- 1 Ultrashort treatment with telacebec alone and with companion drugs in immunocompetent and
- 2 immunosuppressed mouse footpad models of Buruli ulcer
- 4 Oliver Komm^{a,b}, Deepak V. Almeida^a, Paul J. Converse^a, Till F. Omansen^b, Eric L.
- 5 Nuermberger^{a, #}

6

11

13

15

- 7 Center for Tuberculosis Research, Department of Medicine, Johns Hopkins University,
- 8 Baltimore, Maryland, USA^a; Department of Tropical Medicine, Bernhard Nocht Institute for
- 9 Tropical Medicine & I. Dep. of Medicine, University Medical Center Hamburg-Eppendorf,
- 10 Hamburg, Germany^b
- Running head: Telacebec combination therapy for Buruli ulcer
- [#] Address correspondence to: Dr. Eric Nuermberger, enuermb@jhmi.edu

ABSTRACT

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

The antimicrobial treatment of Mycobacterium ulcerans infection, or Buruli ulcer (BU), has a long duration and is therefore burdensome and linked to indirect costs for affected patients. The new antimycobacterial drug telacebec (Q203) has previously shown promising treatmentshortening potential in mouse models of BU. In the present study, we investigated the potential of O203 to reduce the treatment duration further. The first experiment investigated the possibility of cure by one, three or five doses of Q203 (2 mg/kg) with or without a companion drug (bedaquiline, BDQ, clofazimine, CFZ, or clarithromycin, CLR) in immunocompetent BALB/c mice. The second experiment assessed the effect of five doses of Q203 with or without BDQ or CFZ on Mycobacterium ulcerans infection of immunocompromised SCID-beige mice with the aim to evaluate the contribution of host immunity to treatment efficacy. In BALB/c mice, a treatment duration as short as 3 days was sufficient to prevent relapse in nearly all footpads and a single dose of Q203 with or without BDQ or CFZ prevented relapse in approximately 50% of footpads. Unlike in BALB/c mice, a small percentage of SCID-beige mouse footpads were culture-positive after a treatment duration of five days, highlighting an important role of host immunity for M. ulcerans clearance. Our results confirm the marked potency and prolonged bactericidal and sterilizing effects of Q203, even immunocompromised SCID-beige mice.

KEYWORDS Buruli ulcer, Mycobacterium ulcerans, Q203, Telacebec, SCID-beige mice

INTRODUCTION

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

The neglected tropical disease Buruli ulcer (BU), which is caused by Mycobacterium ulcerans and which primarily causes skin and soft tissue lesions, is treated following WHO guidelines by a combination of 10 mg/kg rifampin (RIF, R) and 15-30 mg/kg clarithromycin (CLR, C) daily for 8 weeks, a long and burdensome treatment for the patients (1-5). This newer regimen, compared to the earlier regimen of RIF in combination with 15 mg/kg streptomycin (STR) daily for 8 weeks, avoids the risk of oto- and nephrotoxicity as well as the need for daily injections for 56 days. Nonetheless, the RIF+CLR treatment has disadvantages, such as frequent gastrointestinal intolerance, reduced CLR exposure due to a drug-drug interaction with RIF (6-9), and no reduction in the treatment duration of 8 weeks. Accordingly, even though RIF+CLR is a clinically proven effective therapy for BU (4, 5), the search for a shorter oral treatment for BU is a priority for improved control of BU. Previous studies have sought to improve the efficacy of oral BU treatment. Experiments in mice have tried replacing CLR and/or STR with drugs like clofazimine (CFZ) or oxazolidinones or to give higher doses of rifamycins. Some of these new regimens allowed a reduction of the treatment duration, but no regimen was able to cure mouse footpad infections in less than 4 weeks (10-14). The oxidative phosphorylation pathway was recently identified as a new drug target in Mycobacterium tuberculosis (15). Bedaquiline (BDQ) and telacebec (Q203, Q), two drugs acting at different steps on this pathway, proved to be efficacious for the treatment of BU (16-19). BDQ, an ATP synthase inhibitor, showed treatment shortening effects in regimens for drug-susceptible and multidrug-resistant tuberculosis (TB) (20-22). The recently rediscovered tuberculosis drug CFZ acts by being reduced by NDH-2, then reducing O2 resulting in the production of reactive oxygen species (23). Q203 is a mycobacterial cytochrome bc1:aa3 complex inhibitor that showed efficacy in mouse TB models and in an initial Phase 2 study in

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

TB patients (24, 25). An important advantage of Q203 for the treatment of M. ulcerans is the lack of the alternative terminal cytochrome bd oxidase in this species (26), as a result of a nonsense mutation in the cydA gene (base substitution - 692G>A) (18). Q203 proved to be extremely potent against M. ulcerans in vitro with an MIC between 0.075 and 0.15 ng/ml (16, 18) and also as monotherapy in the mouse footpad infection model (17) as well as in 3- or 4drug combination therapy with rifapentine, CFZ and/or BDQ (16). In both studies no CFU were detected 2 weeks after the end of a 2-week treatment. The addition of RIF at 10 or 20 mg/kg did not add a benefit (17). In the present study using BALB/c mice we investigated if shorter treatment durations (1, 3 or 5 days) of Q203 at 2 mg/kg in monotherapy or 2-drug combination therapy with BDQ, CFZ or CLR, render mouse footpads culture negative, how long it takes to render the footpads negative, and if mice relapse after a 15-week observation period. In the present study using immunocompromised SCID-beige mice we addressed the question of whether the continued reduction in CFU counts after completion of short courses of Q203 treatment in immunocompetent mice (16, 17) is primarily due to a prolonged antimicrobial effect of Q203 or due to the adaptive host immune response. Additionally, we tested the SCID-beige mouse footpads for emergence of bacterial resistance against Q203 and compared the results between groups, to assess the necessity of a companion drug for Q203 to prevent resistance. **RESULTS** Experiment 1: To determine if addition of a companion drug increases the sterilizing activity of Q203 and to find the shortest possible treatment duration In this experiment, BALB/c mice were treated with 2 mg/kg Q203 (Q2) alone or in combination with one of the following drugs: 25 mg/kg BDQ (Q₂BDQ₂₅), 12.5 mg/kg CFZ (Q₂CFZ_{12.5}), or

89

90

91

92

93

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

100 mg/kg CLR (Q₂CLR₁₀₀). Drugs were administered once daily via oral gavage and each regimen was given for 1, 3 and 5 days. Cohorts of mice were followed for up to 15 weeks after the start of treatment (Table S1). Footpad swelling and CFU counts up to 5 weeks post-treatment: On the day after infection, the mean (\pm SD) CFU count was $4.95 \pm 0.38 \log_{10}$ CFU/footpad. At the start of treatment (D0), 34 days after infection, the mean CFU count was $6.62 \pm 0.34 \log_{10}$ CFU/footpad, and the median swelling grade was 2 on a scale of 0-4 (27). Untreated mice subsequently exhibited increased footpad swelling and required euthanasia during the experiment (Fig. 1A), with the exception of one untreated mouse that survived until the end of the experiment without reaching the endpoint for euthanasia despite harboring $6.17 \pm 0.86 \log_{10}$ CFU/footpad at the end of the experiment. In mice treated with R₁₀CLR₁₀₀ for 10 consecutive days, the swelling decreased slightly to an average of 1.7 after one week of treatment and to 1.25 after 2 weeks. The decrease in swelling continued up to Week 4 and was maintained until Week 8, when a gradual increase in swelling started and ultimately resulted in four footpads having a swelling ≥ 3 by Week 14. The mean CFU count in $R_{10}CLR_{100}$ -treated mice decreased to $3.81 \pm 1.35 \log_{10} CFU/footpad$ by Week 2 (Fig. 1B) but increased after the end of treatment to $5.49 \pm 1.21 \log_{10}$ CFU/footpad by Week 15. As previously observed (17), Q203-containing regimens rapidly decreased footpad swelling, resulting in >98% of these mice having a swelling grade ≤1 by Week 1, significantly lower than the R₁₀CLR₁₀₀ control group (p<0.001). Among mice receiving Q203, 36% and 67% of the footpads showed no swelling by Weeks 2 and 4, respectively, with no significant difference between Q203-treated groups. In the first week, the superior effect of Q203 was not yet reflected in the footpad CFU counts. Five weeks after the end of treatment, all Q203containing regimens given for 5 days resulted in culture-negative footpads (Fig. 1C). With the

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

exception of three of 6 footpads from mice treated for 3 days with Q₂CFZ_{12.5}, footpads treated with a O203-containing regimen for three days were all culture-negative. In the groups treated for only one day, 1 mouse (both footpads) that received Q203 alone had positive culture results, all mice receiving Q₂BDQ₂₅ for one day were culture-negative, one mouse receiving Q₂CFZ_{12.5} for one day had a culture-positive footpad, and in the group receiving Q₂CLR₁₀₀ for one day, two mice had both footpads culture-positive at this time point. At Week 6, the CFU counts were significantly higher in mice treated with one dose of Q₂CLR₁₀₀ compared to one dose of Q_2BDQ_{25} (p<0.03). Relapse: Eight BALB/c mice of each treatment group were held for 15 weeks to evaluate relapse. Apart from the relapses observed in all R₁₀CLR₁₀₀-treated mice, we also observed relapse in all but 3 of the 16 footpads treated with a single dose of Q₂CLR₁₀₀. A single dose of Q203 resulted in 9 out of 16 footpads being culture-negative, a single dose of Q203 and BDQ resulted in half (8/16) of the footpads being culture-negative, one dose of Q₂CFZ_{12.5} resulted in 7 of 16 footpads being culture-negative. At Week 15, the CFU counts were significantly higher in mice treated with one dose of Q₂CLR₁₀₀ compared to one dose of Q₂O₃ alone, Q₂BDQ₂₅ or Q₂CFZ_{12.5} (p<0.02). When given for 3 days, Q203 with or without a companion drug prevented relapse in all footpads with the exception of 2 out of 16 footpads from mice treated with Q₂CLR₁₀₀. Q₂ alone or with a companion drug given for 5 days prevented relapse in all footpads (Fig. 1D). The limit of detection was 1 CFU. Experiment 2: To determine the impact of the host immune response on the sterilizing activity of Q203-containing regimens and the selection of Q203-resistant mutants

137

138

139

140

141

142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

158

159

Experiment 2 was conducted to determine if a five-day treatment with 2 mg/kg Q203 (Q2) daily, alone or in combination with 25 mg/kg BDO (O₂BDO₂₅) daily or 12.5 mg/kg CFZ (O₂CFZ_{12.5}) daily can successfully treat M. ulcerans footpad infection in SCID-beige mice without selection of Q203-resistant mutants (Table S2). Footpad swelling and CFU count results up to 4 weeks post-treatment: One day after infection, the mean (\pm SD) CFU count was $4.17 \pm 0.22 \log_{10}$ CFU/footpad. At the start of treatment (D0), 40 days after infection, the mean footpad swelling was 2.5 (Fig. 2A), the mean CFU count was $6.26 \pm 0.35 \log_{10}$ CFU/footpad for the mice allocated to the Q203-containing regimens and 6.56 \pm 0.29 log₁₀ CFU/footpad for the mice allocated to the R₁₀CLR₁₀₀ control group (Fig. 2B). In all Q203-containing treatment groups, a sharp reduction in footpad swelling to a median swelling grade of 0.25 after one week of treatment was observed; in the Q₂CFZ_{12.5} group, 19 of 34 footpads had a swelling grade of 0, and in the Q₂ and Q₂BDQ₂₅ groups, 7 of 34 and 8 of 34 footpads, respectively, had a swelling grade of 0. In contrast, the swelling of untreated mouse footpads continued to increase such that the mice reached the humane endpoint for euthanasia at Week 1, while the control R₁₀CLR₁₀₀ regimen decreased footpad swelling after one week to a median swelling grade of only 1.5, which was still significantly higher than in the Q203containing regimens (p<0.001). After the end of treatment in every group receiving Q203 the median swelling grade was 0, whereas the swelling started to increase in R₁₀CLR₁₀₀-treated mice again, reaching median swelling grades of 2.1 at Week 2 and ≥3 at Week 3, mandating that all R₁₀CLR₁₀₀-treated mice be euthanized per protocol at Week 4. Footpads in the Q203treated groups maintained a swelling grade of approximately 0 until the end of the experiment. At Week 3 only 2 of the 24 footpads from mice treated with a Q203 containing regimen were culture-negative.

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

Trends in footpad CFU counts largely mirrored the swelling grades, although there was not as much discrimination between the R₁₀CLR₁₀₀ control group and the Q203-treated groups at Week 1 and the minimum values were not reached in Q203-treated groups until Week 5 (4 weeks after treatment completion) or later. Three weeks after the end of treatment, when all R₁₀CLR₁₀₀-treated mice were euthanized per protocol, their footpad CFU counts had still not decreased and were near the pre-treatment baseline. In contrast, in Q203-treated mice, footpad CFU counts continued to decline after dosing ended and, by Week 5, all footpads in the Q₂BDQ₂₅-treated group were culture-negative, in the Q₂CFZ_{12.5} group two footpads (from two different mice) were culture-negative, and in the Q203 alone group three footpads (including both footpads in one mouse) were culture-negative (Fig. 2C). The differences between the Q_2BDQ_{25} and the Q203 alone and $Q_2CFZ_{12.5}$ groups were statistically significant (p <0.03). The median footpad swelling grade as well as the mean CFU count in mice treated with any Q203containing regimen was significantly (p<0.0001) lower than those in the R₁₀CLR₁₀₀ control group. The limit of detection was 3 CFU per footpad. Relapse: Five SCID-beige mice of each Q203-containing treatment group were held for 17 weeks after starting treatment. No mouse showed a relapse in footpad swelling. In the group receiving Q203 alone, 2 out of 10 footpads had detectable CFU, in the Q2CFZ_{12.5} group one footpad had detectable CFU, and in the Q₂BDQ₂₅ group 2 of 10 footpads had 1 CFU each (limit of detection, 1 CFU) (Fig. 2D). Resistance detection: The M. ulcerans isolates from the eight SCID-beige footpads with the highest CFU counts at Week 5 (4 weeks post-treatment) were selected for Q203 susceptibility testing. Two of the four footpad isolates from mice receiving Q203 alone showed resistance (40% and 2.2% of total CFU on 3 ng/ml, 40% and 3.9% of total CFU on 0.3 ng/ml, respectively)

(Table S3). The Q203-containing plates from two $Q_2CFZ_{12.5}$ -treated footpads were contaminated and were not assessable, whereas the isolates from the other two footpads from $Q_2CFZ_{12.5}$ -treated mice did not show resistance.

DISCUSSION

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

207

208

209

The standard treatment for BU is an 8-week course of RIF+CLR (3), an improvement compared to the previously recommended combination of RIF+STR, as the new regimen is fully oral and avoids the previously required injections. A shorter regimen remains a high priority in the search for a better treatment of BU (19, 28). Q203, a potent inhibitor of the respiratory chain of M. ulcerans, is a promising candidate for ultra-short, and possibly singledose, treatment for BU (17, 19). It has proven to be capable of significantly shortening the treatment duration in mouse footpad models (16, 17, 19, 29). Furthermore, a clinical trial showed no serious adverse effects from Q203 when taken daily for 2 weeks in a small group of TB patients (24). In January of 2021, the US Food and Drug Administration granted an orphan designation Q203 BU drug for treatment (http://www.koreabiomed.com/news/articleView.html?idxno=10169), providing further incentives for clinical development of Q203 for this indication. In the present study in M. ulcerans-infected BALB/c mice, Q203 given alone or in

In the present study in *M. ulcerans*-infected BALB/c mice, Q203 given alone or in combination with BDQ or CFZ for 3 days was sufficient to render all footpads culture-negative 15 weeks after start of treatment. When Q203 was administered alone or in combination with BDQ, all footpads were culture-negative even five and a half weeks after the last dose. The total dose of Q203 amounted to 6 mg/kg. To our knowledge, this is the lowest total dose of Q203 to render all footpads culture-negative in a mouse footpad infection model.

No tested combination including Q203 at 2 mg/kg rendered all footpads culture-negative after a single dose. For this purpose, higher doses or other companion drugs may be necessary

to achieve a single-dose cure. In BALB/c mice, none of the combinations tested was significantly more effective than Q203 alone.

210

211

212

213

214

215

216

217

218

219

220

221

222

223

224

225

226

227

228

229

230

231

232

233

To compare the sterilizing potency of Q203 and combination regimens in immunocompetent BALB/c mice and immunocompromised SCID-beige mice we analyzed the swelling grades and CFU counts in mice of each strain treated with the same regimen. All Q203containing treatments rapidly resolved footpad swelling in SCID-beige mice, as in BALB/c mice. However, the rate of bacterial killing and the extent of bacterial eradication, while great in both strains, were not as great in SCID-beige mice. Nevertheless, by Week 15, all groups showed a relapse-free survival of at least 80% in SCID-beige mice, this result shows, that a 5day treatment with Q203 alone (total dose of 10 mg/kg) or with a companion drug was sufficient to render the majority of footpads culture-negative and prevent relapse, even in mice lacking an adaptive immune response. At Week 5, the difference between Q2BDQ25, which rendered all footpads culture-negative, and Q203 alone, which rendered 3 of 8 footpads culture negative, and Q₂CFZ_{12.5}, which rendered 2 of 8 footpads culture negative, was statistically significant. However, at the end of the follow-up period, the difference in CFU counts was not significant. The difference between BALB/c mice, in which all footpads were culture-negative, and SCID-beige mice, in which no group had all footpads culture-negative, is striking. This difference illustrates the importance of the immune system in the successful clearance of M. ulcerans. Our results suggest that treatment of BU in immunosuppressed patients, for example in the case of HIV coinfection or patients undergoing iatrogenic immunosuppression, may need to be adjusted to reach a complete cure in all patients when using Q203. The continued reduction in CFU counts supports a conclusion that the prolonged activity of Q203 is primarily due to drug effects, but the incomplete sterilizing activity of the tested regimens in SCID-beige mice illustrates the contribution of the adaptive immune system to the overall treatment response.

Further research might be needed in immunodeficient mice to find a better treatment for patients, who are immune-deficient.

We did observe selection of Q203 resistance during Q203 monotherapy in SCID-beige mice. No resistance was observed when Q203 was combined with CFZ. Although only a limited number of isolates was assessed, this finding suggests that a companion drug might be beneficial to prevent emergence of Q203 resistance in very immunocompromised hosts. However, it should be recognized that SCID-beige mice represent an extreme state of immunosuppression and the majority of these mice receiving monotherapy appeared to be cured. Therefore, the pros and cons of monotherapy versus combination therapy will require careful consideration. The overall positive outcomes in SCID-beige mice, the simplicity of monotherapy and its reduced cost and potential for adverse effects, as well as the very low likelihood of human-to-human transmission of Q203-resistant BU (30, 31), should it occur, would seem to favor monotherapy. Nevertheless, further investigations may be warranted to identify an optimal companion drug that could better assure the efficacy of ultra-short (i.e., 1-3 dose) regimens across the spectrum of host immune competency (11, 32).

In summary, we identified the lowest-yet published total dose of Q203 leading to a complete healing of all affected footpads in this mouse model. Surprisingly, none of the companion drugs evaluated, contributed an additional treatment-shortening effect in BALB/c mice, nor a higher overall rate of cure in SCID-beige mice. Our studies once again show the treatment-shortening capability of Q203 but also raise the question of the optimal treatment for immunosuppressed individuals with BU.

Materials and Methods

Bacterial strain. For this study *M. ulcerans* strain 1059, which was originally isolated from a

patient in Ghana, was used (33).

259

260

261

262

263

264

265

266

267

268

269

270

271

272

273

274

275

276

277

278

279

280

281

282

Antibiotics. Q203 and BDQ were kindly provided by the TB Alliance. CFZ and RIF were purchased from Sigma-Aldrich (St. Louis, MO, USA). CLR was purchased from the Johns Hopkins Hospital Pharmacy. Q203 was prepared in D-α tocopheryl polyethylene glycol 1000 succinate solution. BDQ was formulated in acidified 20% hydroxypropyl-β-cyclodextrin solution. CFZ, RIF and CLR were prepared in a sterile 0.05% (wt/vol) agarose solution in distilled water. Mouse infection. Female 6-week-old Fox Chase SCID-beige and female BALB/c mice (Charles River Laboratories) were inoculated subcutaneously in both hind footpads with 0.03 ml of a culture suspension containing M. ulcerans 1059. Treatment started 5-to-6 weeks (D0) after infection when the mice had footpad swelling of grade 2-3 on a scale of 0 to 4 (27). The CFU counts at implantation were evaluated on six footpads from three mice on the day after infection (D-40 and D-34) and at the start of treatment (D0) to determine the infectious dose and the pretreatment CFU counts, respectively. The response to treatment was determined by plating each hind footpad from mice at predetermined time points. Treatment. Mice were treated 5 days per week with 0.2 ml per dose by gavage. Control mice treated with RIF and CLR were treated for 10 days continuously without interruption. Drug doses were chosen based on mean plasma exposures (similar area under the curve [AUC] in 24h after dose) similar to human exposures at standard doses (16, 17). All animal procedures were conducted in adherence with the Animal Welfare Act and Public Health Service Policy and other national and international guidelines. The procedures were approved by the Johns Hopkins University Animal Care and Use Committee. Experiment 1. BALB/c mice (n=191 mice) were infected 5 weeks before the treatment start. The control regimen was given in the form of RIF₁₀CLR₁₀₀ for 10 consecutive days. One group of mice (6 mice) received no treatment, 6 footpads were collected one week after the beginning of treatment.

284

285

286

287

288

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

304

305

306

To each study group (i.e. Q203-containing groups), 14 BALB/c mice were allocated. Three groups received 2 mg/kg O203 alone, three groups received 2 mg/kg O203 plus 25 mg/kg BDQ, three groups received 2 mg/kg Q203 plus 12.5 mg/kg CFZ, three groups received 2 mg/kg Q203 plus 100 mg/kg CLR. Each treatment was given once daily in one group for 5 consecutive days, in one group for 3 consecutive days and in one group for one day. Six footpads per group were harvested for determination of CFU counts at 1 week and 6 weeks after the start of treatment; and 16 footpads per group were harvested 15 weeks from the start of treatment. Experiment 2. SCID-beige mice (n=76 mice) were infected 6 - 7 weeks before the treatment start. The control regimen was given in the form of R₁₀CLR₁₀₀ treatment for 5 consecutive days. Three mice were held for another week without treatment. During this time the swelling increased and reached the threshold for euthanasia per protocol. At this point the mice were sacrificed and the footpads harvested for CFU counts. To each study group (i.e., Q203-containing groups), 17 SCID-beige mice were allocated. Each group was treated once daily for 5 consecutive days. One group received 2 mg/kg Q203 alone, one group received 2 mg/kg Q203 plus 25 mg/kg BDQ, one treatment group received 2 mg/kg Q203 plus 12.5 mg/kg CFZ. At 1, 3 and 5 weeks after the start of treatment, 8 footpads per group were evaluated for response to treatment. Five additional mice from each treatment group were held for 17 weeks from the start of treatment before each footpad was harvested. Evaluation of treatment response. Treatment response was evaluated by (i) change in footpad swelling, as previously described (27), and (ii) change in log₁₀-transformed CFU counts. The inflammatory swelling was evaluated weekly and scored as a swelling grade between 0 and 4, where a swelling grade of 0 corresponds to a normal footpad, swelling grade 1 to a swollen but non-inflammatory footpad, swelling grade 2 to inflammatory swelling of the footpad, swelling

308

309

310

311

312

313

314

315

316

317

318

319

320

321

322

323

324

325

326

327

328

329

330

grade 3 to swelling of the entire hind foot and swelling grade 4 to an inflammatory leg swelling and ulcerations. To determine footpad CFU counts, the footpad was thoroughly disinfected with 70% alcohol swabs and afterwards the footpad tissue was removed. The tissue was then mechanically finely minced and suspended in 1.5 ml sterile phosphate-buffered saline (PBS). Undiluted and serial 10-fold diluted aliquots (each 0.5ml) were cultured on Middlebrook 7H11selective agar, supplemented with 10% oleic acid-albumin-dextrose-catalase (OADC), at 32°C for up to 10 weeks before CFU counts were calculated. Relapse was evaluated by holding mice for 15 weeks (Experiment 1) or 17 weeks (Experiment 2) from the start of treatment. After the end of treatment, footpads were examined every 2 weeks until the final time point was reached. Mice that re-developed footpad swelling with a lesion index ≥3 were sacrificed and the footpads plated for CFU counts. For mice reaching the final endpoint, the entire footpad homogenate was plated. Resistance testing in SCID-beige mice. To evaluate for selection of Q203-resistant mutants in Experiment 2, isolates were collected from culture plates from each of four mice treated with Q203 alone or Q₂CFZ_{12.5} at the Week 5 time point. Each isolate consisted of colonies from one plate that were pooled, homogenized and then re-plated onto drug-free 7H11 agar and agar containing Q203 concentrations of 0.3 ng/ml and 3 ng/ml (4 times and 40 times MIC, respectively). The isolate was determined to be Q203-resistant if the proportion obtained by dividing the CFU count on Q203-containing plate by the CFU count on drug-free plates was ≥ 0.01. Statistical analysis. GraphPad Prism 9 was used to compare CFU counts and swelling grades at week 1 in Q203-treatment groups to the R₁₀CLR₁₀₀ and untreated control groups using oneway analysis of variance with Dunnett's posttest to adjust for multiple comparisons. For the

following timepoints we used one-way analysis of variance (Kruskal-Wallis test) corrected for multiple comparisons using Dunn's test.

Acknowledgments

This study was supported by the National Institutes of Health (R01-AI113266). O.K. was supported by a personal grant from the Bernhard-Nocht-Institute for Tropical Medicine, Hamburg, Germany. We gratefully thank the TB Alliance for providing Q203 and bedaquiline.

References

339

340 Röltgen K, Pluschke G. 2019. Buruli Ulcer: History and Disease Burden, p 1-41. In 341 1. 342 Pluschke G, Röltgen K (ed), Buruli Ulcer: Mycobacterium ulcerans Disease doi:10.1007/978-3-030-11114-4 1. Springer Copyright 2019, The Author(s). Cham 343 (CH). 344 Chukwu JN, Meka AO, Nwafor CC, Oshi DC, Madichie NO, Ekeke N, Anyim MC, 2. 345 Chukwuka A, Obinna M, Adegbesan J, Njoku M, Soyinka FO, Adelokiki AO, 346 Enemuoh IO, Okolie PI, Edochie JE, Offor JB, Ushaka J, Ukwaja KN. 2017. Financial 347 348 burden of health care for Buruli ulcer patients in Nigeria: the patients' perspective. Int 349 Health 9:36-43. Organization WH. 2017. Report from the meeting of the Buruli ulcer Technical 350 3. Advisory Group. World Health Organization, Geneva, Switzerland. 351 4. Phillips RO, Robert J, Abass KM, Thompson W, Sarfo FS, Wilson T, Sarpong G, 352 Gateau T, Chauty A, Omollo R, Ochieng Otieno M, Egondi TW, Ampadu EO, 353 354 Agossadou D, Marion E, Ganlonon L, Wansbrough-Jones M, Grosset J, Macdonald JM, Treadwell T, Saunderson P, Paintsil A, Lehman L, Frimpong M, Sarpong NF, 355 Saizonou R, Tiendrebeogo A, Ohene SA, Stienstra Y, Asiedu KB, van der Werf TS. 356 2020. Rifampicin and clarithromycin (extended release) versus rifampicin and 357 streptomycin for limited Buruli ulcer lesions: a randomised, open-label, non-inferiority 358 phase 3 trial. Lancet 395:1259-1267. 359 360 5. Friedman ND, Athan E, Walton AL, O'Brien DP. 2016. Increasing Experience with Primary Oral Medical Therapy for Mycobacterium ulcerans Disease in an Australian 361 Cohort. Antimicrob Agents Chemother 60:2692-5. 362

6. O'Brien DP, Friedman D, Hughes A, Walton A, Athan E. 2017. Antibiotic 363 complications during the treatment of Mycobacterium ulcerans disease in Australian 364 patients. Intern Med J 47:1011-1019. 365 7. Koh WJ, Jeong BH, Jeon K, Lee SY, Shin SJ. 2012. Therapeutic drug monitoring in 366 the treatment of *Mycobacterium avium* complex lung disease. Am J Respir Crit Care 367 Med 186:797-802. 368 8. van Ingen J, Egelund EF, Levin A, Totten SE, Boeree MJ, Mouton JW, Aarnoutse RE, 369 Heifets LB, Peloquin CA, Daley CL. 2012. The pharmacokinetics and 370 pharmacodynamics of pulmonary Mycobacterium avium complex disease treatment. 371 372 Am J Respir Crit Care Med 186:559-65. 9. Williams KN, Bishai WR. 2005. Clarithromycin extended-release in community-373 acquired respiratory tract infections. Expert Opin Pharmacother 6:2867-76. 374 375 10. Almeida D, Converse PJ, Ahmad Z, Dooley KE, Nuermberger EL, Grosset JH. 2011. Activities of rifampin, Rifapentine and clarithromycin alone and in combination 376 against Mycobacterium ulcerans disease in mice. PLoS Negl Trop Dis 5:e933. 377 11. Almeida DV, Omansen TF, Li SY, Lee J, Grosset JH, Converse PJ, Nuermberger EL. 378 379 2019. Oxazolidinones Can Replace Clarithromycin in Combination with Rifampin in a 380 Mouse Model of Buruli Ulcer. Antimicrob Agents Chemother 63. 12. Omansen TF, Almeida D, Converse PJ, Li SY, Lee J, Stienstra Y, van der Werf T, 381 Grosset JH, Nuermberger EL. 2019. High-Dose Rifamycins Enable Shorter Oral 382 383 Treatment in a Murine Model of *Mycobacterium ulcerans* Disease. Antimicrob Agents Chemother 63. 384 13. Converse PJ, Tyagi S, Xing Y, Li SY, Kishi Y, Adamson J, Nuermberger EL, Grosset 385 JH. 2015. Efficacy of Rifampin Plus Clofazimine in a Murine Model of 386 Mycobacterium ulcerans Disease. PLoS Negl Trop Dis 9:e0003823. 387

Converse PJ, Almeida DV, Tasneen R, Saini V, Tyagi S, Ammerman NC, Li SY, 388 14. Anders NM, Rudek MA, Grosset JH, Nuermberger EL. 2018. Shorter-course 389 treatment for Mycobacterium ulcerans disease with high-dose rifamycins and 390 clofazimine in a mouse model of Buruli ulcer. PLoS Negl Trop Dis 12:e0006728. 391 Bald D, Villellas C, Lu P, Koul A. 2017. Targeting Energy Metabolism in 392 15. Mycobacterium tuberculosis, a New Paradigm in Antimycobacterial Drug Discovery. 393 mBio 8. 394 Converse PJ, Almeida DV, Tyagi S, Xu J, Nuermberger EL. 2019. Shortening Buruli 16. 395 Ulcer Treatment with Combination Therapy Targeting the Respiratory Chain and 396 397 Exploiting Mycobacterium ulcerans Gene Decay. Antimicrobial Agents and Chemotherapy 63:e00426-19. 398 17. Almeida DV, Converse PJ, Omansen TF, Tyagi S, Tasneen R, Kim J, Nuermberger 399 400 EL. 2020. Telacebec for Ultrashort Treatment of Buruli ulcer in a Mouse Model. Antimicrob Agents Chemother 64. 401 18. Scherr N, Bieri R, Thomas SS, Chauffour A, Kalia NP, Schneide P, Ruf M-T, 402 Lamelas A, Manimekalai MSS, Grüber G, Ishii N, Suzuki K, Tanner M, Moraski GC, 403 404 Miller MJ, Witschel M, Jarlier V, Pluschke G, Pethe K. 2018. Targeting the 405 Mycobacterium ulcerans cytochrome bc1:aa3 for the treatment of Buruli ulcer. Nature Communications 9:5370. 406 19. Thomas SS, Kalia NP, Ruf MT, Pluschke G, Pethe K. 2020. Toward a Single-Dose 407 Cure for Buruli ulcer. Antimicrob Agents Chemother 64. 408 20. Lanoix JP, Betoudji F, Nuermberger E. 2014. Novel regimens identified in mice for 409 treatment of latent tuberculosis infection in contacts of patients with multidrug-410 resistant tuberculosis. Antimicrob Agents Chemother 58:2316-21. 411

21. Diacon AH, Pym A, Grobusch MP, de los Rios JM, Gotuzzo E, Vasilyeva I, Leimane 412 413 V, Andries K, Bakare N, De Marez T, Haxaire-Theeuwes M, Lounis N, Meyvisch P, De Paepe E, van Heeswijk RP, Dannemann B. 2014. Multidrug-resistant tuberculosis 414 and culture conversion with bedaquiline. N Engl J Med 371:723-32. 415 22. Tiberi S, du Plessis N, Walzl G, Vjecha MJ, Rao M, Ntoumi F, Mfinanga S, Kapata N, 416 Mwaba P, McHugh TD, Ippolito G, Migliori GB, Maeurer MJ, Zumla A. 2018. 417 Tuberculosis: progress and advances in development of new drugs, treatment 418 regimens, and host-directed therapies. Lancet Infect Dis 18:e183-e198. 419 23. Yano T, Kassovska-Bratinova S, Teh JS, Winkler J, Sullivan K, Isaacs A, Schechter 420 421 NM, Rubin H. 2011. Reduction of clofazimine by mycobacterial type 2 NADH:quinone oxidoreductase: a pathway for the generation of bactericidal levels of 422 reactive oxygen species. J Biol Chem 286:10276-87. 423 424 24. de Jager VR, Dawson R, van Niekerk C, Hutchings J, Kim J, Vanker N, van der Merwe L, Choi J, Nam K, Diacon AH. 2020. Telacebec (Q203), a New 425 Antituberculosis Agent. N Engl J Med 382:1280-1281. 426 25. Pethe K, Bifani P, Jang J, Kang S, Park S, Ahn S, Jiricek J, Jung J, Jeon HK, Cechetto 427 428 J, Christophe T, Lee H, Kempf M, Jackson M, Lenaerts AJ, Pham H, Jones V, Seo 429 MJ, Kim YM, Seo M, Seo JJ, Park D, Ko Y, Choi I, Kim R, Kim SY, Lim S, Yim SA, Nam J, Kang H, Kwon H, Oh CT, Cho Y, Jang Y, Kim J, Chua A, Tan BH, 430 Nanjundappa MB, Rao SP, Barnes WS, Wintjens R, Walker JR, Alonso S, Lee S, Kim 431 J, Oh S, Oh T, Nehrbass U, Han SJ, No Z, et al. 2013. Discovery of Q203, a potent 432 clinical candidate for the treatment of tuberculosis. Nat Med 19:1157-60. 433 26. Mascolo L, Bald D. 2020. Cytochrome bd in Mycobacterium tuberculosis: A 434 respiratory chain protein involved in the defense against antibacterials. Prog Biophys 435 Mol Biol 152:55-63. 436

27. Dega H, Bentoucha A, Robert J, Jarlier V, Grosset J. 2002. Bactericidal activity of 437 rifampin-amikacin against Mycobacterium ulcerans in mice. Antimicrob Agents 438 Chemother 46:3193-6. 439 28. Barksby R. 2019. WHO takes a new approach to Buruli ulcer. Lancet Infect Dis 440 441 19:473-474. 29. Chauffour A, Robert J, Veziris N, Aubry A, Pethe K, Jarlier V. 2020. Telacebec 442 (Q203)-containing intermittent oral regimens sterilized mice infected with 443 Mycobacterium ulcerans after only 16 doses. PLoS Negl Trop Dis 14:e0007857. 444 30. Exner K, Lemperle G. 1987. [Buruli ulcer--necrotizing infection of the hand of a 445 446 plastic surgeon]. Handchir Mikrochir Plast Chir 19:230-2. 31. 447 Debacker M, Zinsou C, Aguiar J, Meyers WM, Portaels F. 2003. First case of Mycobacterium ulcerans disease (Buruli ulcer) following a human bite. Clin Infect 448 449 Dis 36:e67-8. Gao Y, Hameed HMA, Liu Y, Guo L, Fang C, Tian X, Liu Z, Wang S, Lu Z, Islam 450 32. MM, Zhang T. 2021. Ultra-short-course and intermittent TB47-containing oral 451 regimens produce stable cure against Buruli ulcer in a murine model and prevent the 452 453 emergence of resistance for *Mycobacterium ulcerans*. Acta Pharm Sin B 11:738-749. 454 33. Williamson HR, Benbow ME, Nguyen KD, Beachboard DC, Kimbirauskas RK, McIntosh MD, Quaye C, Ampadu EO, Boakye D, Merritt RW, Small PL. 2008. 455 Distribution of Mycobacterium ulcerans in Buruli ulcer endemic and non-endemic 456 457 aquatic sites in Ghana. PLoS Negl Trop Dis 2:e205. 458

459

461

462

463

464

465

466

467

468

469

470

471

472

473

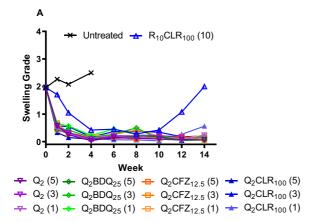
474

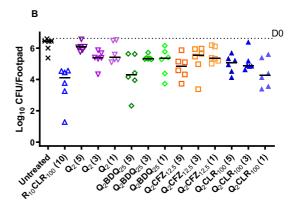
475

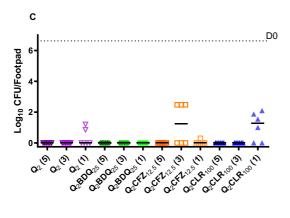
476

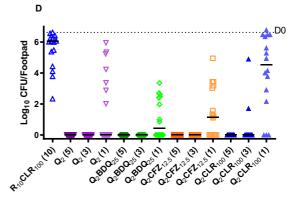
FIGURE LEGENDS Fig 1. Footpad swelling grade and microbiological outcome in BALB/c mice in response to treatment. Median swelling grade in BALB/c mice during and after treatment with the indicated regimens and number of doses in parentheses (A). Log₁₀ CFU/footpad one week after the start of treatment (B). Log₁₀ CFU/footpad six weeks after the start of treatment (C). Log₁₀ CFU/footpad 15 weeks after the start of treatment (D). R, rifampin, CLR, clarithromycin, Q, telacebec, BDQ, bedaquiline, CFZ, clofazimine. Numbers in subscript indicate the dose in mg/kg. D0, day 0 or the beginning of treatment. Fig 2. Footpad swelling grade and microbiological outcome in SCID-beige mice in response to treatment. Median swelling grade in SCID-beige mice during and after treatment with the indicated regimens and number of doses in parentheses (A). Log₁₀ CFU/footpad in SCID-beige mice in the indicated regimens during the study (B). Log10 CFU/footpad five weeks after the start of treatment (C). Log10 CFU/footpad 17 weeks after the start of treatment (D). R, rifampin, CLR, clarithromycin, Q, telacebec, BDQ, bedaquiline, CFZ, clofazimine. Numbers in subscript indicate the dose in mg/kg. D0, day 0 or the beginning of treatment.

477 Fig 1









479 Fig 2

