

1 **Manuscript title: Chlorophyll content and chlorophyll fluorescence as physiological**  
2 **parameters for monitoring *Orobanche foetida* Poir. infestation on faba bean (*Vicia faba***  
3 **L.)**

4 **Short title: Plant response to *Orobanche* parasitism**

5  
6 Moez Amri<sup>aΦ\*</sup>, Zouhaier Abbes<sup>b</sup>, Imen Trabelsi<sup>c</sup>, Michel Edmond Ghanem<sup>a</sup>, Rachid Mentag<sup>d</sup>,  
7 Mohamed Kharrat<sup>b</sup>

8  
9 <sup>a</sup> Plant Physiology and Crop Improvement Program (PPCI), African Integrated Plant and Soil  
10 Research Group (AIPLaS), AgroBioSciences (AgBS), University Mohammed VI Polytechnic  
11 (UM6P), Lot 660, Hay Moulay Rachid Ben Guerir, 43150, Morocco.

12 <sup>b</sup> Carthage University, Field Crop Laboratory, National Institute for Agricultural Research of  
13 Tunisia (INRAT), Menzah 1, Tunis, Tunisia.

14 <sup>c</sup> Carthage University, Laboratory of Agronomic Sciences and Techniques, National Institute  
15 for Agricultural Research of Tunisia (INRAT), Menzah 1, Tunis, Tunisia.

16 <sup>d</sup> Biotechnology Unit, CRRA-Rabat, Regional Center of Agricultural Research of (INRA),  
17 Rabat, Morocco.

18 <sup>Φ</sup> Former research scientist at Regional Field Crop Research Center of Beja (CRRGC) – Tunisia

19 \* Corresponding author: [amrmoez@gmail.com](mailto:amrmoez@gmail.com) ; [moez.amri@um6p.ma](mailto:moez.amri@um6p.ma)

20 **ORCID**

21 Moez Amri <https://orcid.org/0000-0002-4707-0618>

22

23

24

25

## 26 **Abstract**

27 In total, 39 faba bean (*Vicia faba* L.) advanced lines were evaluated for resistance to broomrape  
28 *Orobanche foetida* under highly infested field conditions. The trials were conducted during two  
29 consecutive cropping seasons at Oued-Beja Research Station in Tunisia. The advanced lines  
30 XAR-VF00.13-1-2-1-2-1 and XBJ90.04-2-3-1-1-1-2A expressed high resistance level to *O.*  
31 *foetida* exceeding those recorded for resistance checks Najeh and Baraca. Results showed that  
32 *O. foetida* significantly affected the biomass, grain yield, chlorophyll content index (CCI) and  
33 the maximum quantum efficiency ( $F_v/F_m$  ratio). No significant effect of *O. foetida* parasitism  
34 was observed on host plant water content (WC). *O. foetida* parasitism significantly affected  
35 both CCI and  $F_v/F_m$  ratio. CCI decreases varied from 46.4% for the susceptible check Badi and  
36 4.2% and 9.3% observed for the genotypes Baraca and XBJ90.04-2-3-1-1-1-2A. Compared to  
37 susceptible check, slight decreases of  $F_v/F_m$  ratio were observed for both advanced lines  
38 XBJ90.04-2-3-1-1-1-2A and XAR-VF00.13-1-2-1-2-1. Correlation between CCI and  $F_v/F_m$   
39 with the resistance to broomrape makes this, easy-to-measure, parameter very useful as a  
40 practical screening tool for early parasitism detection, diagnosis and identification and selection  
41 of high resistant plants against this pathogen.

42 **Key words:** *Vicia faba*, *Orobanche foetida*, Resistance,  $F_v/F_m$ , Chlorophyll Content Index,  
43 Seed Yield

## 44 **Introduction**

45 Broomrapes (*Orobanche* spp.) are holoparasitic plants completely dependent on the host for  
46 their nutritional requirements. In the Mediterranean region, where broomrapes are considered  
47 as a serious threat, *Orobanche* causes important damages and yield losses on many legume  
48 crops [1, 2, 3]. In Tunisia, *Orobanche foetida*, *O. crenata*, *O. cumana*, and *Phelipanche ramosa*  
49 were found parasitizing many crops such as faba bean, chickpea, lentil, grass pea, sunflower,  
50 [4, 5]. While *O. crenata* was mentioned as a serious pest for decades, *O. foetida* has been

51 presented as an emerging problem for many legume crops such as faba bean, chickpea, lentil,  
52 grass pea, medick, common and narbon vetch [4, 6, 7]. The *Orobanche* infested area in Tunisia  
53 is estimated now to more than 80,000 ha mostly situated in the main grain legumes production  
54 area (data non-published). As a result, in high infested fields, farmers abandoned planting  
55 legumes especially faba bean which were substituted by non-host crops such as wheat leading  
56 to a strict wheat mono-cropping system. The detrimental effect of *Orobanche* is associated with  
57 their high seed viability (up to 15-20 years) and multiplication rate. In order to slow down and  
58 stop the fast spread of the parasite from invading new agricultural lands, control strategies and  
59 preventive actions should target decreasing of *Orobanche* seed bank in the soil and minimizing  
60 new seed production [8]. Till date, no single control method/technology has shown successful,  
61 and all control strategies resulted in an incomplete protection of the crop [9, 10, 11]. The only  
62 effective method to control *Orobanche* is through an integrated management approach that  
63 should be based, amongst others, on genetic resistance. Farmers should use resistant varieties  
64 and avoid planting contaminated seeds, spreading contaminated manure and soil, control  
65 animals grazing and avoid moving contaminated machinery from infested to non-infested fields  
66 [12].

67 Research is needed for generating new technologies and developing new resistant varieties and  
68 effective screening tools. Many resistance mechanisms were studied focusing mainly on the  
69 physical and biochemical host-parasite interface including *Orobanche* seed germination  
70 stimulant and inhibitors, host plant roots physical barrier and root architecture [2, 13, 14, 15].

71 While avoidance of dispersal of broomrape is virtually difficult, crop resistance and prevention  
72 measures could be the most effective and economical methods to reduce this root parasitic weed  
73 infestations. Genetic resistance coupled with other control methods result somehow in good  
74 control of the parasite with significant decreases of the damages. Such integrated control  
75 strategy could be improved through early detection and monitoring of the underground

76 infestation and the parasite development. Chlorophyll fluorescence, which is a non-destructive  
77 and rapid assessing mean of photochemical quantum yield and photoinhibition, could be used  
78 for early *Orobanche* infestation and estimate its impact on the host plant. It is widely used as a  
79 plant response indicator under many abiotic and biotic constraints such as heat, drought,  
80 waterlogging, salt stress, nitrogen deficiency, pathogen infection and herbicide resistance [16,  
81 17, 18]. However, only few studies were conducted on parasitism effect on host plant  
82 chlorophyll fluorescence [19, 20, 21]. As reported by Maxwell and Johnson [22], the  
83 photochemical processes alterations are usually the first signs in the stressed plant leaves that  
84 could be used to estimate photosynthetic performance under stress conditions. These  
85 photochemical processes alterations appear in the chlorophyll fluorescence kinetics and induce  
86 changes in the established fluorescence parameters and consequently PSII damages.

87 The purpose of this study was to evaluate the response of 39 faba bean advanced lines to *O.*  
88 *foetida* parasitism, identify potential resistance sources and study the effect of the parasite on  
89 plant growth and seed yield in correlation with chlorophyll content and chlorophyll  
90 fluorescence.

## 91 **Material and Methods**

### 92 **Plant material and field trials**

93 A set of 39 small-seeded faba bean advanced lines, developed from crosses performed in  
94 Tunisia, were used for a first-year (2009/2010) screening and evaluation for resistance to *O.*  
95 *foetida*. Three checks were added to the list, two Tunisian varieties Badi and Najeh and a  
96 Spanish variety Baraca. Both varieties Najeh and Baraca, carrying partial resistance to *O.*  
97 *foetida* and *O. crenata* [23, 1] were used as resistance check while Badi was used as susceptible  
98 check (Table 1). The screening was performed under high *O. foetida* infested sick plot at Oued-  
99 Beja Research Station - Tunisia (36°44'N; 9°13'E).

100 Out of the total tested collection, the two best resistant genotypes XAR-VF00.13-1-2-1-2-1 and  
101 XBJ90.04-2-3-1-1-1-2A were selected all with the three checks to conduct the second-year  
102 (2010/2011) evaluation and confirmation trial conducted under infested and non-infested fields.  
103 For both cropping seasons, trials were conducted in a randomized complete block design with  
104 three replications. Each genotype was planted, at a density of 24 seeds per m<sup>2</sup>, in four rows of  
105 4 m length and 50 cm inter-rows spacing. The planting was performed the last week of  
106 November. No fertilizer's supply or herbicide treatments were applied after plant emergence,  
107 only hand weeding was carried out. Monthly rainfall and average temperature distribution for  
108 the two cropping seasons collected from the iMETOS meteorological station (Pessl  
109 instruments) are presented in the Table 2.

## 110 **Measurements**

111 The field response of the studied genotypes to *O. foetida* parasitism and their level of resistance  
112 was evaluated through different parameters that were measured at different host plant  
113 development stages.

114 During the first-year screening 2009/2010, at harvesting time data related to Parasitism Index  
115 (PI), number of emerged *Orobanche* shoots (EOS) per host plant and seed yield (SY) (g.m<sup>-2</sup>)  
116 were recorded. These data were recorded on the two central rows.

$$117 \quad PI = (OIN * OSV)/100$$

- 118 - OIN: *Orobanche* incidence or percentage of plants showing at least on *Orobanche* emerged shoot
- 119 - OSV: *Orobanche* severity (1-9 scale) or level of damage caused by *Orobanche* on the host plant  
120 development and seed production [24]

121 During the second-year evaluation 2010/2011, in addition to OIN, OSV, PI, EOS and SY  
122 mentioned earlier and recorded at harvesting time, other parameters were determined at pod-  
123 setting stage. from both infested and non-infested fields, five random plants from each plot were  
124 carefully dug-up and biomass and water content (WC) were recorded. Plants selected from  
125 infested field were dug-up with all *Orobanche* attachments that were later classified into

126 emerged and non-emerged tubercles. Total emerged and non-emerged *Orobanche* number and  
127 dry weight per plant and number of days to *Orobanche* emergence (NDOE) were also recorded.  
128 Chlorophyll content index (CCI) was measured once a week between 10 am and 1 pm using a  
129 “Hansatech” CL-01 Chlorophyll Content Meter with a non-destructive method on leaves from  
130 the 11<sup>th</sup> main stem node of five random host plants per plot. For every measurement almost the  
131 same part of the leaf was placed between two clips and the chlorophyll content index was  
132 determined in dual wavelength optical absorbance (620 and 940 nm).

133 The maximum quantum efficiency ( $F_v/F_m$  ratio) was measured also once a week, before  
134 *Orobanche* emergence, in both infested and non-infested field between 10 am and 1 pm. from  
135 each plot, the measurements were performed on two random plants from the two central rows  
136 using a Plant Efficiency Analyzer (*Handy-PEA*, *Hansatech instruments Ltd*, *P02.002 v.*). For  
137 each plant, almost the same part/point of the leaflet situated on the 11<sup>th</sup> main stem node was  
138 delimited by measure clip and was maintained in dark during 16 min by closing the clip shutter.  
139 Dark adaptation time was required to obtain a steady state value of the ratio of variable to  
140 maximum fluorescence. After 16 min, chlorophyll fluorescence transients were induced by a  
141 red light of  $1500 \mu\text{mol.m}^{-2}.\text{s}^{-1}$  intensity.

142 Plant sampling, biomass, water content (WC), Chlorophyll content index (CCI) and the  
143 maximum quantum efficiency ( $F_v/F_m$  ratio) measurements were performed only during  
144 2010/2011 cropping season.

#### 145 **Statistical analysis**

146 ANOVA was performed using the SPSS statistical program v.21 and differences among  
147 treatments for all measurements were compared at  $P=0.05$  and using Duncan's multiple-range  
148 test.

#### 149 **Results**

#### 150 **Field evaluation and identification of potential resistance genotypes to *O. foetida***

151 In total, 39 faba bean genotypes all with resistant and susceptible checks were evaluated for  
152 their resistance to broomrape under high *O. foetida* infested field during the cropping season  
153 2009/2010. Results showed high variability in the resistance to *O. foetida* between the  
154 genotypes. Significant differences between genotypes were observed for the number of  
155 emerged *Orobanche* shoots per plant (EOS), parasitism index (PI) and seed yield (SY) (fig 1).  
156 High negative correlation was observed between SY ( $r = 0.644$ ,  $p \leq 0.001$ ) and both EOS ( $r =$   
157  $0.753^{**}$ ) and PI ( $r = 0.770^{**}$ ). Almost 46% of the tested genotypes showed a resistance level  
158 higher than the resistant check Najeh. Both advanced lines XBJ90.04-2-3-1-1-1-2A and XAR-  
159 VF00.13-1-2-1-2-1 expressed a high resistance level to *O. foetida* with respective PI of 1.2 and  
160 2.2. Only 0.9 and 1.4 EOS were observed for these two genotypes. Such resistance observed  
161 for these two genotypes was reflected by a high seed yield with 154.2 257g.m<sup>-2</sup> and 257g.m<sup>-2</sup>  
162 observed respectively for XBJ90.04-2-3-1-1-1-2A and XAR-VF00.13-1-2-1-2-1. They  
163 produced almost two (1.8) and three (2.9) times more than the resistant check Najeh.

#### 164 **Confirmation of the resistance under infested and non-infested field conditions**

165 Among the 39 tested advanced lines during the cropping season 2009/2010, two lines  
166 XBJ90.04-2-3-1-1-1-2A and XAR-VF00.13-1-2-1-2-1 were selected for their high resistance  
167 *O. foetida*. These two lines, all with resistant and susceptible checks, were planted in 2010/2011  
168 for further studies. The trial was conducted under infested and free *O. foetida* fields. ANOVA  
169 showed high differences ( $P \leq 0.01$ ) between the five studied genotypes for OIN, OSV, DOE,  
170 total *Orobanche* tubercles (TOT), and EOS. High OIN was observed for both cultivars Badi  
171 and Baraca with 81.7% and 85%, respectively (Table 3). Moderate incidence was observed for  
172 Najeh (65%) and the advanced line XAR-VF00.13-1-2-1-2-1 (60%). However, the advanced  
173 line XBJ90.04-2-3-1-1-1-2A showed the lowest *Orobanche* incidence (40%). Maximum  
174 infestation was observed for the susceptible genotype Badi with 5 tubercles per plant against  
175 only 1.2, 1.3 and 1.9 tubercles per plant observed on genotypes Najeh, XAR-VF00.13-1-2-1-2-

176 1 and XBJ90.04-2-3-1-1-1-2A (Fig 2). A total *Orobanche* tubercles number of 4.1 (18.4% less  
177 than cv. Badi) was recorded for cv. Baraca which was reported to be resistant to *O. crenata*  
178 (Nadal et al., 2004). At crop maturity, the number of emerged *Orobanche* shoots (EOS) varied  
179 from 0.9 to 2.7 observed respectively for XBJ90.04-2-3-1-1-1-2A and Badi (Table 3). Only,  
180 1.2, 1.4 and 2.6 shoots per plant were recorded respectively for Najeh, XAR-VF00.13-1-2-1-  
181 2-1 and cv. Baraca. *Orobanche* severity varied from 3 to 6.3 for XBJ90.04-2-3-1-1-1-2A and  
182 Badi, respectively. OSV of 4.3, 3 and 4.3 were recorded for the genotypes XAR-VF00.13-1-2-  
183 1-2-1, Najeh and Baraca, respectively (Table 3). Such infestation levels resulted in a significant  
184 negative parasitism impact on plant growth and seed production for the different tested  
185 genotypes. Differences between in the resistance to *O. foetida* were also confirmed by the  
186 number of days to *Orobanche* emergence which varied from 133 days for the susceptible check  
187 cv. Badi to 145 days observed for the advanced line XBJ90.04-2-3-1-1-1-2A (Table 3).  
188 Compared to Badi, a delay of 2.7, 4, 4.3 and 11.7 days was observed for DOE for the genotypes  
189 XAR-VF00.13-1-2-1-2-1, Baraca, Najeh and XBJ90.04-2-3-1-1-1-2A, respectively.

#### 190 ***Orobanche* parasitism effect on biomass and seed yield**

191 Compared to non-infested field, results showed that *O. foetida* has significantly affected the  
192 host plant biomass production ( $P \leq 0.01$ ) for all the studied genotypes (Table 4). A maximum  
193 decrease (68.5%) of biomass production was observed for the susceptible check Badi against  
194 only 15.6% recorded for Najeh. Respective decreases of 31%, 22.5% and 21.2% were recorded  
195 for the Baraca and both advanced lines XAR-VF00.13-1-2-1-2-1 and XBJ90.04-2-3-1-1-1-2A  
196 (Table 4). For all the studied genotypes, *Orobanche* parasitism has significantly affected  
197 biomass production but not the host plant water content (Table 4). Under infested conditions  
198 and contrary to biomass decreases, no significant reduction was observed for WC compared to  
199 non-infested plants. The susceptible check Badi showed the highest decrease (-8.5%).



200 *Orobanche* parasitism effect on host plant biomass was reflected by seed yield losses for all the  
201 studied genotypes. Compared to non-infested plants, seed yield decreases varied from a  
202 minimum of 3.9% recorded for XBJ90.04-2-3-1-1-1-2A to a maximum of 93.9% observed for  
203 the susceptible check Badi. Respective decreases of 77.4%, 39.5% and 28.8% were recorded  
204 for the genotypes Baraca, Najeh and XAR-VF00.13-1-2-1-2-1 (Fig 4). Among all the tested  
205 genotypes, XAR-VF00.13-1-2-1-2-1 was the most productive under *Orobanche* infested  
206 conditions with 228.4 g.m<sup>-2</sup> representing 3 and 4 times the seed yield recorded for the resistant  
207 checks Najeh (78.3 g.m<sup>-2</sup>) and Baraca (53.8 g.m<sup>-2</sup>). Seed production for cv. Baraca which is  
208 reported to be resistant to *O. crenata* varied from 237.8 g.m<sup>-2</sup> to 53.8 g.m<sup>-2</sup> (77.4% less) under  
209 free and *Orobanche* infested fields, respectively.

#### 210 ***Chlorophyll Content Index and chlorophyll fluorescence***

211 Results showed that compared to non-infested plants (free *Orobanche* field), *Orobanche*  
212 parasitism highly affected ( $P \leq 0.01$ ) the host plant chlorophyll content index (CCI) and  $F_v/F_m$   
213 ratio for all the studied genotypes (Fig 4 and 5).

214 Under infested conditions, CCI decreases varied from 23.6% for Baraca to 77.2% recorded for  
215 the susceptible check Badi between the first and last scoring dates. For the same genotypes  
216 respective decreases of 19.4% and 30.8% were observed under free *Orobanche* field. Under  
217 infested conditions, decreases of 43.8%, 49% and 41.9% were recorded respectively for the  
218 other genotypes Najeh, XAR-VF00.13-1-2-1-2-1 and XBJ90.04-2-3-1-1-1-2A, against only  
219 26.6%, 31.8% and 32.5% recorded in non-infested field. Clear differences were observed for  
220 the CCI between infested and non-infested plants. Such differences were highly significant and  
221 more pronounced for the susceptible check Badi and Baraca. Differences in the variation  
222 between 1<sup>st</sup> and last score under infested and non-infested conditions varied from minimum of  
223 4.2% for Baraca and maximum of 46.4% observed for the susceptible check Badi. A difference  
224 of 17.2% was recorded for both Najeh and XAR-VF00.13-1-2-1-2-1 and 9.3% observed for

225 XBJ90.04-2-3-1-1-2. Except Najeh, all the other genotypes showed high significant  
226 difference between infested and non-infested plants before *Orobanche* emergence.  
227 Results, also, showed that, at the end of the experiment and for all tested genotypes, the  
228 maximum quantum efficiency ( $F_v/F_m$  ratio) was significantly affected by *Orobanche* parasitism  
229 (Fig 5). Under infested field conditions,  $F_v/F_m$  decreased by 58.8% (0.789 to 0.325) for the  
230 susceptible check Badi against only 9.9% (0.787 to 0.709) observed for non-infested plants.  
231 Decreases of 46.2%, 14.5%, 5.9% and 4.7% were recorded, respectively, for the genotypes  
232 Baraca, Najeh, XAR-VF00.13-1-2-1-2-1 and XBJ90.04-2-3-1-1-2A. No significant variation  
233 of  $F_v/F_m$  was observed for all the studied genotypes under free *Orobanche* conditions. For the  
234 three checks Badi, Baraca and Najeh, and before *Orobanche* emergence, no significant  
235 differences were observed in  $F_v/F_m$  ratio between infested and non-infested plant (fig 5). For  
236 both selected genotypes XAR-VF00.13-1-2-1-2-1 and XBJ90.04-2-3-1-1-2A significant  
237 differences were recorded between infested and non-infested plants before *Orobanche*  
238 emergence. High decreases of  $F_v/F_m$  ratio were observed before *Orobanche* emergence for both  
239 genotypes Badi and Baraca against only slight decreases recorded for Najeh and both selected  
240 advanced lines XAR-VF00.13-1-2-1-2-1 and XBJ90.04-2-3-1-1-2A. At the end of the  
241 experiment comparison of  $F_v/F_m$  ratio between non-infested and infested plants showed  
242 differences of 6.4% (0.804 vs 0.753) and 9.5% (0.781 vs 0.707) for both genotypes XBJ90.04-  
243 2-3-1-1-2A and XAR-VF00.13-1-2-1-2-1 against 54.2% (0.709 vs 0.325), 39.3% (0.702 vs  
244 0.426) and 16.5% (0.802 vs 0.670) recorded for the susceptible and resistant checks Badi,  
245 Baraca and Najeh, respectively.

## 246 **Discussion**

247 Results from the first-year screening showed high variability for the resistance to *O. foetida* in  
248 the tested collection. Two advanced lines, XBJ90.04-2-3-1-1-2A and XAR-VF00.13-1-2-1-  
249 2-1 were identified and selected for their high resistance level and high yield under heavy *O.*

250 *foetida* infested conditions. Confirmation trials conducted under infested and non-infested  
251 conditions using both genotypes and susceptible and resistant checks during the two cropping  
252 seasons 2009/2010 and 2010/2011 revealed high significant variation ( $P \leq 0.01$ ) between the  
253 studied genotypes in term of resistance to *O. foetida*. In general, resistance to broomrapes is,  
254 not only the capability of the host to limit the parasite development and the damages that causes,  
255 but also the capacity of the host to grow and produce grains under such parasitism attack.  
256 Significant differences were recorded for DOE, TOT, EOS and *Orobanche* incidence and  
257 severity. Under *Orobanche* infested conditions and compared to the susceptible check Badi, the  
258 genotypes Baraca, Najeh, XAR-VF00.13-1-2-1-2-1 and XBJ90.04-2-3-1-1-1-2A showed a low  
259 infestation level. Najeh and both advanced lines XAR-VF00.13-1-2-1-2-1 and XBJ90.04-2-3-  
260 1-1-1-2A showed the highest resistance to *O. foetida* while Baraca, previously reported to be  
261 resistant to *O. crenata* [23], expressed a moderate resistance to *O. foetida*. Furthermore, data  
262 showed that for all five genotypes, biomass and seed production were negatively affected by  
263 the parasite. Compared to non-infested plants, early wilting symptoms were observed for  
264 parasitized plants resulting in a shortening of the reproductive phase and affecting the plant  
265 biomass and grain yield. For Badi, *Orobanche* has severely restrained plant growth, affected  
266 the flowering and pod setting, and resulted in almost complete damage and yield losses. A  
267 moderate effect of the parasite on plant development and seed production was observed for  
268 other tested genotypes Baraca, Najeh, XAR-VF00.13-1-2-1-2-1 and XBJ90.04-2-3-1-1-1-2A.  
269 Results also showed that despite the biomass decreases recorded for different studied  
270 genotypes, no significant effect of *Orobanche* parasitism was observed on the host water  
271 content (WC) which varied from 70.3% to 78.2% under both free and infested conditions. This  
272 could be explained by the fact that due to the parasitic burden and resources sinking the host  
273 plant limited its biomass and dry matter production and allocation in order to keep its  
274 physiological functioning through a normal and optimum water content. *Orobanche* parasitism

275 effects on host plant growth and biomass production and allocation are directly related to the  
276 infestation level. Ennami et al., [25] reported a high negative correlation between faba bean  
277 and lentil plant growth and biomass production and the number and size of *Orobanche*  
278 shoots/tubercles and thus the nutrients sinking level. Other previous studies reported that the  
279 detrimental effect of both *O. foetida* and *O. crenata* on faba bean grain yield can reach up to  
280 90% - 100% depending on infection severity and the broomrape-crop association [26, 27].  
281 Furthermore, Cameron et al. [28] reported that the parasitic plant *Rhinanthus minor*  
282 significantly reduced biomass production in *Phleum bertolinii* and demonstrated that such  
283 decrease was reflected by changes in photosynthetic activities and significant reductions in the  
284 quantum efficiency of PS II and chlorophyll concentration. In fact, the number of leaves of host  
285 plants as well as leaf greenness is very important in plant eco-physiological studies because it  
286 provides information about physiological responses of plants under stress conditions [29, 30].  
287 In our study, results showed that compared to the non-infested plants, CCI was significantly  
288 affected by *O. foetida* for the five studied genotypes. Significant differences in CCI were  
289 observed, between infested and non-infested plants, before *Orobanche* emergence which can  
290 make this parameter very useful for early detection of the underground *Orobanche* infestation.  
291 In addition, decreases in CCI for infested plants could be explained by the parasite nutritional  
292 requirements that limits the normal growth and functioning of the host plant. Similar results  
293 were reported for tomato/*P. ramosa* pathotype [31] and *Mikania micrantha*/*Cuscuta campestris*  
294 [20]. The latter study showed that despite the CCI decrease observed on the *M. micrantha*  
295 leaves, there was no significant effects of *C. campestris* parasitism on chlorophyll a:b ratio.

296 Our results demonstrated also that CCI decreases was associated with photosynthetic  
297 characteristics variation in the host plant leaves. *O. foetida* affected the photosynthetic system  
298 through significant decreases of the leaves CCI and the maximum quantum efficiency ( $F_v/F_m$   
299 ratio) which was increasingly pronounced over time, especially for the susceptible check Badi.

300 For the different studied genotypes, *O. foetida* parasitism effect on faba bean plants resulted in  
301 a significant ( $P \leq 0.01$ ) decrease of the  $F_v/F_m$  ratio as compared to non-infested plants. High  
302 significant difference was observed in  $F_v/F_m$  ratio between infested and non-infested plant  
303 before *Orobanch*e emergence for both advanced lines XAR-VF00.13-1-2-1-2-1 and XBJ90.04-  
304 2-3-1-1-1-2A. Despite *Orobanch*e parasitism effect, these two genotypes were able to maintain  
305 a good functioning of their PSII as normal as for the free-orobanche plants even after  
306 *Orobanch*e emergence. Contrary, this was not the case for the three susceptible and resistant  
307 checks Badi, Baraca and Najeh as no significant differences in  $F_v/F_m$  ratio between infested and  
308 non-infested plant before *Orobanch*e emergence. These genotypes maintained normal  
309 functioning but under an increasing parasitism pressure, important decreases in  $F_v/F_m$  ratio were  
310 recorded at *Orobanch*e emergence for three genotypes, especially, Badi and Baraca. These  
311 results all with the analyses of  $F_v/F_m$  ratio recorded for all five genotypes in both free and  
312 infested *O. foetida* fields, indicated that  $F_v/F_m$  ratio could be used not only for the quantification  
313 of stress caused by *Orobanch*e parasitism and early detection of the underground infestation  
314 but also the screening and identification of high resistant genotypes.

315 Similar results were reported by Mauromicale et al. [31] who showed that  $F_v/F_m$  ratio, which is  
316 proportional to the PS II quantum yield and well correlated with the photosynthesis quantum  
317 yield [32], was significantly reduced by *P. ramosa* attack on tomato. In the same study, the  
318 authors demonstrated that the  $F_v/F_m$  reduction is mainly induced by an effect on the variable  
319 fluorescence ( $F_v$ ) resulted on damage in PS II electron transport. In addition, Jeschke and  
320 Hilpert [33] showed that *Cuscuta reflexa* induced a sink-dependent stimulation of net  
321 photosynthesis on *Ricinus communis*. Shen et al. [20] showed that *C. campestris* infection  
322 decreases host stomatal conductance, transpiration, chlorophyll content, and soluble protein  
323 concentration on *M. micrantha*, which may directly and indirectly reduce the photosynthesis  
324 rate and affect the host plant growth. These results are contrasting with other studies [34, 35]

325 who reported that broomrape affects host biomass and yield and related traits with only minor  
326 disturbance to leaves tissue but no perceptible effects on photosynthetic rate. More recently,  
327 Ennami et al. [25] showed that effective quantum yield of open photosystem II,  $(F_m' - F)/F_m'$ ,  
328 was significantly reduced by *O. crenata* attack on susceptible faba bean and lentil genotypes.

### 329 **Conclusions**

330 Results showed that *O. foetida* can affect faba bean host plants in/through different ways and  
331 at a big range of scales, from the root to the leaves through the whole plant. Compared to non-  
332 infested plant, high significant difference was recorded between different studied genotypes in  
333 response to *O. foetida* parasitism. The genotypes Najeh, XAR-VF00.13-1-2-1-2-1 and  
334 XBJ90.04-2-3-1-1-2A which expressed the highest resistance levels to *O. foetida* showed a  
335 moderate decrease of biomass and seed production. A significant variation in CCI and  $F_v/F_m$   
336 ratio was observed from individual plants between the tested genotypes and between free and  
337 infested plants for the same genotype. The significant positive correlation observed between  
338  $F_v/F_m$  ratio and high resistance level to *O. foetida* may suggest that this physiological parameter  
339 could be potentially used as a practical screening tool integrating physiological trait for plant  
340 selection and early detection of the root parasitic weeds infestation.

### 341 **Acknowledgment and funding support**

342 This work was supported by the Tunisian Ministry of Agriculture, Hydraulic Resources and  
343 Fishery and the Ministry of Higher Education and Scientific Research. The authors are thankful  
344 to all the field crop laboratory's technical staff for their assistance; Leila Dakhli, Fadhel Sellami,  
345 Hadhemi Abidli, Bisma Soltani and Olfa Mlayeh.

### 346 **Author's contribution**

347 M.A., Z.A., I.T., M.K., designed the research. M.A., Z.A., M.K., performed the experiments.  
348 M.A., Z.A., I.T., R.M., M.E.G., M.K., contributed materials/analysis tools. M.A., Z.A., wrote  
349 the paper. I.T., M.E.G., R.M., M.K., revised the paper. All authors approved the final  
350 manuscript.

### 351 **Conflict of interest**

352 The authors declare that there is no conflict of interest.

### 353 **References**

- 354 1. Kharrat M, Abbes Z, Amri M. A new faba bean small seeded variety “Najeh” tolerant to  
355 *Orobanche* registered in the Tunisian catalogue. *Tunisian Journal of Plant Protection*. 2010;  
356 5: 125-130.
- 357 2. Abbes Z, Sellami F, Amri M, Kharrat M. Variation in the resistance of some faba bean  
358 genotypes to *Orobanche crenata*. *Pak. J. Bot.* 2011; 43(4): 2017-2021.
- 359 3. Ennami M, Briache FZ, Gaboun F, Abdelwahd R, Ghaouti L, Belqadi L, Westwood J,  
360 Mentag R. Genetic diversity of Moroccan *Orobanche crenata* populations revealed by  
361 sequence-related amplified polymorphism markers. *The Journal of Agricultural Science*.  
362 2017; 9(4):164–175
- 363 4. Kharrat M. Etude de la virulence de l'écotype de Beja d'*Orobanche foetida* sur différentes  
364 espèces de légumineuses. In: *Proceedings of the 2nd seminar of REMAFEVE/REMALA*  
365 *Network*, pp. 89, 2002; Hammamet, Tunisia.
- 366 5. Amri M, Abbes Z, Bouhadida M, Kharrat M. First Report of the Parasitic Plant *Phelipanche*  
367 *ramosa* on berseem clover (*Trifolium alexandrium* L.) in Tunisia. *Tunisian Journal of Plant*  
368 *Protection*. 2013; 8(2): 127-132.



- 369 6. Millan T, Madrid E, Cubero JI, Amri M, Castro P, Robio J. Chickpea in: Antonio, DeRon  
370 M, (ed.) Grain Legumes, Series Handbook of Plant Breeding. Springer Science+Business  
371 Media, New York. 2015; 10: pp 499.
- 372 7. Nefzi F, Trabelsi I, Amri M, Triki E, Kharrat M, Abbas Z. Response of some chickpea (*Cicer*  
373 *arietinum* L.) genotypes to *Orobanche foetida* Poir. Parasitism. Chilean Journal of  
374 Agricultural Research. 2016; 76(2): 270-278.
- 375 8. Habimana SA, Nduwumuremyi JD, Chinama R. Management of Orobanche in field crops-  
376 A review. Journal of Soil Science and Plant Nutrition. 2014; 14(1): 43-62.
- 377 9. Abbas Z, Sellami F, Amri M, Kharrat M. Effect of sowing date on *Orobanche foetida*  
378 infection and seed yield of resistant and susceptible faba bean cultivars. Acta  
379 Phytopathologica et Entomologica Hungarica. 2010; 45(2): 267-275.
- 380 10. Amri M, Trabelsi I, Abbas Z, Kharrat M. Release of a new faba bean variety "Chourouk"  
381 resistant to the parasitic plants *Orobanche foetida* and *O. crenata* in Tunisia. Intl. J. Agric.  
382 Biol. 2019; 21(3): 499–505
- 383 11. Briache FZ, Ennami M, Mbasani-Mansi J, Lozzi A, Abousalim A, El Rodeny W, Amri M,  
384 Triqui ZEA, Mentag R. Effects of salicylic acid and indole acetic acid exogenous  
385 applications on induction of faba bean resistance against *Orobanche crenata*. The plant  
386 pathology journal. 2020; 36(5): 476 – 490.
- 387 12. Abu-Irmaileh BE, Labrada R. The problem of *Orobanche* spp. in Africa and Near East.  
388 FAO, 2017; [http://www.fao.org/agriculture/crops/thematic-](http://www.fao.org/agriculture/crops/thematic-sitemap/theme/biodiversity/weeds/issues/oro/en/)  
389 [sitemap/theme/biodiversity/weeds/issues/oro/en/](http://www.fao.org/agriculture/crops/thematic-sitemap/theme/biodiversity/weeds/issues/oro/en/)
- 390 13. Trabelsi I, Abbas Z, Amri M, Kharrat M. Performance of faba bean genotypes with  
391 *Orobanche foetida* and *Orobanche crenata* infestation in Tunisia. Chilean Journal of  
392 Agricultural Research. 2015; 75 (01): 27-34.



- 393 14. Trabelsi I, Yoneyama K, Abbes Z, Xie X, Amri M, Kharrat M. Yoneyama K.  
394 Characterization of strigolactones produced by *Orobanche foetida* and *Orobanche crenata*  
395 resistant and -susceptible faba bean genotypes and effect of phosphorous, nitrogen, and  
396 potassium deficiencies on strigolactone production. South African Journal of Botany. 2017;  
397 108: 15-22.
- 398 15. Briache FZ, Ennami M, Mansi JM, Gaboun F, AbdelWahd R, Fatimi ZA, El-Rodeny W,  
399 Amri M, Triqui ZA, Mentag R. Field and controlled conditions screenings of faba bean  
400 genotypes for resistance to *Orobanche crenata* Forsk. and investigation of resistance  
401 mechanisms. J. Plant Dis. Prot. 2019; 126(3): 211–224
- 402 16. Korres NE, Froud-Williams RJ, Moss SR. Chlorophyll fluorescence technique as a rapid  
403 diagnostic test of the effects of the photosynthetic inhibitor chlorotoluron on two winter  
404 wheat cultivars. Annals of Applied Biology. 2003; 143(1): 53-56.
- 405 17. Pavlovic D, Vrbnicanin S, Bozic D, Fischer AJ. Morpho-physiological traits and atrazine  
406 sensitivity in *Chenopodium album* L. Pest Management Science. 2008; 64(2): 101-107.
- 407 18. Amri M, El-Ouni MH, Ben Salem M. Waterlogging affect the development, yield and  
408 components, chlorophyll content and chlorophyll fluorescence of six bread wheat genotypes  
409 (*Triticum aestivum* L.). Bulg. J. Agric. Sci. 2014; 20(3): 647-657.
- 410 19. Cameron DD, Hwangbo JK, Keitha M, Geniez JM, Kraushaardar D, Rowntree J, Seel WE.  
411 Interactions between the hemiparasitic angiosperm *Rhinanthus minor* and its hosts: from the  
412 cell to the ecosystem. Folia Geobotanica. 2005; 40: 217-229.
- 413 20. Shen H, Hong L, Chen H, Ye WH, Cao HL, Wang ZM. The response of the invasive weed  
414 *Mikania micrantha* to infection density of the obligate parasite *Cuscuta Campestris* and its  
415 implications for biological control of *M. micrantha*. Botanical Studies 2011; 52: 89-97.

- 416 21. Walters DR. Photosynthesis in Attacked Plants and Crops. In: Physiological Responses of  
417 Plants to Attack. Crop & Soil Systems Research Group SRUC Edinburgh, (Eds Walters  
418 D.R.), pp. 248. 2015; Wiley-Blackwell, UK
- 419 22. Maxwell K, Johnson GN. Chlorophyll fluorescence - A practical guide. Journal of  
420 Experimental Botany. 2000; 51(345): 659-668.
- 421 23. Nadal S, Moreno JI, Cubero MT. Registration of 'Baraca' Faba Bean. Crop Science. 2004;  
422 44(5). <https://doi.org/10.2135/cropsci2004.1864a>
- 423 24. Abbes Z, Kharrat M, Delavault P, Simier P, Chaibi W. Field evaluation of the resistance of  
424 some faba bean (*Vicia faba* L.) genotypes to the parasitic weed *Orobanche foetida* Poiret.  
425 Crop Protection. 2007; 26: 1777-1784.
- 426 25. Ennami M, Mansi, MJ, Briache FZ, Oussible N, Gaboun F, Ghaouti L, Belqadi L, Ghanem  
427 ME, Aberkani K, Westwood J, Mentag R. Growth-defense tradeoffs and source-sink relation  
428 explain the responses of susceptible and resistant faba bean and lentil genotypes to infection  
429 by *Orobanche crenata*. J. of Crop protection. 2020; 127:  
430 <https://doi.org/10.1016/j.cropro.2019.104924>
- 431 26. Kharrat M, Souissi T. Research on *Orobanche foetida* and *O. crenata* in Tunisia. In:  
432 Integrated management of orobanche in food legumes in the Near East and North Africa.,  
433 Proceedings of the expert consultation on IPM for *Orobanche* in Food Legume Systems in  
434 the Near East and North Africa, (Eds Dahan, R., El Mourid M.), 2004; 106-110.
- 435 27. Trabelsi I, Abbes Z, Amri M, Kharrat M. Response of some selected faba bean (*Vicia faba*  
436 L.) genotypes to *Orobanche* spp. in Tunisia. Plant Production Science. 2016 ; 19: 562-573.
- 437 28. Cameron DD, Geniez JW, Seel LJ, Irving IJ. Suppression of host photosynthesis by the  
438 parasitic plant *Rhinanthus minor*. Annals of Botany. 2008; 101: 573–578.
- 439 29. Yordanov I, Velikova V, Tsonev T. Plant responses to drought, acclimation, and stress  
440 tolerance. Photosynthetica. 2000; 38: 171-186.

- 441 30. Valladares F, Niinemets U. Shade tolerance, a key plant feature of complex nature and  
442 consequences. *Annu. Rev. Ecol. Evol. Syst.* 2008; 39: 237-257.
- 443 31. Mauromicale G, Monaco AL, Longo AMG. Effect of Branched Broomrape (*Orobanche*  
444 *ramosa*) Infection on the Growth and Photosynthesis of Tomato. *Weed Science.* 2008; 56:  
445 574-581.
- 446 32. Butler WL, Kitajima M. Fluorescence quenching in photosystem II of chloroplasts. *Biochim*  
447 *Biophys Acta.* 1975; 376(1):116-25.
- 448 33. Jeschke WD, Hilpert A. Sink-stimulated photosynthesis and sink dependent increase in  
449 nitrate uptake: nitrogen and carbon relations of the parasitic association *Cuscuta reflexa*-  
450 *Ricinus communis*. *Plant, Cell and Environment.* 1997; 20 : 47-56.
- 451 34. Dale H, Press MC. Elevated atmospheric CO<sub>2</sub> influences the interaction between the  
452 parasitic angiosperm *Orobanche minor* and its host *Trifolium repens*. *New Phytol.* 1998;  
453 140: 65-73.
- 454 35. Hibberd JM, Quick WP, Press MC, Scholes JD. Can source–sink relations explain responses  
455 of tobacco to infection by the root holoparasitic angiosperm *Orobanche cernua*? *Plant, Cell*  
456 *and Environment.* 1998; 21: 333–340  
457

## Figures list

**Figure 1:** Parasitism Index (PI), Emerged Orobanche shoots per Plant (EOS) and Seed Yield (SY g.m<sup>-2</sup>) recorded for 39 genotypes under *O. foetida* infested field during the cropping season 2009/2010.

**Figure 2:** Total emerged and non-emerged *O. foetida* tubercles per plant recorded for different studied genotypes at pod setting stage.

bioRxiv preprint doi: <https://doi.org/10.1101/2020.10.19.345479>; this version posted October 19, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

**Figure 3:** Seed yield (g.m<sup>-2</sup>) recorded for different studied genotypes in both *O. foetida* infested and non-infested fields during the cropping season 2010/2011.

**Figure 4:** Chlorophyll content index (CCI) recorded for the different studied genotypes in both *O. foetida* infested and non-infested fields during the cropping season 2010/2011.

**Figure 5:** F<sub>v</sub>/F<sub>m</sub> ratio recorded for the different studied genotypes in both *O. foetida* infested and non-infested fields during the cropping season 2010/2011.

bioRxiv preprint doi: <https://doi.org/10.1101/2020.10.19.345173>; this version posted October 19, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

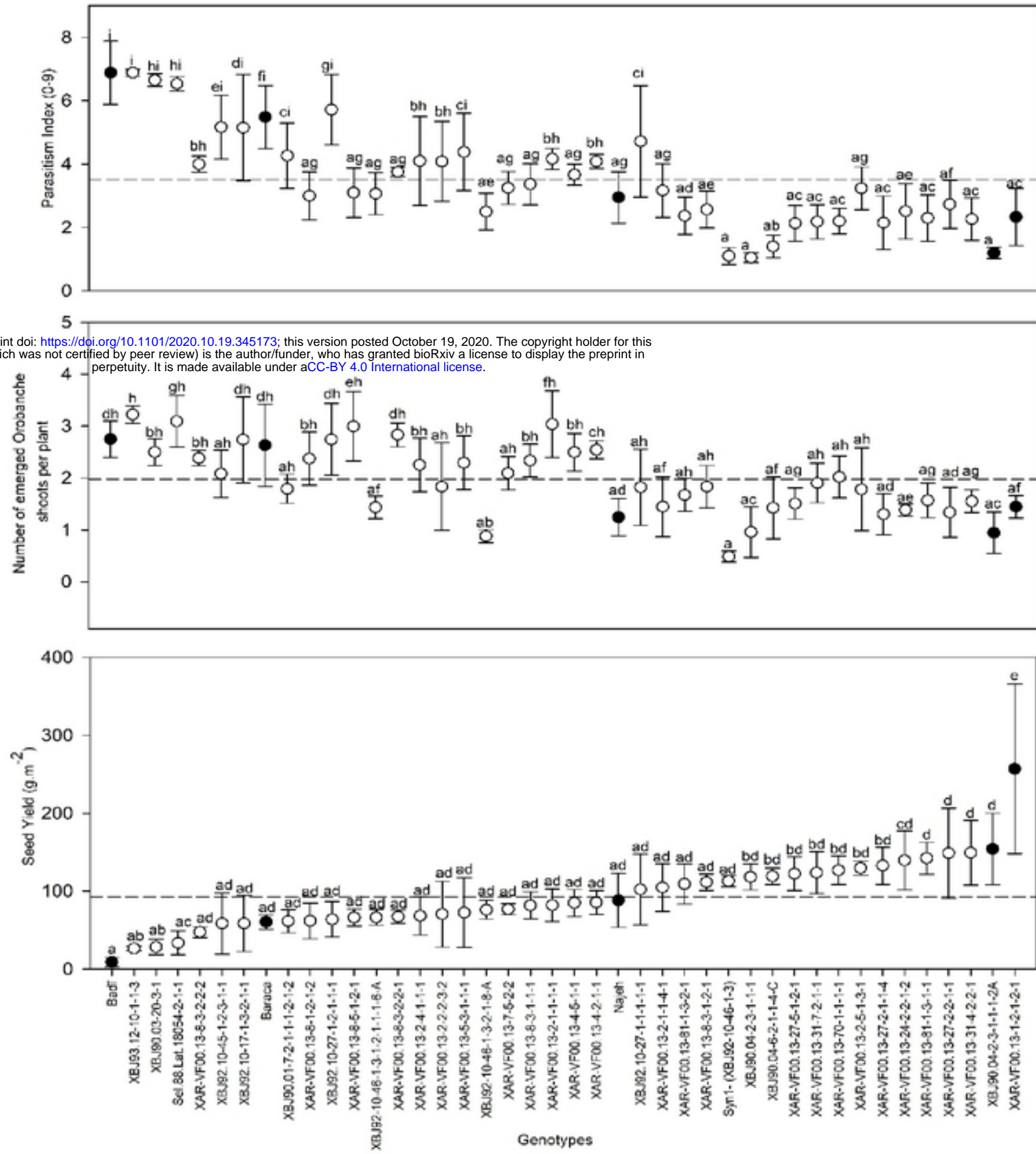


Figure 1: Parasitism Index (PI), Emerged Orobanchae shoots per Plant (EOS) and Seed Yield (SY g.m<sup>-2</sup>) recorded for 39 genotypes under *O. foetida* infested field during the cropping season 2009/2010. Data are three replication means  $\pm$  SE. Data with the same letter(s) are not significantly different at  $P=0.05$  (Duncan test).

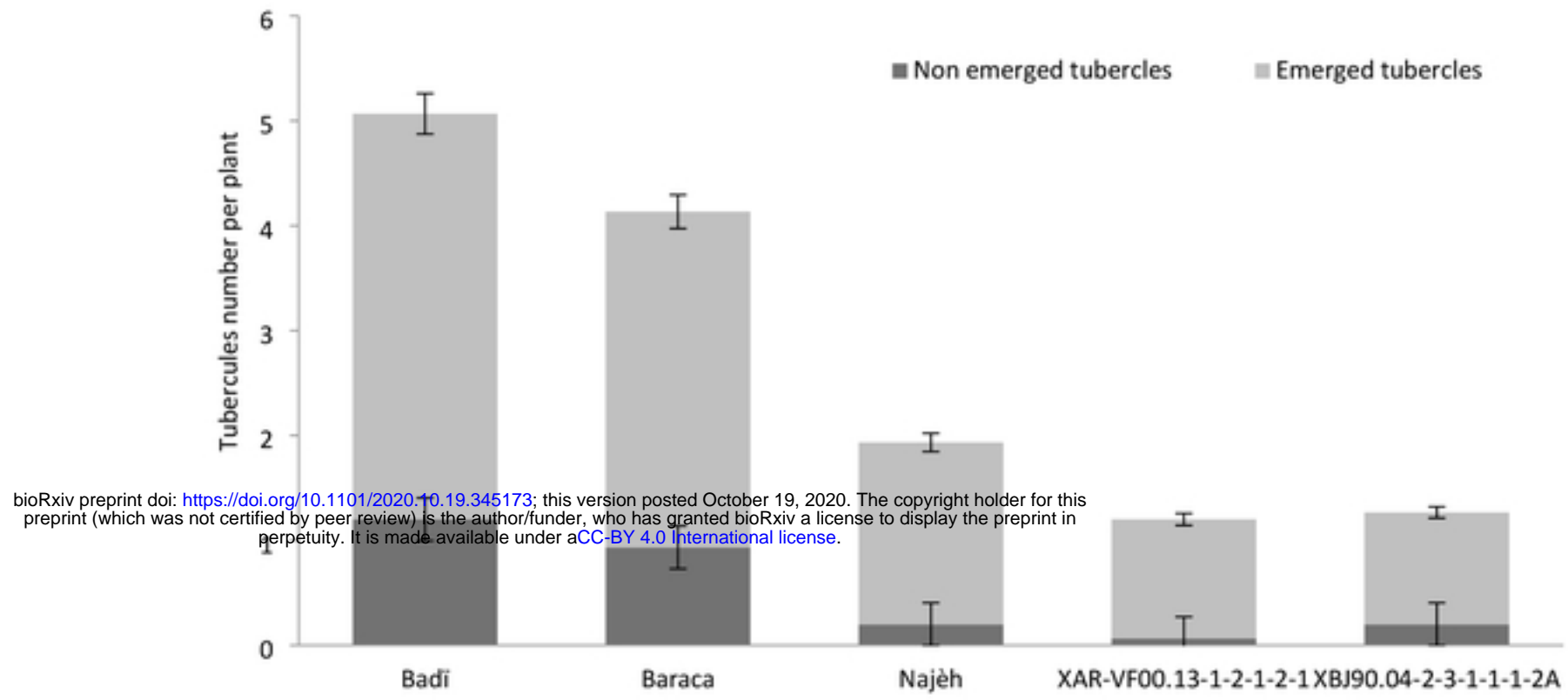


Figure 2: Total emerged and non-emerged *O. foetida* tubercles per plant recorded for different studied genotypes at pod setting stage. Data are three replication means  $\pm$  SE.



bioRxiv preprint doi: <https://doi.org/10.1101/2020.10.19.345173>; this version posted October 19, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

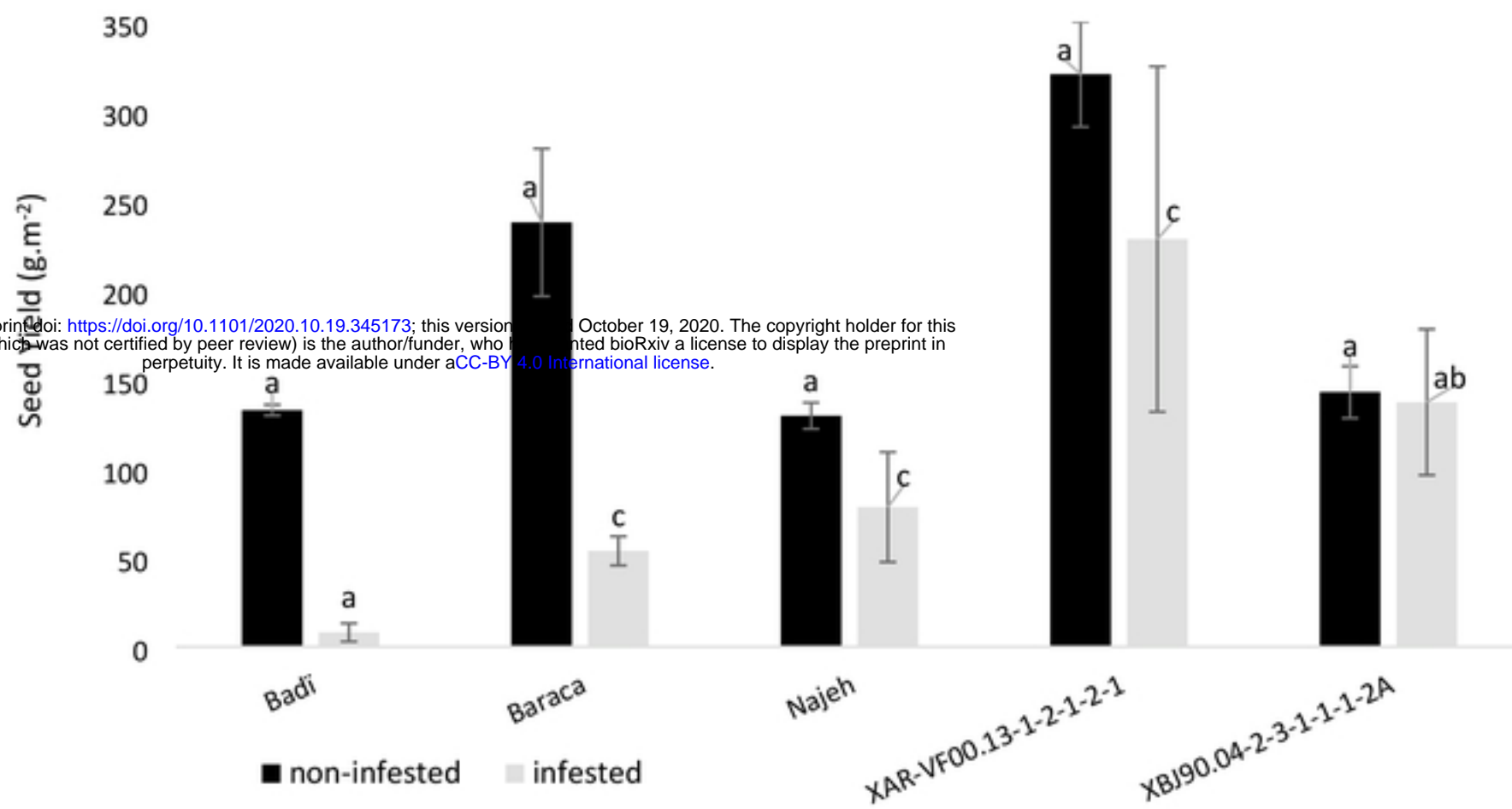


Figure 3: Seed yield (g.m<sup>-2</sup>) recorded for different studied genotypes in both *O. foetida* infested and non-infested fields during the cropping season 2010/2011. Data are three replication means  $\pm$  SE. For each treatment (infested and non-infested) data with the same letter(s) are not significantly different at  $P=0.05$  (Duncan test).

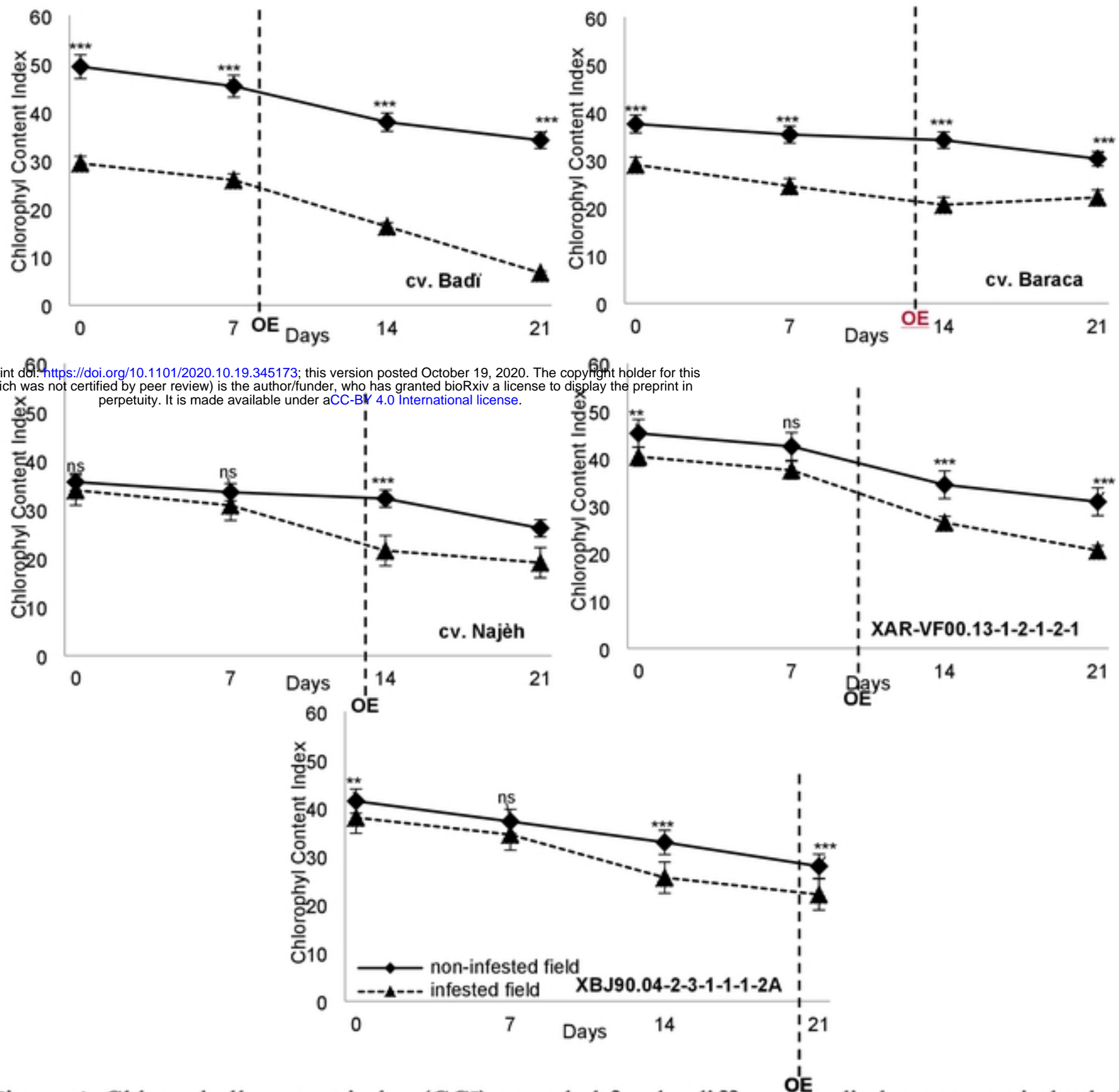


Figure 4: Chlorophyll content index (CCI) recorded for the different studied genotypes in both *O. foetida* infested and non-infested fields during the cropping season 2010/2011. OE: *Orobancha* Emergence. Data are fifteen replications means  $\pm$  SE. Data with the same letter(s) per are not significantly different at  $P=0.05$  (Duncan test).



bioRxiv preprint doi: <https://doi.org/10.1101/2020.10.19.345173>; this version posted October 19, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

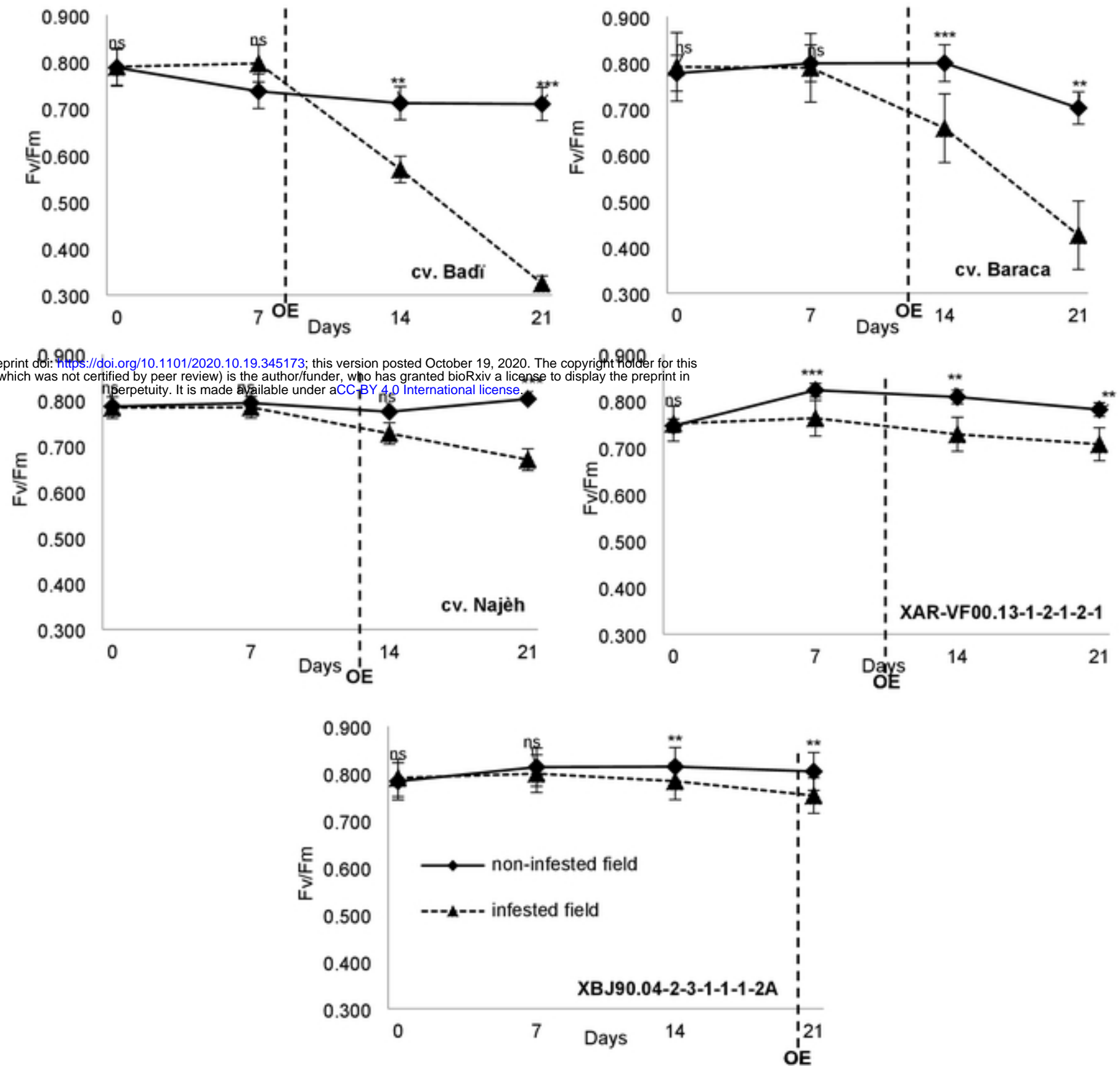


Figure 5:  $F_v/F_m$  ratio recorded for the different studied genotypes in both *O. foetida* infested and non-infested fields during the cropping season 2010/2011. OE: *Orobanchae* Emergence. Data are 6 replications means  $\pm$  SE. ns: non significantly different at  $P \geq 0.05$ ; \*\*\*: significantly different at  $P \leq 0.01$ ; \*\*: significantly different at  $P \leq 0.05$ .

## Tables list

**Table 1:** Origin and main characteristics of different studied genotypes

**Table 2:** Climatic data (monthly minimum, maximum and average temperature (°C) and rain (mm) recorded in Beja research station during the two cropping seasons 2009/2010 and 2010/2011.

**Table 3:** *Orobanche* incidence (%) and *Orobanche* severity (1-9), Number of Days to *Orobanche* Emergence (DOE) and number of Emerged *Orobanche* Shoots per plant (EOS) recorded for different studied genotypes in high *O. foetida* infested field during the cropping season 2010/2011.

**Table 4:** Biomass and water content (WC) recorded for different studied genotypes in both *O. foetida* infested and non-infested fields

bioRxiv preprint doi: <https://doi.org/10.1101/2020.10.19.345173>; this version posted October 19, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

Table 1: Origin and main characteristics of different studied genotypes

Genotypes	Origin	Main characteristics
XBJ90.04-6-2-1-1-4-C	Tunisia	Developed from a cross performed in Beja-Tunisia in 1990
XBJ90.04-2-3-1-1-1-2A	Tunisia	"
XBJ90.04-2-3-1-1-1	Tunisia	"
XBJ90.03-20-3-1	Tunisia	"
XBJ90.01-7-2-1-1-1-2-1-2	Tunisia	"
XBJ92-10-46-1-3-2-1-8-A	Tunisia	Developed from a cross performed in Beja-Tunisia in 1992
XBJ92-10-46-1-3-1-2-1-1-1-6-A	Tunisia	"
XBJ92.10-45-1-2-3-1-1	Tunisia	"
XBJ92.10-27-1-2-1-1-1	Tunisia	"
XBJ92.10-17-1-3-2-1-1	Tunisia	"
Syn1- (XBJ92-10-46-1-3)	Tunisia	"
XBJ93.12-10-1-1-3	Tunisia	Developed from a cross performed in Beja-Tunisia in 1993
XAR-VF00.13-8-5-1-2-1	Tunisia	Developed from cross performed in Ariana-Tunisia in 2000
XAR-VF00.13-8-3-2-2-2	Tunisia	"
XAR-VF00.13-8-3-2-2-1	Tunisia	"
XAR-VF00.13-8-3-1-2-1	Tunisia	"
XAR-VF00.13-8-3-1-1-1	Tunisia	"
XAR-VF00.13-8-1-2-1-2	Tunisia	"
XAR-VF00.13-81-1-3-2-1	Tunisia	"
XAR-VF00.13-81-1-3-1-1	Tunisia	"
XAR-VF00.13-7-5-2-2	Tunisia	"
XAR-VF00.13-70-1-1-1-1	Tunisia	"
XAR-VF00.13-5-3-1-1-1	Tunisia	"
XAR-VF00.13-4-5-1-1	Tunisia	"
XAR-VF00.13-4-2-1-1	Tunisia	"
XAR-VF00.13-31-7-2-1-1	Tunisia	"
XAR-VF00.13-31-4-2-2-1	Tunisia	"
XAR-VF00.13-27-5-1-2-1	Tunisia	"
XAR-VF00.13-27-2-2-1-1	Tunisia	"
XAR-VF00.13-27-2-1-1-4	Tunisia	"
XAR-VF00.13-2-5-1-3-1	Tunisia	"
XAR-VF00.13-24-2-2-1-2	Tunisia	"
XAR-VF00.13-2-4-1-1-1	Tunisia	"
XAR-VF00.13-2-2-2-3-2	Tunisia	"
XAR-VF00.13-2-1-1-4-1	Tunisia	"
XAR-VF00.13-2-1-1-1-1	Tunisia	"
XAR-VF00.13-1-2-1-2-1	Tunisia	"
Sel.88.Lat.18054-2-1-1	Tunisia	Originated from ICARDA
Badi	Tunisia	High yielding variety, released in Tunisia in 2004, susceptible to <i>O. foetida</i> and <i>O. crenata</i> .
Najeh	Tunisia	High yielding variety, released in Tunisia in 2009, developed from a cross performed in Beja-Tunisia.
Baraca	Spain	High yielding variety released in Spain. Derived from the line VF1071 (a selection from F402) as the original source of resistance to <i>O. crenata</i> .

bioRxiv preprint doi: <https://doi.org/10.1101/2020.10.19.345173>; this version posted October 19, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.



Table 2: Climatic data (monthly minimum, maximum and average temperature (°C) and rain (mm) recorded in Beja research station during the two cropping seasons 2009/2010 and 2010/2011.

Cropping season	Temp./Rain	Sep	Oct	Nov	Dec	Jan	Feb	Mar	Apr	May	Jun	Avg	Total
2009-10	Temp. min	17.7	14.1	8.1	7.8	6.6	5.9	6.9	10.4	11.7	15.4	10.5	-
	Temp. max	30.3	24.7	21.5	18.5	15.8	17.6	19.7	22.9	26.7	32.1	23.1	-
	Rain (mm)	89.5	59.4	47.4	64.2	10.7	67.1	78	46.4	27.2	4.8	-	494.7
2010-11	Temp. min	17.3	13.6	10.7	6.2	5.6	5.1	6.4	9.5	12.3	15.2	10.2	-
	Temp. max	30.7	26.7	20.8	17	15.9	15.5	18.3	23.5	26.4	31.5	22.6	-
	Rain (mm)	42.7	82	56.8	61.6	63.2	138.4	58.3	38.6	43.4	7.4	-	592.4

bioRxiv preprint doi: <https://doi.org/10.1101/2020.10.19.345173>; this version posted October 19, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.

Table 3: *Orobanche* incidence (%) and *Orobanche* severity (1-9), Number of Days to *Orobanche* Emergence (DOE) and number of Emerged *Orobanche* Shoots per plant (EOS) recorded for different studied genotypes in high *O. foetida* infested field during the cropping season 2010/2011.

	<i>Orobanche</i> incidence (%)	<i>Orobanche</i> severity (1-9)	Number of days to <i>Orobanche</i> emergence	Emerged <i>Orobanche</i> shoot per plant at harvesting
Badi	100.0±0 <sup>c</sup>	6.3±1.2 <sup>b</sup>	133.0±2.6 <sup>a</sup>	2.7±0.6 <sup>c</sup>
Baraca	76.7±25.2 <sup>bc</sup>	4.3±1.2 <sup>a</sup>	139.7±6.1 <sup>ab</sup>	2.6±1.4 <sup>b</sup>
Najeh	50.0±30 <sup>b</sup>	3.0±0 <sup>a</sup>	141.7±5 <sup>b</sup>	1.2±0.6 <sup>a</sup>
XAR-VF00.13-1-2-1-2-1	70.0±17.3 <sup>bc</sup>	4.3±1.2 <sup>a</sup>	140.3±3.1 <sup>bc</sup>	1.4±0.4 <sup>a</sup>
XBJ90.04-2-3-1-1-1-2A	13.3±5.8 <sup>a</sup>	3±0.0 <sup>a</sup>	145±2 <sup>a</sup>	0.9±0.7 <sup>a</sup>

Values followed by the same letter column are not significantly different at p=0.05 according to Duncan's multiple range mean comparison test

Table 4: Biomass and water content (WC) recorded for different studied genotypes in both *O. foetida* infested and non-infested fields

	Non-infested field		<i>O. foetida</i> infested field	
	Biomass	WC	Biomass	WC
Badi	313.4±122.9 <sup>b</sup>	76,8±4,9 <sup>bc</sup>	98.7±67.4 <sup>a</sup>	70,3±16,3 <sup>a</sup>
Baroca	250,3±67,1 <sup>a</sup>	76,7±2,5 <sup>bc</sup>	151.9±78.9 <sup>ab</sup>	77,1±6,3 <sup>a</sup>
Najeh	219.6±124.1 <sup>a</sup>	73,4±3,9 <sup>a</sup>	185.3±128.1 <sup>b</sup>	76±10,3 <sup>a</sup>
XAR-VF00.13-1-2-1-2-1	218.2±119.7 <sup>a</sup>	74,1±5,5 <sup>ab</sup>	169.2±113.7 <sup>b</sup>	76±7,7 <sup>a</sup>
XBJ90.04-2-3-1-1-1-2A	164.2±63.4 <sup>a</sup>	78,2±2,6 <sup>c</sup>	129.4±73.1 <sup>b</sup>	76±11,4 <sup>a</sup>

Values followed by the same letter per column are not significantly different at p=0.05 according to Duncan's multiple range mean comparison test.

bioRxiv preprint doi: <https://doi.org/10.1101/2020.10.19.345173>; this version posted October 19, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY 4.0 International license.