

1 **Genes encoding teleost orthologs of human haplo-insufficient**
2 **and monoallelic genes remain in duplicate more frequently than**
3 **the whole genome**

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26 **Keywords**

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28 singleton

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31 Abstract

32 Gene dosage is important is an important issue both in cell and evolutionary biology. Most genes
33 are present in two copies in eukaryotic cells. The first outstanding exception is monoallelic gene
34 expression (MA) that concerns genes localized on the X chromosome or in regions undergoing
35 parental imprinting in eutherians, and many other genes scattered throughout the genome. The
36 second exception concerns haploinsufficiency (HI), responsible for the fact that a single func-
37 tional copy of a gene in a diploid organism is insufficient to ensure a normal biological function.
38 One of the most important mechanisms ensuring functional innovation during evolution is Whole
39 genome duplication (WGD). In addition to the two WGDs that have occurred in vertebrate ge-
40 nomes, the teleost genomes underwent an additional WGD, after their divergence from tetrapod.
41 In the present work, we have studied on 57 teleost species whether the orthologs of human MA
42 or HI genes remain more frequently in duplicates or returned more frequently in singleton than
43 the rest of the genome. Our results show that the teleost orthologs of HI human genes remained
44 more frequently in duplicate than the rest of the genome in all the teleost species studied. No
45 signal was observed for the orthologs of genes localized on the human X chromosome or sub-
46 jected to parental imprinting. Surprisingly, the teleost orthologs of the other human MA genes
47 remained in duplicate more frequently than the rest of the genome for most teleost species. These
48 results suggest that the teleost orthologs of MA and HI human genes also undergo selective pres-
49 sures either related to absolute protein amounts and/or of dosage balance issues. However, these
50 constraints seem to be different for MA genes in teleost in comparison with human genomes.

51 Introduction

52 Gene dosage effects are an important phenomenon in cell biology that has evolutionary conse-
53 quences. Indeed, in eukaryote cells, most genes are present in two copies that are transcribed and
54 produce functional proteins. However, there are exceptions. The first outstanding exception is
55 the case of monoallelic gene expression (MA). This is so for the majority of genes that are pre-
56 sent on the X chromosome of eutherian mammals, genes that present a parental imprinting in
57 eutherians, and genes encoding immunoglobulins and olfactory receptors (Chess et al., 2016).
58 Monoallelic expression of genes is under an epigenetic control that is not well understood. For
59 these genes, dysregulation of the mechanism(s) underlying monoallelic expression can lead to
60 expression of both alleles, and to overexpression of the corresponding protein, and thus to severe
61 pathologies (Horsthemke 2010). The second exception concerns haploinsufficiency. Haploinsuf-
62 ficiency is a biological phenomenon responsible for the fact that a single functional copy of a
63 gene in a diploid organism is insufficient to ensure a normal biological function. Haploinsuffi-
64 ciency is detected more frequently in essential genes than in nonessential genes in yeast (Ohnuki
65 & Ohya, 2018). Two non-mutually exclusive theories have been proposed to explain the cause of
66 haploinsufficiency: the “insufficient amounts” hypothesis and the gene dosage balance hypothe-
67 sis (GDBH). The “insufficient amounts” hypothesis states that haploinsufficiency is the conse-
68 quence of a reduced protein amount due to the loss of function of one allele, this amount being
69 insufficient to ensure its biological function (Deutschbauer et al., 2005). This hypothesis does not
70 explain why haploinsufficiency persisted over evolutionary time. The GDBH suggests that the

71 phenotype caused by changes of protein level in a biological process is due to stoichiometric
72 imbalances of protein complexes involved in cellular functions (Veitia, 2002; Papp et al., 2003).
73 This hypothesis predicts that haplo-insufficient genes confer a biological defect when the amount
74 of proteins is halved (such as A in a complex A-B-A) but also in excess in particular cases (such
75 as B in the same complex (Veitia, 2002). In contrast to the “insufficient amounts” hypothesis,
76 this hypothesis proposes an elegant explanation of the conservation of haploinsufficiency during
77 evolution.

78
79 One of the most important mechanism ensuring functional innovation during evolution is gene
80 duplication or the duplication of entire genome (Ohno et al., 1964; Hideki & Kondrashov, 2010).
81 Whole genome duplication (WGD) events have been observed in all taxonomic groups: bacteria
82 (Kuroda et al, 2001), unicellular eukaryotes (Manolis et al., 2004) and in plants (Adams et al,
83 2005). In vertebrates, there have been two rounds of duplication of the ancestral deuterostome
84 genome (Mable et al., 2011). One of the striking features that characterize the teleost genomes is
85 that they underwent an additional WGD, also called the teleost-specific genome duplication
86 (TGD), after divergence from tetrapods (Glasauer & Neuhauss, 2014). This specific WGD event
87 provided important additional genetic material, which strongly contributed to the radiation of
88 teleost fishes (Ravi et al., 2008). Teleosts constitute a monophyletic group of ray finned fishes,
89 and is the widest and most diverse group of vertebrates (Robinson-Rechavi et al., 2001; Taylor et
90 al., 2003; Taylor & Raes, 2014; Christoffels et al., 2004). The high diversity of fish species com-
91 bined with a recent complete duplication makes Clupeocephala a group of great interest for the
92 study of complete genome duplication in the animal kingdom.

93
94 Unlike single-gene duplication events, a WGD provides all at once a large number of new genet-
95 ic material, promoting an increased inter- and intra-specific diversity (Van de Peer et al, 2009,
96 2017). Interestingly, after WGD, all genes do not remain in duplicate with the same probability.
97 Most models predict a rapid return of part of the duplicates to a singleton state (Maere et al,
98 2005), the extra-copies being rapidly pseudogenized (Sankoff et al, 2010). In particular for the
99 rainbow trout, whose genome has duplicated one more time than that of the teleost about 100 my
100 ago, it is estimated that about 48% of the genome remained in duplicate, when the remaining 52%
101 of the genome quickly returned to a singleton state (Berthelot et al, 2014).

102 Understanding the rules explaining why certain genes remain in duplicate when others return to
103 singleton is a challenging issue. It has been shown that certain families of genes are more likely
104 to remain as duplicates in all taxonomic groups studied. This is the case for transcription factors,
105 protein kinases, enzymes and transporters (Conant et al., 2008). Recently, we showed that this is
106 also the case for genes encoding membrane receptors and their ligands (Grandchamp et al.,
107 2019). The first explanation that has been put forward to explain the fact that genes are more
108 often kept in duplicate is that these molecules are involved in key functions common to all or-
109 ganisms. Their quantitative increase would favor these key functions because of an increase in
110 the number of molecules produced (selection for an absolute dosage increase), and/or because of
111 a compensation of a potential loss of function mutation of one of both copies. Another explana-
112 tion is based on the respect of gene dosage balance. This is particularly so for proteins whose
113 function is heavily dependent on interactions with partners.

114
115 In the present work, we have studied on 57 teleost species whether the orthologs of human genes

116 known to present a monoallelic (MA) expression or to be haplo-insufficient (HI) in human re-
117 main more frequently in duplicates or returned more frequently in singleton than the whole ge-
118 nome in fish species or not.

119 Results and Discussion

120 There is a mean number of 13882 human genes on 22836 (60.8%) that possess at least one
121 ortholog in at least one teleost genome. Among them, an average of 9854 (range from 3530 to
122 10868) have returned in singleton, an average of 3135 (range from 2323 to 7066) remained in
123 duplicate, and an average of 893 (range from 337 to 4772) that are in triplicate or more copies.

124 Concerning the 312 human HI genes, 299 (95.8%) possessed at least one ortholog in at least one
125 teleost genome. Among them, an average of 172 (range from 47 to 199 depending on the studied
126 species) have returned in singleton, an average of 85 (range from 68 to 122 depending on the
127 species) remained in duplicate, and an average of 19 (range from 3 to 140) that are in triplicate or
128 more copies. A total of 285 genes remained in duplicate (or more) in at least one species among
129 the 57 teleost species studied. In comparison with the whole genome, this higher percentage of
130 genes returned to singleton and remained in duplicate or more is significantly different for 55
131 species out of 57 (Chi Square analysis, p-value range from 0.058 to 4.2E-6), and for the 57 spe-
132 cies studied (according to a hypergeometric test, p-value range from 0.034 to 8.5E-6). Moreover,
133 in comparison with the whole genome as well, the higher percentage of genes that are in tripli-
134 cate or more copies is significantly higher in the genomes of Rainbow trout, Brown trout, Atlan-
135 tic salmon, Huchen, and Common Carp (p-value range from 1.3E-8 to 8.1E-4) but not in the ge-
136 nome of the other teleosts. These results suggest that the teleost orthologs of HI human genes are
137 also subjected to selective pressures either related to absolute protein amounts and/or of dosage
138 balance issues. This suggests that HI genes in humans undergo similar constraints in in teleosts.

139 Among the 285 genes that remained in duplicate in at least one teleost species, 76 genes re-
140 mained in duplicate or more in at least 80% (45) of species. These genes encode more (from 3 to
141 38 more times) transcription factors than the whole genome: bHLH transcription factor binding
142 (GO:0043425); RNA polymerase II activating transcription factor binding (GO:0001102); acti-
143 vating transcription factor binding (GO:0033613); transcription factor binding (GO:0008134);
144 DNA-binding transcription factor binding (GO:0140297); DNA-binding transcription factor ac-
145 tivity, RNA polymerase II-specific (GO:0000981); DNA-binding transcription factor activity
146 (GO:0003700). This enrichment of GO is completely in accordance with the GO of HI genes
147 previously reported (Veitia, 2002). There was no particularly representative GO among the genes
148 in triplicate in the genome of teleost species. These results are compatible with both direct insuf-
149 ficiency of a transcription factor as well as with balance issues (as they are often multi-subunit
150 complexes). Threshold effects can also be at play because of the strongly nonlinear relationships
151 (sigmoidal or S-shaped) produced by the cooperative binding of a transcription factor to a cis
152 regulatory sequence and the transcriptional response. Thus, depending on the concentration of
153 transcription factor a halved dosage may not be sufficient to cross the threshold required for a
154 normal transcriptional response (Veitia, 2002).

155 Concerning the 206 X human chromosome genes, 176 (82,6%) possessed at least one ortholog in
156 at least one teleost genome. Among them, an average of 116 (range from 32 to 132 depending on
157 the studied species) have returned in singleton, an average of 35 (range from 23 to 79 depending

158 on the species) remained in duplicate, and an average of 7 (range from 0 to 54) that are in tripli-
159 cate or more copies. Concerning the 90 genetic imprinting genes, 51 (56,7%) possessed at least
160 one ortholog in at least one teleost genome. Among them, an average of 35 (range from 12 to 41
161 depending on the studied species) have returned in singleton, an average of 8 (range from 3 to 23
162 depending on the species) remained in duplicate, and an average of 3 (range from 0 to 20) that
163 are in triplicate or more copies. So the teleost orthologs of human genes subjected to genetic im-
164 printing or located on X human chromosome returned to singleton or remained in duplicate (or
165 remain present as triplicates or more copies), in the same proportions than the whole genome.

166
167 Concerning the 580 human MA genes that are not localized on the X chromosome and that are
168 not subjected to parental imprinting, 469 (80,9%) had at least one ortholog in at least one teleost
169 genome. Among them, an average of 265 (range from 87 to 296) have returned to singleton, an
170 average of 118 (range from 87 to 193) remained in duplicate, and an average of 26 (range from 4
171 to 160) that were found in triplicate or more copies. A total of 437 genes remained in duplicate in
172 at least one species among the 57 teleost species studied. In comparison with the whole genome,
173 the difference of percentage of genes remained in duplicate or more is significantly higher for 47
174 species on 57 (Chi Square analysis, p-value range from 0.055 to 6.5E-4), and for 50 species on
175 57 (hypergeometric test, p-value range from 0.044 to 6.2E-4). Moreover, in comparison with the
176 whole genome as well, the difference of percentage of genes that are in triplicate or more copies
177 is significantly higher in the genomes of Rainbow trout, Brown trout, Atlantic salmon, Huchen,
178 and Common Carp (p-value range from 0,056 to 5.3E-3), not in the genome of the other teleosts.
179 We found this result surprising. Indeed, one would have hypothesized that the teleost orthologs
180 of MA human genes returned more frequently to singleton than the whole genome. This suggests
181 that the phenomenon that the regulation -of epigenetic mechanism- of monoallelic expression is
182 not likely to occur for these genes in teleosts. Moreover, this suggests that the constraints to ex-
183 press only one allele in the human does not exist for these genes in teleost. Unlike the HI genes,
184 there is no particularly representative GO among the MA genes.

185
186

187 Material and methods

188 We studied 57 species of fish:

189 Amazon molly (*Poecilia formosa*), Atlantic herring (*Clupea harengus*), Atlantic salmon (*Salmo*
190 *salar*), Ballan wrasse (*Labrus bergylta*), Barramundi perch (*Lates calcarifer*), Blue tilapia (*Oreo-*
191 *chromis aureus*), Blunt-snouted clingfish (*Gouania willdenowi*), Brown trout (*Salmo trutta*), Bur-
192 ton's mouthbrooder (*Haplochromis burtoni*), Channel bull blenny (*Cottoperca gobio*), Channel
193 catfish (*Ictalurus punctatus*), Climbing perch (*Anabas testudineus*) Cod (*Gadus morhua*), Com-
194 mon carp (*Cyprinus carpio common_carp_genome*), Denticle herring (*Denticeps clupeoides*),
195 Eastern happy (*Astatotilapia calliptera*), Electric eel (*Electrophorus electricus*), European seabass
196 (*Dicentrarchus labrax*), Fugu (*Takifugu rubripes*), Gilthead seabream (*Sparus aurata*), Greater
197 amberjack (*Seriola dumerili*), Guppy (*Poecilia reticulata*), Huchen (*Hucho hucho*), Indian glassy
198 fish (*Parambassis ranga*), Indian medaka (*Oryzias melastigma*), Japanese medaka HdrR (*Oryzias*
199 *latipes* ASM223467v1), Japanese medaka HNI (*Oryzias latipes* ASM223471v1), Japanese meda-
200 ka HSOK (*Oryzias latipes* ASM223469v1), Jewelled blenny (*Salarias fasciatus*), Large yellow
201 croaker (*Larimichthys crocea*), Live sharksucker (*Echeneis naucrates*), Lyretail cichlid (*Neo-*

202 lamprologus brichardi), Makobe Island cichlid (*Pundamilia nyererei*), Mexican tetra (*Astyanax*
203 *mexicanus* *Astyanax_mexicanus*-2.0), Midas cichlid (*Amphilophus citrinellus*), Mummichog
204 (*Fundulus heteroclitus*), Nile tilapia (*Oreochromis niloticus*), Northern pike (*Esox lucius*), Or-
205 biculate cardinalfish (*Sphaeramia orbicularis*), Pachon cavefish (*Astyanax mexicanus* *Asty-*
206 *nax_mexicanus*-1.0.2), Pinecone soldierfish (*Myripristis murdjan*), Rainbow trout (*Oncorhyn-*
207 *chus mykiss*), Red-bellied piranha (*Pygocentrus nattereri*), Sailfin molly (*Poecilia latipinna*),
208 Sheepshead minnow (*Cyprinodon variegatus*), Shortfin molly (*Poecilia mexicana*), Siamese
209 fighting fish (*Betta splendens*), Stickleback (*Gasterosteus aculeatus*), Swamp eel (*Monopterus*
210 *albus*), Tetraodon (*Tetraodon nigroviridis*), Tiger tail seahorse (*Hippocampus comes*), Tongue
211 sole (*Cynoglossus semilaevis*), Turbot (*Scophthalmus maximus*), Yellowtail amberjack (*Seriola*
212 *lalandi dorsalis*), Zebra mbuna (*Maylandia zebra*), Zebrafish (*Danio rerio*), Zig-zag eel (*Masta-*
213 *cembelus armatus*).

214

215 These fish species diverged after complete TGD. The human genes were retrieved from EN-
216 SEMBL. The ortholog copy for each gene was established in every one of the 57 fish species.
217 Then, in each species, the fate (singleton vs duplicate) of the entirety of the human gene
218 orthologs was studied. Moreover, a total of 312 human genes known to be haploinsufficient were
219 recovered from ClinGene (<https://www.ncbi.nlm.nih.gov/projects/dbvar/clingen/>), 580 human
220 genes known to be monoallelic (Nag et al., 2015), 206 X human chromosome genes was recov-
221 ered for GeneImprint (<http://www.geneimprint.com/site/genes-by-species>) and 90 genetic im-
222 printing genes (Carrel & Willard, 2005) and the fate of their fish orthologs was recovered. A list
223 of human genes (GRCh38.p13) was generated using BioMart from Ensembl Genes 101. The set
224 of human genes encoding a protein (protein_coding) is selected from the gene type filter. The
225 selected attributes in the homologous category are the different species of teleostens listed in
226 ENSEMBL. Only stable gene IDs were selected. A list of 22836 human genes encoding a protein
227 is listed.

228 We got between 12,918 (Tetraodon) and 14,626 (Brown trout) orthologous genes by fish species
229 (average: 13,882). This does not represent the entire genome of each fish, but allowed us to make
230 strong statistics. Moreover, we compared the global evolution of the whole human genome that
231 had orthologs in fishes with the specific evolution of human MA and HI genes in fish species.
232 We studied whether these fish orthologs of MA and HI genes remained as a duplicate copy, or
233 had return to singleton in the same proportion as whole human ortholog genes.

234 Both Chi Square test statistical analysis and hypergeometric analysis with Benjamini-Hochberg
235 correction were used to test the hypothesis that teleost genes that are orthologs of human MA and
236 HI genes remained more in duplicate than the whole genome. All the statistical tests conducted in
237 our study were performed in R. Moreover, the Panther DataBase (<http://www.pantherdb.org/>)
238 was used to study the gene ontology of teleost genes that are orthologs to human HI genes, and
239 Fisher test with Benjamini-Hochberg correction was used to classify genes according to the fami-
240 ly.

241

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318 Legend figures

319 Figure 1 – Barplot of the global distribution of the genes in each category. Teleost orthologs of
320 human genes localized on the X chromosome, of human haplo-insufficient (HI) genes, of human
321 genes of monoallelic expression (MA, except genes that present a parental imprinting and local-
322 ized on the X chromosome), of human genes that present a parental imprinting. Right: Teleost
323 orthologs of human genes of the whole genome.

324 The yellow bars correspond to the genes that remained in duplicate; the blue bars correspond to
325 the genes returned in singleton. The grey bars correspond to the genes in triplicate or more. The
326 results are presented as mean \pm SEM. * indicates a significant difference compared with the
327 whole genome ($p < 0.05$).

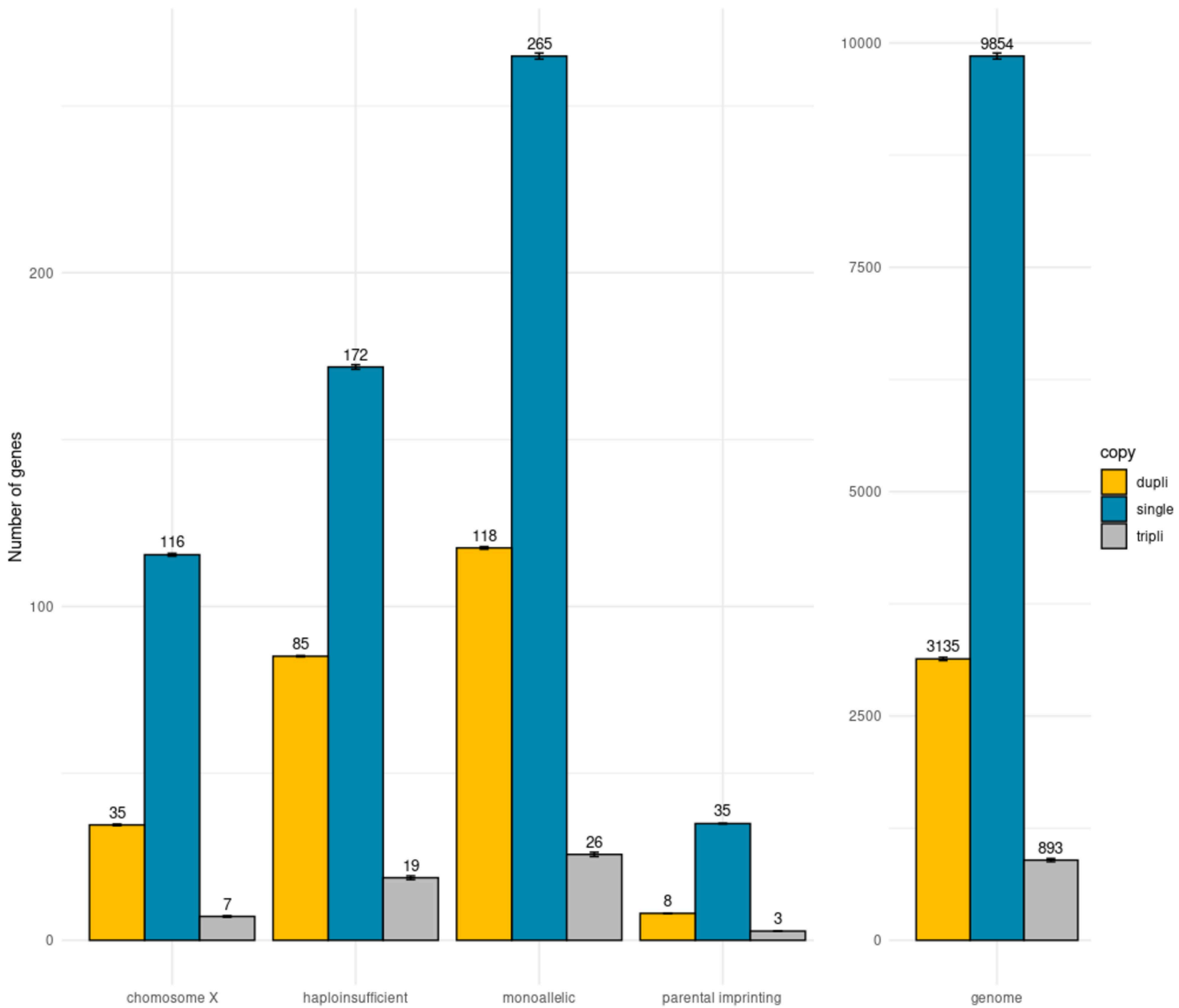
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329 **Suppl Data 1** - Table of statistic tests for each species of teleost and for each category.

330 Each category (HI, MA, X Chromosome, Parental imprinting) have the same construction. By
331 column (category HI for example): (A) Species; (B) Total number of teleost genes returned in
332 singleton. (C) Total number of teleost genes remained in duplicate; (D) Total number of teleost
333 genes in triplicate or more copies; (E) Total number of teleost genes with a human ortholog; (F)
334 Number of teleost orthologs of HI human genes returned in singleton; (G) Number of teleost
335 orthologs of HI human genes remained in duplicate; (G) Number of teleost orthologs of HI hu-
336 man genes in triplicate or more copies; (I) Total number of teleost orthologs to human HI gene;
337 (J) Chi2 value of the repartition of HI orthologs in singleton, in duplicate or more copies in com-
338 parison with the whole genome; (K) P-value of Chi2 test; (L) Chi2 False Discovery Rate (FDR)
339 by Benjamini Hochberg (BH) procedure. (M) P-value of hypergeometric test between singleton
340 and duplicate/more copies. (N) Hypergeometric FDR by BH procedure. (O) P-value of hyperge-
341 ometric test between triplicate or more copies and in duplicate or less copies. (P) Hypergeometric
342 FDR by BH procedure.

343 The same organization of columns is used for the other categories (MA, X chromosome, im-
344 printed genes).

345 Concerning for HI and MA categories, the chi2 test is significant for 55/57 and 47/57 species,
346 respectively, and the hypergeometric test is significant for 57/57 and 50/57 species, respectively,
347 i.e. these orthologs remain more frequently in duplicate than the whole genome. For comparison
348 between triplicate (or more copies) and duplicate (or less copies), the hypergeometric test is sig-
349 nificant for 5/57 (salmonids and carp) and 3/57 species respectively (salmonid and carp as well).



Parental imprinting

Species	Genome				Number of parental imprinting teleost orthologs returned in singleton
	Number of single copy genes	Number of duplicate copy genes	Number of triplicate or more copy genes	Total number of genes	
Amazon molly	10313	2909	712	13934	40
Atlantic herring	10198	2818	624	13640	36
Atlantic salmon	3919	6418	4147	14484	15
Ballan wrasse	9759	3130	1027	13916	35
Barramundi perch	10123	3089	836	14048	40
Blue tilapia	10480	2910	651	14041	38
Blunt-snouted clingfish	10139	2552	549	13240	33
Brown trout	3744	7066	3816	14626	14
Burton's mouthbrooder	10678	2836	514	14028	37
Channel bull blenny	10109	2678	517	13304	39
Channel catfish	10868	2672	590	14130	35
Climbing perch	10638	2979	540	14157	40
Cod	10411	2323	379	13113	35
Common carp	3530	5699	4772	14001	12
Denticle herring	9916	3182	798	13896	34
Eastern happy	10464	2865	713	14042	40
Electric eel	10526	2973	513	14012	36
European seabass	10503	2861	442	13806	37
Fugu	10133	2549	576	13258	31
Gilthead seabream	10711	2870	495	14076	38
Greater amberjack	10750	2957	552	14259	35
Guppy	10623	2662	509	13794	36
Huchen	3797	6234	4510	14541	12
Indian glassy fish	10440	2833	556	13829	40
Indian medaka	10589	2798	531	13918	38
Japanese medaka HdrR	10699	2557	474	13730	38
Japanese medaka HNI	10528	2432	426	13386	39
Japanese medaka HSOK	10609	2563	455	13627	37
Jewelled blenny	9907	2882	881	13670	35
Large yellow croaker	10645	2916	464	14025	40
Live sharksucker	10656	2761	422	13839	36
Lyretail cichlid	10331	2816	640	13787	38
Makobe Island cichlid	10552	2881	537	13970	37
Mexican tetra	10464	3139	621	14224	37
Midas cichlid	10441	2761	595	13797	36
Mummichog	10489	2813	553	13855	38
Nile tilapia	10588	2803	574	13965	36
Northern pike	10447	3354	462	14263	37
Orbiculate cardinalfish	10081	2994	797	13872	36
Pachon cavefish	10333	2844	497	13674	34
Pinecone soldierfish	10486	3042	553	14081	41
Rainbow trout	4274	6304	3445	14023	16
Red-bellied piranha	10767	3201	589	14557	38
Sailfin molly	10238	2843	780	13861	37
Sheepshead minnow	10388	2800	597	13785	39
Shortfin molly	10174	2962	780	13916	39
Siamese fighting fish	10663	2637	432	13732	38

Parental imprinting

Stickleback	10384	2474	337	13195	34
Swamp eel	10771	2672	512	13955	36
Tetraodon	9699	2576	643	12918	32
Tiger tail seahorse	10485	2519	512	13516	38
Tongue sole	10473	2615	555	13643	35
Turbot	10755	2757	340	13852	37
Yellowtail amberjack	10347	3084	735	14166	38
Zebra mbuna	10262	3037	831	14130	39
Zebrafish	10601	2942	552	14095	38
Zig-zag eel	10768	2867	431	14066	38
MAXIMAL	10868	7066	4772	14626	41
MINIMAL	3530	2323	337	12918	12
MEAN	9853.78947	3135.2807	892.82456	13881.89474	34.96491

Parental imprinting

Parental imprinting genes			C
Number of parental imprinting teleost orthologs remained in	Number of parental imprinting teleost orthologs remained in	Total number of parental imprinting genes	Chi2 test
7	1	48	2.16781931651791
5	2	43	1.82790798273002
20	12	47	0.561884391366698
7	3	45	1.25708265096669
5	3	48	3.02984136731787
8	1	47	0.958283980028896
7	2	42	0.0930015339869288
22	11	47	0.433033913803445
9	1	47	0.175350903859632
8	0	47	1.25991609315485
10	3	48	0.432028462558449
7	0	47	2.49787865671492
6	0	41	0.89362774798101
23	14	49	0.0135723202883893
10	4	48	0.00648137825067259
7	0	47	2.77440861277998
6	2	44	1.05585496673543
5	3	45	0.934107006146681
11	2	44	0.871895653983832
7	1	46	1.07320185619677
9	1	45	0.138175058156103
7	3	46	0.0405421597713641
15	20	47	0.00820767112382734
5	3	48	1.59463492246533
5	4	47	0.587584628055566
3	5	46	0.58679384563407
3	3	45	1.72252582204062
4	4	45	0.498274781581108
6	4	45	0.634860547020917
7	0	47	2.17772225095823
6	2	44	0.576815194510765
6	2	46	1.44287002784328
8	2	47	0.258819801348492
6	3	46	1.11609210522594
7	2	45	0.457103939167116
8	1	47	0.676589161381741
10	0	46	0.149711021618182
8	1	46	1.21326465503087
8	3	47	0.364431228885547
7	2	43	0.285783122702426
5	2	48	3.02573776813573
19	10	45	0.547423821515953
9	1	48	0.674580324551083
8	1	46	1.02937426256617
8	0	47	1.47014183202537
8	1	48	1.61775914165671
7	1	46	0.651559243263343

Parental imprinting

6	0	40	0.948025644493705
6	1	43	1.04345241940465
5	2	39	1.01268132801778
7	2	47	0.290012781977809
7	2	44	0.190759310728167
7	0	44	1.05406939973635
7	2	47	1.45588817316256
8	0	47	2.53398985695549
8	0	46	1.35021591208845
7	1	46	0.939691575718443
23	20	49	3.02984
3	0	39	0.00648
8.07018	2.73684	45.77193	0.99444

Parental imprinting

omparison between singleton. duplicate. triplicate and more copies			
Chi2 test p-value	Chi2 test FDR (Benjamini-Hochberg procedure)	Hypergeometric test p-value	Hypergeometric test FDR (Benjamini-Hochberg procedure)
0.140925948344524	0.644340954762652	0.955257133221294	0.977568316880199
0.176374844100671	0.644340954762652	0.943090901659366	0.977568316880199
0.453501987548966	0.654605433377509	0.821149626567003	0.977568316880199
0.262204028175485	0.644340954762652	0.903912791062508	0.977568316880199
0.0817460083949506	0.644340954762652	0.976652019033423	0.977568316880199
0.32761960678852	0.644340954762652	0.87696317803767	0.977568316880199
0.760395453111118	0.817783789194976	0.677908793033684	0.977568316880199
0.51050399575099	0.661971167959833	0.798201872937035	0.977568316880199
0.675399424699246	0.769955344157141	0.715742341253019	0.977568316880199
0.261666973605296	0.644340954762652	0.906801386400846	0.977568316880199
0.510995287547941	0.661971167959833	0.305130226439542	0.977568316880199
0.113999761623482	0.644340954762652	0.966445334353108	0.977568316880199
0.344496112381048	0.644340954762652	0.876163779905064	0.977568316880199
0.907256047031207	0.935834023177937	0.529996518795498	0.977568316880199
0.935834023177937	0.935834023177937	0.52246887814117	0.977568316880199
0.0957820280380141	0.644340954762652	0.972841898421625	0.977568316880199
0.304162478404626	0.644340954762652	0.888929778844751	0.977568316880199
0.333797991174305	0.644340954762652	0.876050192897145	0.977568316880199
0.350431045572671	0.644340954762652	0.220448902183295	0.977568316880199
0.300223340694318	0.644340954762652	0.890464590598548	0.977568316880199
0.710102033877591	0.77838107559659	0.699791527988626	0.977568316880199
0.840424253853623	0.887114490178824	0.636418344382686	0.977568316880199
0.927813452927157	0.935834023177937	0.540566437116734	0.977568316880199
0.206665180631731	0.644340954762652	0.92946446276751	0.977568316880199
0.443354650907187	0.654605433377509	0.825378511102819	0.977568316880199
0.443661603667878	0.654605433377509	0.827044317640085	0.977568316880199
0.189368240632462	0.644340954762652	0.939971071860061	0.977568316880199
0.480259150580241	0.661971167959833	0.8105351216456	0.977568316880199
0.425578018375252	0.654605433377509	0.832467554416157	0.977568316880199
0.140021555814936	0.644340954762652	0.956952948923234	0.977568316880199
0.447563377613561	0.654605433377509	0.825601881717692	0.977568316880199
0.229675471658379	0.644340954762652	0.920118766135543	0.977568316880199
0.610932128750294	0.725481902890975	0.746342142838567	0.977568316880199
0.29076148354252	0.644340954762652	0.892966671281818	0.977568316880199
0.498980570368659	0.661971167959833	0.800323737955721	0.977568316880199
0.410763714242623	0.654605433377509	0.84011592531749	0.977568316880199
0.69881166945096	0.77838107559659	0.704868334360213	0.977568316880199
0.270686527802615	0.644340954762652	0.901511023165936	0.977568316880199
0.546055484693868	0.691670280612232	0.775733066107286	0.977568316880199
0.592935564166668	0.719092067180853	0.757192802973317	0.977568316880199
0.0819530397653641	0.644340954762652	0.977568316880199	0.977568316880199
0.459372233949129	0.654605433377509	0.817877385042016	0.977568316880199
0.411459240677466	0.654605433377509	0.838292761124089	0.977568316880199
0.31030568070788	0.644340954762652	0.884573287417485	0.977568316880199
0.225323317138451	0.644340954762652	0.921880265759968	0.977568316880199
0.203404526576614	0.644340954762652	0.92939124460366	0.977568316880199
0.419555760475738	0.654605433377509	0.83793982459297	0.977568316880199

Parental imprinting

0.330222362168513	0.644340954762652	0.882391184023231	0.977568316880199
0.307019887699219	0.644340954762652	0.88966933324267	0.977568316880199
0.314261331664772	0.644340954762652	0.886970449014357	0.977568316880199
0.590212339400406	0.719092067180853	0.757409644833045	0.977568316880199
0.6622853998245	0.769955344157141	0.724148168991361	0.977568316880199
0.304571725611271	0.644340954762652	0.890643225663272	0.977568316880199
0.227585490213454	0.644340954762652	0.91949956334093	0.977568316880199
0.111418169974796	0.644340954762652	0.966160463046428	0.977568316880199
0.245240374315873	0.644340954762652	0.91368190937084	0.977568316880199
0.332357173110182	0.644340954762652	0.876576965710438	0.977568316880199
0.93583	0.93583	0.97757	0.97757
0.08175	0.64434	0.22045	0.97757
0.39979	0.68383	0.81934	0.97757

Parental imprinting

Comparison between tripli or more copies and dupli or less copies	
Hypergeometric test p-value	Hypergeometric test FDR (Benjamini-Hochberg procedure)
0.919697883721407	1
0.591671556911165	1
0.731618535243889	1
0.655280844788823	1
0.55040331717341	1
0.89300832429622	1
0.524685862083034	1
0.714964489765281	1
0.82750852910998	1
1	1
0.324438851863797	1
1	1
1	1
0.832830330656687	1
0.296585625240612	1
1	1
0.482545496528408	1
0.173894245758489	1
0.575679270258282	1
0.807846579527661	1
0.831273389308077	1
0.240387740607254	1
0.0626763623704843	1
0.303860354974478	1
0.103473462396853	1
0.020560745644604	1
0.171765715980766	1
0.0624317485962584	1
0.329504317689732	1
1	1
0.389906623547002	1
0.636665677356488	1
0.544224894048767	1
0.325558848240955	1
0.583885952588294	1
0.853047315048397	1
1	1
0.780655009805815	1
0.511810362172052	1
0.466686896846573	1
0.567497718467925	1
0.69752313275528	1
0.862731684825785	1
0.930658132592561	1
1	1
0.937562813354077	1
0.770725733121052	1

Parental imprinting

1	1
0.800068676788555	1
0.584939098783418	1
0.536228923097732	1
0.53943407032701	1
1	1
0.708496608567602	1
1	1
1	1
0.761616606692177	1
1	1
0.02056	1
0.64594	1