925 generalist stick insects (Phasmida). *Oikos*, 116, 1853-1862.

926 <https://doi.org/10.1111/j.0030-1299.2007.16037.x>

927 Feduraev, P., Chupakhina, G., Maslennikov, P., Tacenko, N., & Skrypnik, L. (2019).

928 Variation in phenolic compounds content and antioxidant activity of different

929 plant organs from *Rumex crispus* L. and *Rumex obtusifolius* L. at different

930 growth stages. *Antioxidants*, 8, 237. <https://doi.org/10.3390/antiox8070237>

931 Hagerman, A. E. (1987). Radial diffusion method for determining tannin in plant

932 extracts. *Journal of Chemical Ecology*, 13, 437-449.

933 <https://doi.org/10.1007/BF01880091>

934 Julkunen-Tiitto, R. (1985). Phenolic constituents in the leaves of northern willows:

935 methods for the analysis of certain phenolics. *Journal of Agricultural and*

936 *Food Chemistry*, 33, 213-217. <https://doi.org/10.1021/jf00062a013>

937 Lackner, S., Lackus, N. D., Paetz, C., Köllner, T. G., & Unsicker, S. B. (2019).

938 Aboveground phytochemical responses to belowground herbivory in poplar

939 trees and the consequence for leaf herbivore preference. *Plant, Cell &*

940 *Environment*, 42, 3293-3307. <https://doi.org/10.1111/pce.13628>

941 Makuchi, T., & Sakai, H. (1984). Seedling survival and flowering of *Rumex obtusifolius*

942 L. in various habitats. *Weed Research*, 29, 123–130.

943 <https://doi.org/10.3719/weed.29.123>

944 Miyagi, A., Noguchi, K., Tokida, T., Usui, Y., Nakamura, H., Sakai, H., Hasegawa, T., &

- 945 Kawai-Yamada, M. (2019). Oxalate contents in leaves of two rice cultivars
946 grown at a free-air CO₂ enrichment (FACE) site. *Plant Production Science*,
947 22, 407–411. <https://doi.org/10.1080/1343943X.2019.1598272>
- 948 Miyagi, A., Takahashi, H., Takahara, K., Hirabayashi, T., Nishimura, Y., Tezuka, T.,
949 Kawai-Yamada, M., & Uchimiya, H. (2010). Principal component and
950 hierarchical clustering analysis of metabolites in destructive weeds;
951 polygonaceous plants. *Metabolomics*, 6, 146–155.
952 <https://doi.org/10.1007/s11306-009-0186-y>
- 953 Ohsaki H. (2020). Effects of plant-plant interactions on resource usage of herbivores.
954 MS thesis, Hirosaki University, Hirosaki (in Japanese)
- 955 Sato, Y., Kawagoe, T., Sawada, Y., Hirai, M. Y., & Kudoh, H. (2014). Frequency-
956 dependent herbivory by a leaf beetle, *Phaedon brassicae*, on hairy and
957 glabrous plants of *Arabidopsis halleri* subsp. *gemmifera*. *Evolutionary*
958 *Ecology*, 28, 545-559. <https://doi.org/10.1007/s10682-013-9686-3>
- 959 Shibaeva, T. G., Mamaev, A. V., & Sherudilo, E. G. (2020). Evaluation of a SPAD-502
960 Plus Chlorophyll Meter to Estimate Chlorophyll Content in Leaves with
961 Interveinal Chlorosis. *Russian Journal of Plant Physiology*, 67, 690-696.
962 <https://doi.org/10.1134/S1021443720040160>
- 963 Shirahama, S., Yamawo, A., & Tokuda, M. (2017). Dimorphism in trichome production
964 of *Persicaria lapathifolia* var. *lapathifolia* and Its multiple effects on a leaf
965 beetle. *Arthropod-Plant Interactions*, 11, 683-690.
966 <https://doi.org/10.1007/s11829-017-9520-x>
- 967 Suzuki, N. (1985). Resource utilization of three chrysomelid beetles feeding on *Rumex*
968 plants with diverse vegetational background. *Japanese Journal of Ecology*,

969 35, 225-234. https://doi.org/10.18960/seitai.35.2_225

970 Van Evert, F. K., Samsom, J., Polder, G., Vijn, M., Dooren, H. J. V., Lamaker, A., van
971 der Heijden, G. W., Kempenaar, C., van der Zalm, T., & Lotz, L. A. P. (2011).
972 A robot to detect and control broad-leaved dock (*Rumex obtusifolius* L.) in
973 grassland. *Journal of Field Robotics*, 28, 264-277.
974 <https://doi.org/10.1002/rob.20377>

975