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1 Infective prey leads to a partial role reversal in a

2 predator-prey interaction

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13 Abstract

14	An infective prey has the potential to infect, kill and consume its predator. Such a
15	prey-predator relationship fundamentally differs from the classical Lotka-Volterra
16	predator-prey premise because the prey can directly profit from the predator as a
17	growth resource. Here we present a population dynamics model of partial role
18	reversal in the predator-prey interaction. We parametrize the model to represent the
19	predator-prey interaction of sea cucumber Apostichopus japonicus and bacterium
20	Vibrio splendidus. We observe that two major factors stabilize the predator-prey
21	interaction. First, the partial role reversal in the predator-prey community stabilizes
22	the predator-prey interaction. Second, if the predator is a generalist and follows the
23	type I functional response in attacking the prey, the predator-prey interaction is
24	stable. We also analysed the conditions for species extinction. The extinction of the
25	prey, V. splendidus, may occur when its growth rate is low, or in the absence of
26	infectivity. The extinction of the predator, A. japonicus, may follow if either the
27	infectivity of the prey is high or a moderately infective prey is abundant. We conclude
28	that partial role reversal is an underestimated subject in predator-prey studies.

29

30 Introduction

The ability of a prey to utilize the predator as a food source is referred to as a role reversal in predator-prey interaction [1-3]. The prey may become an enemy to the predator. If the role reversal is not complete the predator continues to hunt the prey while becoming vulnerable to the predation itself. In the partial role reversal the

35	3 growth of the prey population relies on the prey's normal growth rate and on the
36	additional resource acquiring by the infectivity, in particular, by its efficiency in killing
37	and converting the predator into nutrition.
38	A partial role reversal in the aquatic environment can occur in the aquaculture of sea
39	cucumbers (Apostichopus japonicus) which feeds bottom sediments inhabited by the
40	opportunistic, potentially infective bacterium Vibrio splendidus. The sea cucumber
41	belongs to the class Holothuroidea in the Phylum Echinodermata. It is a bottom
42	dwelling marine deposit feeder that uses its tentacled mouth to consume the topmost
43	sediment layer [4,5]. The sediment contains plant and animal debris, protozoa,
44	diatoms and a diverse selection of bacteria [6-10]. The sediment also hosts the
45	bacteria V. splendidus [10,11] which has been associated with seasonal epidemics of
46	high mortality among the cultured sea cucumbers [12,13]. On the other hand, V.
47	splendidus can also coexist in the gut of healthy sea cucumbers [14,15]. Because
48	bacteria generally form an important food source for the sea cucumber, A. japonicus
49	[5] can be treated as a predator to V. splendidus. The interaction is not tight in the
50	sense of traditional Lotka-Volterra predator interaction since both species can also
51	consume other resources.
52	We address the problem of partial role reversal in the predator-prey interaction by

53 presenting a predator-infective prey model to analyse the dynamics and coexistence 54 of the species in the community. After presenting the basic framework of the model 55 we parametrize the model for an opportunistic pathogenic bacteria and the 56 commercially cultivated sea cucumber, an economically important species in 57 aquaculture. The sea cucumber is appreciated as a delicacy and aphrodisiac widely in

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58	Asia. Even though the catches from the wild populations have drastically declined, the
59	production of cultured sea cucumbers in year 2014 was over 200000 tonnes in China
60	alone. [16]. According our results the species most likely coexist at a stable
61	equilibrium.
62	We also analysed the conditions for species extinctions. For the predator the
63	extinction depends on the infectivity of the prey, and its population size as well as the
64	attack rate of the predator. The possibility of recognizing the effects of an infective
65	prey within a food web is thus meaningful both scientifically and economically.
66	

67 Conceptual model description

68 The predator (S) and the prey (C) interact according to a conventional predator-prey 69 model (Fig 1). The predator population grows by consuming the prey (i). However, 70 both species also use other resources for growth ((vi) and (vii)), meaning that each of 71 them can survive as a single species population. Thus, we are dealing with a generalist 72 predator. However, in a special case the predator can be specialist. As the prey is also 73 pathogenic to the predator, a part of the predator population is infected (ii), 74 increasing the population size of infected predators (I). An infected predator can 75 recuperate (iii), die naturally (iv), or become a growth resource for the pathogenic 76 prey (v).

77

78

79 Fig 1. A schematic presentation of the predator-infective prey model.

5

80	The predator (S) and	the prey (C) interact	according to a conven	tional predator-prey
00				

- 81 model. However, the prey is pathogenic to the predator, and a part of the predator
- 82 population is infected (I). An infected predator can recover, die naturally, or become
- 83 consumed by the pathogenic prey.
- 84
- 85 A distinctive aspect in our predator-prey interaction is that both the prey and the
- 86 predator are only a part of a food web. Both species have a base growth rate that is
- 87 independent of their mutual interaction, and they both are able to grow
- 88 independently according to the respective carrying capacity of the environment ((vi)
- 89 and (vii) in Fig 1).
- 90

91 Modelling partial role reversal in predator-prey

92 interaction

Let C, S and I denote the abundances of the prey, predator and infected predator
populations, respectively. The differential equation model for the dynamics of the

95 populations are given as

96
$$\frac{dC}{dt} = r_C C \left(1 - \frac{C}{K_C}\right) - CaS + e_{IC} \mu_{inf} I$$
(1)

97

98
$$\frac{dS}{dt} = r_S S \left(1 - \frac{S}{K_S} \right) - e_{SI} \alpha C a S + (1 - \alpha) e_{CS} C a S - \mu_I S + \beta I$$
(2)

99

100
$$\frac{dI}{dt} = e_{SI}\alpha CaS - I(\mu_I + \mu_{inf} + \beta)$$
(3)

6

101

102 where the increases of the prey and predator abundances are both defined as logistic 103 growth. Parameters r_C , K_C , r_S and K_S are the growth rates and carrying capacities of 104 the prey and predator, respectively. Parameter a ($0 \le a \le 1$) denotes the attack rate of 105 the predator S. This can be interpreted either as a fraction of the feeding area grazed 106 during a time step, or it can equally be interpreted as the prey selectivity coefficient of 107 the predator. Thus, the total number of the prey harvested by the predator is aC, and 108 the harvesting is described as Type I functional response. Parameter α denotes the 109 fraction of the infective prey from the total prey population. Thus, from the predation 110 rate *CaS* fraction $(1 - \alpha)$ increases the growth rate of the healthy predator 111 population with a prey to predator growth conversion efficiency $e_{\rm CS}$. The rest of the 112 harvested prey, αCaS , infects the predators at a conversion rate e_{SI} . The infected the 113 predators end up in the infected predator population I. Parameter β denotes the 114 recovery rate of the infected predators. Parameters μ_{inf} and e_{IC} denote the predator 115 infection mortality and predator to prey conversion efficiency, respectively. Finally, μ_I denotes the natural predator mortality. 116 117 Our model follows the basic structure of the traditional predator-prey Lotka-Volterra model in that the predation is modelled as Type I functional response, and that the 118 119 healthy harvest is used for the growth of the of the predator. The model differs from 120 the Lotka-Volterra model in that both the susceptible predator and the infective prey 121 have their own logistic growth functions implying that they are generalists rather than

122 specialists. The infected predator population grows only at the expense of the

123 infections of the healthy predators. The infected predators also serve as a resource for

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124 the prey growth as they become diseased.

125 **Parametrization of the model**

- 126 Most of the parameters were chosen according to the suitable values found from the
- 127 literature. Conversion efficiencies were calculated as the ratio of dry weights of the
- 128 predator and the prey multiplied by ecological efficiency. Ecological efficiency was set
- to 0.25 for the sea cucumber, and to 0.5 for the bacteria [17,18].
- 130 Because we could not find the dry weight of V. splendidus, we used the dry weight of
- 131 E. coli [19]. The dry weight of A. japonicus is calculated according to the article by Sun
- 132 et al., where it was stated that the dry weight of A. japonicus equals 0.075×wet weight
- 133 [4]. The mean wet weight m_{Ai} was set at 150g [20].

134 Prey to predator conversion efficiency is calculated as $e_{CS} = \frac{m_{bact}}{m_{Aj}} 0.25 = \frac{0.28 \cdot 10^{-12}g}{150g \cdot 0.075}$ 135 $0.25 = 6.22 \cdot 10^{-15}$.

136 Predator to prey conversion efficiency is $e_{IC} = \frac{m_{Aj}}{m_{bact}} 0.5 = \frac{150g \cdot 0.075}{0.28 \cdot 10^{-12}g} 0.5 = 1.00 \cdot 10^{13}.$

137 According to the empirical study by Lysenkov et al. the natural population density of

138 A. japonicus is 0.14 individuals per square meter, even though the observed density

139 has fallen to 0.023 individuals per square meter because of illegal harvesting [20].

140 Therefore, the area of the feeding unit is set to $A_{KS} = \frac{1m^2}{0.14} = 7m^2$ and depth of

141 foraging to 1*cm*. We calculated the predator attack rate using the formula

142
$$a = \frac{m_{Aj}f}{\rho V} = \frac{150g \cdot 5.3 \cdot 10^{-3}g^{-1}h^{-1}mg}{1gcm^{-3} \cdot 70000cm^2 \cdot 1cm} = 1.14 \cdot 10^{-5}h^{-1} = 2.73 \cdot 10^{-4}d^{-1}.$$

143 where m_{A_i} is the wet weight of the sea cucumber and f is the amount of sediment

8

144	eaten hy the sea	cucumber per hour	ner gram of sea c	ucumber [4]	n is the density of
T-4-4	eaten by the sea	cucumber per nour	per grann or sea c	ucumber [4],	p is the density of

- 145 the sediment as given by Kennish [9], and V is the volume of the feeding unit. The
- 146 resulting attack rate is the nominal portion of the available prey eaten within a time
- 147 step. Because the actual attack rate depends also from the selectivity of the predator,
- 148 a range of attack rate values around the nominal value was used in model analysis and
- 149 numerical simulations.
- 150 We have taken the carrying capacity of the bacteria K_c from the literature [7,11]. For
- 151 the sea cucumber carrying capacity K_s we tested a range of values, but for consistency
- 152 in the shown simulation results K_s is always 10000.
- 153 The symbols and parameters used in the model and are shown in Table 1.

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- 156

157 **Table 1.** Symbols and parameter values

Parameter		Value	Unit
Susceptible predator	S	A. japonicus,	individ. [<i>i</i>]
Infected predator	Ι	A. japonicus,	individ. [<i>i</i>]
Infective prey	С	V. splendidus,	individ. [<i>i</i>]
Infective prey growth rate	r _c	0.5, 5.0, 50	[<i>i/i</i> · <i>d</i> ⁻¹]
Susceptible predator growth	r _s	0.02	$[i/i \cdot d^{-1}]$
rate			
Prey K	K _c	1.10^{13}	[<i>i</i>]

Predator K	Ks	10000	[<i>i</i>]
Predator infection mortality	μ_{inf}	0.8	[<i>i/i</i> · <i>d</i> ⁻¹
due to the prey			
Predator mortality	$\mu_{\rm I}$	0.01	[<i>i/i</i> · <i>d</i> ⁻¹]
Predator attack rate	а	1.0.10-1110.0.10-4	[<i>i/i²·d</i> -
Prey to predator conversion	e _{CS}	6.22·10 ⁻¹⁵	[<i>i</i> / <i>i</i>]
efficiency			
Predator to prey conversion	e _{IC}	1.0.1013	[<i>i</i> / <i>i</i>]
efficiency			
Infectivity of the prey	e _{SI}	10-1310-9	[<i>i</i> / <i>i</i>]
Proportion of infective prey	α	0.0011.0	[<i>i</i> / <i>i</i>]
Infected predator recovery	β	0.2	[<i>i/i</i> · <i>d</i> ⁻¹

159

160 Model analyses

161

162 **Population equilibria**

163 The equilibrium of the community is the starting point of the analysis of community

164 behaviour. The equilibrium is defined by assuming the time derivatives in the

165 population equations (1)-(3) equal to zero:

166

167
$$0 = r_C C \left(1 - \frac{c}{\kappa_c}\right) - CaS + e_{IC} \mu_{inf} I$$
(4)

168

169
$$0 = r_{S}S(1 - \frac{S}{K_{S}}) - e_{SI}\alpha CaS + (1 - \alpha)e_{CS}CaS - \mu_{I}S + \beta I$$
(5)

170

171
$$0 = e_{SI} \alpha C a S - I(\mu_I + \mu_{inf} + \beta)$$
(6)

172

173 Inserting I from eq. (6) into (4) and (5) and dividing the resulting equations by C and S,

174 respectively, we get

175
$$0 = r_{c} \left(1 - \frac{c}{\kappa_{c}} \right) - aS + e_{IC} \mu_{inf} zS$$
(7)

176
$$0 = r_S \left(1 - \frac{s}{K_S}\right) - e_{SI} \alpha C a + e_{CS} (1 - \alpha) C a - \mu_I + \beta z C$$
(8)

177 where
$$z = e_{SI}\alpha a/(\mu_I + \mu_{inf} + \beta)$$
.

178 Linear equations (7) and (8) can be presented in a matrix form

179
$$A\begin{bmatrix} C\\S\end{bmatrix} = \begin{bmatrix} r_C\\r_S - \mu_I \end{bmatrix}$$
(9)

180 where

181
$$A = \begin{bmatrix} a_{11} & a_{12} \\ a_{21} & a_{22} \end{bmatrix} = \begin{bmatrix} \frac{r_c}{K_c} & a - e_{IC}\mu_{inf}z \\ e_{SI}\alpha a - e_{CS}(1 - \alpha)a - \beta z & \frac{r_s}{K_s} \end{bmatrix}$$
(10)

182 The solution of eqs. (9) and (10) is given as

183
$$\dot{C} = \frac{1}{a_{11}a_{22} - a_{12}a_{21}} (a_{22}r_C - a_{12}(r_S - \mu_I))$$
(11)

184
$$\dot{S} = \frac{1}{a_{11}a_{22} - a_{12}a_{21}}(-a_{21}r_C + a_{11}(r_S - \mu_I))$$
 (12)

185

186 Infected predators are then calculated as
$$I = z\dot{C}\dot{S} = \dot{C}\dot{S}e_{SI}\alpha a/(\mu_I + \mu_{inf} + \beta)$$
 (eq.

187 (6)).

188 The equilibrium states of interest are the following:

	11	
189	a)	Both species coexists at a general equilibrium: \dot{C} , \dot{S} , $\dot{I} > 0$. Stability of this
190		equilibrium represent continuing coexistence of the species.
191		In the absence of species interaction the environmental carrying capacity of
192		the prey is equal to K _C and that of the predator is equal to $(r_S-\mu_I)K_S/r_S$.
193	b)	Infective prey exists but is zero: $\dot{C} = 0$, $\dot{S} > 0$, $\dot{I} = 0$. This represent an extinction
194		of the prey.
195	c)	Predator is absent: $\dot{C} > 0$, $\dot{S} = \dot{I} = 0$. This represents an extinction of the
196		predator.

197 Analytical results

The analysis of the model presents us three possible outcomes. Either the prey or the
predator drives the other to extinction, or both populations coexist in a stable
equilibrium.

201 Numerical simulations

202 The numerical simulations of the model (1)-(3) were performed out using Matlab

203 R2020b. Numerical simulations were in accordance with the analytical results (Section

204 "Population equilibria"). Both the prey and the predator were able to drive the other

205 to extinction. All simulation results with positive coexisting populations were locally

206 stable. The parameters used in the simulations are described in Section

207 "Parametrization of the model". Because simulations exemplify the partial role

- 208 reversal between A. japonicus and V. splendidus, respective conversion efficiencies
- 209 were used throughout, as well as the high mortality rates shown to be associated with
- 210 the infection [12,13]. The effects of infectivity of the prey e_{SI} , proportion of the

12

211 infective prey α , and the attack rate α were tested using wide parameter ranges.

212 Though the outbreaks caused by V. splendidus have been associated with high

213 mortality rates, we tested the model also with low infection mortality rates and high

- 214 recovery rates. Even when infection mortality μ_{inf} and infected predator recovery β
- were 0.3 and 0.7, respectively, the results remained qualitatively same. Initial
- 216 population sizes did not affect the results, and the model gives consistent results.

217 **Results**

218 For an opportunist prey with a high environmental growth rate the level of infectivity,

219 e_{sl,} is not crucial (Fig 2). The prey population size will settle around the level of

220 carrying capacity K_c. A low infectivity e_{sl} combined with a high environmental growth

rate r_c of the prey can be beneficial also for the predator because the predator is able

to sustain population levels above the environmental carrying capacity $(r_S - \mu_I)K_S/$

223 r_s . Rising the level of infectivity, however, decreases the predator population. To the

224 contrary, the population size of a prey with slow environmental growth depends on

the level of infectivity. Low infectivity leads to the extinction of the slowly growing

226 prey.

227

228 Fig 2. Infectivity affects both prey and predator population sizes.

For low infectivity e_{sl} a higher outside prey growth rate r_c supports larger prey population than a lower growth rate, but this is reversed if e_{sl} increases enough. After the turning point (o), where high and low growth rates of the prey provide equal population sizes, an increase in infectivity e_{sl} of the prey results in greater prey and

13

233	13 lesser predator population sizes. If the infectivity is increased even more, the trend of
234	the prey population turns into decreasing. At the extinction of the prey (at low
235	infectivity values and low prey growth rate) the predator population size settles
236	down at its environmental carrying capacity $\frac{(r_S - \mu_I)K_S}{r_S} = 5000$. Subfigures A and B
237	show the full scale of the population sizes, whereas subfigure C displays a closer view
238	to the predator population at the turning point (o). Red, purple and blue lines are fast
239	(r_c =50), medium (r_c =5) and slow (r_c =0.5) growth rates. The predator's attack rate
240	$a=3.0\cdot10^{-4}$ and the infective proportion of the prey $\alpha=0.001$. Infectivity e_{s_1} ranges from
241	10 ⁻¹³ to 10 ⁻⁹ .
242	
243	High infectivity e _{st} increases the population size of a slowly growing prey because
244	increasing infectivity allows the prey to reach a higher prey population size as
245	compared to a prey with a higher growth rate r_c (Fig 2). However, due to the high
246	mortality μ_{inf} associated with the infection, a too high level of infectivity causes the

247 extinction of the predator and a decline in the prey population size. Likewise, in the

case of fast growing prey, very high infectivity leads the prey population size to settle

249 at the environmental carrying capacity.

250

251 Fig 2 also illustrates the presence of a turning point such that the order of the

252 population sizes will change with the change of a parameter. When $e_{SI} = 1.22e-10$,

253 making $a_{12} = 0$ (eq. 8), the equilibrium population size of the prey equals to its

environmental carrying capacity $\dot{C} = K_C$. At the same value of infectivity the

equilibrium population size of the predator will be $\dot{S} = 4865$. Below the turning point

	14
256	slow prey growth rates supports lower prey population sizes than higher growth rates.
257	When the parameter e_{s_l} passes the turning point then the order of the population
258	sizes is reversed. The effect of the turning point to the population sizes of the
259	predator is opposite. Note that the turning point is not a uniquely defined concept but
260	always related a chosen parameter. This is because the condition $a_{12} = 0$ can become
261	true for choosing appropriate values for e_{SI} , $lpha$, e_{IC} and μ_{inf} . A comparable analysis can
262	be carried out for the solutions with $a_{21} = 0$.
263	
264	Infectivity e_{s_I} and the proportion of infective prey in the total prey population α have
265	parallel but not completely interchangeable effects on the population sizes of the prey
266	and predator (Fig 3). If the prey is very weakly infective (e_{si} =10 ⁻¹³ , Figs 3A,B), the
267	predator will survive any proportion of the infective prey, and can even completely
268	eradicate a slow growing prey. In contrast, if the infectivity is high (e_{SI} =10 ⁻¹¹ , Figs
269	3C,D), then the predator will become extinct even at relatively low infective prey
270	densities. This happens regardless of the prey growth rate.
271	
272	Fig 3. Proportion of infective prey $lpha$ affects the population in the same way as
273	infectivity <i>e</i> _{si} .
274	Low infectivity of prey, e _{si} , supports lower prey population sizes (A) and higher
275	predator population sizes (B) than high infectivity (C and D, respectively). Slowly
276	growing prey with low infectivity can proliferate only if the majority of the prey are

277 infective (A). Even high proportions of infective prey cause only a slight decrease in

278 predator population (B). High infectivity e_{SI} prevents the extinction of the prey. Highly

279	15 infective prey thrives best when it forms relatively small part of the prey population
280	(C) because the predator becomes extinct if the majority of the prey are infective
281	$(lpha\gtrsim 0.4)~$ (D). The whole range of final population sizes in subfigures A and B fit
282	within the dotted lines in subfigures C and D, respectively. Red, purple and blue lines
283	are fast (r_c =50), medium (r_c =5) and slow (r_c =0.5) growth rates. Infectivity values are
284	e_{SI} =10 ⁻¹³ in subfigures A and B, and e_{SI} =10 ⁻¹¹ in subfigures C and D.
285	
286	If attack rate approaches zero both the prey and the predator population sizes tend
287	towards the environmental carrying capacity regardless of the infectivity (Fig 4).
288	Increasing attack rate may have different effects on the prey and predator sizes. When
289	the value of the infectivity remains low an increase in the attack rate benefits the
290	predator (Fig 4B). High growth rate of the prey results in larger predator population
291	than low growth rate. If the value of the infectivity is increased slightly (moderate
292	infectivity) an increment in growth rate decreases predator population sizes (Fig 4D).
293	In both cases increasing attack rate decreases the prey population size (Figs 4A,C).
294	
295	Fig 4. An increase in the infectivity may reverse the effect of predator attack rate on
296	the predator population size.
297	In subfigures A and B the prey's infectivity e_{st} is weak. Increasing the predator attack
298	rate a decreases prey and increases predator population sizes. In contrast, e_{sl} in
299	subfigures C and D is slightly greater, and increasing attack rate decreases the

300 population levels of the prey as well as of the predator. As the attack rate decreases,

301 the population sizes approach their respective environmental carrying capacities. Red,

16

302 purple and blue lines are fast (r_c =50), medium (r_c =5) and slow (r_c =0.5) growth rates.

303 The infectivity values are $e_{sl}=10^{-13}$ in subfigures A and B, and $e_{sl}=10^{-10}$ in subfigures C

304 and D.

305 Extinction of the species

306 We consider here the possibility of extinction of the predator or the prey. The

307 questions of interest are: 1) Under which conditions the predator can drive the prey to

- 308 extinction such that the species community would approach lie at a "predator only"
- 309 equilibrium $\dot{C} = 0, \dot{S} > 0, \dot{I} = 0.2$) Alternatively, we ask under what conditions the prey
- 310 can eradicate the predator such that the species community would ultimately lie the
- 311 "prey only" equilibrium $\dot{C} > 0, \dot{S} = \dot{I} = 0$.

312 Consider first the "predator only " equilibrium $\hat{C} = 0, \hat{S} = \frac{r_s - \mu_l}{r_s} K_s, \hat{I} = 0$, that is, the

313 sea cucumber lies at its carrying capacity and *V. splendidus* has been driven to

- 314 extinction. If the equilibrium is locally stable then the extinction of *V. splendidus* is
- 315 expected to occur. The local stability of the linearized dynamics at the equilibrium can
- 316 be analysed studying the properties of the following Jacobian matrix

$$317 \quad J = \begin{bmatrix} J_{11} & J_{12} & J_{13} \\ J_{21} & J_{22} & J_{23} \\ J_{31} & J_{32} & J_{33} \end{bmatrix} = \begin{bmatrix} r_C - \frac{a(r_S - \mu_I)}{r_S} K & 0 & e_{IC}\mu_{inf} \\ \frac{r_S - \mu_I}{r_S} K [e_{CS}(1 - \alpha) - e_{SI}\alpha] & -r_S + \mu_I & \beta \\ ae_{SI}\alpha \frac{r_S - \mu_I}{r_S} K_S & 0 & -(\mu_I + \mu_{inf} + \beta) \end{bmatrix}$$

318

319 Recall that if the real parts of the eigenvalues of J are all negative the system is locally 320 stable. It can be shown that the first eigenvalue is $\lambda_1 = -r_S + \mu_I$ which we assume to 321 be negative. The remaining two eigenvalues depend on the submatrix where line 2

17

322 and column 2 are deleted in matrix J. The eigenvalues λ_2 , λ_3 both have negative real

323 pars if and only if [21]

324
$$J_{11} + J_{33} = \left(r_C - a \frac{r_S - \mu_I}{r_S} K_S\right) - (\mu_I + \mu_{inf} + \beta) < 0$$

325 and

326
$$J_{11}J_{33} - J_{13}J_{31} = -\left(r_C - a\frac{r_S - \mu_I}{r_S}K_S\right)(\mu_I + \mu_{inf} + \beta) - e_{IC}\mu_{inf}ae_{SI}a\frac{r_S - \mu_I}{r_S}K_S > 0$$

In this case extinction occurs. For example, if the proportion of infective prey is low $(\alpha \approx 0)$ and the growth rate is low $(r_C < a \frac{r_S - \mu_I}{r_S} K_S)$ then the both conditions become true and the predator will eradicate the prey. High proportion of infective prey, high energetic efficiency and high carrying capacity may protect the prey from extinction. 331

332 The extinction of the prey depends crucially also on the attack rate of the predator

333 (Fig 5). There is a threshold value or a minimum attack rate at which the predator can

334 cause the extinction of the prey.

335 Fig 5. Predator attack rate affects the extinction of prey.

336 Extinction of prey is possible if the predator's attack rate is higher than the threshold

337 defined by the prey's growth rate. Fast growing prey survives higher predator attack

338 rate than slow growing. If the prey's infectivity e_{si} is high, only a small fraction α of the

339 prey population needs to be infective to escape the extinction. Prey growth rate

 $340 r_c=10.$

341

- 342 If the prey is a specialist such that it consumes only the predator ($r_{c}=0$), then the
- 343 extinction can be due to low infectivity or insufficient infective population. Yet, even a

18

344 low prey growth rate may keep the prey population alive, if the prey is infective

345 enough. Because infective prey can survive when its outside growth rate $r_c=0$, a

346 positive growth rate guarantees the survival also in the case of relatively low

- 347 infectivity, if the infective prey forms large part of the predator's diet.
- 348
- 349 Extinction of the predator leads to the "prey only" equilibrium $\dot{C} = K_C \dot{S} = I = 0$. The

350 linearized dynamics of the predator-prey interaction at the equilibrium can be

351 presented as

352
$$J = \begin{bmatrix} J_{11} & J_{12} & J_{13} \\ J_{21} & J_{22} & J_{23} \\ J_{31} & J_{32} & J_{33} \end{bmatrix} = \begin{bmatrix} -r_c & -aK_c & e_{IC}\mu_{inf} \\ 0 & r_s - K_c a(e_{SI}\alpha - e_{CS}(1-\alpha)) - \mu_I & \beta \\ 0 & ae_{SI}\alpha K_c & -(\mu_I + \mu_{inf} + \beta) \end{bmatrix}$$

353

354 The first eigenvalue of the Jacobian matrix J is $\lambda_1 = -r_c$. The remaining two

355 eigenvalues depend on the submatrix with line 1 and column 1 deleted in the Jacobian

356 matrix J. The real parts of the eigenvalues λ_2 , λ_3 are both negative if and only if [21]

357
$$J_{22} + J_{33} = r_S - K_C a (e_{SI} \alpha - e_{CS} (1 - \alpha)) - \mu_I - (\mu_I + \mu_{inf} + \beta) < 0$$

358 and

359
$$J_{22}J_{33} - J_{23}J_{32} = -[r_S - K_C a(e_{SI}\alpha - e_{CS}(1 - \alpha)) - \mu_I](\mu_I + \mu_{inf} + \beta) - \beta a e_{SI} \alpha K_C > 0$$

360 The proportion of the infective prey α is crucial. Assume that α =0. Then $J_{22}J_{33} - J_{23}$

361
$$J_{32} = -[r_S + K_C a - \mu_I](\mu_I + \mu_{inf} + \beta) < 0$$
 indicating that the predator does not become

362 extinct. Assume next that $\alpha = 1$. Then $J_{22} + J_{33} < 0$ if

363
$$r_S - \mu_I - (\mu_I + \mu_{inf} + \beta) < K_C a e_{SI}$$

364 and
$$J_{22}J_{33} - J_{23}J_{32} > 0$$
 if $-K_C ae_{SI}(\mu_I + \mu_{inf}) - (r_S - \mu_I)(\mu_I + \mu_{inf} + \beta) > 0$,

365 indicating that the predator will become extinct.

19

366

367	Under these conditions the predator becomes extinct. Fig 6 presents how the change
368	in infectivity \mathbf{e}_{SI} together with the different infective prey proportions affect the
369	extinction of predator. If the prey's growth rate r_c is at all positive, the prey can drive
370	the predator to extinction. Interestingly, if $r_c>0$ the level of growth rate does not
371	affect the results.
372	
373	Fig 6. Extinction of predator depends on the volume of infective prey consumption.
374	If the infective prey forms a large fraction of the available prey population, lower prey
375	infectivity \mathbf{e}_{SI} and predator attack rate are needed to eradicate the predator. To the
376	contrary, if the infective fraction α is small, or the infectivity e_{SI} is low, the predator
377	prevails. Using the parameters for A. japonicus and V. splendidus the predator survives
378	always if α <0.011. At very low attack rate or infectivity the predator does not consume
379	enough of the infective prey to suffer extinction. In the case of a specialist prey, when
380	r_c =0, it can not drive the predator to extinction. However, as long as r_c >0, then the
381	extinction does not depend on the rate of growth.
382	

383 **Discussion and conclusions**

We have presented a new predator-prey model with partial role reversal where the predator can become a target of attacks by the prey such that the prey can use the predator as a resource for growth. We parametrized the model using sea cucumber *A*. *japonicus* and a bacterium species *V. splendidus* as a model system.

388	20 A distinctive feature in our model is that both the prey and the predator are only a
389	part of a food web. Both species have an environmental growth rate that is
390	independent of their mutual interaction, and they both are able individually to grow
391	to their respective carrying capacity. Thus, both the prey and the predator are
392	generalists. The predator can benefit from the infective prey through eating the
393	increasing prey population. The final size of the predator population depends on the
394	proportion of infective prey in its diet. A small proportion of very infective prey
395	functions similarly with a large proportion of less infective prey.
396	Overall, the partial role reversal in the predator prey community stabilizes the
397	predator-prey interaction. The same effect is observed if we add logistic growth to
398	both species in the Lotka-Volterra equation. We did not observe in our extensive
399	simulations any signs of instability in the coexistence solutions
400	We also analysed the conditions for species extinction. A generalist predator becomes
401	eradicated if some part of the prey population is infective, the infection can cause
402	mortality, and the abundance of the infective prey is high. For a specialist predator the
403	extinction depends on the infectivity of the prey, and its population size as well as the
404	attack rate of the predator. The extinction of the prey is possible if its infectivity is low
405	and either it the infective prey forms only a small fraction of total prey population or

406 the growth rate is low.

The infective prey-predator model departs from a predator-non infective prey model
by quantifying the infectivity of the prey and the mortality of the infection. In
principle, this resembles a fatal infectious disease. However, the predator is also able
to consume the prey regardless of its pathogenicity, and can therefore benefit from

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411	the growing pathogenic prey population. It is noteworthy that infection does not need
412	to be a bacterial infection. Any similar situation can play the part of a disease in the
413	model framework. The examples include a shoal of young pikes can that attract a
414	growing number of sticklebacks [3]. In these examples, the infectivity is taken as a
415	number that describes the ability of the prey to find potential victims among the
416	predators and to attract the rest of the prey population to the site. The infection
417	mortality describes the probability of death of an infected or attacked predator.
418	The system consisting of a predator and an infective prey remains mostly an
419	unresearched subject. The population model presented here describes the process of
420	role reversal using several parameters. However, of these parameters only a few were
421	in the model markedly involved in the role reversal process. These include the prey to
422	predator and predator to prey conversion efficiencies, and the infectivity of the prey.
423	This implies that it would be possible to address the subject empirically by studying a
424	suitable pair of model organisms.
425	Aquaculture provides many opportunities to find both scientifically and economically
426	interesting targets for basic and applies research. Also, agriculture can be considered
427	as a field that would benefit from the research.
428	For the purposes of enhancing sea cucumber cultivation, factorial experiments
429	manipulating the growth conditions of the non-pathogenic vs. pathogenic bacteria as
430	food could be set up. One avenue on this would be for example a biological control
431	[22] of Vibrio using specific lytic bacteriophages continuously of periodically added to
432	the culturing ponds. Bacteriophages are commonly species specific and can be mass-
433	produced in bioreactors. In practise, Vibrio phages could be isolated from raw water

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- 434 samples by filtering out the bacteria and adding the filtrate on *Vibrio* pure culture to
- 435 amplify only *Vibrio*-specific phages.

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438 **References**

439 1. Barkai A, McQuaid C. Predator-prey role reversal in a marine benthic

440 ecosystem. Science 1988; 242(4875): 62-64.

- 441
 2. Sánchez-Garduño F, Miramontes P, Marquez-Lago T. Role reversal in a
 442 predator-prev interaction. R Soc Open Sci 2014;1:140186.
- 3. Nilsson J, Flink H, Tibblin P. Predator–prey role reversal may impair the
- recovery of declining pike populations. Journal of Animal Ecology 2019; 88(6): 927-939.
- 446 4. Sun J, Zhang L, Pan Y, Lin C, Wang F, Kan R, et al. Feeding behavior and
- 447 digestive physiology in sea cucumber *Apostichopus japonicus*. Physiology &
 448 Behavior 2015; 139: 336-343.
- 5. Xu Q, Hamel J-F, Mercier A. Chapter 10 Feeding, digestion, nutritional
 physiology, and bioenergetics. Developments in Aquaculture and Fisheries
 Science 2015; 39: 153-175.
- 452 6. Bratbak G. Bacterial biovolume and biomass estimations. Applied and
 453 Environmental Microbiology 1985; 49(6),:1488-1493.
- 454 7. Watson SW, Novitsky TJ, Quinby HL, Valois FW. Determination of bacterial

	23		number and biomass in the maxime environment. Applied and Environmental
455			number and biomass in the marine environment. Applied and Environmental
456			Microbiology 1977; 33(4): 940-946.
457	8	3.	Liu X, Zhou Y, Yang H, Ru S. Eelgrass detritus as a food source for the sea
458			cucumber Apostichopus japonicus Selenka (Echinodermata: Holothuroidea) in
459			coastal waters of north China: An experimental study in flow-through systems.
460			PLOS ONE 2013; 8(3): e58293
461	Q	9.	Kennish MJ. Encyclopedia of estuaries. 2016, Springer Netherlands, ISBN 978-
462			94-017-8800-7.
463	1	10.	Xu H, Wang L, Bao X, Jiang N, Yang X, Hao Z, et al. Microbial communities
464			in sea cucumber (Apostichopus japonicus) culture pond and the effects of
465			environmental factors. Aquaculture Research 2019; 50: 1257-1268.
466	1	11.	Vezzulli L, Pezzati E, Stauder M, Stagnaro L, Venier P, Pruzzo, C. Aquatic
467			ecology of the oyster pathogens Vibrio splendidus and Vibrio aestuarianus.
468			Environmental Microbiology 2015; 17(4): 1065-1080.
469	1	12.	Deng H, He C, Zhou Z, Liu C, Tan K, Wang N, et al. Isolation and
470			pathogenicity of pathogens from skin ulceration disease and viscera ejection
471			syndrome of the sea cucumber <i>Apostichopus japonicas</i> . Aquaculture 2009; 287
472			(1-2): 18-27.
473	1	13.	Liu N, Zhang S, Zhang W, Li C. Vibrio sp. 33 a potential antagonist of Vibrio
474			splendidus pathogenic to sea cucumber (Apostichopus japonicus). Aquaculture
475			2017; 470: 68-73.
476	1	14.	Zhang Z, Lv Z, Zhang W, Shao Y, Zhao X, Guo M, et al. Comparative analysis
477			of midgut bacterial community under Vibrio Splendidus infection in
478			Apostichopus japonicus with hindgut as a reference. Aquaculture 2019a; 513:

479	24	734427.
480	15	. Zhang Z. Zhang W. Hu Z. Li C. Shao Y. Zhao X. et al. Environmental factors
481	10	promote pathogen-induced skin ulceration syndrome outbreak by readjusting
482		the hindgut microbiome of <i>Apostichopus japonicus</i> . Aquaculture 2019 b; 507:
483		155-163.
484	16	. Han Q, Keesing JK, Liu D. A Review of sea cucumber cquaculture, ranching,
485		and stock enhancement in China. Reviews in Fisheries Science & Aquaculture
486		2016; 24(4): 326-341.
487	17	. Yang H, Yuan X, Zhou Y, Mao Y, Zhang T, Liu Y. Effects of body size and
488		water temperature on food consumption and growth in the sea cucumber
489		Apostichopus japonicus (Selenka) with special reference to aestivation.
490		Aquaculture Research 2005; 36: 1085-1092.
491	18	8. Yingst JY. The utilization of organic matter in shallow marine sediments by an
492		epibentic deposit-feeding holothurian J Exp Mar Biol Ecol 1976; 23: 55-69.
493	19	. Neidhardt FC, Curtiss IR, Ingraham JL, Lin, ECC, Low KB, Magasanic B, et
494		al., editors. Eschericia coli and salmonella: Cellular and molecular biology. 2^{nd}
495		ed. Washington D.C.: ASM Press; 1996.
496	20	. Lysenko VN, Zharikov VV, Lebedev AM. The current status of populations of
497		the sea cucumber Apostichopus japonicus (Selenka, 1867) in the Far Eastern
498		marine reserve. Russian Journal of Marine Biology 2018; 44(2): 164-171.
499	21	. Edelstein-Keshet, L. Mathematical models in biology. 1^{st} ed, Random House,
500		NY, 1988.
501	22	. Merikanto I, Laakso JT, Kaitala V. Outside-host phage therapy as a biological

control against environmental infectious diseases. Theoretical Biology and

503 Medical Modelling 2018; 15(7).

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