bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

Clostridium difficile Toxin B activates the NLRP3 inflammasome in human macrophages, demonstrating a novel regulatory mechanism for the Pyrin inflammasome

Short title: Novel regulatory mechanism for Pyrin inflammasome activation in human macrophages

Matthew S.J. Mangan[1,2], Friederike Gorki[1*], Karoline Krause[3*], Alexander Heinz[4], Thomas Ebert[5], Dieter Jahn[6,7], Karsten Hiller[4,8], Veit Hornung[5], Marcus Mauer[3], Florian I. Schmidt[1], Ralf Gerhard[9], Eicke Latz[1,2,10,11]

1. Institute of Innate Immunity, University Hospital Bonn, University of Bonn, 53127 Bonn, Germany.
2. German Center for Neurodegenerative Diseases, 53127, Bonn, Germany.
3. Dermatological Allergology, Allergie-Centrum-Charité, Department of Dermatology and Allergy, Charité – Universitätsmedizin Berlin, Berlin, Germany.
4. Department of Bioinformatics and Biochemistry, Braunschweig Integrated Center of Systems Biology (BRICS), Technische Universität Braunschweig, Braunschweig, Germany.
5. Gene Center and Department of Biochemistry, Ludwig-Maximilians-Universität München, 81377 Munich, Germany.
6. Institute for Microbiology, Technische Universität Braunschweig, Braunschweig, Germany.

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

7. Braunschweig Integrated Center of Systems Biology (BRICS), Technische Universität Braunschweig, Braunschweig, Germany.
8. Computational Biology of Infection Research, Helmholtz Centre for Infection Research, Braunschweig, Germany.
9. Institute of Toxicology, Hannover Medical School, 30625 Hannover, Germany.
10. Department of Infectious Diseases & Immunology, UMass Medical School, Worcester, MA 01605, USA.
11. Centre of Molecular Inflammation Research, Norwegian University of Science and Technology, 7491 Trondheim, Norway.

* contributed equally

Correspondence should be addressed to: Eicke Latz (eicke.latz@uni-bonn.de) or Matthew S.J. Mangan (matthew.mangan@ukbonn.de)

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

Abstract

Pyrin is a cytosolic immune sensor that forms an inflammasome when bacterial virulence factors inhibit RhoA, triggering the release of inflammatory cytokines, including IL-1β. Gain of function mutations in the MEFV gene encoding Pyrin cause auto-inflammatory disorders, such as familial Mediterranean fever (FMF) and Pyrin associated auto-inflammation with Neutrophilic Dermatosis (PAAND). To precisely define the role of Pyrin in detecting pathogen virulence factors in relevant human immune cells, we investigated how the Pyrin inflammasome response was initiated and regulated in monocyte-derived macrophages (hMDM) compared to human monocytes. Unlike monocytes and murine macrophages, we determined that hMDM failed to activate Pyrin in response to known Pyrin activators *Clostridioides difficile* (*C. difficile*) toxins A or B (TcdA or TcdB). In contrast, TcdB activated the NLRP3 inflammasome in hMDM. Notably, we ascertained that the Pyrin inflammasome response could be re-enabled in hMDM by prolonged priming with either LPS or type I or II interferons, and required an increase in Pyrin expression. These data demonstrate an unexpected redundancy in detecting these toxins by inflammasome sensors.

Introduction

Inflammasome-forming proteins are cytosolic sensors that mediate a post-translational inflammatory response to pathogens or cell stress. Upon detecting a stimulus, these sensors oligomerize and recruit the adapter protein ASC,

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

enabling activation of caspase-1. Active caspase-1 then mediates cleavage and release of pro-inflammatory cytokines including IL-1β, and pro-inflammatory cell death by cleaving the pore-forming molecule gasdermin D [1]. Notably, rather than relying solely on direct detection of pathogens, some inflammasome sensors have instead evolved to detect infection or cellular stress by monitoring disruption of cellular homeostasis [2]. A leading example of this is the Pyrin inflammasome, encoded by the *MEFV* gene, which is activated in response to inhibition of RhoA [3]. This small G protein controls cytoskeletal rearrangement and is essential for immune cell migration and phagocytosis, amongst other functions [4]. Unsurprisingly, given its role in fundamental cellular processes, RhoA is a target of numerous bacterial virulence factors from pathogenic bacteria, including *Clostridium difficile* toxins A and B (TcdA and TcdB, respectively) [5]. Another prominent sensor of homeostasis is the NLRP3 inflammasome, which detects a wide range of events that converge to cause either mitochondrial dysfunction or loss of osmotic control of the cytosol [6].

Due to their high inflammatory potential, inflammasome forming sensors are strictly regulated by post-translational controls. Under resting conditions, Pyrin is maintained in an inactivate state by two distinct mechanisms regulated by RhoA signaling. The most well-characterized of these is phosphorylation of Pyrin at residues Ser208 and Ser242 by PKN1/2, members of the PKC superfamily [7–10]. Phosphorylation of Pyrin enables the subsequent binding of 14-3-3 proteins, which sequesters Pyrin in an inactive state. The second mechanism regulating Pyrin activation is less well understood, but is defined by

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

a requirement for microtubule dynamics [9]. Pretreatment with colchicine, a microtubule destabilization agent, inhibits Pyrin activation but does not prevent dephosphorylation of Pyrin, demonstrating that these two regulatory mechanisms are mutually exclusive. Interestingly, human and mouse Pyrin share these regulatory mechanisms, although murine Pyrin does not contain the C-terminal B30.2 domain due to a frameshift mutation [11].

Understanding the regulatory mechanisms governing Pyrin is particularly important as mutations in the *MEFV* gene cause the hereditary auto-inflammatory disorder, Familial Mediterranean Fever (FMF). FMF is characterized by recurrent attacks of fever, serositis, abdominal pain and can, over time, cause secondary AA amyloidosis, leading to kidney failure [12]. Mutations linked to FMF are mostly amino acid substitutions in the C-terminal B30.2 domain [13]. Though the function of the B30.2 domain is still relatively unclear, these mutations are suggested to perturb the colchicine-dependent regulatory mechanism of Pyrin. This is supported by studies showing that Pyrin variants containing FMF mutations are resistant to inhibition by colchicine and that dephosphorylation is sufficient to activate the FMF form of Pyrin [14,15]. However, as the B30.2 domain has been lost in the mouse, it is difficult to model this disease. Thus, there is a need for further research into the effects of these mutations in human cell types to understand how they alter Pyrin regulation.

C. difficile infection is a leading cause of hospital-associated mortality through diarrhea triggered by antibiotic therapy-mediated dysbiosis and

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

126 pseudomembranous colitis. All of these effects are entirely dependent on the
127 expression of TcdA and TcdB, although TcdB is sufficient to cause disease [16].
128 Yet, the role of the inflammasome in the immune response to *C. difficile*
129 infection is controversial. *In vivo* experiments in ASC KO mice examining the
130 effect of TcdA and TcdB directly, rather than through *C. difficile* infection,
131 demonstrated that inflammasome activation in response to these toxins
132 increased tissue damage in an IL-1β dependent manner [17]. However, a
133 subsequent study demonstrated a role for caspase-1 in controlling *C. difficile*
134 infection [18]. Experiments in both NLRP3 KO and Pyrin KO mice found that
135 neither sensor impacted the severity of the disease [18,19]. It is conceivable
136 that the pathology of the human pathogen *C. difficile* is not fully recapitulated in
137 the murine model. Therefore, investigations into the inflammasome response
138 to *C. difficile* in human cells are very important, and have so far included
139 PBMCs, monocytes, and neutrophils [11].
140
141 In this study, we assessed the inflammasome response to *C. difficile* and its
142 toxins, TcdA and TcdB, in a human macrophage model using M-CSF
143 monocyte-derived macrophages. We discovered that only TcdB triggered an
144 inflammasome response in these cells and that, in contrast to monocytes, this
145 was entirely independent of Pyrin and instead occurred through NLRP3.
146 Furthermore, we found that prolonged exposure to LPS or type-I or –II
147 interferons was sufficient to reactivate the Pyrin inflammasome by increasing
148 Pyrin expression. These results demonstrate that Pyrin is held in an inactive
149 state when monocytes differentiate to macrophages by a hitherto
150 uncharacterized regulatory mechanism.

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

Results

Inflammasome activation by *C. difficile* in hMDM is dependent on the expression of its toxins and can be blocked by NLRP3 inhibition

To investigate whether *C. difficile* or its secreted virulence factors could elicit an inflammasome response in primary hMDM, we treated the cells with either conditioned supernatant from *C. difficile* or infected hMDM with the bacteria directly. As production of TcdA and TcdB is crucial for both *C. difficile*-driven pathology and the activation of the Pyrin inflammasome, two different strains of *C. difficile* were used: one proficient for production of both toxins A and B, and one deficient for both. We also assessed the effect of prior exposure to TLR ligands on the inflammasome response by priming one hMDM group with LPS before exposure to the supernatant or bacteria. In addition, we pre-incubated one group of cells with the potent and highly specific NLRP3 inhibitor CP-456,773 (also known as CRID3 or MCC950) [20] to determine if the response was NLRP3 dependent.

We determined that both *C. difficile* conditioned supernatant and direct infection induced the release of two inflammasome-dependent cytokines, IL-1β and IL-18, from hMDM. The cytokine release was entirely toxin-dependent in the cells treated with the conditioned supernatant (Fig. 1a) and primarily reliant on toxin expression when the cells were infected directly (Fig. S1a). Notably, IL-18, which is transcribed independent of TLR stimulation [21], was still secreted

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

exclusively when the cells were pretreated with LPS. The secretion of both cytokines was inhibited by CP-456,773 [20]. Thus, both the requirement for pre-priming and sensitivity to CP-456,773 suggested that NLRP3 mediated the response to *C. difficile* in hMDM.

Figure 1. TcdB, but not TcdA, triggers CP-456,773 sensitive IL-1β release in hMDM but not monocytes

(A) hMDM incubated with conditioned supernatant from toxin proficient (TcdA+,TcdB+) or toxin deficient (TcdA-,TcdB-) *C. difficile* for 4h, then the supernatant assessed for IL-1β and IL-18. (B) LPS primed (10ng/ml, 3h) monocytes or hMDM were treated with a dose titration of either TcdB or TcdA for 3h. Supernatants were harvested and assessed for IL-1β. (C) Immunoblot of lysate from LPS treated (10ng/ml, 3h) monocytes or hMDM probed for either Pyrin or actin. Representative of 3 independent experiments. (D) LPS primed (10ng/ml, 3h) monocytes or hMDM were pre-incubated for 15 min with the vehicle alone, CP-456,773 (2.5 μM), VX-765 (40 μM) or colchicine (2.5 μM) then stimulated with TcdB (20 ng/ml), Nigericin (8 μM) or LFN-PrgI/PA (25 ng/ml) for 2.5h. The supernatant was assessed for IL-1b and LDH activity or (E) harvested and precipitated and the cells lysed. Both the precipitated supernatant and cell lysate were analysed by immunoblot for IL-1β and caspase-1. Mean and SEM shown for three donors or immunoblots representative of 3 independent experiments.

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

C. difficile toxin B, but not toxin A, mediates NLRP3-dependent inflammasome activation in hMDM

We observered that hMDM released IL-1β and IL-18 only in response to the toxin-proficient bacteria. To determine whether one or both toxins could trigger an inflammasome response, we incubated the cells with either recombinant toxin A (TcdA) or toxin B (TcdB). As a control, we incubated monocytes with both toxins, as they have previously been shown to respond to both in a Pyrin inflammasome-dependent manner. Monocytes, as predicted, released IL-1β in response to both TcdA and TcdB (Fig. 1b). Surprisingly, and in contrast to the monocytes, we found that TcdB, but not TcdA, induced an inflammasome response in the hMDM (Fig. 1b). Notably, TcdB triggered IL-1β release from hMDM at concentrations as low as 1 ng/ml. To ensure that the lack of TcdA-mediated inflammasome activation was not due to a failure of toxin uptake, we assessed TcdA mediated Rac modification. To do so, we used a previously described monoclonal antibody that no longer recognizes Rac when its epitope is modified by the toxin [22]. The antibody was unable to bind to Rac in both monocytes and hMDM treated with the active forms of TcdB and TcdA, but not with mutants lacking glucosyltransferase activity, indicating that the cells took up the toxin (Fig. S1b).

Given the disparity in inflammasome response between monocytes and hMDM, we assessed whether Pyrin was differentially expressed in the two cell types. Pyrin levels were comparable in hMDM and monocytes (Fig. 1c), indicating that Pyrin expression was not limiting.

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

To further investigate the TcdB-mediated inflammasome response, we pre-incubated LPS primed hMDM or monocytes with inhibitors against NLRP3 (CP-456,773), Pyrin (colchicine) or caspase-1 (VX-765) and then treated the cells with TcdB. As specificity controls, we used Nigericin, a potassium ionophore that activates NLRP3, as well as the NLRC4 activator needletox. The latter contains the *Salmonella typhimurium* T3SS needle protein PrgI fused to the N-terminus of anthrax lethal factor, which is delivered to the cytosol by anthrax protective antigen (PA). In hMDM, we determined that both TcdB dependent IL-1β secretion and LDH release as a measure of pyroptosis were inhibited by CP-456,773 but not colchicine. This provides further evidence that TcdB is activating NLRP3 rather than Pyrin in these cells (Fig. 1d). In contrast, and as demonstrated previously, the response to TcdB in monocytes was not inhibited by CP-456,773 but was inhibited by colchicine, indicating a dependence on Pyrin (Fig. 1d). To ensure that the observed IL-1β release was accompanied by caspase-1 activation, we also assessed the cleavage of IL-1β and caspase-1. In agreement with the IL-1b and LDH release results, we found that TcdB-mediated IL-1β and caspase-1 cleavage were CP-456,773 sensitive in hMDM, but not in monocytes (Fig. 1e). We also monitored the actin cytoskeleton after treatment by TcdB by staining the cells with Phalloidin. As predicted, TcdB disrupted the actin cytoskeleton similarly in both cell types (Fig. S1c).

The inflammasome response to TcdB in human macrophage cell lines is entirely dependent on NLRP3

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

Our results thus far demonstrated that NLRP3 is the responding inflammasome sensor to TcdB in hMDM, but relied solely on compound-based inhibition. Therefore, we used genetically modified human macrophage cell lines to determine whether NLRP3 or Pyrin mediates the inflammasome response to TcdB. Initially, we sought a model cell line that had a robust inflammasome response to TcdB. We investigated whether the BLaER1 cell line, a recently established monocyte/macrophage cell line [23], would similarly respond to TcdB to the hMDM. We found that WT BLaER1 cells released IL-1β in response to treatment with TcdB, as well as activating caspase-1 as measured by a caspase-1 activity assay (Fig. 2a). These data demonstrate that TcdB activates an inflammasome response in BLaER1 similar to hMDM.

Figure 2. NLRP3, not Pyrin, is the responding inflammasome sensor to TcdB in human macrophages

(A) Differentiated WT BLaER1 cells were primed with LPS (100 ng/ml, 3h), pre-incubated with either CP-456,773 (2.5 μM, 15 min), then activated with Nigericin (8 μM), TcdB (20ng/ml) or with LFN-PrgI/PA (25 ng/ml each) for 2h. IL-1β and caspase-1 activity were assessed from the harvested supernatants. (B) ASC-BFP transduced WT BLaER1 cells treated as in (A). Cells were then fixed and the number of ASC specks quantified by microscopy. (C) NLRP3 expression in differentiated BLaER1 cells (+/- 100ng/ml LPS 3h) was assessed by immunoblot. (D) Differentiated Casapse-4, NLRP3 double deficient BLaER1 cells reconstituted with either NLRP3, the NLRP3 walker A/B mutant (NLRP3 WAB) or the vector control treated as in (A) and the supernatants assessed for

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

IL-1β or TNFα. Mean and SD of three technical replicates shown, representative of 3 independent experiments. (E) Immunoblot of Pyrin expression in differentiated BLaER1 cells. Representative of 3 independent experiments. (F) Differentiated Pyrin deficient BLaER1 cells reconstituted with either Pyrin or the vector control treated as in (A) and the supernatants assessed for IL-1β or TNFα. Mean and SD of three technical replicates shown, representative of 3 independent experiments. (G) LPS primed WT THP-1s or the listed KOs were activated with inflammasome activators for either 1.5h (Nigericin, LFN-PrgI/PA) or for 8h (TcdB). Supernatants were assessed for IL-1β. Where used, CP-456,773 (2.5 μM) and VX-765 (40 μM) were pre-incubated with the cells for 15 min prior to addition of the inflammasome activators. Mean and SEM of 3 independent experiments shown.

We next used the BLaER1 cell line to determine the propensity of TcdB to trigger ASC speck formation, another hallmark of inflammasome activation. Accordingly, we generated a BLaER1 cell line overexpressing ASC-mCherry and stimulated it with either TcdB or Nigericin in the presence or absence of CP-456,773. This experiment was also performed in the presence of the caspase-1 inhibitor VX-765 to prevent cell death of inflammasome-activated cells. Following stimulation, the cells were fixed, and the number of ASC specks was quantified by microscopy, followed by normalization to the number of nuclei in each image. Consistent with our other data, we found that TcdB and Nigericin also caused CP456,773-sensitive ASC speck formation in these cells (Fig. 2b).

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

299 To determine the sensor responsible for the TcdB mediated inflammasome
300 response, we used either NLRP3 or Pyrin knock-out BLaER1 cells. As the
301 NLRP3 KO cells we used were on a caspase-4 KO background, we first
302 ensured that caspase-4 played no role in the response. Accordingly, we tested
303 the inflammasome response to TcdB in the caspase-4 KO BLaER1 cells, but
304 found no difference in IL-1β secretion, demonstrating caspase-4 was not
305 required for the response to TcdB (Fig. S2a). The NLRP3, caspase-4 double
306 KO cells were then reconstituted with either NLRP3, the inactive Walker A/B
307 NLRP3 mutant, or a vector alone control. NLRP3 expression was confirmed by
308 immunoblot (Fig. 2c). As previously, we primed these cells with LPS, incubated
309 them with either TcdB, Nigericin or needletox, and assessed IL-1β release to
310 the supernatant. In agreement with our findings in hMDM, only the cells
311 reconstituted with active NLRP3, but not the walker A/B mutant, were able to
312 respond to TcdB and Nigericin (Fig. 2d). In contrast, all cell lines responded
313 equally to the NLRC4 trigger needletox (Fig. 2d) and secreted similar levels of
314 TNFa in response to LPS (Fig. 2d). These results confirm that the TcdB-
315 mediated inflammasome response was dependent on the expression of active
316 NLRP3.
317
318 To confirm that Pyrin was not required for the TcdB-driven inflammasome
319 response, we reconstituted Pyrin KO BLaER1 cells with either Pyrin or vector
320 control, and confirmed expression levels by immunoblot (Fig. 2e). We found the
321 inflammasome response to TcdB was unaffected by the absence of Pyrin,
322 indicating that Pyrin does not play a role in the inflammasome response to TcdB

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

in these cells (Fig. 2f). Similarly, Pyrin expression did not affect the inflammasome response to Nigericin or needletox (Fig. 2f).

BLaER1 cells can readily secrete IL-1β in response to TLR4 stimulation by LPS alone. To rule out that LPS contamination of the TcdB was responsible for the observed IL-1β release, we stimulated the cells with TcdB or Nigericin following pre-incubation of the cells with TAK-242, a TLR4 inhibitor. TAK-242 did not block TcdB-mediated IL-1β release (Fig. S2b), but TAK-242 completely abolished LPS mediated TNFa secretion when applied before LPS stimulation (Fig. S2c), demonstrating that NLRP3 mediated activation by TcdB is not due to LPS contamination or TLR4 activation.

Having established the requirement for NLRP3 in response to TcdB in the BLaER1 cell line, we sought to determine if this was true in the other commonly used human macrophage cell line, THP-1. We first titrated TcdB on PMA differentiated THP-1 cells and found that it required both a higher concentration of TcdB (2ug/ml) to trigger IL-1β release as well as a longer incubation time (8h). Having established this we tested several THP-1 CRISPR knock-out cell lines to determine the requirements for TcdB-mediated inflammasome activation. PMA-differentiated WT THP-1 cells or THP-1 cells deficient in ASC, caspase-1, NLRP3 or Pyrin were primed with LPS (200 ng/ml, 3h), then incubated with TcdB, Nigericin and needletox. WT cells were also pre-incubated with CP-456,773 to determine if it had the same effect as ablation of NLRP3. Echoing our previous data, the TcdB-triggered inflammasome response in THP-1 cells was CP-456,773-sensitive and NLRP3-dependent

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

(Fig. 2g). As anticipated, this response also required ASC and caspase-1, but not Pyrin. Nigericin similarly was NLRP3-dependent, while the NLRC4 trigger only required ASC and caspase-1, demonstrating the effect of NLRP3 ablation was specific (Fig. 2g). Furthermore, the different KO lines secreted similar amounts of TNFα in response to LPS (Supp. Fig 2d). Collectively, these results show an absolute requirement for NLRP3, but not Pyrin, in the TcdB-mediated inflammasome response in human THP-1 cells.

Having established that TcdB activates NLRP3, but not Pyrin, we next determined whether NLRP3 activation required the activity of the TcdB glucosyltransferase domain (GTD) against Rho, as found for Pyrin. Thus, we incubated LPS-primed hMDM with TcdB or a variant of TcdB containing inactivating mutations in the glucosyltransferase domain, D286N and D288N (TcdB NXN), which does not inactivate Rho, Rac or Cdc42. We determined that both TcdB and TcdB NXN induced IL-1β release (Supp. fig. 2e). Interestingly, the NXN variant of TcdB was a more proficient inflammasome activator than the WT toxin, suggesting that the activity of the GTD domain opposes TcdB-mediated NLRP3 activation. This demonstrates that in contrast to Pyrin activation, GTD activity, and subsequent Rho inhibition are not required for the TcdB mediated inflammasome response in hMDM.

TcdA and TcdB elicit an NLRP3-independent inflammasome response in BMDM and peritoneal macrophages

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

372 The observation that neither TcdB nor TcdA could activate Pyrin in hMDM was
373 unexpected. To test if this was specific to human cells, we next analyzed toxin-
374 dependent inflammasome responses in differentiated macrophages from bone
375 marrow (BMDM) as well as isolated peritoneal macrophages (PM) from either
376 WT or NLRP3 KO mice. We stimulated them with either TcdA or TcdB, using
377 Nigericin and poly(dA:dT) transfection as specificity controls for NLRP3 and
378 AIM2, respectively.
379
380 We determined that TcdA and TcdB triggered IL-1β secretion in BMDM from
381 both WT and NLRP3 KO cells, indicating that the response was NLRP3
382 independent (Fig. 3a). This was also true in peritoneal macrophages, which
383 showed little difference in toxin-mediated IL-1β secretion between WT and
384 NLRP3 KO cells (Fig. 3b). In contrast, the inflammasome response to Nigericin
385 was ablated entirely in the NLRP3 KO. At the same time, there was no
386 difference in IL-1β release in response to transfected dA:dT and no difference
387 in TNFa secretion in response to LPS. We also assessed IL-1β and caspase-1
388 cleavage in WT and NLRP3 KO BMDM following LPS priming and stimulation
389 with TcdB, Nigericin or dA:dT. Similarly, we found no differences between the
390 two genotypes when stimulated with TcdB or the specificity control dA:dT, while
391 Nigericin mediated IL-1β and caspase-1 cleavage were ablated in the NLRP3
392 KO (Fig. 3c).
393
394 **Figure 3. Pyrin is the responding sensor to TcdB in murine macrophages**
395

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

WT and NLRP3 deficient BMDM (A) or peritoneal macrophages (PM) (B) were primed with LPS (200 ng/ml, 3h), then activated with Nigericin and TcdB for 2h or dA:dT for 4h. (C) IL-1β and caspase-1 immunoblot of precipitated supernatant or cell lysate from WT and NLRP3 deficient BMDM treated as in (A). (D) IL-1β and caspase-1 immunoblot of precipitated supernatant or cell lysate from LPS primed WT BMDM either untreated or pretreated with CP-456,773 (2.5 μM, 30 mins), then stimulated as in (A). Mean and SEM of 3 independent experiments shown, immunoblots are representative of 3 independent experiments.

Given that we rely on CP-456,773 to determine the role of NLRP3 in response to TcdB in human primary macrophages, we also investigated whether, despite the lack of NLRP3 dependence, CP-456,773 could affect the response in BMDM. Therefore, WT BMDM were primed with LPS pre-incubated in the presence or absence of CP-456,773 and then incubated with TcdB, Nigericin and dA:dT. However, as with the NLRP3 KO cells, CP-456,773 did not affect TcdB- or dA:dT-mediated IL-1β or caspase-1 cleavage but completely inhibited Nigericin-mediated cleavage of both (Fig. 3d). Therefore, the TcdB-mediated inflammasome response in murine macrophages is both NLRP3-independent and insensitive to CP-456,773. Collectively these results show that the inflammasome response to both TcdA and TcdB in murine macrophages is independent of NLRP3 and likely depends on Pyrin.

Prolonged incubation with LPS or type I and II interferons increases Pyrin expression in hMDM

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

It was surprising that, in contrast to monocytes, neither TcdA nor TcdB triggered a Pyrin inflammasome response in hMDM. Thus, we next investigated whether inflammatory conditions increased the expression of inflammatory signaling molecules and thus potentially enable their activation. Pro-inflammatory signaling molecules activating either the NF-κB or IRF transcription factors have been demonstrated to increase Pyrin expression in PBMCs [24]. To determine whether either of these pathways could increase Pyrin expression in hMDM, we treated the cells with LPS, Pam3CS4K, TNFα, IFN-β and IFN-γ, as well as IL-4 and IL-10 for 5 or 18 hours and assessed Pyrin expression by immunoblot. Notably, only LPS increased Pyrin expression after 5h, while LPS, IFN-β and IFN-γ but not TNFα, IL-1β or Pam3CS4K increased expression after 18h (Fig. 4a). Thus, only stimuli that signal through IRF family transcription factors triggered an increase in Pyrin expression. To ensure that these molecules were functional in our system, we assessed the expression of IL-1β, a target of NF-kB, in response to LPS and Pam3CS4K. We determined that both were able to increase IL-1β expression (Fig. 4a), demonstrating that they were functional but unable to cause an increase in Pyrin expression.

Figure 4. LPS type I and type II interferon increase Pyrin expression and enable Pyrin activation in human macrophages

(A) Pyrin and IL-1β expression in hMDM treated with either IFN-γ (200 U/ml), LPS (10ng/ml), TNFα (50ng/ml), IL-10 (100ng/ml), IFN-β (5000 U/ml), IL-4 (1000U/ml) or Pam3Cys4K (20ng/ml) for either 5 or 18h. Representative of 3

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

446 independent experiments. (B) Pyrin (MEFV) or IL-1β transcript from hMDM
447 treated LPS (10ng/ml) or Pam3Cys4K (20ng/ml) for 12h. Mean and SEM the
448 fold change of three experimental replicates shown.
449
450 Previous studies had shown that the Pyrin promoter contains an IRSE element
451 that can be activated by both TRIF and IFN signaling [24], suggesting the
452 increase in Pyrin expression observed can be due to increased transcription.
453 We investigated this by stimulating the cells with either LPS or Pam3 for 12h,
454 then assessing mRNA copy number for Pyrin by qPCR, using IL-1β as a control.
455 We observed that LPS, but not Pam3CS4K, caused an increase in *MEFV*
456 transcript compared to the untreated cells (Fig. 4b). In contrast, both LPS and
457 Pam3CS4K increased IL-1β transcription, demonstrating that the increase in
458 *MEFV* transcript was specific to LPS (Fig. 4b). It is, therefore, likely that the
459 increase in Pyrin expression is driven by increased gene transcription.
460
461 **LPS and interferons prime activation of the Pyrin inflammasome in hMDM**
462
463 We tested whether increased Pyrin expression would be sufficient to enable
464 Pyrin inflammasome activation. Accordingly, we primed hMDM with LPS for
465 either 3 or 18 hours, pre-incubated them with DMSO, CP-456,773, VX-765 or
466 colchicine, and then treated them with TcdA or Nigericin. As we had observed
467 previously, TcdA did not trigger an inflammasome response after 3h of LPS
468 priming (Fig. 5a). In contrast, after 18 h, TcdA triggered robust secretion of IL-
469 1β that was sensitive to colchicine, and thus dependent on Pyrin (Fig. 5a). By
470 comparison, Nigericin mediated CP-456,773-sensitive IL-1β release after both

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

3 and 18 hours of LPS priming but was not affected by colchicine (Fig. 5a). Similarly, priming the cells for 18h, but not 3h, with IFN-β was sufficient to trigger Pyrin-dependent IL-18 secretion, demonstrating that it could also prime a Pyrin response (Fig. 5b).

Figure 5. Increased Pyrin expression is necessary and sufficient for Pyrin reactivation in human macrophages

(A) LPS primed (10ng/ml, 3 or 18h) hMDM were pre-incubated with compounds as noted previously, then stimulated with either TcdA (200 ng/ml) or Nigericin (8 μM) for 2.5h. The supernatant was harvested and assessed for IL-1β release. (B) IFN-β (5000U/ml, 3 or 18h) primed hMDM with either for 3h or 18h were pre-incubated with compounds as before then stimulated with TcdB (20 ng/ml) for 2.5h. The supernatant was harvested and assessed for IL-18 release. (C) hMDM primed with either LPS (10ng/ml) or Pam3Cys4K (20ng/ml) for 3h or 18h were treated as in (B). The supernatant was harvested and assessed for IL-1β release. (D) hMDM were transfected with siRNA targeting Pyrin mRNA (#59, #60) or the scrambled control and assessed for Pyrin expression by immunoblot or stimulated a previously following primed with LPS (10ng/ml) for 18h. (E) Expression of Pyrin in hMDM incubated with pepinhTRIF or the scrambled control peptide (25μg/ml, 5h), then treated with LPS (10ng/ml, 18h). Representative of 3 donors. IL-1β secretion from cell pretreated with pepinhTRIF or the scrambled control peptide (25μg/ml) in response to LPS priming (10ng/ml, 18h) followed by stimulation with either TcdA (200ng/ml) or nigericin (8μM) for 2.5h. (F) LPS primed (100ng/ml, 3h) differentiated Casapse-

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

4, NLRP3 double deficient BLaER1 cells reconstituted with either Pyrin or the vector control were incubated with Nigericin (8μM) TcdB (20ng/ml) or LFN-PrgI in the presence of protective antigen (PA) (25ng/ml) for 2.5h in the presence or absence of CP-456,773 (2.5μM, 15 min incubation). Supernatants were assessed for IL-1β. For (A)-(E) Mean and SEM of 3-4 experimental replicates shown. For (F) Mean and SD of three technical replicates shown, representative of 3 independent experiments.

We next sought to determine whether restoration of the Pyrin inflammasome response was specific to a stimulus that increased Pyrin expression. We thus compared the ability of TcdB to activate Pyrin in hMDM primed with Pam3CS4K to those primed with LPS. As done previously, hMDM were pre-incubated with different inhibitors to determine the responding inflammasome sensor. TcdB triggered CP-456,773-dependent inflammasome activation after three hours of priming with either LPS or Pam3CS4K (Fig. 5c). Comparative stimulation after 18 hours of priming led to a Pyrin-dependent response in the LPS primed cells, while the Pam3CS4K-primed cells failed to activate any inflammasome response (Fig. 5c), demonstrating that increased Pyrin expression is linked to Pyrin reactivation.

Increased Pyrin expression is necessary for the Pyrin response in hMDM

To determine whether the increase in Pyrin expression was a requirement for Pyrin activation, we transfected hMDM with two distinct siRNA against Pyrin or a scrambled control 24h before priming with LPS. Notably, both Pyrin targeting

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

siRNAs effectively prevented the LPS mediated increase in Pyrin expression. Still, they did not reduce it further than the untreated control, while the scrambled control had no effect (Fig. 5d). siRNA transfected hMDM were primed with LPS for 18h and then stimulated with TcdA or Nigericin. We observed that the two Pyrin siRNAs, but not the scrambled control, prevented TcdA-mediated IL-1β release. In contrast, neither of the Pyrin-targeting siRNAs, nor the control siRNA, affected Nigericin-mediated inflammasome activation (Fig. 5d), establishing decreasing Pyrin expression is sufficient to inhibit Pyrin activation specifically.

As Pyrin expression increased after treatement with LPS or interferons, but not Pam3Cys4K or TNFα, we hypothesised that the increase in LPS-dependent Pyrin expression likely required the TRIF signaling pathway. We tested our hypothesis by investigating whether blocking TLR4-mediated TRIF signaling prevents the LPS-dependent increase in Pyrin expression and restores the Pyrin inflammasome response. To this end, we pretreated hMDM with pepinhTRIF, a peptide that prevents the interaction of TRIF with its downstream interaction partners. We then incubated these cells with LPS for 18h before assessing Pyrin expression and activation. We found that treatment with pepinhTRIF, but not a control peptide, reduced LPS mediated Pyrin expression (Fig. 5e). We then stimulated the cells with either TcdA or Nigericin. We found that only the TcdA-mediated inflammasome response was inhibited by pretreatment with the pepinhTRIF, whereas the Nigericin-mediated IL-1β secretion was unaffected (Fig. 5e). These results demonstrate that the LPS-

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

stimulated increase in Pyrin expression is TRIF-mediated, and blocking this is sufficient to prevent Pyrin activation in these cells.

Given that increased Pyrin expression is necessary for Pyrin reactivation in hMDM, we next sought to determine whether Pyrin overexpression alone would enable a Pyrin inflammasome response. For this experiment we used the caspase-4, NLRP3 KO BLaER1 cells, which otherwise do not mount an inflammasome response to TcdB. We overexpressed Pyrin in the caspase-4, NLRP3 KO BLaER1 cells using reconstitution with a vector alone as a control. We then primed these cells with LPS and stimulated them with TcdB, Nigericin or needletox. Markedly, TcdB mediated an inflammasome response in the Pyrin-reconstituted cells, but not in cells transduced with the vector alone Notably, this was not inhibited by CP-456,773 (Fig. 5f). In contrast, Pyrin reconstitution had no effect on either NLRP3 or NLRC4 activation (Fig. 5f). This differed from our earlier observations where TcdB triggered an NLRP3 dependent inflammasome response in Pyrin KO BLaER1 cells overexpressing Pyrin. However, in this experiment we did not determine whether Pyrin overexpression in the Pyrin KO BLaER1 cells resulted in NLRP3 independent IL-1β secretion in response to TcdB. Furthermore, when NLRP3 is present, the NLRP3 inflammasome response to TcdB might predominate in BLaER1. These results demonstrate that an increasing Pyrin expression is sufficient to enable Pyrin inflammasome response, and regulation of Pyrin expression is the primary factor controlling Pyrin inflammasome activation in hMDM.

Dephosphorylation of Pyrin is insufficient for activation in hMDM

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

570

571 Pyrin does not form an inflammasome in response to TcdA or TcdB in hMDM
572 in the absence of a priming stimulus. This suggests that the toxin cannot
573 disengage one or both of the two known inhibitory mechanisms controlling
574 Pyrin: either dephosphorylation of the S204 and S242 residues, or the less well
575 characterized mechanism related to the B30.2 domain. To determine the
576 involvement of phosphorylation in regulating Pyrin activation in hMDM we
577 tested whether TcdB mediated dephosphorylated of Pyrin at serine 242 after
578 priming with LPS for 3h or 18h. Notably, we found that Pyrin was
579 dephosphorylated after treatment with TcdB regardless of the length of LPS
580 priming, demonstrating that this is unlikely to be the mechanism preventing
581 Pyrin activation (Fig. 6).

582

583 **Figure 6. The B30.2 domain regulates Pyrin activation in human**
584 **macrophages, and is disrupted by FMF mutations**

585

586 (A) LPS primed (10ng/ml, 3h or 18h) hMDM were treated with TcdB (5 ng/ml,
587 1h), then lysed and assessed for phosphorylation of Pyrin (S242), Pyrin or actin
588 by immunoblot. Representative of 3 independent experiments.

589

590 **Discussion**

591

592 Pyrin responds to virulence factors and endogenous molecules that inhibit the
593 RhoA signaling pathway. Gain of function mutants in the MEFV gene cause
594 multiple genetic autoinflammatory disorders [11]. Understanding Pyrin

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

595 regulation, particularly in the context of different human cell types, is therefore
596 critical to understand how auto-inflammation-associated mutations may disrupt
597 these mechanisms. In this study, we investigated the inflammasome response
598 to the disease-causing *C. difficile* toxins TcdA and TcdB in hMDM. We found
599 that under steady state conditions, and in spite of expressing similar levels of
600 Pyrin to monocytes, hMDM do not mount a Pyrin inflammasome response.
601 Notably, Pyrin activation could be re-established in these cells by further
602 increasing in Pyrin expression through prolonged priming with either LPS or
603 type I or II interferon. Surprisingly, in the absence of Pyrin activation, TcdB
604 instead activated NLRP3 in these cells, thus demonstrating a redundancy in the
605 inflammasome system to detect this toxin. TcdB similarly triggered NLRP3
606 activation in both human macrophage cell lines we tested, BLaER1 and THP1,
607 rather than Pyrin. Notably, unlike Pyrin activation, NLRP3 activation by TcdB
608 did not require the enzymatic activity of the toxin, demonstrating that it activates
609 Pyrin and NLRP3 through two distinct pathways. These findings are in contrast
610 to monocytes, which responded to both toxins in a Pyrin inflammasome
611 dependent manner, as has been shown previously [3]. This demonstrates a cell
612 type specific divergence in the inflammasome response to TcdB.
613
614 The results of our study demonstrate that regulatory mechanisms governing
615 Pyrin activation in hMDM prevent Pyrin activation unless the cell has been
616 exposed to a prior pro-inflammatory stimulus. Notably, in the case of Pyrin,
617 licensing required prolonged stimulation with LPS or interferons, but not other
618 inflammatory stimuli including TNFα. This contrasts with the data from a
619 previous study in PBMCs, where TNFα and Pam3CS4K stimulation also

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

620 increased Pyrin expression [24]. Increased expression of Pyrin was necessary
621 to prime a Pyrin dependent inflammasome response in hMDM, as preventing
622 increased Pyrin expression after these stimulation with LPS was sufficient to
623 inhibited Pyrin activation. Furthermore, overexpression of Pyrin in BLaER1 cells
624 was sufficient to enable a Pyrin response. These results suggest that, similar
625 to the NLRP3 and AIM2 inflammasomes, control of Pyrin expression through
626 inflammatory signals regulates Pyrin activation. Although the promoter region
627 of the MEFV gene contains elements that are recognized by either interferon
628 driven transcription factors or by NF-kB [24], increased Pyrin expression was
629 specific to activation of the TRIF/interferon pathway. This suggests that Pyrin is
630 specifically reactivated in hMDM by pathways associated with cell host defense
631 and gram-negative bacteria and may be important in the response to these
632 pathogens. An example of this is an increase in Pyrin expression seen in
633 response to *Francisella novidica* infection, where, unlike in mice, Pyrin is the
634 responding sensor to *F. novicida* in hMDM [25]. This also has implications for
635 Pyrin associated auto-inflammatory conditions, where, compared to NLRP3
636 driven auto-inflammatory conditions, IL-1β blocking therapy is only somewhat
637 successful in preventing disease [26]. Our data suggests that interferon
638 signaling could also play a role in triggering auto-inflammation in these patients,
639 as well as DAMPs released after tissue injury that stimulate TLR4. Further
640 research is needed to understand the contribution of these sensors to Pyrin
641 based auto-inflammatory conditions to determine if blocking these priming
642 signals helps to control these diseases.
643

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

644 Notably, we found Pyrin expression was comparible between hMDM and
645 monocytes under unstimulated conditions, indicating that the additional
646 regulatory steps in hMDM are post-translational. The most well characterized
647 of these is the phosphorylation of Pyrin. However, in spite of failing to trigger
648 formation of a Pyrin inflammasome, TcdB still caused dephosphorylation of the
649 S242 residue, demonstrating that this is not the mechanism preventing Pyrin
650 activation. An alternative mechanism controlling Pyrin activation could be the
651 control mechanism related to the colchicine sensitive requirement for Pyrin
652 activation, which requires the B30.2 domain [15]. Supporting this is the finding
653 that, unlike in human macrophages, TcdA and TcdB activate Pyrin in murine
654 macrophages, where a frame shift mutation has removed the B30.2 domain.
655 Another option would include the protein proline serine threonine phosphatase-
656 interacting protein 1 (PSTPIP1), which binds to Pyrin and facilitates its
657 oligomerisation and inflammasome formation [27]. However, further
658 investigations into how this control mechanism functions are required to gain
659 further information into whether this is indeed the control mechanism in
660 macrophages.
661
662 These findings demonstrate yet another point of divergence between human
663 monocytes and macrophages, as Pyrin is inactive in these cells. These
664 observations join a growing list of differences in inflammasome activation
665 between these cell types, which also differ in their responses to LPS, which
666 activates NLRP3 in monocytes without a need for a second stimulus, as well as
667 dsDNA, which triggers a STING- and NLRP3-dependent inflammasome
668 response in monocytes rather than activating AIM2 [23,28]. That Pyrin is active

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

669 in monocytes is unsurprising, as they are migratory cells that rely on actin
670 rearrangement to reach sites of infection where they contribute to the immune
671 response and clearing the pathogen. Therefore, inhibition of migration
672 represents a disruption of a basic function of these cells. This is consistent with
673 Pyrin activation in neutrophils, another migratory immune cell type. Conversely,
674 our results demonstrate that Pyrin is unable to respond in hMDM, even though
675 inactivation of Rho will also impact the immune response and viral clearance.
676 This suggests a requirement for more nuanced control of Pyrin activation in
677 hMDM, which may be a requirement to limit Pyrin driven auto-inflammation.
678 Limiting the Pyrin response to situations requiring pre-exposure to pro-
679 inflammatory molecules such as LPS or interferons may be necessary to
680 ensure that Pyrin is not aberrantly activated by endogenous molecules such as
681 bile acids [29], which may be present in tissues but not in the circulation [30].
682 How widespread this mechanism is will be elucidated by research focusing on
683 Pyrin regulation in other cell types. Notably, the regulation of Pyrin observed in
684 human macrophages was not evident in murine macrophages, which, similar
685 to human monocytes, responded to TcdA and TcdB in a Pyrin-dependent
686 manner. However, murine Pyrin lacks the B30.2 domain, and so may have lost
687 the regulatory mechanism preventing Pyrin activation in hMDM. This
688 demonstrates a further divergence in inflammasome responses between the
689 two species, in addition to NLRC4 activation and NLRP3 responses [31,32].
690
691 It was interesting to note that Pyrin activation was restricted in the hMDM in
692 spite of similar basal Pyrin expression to monocytes. The expression observed
693 was most likely due to M-CSF, which increases Pyrin expression in

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

694 monocytes/macrophages [33]. However, it is unclear why Pyrin is present but
695 stringently regulated. One possible explanation for this is that Pyrin has
696 additional roles in the cell aside from forming an inflammasome. It has been
697 suggested previously that Pyrin operates as a specialized adapter for
698 autophagic machinery [34]. This capacity it associates with autophagic
699 adapters ULK1 and Beclin1 to target NLRP3 and capase-1 for autophagic
700 degradation. Further investigation would be required to understand how M-CSF
701 driven Pyrin expression controls this phenomenon.
702
703 Our results have potential implications for FMF pathogenesis. Whilst defining
704 which mutations can cause FMF is still an area of ongoing research, the most
705 well characterized FMF-causing mutations occur in the B30.2 domain [35]. If
706 indeed the mechanism holding Pyrin inactive requires the less well
707 characterized mechanism involving the B30.2 domain, our data suggest that
708 this control mechanism will be disrupted by these mutations. Further research
709 investigating the effect of FMF mutations on regulation of Pyrin in macrophage
710 will elucidate whether these mutations indeed alter this regulation.
711
712 Our findings have implications for the pathogenesis of *C. difficile* infection. TcdB
713 triggered either a Pyrin- or NLRP3-dependent inflammasome response
714 depending on the cell type, while TcdA did not trigger an inflammasome
715 response in macrophages. This redundancy is quite intriguing, and suggests
716 that inflammasome-mediated detection of TcdB is important in the response to
717 the bacteria. Given that strains of *C.difficile* exist that express only one of TcdA
718 or TcdB, as well as both [36], it is also possible that disease severity alters

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

719 depending on whether the bacteria express TcdB, as early NLRP3 activation in
720 the macrophages prior to systemic penetration by the toxin may dictate the
721 speed of the immune response or conversely enhance tissue damage .
722 Furthermore, the detection mechanism for each sensor has different
723 requirements, as Pyrin detects the activity of the glycosyltransferase domain
724 through RhoA inactivation, whilst NLRP3 mediated TcdB detection is
725 independent of glycosyltransferase activity. Furthermore, our data demonstrate
726 that Pyrin activation is differentially regulated in human and mouse, and so the
727 inflammasome response to this infection may differ from what has been shown
728 in mouse models. Further studies in a model more closely related to humans,
729 such as pigs, is needed to determine the role of inflammasomes in *C.difficile*
730 infection, and determine whether the current inflammasome inhibitors being
731 developed represent new treatments to prevent *C. difficile*-associated
732 pathology or pose an increased risk of *C. difficile* infection.
733
734 **Materials and methods**
735
736 Ethics statement
737 Ethics for the use of human material was obtained according to protocols
738 accepted by the institutional review board at the University Clinic Bonn; local
739 ethics votes Lfd. Nr. 075/14). No consent was taken as all donors were
740 anonymous.
741
742 Reagents

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

743 LPS (Eb-ultrapure, 0111:B4), Pam3CS4K, pepinhTRIF and TAK-242 were
744 obtained from InvivoGen, Nigericin was obtained from Invitrogen and *Bacillus*
745 *anthracis* protective antigen was obtained from List Biological Laboratories.
746 Colchicine was obtained from Sigma, and VX-765 was obtained from
747 Sellekchem. DRAQ5 was purchased from eBioscience. TNFα, IFN-β, IFN-γ, M-
748 CSF, IL-3, IL-4 and IL-10 were purchased from Immunotools. TcdA and TcdB
749 from C. difficile strain VPI10463 were recombinantly produced and supplied by
750 Prof. Ralf Gerhard [37]. Both toxins are identical to TcdA and TcdB from strain
751 cdi630, which was used for infection assay. The HTRF kits for human IL-1β and
752 TNFa were obtained from Cisbio, the ELISA kit for human and mouse IL-1β
753 was obtained from R&D Systems. Both were used according to the
754 manufacturer's instructions. For the *Clostridioides difficile* supernatant transfer
755 and co-culture, the following reagents were used: Butyric acid (Sigma-Aldrich:
756 W222119-1KG-K), Various amino acids (Roth or Sigma), Iron sulfate
757 heptahydrate (Sigma-Aldrich: 215422-250G), Triton-X 100 (Roth: 3051.2), M-
758 Per (Sigma-Aldrich: 78501), Protease inhibitor (Sigma-Aldrich: 11836170001).
759
760 Cells lines and tissue culture
761 The BLaER1 cells and THP-1 cells were maintained in complete RPMI (RPMI
762 containing 10% heat-inactivated fetal calf serum, 1% Pen/Strep and 1%
763 GlutaMAX, all obtained from Thermo Fisher). To trans-differentiate the BLaER1
764 cells to a macrophage-like cells they were resuspended at $1x10^6$ cells/ml in
765 complete RPMI with 10 ng/ml IL-3, 25 U/ml M-CSF and 100 nM β-Estradiol,
766 and then 100ul was plated in poly-L-lysine (Sigma, P8920) coated 96 well
767 plates and incubated for 6 days to differentiate the cells. On day 6 the cells

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

were used for experiments. The BLaER1 CRISPR KO cell lines were obtained from the laboratory of Prof. Veit Hornung and were generated as described in [23]. THP-1 cells were differentiated in full medium containing 50 ng/mL PMA. The THP-1 CRISPR KO cells were generated using transduction with lentiviruses based on pLenti CRISPR v2 [38] using sgRNAs: Caspase- 1- TACCATGAGACATGAACACC, ASC – GCTGGATGCTCTGTACGGGA, NLRP3 - CAATCTGAAGAAGCTCTGGT and Pyrin – TCTGCTGGTCACCTACTATG, followed by selection in 0.75 µg/mL puromycin. Monoclonal cell lines were generated by limited dilution and verified by Sanger sequencing and immunblot.

Primary cell isolation and differentiation

Monocytes were purified from buffy coat preparations from healthy donors (obtained according to protocols accepted by the institutional review board at the University Clinic Bonn; local ethics votes Lfd. Nr. 075/14). All donors were anonymous. The buffy coat was mixed in a 2:3 ratio with PBS, layered onto a ficoll gradient, and centrifuged at 700g for 20 min without brake. The PBMC layer was extracted from the interface. After washing, it was incubated with CD14 conjugated magnetic beads (Milltenyi Bioscience) and purified using MACS columns (Milltenyi Bioscience) as per the manufacture's protocol. The cells were then counted and resuspended for direct use or cultured for 3 days in RPMI containing 50 U/ml M-CSF at $2x10^6$ cells/ml to generated hMDM. After the three-day differentiation, hMDM were harvested and plated for experiments, then left to adhere overnight in RPMI containing 25 U/ml M-CSF.

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

793 Inflammasome stimulation assays

794 Primary monocytes/hMDM: Cells were harvested and seeded the day before
795 the assay. Before the experiment, the media was removed, and fresh media
796 with or without a TLR stimulus was added (LPS 10ng/ml, Pam3CS4K 25 ng/ml)
797 and incubated for 3h. Next, compounds were added and incubated for 15 min
798 (CP-456,773 2.5 uM, VX-765 40 uM, Colchicine 2.5 uM). Inflammasome
799 activators were subsequently added and incubated for 2.5h (TcdB 20 ng/ml,
800 Nigericin 8 μM, PrgI/PA 25 ng/ml, respectively). Plates were centrifuged at 450g
801 for 5 min, then the supernatant harvested for a cytokine or immunoblot analysis,
802 and the cells lysed in RIPA buffer were applicable.
803
804 BLaER1 cells: Differentiated BLaER1 cells were seeded in 96 well plates pre-
805 coated in the poly-L-lysine. Before the experiment, the media was removed and
806 fresh media with or without LPS (100 ng/ml) and incubated for 3h. Next,
807 compounds were added and incubated for 15 min (CP-456,773 2.5 μM, VX-765
808 40 μM, Colchicine 2.5 μM). Inflammasome activators were subsequently added
809 and incubated for 2.5h (TcdB 20 ng/ml, Nigericin 8 uM, PrgI/PA 25 ng/ml,
810 respectively). Plates were centrifuged at 450g for 5mins, and then the
811 supernatant was harvested for cytokine analysis.
812
813 THP1s were seeded in 24 well plates in the presence of 50 ng/mL PMA ($2 \cdot 10^5$
814 per well). Medium was replaced after 16 h; 24 h after this, cells were primed
815 with 100 ng/mL LPS for 3h, followed by treatment with 8 uM Nigericin (NLRP3)
816 or 100 ng/mL PA + 200 ng/mL LFn-PrgI (needletox, NLRC4) for 1.5 h, or 2
817 μg/mL TcdB for 8h. Supernatants were cleared by centrifugation at 4° C, 1000g

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

for 10 min and IL-1β and TNFα levels were quantified by ELISA. Where indicated, cells were treated with 2.5 μM CP-456,773 (CRID3, MCC950), or 40 μM Vx-765 30 min before and during stimulation.

Sample preparation and immunoblotting

The supernatant from primary human monocytes, hMDM, or BMDMs ($2x10^6$ cells/well in a 6 well plate) was harvested following inflammasome stimulation and the cells lysed in RIPA buffer (20 mM Tris-HCl, pH 7.4, 150 mM NaCl, 1 mM EDTA, 1% Triton X-100, 0.1% SDS, 0.5% deoxycholate, cOmplete protease and PhosSTOP (Roche) inhibitor). First, DNA was disrupted by sonication, then the lysate equivalent to $2x10^5$ cells was mixed at a 1:4 ratio with 4x LDS buffer containing 10% sample reducing agent (Invitrogen). Samples were heated at 95°C for 5 minutes and collected by centrifugation before loading.

Protein from supernatants was precipitated by adding an equal volume of methanol and 0.25 volumes of chloroform, centrifuged for 3 min at 13,000g. Next, the upper phase was discarded, the same volume of methanol of the previous step was added, and the sample was centrifuged for 3 min at 13,000g. The pellet was then dried and resuspended in 1x LDS-sample buffer containing a 10% sample reducing agent (Invitrogen). Samples were heated at 95°C for 5 minutes and collected by centrifugation before loading.

Proteins were separated by 4–12% SDS-PAGE in precast gels (Novex; Invitrogen) with MOPS buffer for proteins above 50kDa or MES buffer for

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

proteins below 50kDa (Novex; Invitrogen). Proteins were transferred onto Immobilon-FL PVDF membranes (Millipore), and nonspecific binding was blocked with 3% BSA in Tris-buffered saline for 1 h, followed by overnight incubation with specific primary antibodies in 3% BSA in Tris-buffered saline with 0.1% Tween-20. For the phosphor-Pyrin immunoblots the transferred membranes were instead blocked in Tris-buffered saline containing 1% milk powder.

Primary antibodies were used as follows: NLRP3 (1:5,000; Cryo-2), human caspase-1 for lysate analysis (1:1,000; Bally-1), murine caspase-1 (1:1000; casper-1) from Adipogen, Pyrin (1:1000, MEFV polyclonal 24280-1-AP) from Proteintech, phospho-Pyrin S241 (1:500; ab200420) from Abcam, human IL-1β (1:1000, AF-201-NA) and murine IL-1β (1:1000, AF-401-NA) from R&D bioscience, Rac (1:1000, clone 102) from BD transduction laboratories, human caspase-1 for supernatant analysis (1:1000, D57A2), Rac1/2/3 (1:1000, rabbit polyclonal #2465) from CST, actin (mouse or rabbit, both 1:1,000 dilution) from LI-COR Biosciences. Membranes were then washed and incubated with the appropriate secondary antibodies (IRDye 800CW, IRDye 680RD or HRP; 1:25,000 dilution; LI-COR Biosciences). In the case of caspase-1 detection or Pyrin detection, the membranes were incubated with washed and analyzed with an Odyssey CLx imaging system (LI-COR Biosciences) and ImageStudio 3 Software (LI-COR Biosciences). For the phosphor-Pyrin immunoblots the membrane was developed using western lighting plus-ECL and analysed on a VersaDoc (Biorad).

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

868 Cytokine measurements

869 Cytokines were measured either by ELISA or by HTRF as per the
870 manufactures' instructions. The kits used for ELISA were the human IL-1β
871 (DY201), human TNFα (DY210), mouse IL-1β (DY401) or mouse TNFa
872 (DY410), all from RnD biosystems. HTRF kits used were human IL-1β
873 (62IL1PEC) or human TNFα (62TNFPEB). All assays were read using a
874 SpectraMAX i3 (molecular devices) using the additional HTRF cartridge.
875 Human IL-18 measurements were performed using a cytokine bead array that
876 was generated in our laboratory. The xMAP antibody coupling kit from Luminex
877 (40-50016) was used to conjugate the capture IL-18 antibody (D0044-3, MBL)
878 to the beads. IL-18 was then measured following the standard Luminex protocol
879 and measured on a Magpix multiplexing unit (Luminex).

880

881 siRNA transfection

882 hMDM were harvested by centrifugation (350g, 5 min) and washed twice in
883 PBS. The cells were then aliquoted to have 1.2 x 10^6 cells per reaction and
884 centrifuged again for 2 min at 3000 rpm. The supernatant was discarded, and
885 the cell pellet resuspended in 10.5 μL Buffer R with siRNA at 10nM. 10 μL of
886 reaction mix was loaded into the neon electroporation, and the pipette plugged
887 into place within the electroporation tube containing 3ml Buffer E. The
888 electroporation settings were: 1400 V, 20 ms and 2 pulses. Subsequently, the
889 cells were transferred into 2 mL pre-warmed antibiotic-free RPMI. After
890 counting, the appropriate number of cells was seeded in 12-well or 96-well
891 tissue culture plates and incubated for 24 h before experiments.

892

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

893 Immunofluorescence and microscopy

894 Following treatment were washed once in PBS, then fixed in 2% PFA at 4C
895 overnight (for BLaER1 ASC speck analysis) or 4% PFA on ice for 30 min. The
896 cells were then washed twice in PBS containing 20 mM Glycine, then twice in
897 PBS. To stain for intracellular targets, the cells were then permeabilized in 0.1%
898 Triton X-100 for 5 min, then blocked in intracellular staining solution (PBS +10%
899 goat serum, 1% HI-FBS and 0.1% Triton X-100) for 30 mins RT. Next, we used
900 Alexa-647 conjugated Phalloidin (Invitrogen, A22287) for 30 min RT in an
901 intracellular staining solution to stain actin. The cells were then washed (3x
902 5min) and incubated with DAPI (1μg/ml, 10min) before being washed and
903 imaged. For ASC speck detection, the fixed cells were incubated with DRAQ5
904 (eBioscience, 65-0880-96) for 5 min (1:2000 dilution), then imaged directly. All
905 imaging was performed with an Observer.Z1 epifluorescence microscope, 20×
906 objective (dry, PlanApochromat, NA 0.8; ZEISS), Axiocam 506 mono, and ZEN
907 Blue software (ZEISS). Image analysis of all ASC speck experiments was done
908 using a cell profiler pipeline optimized to detect either ASC specks or nuclei. A
909 minimum of 6 images was analyzed for each condition in each experiment.
910
911 Retroviral transduction and fluorescent activated cell sorting
912 To produce the virus-containing supernatant $0.4x10^6$ HEK293T cells were
913 plated in 2 mL complete DMEM in one well of a 6 well dish. After 16 - 24 h,
914 HEK293T cells were transfected with retroviral constructs encoding the gene of
915 interest (2 µg per well), the retroviral packaging plasmids gag-pol (1 µg well),
916 and VSV-G (100 ng/well) using GeneJuice transfection reagent (Novagen,
917 70967). Cells were incubated at 37C, 5% CO2 for approximately 12 h, and then

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

the media was exchanged with RPMI containing 30% HI-FBS, and cells were incubated for another 34 h. After 36 h, the viral supernatant was collected using a 10 mL Luer-lock syringe attached to a blunt 18G needle and then filtered using a 0.45 mm filter unit into a 50 mL falcon. The medium on target cells was removed, and viral supernatant was added to the cells at a 2:1 ratio with complete RPMI. 8 µg/ml polybrene was then added to the diluted virus-containing supernatant. The cells were then centrifuged at 800g for 45 min at 37°C, then harvested and plated in 24 well plates before being incubated for approximately 24 hr at 37°C, 5% CO2. Following incubation, the cells were collected by centrifugation, and the virus-containing medium was removed and replaced by complete RPMI. Transduced cells were passaged three times before frozen stocks were prepared. Cells were sorted for equal expression of Pyrin, and NLRP3 variants using fluorescence assisted cell sorting on a FACS Aria cell sorter for equivalent expression of the fluorescent protein used as a marker of transduction.

C.difficile co-culture and supernatant generation

Experiments were performed with *C. difficile* DSM 28645 and DSM 29688 obtained from the German Collection of Microorganisms and Cell Cultures (DSMZ, Braunschweig, Germany). Main cultures were cultivated in RPMI 1640, supplemented 10% FBS, 0.014 mM iron sulfate, 4.16 mM cysteine, 4.33 mM proline, 1.11 mM valine, 1.12 mM leucine, 0.72 mM isoleucine, 0.22 mM tryptophane, 0.57 mM methionine and 0.22 mM histidine at 2% O_2, 5% CO_2, 37°C, 40-50% humidity using O2 Control InVitro Glove Box (Coy Labs, USA).

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

PBMCs were isolated from three different donors and differentiated into hMDM as described above. Two days before the experiment, cells were seeded in RPMI medium supplemented with 10% HI-FBS at $3.3x10^5$ cells per well in 24 well plates and incubated at normoxic conditions at 37°C for 24 h. On the following day, the cells were placed into a hypoxia chamber (2% O_2, 5% CO_2, 37°C, 40-50% humidity) for another 24 h. On the same day, *C. difficile* main cultures of DSM 28645 (toxin-producing) and DSM 29688 (non-toxigenic) were inoculated at an OD_{600nm} of ~0.01 and incubated for 24 h. On the day of the experiment the medium was removed, and the cells were washed with 500 µl PBS. Following, the cells were treated with 375 µl RPMI or 375 µl RPMI containing 10 ng/ml LPS for 2 h. The OD_{600nm} of both *C. difficile* cultures was determined, and the number of bacterial density was determined using the following formulation:

C. difficile per ml main culture = 26,445,593 x OD_{600nm}

The cells were centrifuged at 2500g for 10 min. After centrifugation, the supernatant was passed through a 0.2 µM sterile filter. The pellet was resuspended in RPMI supplemented with 5 mM butyrate and with lower concentrations of glycine (reduced to 0.033 mM), cysteine (0 mM), proline (0 mM), isoleucine (0.095 mM), leucine (0.095 mM), methionine (0.026 mM), serine (0 mM), threonine (0.042 mM) and valine (0.042 mM) and the bacterial density was adjusted to a multiple of infection (MOI) of 300. The cells were treated with 375 µl sterile-filtered *C. difficile* supernatant, living *C. difficile* (300 MOI) or RPMI. The cells were additionally treated with 10 ng/ml LPS, 2 µM CP-

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

456,773 or a combination of both. A lysis control was included by the addition of 0.5% triton-X100 in RPMI. After 3 or 6 hours, the cell supernatant was collected, centrifuged for 10 min at 2500g and frozen at -80°C. The cells were washed two times with 750 µl PBS and lysed by the addition of 80 µl M-PER with cOmplete Mini Protease Inhibitor for 5 minutes. The lysed cell suspensions were collected and stored at -80°C.

Caspase-1 activity assay

The caspase-1 activity assay was performed using the Caspase-Glo 1 inflammasome assay from Promega (G9951) as per the manufactures instructions. Briefly, cell-free supernatant from inflammasome stimulated cells was mixed with equal amounts for reconstituted caspase-1 reagent and the Luminescence signal read on a SpectraMAX i3 (molecular devices) at 30, 60 and 90 min post mixing.

qPCR

qPCR quantifications were performed essentially as previously described [39] with the following changes: 500 ng of RNA was used for the RT-PCR and the qPCR was performed using QuantStudio 6 PCR System (Thermo Fisher Scientific). The primer sequences were as follows: Hprt, forward 5′-TCAGGCAGTATAATCCAAAGATGGT-3′ and reverse 5′-AGTCTGGCTTATATCCAACACTTCG-3′; MEFV, forward 5′-GGAAGGCCACCAGACACGG-3′ and reverse 5′-GTGCCCAGAAACTGCCTCGG-3′.

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

Acknowledgements

We thank C.C. Kolbe for setting up and optimising the IL-18 Cytokine Bead Array assay, H. Tatge for purification of the different *C.difficile* toxins as well as both the Microscopy Core Facility and Flow Cytometry Core Facility at the University of Bonn for providing help, service and devices funded by the Deutsche Forschungsgemeinschaft (DFG – German Research Foundation) – Projektnummer 13123509. This work was funded by the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation) – Project-ID 369799452 (TRR237) (E.L.), Project-ID 414786233 (SFB1403) (E.L.), Project-ID 432325352 (SFB1454) (E.L.), DFG (GE1017/5-1) (R.G) and Germany's Excellence Strategy – ImmunoSensation2 – Project-ID 390873048 (EXC2151) (E.L.), The work was also supported by the Helmholtz-Gemeinschaft, Zukunftsthema 'Immunology and Inflammation' (ZT-0027) (E.L.) and the German-Israeli Foundation for Scientific Research and Development, Grant No: 1085 (V.H.) This project was partially funded by the Niedersächsiches Ministerium für Wissenschaft und Kultur (MWK), Niedersächsisches Vorab [grant number ZN3380] (K.H.)

References

1. Zheng D, Liwinski T, Elinav E. Inflammasome activation and regulation: toward a better understanding of complex mechanisms. Cell Discov. 2020;6: 36. doi:10.1038/s41421-020-0167-x

2. Liston A, Masters SL. Homeostasis-altering molecular processes as mechanisms of inflammasome activation. Nat Rev Immunol. 2017;17: 208–214. doi:10.1038/nri.2016.151

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

1019 3. Xu H, Yang J, Gao W, Li L, Li P, Zhang L, et al. Innate immune sensing of
1020 bacterial modifications of Rho GTPases by the Pyrin inflammasome. Nature.
1021 2014;513: 237–241. doi:10.1038/nature13449

1022 4. Hodge RG, Ridley AJ. Regulating Rho GTPases and their regulators. Nat
1023 Rev Mol Cell Bio. 2016;17: 496–510. doi:10.1038/nrm.2016.67

1024 5. Popoff MR. Bacterial factors exploit eukaryotic Rho GTPase signaling
1025 cascades to promote invasion and proliferation within their host. Small
1026 Gtpases. 2014;5: e983863. doi:10.4161/sgtp.28209

1027 6. Mangan MSJ, Olhava EJ, Roush WR, Seidel HM, Glick GD, Latz E.
1028 Targeting the NLRP3 inflammasome in inflammatory diseases. Nat Rev Drug
1029 Discov. 2018;17: 588–606. doi:10.1038/nrd.2018.97

1030 7. Akula MK, Shi M, Jiang Z, Foster CE, Miao D, Li AS, et al. Control of the
1031 innate immune response by the mevalonate pathway. Nat Immunol. 2016;17:
1032 922–929. doi:10.1038/ni.3487

1033 8. Masters SL, Lagou V, Jéru I, Baker PJ, Eyck LV, Parry DA, et al. Familial
1034 autoinflammation with neutrophilic dermatosis reveals a regulatory
1035 mechanism of pyrin activation. Sci Transl Med. 2016;8: 332ra45-332ra45.
1036 doi:10.1126/scitranslmed.aaf1471

1037 9. Gao W, Yang J, Liu W, Wang Y, Shao F. Site-specific phosphorylation and
1038 microtubule dynamics control Pyrin inflammasome activation. Proc National
1039 Acad Sci. 2016;113: E4857–E4866. doi:10.1073/pnas.1601700113

1040 10. Park YH, Wood G, Kastner DL, Chae JJ. Pyrin inflammasome activation
1041 and RhoA signaling in the autoinflammatory diseases FMF and HIDS. Nat
1042 Immunol. 2016;17: 914–921. doi:10.1038/ni.3457

1043 11. Heilig R, Broz P. Function and mechanism of the pyrin inflammasome. Eur
1044 J Immunol. 2018;48: 230–238. doi:10.1002/eji.201746947

1045 12. Rigante D, Manna R. FAMILIAL MEDITERRANEAN FEVER:
1046 ASSESSING THE OVERALL CLINICAL IMPACT AND FORMULATING
1047 TREATMENT PLANS. Mediterr J Hematology Infect Dis. 2019;11: e2019027–
1048 e2019027. doi:10.4084/mjhid.2019.027

1049 13. Torre-Minguela C de, Castillo PM del, Pelegrín P. The NLRP3 and Pyrin
1050 Inflammasomes: Implications in the Pathophysiology of Autoinflammatory
1051 Diseases. Front Immunol. 2017;8: 43. doi:10.3389/fimmu.2017.00043

1052 14. Magnotti F, Lefeuvre L, Benezech S, Malsot T, Waeckel L, Martin A, et al.
1053 Pyrin dephosphorylation is sufficient to trigger inflammasome activation in
1054 familial Mediterranean fever patients. Embo Mol Med. 2019;11: e10547.
1055 doi:10.15252/emmm.201910547

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

15. Gorp HV, Saavedra PHV, Vasconcelos NM de, Opdenbosch NV, Walle LV, Matusiak M, et al. Familial Mediterranean fever mutations lift the obligatory requirement for microtubules in Pyrin inflammasome activation. Proc National Acad Sci. 2016;113: 14384–14389. doi:10.1073/pnas.1613156113

16. Lyras D, O'Connor JR, Howarth PM, Sambol SP, Carter GP, Phumoonna T, et al. Toxin B is essential for virulence of Clostridium difficile. Nature. 2009;458: 1176–1179. doi:10.1038/nature07822

17. Ng J, Hirota SA, Gross O, Li Y, Ulke–Lemee A, Potentier MS, et al. Clostridium difficile Toxin–Induced Inflammation and Intestinal Injury Are Mediated by the Inflammasome. Gastroenterology. 2010;139: 542-552.e3. doi:10.1053/j.gastro.2010.04.005

18. Liu Y-H, Chang Y-C, Chen L-K, Su P-A, Ko W-C, Tsai Y-S, et al. The ATP-P2X7 Signaling Axis Is an Essential Sentinel for Intracellular Clostridium difficile Pathogen-Induced Inflammasome Activation. Front Cell Infect Mi. 2018;8: 84. doi:10.3389/fcimb.2018.00084

19. Saavedra PHV, Huang L, Ghazavi F, Kourula S, Berghe TV, Takahashi N, et al. Apoptosis of intestinal epithelial cells restricts Clostridium difficile infection in a model of pseudomembranous colitis. Nat Commun. 2018;9: 4846. doi:10.1038/s41467-018-07386-5

20. Coll RC, Robertson AAB, Chae JJ, Higgins SC, Muñoz-Planillo R, Inserra MC, et al. A small-molecule inhibitor of the NLRP3 inflammasome for the treatment of inflammatory diseases. Nat Med. 2015;21: 248–255. doi:10.1038/nm.3806

21. Dinarello CA, Novick D, Kim S, Kaplanski G. Interleukin-18 and IL-18 Binding Protein. Front Immunol. 2013;4: 289. doi:10.3389/fimmu.2013.00289

22. Genth H, Huelsenbeck J, Hartmann B, Hofmann F, Just I, Gerhard R. Cellular stability of Rho-GTPases glucosylated by Clostridium difficile toxin B. Febs Lett. 2006;580: 3565–3569. doi:10.1016/j.febslet.2006.04.100

23. Gaidt MM, Ebert TS, Chauhan D, Schmidt T, Schmid-Burgk JL, Rapino F, et al. Human Monocytes Engage an Alternative Inflammasome Pathway. Immunity. 2016;44: 833–846. doi:10.1016/j.immuni.2016.01.012

24. Centola M, Wood G, Frucht DM, Galon J, Aringer M, Farrell C, et al. The gene for familial Mediterranean fever, MEFV, is expressed in early leukocyte development and is regulated in response to inflammatory mediators. Blood. 2000;95: 3223–3231. doi:10.1182/blood.v95.10.3223

25. Gavrilin MA, Mitra S, Seshadri S, Nateri J, Berhe F, Hall MW, et al. Pyrin Critical to Macrophage IL-1β Response to Francisella Challenge. J Immunol. 2009;182: 7982–7989. doi:10.4049/jimmunol.0803073

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

1095 26. Hentgen V, Vinit C, Fayand A, Georgin-Lavialle S. The Use of
1096 Interleukine-1 Inhibitors in Familial Mediterranean Fever Patients: A Narrative
1097 Review. Front Immunol. 2020;11: 971. doi:10.3389/fimmu.2020.00971

1098 27. Yu J-W, Fernandes-Alnemri T, Datta P, Wu J, Juliana C, Solorzano L, et
1099 al. Pyrin Activates the ASC Pyroptosome in Response to Engagement by
1100 Autoinflammatory PSTPIP1 Mutants. Mol Cell. 2007;28: 214–227.
1101 doi:10.1016/j.molcel.2007.08.029

1102 28. Gaidt MM, Ebert TS, Chauhan D, Ramshorn K, Pinci F, Zuber S, et al.
1103 The DNA Inflammasome in Human Myeloid Cells Is Initiated by a STING-Cell
1104 Death Program Upstream of NLRP3. Cell. 2017;171: 1110-1124.e18.
1105 doi:10.1016/j.cell.2017.09.039

1106 29. Alimov I, Menon S, Cochran N, Maher R, Wang Q, Alford J, et al. Bile acid
1107 analogues are activators of pyrin inflammasome. J Biol Chem. 2019;294:
1108 3359–3366. doi:10.1074/jbc.ra118.005103

1109 30. Chiang JYL. Comprehensive Physiology. Compr Physiol. 2013;3: 1191–
1110 1212. doi:10.1002/cphy.c120023

1111 31. Swanson KV, Deng M, Ting JP-Y. The NLRP3 inflammasome: molecular
1112 activation and regulation to therapeutics. Nat Rev Immunol. 2019;19: 477–
1113 489. doi:10.1038/s41577-019-0165-0

1114 32. Yang J, Zhao Y, Shi J, Shao F. Human NAIP and mouse NAIP1 recognize
1115 bacterial type III secretion needle protein for inflammasome activation. Proc
1116 National Acad Sci. 2013;110: 14408–14413. doi:10.1073/pnas.1306376110

1117 33. Seshadri S, Duncan MD, Hart JM, Gavrilin MA, Wewers MD. Pyrin Levels
1118 in Human Monocytes and Monocyte-Derived Macrophages Regulate IL-1β
1119 Processing and Release. J Immunol. 2007;179: 1274–1281.
1120 doi:10.4049/jimmunol.179.2.1274

1121 34. Kimura T, Jain A, Choi SW, Mandell MA, Schroder K, Johansen T, et al.
1122 TRIM-mediated precision autophagy targets cytoplasmic regulators of innate
1123 immunity. J Cell Biol. 2015;210: 973–989. doi:10.1083/jcb.201503023

1124 35. Touitou I. The spectrum of Familial Mediterranean Fever (FMF) mutations.
1125 Eur J Hum Genet. 2001;9: 473–483. doi:10.1038/sj.ejhg.5200658

1126 36. Aktories K, Schwan C, Jank T. Clostridium difficile Toxin Biology. Annu
1127 Rev Microbiol. 2016;71: 1–27. doi:10.1146/annurev-micro-090816-093458

1128 37. Burger S, Tatge H, Hofmann F, Genth H, Just I, Gerhard R. Expression of
1129 recombinant Clostridium difficile toxin A using the Bacillus megaterium
1130 system. Biochem Bioph Res Co. 2003;307: 584–588. doi:10.1016/s0006-
1131 291x(03)01234-8

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

38. Sanjana NE, Shalem O, Zhang F. Improved vectors and genome-wide libraries for CRISPR screening. Nat Methods. 2014;11: 783–784. doi:10.1038/nmeth.3047

39. Nardo DD, Labzin LI, Kono H, Seki R, Schmidt SV, Beyer M, et al. High-density lipoprotein mediates anti-inflammatory reprogramming of macrophages via the transcriptional regulator ATF3. Nat Immunol. 2014;15: 152–160. doi:10.1038/ni.2784

Supp figure 1.

(A) hMDM were incubated with either toxin proficient (TcdA+,TcdB+) or toxin deficient (TcdA-,TcdB-) *C. difficile* (MOI 100:1) for 6h, then the supernatant assessed for IL-1b and IL-18. Mean and SEM of three experimental replicates shown. (B) Rac glucosylation status in either monocytes or hMDM following treatment with the listed toxins (NXN variants - lack glucosyltransferase activity). Representative of 3 experiments. (C) Actin staining following incubation of monocytes or macrophages with or without LPS and TcdB. Treated cells were fixed and stained with Phalloidin 647 to detect actin (red) or with DAPI to detect nuclei (blue). Images are representative from 3 separate donors.

Supp figure 2.

A) Differentiated Casapse-4 deficient BLaER1 cells were stimulated as in Fig 2A. IL-1β was assessed from the harvested supernatants. (B) LPS-primed differentiated WT BLaER1 cells were pre-incubated with TAK242 (2uM, 30 min) then activated with TcdB (20ng/ml) or Nigericin (8uM) for 2h. Harvested supernatant was assessed for IL-1β. (C) Differentiated WT BLaER1 cells were

bioRxiv preprint doi: https://doi.org/10.1101/2021.07.07.451422; this version posted July 7, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

pre-incubated with TAK242 then stimulated with LPS for 4h. TNFa was assessed from the supernatant. (D) TNFa was measured for THP-1 cells from Fig. 2G. Mean and SEM shown for three independent experiments. (E) LPS primed (10ng/ml, 3h) human macrophages were treated either TcdB or the TcdB NXN mutant lacking glucosyltrasferase activity (20ng/ml, 2.5h). Supernatant was harvested and assessed for IL-1β. For A-C the mean and SD of three technical replicates shown, representative of 3 independent experiments. For D and E. the mean and SEM shown for three independent experiments.

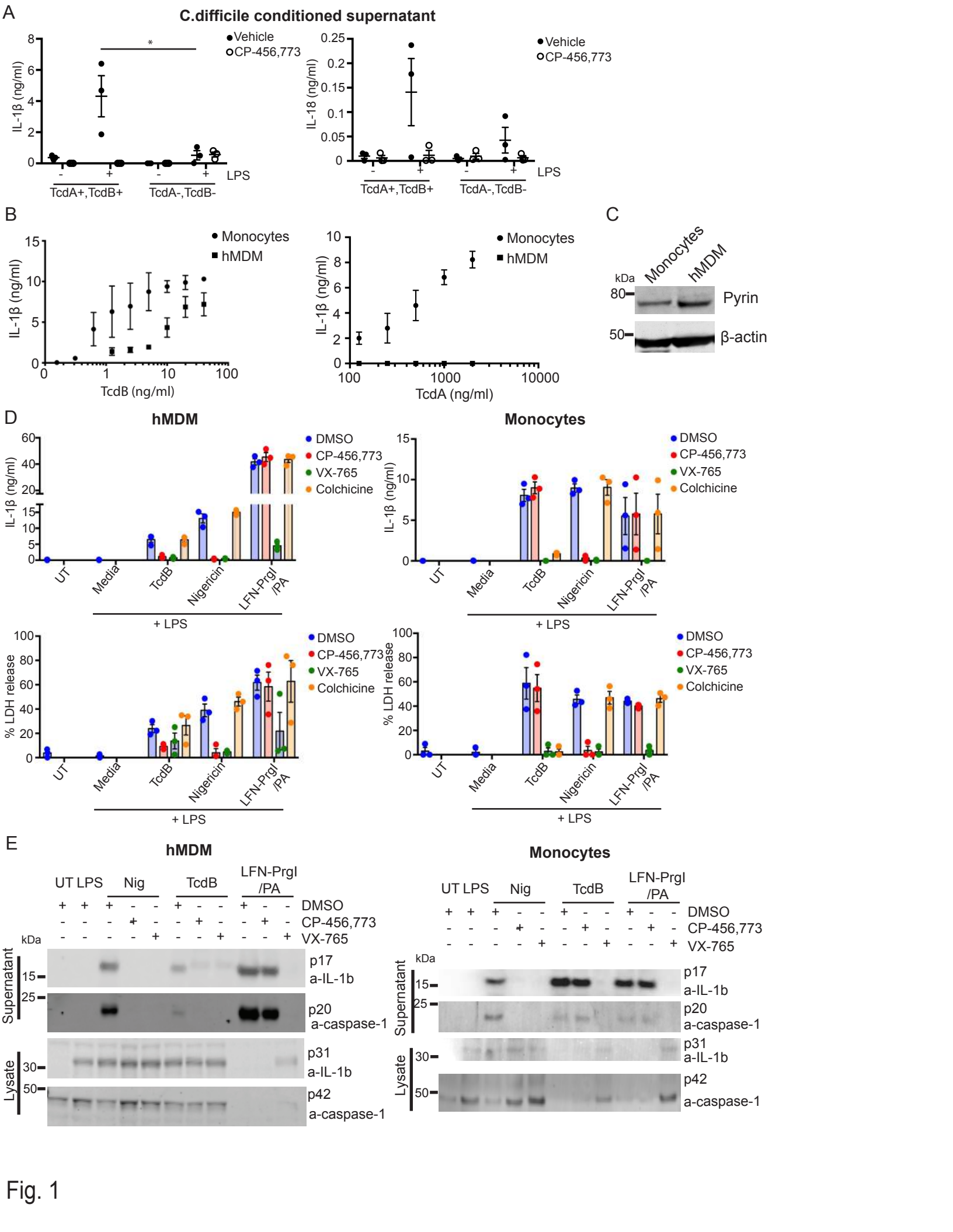
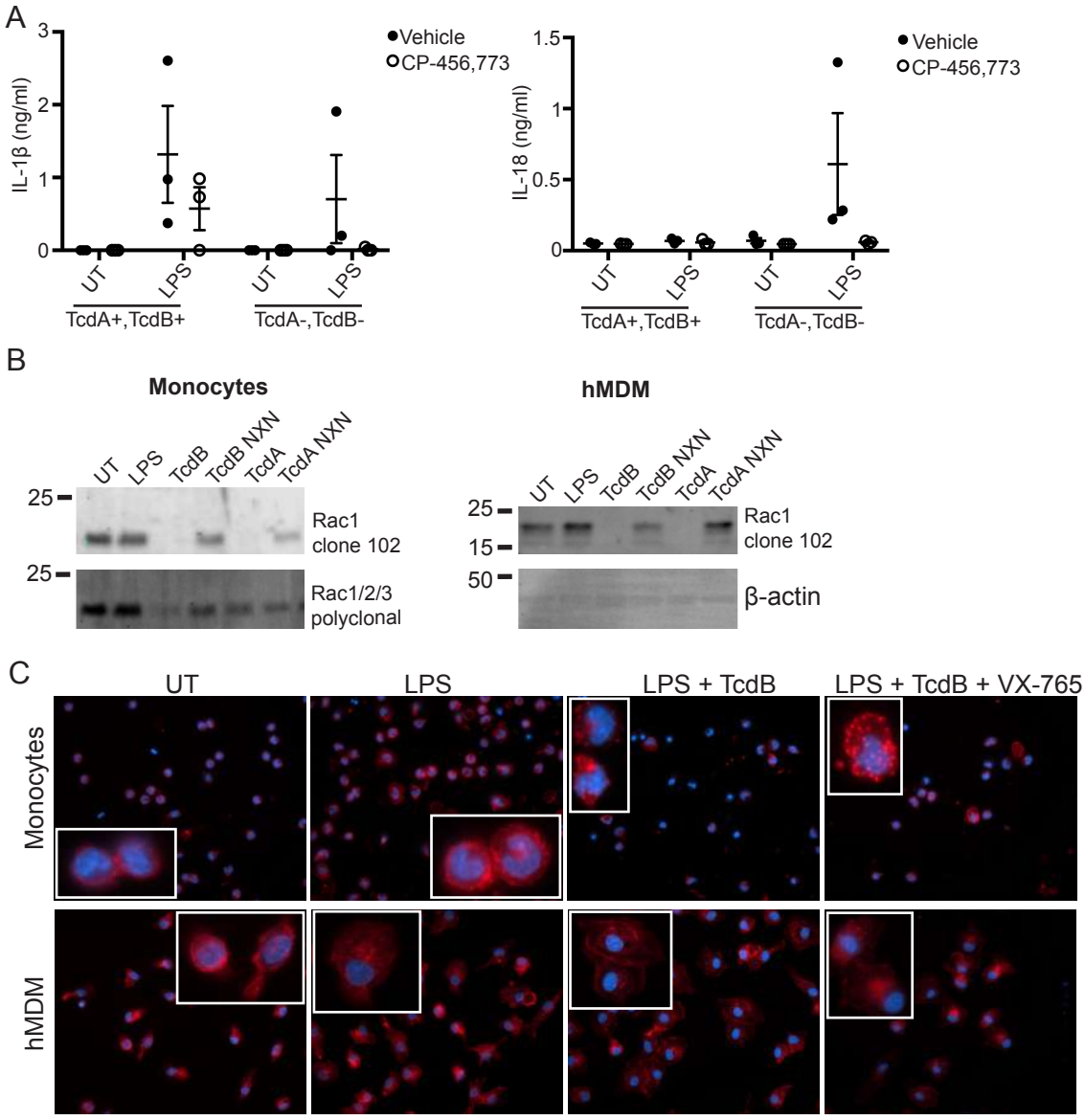

A
C.difficile conditioned supernatant
IL-1β (ng/ml)
Vehicle
CP-456,773
LPS
TcdA+,TcdB+
TcdA-,TcdB-
IL-18 (ng/ml)
B
Monocytes
hMDM
TcdB (ng/ml)
TcdA (ng/ml)
C
Monocytes
hMDM
kDa
Pyrin
β-actin
D
hMDM
Monocytes
DMSO
CP-456,773
VX-765
Colchicine
UT
Media
TcdB
Nigericin
LFN-PrgI /PA
+ LPS
% LDH release
E
hMDM
Monocytes
UT LPS
Nig
TcdB
LFN-PrgI /PA
DMSO
CP-456,773
VX-765
Supernatant
Lysate
p17 a-IL-1b
p20 a-caspase-1
p31 a-IL-1b
p42 a-caspase-1

Fig. 1


A
IL-1β (ng/ml)
Vehicle
CP-456,773
UT
LPS
TcdA+,TcdB+
TcdA-,TcdB-
IL-18 (ng/ml)
B
Monocytes
hMDM
UT
LPS
TcdB
TcdB NXN
TcdA
TcdA NXN
Rac1
clone 102
Rac1/2/3
polyclonal
β-actin
C
UT
LPS
LPS + TcdB
LPS + TcdB + VX-765
Monocytes
hMDM

Supp. fig. 1

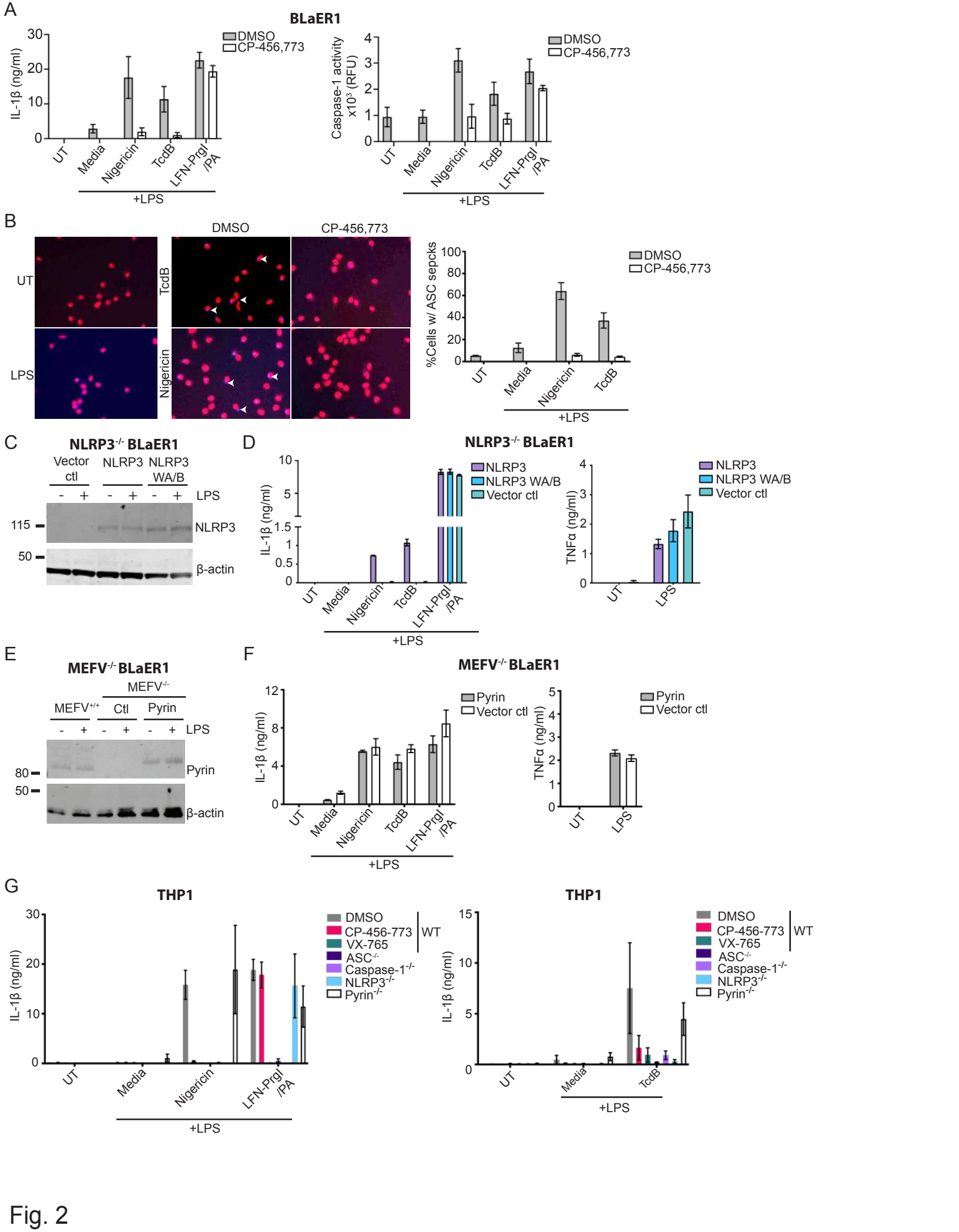
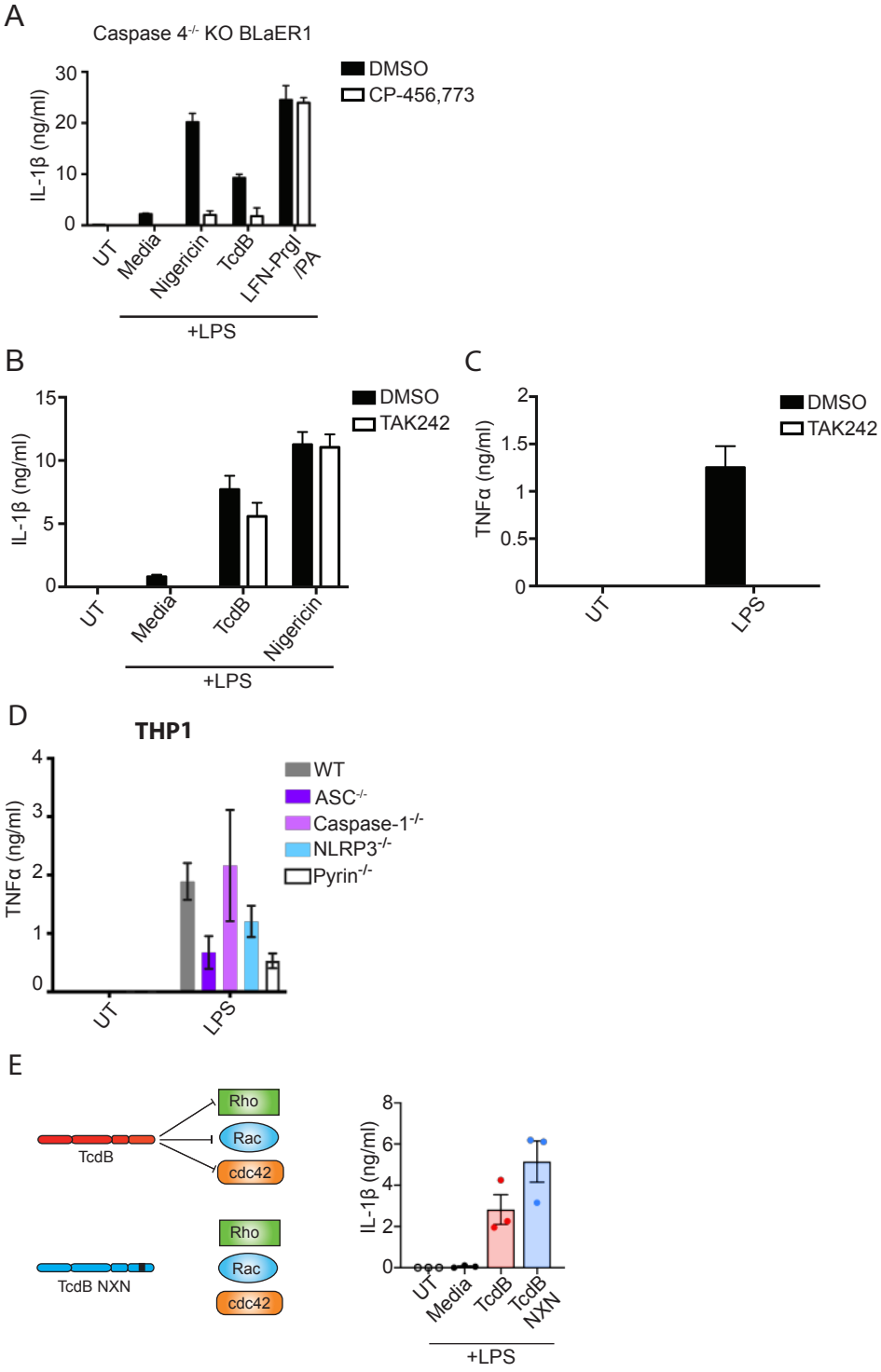

A
BLaER1
IL-1β (ng/ml)
DMSO
CP-456,773
UT
Media
Nigericin
TcdB
LFN-PrgI /PA
+LPS
Caspase-1 activity x10³ (RFU)
B
DMSO
CP-456,773
UT
LPS
TcdB
Nigericin
%Cells w/ ASC sepcks
C
NLRP3-/- BLaER1
Vector ctl
NLRP3
NLRP3 WA/B
LPS
115
50
NLRP3
β-actin
D
NLRP3-/- BLaER1
NLRP3
NLRP3 WA/B
Vector ctl
TNFα (ng/ml)
E
MEFV-/- BLaER1
MEFV+/+
MEFV-/-
Ctl
Pyrin
80
50
F
MEFV-/- BLaER1
G
THP1
WT
VX-765
ASC-/-
Caspase-1-/-
NLRP3-/-
Pyrin-/-

Fig. 2


A
Caspase 4-/- KO BLaER1
IL-1β (ng/ml)
30
20
10
0
DMSO
CP-456,773
UT
Media
Nigericin
TcdB
LFN-Prgl /PA
+LPS
B
IL-1β (ng/ml)
15
10
5
0
DMSO
TAK242
UT
Media
TcdB
Nigericin
+LPS
C
TNFα (ng/ml)
2
1.5
1
0.5
0
DMSO
TAK242
UT
LPS
D
THP1
TNFα (ng/ml)
4
3
2
1
0
WT
ASC-/-
Caspase-1-/-
NLRP3-/-
Pyrin-/-
UT
LPS
E
Rho
Rac
cdc42
TcdB
Rho
Rac
cdc42
TcdB NXN
IL-1β (ng/ml)
8
6
4
2
0
UT
Media
TcdB
TcdB NXN
+LPS

Supp. fig. 2

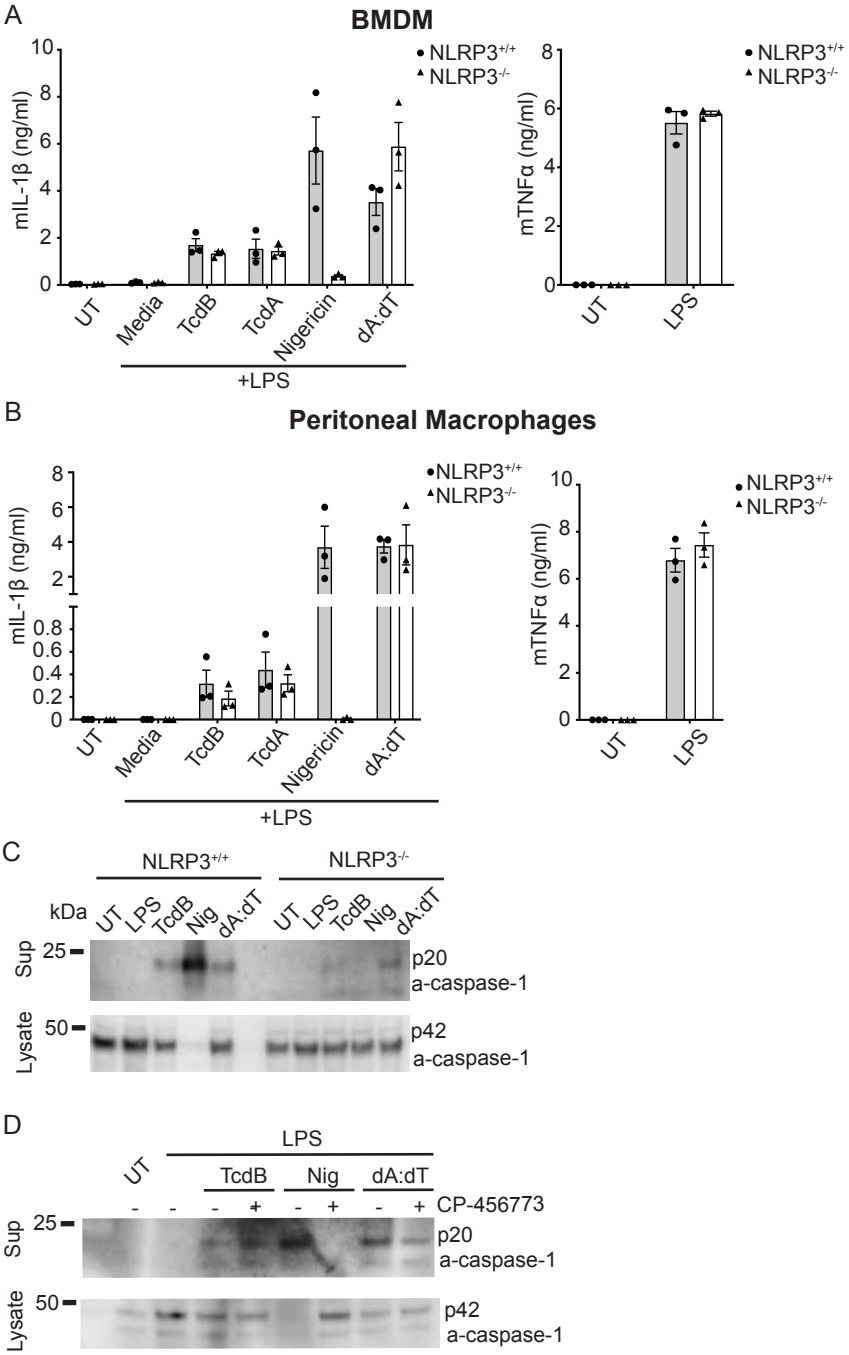
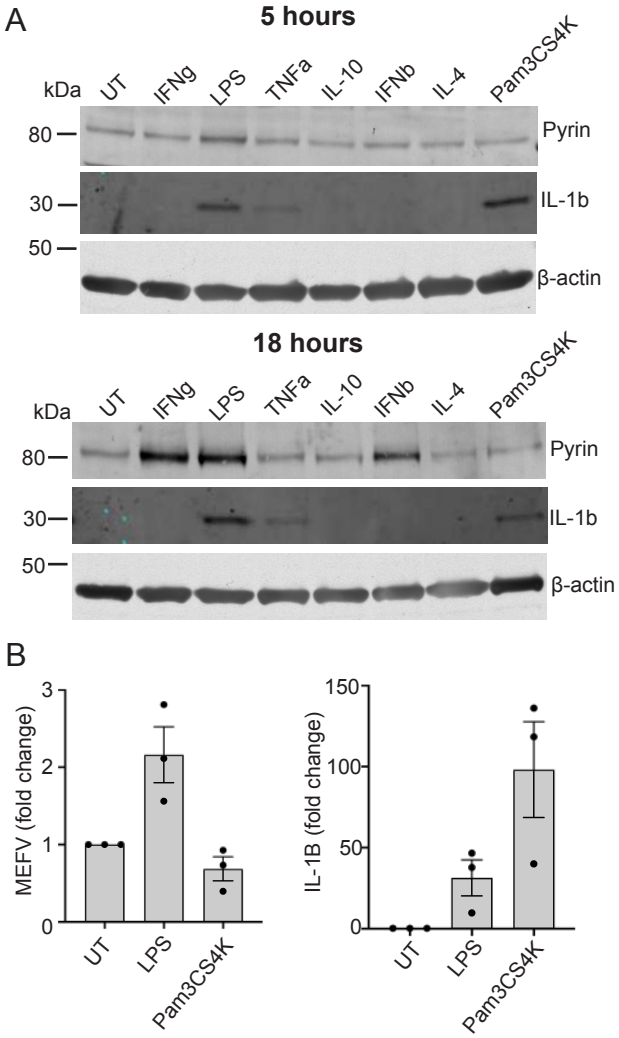

A
BMDM
mIL-1β (ng/ml)
NLRP3+/+
NLRP3-/-
UT
Media
TcdB
TcdA
Nigericin
dA:dT
+LPS
mTNFα (ng/ml)
LPS
B
Peritoneal Macrophages
C
NLRP3+/+
NLRP3-/-
kDa
Sup
Lysate
p20
a-caspase-1
p42
a-caspase-1
D
LPS
TcdB
Nig
dA:dT
CP-456773

Fig. 3


A
5 hours
kDa
UT
IFNg
LPS
TNFa
IL-10
IFNb
IL-4
Pam3CS4K
80
Pyrin
30
IL-1b
50
β-actin
18 hours
kDa
UT
IFNg
LPS
TNFa
IL-10
IFNb
IL-4
Pam3CS4K
80
Pyrin
30
IL-1b
50
β-actin

B
MEFV (fold change)
3
2
1
0
UT
LPS
Pam3CS4K
IL-1B (fold change)
150
100
50
0
UT
LPS
Pam3CS4K

Fig 4

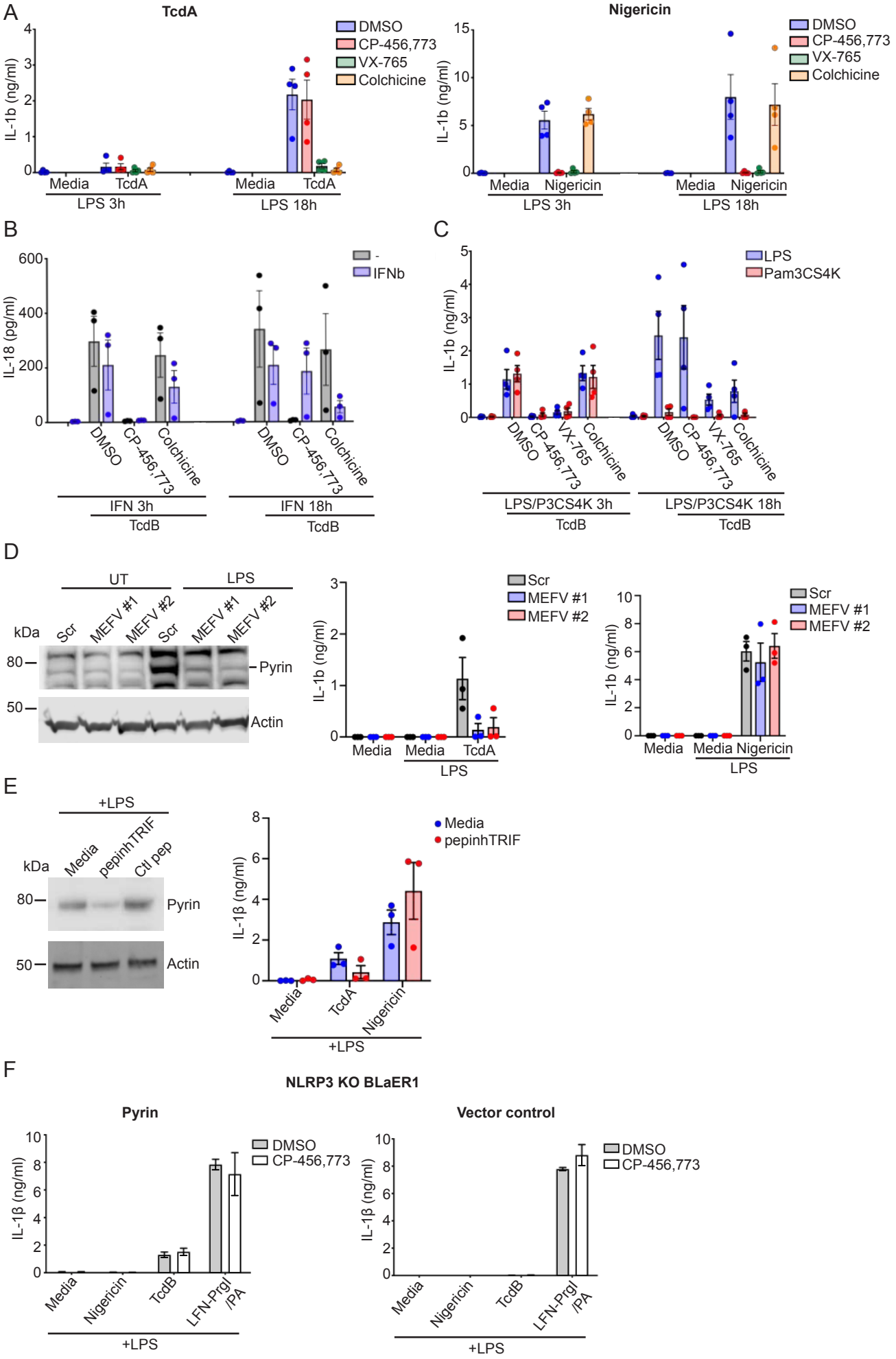

A
TcdA
IL-1b (ng/ml)
DMSO
CP-456,773
VX-765
Colchicine
Media
TcdA
LPS 3h
LPS 18h
Nigericin
B
IL-18 (pg/ml)
IFNb
IFN 3h
IFN 18h
TcdB
C
IL-1b (ng/ml)
LPS
Pam3CS4K
LPS/P3CS4K 3h
LPS/P3CS4K 18h
D
UT
LPS
kDa
Scr
MEFV #1
MEFV #2
Pyrin
Actin
E
+LPS
Media
pepinhTRIF
Ctl pep
IL-1β (ng/ml)
TcdA
Nigericin
F
NLRP3 KO BLaER1
Pyrin
Vector control
TcdB
LFN-PrgI /PA

Fig. 5

A

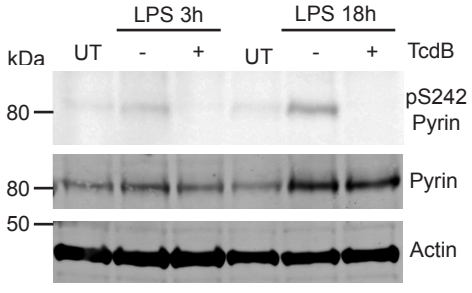


Fig. 6