

# 1 Manuscript Title

## 2 Multi-Scale Modeling of Intensive Macroalgae Cultivation and Marine 3 Nitrogen Sequestration

4

5 Meiron Zollmann<sup>a\*</sup>, Boris Rubinsky<sup>b</sup>, Alexander Liberzon<sup>c</sup> and Alexander Golberg<sup>a</sup>.

6 <sup>a</sup>Porter School of Environmental and Earth Sciences, Tel Aviv University, Israel; and <sup>b</sup>Department  
7 of Mechanical Engineering, University of California at Berkeley, CA, US; and <sup>c</sup>School of  
8 Mechanical Engineering, Tel Aviv University, Israel.

9 \*Corresponding author: Meiron Zollmann

10 **Email:** [meironz@mail.tau.ac.il](mailto:meironz@mail.tau.ac.il)

### 11 **Keywords**

12 Nearshore Cultivation; Biosequestration; Estuarine Bioremediation; Biorefinery Feedstock; Ulva;

### 13 **Abstract**

14 Multi-scale macroalgae growth models are required for the efficient design of sustainable,  
15 economically viable and environmentally safe farms. Here, we develop a multi-scale model for  
16 Ulva sp. macroalgae growth and nitrogen sequestration in an intensive cultivation farm, regulated  
17 by temperature, light and nutrients. The model incorporates a range of scales by incorporating  
18 spatial effects in two steps: light extinction at the reactor scale (1 m) and nutrient absorption at  
19 the farm scale (1 km). The model was validated on real data from an experimental reactor  
20 installed in the sea. Biomass production rates, chemical compositions and nitrogen removal were  
21 simulated under different seasons, levels of dilution in the environment and water-exchange rate  
22 in the reactor. This multi-scale model provides an important tool for environmental authorities and  
23 seaweed farmers who desire to upscale to large bioremediation and/or macroalgae biomass  
24 production farms, thus promoting the marine sustainable development and the macroalgae-based  
25 bioeconomy.

26

### 27 **Introduction**

28

29 Marine conservation and sustainable development is essential for achieving the United Nations'  
30 Sustainable Development Goals<sup>1</sup>. Large scale seaweed farms (> 1 km) could proffer a  
31 sustainable and environmentally safe means for biomass production for biorefineries, to supply  
32 the soaring demand for food, energy and raw materials, without expanding agricultural lands or  
33 freshwater requirements<sup>2-5</sup>. Furthermore, seaweed aquaculture can be utilized for eutrophication  
34 mitigation<sup>6-9</sup>, thus contributing to the international effort to abate nutrient over-enrichment in  
35 coastal ecosystems<sup>10,11</sup> (i.e. the Mediterranean Action Plan<sup>5</sup>). However, the implementation of  
36 commercial cultivation of seaweed beyond East Asia countries is limited, because of a lack of  
37 farming tradition, undeveloped markets, and a questionable economic viability<sup>12</sup>. Large-scale  
38 commercial macroalgae cultivation, which is considered a new technology in most countries,  
39 could be advanced using multi-scale models. The use of multi-scale models to promote new  
40 technologies in reduced time and cost was demonstrated in the Carbon Capture Simulation  
41 Initiative (CCSI)<sup>13</sup>. The CCSI, a partnership among national laboratories, industry, and  
42 universities, was established to enable accelerated commercialization of carbon capture  
43 technologies by developing multiscale models and simulation tools, used to improve design and

44 reduce scale-up risk. Similarly, advances in cultivations of seaweed from small-scale activities to  
45 large scale implementation could also benefit from the availability of multiple scale models. We  
46 propose that these multi-scale models could facilitate the design and optimization of large  
47 seaweed farms by incorporating in the large scale models data from cultivation activities in a  
48 small scale<sup>14,15</sup>, and demonstrate it in a study with mathematical and experimental parts.

49 Current macroalgae growth and nutrient dynamics models were developed for specific  
50 applications. For example, long-term ecological models that attempt to predict macroalgal  
51 productivity and seasonal blooms in prone ecosystems<sup>16,17,26,27,18–25</sup> or “black box” culture models  
52 that focus mostly on on-shore photobioreactors or tanks<sup>9,28</sup> or offshore extensive cultivation<sup>29–31</sup>.  
53 These models, which pursue a basic understanding of the thermodynamics of individual algae  
54 thalli and photobioreactors<sup>32</sup>, can provide a general idea about productivity and seasonal effects  
55 on algae growth. However, they do not incorporate spatial effects at the scale of the farm and its  
56 environment and therefore cannot predict how the algae would behave in a real-life large-scale  
57 farm. On the other hand, as proposed above, multi-scale models that extend from the scale of a  
58 single plant to the scale of the farm could be used for the design of real-life scale seaweed  
59 farms<sup>15</sup>. Such a multi-scale model could incorporate available small-scale mathematical models  
60 and small-scale experimental data. This challenging task involves the combination of multiple  
61 biological, engineering and environmental factors and is the focus of this research.

62 Recently, some studies have proposed to apply intensified macroalgae cultivation, usually done  
63 in photobioreactors, also at near- and off-shore seaweed farms<sup>33,34</sup>. Intensified cultivation  
64 systems rely on frequent harvesting and could benefit from temporal multi-scale models that can  
65 predict biomass production and chemical composition in a time scale of days. As a case study,  
66 we used data from a Mediterranean Sea near-shore intensive growth experimental reactor used  
67 for free-floating *Ulva* species cultivation, which was described by Chemodanov et al.<sup>34</sup>. This  
68 reactor employs airlift pumps and bottom aeration and is suitable for shallow coastal areas or  
69 estuarine systems, in which macroalgae have a natural important role in nutrient cycling<sup>22</sup>. As  
70 these environments are the most prone to harmful eutrophication<sup>35,36</sup> which is responsible for  
71 significant environmental and economic damages<sup>36</sup>, the added value of nutrient bio-sequestration  
72 may increase the economic viability of seaweed cultivation in such locations.

73 In this study we develop a theoretical multi-scale model for macroalgae growth and nitrogen  
74 sequestration in an intensive cultivation seaweed farm, which is regulated by temperature, light  
75 and nutrients (**Fig. 1**). The model is used to simulate farm-scale biomass production and nitrogen  
76 removal in a nutrient-enriched environment, at a temporal and spatial resolution and scale that is  
77 not available today. Specifically, the model predicts farmed seaweed biomass and sequestered N  
78 in different seasons. The model incorporates the required nutrient concentrations and how is the  
79 spatial distribution of biomass composition and productivity affected by levels of airlift pumping  
80 and dilution in the environment. Our model enables the investigation of farm spatial and temporal  
81 responses to environmental variations and provides useful insights on the effects of farm design  
82 and operation on the compliance with environmental and commercial requirements (i.e uniform  
83 biomass composition and minimal energy consumption). Altogether, this multi-scale model  
84 provides an important tool for environmental authorities and seaweed farmers who desire to  
85 upscale to large bioremediation and/or macroalgae biomass production farms, thus promoting the  
86 macroalgae-based bioeconomy.

87

## 88 **Results and Discussion**

89

90 **Calibrated model.** The calibration process started with light extinction parameters ( $K_a$  and  $K_0$ )  
91 and continued to growth function parameters (parameters of eq 1 and SI appendix, eq S1). Based  
92 on a scan of 600 parametric combinations within a pre-defined range, which was built based on  
93 literature values (SI appendix, Table S2), we manually fitted parametric combinations that provide

94 both good RMSREs (<15%) and experiment-specific good relative errors (<20%). We used both  
95 criteria to prevent over- or under dominance of specific returns and environmental conditions (i.e  
96 three returns with a low error and one with a high error). The chosen parametric combination  
97 yielded  $RMSRE_1 = 10.3\%$  for the first step and  $RMSRE_2 = 13.7\%$  for the second step (SI appendix,  
98 **Figs. S5-8** and Table S3).

99 *Light extinction parameters.* We found that the model is not sensitive to  $K_0$  in the examined range  
100 as the optical path in water is short. The best fit between in- and ex-situ light intensity  
101 measurements were found using a light extinction coefficient of  $K_a = 0.15$  (SI appendix, **Fig. S10**),  
102 which is higher than the previously used  $K_a = 0.01^9$  for *Ulva*, but similar to values used for other  
103 algae species<sup>37</sup>. The higher value better represents the significant effect of biomass density on  
104 light extinction.

105 *Growth function parameters.*  $f_{Temp}$  parameters,  $T_{opt}$  and  $T_{max}$ , were adjusted to 18 and 31.5°C,  
106 fitting the literature optimal temperature range of 15-20°C<sup>38,39</sup>.  $K_I$  was adjusted to 20  $\mu\text{mol photons}$   
107  $\text{m}^{-2}\cdot\text{s}^{-1}$  (SI appendix, **Fig. S11**). However,  $K_I$  is a flexible parameter and is known to decrease  
108 when the *Ulva* is acclimated to low light intensities<sup>40</sup>.  $\lambda_{20}$  was adjusted to 2.2%  $\text{day}^{-1}$  (0.16% light  
109  $\text{hour}^{-1}$ , SI appendix, **Fig. S11**), which is low compared to literature values (5-6.5%  $\text{day}^{-1}$ ). A  
110 limitation of this study is that the calibration system was mostly P-limited (N:P >20<sup>2</sup>), a fact that is  
111 not represented in the model and may lead to underestimations of biomass production under P-  
112 saturation conditions. Furthermore, the agreement between modelled and measured final  $N_{int}$   
113 was low, which may be a result of the P-limitation, as high N:P ratio can inhibit N uptake<sup>39</sup>.

114 **Sensitivity analysis.** The parameter with the largest total effect on the total biomass production  
115 and N bio-sequestration (Sobol sensitivity index of 0.35-0.4 in the range of 0 to 1) is  $K_a$ .  $K_I$  and  
116  $\lambda_{20}$ , with total sensitivity indexes of 0.15-0.28 and 0.09-0.1, respectively, have a moderate effect,  
117 and  $\mu_{max}$  has a weak effect (~0.02) on total biomass production and N bio-sequestration.  $N_{env}$ , in  
118 comparison, is highly sensitive only to  $d$  (sensitivity index of 0.97). The effect of other parameters  
119 within examined range is negligible (<0.01) (**Fig. 2**). This analysis shows that our multi-scale  
120 model is sensitive to parameters related to light ( $f_I$ ), which, in the simulated climate, limits growth  
121 only in winter when days are short and sky may be cloudy, and when biomass density in reactor  
122 is high. The sensitivity of the model to parameters related to N ( $\psi_{N_{ext}}$  and  $f_{N_{int}}$ ), on the other  
123 hand, is low, as both reach a steady state relatively rapidly in N rich environments and affect  
124 model outcomes only when  $N_{ext}$  and  $N_{int}$  are low (i.e  $N_{ext}$  below  $K_S$  or  $N_{int}$  below  $N_{int\text{crit}}$ ). The  
125 low sensitivity to N related parameters can be understood in greater depth by the time-scale  
126 separation idea<sup>41</sup>. In diluting environments ( $d > 0$ ), small changes in  $d$  have significant effects on  
127 the results of the multi-scale model as they force rapid  $N_{env}$  and  $N_{ext}$  attenuation regardless of  
128 biomass uptake. contrarily, small changes in  $Q_P$  have no effect on model results as throughout  
129 the examined range N supply does not limit growth. The model was found to be insensitive to  
130  $f_{Temp}$  and  $f_S$  related parameters in the simulated environmental conditions, but this finding should  
131 be examined with a wider range of temperatures and salinities. Model sensitivity to  $\lambda_{20}$  was  
132 higher than the sensitivity to  $\mu_{max}$  probably due to the dependence of  $\mu$  also on other parameters  
133 (T, S, I and  $N_{int}$ ), that lessen the direct effect of  $\mu_{max}$  on model results.

134 **Seasonal trends in biomass production and nitrogen removal.** Productivity and N  
135 sequestration vary significantly seasonally, ranging between 0 and 26.8  $\text{gDW}\cdot\text{day}^{-1}\cdot\text{m}^{-2}$  (0-30  
136  $\text{gDW}\cdot\text{day}^{-1}\cdot\text{m}^{-3}$ ) and between 0.2 and 1.2  $\text{gN}\cdot\text{day}^{-1}\cdot\text{m}^{-2}$  (0.2-1.3  $\text{gN}\cdot\text{day}^{-1}\cdot\text{m}^{-3}$ ), with average values  
137 of 13.3  $\text{gDW}\cdot\text{day}^{-1}\cdot\text{m}^{-2}$  (14.9  $\text{gDW}\cdot\text{day}^{-1}\cdot\text{m}^{-3}$ ) and 0.7  $\text{gN}\cdot\text{day}^{-1}\cdot\text{m}^{-2}$  (0.8  $\text{gN}\cdot\text{day}^{-1}\cdot\text{m}^{-3}$ ) (**Fig. 3**). In a  
138 farm of 100 chained reactors (cultivation area of 200  $\text{m}^2$ ), this translates into annual productivity  
139 of 1210  $\text{gC}\cdot\text{m}^{-2}\cdot\text{year}^{-1}$ , almost four times the estimated average productivity of terrestrial biomass  
140 in the Middle East (290  $\text{gC}\cdot\text{m}^{-2}\cdot\text{year}^{-1}$ ,<sup>42</sup>) and N sequestration of 249  $\text{gN}\cdot\text{m}^{-2}\cdot\text{year}^{-1}$ . Peak  
141 production is expected from the end of February till the middle of March, and a second production  
142 peak is found in November. Production during the summer is very low, which is explained by high  
143 water temperatures (SI appendix, **Fig. S12**). Therefore, effective bio-sequestration cannot be

144 applied during the summer in the modeled conditions. The apparent differences in N  
145 sequestration between the diluting environment ( $d>0$ ), in which high and low  $N_{env}$  water is mixed,  
146 and the non-diluting environment ( $d=0$ ) is discussed below in the spatial effects section.

147 To reduce environmental N levels below a defined, environmentally benign, level, different  
148 seasons require different sizes of the seaweed farm. Considering that a  $10\ \mu\text{M}$  threshold prevents  
149 extreme eutrophication<sup>43</sup>, to avoid damage to the environment, in winter, the dimension of the  
150 farm should be  $1,462\ \text{m}^2$ , in spring the farm should be  $914\ \text{m}^2$  and in the fall  $1,192\ \text{m}^2$  (**Fig. 4a**).  
151 From the perspective of the model, these dimensions of the farm are between 600 to 900 reactor  
152 size macro elements, i.e. the assumption that the single element control volume used in the  
153 analysis is small relative to the entire domain of analysis is acceptable. As important, these  
154 results demonstrate the value of this analysis. They provide a measure on how to design a large  
155 seaweed farm that is safe for the environment.

156 Following are additional examples of how this multi-scale model can be used to design large  
157 seaweed farms. A farm designed according to winter N sequestration abilities will produce  $7.1$   
158 tons  $\text{DW}\cdot\text{year}^{-1}$ , whereas farms designed according to spring or autumn sequestration abilities will  
159 produce only  $4.4$  or  $5.8$  tons  $\text{DW}\cdot\text{year}^{-1}$ , respectively. As a general trend, in high  $N_{env}$  levels, the  
160 relationship between added reactors and N sequestration is linear, but in lower N levels, closer to  
161  $K_S$ , uptake is slower, and more reactors are needed per sequestration unit. **Figs. 4b-d** present N  
162 and biomass dynamics in the last reactor in a farm designed to achieve the threshold in all  
163 seasons (731 reactors). Fixed year-round cultivation cycles result in time and space non-uniform  
164 chemical composition. However, uniform chemical composition can be achieved by adjusting  
165 lengths of cultivation cycles to environmental conditions, specifically, temperature, day length and  
166  $N_{env}$ . Shortening autumn and spring cultivation cycles to 11 and nine days, respectively, for  
167 example, will enable the production of biomass with constant  $N_{int}$ , although won't comply with the  
168 defined  $10\ \mu\text{M}$  threshold during the spring (SI appendix, **Fig. S13**). However, shorter cultivation  
169 cycles come at an expense of higher labor demand and do not necessarily grant higher  
170 accumulated yields.

171 **Spatial effects controlled by dilution and pumping.** In our model, spatial effects on biomass  
172 composition and growth rate appear only when  $N_{env}$  decreases to limiting levels. The rate of this  
173 decrease can be controlled by airlift pumping flow and is accelerated in a diluting environment.

174 *Pumping flow.*  $Q_p$  can be manipulated to control N flux into reactors and thus also chemical  
175 composition and growth rate of the algae (**Fig. 5**). The immediate effect of  $Q_p$  is on the  $N_{env}$  vs  
176  $N_{ext}$  dynamics. High  $Q_p$  minimizes differences between  $N_{env}$  and  $N_{ext}$ , which leads to a faster  
177 reduction in  $N_{env}^1$  and slower reduction in  $N_{ext}^2$  compared to the trajectories of  $N_{env}$  and  $N_{ext}$  with  
178 lower  $Q_p$ <sup>3,4</sup>. Simulating reactors without pumps ( $Q_p = 0$ , dark blue line) decouples  $N_{ext}$  from  $N_{env}$   
179 and eliminates the spatial effects of nutrient absorption. Thus, although  $N_{env}$  does not change,  
180 rapid depletion of  $N_{ext}$  leads to a decrease in  $N_{int}$  which is followed by a decrease in produced  
181 biomass. Therefore, in the described system pumping is essential. High  $Q_p$  promotes bio-  
182 sequestration but may result in a steeper spatial gradient of  $N_{int}$ <sup>5</sup> compared to low  $Q_p$ <sup>6</sup>. Finally,  
183  $Q_p$  can be manipulated according to farm design requirements, controlling farm size and biomass  
184 composition. It should be mentioned that water exchange by pumping has additional important  
185 contributions, such as the supply of inorganic carbon, removal of waste material which may inhibit  
186 growth, and temperature control<sup>34,44</sup>. Furthermore, in an estuarine environment, pumping water  
187 from 1-2 m below the surface can increase salinity, which is crucial for the growth of marine  
188 macroalgae species. However, water pumping is an energy-consuming component of seaweed  
189 farms and should be optimized to minimize its carbon footprint. Previous trials to cultivate *Ulva* in  
190 the described reactors without water exchange were unsuccessful in our group<sup>34</sup>. However, a  
191 thorough review of seaweed cultivation<sup>44</sup> mentioned that water exchange in *Ulva* cultivation can

192 be reduced to 10% day<sup>-1</sup>, equivalent to 15 l-hour<sup>-1</sup> in our work, without a significant change in  
193 yield.

194 *Dilution.* In highly diluting environments, bio-sequestration would be usually ineffective. However,  
195 such environments are not prone to eutrophication and do not require nutrient removal. **Fig. 6**  
196 presents the spring system dynamics in a 100-reactors farm, subjected to 5% dilution between  
197 each two reactors, similar to dilution rates used in literature<sup>30</sup>. Compared to the first reactor  
198 (darkest green), which is not affected by dilution, downstream reactors meet lower  $N_{env}$   
199 concentrations which are translated into lower  $N_{ext}$  and gradually into lower  $N_{int}$  and lower  
200 biomass production. In the simulated conditions ( $N_{env_0} = 500\mu\text{M}$ ), annual decrease in biomass  
201 production due to dilution (968 to 962 kgDW, 0.6%) is significantly smaller than the annual  
202 decrease in N sequestration (50 to 32 kgDW, 36%) (**Fig. 3**). This difference can be explained by  
203 the production of low protein biomass in the downstream, diluted, areas. Larger farms may not be  
204 practical in high-dilution locations, as downstream  $N_{env}$  concentrations would not allow any  
205 growth beyond what the initial  $N_{int}$  allows. However, using high-protein upstream biomass as a  
206 continuous seeding feedstock for further cultivation may enable sustainable low protein biomass  
207 production in such an environment. Following a similar concept, previous works suggested  
208 performing a two-step cultivation process, starting with high biomass production in a nutrient-rich  
209 environment and finishing with carbohydrate accumulation in nutrient-limited environment<sup>45</sup>. As  
210 opposed to the protein-rich biomass that is produced in N enriched environments and can be  
211 used for food and feed applications, such carbohydrate-rich biomass is advantageous for the  
212 extraction of different polysaccharides (i.e starch, ulvan and cellulose) and can be processed into  
213 various forms of biofuels and chemicals<sup>4</sup>.

214 A few previous studies assessed the effectiveness of eutrophication bioremediation in China by  
215 macroalgae cultivation. Generally, this was examined by comparing N and P open sea levels in  
216 cultivation season and off-season, by calculating how much nutrients were removed based on  
217 published data and biomass composition analysis, and by following eutrophication symptoms,  
218 such as hypoxia and harmful algal blooms<sup>7,46,47</sup>. One study, by Fan et al.<sup>6</sup>, advanced into actively  
219 increasing nutrient removal by ecological engineering, specifically artificial upwelling, which is the  
220 pumping of nutrient-rich deep water to the surface. Fan et al.<sup>6</sup> found that artificial upwelling can  
221 increase the average yield of kelp seaweed by 55 g per plant, and developed a few useful  
222 recommendations regarding the conditions in which intensified cultivation can be worthwhile.  
223 Although in a different setup and framework, our work strengthens their recommendation to  
224 optimize pump operation according to algae requirements (nutrients, water exchange and salinity  
225 and temperature control), environmental conditions and regulations, and energy costs. These  
226 considerations change seasonally and spatially, even within the farm itself. Our model, developed  
227 especially for this cause, can help relating to spatial differences during the design and the  
228 operation of seaweed farms.

229 The environmental significance of this work relates to two major environmental issues: climate  
230 change and water pollution. The model developed in this work can be used to quantify and  
231 optimize the environmental significance of large-scale seaweed farms, specifically eutrophication  
232 mitigation. Thus, bioremediation by seaweed farms can be advanced from an unplanned external  
233 benefit to an inherent part of coastal development. Furthermore, if eutrophication mitigation is  
234 compensated by the authorities, this model can play a key role and incentivize the establishment  
235 of new seaweed farms, accompanied by additional environmental and economic benefits, on the  
236 local (i.e. marine conservation and economic development) and global (i.e. carbon sequestration,  
237 sustainable biomass supply and mitigation of fresh water stress) scales. In addition, with some  
238 modifications, this model can be used to model fish cages and integrated multi-trophic  
239 aquaculture (IMTA) and promote sustainable aquaculture and marine development.

## 240 **Conclusions**

241 We developed a multi-scale model for *Ulva* sp. macroalgae growth and nitrogen sequestration in  
242 an intensive cultivation farm, regulated by temperature, light and nutrients. The model enables  
243 spatial simulations by incorporating light extinction effects at the reactor scale (1 m) and nutrient  
244 absorption effects at the farm scale (1 km). Specifically, we simulated: 1. year-round  
245 productivities and N sequestration in the farm; 2. the farm size required for eutrophication  
246 mitigation in different seasons; and 3. spatial distribution of biomass production, chemical  
247 composition and environmental N along the farm in different dilution rates in the environment and  
248 in different airlift pumping flows.

249 The high-resolution spatial and temporal model developed in this work, is an important step  
250 toward implementing precision agriculture techniques in seaweed aquaculture. Such advanced  
251 techniques are expected to improve productivities, efficiencies and accompanied environmental  
252 benefits, leading the way to sustainable marine development, accompanied by multiple economic  
253 and environmental benefits regarding climate change and water pollution mitigation.

254 Future studies need to validate the model on higher-resolution data of all state variables and  
255 engage in uncertainty quantification in different scales. In general, the robustness of the model  
256 will increase by further calibrating it with wider and more diverse empiric data sets, that will raise  
257 additional important constraining factors. Future efforts to improve the model should include  
258 adjusting it to P limited environments and relating to various phenomena that cause uncertainty in  
259 macroalgae cultivation. These phenomena include, for example, an unexplained decline in  
260 biomass, sudden sporulation, age, and history effect on the growth rate, water flow effects on  
261 growth and chemical composition and pest damage. By improving the ability to understand and  
262 describe both temporal and spatial phenomena in a seaweed farm in a resolution of days, these  
263 improved models should help to optimize the design of seaweed farms to combine environmental  
264 improvement and commercial viability.

265

## 266 **Materials and Methods**

267 Our model incorporates multi-scale spatial effects: light extinction at the reactor scale and nutrient  
268 absorption at the farm scale, into a mathematical model of the *Ulva* sp. macroalgae metabolism<sup>2</sup>  
269 (See schematic description in **Fig. 1**). The spatial effects employ the following multiscale  
270 procedures: 1. from a single thallus scale (1 cm) to a reactor scale (1 m), relating to light  
271 extinction in the reactor, and 2. from a reactor scale to a farm-scale (1 km), relating to nutrient  
272 absorption in the farm.

273 The model was calibrated using experimental data from the reactor scale and qualified with a  
274 sensitivity analysis. Thereafter, biomass production rates, chemical compositions and farm-scale  
275 nitrogen removal was simulated under different seasons, levels of dilution in the environment (0-  
276 5% dilution ratio between every two reactors) and water-exchange rate in the reactor (0, 15 and  
277 460 l-hour<sup>-1</sup>). The entire code of this project is available as an open source in  
278 <https://doi.org/10.5281/zenodo.4062432>.

279 **Model assumptions.** The *Ulva* metabolic model assumes that the dynamics of the limiting  
280 nutrient, in this case nitrogen (N), under the constraining effects of environmental conditions (light  
281 intensity (I), temperature (T) and salinity (S)) predicated the dynamics of biomass growth and  
282 chemical composition. In the marine environment, the limiting nutrient is usually N<sup>48</sup> and our  
283 model focuses on N limited environments. However, similar models can be developed also for  
284 other elements such as phosphorus (P) and ferrous that may limit growth too in some marine  
285 environments. Our model also assumes that the organic carbon reserve, depending on carbon  
286 uptake and photosynthesis rates, is not limiting within the modelled conditions. The model follows  
287 the Droop Equation concept, in which the effect of the external, environmental, nutrient  
288 concentration on growth is mediated by internal nutrient concentrations (“cell quota”)<sup>16,49</sup>. This is

289 rather important as changes in internal N concentration occur gradually in a typical time scale of  
290 days whereas significant changes in environmental N concentrations may occur much faster, on  
291 a time scale of hours<sup>50</sup>.

292 Our multi-scale model relates to cultivation in semi-closed reactors with controlled water  
293 exchange. This leads to the differentiation between nutrient concentrations inside the reactor that  
294 interact with the biomass directly, named here external N, and nutrient concentrations outside the  
295 reactor that are affected only secondarily named here environmental N. Environmental N is the  
296 connecting agent that passes onwards in the flow the accumulating signal of changing N  
297 concentrations, which is translated into spatial differences in biomass composition and growth  
298 rate.

299 We used as a reference a cultivation reactor (cage) described by Chemodanov et al.<sup>34</sup>. Each  
300 reactor is assumed to be well-mixed by bottom aeration and is connected to an airlift pump that  
301 supplies the reactor with fresh seawater and nutrients. We also assume water flow through  
302 reactor boundaries is negligible.

303 We simulate the large-scale farm as composed of a continuum of macroscopic reactor size  
304 elements (compartments). This type of mass transfer model is commonly used in pharmaceuticals  
305 which studies mass transfer through macroscopic units referred to as compartment<sup>51</sup>. The model  
306 assumes that the conditions in each reactor size control volume (compartment) can be accurately  
307 represented by one average value (external N) and that the domain of analysis (farm) is much  
308 larger than the macroscopic reactor size element.

309 We define our large-scale farm model as a 3D model (SI appendix, **Fig. S1**). The x-axis is the  
310 direction of the flow and all simulations relate to one row of reactors in this direction. Each reactor  
311 constitutes an N sink, causing the spatial change of environmental N concentrations in the  
312 direction of the flow (x). By assuming the width of this change is small concerning the distance  
313 between the rows, this model becomes applicable also to multiple rows of reactors, with no  
314 variation in the y-axis. Finally, although light extinction increases with depth, potential variations in  
315 biomass with depth (z-axis) can be averaged out due to the well-mixed reactors' assumption.

316 **Model Governing Equations.** The multi-scale model is based on four governing ordinary  
317 differential equations (ODEs), describing the mass balance of four state variables: biomass  
318 density in a reactor ( $m$ , g Dry Weight (DW)·l<sup>-1</sup>, eq 1), biomass internal concentration of N  
319 ( $N_{int}$ , % gN · gDW<sup>-1</sup>, eq 2), external concentration of N in the reactor ( $N_{ext}$ , μmol - N · l<sup>-1</sup>, eq 3)  
320 and the environmental N concentration outside the reactor ( $N_{env}$ , μmol - N · l<sup>-1</sup>, eq 4) under  
321 varying temperatures, light intensities and salinities.

$$\frac{\partial m}{\partial t} = (\mu - \lambda)m, \quad (1)$$
$$\mu = \mu_{max} f_{Temp} f_S \min \{f_{N_{int}}, f_{P_{int}}, f_I\}$$

Initial Condition (I.C):  $m_{(x,t=0)} = m_0$

322 Where  $\mu$  (h<sup>-1</sup>) is biomass specific growth rate formulated of  $\mu_{max}$  (h<sup>-1</sup>), the maximum specific  
323 growth rate, and  $f_{Temp}$ ,  $f_S$ ,  $f_{N_{int}}$ ,  $f_{P_{int}}$  and  $f_I$ , which are the T, S,  $N_{int}$ ,  $P_{int}$ , and I growth functions<sup>2</sup>  
324 (see more in SI).  $\lambda$  is biomass specific losses rate as a function of T and is formulated of  $\lambda_{20}$  (h<sup>-1</sup>),  
325 the specific rate of biomass losses and  $\theta$ , an empiric factor of biomass losses<sup>2</sup>.  $\lambda$  does not  
326 include losses by grazing, sporulation and fragmentation by storms, which vary between different  
327 environments and are highly affected by extreme events. We adjusted daily specific growth and  
328 losses rates to hourly rates, assuming for simplicity that growth and biomass losses occur only  
329 during light hours (see details in SI). This assumption ignores night growth that occurs due to  
330 metabolites produced during light-time photosynthesis<sup>52</sup>, and thus distorts growth distribution  
331 throughout the day. However, the assumption does not affect total daily growth and therefore  
332 does not impair the model accuracy at a temporal resolution of days to weeks.

$$\frac{\partial N_{int}}{\partial t} = \psi_{N_{ext}} - N_{int}\mu_m \quad (2)$$

$$\psi_{N_{ext}} = \frac{N_{intmax} - N_{int}}{N_{intmax} - N_{intmin}} \frac{V_{max} N_{ext}}{K_S + N_{ext}}$$

I.C:  $N_{int}(x,t=0) = N_{int0}$

333 Where  $\psi_{N_{ext}}$  ( $\mu\text{mol}\cdot\text{N}\cdot\text{gDW}^{-1}\cdot\text{h}^{-1}$ ) is the N uptake function, formulated of  $N_{intmax}$  and  $N_{intmin}$  ( $\% \text{gN} \cdot$   
 334  $\text{gDW}^{-1}$ ), the maximum and minimum  $N_{int}$  concentrations, respectively,  $V_{max}$  ( $\mu\text{mol}\cdot\text{N}\cdot\text{gDW}^{-1}\cdot\text{h}^{-1}$ ),  
 335 the maximum N uptake rate and  $K_S$  ( $\mu\text{mol}\cdot\text{N}\cdot\text{l}^{-1}$ ), the N half-saturation uptake constant.  $-N_{int}\mu_m$   
 336 describes  $N_{int}$  dilution in biomass by growth.

$$\frac{\partial N_{ext}}{\partial t} = \frac{Q_p(N_{env} - N_{ext})}{V_{cage}} - \psi_{N_{ext}} m \quad (3)$$

I.C:  $N_{ext}(x,t=0) = N_{ext0}$

337 Where  $Q_p$  ( $\text{l}\cdot\text{h}^{-1}$ ) is the airlift pumping flow and  $V_{cage}$  ( $\text{m}^3$ ) is the reactor volume. The change in  $N_{ext}$   
 338 is the sum of N in incoming airlift pump flow, N in reactor overflow and N uptake by the biomass  
 339 in the reactor.

$$\frac{\partial N_{env}}{\partial t} = \frac{[-Q_s(N_{env_{x-1}}(1-d) - N_{env_x}) - Q_p(N_{env_x} - N_{ext_x})]}{V_{cage}} \quad (4)$$

I.C:  $N_{env}(x,t=0) = N_{ext0}$ , Boundary Condition (B.C):  $N_{env}(x=0,t) = N_{ext0}$

340 Where  $N_{env_x}$  is  $N_{env}$  below reactor x at time t,  $N_{env_{x-1}}$  is  $N_{env}$  below reactor x-1 at time t,  $d$  (%) is  
 341 the dilution ratio between every two reactors and  $Q_s$  ( $\text{l}\cdot\text{h}^{-1}$ ) is the stream flow through an area  
 342 equivalent to the reactor narrow-side cross-section. Thus, the change in  $N_{env}$  is the sum of  
 343 incoming N flows (upstream flow and reactor overflow) and outflowing flows (downstream flow  
 344 and airlift pumping into the reactor). All four ODEs were solved numerically with hourly time steps.

345 **Scale Elements in Model.** The multi-scale model has two scale elements: 1. light extinction at the  
 346 reactor scale that requires dynamic averaging of light intensity per biomass unit, and 2. nutrient  
 347 absorption at the farm scale that requires following the dynamics of environmental N.

348 *Single Thallus to Reactor.* In the metabolic model of a single thallus scale, growth is affected  
 349 directly by incident light intensity (eq 5). In transition to a reactor scale, light intensity is averaged  
 350 per biomass unit, as formulated by Oca et al.<sup>9</sup> (eq 6). This formulation considers water depth in  
 351 the reactor, biomass density and light extinction coefficients of both water and biomass. In both  
 352 equations, we multiplied  $I_0$  by a 0.43 PAR constant, representing the ratio of the sunlight which is  
 353 suitable for photosynthesis<sup>53</sup>.

$$f(I) = \frac{I}{K_I + I} PAR \quad (5)$$

354 Where  $I$  and  $K_I$  ( $\mu\text{mol photons}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ ) are incident light intensity and light half-saturation  
 355 constant, respectively.

$$f(I) = \frac{I_{average}}{K_I + I_{average}} PAR \quad (6)$$

$$I_{average} = \frac{I_0}{K_0 Z + K_a SD} [1 - \exp(-(K_0 Z + K_a SD))]$$

356 Where  $I_{average}$  and  $I_0$  ( $\mu\text{mol photons}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ ) are average photon irradiance in the reactor and  
 357 incident photon irradiance at water surface, respectively,  $SD$  ( $\text{gDW}\cdot\text{m}^{-2}$ ) is stocking density of  
 358 biomass per unit of water surface in the reactor,  $K_0$  ( $\text{m}^{-1}$ ) is water light extinction coefficient,  $Z$  (m)  
 359 is maximum water depth in the reactor and  $K_a$  ( $\text{m}^2\cdot\text{gDW}^{-1}$ ) is *Ulva* light extinction coefficient.



360 *Reactor to Farm*. In a single well-mixed reactor, nutrient reduction by biomass is local and does  
361 not accumulate along the stream. Therefore, eq 4, describing changes in  $N_{env}$ , is redundant.  
362 However, in a seaweed farm, spatial variations in  $N_{env}$  cannot be described without eq 4 that  
363 connects the reactors and the environment. Eq 3, describing changes in  $N_{ext}$ , was derived from  
364 the Convection–Diffusion equation<sup>54</sup> (eq 7). Eq 4, describing changes in  $N_{env}$ , is based on the  
365 same equation, without the uptake term.

$$\frac{\partial N_{ext}}{\partial t} = \nabla \cdot (D \nabla N_{ext}) - \nabla \cdot (v N_{ext}) - \psi_{N_{ext}} m \quad (7)$$

366 Where  $D$  ( $m^2 \cdot s^{-1}$ ) is the average diffusivity coefficient of dissolved inorganic N species and  $v$  ( $m \cdot s^{-1}$ )  
367 is the velocity field in which the dissolved nitrogen is moving. Both eq 3 and eq 4 are derived  
368 from this equation, with specific simplifying assumptions: 1.  $D$  constant in space; 2.  
369 incompressible velocity flow, and 3. zero net diffusivity, as the reactor is well-mixed and there is  
370 no concentration gradient ( $\nabla N_{ext} = 0$ ). Therefore,  $N_{ext}$  in the reactor is affected only by the N  
371 supply by airlift pump (normalized to reactor volume) and N uptake by algae. Eq 4, describing  
372 changes in  $N_{env}$ , follows the same principal form but without the uptake term.

373 **Model Calibration.** We calibrated the model parameters using experimental growth data of *Ulva*  
374 cultivation in a single well-mixed sea-based reactor from Chemodanov et al.<sup>34</sup> (SI appendix, **Figs.**  
375 **S3-4** and Table S1). Model was calibrated by manipulating model parameters and minimizing two  
376 types of root mean square relative errors (RMSRE): 1. RMSRE between biomass growth  
377 projected values (PV) based on in-situ I measurements compared to biomass growth PV based  
378 on ex-situ measurements, and 2. RMSRE between ex-situ I based PV and experimentally  
379 measured growth values. See full description of calibration process in SI appendix.

380 **Sensitivity analysis.** To examine how each parameter, in a defined range (SI appendix, Table  
381 S2), influences model simulations output, we analyzed farm-scale sensitivity of state variables  
382 using SALib, the Sensitivity Analysis Library in Python<sup>55</sup>. Specifically, the analysis focused on the  
383 projected values of total produced biomass, total accumulated  $N_{int}$  and average final  $N_{env}$ , under  
384 the simulation frame of a 100-reactors' farm and one cultivation period per season, that should  
385 suffice to observe both temporal and spatial effects of the different parameters. First, 10 values  
386 and 420 random parametric combinations of all model parameters (SI appendix, Table S2) were  
387 generated using the Saltelli method<sup>56,57</sup>. Next, each combination was run through the model,  
388 producing an array of possible biomass production, N accumulation and final  $N_{env}$  results. Finally,  
389 the results were analyzed using the Sobol analysis<sup>58</sup>, giving each parameter a first order and total  
390 sensitivity index between zero and one.

391 **Model Simulations.** The model was applied to simulate year-round cultivation of *Ulva* sp. in a  
392 row of cultivation reactors in a nutrient-enriched estuary environment located in a semi-arid  
393 climate. Data regarding nutrient concentrations, salinities, water temperature and flow was taken  
394 from the long-term study of Suari et al.<sup>59</sup> on the Alexander estuary, located in the center of Israel  
395 (SI appendix, **Fig. S12** and Table S4). I data was extracted from the IMS database from the Israel  
396 Meteorological Services (<https://ims.data.gov.il/he/ims/6>). Although S varies with depth and can  
397 change dramatically according to flesh flood events and formation of sandbar breaches<sup>59</sup>, effect  
398 on growth was minor and we used a constant value of  $S=30$  PSU. All constraining environmental  
399 factors except nutrients were assumed to be constant in space. Each cultivation cycle started with  
400 a constant set of initial conditions ( $m_0, N_{int,0}, N_{ext,0}$  and  $N_{env,0}$ ) which applied to all reactors.  
401 Harvesting back to initial biomass was performed every two weeks, and accumulated biomass  
402 production was calculated. In addition, N removal from the environment was calculated as the  
403 difference between total N in final and initial biomass. Specific simulations of seasonal N removal  
404 capacity were used to project the number of reactors needed to achieve a 10  $\mu M$ -N level  
405 threshold, which is below levels found in extremely eutrophicated zones<sup>43</sup>, in each season.  
406 Finally, a spatial perspective was added by examining the system dynamics under various

407 pumping levels and in a diluting environment, in which the enriched  $N_{env}$  water is diluted by  
408 mixing with lower  $N_{env}$  water (i.e. 5% dilution between each two reactors).

409

## 410 Acknowledgments

411

412 The authors thank the TAU-Berkeley fund for funding and Tzachi Yaffe for assistance in the  
413 graphic design of figures.

414

## 415 Contributions

416 Meiron Zollmann has designed and performed the research, analyzed the data and wrote the  
417 paper. Boris Rubinsky, Alexander Liberzon and Alexander Golberg have designed and performed  
418 the research, contributed analytic tools and wrote the paper.

## 419 Ethics declarations

420 The authors declare no competing interests.

421

## 422 References

423

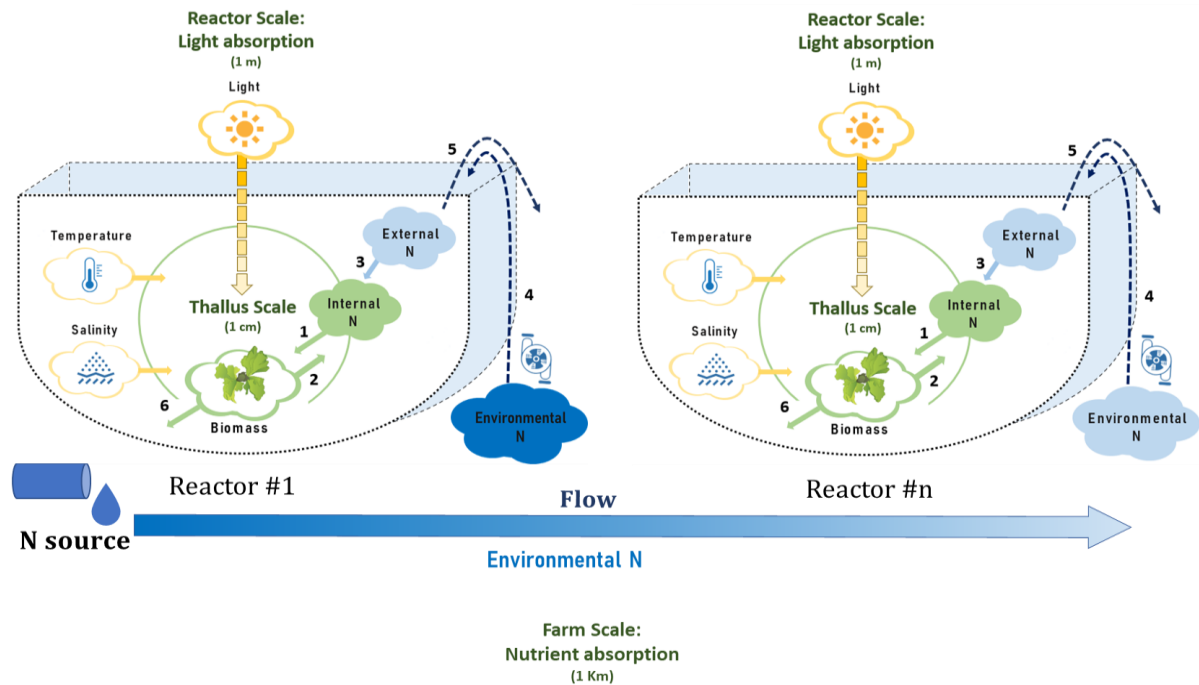
- 424 1. Thomsen, M. & Zhang, X. Life cycle assessment of macroalgal ecoindustrial systems. in  
425 *Sustainable Seaweed Technologies. Cultivation, Biorefinery, and Applications* (Elsevier,  
426 2020).
- 427 2. Lehahn, Y., Ingle, K. N. & Golberg, A. Global potential of offshore and shallow waters  
428 macroalgal biorefineries to provide for food, chemicals and energy: feasibility and  
429 sustainability. *Algal Res.* **17**, 150–160 (2016).
- 430 3. Fernand, F. *et al.* Offshore macroalgae biomass for bioenergy production: Environmental  
431 aspects, technological achievements and challenges. *Renew. Sustain. Energy Rev.*  
432 (2016). doi:<http://dx.doi.org/10.1016/j.rser.2016.10.046>
- 433 4. Zollmann, M. *et al.* Green technology in green macroalgal biorefineries. *Phycologia* **58**,  
434 (2019).
- 435 5. Desa, U. N. *TRANSFORMING OUR WORLD: THE 2030 AGENDA FOR SUSTAINABLE*  
436 *DEVELOPMENT UNITED NATIONS UNITED NATIONS TRANSFORMING OUR*  
437 *WORLD: THE 2030 AGENDA FOR SUSTAINABLE DEVELOPMENT.* (2016).
- 438 6. Fan, W. *et al.* Nutrient Removal from Chinese Coastal Waters by Large-Scale Seaweed  
439 Aquaculture Using Artificial Upwelling. *Water* **11**, 1754 (2019).
- 440 7. Xiao, X. *et al.* Nutrient removal from Chinese coastal waters by large-scale seaweed  
441 aquaculture. *Nat. Publ. Gr.* (2017). doi:10.1038/srep46613
- 442 8. Rose, J. M. *et al.* Nutrient Bioextraction. *Encycl. Sustain. Sci. Technol.* (2015).  
443 doi:10.1007/978-1-4939-2493-6\_944-1
- 444 9. Oca, J., Cremades, J., Jiménez, P., Pintado, J. & Masaló, I. Culture of the seaweed *Ulva*  
445 *ohnoi* integrated in a *Solea senegalensis* recirculating system: influence of light and  
446 biomass stocking density on macroalgae productivity. *J. Appl. Phycol.* **31**, 2461–2467  
447 (2019).
- 448 10. Kanter, D. R., Chodos, O., Nordland, O., Rutigliano, M. & Winiwarer, W. Gaps and  
449 opportunities in nitrogen pollution policies around the world. *Nat. Sustain.* 1–8 (2020).  
450 doi:10.1038/s41893-020-0577-7
- 451 11. Boesch, D. F. *Challenges and Opportunities for Science in Reducing Nutrient Over-*  
452 *enrichment of Coastal Ecosystems.* **25**, (2002).
- 453 12. Radulovich, R. *et al.* Farming of seaweeds. *Seaweed Sustain.* 27–59 (2015).  
454 doi:10.1016/B978-0-12-418697-2.00003-9
- 455 13. Miller, D. C. *et al.* Carbon Capture Simulation Initiative: A Case Study in Multiscale  
456 Modeling and New Challenges. *Annu. Rev. Chem. Biomol. Eng.* **5**, 301–323 (2014).

- 457 14. Wood, D., Capuzzo, E., Kirby, D., Mooney-McAuley, K. & Kerrison, P. UK macroalgae  
458 aquaculture: What are the key environmental and licensing considerations? *Mar. Policy*  
459 **83**, 29–39 (2017).
- 460 15. Van Der Molen, J. *et al.* Modelling potential production and environmental effects of  
461 macroalgae farms in UK and Dutch coastal waters. *biogeosciences Discuss.* (2017).  
462 doi:10.5194/bg-2017-195
- 463 16. Martins, I. & Marques, J. C. A Model for the Growth of Opportunistic Macroalgae  
464 (*Enteromorpha* sp.) in Tidal Estuaries. *Estuar. Coast. Shelf Sci.* **55**, 247–257 (2002).
- 465 17. Port, M. A. (Alex). Measuring and modelling estuarine macroalgae blooms and water  
466 column nutrients. (2016).
- 467 18. Martins, I. *et al.* Significant variations in the productivity of green macroalgae in a  
468 mesotidal estuary: Implications to the nutrient loading of the system and the adjacent  
469 coastal area. *Mar. Pollut. Bull.* **54**, 678–690 (2007).
- 470 19. Port, A., Bryan, K. R., Pilditch, C. A., Hamilton, D. P. & Bischof, K. Algebraic equilibrium  
471 solution of tissue nitrogen quota in algae and the discrepancy between calibrated  
472 parameters and physiological properties. *Ecol. Modell.* **312**, 281–291 (2015).
- 473 20. Nixon, S. W. & Brush, M. J. Modeling the role of macroalgae in a shallow sub-estuary of  
474 Narragansett Bay, RI (USA). *Ecol. Modell.* **221**, 1065–1079 (2010).
- 475 21. Aldridge, J. N. & Trimmer, M. Modelling the distribution and growth of ‘problem’ green  
476 seaweed in the Medway estuary, UK. in *Eutrophication in Coastal Ecosystems* 107–122  
477 (Springer Netherlands, 2009). doi:10.1007/978-90-481-3385-7\_10
- 478 22. Lavaud, R., Filgueira, R., Nadeau, A., Steeves, L. & Guyondet, T. A Dynamic Energy  
479 Budget model for the macroalga *Ulva lactuca*. *Ecol. Modell.* **418**, 108922 (2020).
- 480 23. Duarte, P. & Ferreira, J. G. A model for the simulation of macroalgal population dynamics  
481 and productivity. *Ecol. Modell.* **98**,
- 482 24. Seip, K. L. A computational model for growth and harvesting of the marine alga  
483 *ascophyllum nodosum*. *Ecol. Model.* **8**, 189–199 (1980).
- 484 25. Aveytua-Alcázar, L., Camacho-Ibar, V. F., Souza, A. J., Allen, J. I. & Torres, R. Modelling  
485 *Zostera marina* and *Ulva* spp. in a coastal lagoon. *Ecol. Modell.* **218**, 354–366 (2008).
- 486 26. Solidoro, C., Pecenic, G., Pastres, R., Franco, D. & Dejak, C. Modelling macroalgae (*Ulva*  
487 *rigida*) in the venice lagoon: Model structure identification and first parameters estimation.  
488 *Ecol. Modell.* **94**, 191–206 (1997).
- 489 27. Ren, J. S., Barr, N. G., Scheuer, K., Schiel, D. R. & Zeldis, J. A dynamic growth model of  
490 macroalgae: Application in an estuary recovering from treated wastewater and  
491 earthquake-driven eutrophication. *Estuar. Coast. Shelf Sci.* **148**, 59–69 (2014).
- 492 28. Friedlander, M., Galai, N. & Farbstein, H. A model of seaweed growth in an outdoor  
493 culture in Israel. *Hydrobiologia* **204**, (1990).
- 494 29. Broch, O. J., Slagstad, D., Broch, O. J. & Slagstad, D. Modelling seasonal growth and  
495 composition of the kelp *Saccharina latissima*. *J Appl Phycol* **24**, 759–776 (2012).
- 496 30. Petrell, R. J., Tabrizi, K. M., Harrison, P. J. & Druehl, L. D. *Mathematical model of*  
497 *Laminaria production near a British Columbian salmon sea cage farm. Journal of Applied*  
498 *Phycology* **5**, (1993).
- 499 31. Hadley, S., Wild-Allen, K., Johnson, C. & Macleod, C. Modeling macroalgae growth and  
500 nutrient dynamics for integrated multi-trophic aquaculture. *J. Appl. Phycol.* **27**, 901–916  
501 (2015).
- 502 32. Zollmann, M., Traugott, H., Chemodanov, A., Liberzon, A. & Golberg, A. Exergy efficiency  
503 of solar energy conversion to biomass of green macroalgae *Ulva* (Chlorophyta) in the  
504 photobioreactor. *Energy Convers. Manag.* **167**, (2018).
- 505 33. Golberg, A. & Liberzon, A. Modeling of smart mixing regimes to improve marine  
506 biorefinery productivity and energy efficiency. *Algal Res.* **11**, 28–32 (2015).
- 507 34. Chemodanov, A. *et al.* Feasibility study of *Ulva* sp. (Chlorophyta) intensive cultivation in a  
508 coastal area of the Eastern Mediterranean Sea. *Biofuels, Bioprod. Biorefining* **13**, 864–877  
509 (2019).
- 510 35. McCrackin, M. L., Jones, H. P., Jones, P. C. & Moreno-Mateos, D. Recovery of lakes and

- 511 coastal marine ecosystems from eutrophication: A global meta-analysis. *Limnol.*  
512 *Oceanogr.* **62**, 507–518 (2017).
- 513 36. Gren, I. M. & Limburg, K. Nutrient Recycling and Waste Treatment as a Service from  
514 Estuarine and Coastal Ecosystems. in *Treatise on Estuarine and Coastal Science* **12**,  
515 181–198 (Elsevier Inc., 2012).
- 516 37. Béchet, Q., Shilton, A. & Guieysse, B. Modeling the effects of light and temperature on  
517 algae growth: State of the art and critical assessment for productivity prediction during  
518 outdoor cultivation. *Biotechnology Advances* **31**, 1648–1663 (2013).
- 519 38. Steffensen, D. A. The effect of nutrient enrichment and temperature on the growth in  
520 culture of *Ulva lactuca* L. *Aquat. Bot.* **2**, 337–351 (1976).
- 521 39. Fan, X. *et al.* The effect of nutrient concentrations, nutrient ratios and temperature on  
522 photosynthesis and nutrient uptake by *Ulva prolifera*: Implications for the explosion in  
523 green tides. *J. Appl. Phycol.* **26**, 537–544 (2014).
- 524 40. Sand-Jensen, K. *Photosynthetic responses of Ulva lactuca at very low light.* *int-res.com*  
525 **50**, (1988).
- 526 41. Gunawardena, J. Time-scale separation – Michaelis and Menten's old idea, still bearing  
527 fruit. *FEBS J.* (2013).
- 528 42. Monfreda, C., Ramankutty, N., & Foley, J. A. Farming the planet: 2. Geographic  
529 distribution of crop areas, yields, physiological types, and net primary production in the  
530 year 2000. *Global Biogeochem. Cycles* **22(1)**, (2008).
- 531 43. Boynton, W. R., Murray, L., Hagy, J. D., Stokes, C. & Kemp, W. M. A comparative  
532 analysis of eutrophication patterns in a temperate coastal lagoon. *Estuaries* **19**, 408–421  
533 (1996).
- 534 44. Friedlander, M. Israeli R & D activities in seaweed cultivation. *Isr. J. Plant Sci.* **56**,  
535 15–28 (2008).
- 536 45. Korzen, L., Pulidindi, I. N., Israel, A., Abelson, A. & Gedanken, A. Marine integrated  
537 culture of carbohydrate rich *Ulva rigida* for enhanced production of bioethanol. *RSC Adv.*  
538 **5**, 59251–59256 (2015).
- 539 46. Wu, H. *et al.* Bioremediation efficiency of the largest scale artificial *Porphyra yezoensis*  
540 cultivation in the open sea in China. *Mar. Pollut. Bull.* **95**, 289–296 (2015).
- 541 47. He, P. *et al.* Bioremediation efficiency in the removal of dissolved inorganic nutrients by  
542 the red seaweed, *Porphyra yezoensis*, cultivated in the open sea. *Water Res.* **42**, 1281–  
543 1289 (2008).
- 544 48. Zollmann, M., Traugott, H., Chemodanov, A., Liberzon, A. & Golberg, A. Deep Water  
545 Nutrient Supply for an Offshore *Ulva* sp. Cultivation Project in the Eastern Mediterranean  
546 Sea: Experimental Simulation and Modeling. *Bioenergy Res.* **12**, 1113–1126 (2019).
- 547 49. Lemesle, V. & Mailleret, L. A Mechanistic Investigation of the Algae Growth “Droop”  
548 Model. *Acta Biotheor.* **56**, 87–102 (2008).
- 549 50. Pérez-Mayorga, D. M. *et al.* Nitrogen uptake and growth by the opportunistic macroalga  
550 *Ulva lactuca* (Linnaeus) during the internal tide. *J. Exp. Mar. Bio. Ecol.* **406**, 108–115  
551 (2011).
- 552 51. Rubinsky, B. The Energy Equation for Freezing of Biological Tissue. *J. Heat Transfer* **111**,  
553 988–997 (1989).
- 554 52. Fort, A. *et al.* Extensive variations in diurnal growth patterns and metabolism among *ulva*  
555 spp. Strains. *Plant Physiol.* **180**, 109–123 (2019).
- 556 53. Mottus, M., Sulev, M., Baret, F., Reinart, A. & Lopez, R. Photosynthetically Active  
557 Radiation : Measurement and Modeling. (2011).
- 558 54. Thomas, S. *Introduction to Climate Modelling.* (Springer Science & Business Media,  
559 2011).
- 560 55. Herman, J. & Usher, W. SALib: An open-source Python library for Sensitivity Analysis. *J.*  
561 *Open Source Softw.* **2**, 97 (2017).
- 562 56. Saltelli, A. Making best use of model evaluations to compute sensitivity indices. *Comput.*  
563 *Phys. Commun.* **145**, 280–297 (2002).
- 564 57. Saltelli, A. *et al.* Variance based sensitivity analysis of model output. Design and estimator

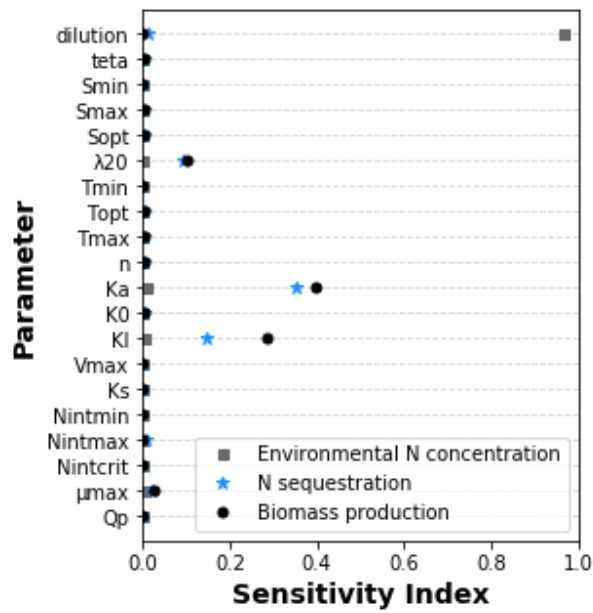
- 565 for the total sensitivity index. *Comput. Phys. Commun.* **181**, 259–270 (2010).  
566 58. Sobol, I. M. Global sensitivity indices for nonlinear mathematical models and their Monte  
567 Carlo estimates. *Math. Comput. Simul.* **55**, 271–280 (2001).  
568 59. Suari, Y. *et al.* Sandbar Breaches Control of the Biogeochemistry of a Micro-Estuary  
569 RIME-restoration of Israeli micro estuaries View project Effect of water circulation on the  
570 Yarqon River ecosystem View project. *Front. Mar. Sci.* (2019).  
571 doi:10.3389/fmars.2019.00224  
572  
573

574 **Figures**

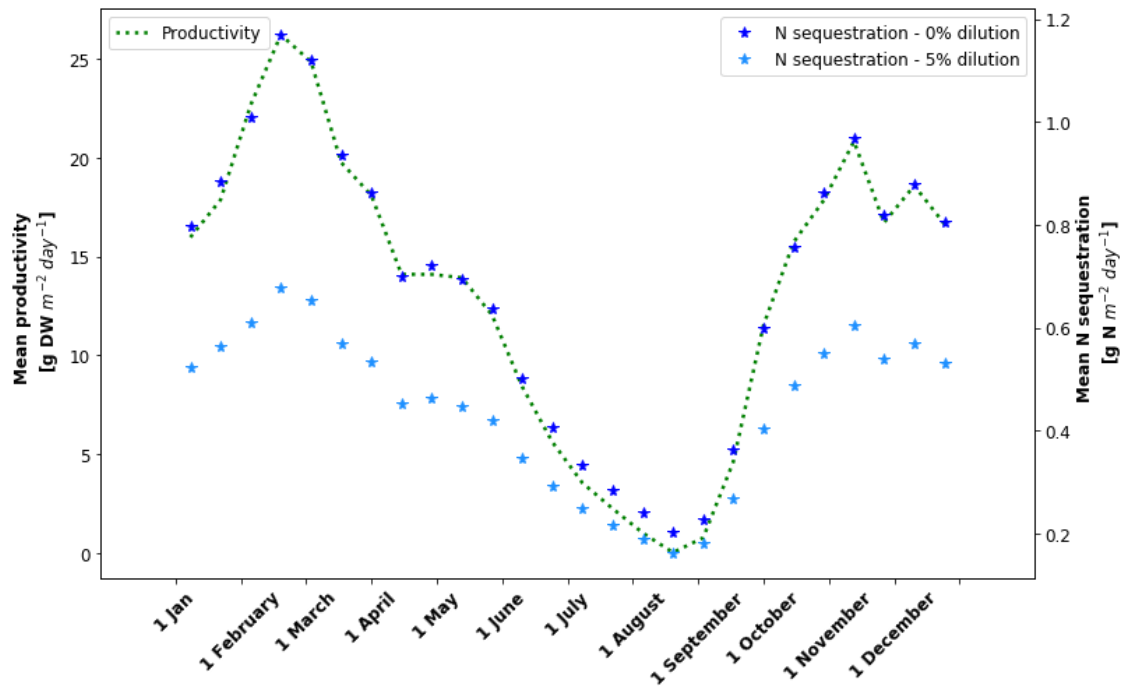


575

576 **Fig. 1.** A schematic description of the multi-scale model. The thallus scale (1 cm, green circle) is  
 577 composed of a simple metabolic model of Ulva, in which the production of new biomass (Ulva  
 578 icon) is affected by internal nitrogen (N, full green cloud) and by constraining environmental  
 579 conditions, including light intensity, salinity and temperature (yellow clouds). The reactor scale (1  
 580 m, U shape pictures) adds light extinction effects (yellow graduated arrow), the concentration of  
 581 external N in the reactor and the concentration of environmental N outside the reactor (dark/light  
 582 blue clouds, depending on N concentration). The farm-scale (1 km, row of reactors starting at  
 583 Reactor #1 and counting downstream to Reactor #n) adds the nutrient reduction caused by  
 584 absorption in reactors along with the flow (Blue graduated arrow). Green and blue clouds  
 585 represent the model state variables. Numbers represent the following processes: 1. Biomass  
 586 growth; 2. Dilution of internal N by growth; 3. N uptake; 4-5. Water exchange by airlift pumping  
 587 and overflow, and 6. Biomass losses.



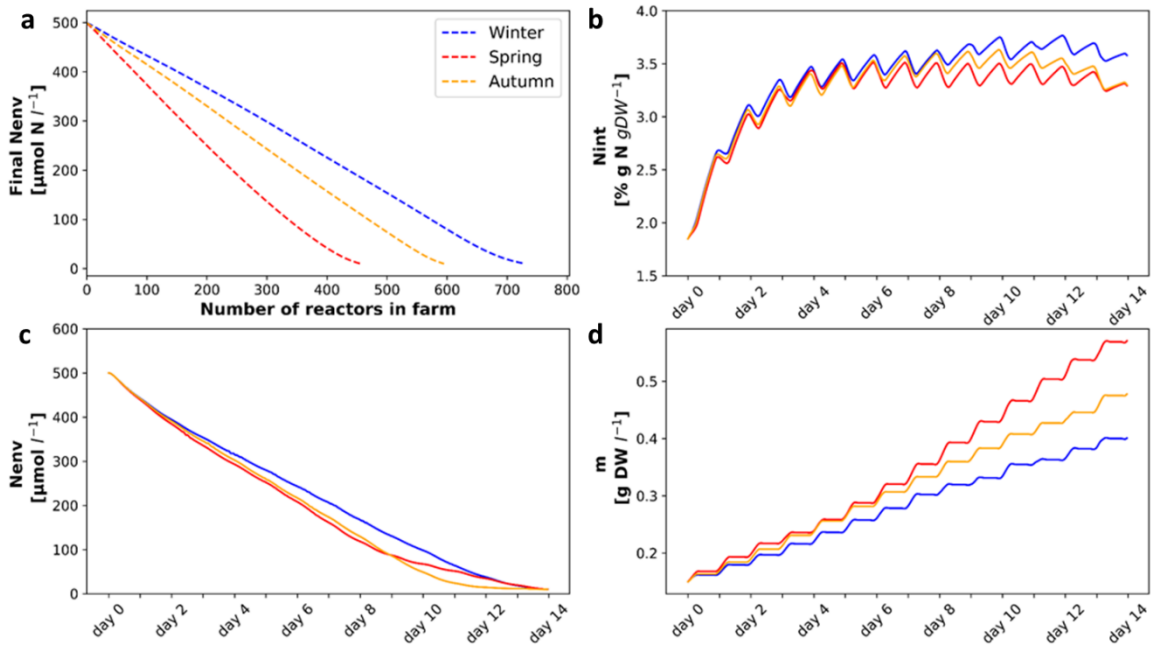
588 **Fig. 2.** Illustrated sensitivity of simulated biomass production (black circles), N sequestration (blue  
589 stars) and final environmental N levels (grey squares) to model parameters, as measured by the  
590 Sobol method.



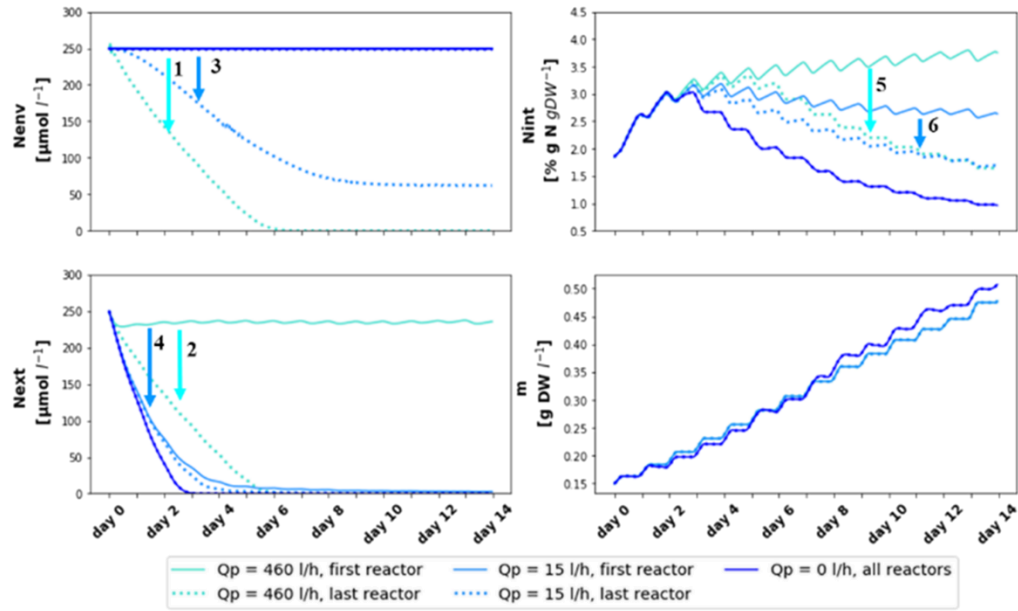
591 **Fig. 3.** Mean productivity ( $\text{gDW}\cdot\text{m}^{-2}\cdot\text{day}^{-1}$ , green dashed line) and mean nitrogen sequestration  
592 ( $\text{gN}\cdot\text{m}^{-2}\cdot\text{day}^{-1}$ ) in a non-diluting environment ( $d=0$ , dark blue stars) and a diluting environment ( $d$   
593  $=0.05$ , 5% dilution between each two reactors, light blue stars) vs week of the year for a farm of  
594 100 reactors.

595





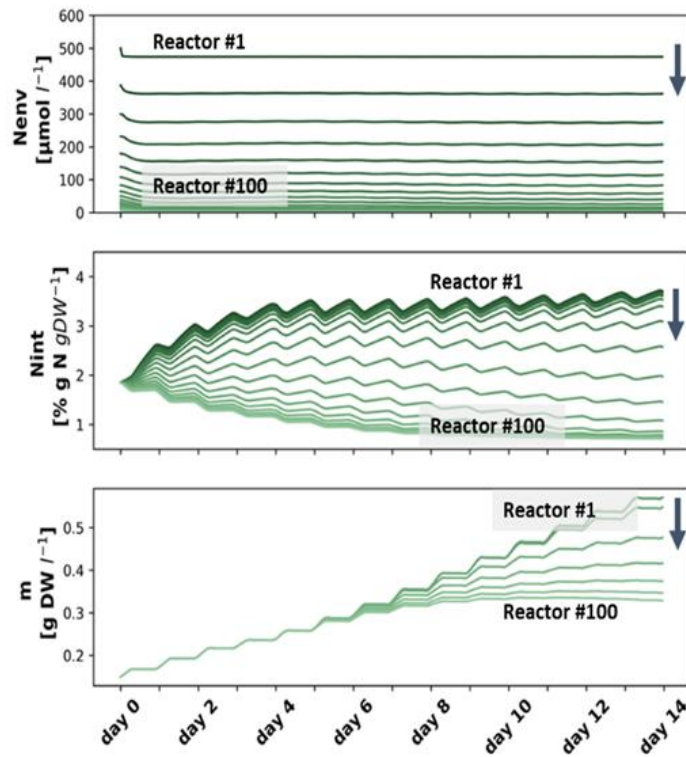
596 **Fig. 4.** (a) Final  $N_{env}$  concentration ( $\mu\text{M-N}$ ) as a function of the number of reactors in different  
597 seasons (Winter in blue, Spring in red and Autumn in yellow), (b-d)  $N_{env}$ ,  $N_{int}$  and  $m$  dynamics  
598 along 14 days' cultivation periods in different seasons, for the last reactor in a farm of 731  
599 reactors.



600

601 **Fig. 5.**  $N_{env}$ ,  $N_{ext}$ ,  $N_{int}$  and  $m$  dynamics along a 14 days cultivation period, simulating  $Q_p$  values of  
 602 0, 15 and 460 l·hour<sup>-1</sup>. Arrows highlight differences between first and last reactor:  $N_{env}$  differences  
 603 for  $Q_p=460$  l·hour<sup>-1</sup> (1) and for  $Q_p=15$  l·hour<sup>-1</sup> (3),  $N_{ext}$  differences for  $Q_p=460$  l·hour<sup>-1</sup> (2) and for  
 604  $Q_p=15$  l·hour<sup>-1</sup> (4) and  $N_{int}$  differences for  $Q_p=460$  l·hour<sup>-1</sup> (5) for  $Q_p=460$  l·hour<sup>-1</sup> (6). Simulation  
 605 parameters and IC: 731 reactors, spring season,  $N_{env0} = 250$  μM-N,  $d = 0$ .

606



607

608 **Fig. 6.**  $N_{env}$ ,  $N_{int}$  and  $m$  dynamics along a 14 days cultivation period in a diluting environment in a  
609 farm of 100 chained reactors. The lines represent  $N_{env}$ ,  $N_{int}$  or  $m$  in each fifth reactor, starting  
610 from  $x=1$  (darkest green) and progressing downstream along the arrows towards the last reactor  
611 ( $x=100$ , lightest green). Simulation parameters and IC: spring season,  $N_{env_0} = 500 \mu\text{M-N}$ ,  $d =$   
612 0.05.

613