1	Large herbivorous wildlife and livestock differentially influence the
2	relative importance of different sources of carbon for riverine food webs
3	
4	Frank O. Masese <sup>1,2#</sup> , Thomas Fuss <sup>2,3</sup> , Lukas Thuile Bistarelli <sup>2,3</sup> , Caroline
5	Buchen-Tschiskale <sup>4,5</sup> and Gabriel Singer <sup>2,3</sup>
6	<sup>1</sup> University of Eldoret, Department of Fisheries and Aquatic Science, P.O. Box 1125-30100
7	Eldoret, Kenya
8	<sup>2</sup> Department of Ecohydrology, Leibniz-Institute of Freshwater Ecology and Inland Fisheries
9	(IGB), Berlin, Germany
10	<sup>3</sup> Department of Ecology, University of Innsbruck, Technikerstrasse 25, A-6020 Innsbruck,
11	Austria
12	<sup>4</sup> Thünen Institute of Climate-Smart Agriculture, Bundesallee 65, 38116 Braunschweig,
13	Germany
14	$^{5}$ Leibniz Centre for Agricultural Landscape Research (ZALF), Eberswalder Straße 84, 15374
15	Müncheberg, Germany
16	
17	Corresponding author email: <u>fmasese@uoeld.ac.ke</u> , <u>f.masese@gmail.com</u>

#### 19 Abstract

In many regions around the world, large populations of native wildlife have declined or been 20 21 replaced by livestock grazing areas and farmlands, with consequences on terrestrial-aquatic 22 ecosystems connectivity and trophic resources supporting food webs in aquatic ecosystems. 23 The river continuum concept (RCC) and the riverine productivity model (RPM) predict a shift 24 of carbon supplying aquatic food webs along the river: from terrestrial inputs in low-order 25 streams to autochthonous production in mid-sized rivers. Here, we studied the influence of 26 replacing large wildlife (mainly hippos) with livestock on the relative importance of C3 27 vegetation, C4 grasses and periphyton on macroinvertebrates in the Mara River, which is an African montane-savanna river known to receive large subsidy fluxes of terrestrial carbon 28 and nutrients mediated by LMH, both wildlife and livestock. Using stable carbon ( $\delta^{13}$ C) and 29 nitrogen ( $\delta^{15}$ N) isotopes, we identified spatial patterns of the relative importance of 30 31 allochthonous carbon from C3 and C4 plants (woody vegetation and grasses, respectively) 32 and autochthonous carbon from periphyton for macroinvertebrates at various sites of the Mara River and its tributaries. Potential organic carbon sources and invertebrates were 33 sampled at 80 sites spanning stream orders 1 to 7, various catchment land uses (forest, 34 35 agriculture and grasslands) and different loading rates of organic matter and nutrients by LMH (livestock and wildlife, i.e., hippopotamus). The importance of different sources of 36 37 carbon along the river did not follow predictions of RCC and RPM. First, the importance of C3 38 and C4 carbon was not related to river order or location along the fluvial continuum but to 39 the loading of organic matter (dung) by both wildlife and livestock. Notably, C4 carbon was important for macroinvertebrates even in large river sections inhabited by hippos. Second, 40 even in small 1<sup>st</sup> -3<sup>rd</sup> order forested streams, autochthonous carbon was a major source of 41

- 42 energy for macroinvertebrates, and this was fostered by livestock inputs fuelling aquatic
- 43 primary production throughout the river network. Importantly, our results show that
- replacing wildlife (hippos) with livestock shifts river systems towards greater reliance on
- 45 autochthonous carbon through an algae-grazer pathway as opposed to reliance on
- 46 allochthonous inputs of C4 carbon through a detrital pathway.
- 47 Keywords: aquatic-terrestrial connectivity, C4 carbon, organic matter, nutrient loading,
- 48 MixSIAR models, resource subsidies
- 49
- 50

# 51 1. Introduction

52	While resource transfers from terrestrial to riverine aquatic ecosystems can occur passively
53	through wind and atmospheric deposition, overland and riverine flow (Abrantes and
54	Sheaves, 2010; Lamberti et al., 2010; Wipfli et al., 2007), animals are major active resource
55	vectors through defecation (egestion), urination (excretion) and in the form of carcasses
56	(Naiman and Rogers, 1997; Subalusky et al., 2015; Subalusky et al., 2017). Resource subsidies
57	by large mammalian herbivores (LMH) are particularly interesting because of their capacity
58	to transfer large amounts of high-quality, nutrient-rich resources that strongly influence
59	abiotic conditions, productivity, community structure and trophic interactions in recipient
60	aquatic ecosystems (Vanni, 2002; Masese et al., 2018; Stears et al., 2018).
61	Along their longitudinal gradient, rivers are postulated to exhibit a prominent shift in trophic
62	resources supporting aquatic communities, influencing their structure and functional
63	organization besides physical and chemical factors (Vannote et al., 1980; Atkinson et al.,
64	2017). In temperate headwater streams, allochthonous carbon from C3-plants (woody
65	vegetation) dominates the base of food webs. As streams widen, however, in-stream
66	primary production grows, while allochthonous inputs decrease to subsidies leaking from
67	upstream (Vannote et al., 1980; Thorp and Delong, 1994). In contrast, in Afrotropical
68	savanna rivers, mega-herbivores including hippos (Hippopotamus amphibius), elephants,
69	buffalo, other ungulates, such as wildebeest, transfer substantial amounts of terrigenous
70	carbon and nutrients to entire river networks (Naiman and Rogers, 1997; Mosepele et al.,
71	2009, Hulot et al., 2019), challenging existing models of riverine ecosystem functioning.
72	Notably, this allochthonous carbon is mainly derived from C4 plants in surrounding
73	grasslands and seems to critically need grazing LMH as subsidy vectors (Marwick et al.,

74	2014). Indeed, hippo egestion can account for up to 88% of total carbon inputs into lowland
75	rivers (Subalusky et al., 2015), which may play a similar role as leaf litter in headwater
76	streams in stimulating microbial processes and supporting food webs (Webster, Benfield et
77	al. 1999). Large quantities of hippo dung have also been found to be detrimental to aquatic
78	biodiversity, with fish kills occurring during phases of reduced flow and high hippo density
79	(Dutton et al., 2018a; Stears et al., 2018).
80	Populations of native LMH have declined in their natural range and have been replaced by
81	human settlements, farmlands and livestock (Doughty et al., 2013; Ogutu et al., 2016;
82	Veldhuis et al., 2019). Large herds of livestock (mainly cattle, goats and sheep) may
83	compensate losses of resource subsidies to aquatic ecosystems arising from wildlife declines
84	(Iteba et al., 2021; Masese et al., 2020), but the quality, quantity (rate), timing and location
85	of inputs can substantially differ (Subalusky and Post, 2019), with ensuing implications for
86	aquatic ecosystem functioning. For instance, nutrient-rich cattle dung was found to increase
87	primary production relative to hippo dung (Masese et al., 2020). And, since high-quality algal
88	carbon contributes higher proportions of carbon to metazoan biomass relative to woody
89	vegetation and grasses (Thorp and Delong, 2002; Douglas et al., 2005; Lau et al., 2009),
90	livestock may shift aquatic food webs to be more reliant on autochthonous sources of
91	carbon. This may be facilitated by the increased transportability of cattle dung due to
92	smaller particle sizes and faster leaching and remineralization (Masese et al., 2020), while
93	hippo faeces contain large particles that settle on the river bottom and decompose slowly
94	(Dawson et al., 2016; Dutton et al., 2020).
95	The Mara River (Kenya, Tanzania) traverses a landscape gradient, that makes it well suited to

95 The Mara River (Kenya, Tanzania) traverses a landscape gradient, that makes it well suited to
96 study the influence of land use change and altered LMH populations on the terrestrial-

97	aquatic linkages achieved through resource subsidies. The forest in the upper reaches
98	contrasts mixed small-scale and large-scale agriculture and human settlements in mid-
99	reaches (Mati et al., 2008). The region hosts livestock (mainly cattle, goats and sheep) at
100	varying densities, grazing in an overlapping distribution with wildlife (Lamprey and Reid
101	2004; Veldhuis et al., 2019). In the lower reaches, protected savanna grasslands in the
102	Kenyan Maasai Mara National Reserve (MMNR) harbour over a million migratory herbivores
103	(Ogutu et al., 2016), including >4000 hippos (Kanga et al., 2011). Per-capita input of organic
104	matter and nutrients into streams and rivers by individual livestock is small compared to
105	hippos (Sualusky et al., 2015; Masese et al., 2020) due to smaller body sizes and shorter time
106	of presence in or near water, but livestock can appear in large numbers (Iteba et al., 2021).
107	Also, resource subsidies of livestock and wildlife differ in quality, with likely impacts on
108	aquatic ecosystem functioning. A good assessment of subsidies transferred to the Mara
109	River critically needs comparative data for livestock and wildlife and how their inputs
110	interact with river size.
111	Here, we investigate carbon sources fuelling aquatic food webs at various sites along the
112	Mara River, specifically the interaction between river size and subsidy fluxes driven by
113	livestock and large wildlife (hippos) at various densities. In the greater Mara River network,
114	we sampled 80 sites in streams differing in catchment land use and LMH abundance. We
115	used natural abundance stable isotope analysis (SIA) of carbon ( $\delta^{13}$ C) and nitrogen ( $\delta^{15}$ N) to
116	distinguish three main sources of carbon (C3 vegetation, C4 grasses and periphyton) for
117	macroinvertebrates. Contrary to predictions of the river continuum concept (RCC) and the

riverine productivity model (RPM), we hypothesise that LMH have an overriding influence

119 over stream size in determining sources of carbon for food webs in savanna rivers.

- 120 Specifically, in lower sections of the river, autochthonous production and C3 carbon should
- 121 decrease in importance while C4 carbon subsidies by livestock and large wildlife should
- 122 become more prominent in the diets of invertebrates.

#### 123 2. Materials and Methods

#### 124 **2.1 Study area**

125 This study was conducted in the Kenyan part of the Mara River catchment. Two perennial

126 tributaries, the Nyangores and Amala rivers drain the Mau Forest, which is the most

127 extensive tropical moist broadleaf forest in East Africa, before joining to form the Mara

mainstem in the lowlands (Fig. 1). In the middle reaches, several seasonal tributaries,

including the Talek, Olare Orok, Ntiakntiak, Molibany and the Sand, drain the semi-arid

130 livestock grazing lands and wildlife conservancies outside the Maasai Mara National Reserve

131 (MMNR). Annual rainfall varies from about 2000 mm in the highlands to around 1000 mm in

the lowland savanna (Jackson and McCarter, 1994). January to March is typically a dry

133 period, while March-July and October-November are wet periods known as long and short

134 rains, respectively.

The Kalenjin ethnic group live on the highlands while the Maasai pastoralists occupy the middle and lower portions of the basin. The basin hosts substantial numbers of livestock but densities differ across the catchment as the Kalenjin have diversified to crop farming and husbandry of small herds of improved breeds of cattle, while the Maasai's large herds of cattle are the mainstay of their livelihoods (Lamprey and Reid, 2004). Agricultural expansion is ongoing across the basin (Mati et al., 2008).

141	The middle reaches of the Mara River and its tributaries within the MMNR host >4,000
142	hippos (Kanga et al., 2011), which graze in savanna grasslands at night and rest in or near the
143	river during the day, transferring ~36,000 kg of organic matter in the form of dung per day
144	from the terrestrial to the aquatic domain (Kanga et al., 2011; Subalusky et al., 2015). In the
145	same region, Maasai pastoralists graze nearly 200,000 cattle and large numbers of goats and
146	sheep in communal lands adjoining the MMNR and utilize rivers as watering points (Lamprey
147	and Reid, 2004; Ogutu et al., 2016; Veldhuis et al., 2019). In communal conservancies
148	outside the MMNR, people graze their livestock in a manner that allows livestock to co-exist
149	with wildlife (Kanga et al., 2013). This distribution results in a displacement pattern with
150	hippo areas inside the reserve, mixed hippo and livestock (mainly cattle) areas in the
151	conservancies and only livestock grazing areas outside the conservancies (Kanga et al.,
152	2013).

#### 153 2.2 Study design

A total of 80 sites were selected for sampling during the beginning of the dry season in 154 155 January 2018. All sites were sampled only once unless specified otherwise. In most cases, we 156 sampled at confluences, i.e., working in both tributaries and a third downstream mainstem 157 site after effective mixing of water from the two tributaries. Sites were selected depending on both catchment-scale (mainly land use) and reach-scale influences by human activities, 158 159 cattle and wildlife. Sites were then grouped into 5 broad regions: forested (19), agricultural 160 (26), low-density livestock (15), high-density livestock (12), and wildlife (i.e., hippo) (10) sites. 161 Forest sites had C3 vegetation dominating the catchment and riparian areas, and were used 162 as a reference for the human and LMH influences. Agricultural sites were in farming areas (crop cultivation), although most farmers in the area also own low numbers (<20 per km<sup>2</sup>) of 163

164	livestock (mainly cattle). The low-density livestock sites were also in agricultural areas but
165	had a higher density of livestock (20-50 per km <sup>2</sup> ). The high-density livestock sites were
166	located in conservancies outside the MMNR where the only land use activity is the grazing of
167	large herds of cattle (> 100 heads per km <sup>2</sup> ), goats and sheep; although some wildlife such as
168	zebra, wildebeest and other herbivores (but rarely hippos) also occur in these areas. Finally,
169	hippo sites were located on the Mara River mainstem and tributaries in the MMNR
170	downstream of river sections inhabited by large populations of hippos (Kanga et al., 2011).

## 171 **2.3 Sample collection**

172 At each site, samples of the dominant riparian plants were collected by hand into aluminium

173 envelopes. Macrophytes were rare and not considered to be an important resource. We

174 collected triplicate samples of cattle and hippo dung when encountered at a site. Triplicate

175 periphyton samples were scrubbed from submerged surfaces after gentle washing to remove

invertebrates and debris. At each site macroinvertebrates were collected from riffles, runs,

177 pools and marginal vegetation using a dip net (500 μm mesh-size) and sorted alive into

178 Eppendorf vials. Plant, dung, periphyton slurry (in HDPE bottles) and invertebrates were

179 frozen for transport to the laboratory pending further processing.

## 180 **2.3.1** Physical and chemical variables

To explore relationships between catchment land use and the importance of various sources
of carbon for invertebrates in the Mara River, sub-catchments for all sites were delineated
based on a digital elevation model of Kenya (90 m resolution, Shuttle Radar Topography
Mission) and their land cover analyzed according to the major types; forest (both natural and

185 plantation), agriculture (cropland) and settlements, grasslands (grazing lands) and

186 shrublands.

205

from crabs and Mollusca.

- 187 At each site, we measured average stream width at 20-30 randomly located transects and
- average depth and velocity at a minimum of 100 randomly located points. Discharge was
- 189 estimated using the velocity-area method at one transect. Dissolved oxygen concentration
- 190 (DO), pH, temperature and electrical conductivity were measured in situ using a WTW
- 191 Multiprobe 3320 (pH320, OxiCal-SL, Cond340i; Weilheim, Germany).

#### **2.3 Sample preparation and stable isotope analysis**

193 On return to the laboratory, samples were immediately prepared for isotope analysis or 194 preserved at -20°C. To clean periphyton, the slurry was decanted onto a petri dish and excess water evaporated in an oven at 60 °C for 48 h. The dry sample was ground using a mortar and 195 196 pestle, an aliquot was weighed into a tin cup. Macroinvertebrate specimens were thawed 197 and examined under a dissecting microscope to confirm identity and remove stomach contents before drying at 60 °C for 48 h. These samples were ground with an IKA A 11 Basic 198 199 mill (IKA-Werke GmbH & Co. KG, 79219, Staufen, Germany) and weighed into tin cups. 200 Macroinvertebrates were identified mostly to the genus level. Several individuals of a given 201 taxon from each site were pooled to produce sufficient dry tissue mass except for species 202 with larger body sizes such as crabs (Potamonautes spp.), dragonflies and damselflies 203 (Odonata), some beetles (Coleoptera) and bugs (Hemiptera). To avoid contamination by carbonates, which can be enriched in <sup>13</sup>C compared to living tissue, we only used soft tissues 204

206	Stable isotope analysis was done on a Thermo Finnigan DeltaPlus Advantage isotope ratio
207	mass spectrometer (IRMS, Thermo Scientific) coupled to an ECS 4010 EA elemental analyzer
208	(Costech Analytical Technologies). Stable isotope ratios ( $^{13}$ C/ $^{12}$ C and $^{15}$ N/ $^{14}$ N) are expressed
209	in parts per mil (‰) deviations to the international standard, as defined by the equation:
210	$\delta^{13}$ C, $\delta^{15}$ N = [(R <sub>sample</sub> /R <sub>reference</sub> ) - 1] × 10 <sup>3</sup> , where R = ${}^{13}$ C/ ${}^{12}$ C for carbon and ${}^{15}$ N/ ${}^{14}$ N for
211	nitrogen. $\delta^{13}$ C values were normalized to the international scale Vienna Pee Dee Belemnite
212	(VPDB) and $\delta^{15}$ N values to atmospheric nitrogen, by analyzes of the international standards
213	USGS40 and USGS41 (L-glutamic acid) within the sequence (Qi et al. 2003). In addition, a
214	laboratory standard ( $\delta^{13}$ C = -27.11 ‰, C% = 48.12; $\delta^{15}$ N = 0.48 ‰, N% = 2.17), which was
215	calibrated directly against USGS40 and USGS41, was measured at different positions within
216	the run. Precision, defined as the standard deviation ( $\pm 1\sigma$ ) of the laboratory control standard
217	along the run was better than $\pm 0.79\%$ for C and $\pm 0.55\%$ for N.

#### 2.4 Functional feeding groups and trophic groups 218

219 Macroinvertebrate taxa were assigned to functional feeding groups (FFGs) and trophic groups based on (Masese et al., 2014) and references therein. FFGs considered included 220 221 shredders feeding on coarse particulate organic matter (CPOM), scrapers feeding on autochthonous production (periphyton), collector-filterers feeding on suspended fine 222 particulate organic matter (FPOM), collector-gatherers feeding on deposited FPOM and 223 224 predators feeding on live animal tissue (Merritt et al., 2017). Shredders, scrapers and collectors were also subsumed as herbivores. 225

226

## 227 **2.5 Data analysis**

All physical and chemical data were transformed using natural-log transformations before 228 229 analysis. We used one-way analysis of variance (ANOVA) to test for differences in water quality, stream size and land use variables among the five regions, followed by post hoc 230 Tukey's Honestly Significant Difference (HSD) multiple comparisons of means. 231 We used MixSIAR Bayesian mixing model to estimate the contributions of different basal 232 233 sources to consumer diets (Parnell et al., 2010; Stock and Semmens, 2013). Models were run 234 for each site separately for each FFG and common family. Only C3 and C4 plants and 235 periphyton were included in the models as possible sources of carbon and nitrogen (Masese 236 et al.; 2015, 2018). For periphyton, we calculated standard deviations of source values for input into MixSIAR from triplicate samples collected from the same site. Because the isotopic 237 238 values of vegetation are influenced by elevation, for C3 and C4 sources we grouped sites into 239 three groups defined by elevation: headwaters (> 2800 m a.s.l.), middle reaches (2000-2800 m a.s.l.) and lower reaches in the MMNR and surrounding areas (<2000 m a.s.l.). Trophic 240 enrichment factors (TEFs) were set to zero after manually correcting for trophic fractionation 241 in the stable isotope composition of consumers. For  $\delta^{15}N$ , we used a trophic fractionation of 242 243  $0.6 \pm 1.7$  ‰ and  $1.8 \pm 1.7$  ‰ for herbivorous and predatory macroinvertebrates, respectively (Bunn et al., 2013). For  $\delta^{13}$ C, we used a trophic fractionation of 0.5 ± 1.3 ‰ (McCutchan Jr et 244 245 al., 2003). Concentration dependencies were set to zero.

For each site, river distance from the source (RDS) was calculated as the square root of the drainage area (Rasmussen et al., 2009) as a general measure reflecting the linear dimension of a watershed. This is based on the finding that the average length of stream paths leading to a point in the drainage can be expressed as a power function of the drainage area

250	(Gregory and Walling, 1973) with the exponent 0.5 (Smart, 1972). We used RDS and LMH
251	(livestock and hippos) density per sampling site as independent variables against which
252	longitudinal changes in fractional contributions of periphyton, C3 and C4 carbon to
253	invertebrates in the river were explored using simple linear regression (SLR). Relationships
254	were tested separately for the different FFGs.
255	Data on livestock (cattle, sheep, goats, and donkeys) and wildlife (ungulates and hippos)
256	numbers were obtained from Development Plans for Bomet and Narok Districts (Plan, 2007,
257	2008), Ministry of Agriculture and Livestock Production, and Kenya National Bureau of
258	Statistics reports (KNBS-IHBS, 2007; KNBS-LS, 2009; KNBS, 2016, 2018), and other
259	unpublished and published reports (Ottichilo et al., 2000; Lamprey and Reid, 2004; Kanga et
260	al., 2011; Kiambi et al., 2012; Ogutu et al., 2016). For livestock and other LMH, density was
261	expressed as the number of individuals per ${\sf km}^2$ in the catchment area of the river sampling
262	site, while for hippos, count data were expressed as the number of individuals per river km.
263	We used generalized additive models (GAMs) (Wood, 2017) to assess how the relative
264	importance of C3 carbon, C4 carbon and periphyton for individual FFGs and all FFGs
265	combined varied with stream size (RDS) and density of LMH. GAMs incorporate smooth
266	functions that are more flexible in modelling nonlinear relationships (Hastie and Tibshirani,
267	1990). Typically, the importance of different sources of carbon varies non-linearly with
268	stream order or distance from the source (Vannote et al., 1980). GAMs were built using
269	penalized cubic regression splines with degrees of freedom automatically identified based on
270	the generalized cross-validation score (GCV) using the mgcv-package (Wood and Wood,
271	2015) in the R platform (R-Development-Core-Team, 2017). GAMs were used further to
272	investigate the influence of region, stream size (RDS) and LMH density, including potential 13

- interactions, on the relative importance of the three sources of carbon for FFGs in the river.
- 274 GAMs included region, RDS and LMH density as fixed effects, and individual sampling sites as
- a random effect. All analyses were performed using R 3.4.4 (Team, 2017).

#### 276 **3. Results**

## 277 **3.1 Physico-chemical variables**

- 278 There was spatial variation in many physical and chemical variables (Table 1). LMH density
- increased with the proportion of forest cover and decreased with the proportion of
- 280 grassland. Electrical conductivity and water temperature increased with LMH density but
- were low at the forested, low livestock density and agricultural regions/ sites (Table 1). Apart
- 282 from the hippo sites, there were no differences in river distance from the source (RDS), river
- width, depth, or discharge among the five regions.

#### 284 **3.2 Basal resources**

The C3 and C4 plants collected at the various sites did not differ significantly in  $\delta^{13}$ C and  $\delta^{15}$ N 285 286 (Table 1), nevertheless mean isotopic signatures were calculated for each of three elevation levels for use in the MixSIAR models. The  $\delta^{15}$ N values of C4 plants were generally higher than 287 those of C3 plants.  $\delta^{13}$ C, but not  $\delta^{15}$ N, of cattle dung, varied considerably among sites and 288 regions (Table 2). In contrast,  $\delta^{13}$ C and  $\delta^{15}$ N of hippo dung showed little variation among 289 regions with hippos.  $\delta^{13}$ C and  $\delta^{15}$ N of periphyton were highly variable among sites and 290 regions (Figure 2, Table 1); specifically, agricultural and livestock sites had elevated  $\delta^{15}$ N 291 292 values compared to forested areas and the MMNR, where hippos had a strong influence

- 293 (Figure 2). In MixSIAR models, site-specific isotopic signatures were used for all resources
- 294 with detectable variation among regions.

## 295 3.3 Longitudinal trends in basal resources

296 Longitudinal patterns in  $\delta^{13}$ C of periphyton in the Mara River were wedge-shaped (Figure 2).

297 Periphyton  $\delta^{13}$ C responded to both measures of stream size RDS (r = 0.23, p < 0.01) and

stream order (r = 0.35, p < 0.01). At shorter RDS, agricultural and low density (LD) livestock

sites had the widest range of periphyton  $\delta^{13}$ C, while values at forested sites were invariably

low (-25.9±2.6‰, Table 2). The narrowest ranges of  $\delta^{13}$ C were recorded for high density (HD)

301 livestock and hippo sites, which appeared at longer RDS. At these sites,  $\delta^{13}$ C (HD livestock: -

302 23.2±1.2‰, hippo sites: -22.7±1.8‰) was on average higher than at forested sites.  $\delta^{13}$ C of

periphyton was positively related to agricultural land use (r = 0.24, p < 0.05) and negatively

to forest cover (r = -0.44, p < 0.01). Overall, periphyton  $\delta^{13}$ C responded positively to LMH

density (r = 0.52, p < 0.01), which itself was highly positively correlated with % grassland

306 cover in the catchment (r = 0.89, p < 0.01).

307 Longitudinal patterns of periphyton  $\delta^{15}$ N were similar to  $\delta^{13}$ C, yet we could not detect a

308 significant change of the mean value with any measure of stream size. Periphyton  $\delta^{15}$ N was

309 higher at agricultural and LD livestock sites (Figure 2) with shorter RDS, but lower at both the

310 forested sites with short RDS and hippo sites at longer RDS. Unexpectedly,  $\delta^{15}$ N at LD

311 livestock sites also surpassed HD livestock sites.  $\delta^{15}$ N of periphyton was positively related to

agricultural land use (r = 0.29, p < 0.01) and negatively to forest cover (r = -0.31, p < 0.01).

313 Interestingly, there was no significant relationship between  $\delta^{15}$ N of periphyton and LMH

314 density.

## 315 **3.4 Importance of different carbon sources for invertebrates**

316	A total of 47 taxa in the five macroinvertebrate FFGs were collected in the study area (Table
317	S1). The importance of C3 vegetation, C4 grasses and periphyton for invertebrates differed
318	among the five regions, but patterns were similar for scrapers and collectors (Figure 3).
319	Overall, periphyton was either the major or second-most important source of carbon for all
320	FFGs at forested, agricultural and livestock sites, and predators were more reliant on this
321	energy pathway than the rest of the FFGs (Figure 3). Except for shredders in forested
322	streams, the importance of C3 vegetation was reduced for the rest of FFGs in the other
323	regions and was lowest at the hippo sites. On the contrary, the importance of C4 carbon
324	responded strongly to LMH density and was the most important source of carbon (>50%) for
325	all FFGs at hippo sites, except shredders (Figure 3).
325 326	all FFGs at hippo sites, except shredders (Figure 3). Source contributions followed similar patterns for all FFGs except predators and shredders
326	Source contributions followed similar patterns for all FFGs except predators and shredders
326 327	Source contributions followed similar patterns for all FFGs except predators and shredders (Figure 3). Predators seemed to draw most of their nutrition from the periphyton pathway at
326 327 328	Source contributions followed similar patterns for all FFGs except predators and shredders (Figure 3). Predators seemed to draw most of their nutrition from the periphyton pathway at all sites except those with hippos. Shredders, on the other hand, displayed an overreliance
326 327 328 329	Source contributions followed similar patterns for all FFGs except predators and shredders (Figure 3). Predators seemed to draw most of their nutrition from the periphyton pathway at all sites except those with hippos. Shredders, on the other hand, displayed an overreliance on C3 vegetation as their main source of carbon at all sites, which is consistent with their

## 333 **3.5 Longitudinal trends in the importance of different carbon sources**

334 We explored relationships between stream size (i.e., river distance from the source, RDS)

and LMH density as predictors of the importance of the three differentiated sources of

carbon (C3 vegetation, C4 grasses and periphyton) for macroinvertebrates in the Mara River

337	(Figure 4). The relative importance (fractional contribution) of the three sources of carbon
338	changed with stream size (RDS) and LMH density (Figure 4a, b). Periphyton was the most
339	important source of energy in headwater and mid-sized streams, contributing on average
340	more than 40% to invertebrate biomass, but decreased to the second most important source
341	of carbon further downstream. C3 vegetation (woody vegetation) was almost as important
342	as periphyton in headwater streams, contributing nearly 40%, but its importance declined
343	from mid-sized streams onwards, becoming the least important source of carbon with
344	contributions <20% in large river sections (Figure 4a). C4 plants (grasses) were not relevant
345	in small streams contributing less than 20% to invertebrates' energy requirements, but their
346	importance markedly increased in mid-sized and large river sections, where they became the
347	dominant source of energy with a contribution of nearly 60% to the total energy
348	requirements for macroinvertebrates. The influence of LMH on the longitudinal importance
349	of carbon sources for macroinvertebrates in the river (Figure 4b) mirrored that of RDS,
350	although patterns for C4 carbon were stronger. Patterns with stream order as a proxy for
351	stream size were identical to those with RDS (data not shown).
352	We also explored longitudinal relationships between stream size (i.e., river distance from the
353	source, RDS) and LMH density as predictors of the importance of the three sources of carbon
354	(C3 vegetation, C4 grasses and periphyton) for individual FFGs (Figure 5). There were
355	significant positive relationships (linear regressions, $p < 0.05$ ) between RDS and the
356	importance of C4 to all FFGs except shredders (Figure 5g, h). In parallel, C3 carbon from
357	terrestrial vegetation became less important with RDS for collector-filters (Figure 5a, b) and
358	collector-gatherers (Figure 5c, d). Also, LMH density had strong influences on the importance
359	of carbon sources for all FFGs except shredders, by reducing the contribution of C3 while

360	increasing that of C4 carbon.	As expected, shredders w	vere mainly reliant on C3 vegetation
500	mercusing that of eq carbon.	As capetica, sincualis w	rere manny rename on es vegetation

361 and were neither influenced by stream size nor LMH density (Figure 5g, h).

362	We found a significant influence of stream size and region on the relative importance of C3
363	and C4 carbon for combined and individual FFGs, except shredders, in the river (Table 2). The
364	independent effects of LMH density on the relative importance of C4 carbon was limited to
365	scrapers and predators only, implying increasing LMH density did not necessarily increase
366	the importance of C4 carbon for invertebrates. This is in agreement with observations that
367	livestock increased the importance of periphyton while hippos increased the importance of
368	C4 carbon for most FFGs, but increasing densities of both livestock and hippos decreased the
369	importance of C3 carbon for invertebrates (Figure 3). There were significant interactions
370	between LMH density and region in the importance of C4 carbon for all FFGs, except
371	shredders. Overall, patterns in the importance of the three sources of carbon for predators
372	were similar to those of all FFGs combined.
373	To eliminate potential confounding by organic matter and nutrient inputs by other
0,0	
374	agricultural activities not associated with LMH, the influence of stream size and LMH density
375	on the relative importance of carbon sources for invertebrate FFGs were evaluated in
376	forested sites only where LMH (both livestock and wildlife) were limited (Supplementary
377	Information, Figures S1). Remarkably, no major effect of stream size was noted for most
378	FFGs, further reinforcing the important role of LMH as drivers of food resources for
379	macroinvertebrates in the river.

380

381

## 382 4. Discussion

383	This study shows that the importance of different sources of carbon for consumers in
384	savanna rivers is spatially variable in response to changes in abundance and distribution of
385	large mammalian herbivores, both livestock and wildlife, and stream size or river distance
386	from the source. Overall, (i) periphyton was the most important source of energy for
387	macroinvertebrates in low order and mid-sized rivers, (ii) periphyton and C4 vegetation
388	became more important carbon sources at the cost of C3 plants at high-density livestock and
389	hippo sites, respectively, and (iii) lower sections of the river hosting large populations of
390	LMH relied largely on C4 vegetation with very little contributions of C3 plants. Our findings
391	thus confirm our hypothesis of an overriding influence of LMH over stream size or location
392	along the fluvial continuum in determining sources of carbon for food webs in savanna
393	rivers.
394	Although previous studies in African savanna rivers have shown that LMH foster the
395	importance of C4 carbon as a major source of carbon for aquatic consumers (Masese et al.

2015, McCauley et al., 2015, Dawson et al., 2020), no study has evaluated the importance of

these resource subsidies on entire gradients of rivers and at the scale that we have done in

this study. The many stream sites sampled that differed in size and LMH density enabled us

to evaluate the interaction of stream size with subsidy quantity as determinants for the base

400 of riverine food webs. Our results depart from predictions on the importance of resources

401 along the river continuum. While a reduction in the contribution of C3 carbon is

402 hypothesized in mid-sized rivers as periphyton becomes dominant (Vannote et al., 1980;

403 Thorp and Delong, 1994), an increase in the importance of fresh C4 vegetation inputs to mid-

404	sized and large rivers is less intuitive and not hypothesized in existing models of riverine
405	ecosystem functioning. Instead, mid-sized and large rivers are thought to be
406	disproportionately reliant on autochthonous production, and leakage of unutilized materials
407	from upstream supply food webs of large rivers (Vannote et al., 1980, Thorp and Delong,
408	2002; Thorp et al., 2006).
409	Although LMH density and the proportion of C4 grasses in the catchments were strongly
410	correlated ( $r = 0.89$ ), the increased importance of C4 carbon in the diet of most
411	macroinvertebrates in this study was assumed to be mediated via the vectoring role of LMH
412	when they defecate in the river. Indeed, in the absence of LMH, the contribution of C4
413	carbon to riverine food webs is less than expected based on areal landcover (Abrantes and
414	Sheaves, 2010; Marwick et al., 2014), and given lower C4 vegetation height direct riparian
415	inputs were minimal. Without LMH-mediation, C4 carbon would hardly reach aquatic food
416	webs and dominance of C3 carbon could be expected in larger rivers, either supplied via
417	leakage from upstream or through direct deposition from riparian vegetation. Headwater
418	streams and riparian areas of most savanna rivers contain more C3 than C4 vegetation
419	(Table 1), implying that LMH are the critical vectors for C4 carbon, especially during the dry
420	season.
101	Although both livesteck and hinnes largely facilitated the disprepertienate contribution of

Although both livestock and hippos largely facilitated the disproportionate contribution of
C4 carbon from grasses to invertebrates in the river, there were interesting differences in
the contribution of other sources (C3 vegetation and periphyton) between sites receiving
inputs by livestock and hippos, which can be linked to differences in the quality, amount and
characteristics of the inputs these two types of LMH transfer into rivers (Iteba et al., 2021;
Masese et al., 2020). While increasing livestock density increased both the importance of

427	periphyton and C4 carbon for macroinvertebrates in the river, hippo populations only
428	increased the importance of C4 carbon further and reduced that of periphyton further
429	(Figure 3). Although hippo dung has been reported to fertilize aquatic ecosystems increasing
430	primary and secondary production (Grey and Harper, 2002; Mosepele et al., 2009), other
431	studies have shown that large particles in hippo dung can have detrimental effects on
432	benthic production, especially during the dry season when they settle at the bottom of hippo
433	pools and downstream sections of rivers (Dawson et al., 2016; Dutton et al., 2018a, b).
434	Hippos also spend long periods in the water for thermoregulation, approximately 12 h of
435	daytime (Subalusky et al., 2015), and their wallowing activity is known to increase turbidity
436	that can further limit primary production (Dutton et al., 2018). Comparative studies with
437	cattle dung have also shown that the weakly digested hippo dung with high C: nutrients (N
438	and P) ratios support heterotrophic microbial activity, while cattle dung with lower
439	stoichiometric ratios can increase primary production faster (Subalusky et al., 2018; Masese
440	et al., 2020). Thus, replacing wildlife (hippopotamus) with livestock (cattle) in savanna
441	landscapes will most likely weaken detrital food web links based on allochthonous C3 and C4
442	material (Dawson et al., 2020) and foster an algal-grazer food chain by increasing the
443	abundance of autochthonous material.
444	The proportional acquisition of energy from terrestrial C3 and C4 plants and periphyton

showed specific patterns for different FFGs (Zeug and Winemiller, 2008; Pingram et al.,

446 2014). Further, the importance of different sources was influenced by land use and the

447 density of LMH. For instance, in forested streams, there was no effect of stream size on the

relative importance of the three sources (C3 and C4 producers and periphyton) for most of

the FFGs (Figure S1). There was significant partitioning among FFGs on the utilization of

450	various resources, with scrapers and predators relying more on periphyton while collector-
451	gatherers and, as expected, shredders relying more on C3 vegetation (Figure 1). However,
452	facultative shredders, such as Potamonautidae (freshwater crabs) that dominated
453	invertebrates most forested and agricultural streams, can obtain a significant amount of
454	energy from periphyton during their omnivorous feeding (Masese et al., 2014). The
455	importance of trophic sources was also related to the trophic position of the
456	macroinvertebrates, with autochthonous production becoming more important with an
457	increase in trophic level (Figures S1).

# 458 **4.1 Longitudinal trends of the relative importance of trophic resources**

459	Even in the absence of LMH (both livestock and wildlife) longitudinal patterns of organic
460	matter and nutrient loading into the Mara River and its tributaries (Figures 4 and 5) departed
461	significantly from existing predictions for river continuum (Vannote et al., 1980; Thorp and
462	Delong, 2002; Thorp et al., 2006). Periphyton, and not leaf litter from C3 vegetation, was the
463	most important source of energy for invertebrates in low-order forested streams by
464	contributing nearly 50%, on average, of the energy requirements of all invertebrates while
465	C3 vegetation contributed no more than 40%. These results have been supported by
466	previous studies in tropical and temperate regions whereby carbon from autochthonous
467	production contributes significantly to aquatic animal biomass in mid-sized and large rivers
468	despite forming a small proportion of available food resources (Douglas et al., 2005; Hayden
469	et al., 2016).

The importance of C3 vegetation for invertebrates declined predictably in mid-sized rivers,
but an expected rebound in lower reaches of the river where turbidity limits primary

472	production and the river becomes heterotrophic with an overreliance on fine particulate
473	organic matter, mainly from C3 vegetation, escaping from upstream (Vannote et al., 1980),
474	did not occur. Instead, the importance of C3 carbon further plummeted, while that of C4
475	carbon, mediated by large populations of livestock and large wildlife (mainly hippos), gained
476	more importance. Moreover, the expected increase in the importance of autochthonously
477	produced carbon in large rivers (Thorp and Delong, 1994, 2002) did not occur, further
478	highlighting the predominant role played by LMH as drivers of ecosystem productivity and
479	functioning in savanna rivers (Stears et al., 2018; Subalusky et al., 2018; Dawson et al., 2020;
480	Masese et al., 2020).
481	Recognizing that carbon flow in riverine food webs is context-dependent, both temporally in
482	terms of flow variability and spatially in terms of the strength of lateral and longitudinal
483	connectivity, it is conceivable that the patterns we report here are transient. However,
484	studies during both the dry and wet seasons, have indicated the overriding influence of LMH
485	inputs on river water quality and biogeochemistry (Stears et al., 2018; Dutton et al., 2018b,
486	2020), ecosystem productivity (Subalusky et al., 2017, 2018), community composition and
487	diversity of invertebrates and fishes (Masese et al., 2018; Stears et al., 2018). The
488	discrepancies between the results of this study, on the relative importance of different
489	sources of carbon for supporting riverine food webs on the fluvial continuum of savanna
490	rivers, and existing models of riverine ecosystem functioning suggests that a new model is
491	needed that recognizes the intimate linkage between terrestrial and aquatic ecosystems in
492	savanna landscapes mediated by the active transfer of organic matter by LMH.

## 494 **5. Conclusions**

495	An understanding of terrestrial-aquatic ecosystems connectivity and the main sources of
496	energy supporting riverine food webs is critical for the conservation and management of
497	African savanna rivers. The novel findings of this study show the distinct functioning of these
498	rivers in terms of the major energy sources fuelling food webs along their riverine continuum
499	and highlight the important role played by LMH as vectors enhancing aquatic productivity
500	through strengthened terrestrial-aquatic subsidy fluxes. We show that terrestrial and
501	autochthonous sources of energy are relatively important along the longitudinal gradient of
502	savanna rivers, and identify LMH as critically enhancing the contribution of C4 carbon far
503	above potential contributions in their absence. The findings of this study also show that
504	different resources are partitioned among macroinvertebrates depending on their feeding
505	modes and location on the fluvial continuum. At a local scale, this study presents further
506	evidence on the species-specific influences of different species of LMH on the functioning of
507	aquatic ecosystems in African savannas. It also suggests that replacing native populations of
508	large herbivores (such as hippos) with livestock (such as cattle) may alter the major sources
509	of energy supporting aquatic communities in savanna rivers, with consequences on
510	ecosystem structure and functioning.

## 511 Acknowledgements

512 We are grateful to Mary Kiplagat (University of Eldoret) and Evans Ole Keshe (Mario Tours) 513 for assistance in the field. We are grateful to Tobias Goldhammer, Sarah Krocker and Claudia 514 Schmalsch for help during the analysis of water chemistry samples at IGB, Berlin. We thank 515 Susanne Remus for stable isotope analysis at the ZALF. We thank Narok County for granting

516	us access to	the Mara	Triangle.	This is a	publication	of the	MaMeMa	Project and	d was funded,
-----	--------------	----------	-----------	-----------	-------------	--------	--------	-------------	---------------

- 517 in part, by an Alexander von Humboldt Postdoc fellowship to FOM; and an ERC Research
- 518 grant to GS.
- 519

520 R	eferences
-------	-----------

- 521 Abrantes, K. G., Sheaves, M., 2010. Importance of freshwater flow in terrestrial-aquatic
- 522 energetic connectivity in intermittently connected estuaries of tropical Australia.
- 523 Marine Biology 157, 2071-2086.
- 524 Anderson, C. Cabana, G., 2005. δ15N in riverine food webs: effects of N inputs from
- agricultural watersheds. Canadian Journal of Fisheries and Aquatic Sciences 62, 333340.
- 527 Atkinson, C. L., Capps, K. A., Rugenski, A. T., Vanni, M. J., 2017. Consumer-driven nutrient
- 528 dynamics in freshwater ecosystems: From individuals to ecosystems. Biological
- 529 Reviews 92, 2003-2023.
- 530 Bunn, S. E., Leigh, C., Jardine, T. D., 2013. Diet-tissue fractionation of δ15N by consumers
- from streams and rivers. Limnology and Oceanography 58(3): 765-773.
- 532 Dawson, J., Pillay, D., Perissinotto, R., Richoux' N. B., 2020. Fatty acid analyses provide novel
- 533 insights on hippo defecation and consequences for aquatic food webs. Scientific
- 534 Reports 10(1): 1-15.
- 535 Dawson, J., Pillay, D., Roberts, P. J., Perissinotto, R., 2016. Declines in benthic
- 536 macroinvertebrate community metrics and microphytobenthic biomass in an estuarine
- 537 lake following enrichment by hippo dung. Scientific Reports 6, 37359.

538	Doughty, C.E.,	Wolf, A.,	, Malhi, Y.	, 2013.	The	impact of	f large	anima	extinctions	on	nutrient
-----	----------------	-----------	-------------	---------	-----	-----------	---------	-------	-------------	----	----------

- 539 fluxes in early river valley civilizations. Ecosphere 4, art148.
- 540 Douglas, M.M., Bunn, S.E., Davies, P.M., 2005. River and wetland food webs in Australia's
- 541 wet-dry tropics: general principles and implications for management. Marine and
- 542 Freshwater Research 56, 329-342.
- 543 Dutton, C.L., Subalusky, A.L., Hamilton, S.K., Rosi, E.J., Post, D.M., 2018a. Organic matter
- 544 loading by hippopotami causes subsidy overload resulting in downstream hypoxia and
  545 fish kills. Nature Communications 9, 1951.
- 546 Dutton, C.L., Subalusky, A.L., Anisfeld S.C., Njoroge, L., Rosi E. J., Post, D.M., 2018b. The
- 547 influence of a semi-arid sub-catchment on suspended sediments in the Mara River,
- 548 Kenya. PloS one 13, e0192828.
- 549 Dutton, C. L., Subalusky, A.L., Hamilton, S.K., Bayer, E.C., Njoroge, L., Rosi, E.J., Post, D.M.,
- 550 2021. Alternative Biogeochemical States of River Pools Mediated by Hippo Use and
  551 Flow Variability. Ecosystems, 24, 284-300.
- Gregory, K.J. Walling, D. E., 1973. Drainage basin form and process: a geomorphological
  approach. Wiley, New York, p 456.
- 554 Grey, J., Harper, D., 2002. Using stable isotope analyses to identify allochthonous inputs to
- Lake Naivasha mediated via the hippopotamus gut. Isotopes in environmental and
  health studies 38, 245-250.
- 557 Hastie, T.J., Tibshirani, R.J., 1990. Generalized additive models, CRC press.
- 558 Hayden, B., McWilliam-Hughes, S.M., Cunjak, R.A., 2016. Evidence for limited trophic
- 559 transfer of allochthonous energy in temperate river food webs. Freshwater Science 35,
- 560 544-558.

561 Hulot	, F.D., P	rijac, A.	, Lefebvre,	, JP.,	, Msiteli-Shumba	, S.	, Kativu	, S.,	2019.	A first	assessment
-----------	-----------	-----------	-------------	--------	------------------	------	----------	-------	-------	---------	------------

- 562 of megaherbivore subsidies in artificial waterholes in Hwange National Park,
- 563 Zimbabwe. Hydrobiologia 837, 161-175.
- Iteba, J.O., Thomas, H., Singer, Singer, G.A., Masese, F.O., 2021. Livestock as vectors of
- organic matter and nutrient loading in aquatic ecosystems in African savannahs. PLOS
  ONE. In Revision.
- 567 Jackson, A., Mccarter P., 1994. A Profile of the Mau Forest Complex. KEFCON, Nairobi.
- 568 Kanga, E.M., Ogutu, J.O., Olff, H., Santema, P. 2011. Population trend and distribution of the
- 569 Vulnerable common hippopotamus Hippopotamus amphibius in the Mara Region of
- 570 Kenya. Oryx 45, 20-27.
- 571 Kanga, E.M., Ogutu, J.O., Piepho, H.-P., Olff, H., 2013. Hippopotamus and livestock grazing:
- 572 influences on riparian vegetation and facilitation of other herbivores in the Mara

573 Region of Kenya. Landscape and Ecological Engineering 9, 47-58.

- 574 Kiambi, S., Kuloba, B., Kenana, L., Muteti, D., Mwenda, E., 2012. Wet season aerial count of
- 575 large herbivores in Masai Mara National Reserve and the adjacent community areas
- 576 (June 2010). Narok: Mara Research Station, Kenya Wildlife Service.
- 577 KNBS-IHBS, 2007. Kenya National Bureau of Statistics (KNBS)-Kenya Integrated Household
- 578 Budget Survey (KIHBS)—2005/06.
- 579 KNBS-LS. 2009. Livestock population by type and district. Nairobi: Census KNBS.
- 580 KNBS 2016. Livestock statistics. Kenya National Bureau of Statistics.
- 581 KNBS (2018). Economic Survey 2018.
- Lamberti, G.A., Chaloner, D.T., Hershey, A.E. 2010. Linkages among aquatic ecosystems.
- Journal of the North American Benthological Society 29, 245-263.

584	Lamprey, R.H., Reid R.S.	., 2004. Expansion of human settlement in Kenya's Maasai M	ara:
-----	--------------------------	--	------

- 585 what future for pastoralism and wildlife? Journal of Biogeography 31, 997-1032.
- Lau, D.C., Leung, K.M., Dudgeon, D., 2009. Are autochthonous foods more important than
- 587 allochthonous resources to benthic consumers in tropical headwater streams? Journal
- of the North American Benthological Society 28, 426-439.
- Lee, K.Y., Graham, L., Spooner D.E., Xenopoulos, M., 2018. Tracing anthropogenic inputs in
- stream foods webs with stable carbon and nitrogen isotope systematics along an
  agricultural gradient. PloS one 13(7).
- 592 Marwick, T.R., Borges, A.V., Van Acker, K., Darchambeau, F., Bouillon, S., 2014.
- 593 Disproportionate contribution of riparian inputs to organic carbon pools in freshwater 594 systems. Ecosystems 17, 974-989.
- 595 Masese, F.O., Abrantes, K.G., Gettel, G.M., Bouillon, S., Irvine, K., McClain M.E., 2015. Are

596 Large Herbivores Vectors of Terrestrial Subsidies for Riverine Food Webs? Ecosystems
597 18, 686-706.

598 Masese, F.O., Abrantes, K.G., Gettel, G.M., Bouillon, S., Irvine, K., McClain M.E., 2018.

599 Trophic structure of an African savanna river and organic matter inputs by large

- 600 terrestrial herbivores: A stable isotope approach. Freshwater Biology 63, 1365-1380.
- Masese, F.O., Kiplagat, M.J., Gonzalez-Quijano, C.R., Subalusky, A.L., Dutton, C.L., Post, D.M.,
- 602 Singer, G.A., 2020. Hippopotamus are distinct from domestic livestock in their resource
- subsidies to and effects on aquatic ecosystems. Proc Biol Sci 287(1926), 20193000.
- 604 Masese, F.O., Kitaka, N., Kipkemboi, J., Gettel, G.M., Irvine, K., McClain, M.E., 2014.
- 605 Macroinvertebrate functional feeding groups in Kenyan highland streams: evidence for
- a diverse shredder guild. Freshwater Science 33, 435-450.

607	Mati, B.M.	, Mutie, S.	, Gadain, H	., Home,	P., Mtalo	, F.	, 2008. lm	pacts of	land-use,	/cover
-----	------------	-------------	-------------	----------	-----------	------	------------	----------	-----------	--------

- 608 changes on the hydrology of the transboundary Mara River, Kenya/Tanzania. Lakes &
- 609 Reservoirs: Research & Management 13, 169-177.
- 610 McCauley, D.J., Dawson, T.E., Power, M.E., Finlay, J.C., Ogada, M., Gower, D.B., Caylor, K.,
- 611 Nyingi, W.D., Githaiga, J.M., Nyunja, J., Joyce, F.H., Lewison, R.L., Brashares, J.S., 2015.
- 612 Carbon stable isotopes suggest that hippopotamus-vectored nutrients subsidize
- 613 aquatic consumers in an East African river. Ecosphere 6, 1-11.
- 614 McCutchan Jr, J.H., Lewis Jr, W.M., Kendall, C., McGrath, C.C., 2003. Variation in trophic shift
- for stable isotope ratios of carbon, nitrogen, and sulfur. Oikos 102, 378-390.
- 616 Merritt, R.W., Cummins, K.W., Berg, M.B., 2017. Trophic relationships of
- 617 macroinvertebrates. Methods in Stream Ecology, Volume 1, Elsevier: 413-433.
- Mosepele, K., Moyle, P.B., Merron, G.S., Purkey, D.R., Mosepele, B. 2009. Fish, Floods, and
- 619 Ecosystem Engineers: Aquatic Conservation in the Okavango Delta, Botswana.
- 620 BioScience 59, 53-64.
- Naiman, R.J., Rogers, K.H., 1997. Large animals and system-level characteristics in river
- 622 corridors. BioScience 47, 521-529.
- 623 Ogutu, J.O., H.-P. Piepho, H.-P., Said, M.Y., Ojwang, G.O., Njino, L., Kifugo, S.C., Wargute,
- P.W., 2016. Extreme Wildlife Declines and Concurrent Increase in Livestock Numbers in
  Kenya: What Are the Causes? PLOS ONE 11, e0163249.
- 626 Ottichilo, W.K., Grunblatt, J., Said, M.Y., Wargute, P.W., 2000. Wildlife and livestock
- 627 population trends in the Kenya rangeland. Wildlife conservation by sustainable use,
- 628 Springer: 203-218.

- 629 Parnell, A.C., Inger R., Bearhop, S., Jackson, A.L., 2010. Source partitioning using stable
- isotopes: coping with too much variation. PLOS ONE 5(3).
- 631 Plan, D.D., 2007. Effective management for sustainable economic growth and poverty
- 632 reduction, Bomet District 2002-2008.
- Plan, D.D., 2008. Effective management for sustainable economic growth and poverty
- 634 reduction, Narok District 2002-2008.
- 635 Qi, H., Coplen, T.B., Geilmann, H., Brand, W.A., Böhlke, J.K., 2003. Two new organic
- reference materials for δ13C and δ15N measurements and a new value for the δ13C of
- 637 NBS 22 oil. Rapid Communications in Mass Spectrometry 17, 2483-2487.
- Rasmussen, J.B., Trudeau, V., Morinville, G., 2009. Estimating the scale of fish feeding
- 639 movements in rivers using  $\delta$ 13C signature gradients. Journal of animal ecology 78, 674-
- 640 685.
- 641 Smart, J., 1972. Quantitative characterization of channel network structure. Water
- 642 Resources Research 8, 1487-1496.
- 643 Stears, K., McCauley, D.J. Finlay, J.C., Mpemba, J., Warrington, I.T., Mutayoba, B.M., Power,
- 644 M.E., Dawson T.E., Brashares J.S., 2018. Effects of the hippopotamus on the chemistry
- and ecology of a changing watershed. Proceedings of the National Academy of
- 646 Sciences. 115(22), E5028-E5037. https://doi.org/10.1073/pnas.1800407115
- 647 Stock, B.C., Semmens, B.X., 2013. MixSIAR GUI user manual, version 1.0. Accessible online at:
- 648 http://conserver. iugo-cafe. org/user/brice. semmens/MixSIAR.
- 649 Subalusky, A.L., Dutton, C.L., Njoroge, L., Rosi E.J., Post D.M., 2018. Organic matter and
- 650 nutrient inputs from large wildlife influence ecosystem function in the Mara River,
- 651 Africa. Ecology 99, 2558-2574.

652	Subalusky, A.L.	, Dutton, C.L	., Rosi-Marshall,	E.J., Post D.M.	, 2015. The hippopotamus
-----	-----------------	---------------	-------------------	-----------------	--------------------------

653 conveyor belt: vectors of carbon and nutrients from terrestrial grasslands to aquatic

654 systems in sub-Saharan Africa. Freshwater Biology 60, 512-525.

- 655 Subalusky, A.L., Dutton C.L., Rosi E.J., Post, D.M., 2017. Annual mass drownings of the
- 656 Serengeti wildebeest migration influence nutrient cycling and storage in the Mara
- 657 River. Proceedings of the National Academy of Sciences 114,7647-7652.
- Subalusky, A.L., Post, D.M. 2019. Context dependency of animal resource subsidies. Biol Rev
  Camb Philos Soc 94, 517-538.
- 660 Team, R.C., 2017. R: A language and environment for statistical computing (Version 3.4.
- 661 2)[Computer software]. Vienna, Austria: R Foundation for Statistical Computing.
- 662 Thorp, J.H., Delong, M.D., 1994. The Riverine Productivity Model: An Heuristic View of
- 663 Carbon Sources and Organic Processing in Large River Ecosystems. Oikos 70, 305-308.
- 664 Thorp, J.H., Delong, M.D., 2002. Dominance of autochthonous autotrophic carbon in food
- 665 webs of heterotrophic rivers. Oikos 96, 543-550.
- 666 Thorp, J.H., Thoms, M.C., Delong, M.D., 2006. The riverine ecosystem synthesis:
- 667 biocomplexity in river networks across space and time. River Research and
- 668 Applications 22, 123-147.
- Vanni, M. J., 2002. Nutrient cycling by animals in freshwater ecosystems. Annual Review of
  Ecology and Systematics 33, 341-370.
- Vannote, R. L., Minshall, G.W., Cummins, K.W., Sedell, J.R., Cushing, C.E. 1980. The river
- 672 continuum concept. Canadian journal of fisheries and aquatic sciences 37, 130-137.

673	Veldhuis, M.P.	, Ritchie, M.E., C	Ogutu, J.O., Morrison,	T.A., Beale, C.M.,	Estes, A.B. Mwakilema,

- 674 W., Ojwang, G.O., Parr C.L., Probert J. 2019. Cross-boundary human impacts
- 675 compromise the Serengeti-Mara ecosystem. Science 363, 1424-1428.
- 676 Webster, J.R., Benfield, E.F., Ehrman, T.P., Schaeffer, M.A., Tank, J.L., Hutchens J.J.. D'Angelo
- D.J., 1999. What happens to allochthonous material that falls into streams? A synthesis
- of new and published information from Coweeta. Freshwater Biology 41, 687-705.
- 679 Wipfli, M. S., Richardson, J. S., Naiman, R. J., 2007. Ecological Linkages Between Headwaters
- and Downstream Ecosystems: Transport of Organic Matter, Invertebrates, and Wood
- 681 Down Headwater Channels 1. JAWRA Journal of the American Water Resources
- 682 Association 43, 72-85.
- 683 Wood, S. Wood, M. S., 2015. Package 'mgcv'. R package version: 1-7.
- 684 Wood, S. N., 2017. Generalized additive models: an introduction with R, CRC press.

## 685 Figure captions

**Figure 1**. Map of the Mara River catchment showing the position of the sampling sites.

**Figure 2**. Longitudinal trends in the mean (±SD) of (a)  $\delta^{13}$ C and (b)  $\delta^{15}$ N values (‰) of

688 periphyton with increasing stream size in the Mara River and its tributaries. FOR = forested

sites, AGR = agriculture sites, LD\_Livestock = low density livestock sites, HD\_Livestock = high

- 690 density livestock sites, and Hippos = hippopotamus sites.
- 691 **Figure 3**. Box-and-whisker plots of fractional contributions of C3 vegetation (grey), C4
- 692 grasses (light grey) and periphyton (dark grey) to all macroinvertebrates FFGs in the Mara
- River. (a) all taxa, (b) collector-gatherers, (c) predators, (d) collector-filterers (e) shredders,
- and (f) scrapers. C3 = C3 plants; C4 = C4 plants.
- 695 Figure 4. Longitudinal trends in source contributions to macroinvertebrates along the Mara 696 River. Longitudinal variability in the relative importance (fractional contribution) of C3 697 vegetation (sky blue), C4 grasses (orange) and periphyton (dark green) as sources of carbon 698 for macroinvertebrates in the Mara River in response to changes in (a) river distance from 699 the source (RDS in km) and (b) density of large mammalian herbivores (LMH Density). To test 700 the significance of the relationships, we fitted a GAM model with a smoothing function. The black line with shaded area represents smoother mean and s.e.; smoother significance,  $R^2$ 701 702 and GCV are supplied in the figures.

**Figure 5**. Source contributions to FFGs along the Mara River. Longitudinal trends in fractional

contributions of trophic source (C3= C3 vegetation, C4 = C4 grasses and periphyton) to

705 macroinvertebrate functional feeding groups (FFGs) in the Mara River. Source contributions

706	are assessed in res	ponse to change	s in river d	istance from	source (RDS	) as a measure of

stream size (a, c, e, g, i) and density of large mammalian herbivores (LMH), b, d, f, h, j). a, b =

collector-filterers, c, d = collector-gatherers, e, f = scrapers, g, h = shredders, and I, j =

- predators. To test the significance of the relationships, we fitted a GAM model with a
- smoothing function. The black line with shaded area represents smoother mean and s.e.;
- smoother significance,  $R^2$  and GCV are supplied in the figures. Note changes on the x-axis
- 712 and y-axis.

**Table 1.** River size, the density of large mammalian herbivores, catchment land use characteristics and  $\delta^{13}$ C and  $\delta^{15}$ N values of the main carbon

sources (periphyton, C3 and C4 carbon, dung) across 80 sites of the Mara River, Kenya, grouped into five regions: Forested, Agricultural, with

<sup>716</sup> low density (LD) livestock, with high density (HD) livestock, and with a dominance of hippos. All values are means (± SD) and  $\delta^{13}$ C and  $\delta^{15}$ N

717 values are in ‰ units.

		Regions					
Characteristics	Forested	Agricultural	LD livestock	HD livestock	Hippos	F - value	p - value
Stream and sub-catchment characteri	stics						
Stream order	4.0±1.5 °	3.7±1.6ª	4.8±2.1 <sup>ab</sup>	4.0±1.9 <sup>ª</sup>	$6.4 \pm 1.6^{b}$	5.26	0.001
River distance from source (RDS)	12.5±6.4 °	12.2±9.2 ª	26.0±17.4 <sup>b</sup>	17.2±15.8 <sup>ªb</sup>	52.6±23.2 <sup>b</sup>	7.37	0.001
LMH density (individuals / km²)	9.3±4.1ª	28.0±16.8 <sup>b</sup>	44.8±12.5 <sup>b</sup>	102.9±17.7 <sup>°</sup>	111.9±10.8 <sup>°</sup>	11.21	0.001
% Agriculture	27.1±24.9ª	61.1±22.2 <sup>b</sup>	63.5±23.9 <sup>b</sup>	1.9±0.5°	22.4±17.3 <sup>ªb</sup>	18.91	0.001
% Grasslands	5.6±5.2ª	7.2±5.1ª	16.5±13.9 <sup>b</sup>	59.9±10.8 <sup>°</sup>	49.2±16.0 <sup>c</sup>	93.21	0.001
% Forest	64.7±28.5ª	39.3±22.9 <sup>b</sup>	25.4±15.4 <sup>b</sup>	37.2±11.5 <sup>b</sup>	27.97±8.9 <sup>b</sup>	9.47	0.001
River width (m)	7.9±5.0ª	7.1±6.1ª	8.7±5.8ª	6.6±3.5ª	15.84±9.9 <sup>b</sup>	4.35	0.003
Water depth (m)	0.20±0.1ª	0.2±0.1ª	0.2±0.2ª	0.2±0.1ª	$1.0\pm0.4^{b}$	5.45	0.001
River discharge (m <sup>3</sup> /s)	0.23±0.2ª	0.4±0.2ª	0.5±0.4ª	0.3±0.2ª	17.4±6.9 <sup>b</sup>	7.57	0.001
pH (units)	7.6±0.3ª	7.6±0.3ª	7.7±0.2ª	7.6±0.1ª	7.8±0.0ª	0.40	0.804
Temperature (°C)	15.8±1.8ª	18.5±3.2 <sup>b</sup>	20.3±2.5 <sup>b</sup>	23.7±2.1 <sup>°</sup>	23.9±2.2°	27.9	0.001
Dissolved oxygen (mgL <sup>-1</sup> )	7.8±0.4ª	7.0±1.1 <sup>bc</sup>	6.3±0.9 <sup>°</sup>	7.3±1.0 <sup>ªb</sup>	8.0±1.1ª	7.36	0.001
Electrical conductivity (µScm <sup>-1</sup> )	74.1±29.4ª	103.2±50.0ª	260.3±128.9 <sup>b</sup>	309.0±172.8 <sup>b</sup>	325.0±178.7 <sup>b</sup>	18.12	0.001
Stable isotopes of basal resources							
$\delta^{13}$ C of C3 vegetation	-26.56±2.08ª	-27.14±3.37ª	-27.62±1.37ª	-25.36±1.98ª	-26.13±1.97ª	1.05	0.407
$\delta^{13}$ C of C4 vegetation	-13.22±6.27ª	-12.08±1.56ª	-16.44±7.95ª	-11.04±2.13ª	-15.64±11.01ª	1.32	0.294
$\delta^{13}$ C of periphyton	-25.93±2.58ª	-24.34±2.64 <sup>ªb</sup>	-23.7±2.15 <sup>ab</sup>	-23.21±1.23 <sup>b</sup>	-22.66±1.80 <sup>b</sup>	4.24	0.004
$\delta^{13}$ C of cattle dung	-20.96±7.31ª	-15.78±2.36 <sup>ªb</sup>	-13.49±1.73 <sup>b</sup>	-14.27±5.01 <sup>b</sup>	-13.58±2.85 <sup>b</sup>	5.08	0.002
$\delta^{13}$ C of hippo dung	-	-	-	-11.89±2.14ª	-14.13±2.72ª	2.11	0.169
$\delta^{15}$ N of C3 vegetation	4.06±1.69ª	5.47±4.96ª	5.24±2.65ª	4.61±2.36ª	3.37±2.91ª	1.83	0.119
$\delta^{15}$ N of C4 vegetation	6.09±2.59 <sup>ª</sup>	5.98±3.07ª	4.76±4.79ª	5.57±3.25ª	4.81±2.74 <sup>ª</sup>	0.15	0.988
$\delta^{15}$ N of periphyton	7.13±0.84 <sup>ª</sup>	7.56±1.49 <sup>ab</sup>	8.53±1.44 <sup>b</sup>	8.11±0.79 <sup>b</sup>	7.29±0.71 <sup>ab</sup>	3.16	0.019
$\delta^{15}$ N of cattle dung	6.79±1.79 <sup>ª</sup>	5.37±1.29 <sup>ª</sup>	7.29±0.94ª	5.89±1.29 <sup>ª</sup>	4.55±0.20 <sup>ª</sup>	1.34	0.257
$\delta^{15}$ N of hippo dung		-	-	6.56±1.96 <sup>ª</sup>	4.93±0.63 <sup>ª</sup>	2.19	0.158

- 719 **Table 2**. Summary of generalized additive mixed models (GAMs) of the effects of stream size (river distance from source {RDS}), region and
- 720 density of large mammalian herbivores (LMH density) (stream size) on the relative importance (fractional contribution) of C3 vegetation (C3
- carbon), C4 grasses (C4 carbon) and periphyton as sources of carbon/energy for macroinvertebrate FFGs (collector-filterers, collector-
- gatherers, scrapers, predators and shredders) and all combined taxa (FFGs) in the Mara River, Kenya. RDS, region and LMH density were
- treated as fixed factors while each study site was used a random factor. In all cases, d.f. = 1.

Source Contributions							
Sources	Collector-filterers	Collector-gatherers Scrapers		Predators	Shredders	All taxa	
C3 carbon							
RDS F <i>, p-</i> value	21.21, <0.001***	17.01, <0.001***	21.14, <0.001***	12.48, <0.001***	0.26, 0.612	60.1, <0.001***	
Region F <i>, p-</i> value	10.47 <i>,</i> 0.002**	23.95 <i>,</i> <0.001***	24.32 <i>,</i> <0.001***	5.02 <i>,</i> 0.028*	0.053, 0.819	39.7 <i>,</i> <0.001***	
LMH density F, <i>p</i> -value	1.30, 0.260	3.32, 0.073	3.62, 0.062	0.02, 0.878	1.65, 0.208	6.12, 0.014*	
RDS x region F, <i>p</i> -value	2.15, 0.150	1.80, 0.185	3.45, 0.068	3.87, 0.053	1.51, 0.228	9.11, 0.003**	
RDS x LMH density F, <i>p</i> -value	0.09, 0.761	0.04, 0.846	0.001, 0.972	1.52, 0.222	1.72, 0.199	0.16, 0.691	
Region x LMH density F, <i>p</i> -value	0.03, 0.857	0.003, 0.957	0.04, 0.848	5.54, 0.022*	0.83, 0.371	0.24, 0.624	
RDS x region x LMH density F, <i>p</i> -value	0.07, 0.796	0.06, 0.805	0.001, 0.972	0.01, 0.905	0.058, 0.812	0.02, 0.887	
Adj. R2	0.692	0.365	0.401	0.227	-0.024	0.271	
Scale est.	0.001	0.0004	0.0005	0.001	0.019	0.014	
Residual deviance	0.68	0.949	0.903	0.817	0.774	5.44	
C4 carbon							
RDS F, <i>p</i> -value	67.10, <0.001***	95.89, <0.001***	120.7, <0.001***	129.6, <0.001***	0.001, 0.969	481.1, <0.001***	
Region F <i>, p-</i> value	45.18, <0.001***	60.52, <0.001***	74.21 <i>,</i> <0.001***	111.7, <0.001***	0.60, 0.445	272.3, <0.001***	
LMH density F, p-value	1.13, 0.293	1.28, 0.262	3.45, 0.068	11.24, 0.001**	4.43, 0.043*	13.7, <0.001***	
RDS x region F, <i>p</i> -value	1.58, 0.215	1.63, 0.207	0.55, 0.463	7.24, 0.009**	3.00, 0.093	9.82, 0.002**	
RDS x LMH density F, <i>p</i> -value	0.56, 0.460	0.80 <i>,</i> 0.376	1.78, 0.187	18.06, <0.001***	0.09, 0.766	13.5 <i>,</i> <0.001***	
Region x LMH density F <i>, p-</i> value	7.22, 0.010*	5.21, 0.026*	6.25 <i>,</i> 0.015*	3.36, 0.072	0.36, 0.550	30.3 <0.001***	
RDS x region x LMH density F, <i>p</i> -value	0.96, 0.333	0.03, 0.874	1.74, 0.192	8.85 <i>,</i> 0.004**	3.92, 0.056	12.1 <0.001***	
Adj. R2	0.13	0.699	0.747	0.795	0.121	0.739	
Scale est.	0.031	0.002	0.002	0.001	<0.001	0.004	
Residual deviance	0.51	0.351	0.300	0.315	0.066	1.43	

Periphyton						
RDS F, p-value	0.05, 0.829	2.87, 0.095	2.24, 0.140	10.57, 0.002**	0.28, 0.599	10.9, 0.001**
Region F, <i>p-</i> value	0.10, 0.754	0.0004, 0.984	0.06 <i>,</i> 0.805	15.05, <0.001***	0.0003 <i>,</i> 0.976	4.5, 0.034*
LMH density F <i>, p-</i> value	1.74, 0.194	1.48, 0.229	0.58, 0.449	3.08, 0.084	0.56, 0.458	0.20, 0.653
RDS x region F, <i>p-</i> value	4.29 <i>,</i> 0.043*	2.96 <i>,</i> 0.090	3.41 <i>,</i> 0.069	11.39 <i>,</i> 0.001**	0.66, 0.424	18.4, <0.001***
RDS x LMH density F <i>, p</i> -value	0.05, 0.816	0.89, 0.348	0.49 <i>,</i> 0.483	1.49, 0.227	2.68, 0.111	3.03, 0.085
Region x LMH density F, <i>p</i> -value	1.42, 0.247	1.32 <i>,</i> 0.256	2.43, 0.125	1.22, 0.273	0.75, 0.392	5.14 <i>,</i> 0.024*
RDS x region x LMH density F, p-value	0.04, 0.841	0.08, 0.771	0.30, 0.584	3.32, 0.073	0.208, 0.651	0.172, 1.91
Adj. R2	0.013	0.037	0.035	0.992	-0.050	0.113
Scale est.	0.002	0.002	0.002	0.001	0.014	0.015
Residual deviance	1.01	1.14	1.15	0.349	0.571	5.95

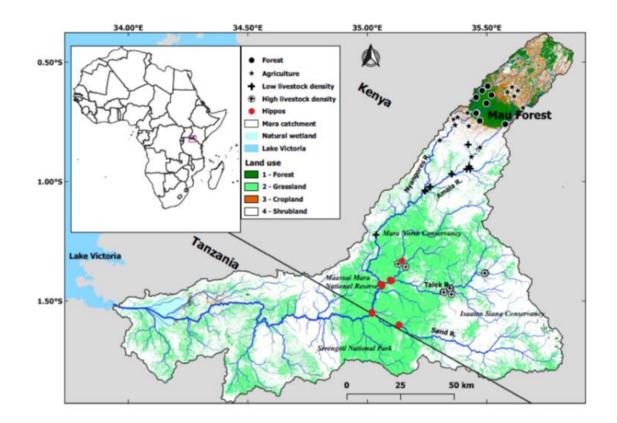


Figure 1

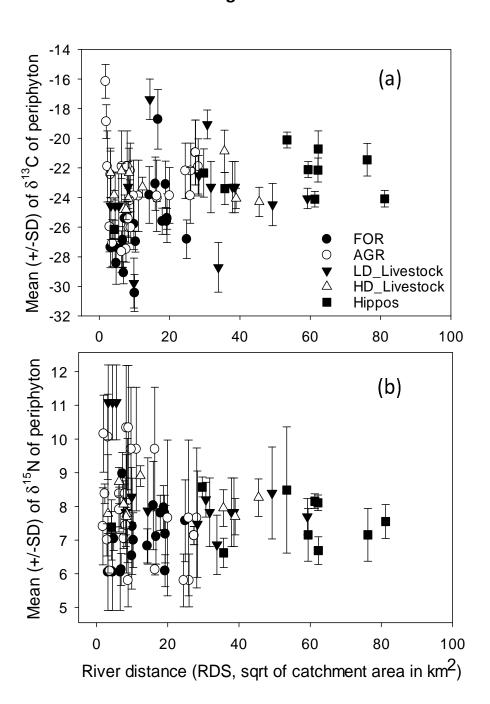
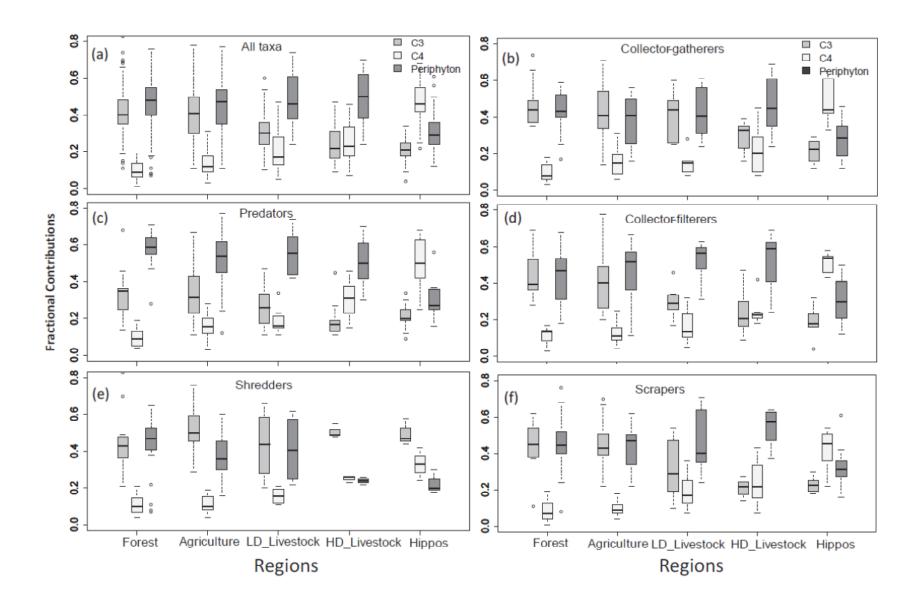


Figure 2



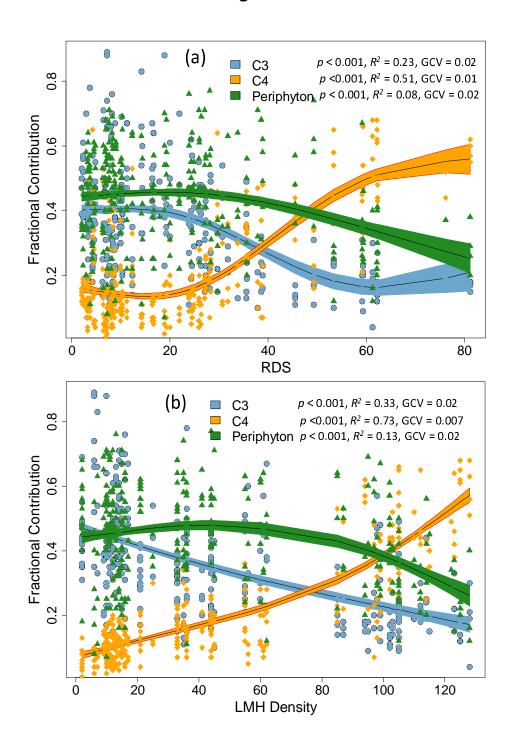


Figure 4

