

1 Metabolomic profiling identifies complex lipid species associated with response to weight loss
2 interventions

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21 Abstract

22 Obesity is an epidemic internationally. While weight loss interventions are efficacious, they are
23 compounded by heterogeneity with regards to clinically relevant metabolic responses. Thus, we
24 sought to identify metabolic pathways and biomarkers that distinguish individuals with obesity
25 who would most benefit from a given type of intervention. Liquid chromatography mass
26 spectrometry-based profiling was used to measure 765 metabolites in baseline plasma from
27 three different weight loss studies: WLM (behavioral intervention, N=443), STRRIDE-PD
28 (exercise trial, N=163), and CBD (surgical cohort, N=125). The primary outcome was percent
29 change in insulin resistance (as measured by the Homeostatic Model Assessment of Insulin
30 Resistance [% Δ HOMA-IR]) over the intervention. Overall, 92 individual metabolites were
31 associated with % Δ HOMA-IR after adjustment for multiple comparisons. Concordantly, the most
32 significant metabolites were triacylglycerols (TAGs; $p=2.3e-5$) and diacylglycerols (DAGs;
33 $p=1.6e-4$), with higher TAG and DAG levels associated with a greater improvement in HOMA-
34 IR. In tests of heterogeneity, 50 metabolites changed differently between weight loss
35 interventions; we found amino acids, peptides, and their analogues to be most significant ($4.7e-$
36 3) in this category. Our results highlight novel metabolic pathways associated with heterogeneity
37 in response to weight loss interventions, and related biomarkers which could be used in future
38 studies of personalized approaches to weight loss interventions.

39 Introduction

40 Obesity is a major epidemic in the developed world and is an increasing problem in developing
41 countries, with a range of consequences including dyslipidemia, hypertension, cardiovascular
42 disease (CVD), stroke, type 2 diabetes mellitus (T2DM), and overall mortality.[1–4] In the United
43 States, one third of adults are obese and approximately 300,000 deaths are attributed to obesity
44 every year.[5,6] Behavioral, pharmacologic, exercise, dietary, and surgical intervention methods
45 have been attempted to curb the obesity epidemic. Ideally, these interventions would be
46 effective in all adults equally; however even when accounting for the amount of weight loss,
47 individuals show heterogeneity in improvement in obesity-related co-morbidities and CVD risk
48 factors.[7] Further, compliance with obesity intervention protocols is low unless expensive and
49 protracted intervention programs run by a trained interventionist are employed. Surgical obesity
50 interventions can partially overcome compliance issues; however, these are costly, can have
51 short- and long-term complications, and are characterized by frequent weight regain. In
52 recognition of these issues, the American Heart Association (AHA), the American College of
53 Cardiology (ACC), and The Obesity Society (TOS) released a call for researchers to focus on
54 determining the “the best approach to identify and engage those who can benefit from weight
55 loss”.[8]

56
57 Blood-based biomarkers, by serving as more granular snapshots into an individual’s unique
58 biochemistry, could distinguish individuals who would benefit the most from surgical
59 interventions from those who can benefit from lower cost, less-invasive solutions. Our previous
60 work has demonstrated a clear disconnect between amount of weight loss during lifestyle
61 obesity interventions and improvement in insulin resistance[9], as well as marked inter-individual
62 heterogeneity in amount of weight loss and metabolic responses to a given weight loss
63 intervention. In this study, we investigate an omics-driven personalized approaches to

64 understanding the molecular mechanisms of obesity and identifying biomarkers of response
65 among diverse weight loss interventions including behavioral, exercise and surgical
66 interventions.

67

68

69 **Methods**

70 **Study Populations and Study approval.** The **WLM** cohort has been described previously[10]
71 (NCT00054925); all participants provided written consent. Approval from the Duke University
72 Institutional Review Board was given. Notably, overweight or obese adults with hypertension,
73 dyslipidemia, or both were recruited from clinical research centers at Duke University, Johns
74 Hopkins University, Pennington Biomedical Research Center, or the Kaiser Permanente Center
75 for Health Research between 2003 and 2009. This study only involves samples collected during
76 phase 1 of the WLM intervention in which all participants were involved in a group-based
77 behavioral intervention. A trained interventionist led 20 weekly group sessions with the goals for
78 participants to reach 180 minutes per week of moderate physical activity (e.g., walking), reduce
79 caloric intake, adopt the Dietary Approaches to Stop Hypertension (DASH) dietary pattern, and
80 lose approximately ~0.5-1 kg per week. DASH was chosen because it reduces CVD risk
81 factors.[11–16] Only participants who lost at least 4 kg during the 6-month weight loss program
82 were considered for this study. Relevant for this study, blood plasma was drawn at baseline and
83 6-month follow-up, which we used for non-targeted metabolomics.

84

85 The **STRIDE-PD** study[17,18] (NCT00962962) compared three 6-month exercise-only groups
86 between 2009 and 2013; differing in amount or intensity to a fourth lifestyle intervention group:
87 diet plus exercise similar to the first 6-months of the Diabetes Prevention Program (DPP). All
88 participants in STRIDE-PD provided written consent and approval from the Duke University
89 Institutional Review Board was given. Sedentary, moderately overweight/obese ($25 < \text{BMI} < 35$
90 kg/m^2), nonsmoking adults between the ages of 45 and 75 years with prediabetes, but no
91 history of diabetes mellitus (T2DM) or CVD, were randomly assigned to one of four groups. The
92 STRIDE-PD study defined prediabetes as high-normal to impaired fasting glucose (95-125
93 mg/dL). The four groups were: 1) low-amount/moderate-intensity exercise; 2) high-

94 amount/moderate-intensity exercise; 3) high-amount/vigorous-intensity exercise; 4) diet and
95 exercise clinical lifestyle intervention. Relevant for this study, the blood plasma which we used
96 for non-targeted metabolomics was drawn at baseline and 6-month follow-up.

97

98 The **CBD** cohort was a surgical weight loss cohort of individuals who underwent either Roux-en-
99 Y gastric bypass surgery (RYGB) or adjustable gastric banding surgery (AGB) at St Luke's
100 Roosevelt Hospital Center between 2006 and 2014. All participants signed a consent form prior
101 to engaging in various research studies aiming to identify changes in gut hormones and
102 metabolism after bariatric surgery (NCT01516320, NCT02287285, NCT02929212,
103 NCT00571220) [19–23]. Approval from the Columbia University Institutional Review Board was
104 given. As such, each participant had fasting blood plasma drawn at baseline, with body weight
105 and HOMA-IR measured at multiple follow-up visits up to a year.

106

107 **Metabolomic profiling.** Four complementary liquid chromatography tandem mass
108 spectrometry (LC-MS)-based methods were used to measure plasma lipids and polar
109 metabolites as previously described [24–28]. The methods are characterized by the
110 chromatography stationary phase and MS ionization mode used and are referred to as C8-pos,
111 C18-neg, HILIC-pos, and HILIC-neg. The C8-pos, C18-neg, and HILIC-pos methods were
112 configured on LC-MS systems comprised of Nexera X2 U-HPLCs (Shimadzu) coupled to Q
113 Exactive series orbitrap mass spectrometers (Thermo Fisher Scientific) for high resolution
114 accurate mass (HRAM) profiling of both hundreds of identified metabolites and thousands of
115 unknowns, while the HILIC-neg method was operated on both a Nexera X2-Q Exactive system
116 for HRAM profiling and a UPLC (Waters) coupled to a QTRAP 5500 (SCIEX) for targeted
117 profiling.

118

119 The **C8-pos** method measures polar and nonpolar lipids. Lipids were extracted from 10 μ L
120 plasma using 190 μ L of isopropanol, separated using reversed phase C8 chromatography, and
121 analyzed HRAM, full-scan MS in the positive ion mode. The **C18-neg** method measures free
122 fatty acids, oxidized fatty acids and lipid mediators, bile acids, and metabolites of intermediate
123 polarity; these metabolites were extracted from 30 μ L plasma using 90 μ L of methanol, then
124 separated using reversed phase C18 chromatography, and analyzed using HRAM, full-scan MS
125 in the negative ion mode. The **HILIC-pos** method measures amino acids, amino acid
126 metabolites, acylcarnitines, dipeptides, and other cationic polar metabolites; these metabolites
127 were extracted from 10 μ L plasma using 90 μ L of 25% methanol/75% acetonitrile, then
128 separated using hydrophilic interaction liquid chromatography (HILIC), and analyzed using
129 HRAM, full-scan MS in the positive ion mode. The **HILIC-neg** method measures sugars, organic
130 acids, purines, pyrimidines, and other anionic polar metabolites; these metabolites were
131 extracted from 30 μ L plasma using 120 μ L of methanol containing internal standards and
132 analyzed using either HRAM, full-scan MS in the negative ion mode or targeted multiple
133 reaction monitoring using the QTRAP 5500 triple quadrupole MS system.

134
135 Raw data from Q Exactive series mass spectrometers were processed using TraceFinder
136 software (Thermo Fisher Scientific) to detect and integrate as subset of identified metabolites
137 and Progenesis QI software (v 2.0, Nonlinear Dynamics) to detect, de-isotope, and integrate
138 peak areas from both identified and unknown metabolites. MultiQuant (SCIEX) was used to
139 integrate peak areas of metabolites measured using the QTRAP 5500. Identities of metabolites
140 were confirmed by matching measured retention times (RT) and mass-to-charge ratios (m/z) to
141 authentic reference standards. Since reference standards are not available for all lipids,
142 representative lipids from each lipid class were used to characterize RT and m/z ratio patterns.
143 Lipid identities are reported at the level of lipid class, total acyl carbon content, and total double
144 bond content since the LC-MS method does not discretely resolve all isomeric lipids from one

145 another. Unknown features (unnamed metabolites) were not used in these analyses.

146 Coefficients of variation (CVs) and missingness are reported in Supplemental Tables 1 & 2 for
147 each metabolite.

148

149 **Statistical analysis.** Percent change in Homeostatic Model Assessment of Insulin Resistance
150 (HOMA-IR) over the intervention time period is the outcome used to represent metabolic health
151 for this study. HOMA-IR is calculated from clinically determined blood glucose and insulin levels
152 as previously described.[10,11] Individuals were excluded if HOMA-IR percent change was
153 implausible, i.e., greater than five standard deviations from the cohort mean (N=2 removed).
154 Metabolites measured at baseline in blood plasma by LC-MS are in the form of the natural log of
155 the MS peak's area under the curve (AUC). The primary study was association of baseline
156 metabolites with percent change in HOMA-IR over the obesity intervention time period. The
157 secondary study was of heterogeneity between cohorts in the primary study.

158

159 **Within cohort analysis.** Two statistical analyses were performed for this study within each of
160 the three cohorts. 1) A univariate linear association model between percent change of HOMA-IR
161 and baseline metabolite, in order to find metabolites that effect HOMA-IR. 2) The same model
162 as before with the addition of covariates for age, sex, race, baseline clinical triglycerides, and
163 percent change in weight over the follow-up time period in order to determine if the individual
164 metabolite effects HOMA-IR independent of traditional clinical measures known to be
165 associated with insulin resistance.

166

167 **Meta-analysis.** For both the primary univariate model and the full model, the three cohorts then
168 were meta-analyzed together using an inverse-variance weighted random effects model
169 implemented in R's meta library.[29] To account for the multiple tests, a false discovery rate
170 (FDR) correction was applied to the random effects p-values from the meta-analysis of the three

171 cohorts.[30] A Cochran q-test of heterogeneity also was performed to find metabolites having
172 different effects between the three cohorts.[31] Metabolites were filtered for only known
173 metabolites with less than 25% missingness after meta-analysis (N=199 removed).

174

175 **Metabolite set analysis.** We used a variation of the Gene Set Enrichment Analysis (GSEA)
176 method [32,33] to determine in an unbiased manner if any particular group of metabolites was
177 overrepresented at the beginning or end of a sorted list of our 765 metabolite results. Lists of
178 metabolite results were either sorted by z-score from the random effects meta-analysis (primary
179 analysis) or by Cochran q-test statistic from the heterogeneity analysis (secondary analysis).
180 The goal of GSEA is to determine whether the metabolites are randomly distributed throughout
181 this sorted list or mainly found at the beginning or end by performing a random walk along the
182 list and recording the maximum enrichment score (ES) along the way. Metabolite sets
183 associated with the outcome will have positive or negative ES depending on the direction of
184 effect; while metabolite sets unrelated to the outcome will have ES near zero. The 22 sets from
185 HMDB's taxonomic sub-classification of metabolites were used, with only sets with at least five
186 metabolites in our data were tested. P-values were determined by one million permutations and
187 FDR multi-test correction.

188

189 **Results**

190

191 **Study Populations.** This study includes three intervention study cohorts: Weight Loss

192 Maintenance (WLM) cohort, Studies of Targeted Risk Reduction Interventions through Defined

193 Exercise in individuals with Pre-Diabetes (STRRIDE-PD), and Columbia Bariatric and Diabetes

194 (CBD) cohort. Table 1 describes the demographics of these three cohorts.

195

196 Table 1: Baseline Characteristics of the Study Populations

Clinical Characteristics	WLM	STRRIDE-PD	CBD
N	443	163	125
Female (%)	62%	63%	82%
Race (% AA)	37%	18%	50%
Race (% EA)	62%	78%	49%
Age (years)	56 ± 8.7	59 ± 7.5	42 ± 10
Weight (kg)	96 ± 16	86 ± 12	121 ± 22
Weight Loss (kg)	-8.5 ± 3.8	-2.7 ± 4.0	-35 ± 16
BMI (kg/m ²)	34 ± 4.7	30 ± 2.8	44 ± 5.8
HOMA-IR	2.4 ± 1.6	2.0 ± 1.5	9.6 ± 5.8
HOMA-IR, Percent Change (%)	-16 ± 105	-16 ± 42	-71 ± 21
Triglycerides (mg/dL)	127 ± 71	123 ± 64	158 ± 108
Total Cholesterol (mg/dL)	185 ± 35	190 ± 32	187 ± 39
HDL Cholesterol (mg/dL)	43 ± 14	46 ± 14	48 ± 12
LDL Cholesterol (mg/dL)	110 ± 27	119 ± 27	108 ± 31

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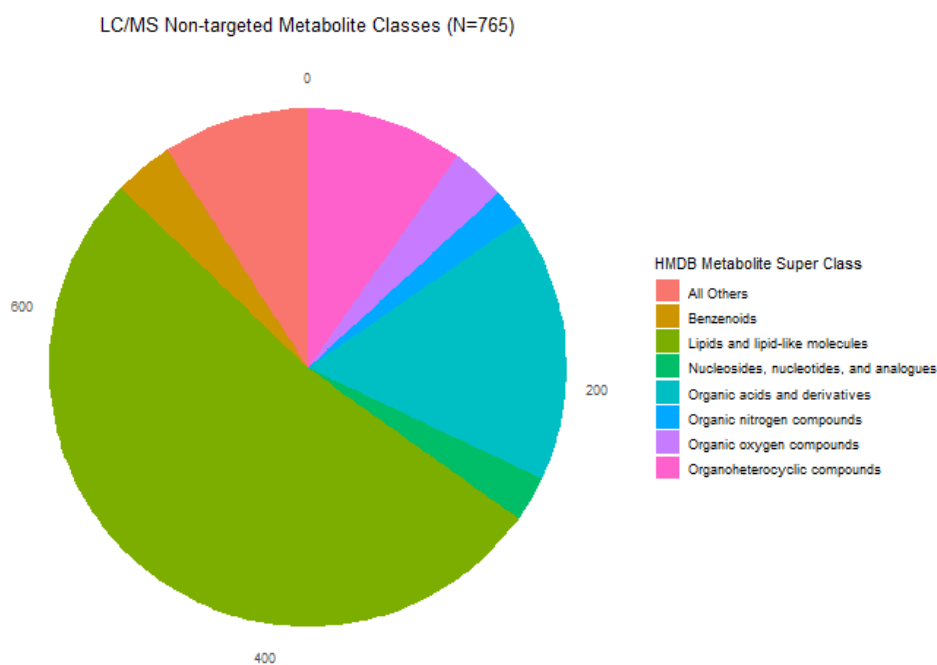
198 Unless otherwise stated, measures are at baseline timepoint. AA: African American. EA:
199 European Ancestry. BMI: body mass index. HOMA-IR: Homeostatic Model Assessment of
200 Insulin Resistance. HDL: high density lipoprotein cholesterol. LDL: low density lipoprotein
201 cholesterol. Weight loss is measured over the follow-up period. Values are listed as mean \pm
202 standard deviation. “cm” is centimeters. “kg” is kilograms. “m” is meters. “mg” is milligrams. “dL”
203 is deciliters. “%” is percent.

204

205 **Metabolomic profiling.** Four complementary liquid chromatography tandem mass
206 spectrometry (LC-MS)-based methods were used to measure plasma lipids and polar
207 metabolites. Figure 1 shows the Human Metabolome Database’s (HMDB’s)[34–37] superclass
208 and subclass taxonomic identifications for the 765 known metabolites that were identified from
209 all LC-MS methods in this study. Note that HMDB IDs ascribed to lipids are representative of
210 one or more isomers sharing the same chemical formula.

211

212 Fig. 1: LC-MS Non-targeted Metabolite Classes



213

214 Pie graph of the Human Metabolome Database's (HMDB's) superclass taxonomic identifications
215 for the 765 known metabolites from all LC-MS methods in this study. Nearly half of known
216 metabolites were lipids or lipid-like molecules shown in forest green color.

217

218 **Association of baseline metabolites with percent change in HOMA-IR.** Meta-analysis of the
219 univariate linear regression models in the three weight loss intervention cohorts identified 92
220 baseline metabolites (Figure 2, Supplemental Table 1) associated with percent change in
221 HOMA-IR after adjustment for multiple comparisons (FDR<5%). The top metabolites were
222 dominated by triacylglycerol (TAG) and diacylglycerol (DAG) species (in light-orange and red,
223 respectively, in Figure 2) and are listed in Table 2. As evidenced by the negative beta
224 coefficient, higher baseline levels of metabolites at baseline were associated with greater
225 reduction in HOMA-IR. For instance, the top result is C34:0 DAG with a FDR adjusted p-value of
226 5.42e-6 and an effect size of -36.08% change in HOMA-IR over the intervention. To determine if
227 an individual metabolite had an effect on HOMA-IR independent of traditional clinical measures
228 known to be associated with insulin resistance, we looked for nominal significance in the full
229 model. In the full model adjusted for age, sex, race, baseline triglycerides, and percent change
230 in weight over the follow-up time period, 90 of the 92 significant metabolites retained the same
231 direction of effect, 38 of these remained nominally associated with percent change in HOMA-IR
232 (Supplemental Table 2). Table 2 compares the effect size estimates of the univariate model with
233 the full model for the top 10 metabolites, demonstrating significance and consistency of
234 magnitude and directionality of effect.

235

236 Table 2: Comparison of effect sizes between univariate and full models

Metabolite	Univariate Model				Full Model		
	Pvalue_FDR	N	Pvalue	Beta	Beta	Pvalue	N
C34:0 DAG	5.42E-06	580	1.44E-08	-36.08	-25.03	8.90E-03	571
C50:1 TAG	5.42E-06	677	1.92E-08	-14.55	-9.21	4.42E-03	661
C30:0 DAG	1.19E-05	676	8.83E-08	-9.02	-5.26	1.67E-01	660
C43:1 TAG	1.19E-05	662	9.07E-08	-5.35	-3.37	1.80E-01	646
C48:0 TAG	1.19E-05	677	1.26E-07	-7.80	-4.79	2.32E-03	661
C50:2 TAG	1.19E-05	677	1.16E-07	-14.34	-8.11	2.87E-02	661
C56:1 TAG	1.23E-05	677	1.52E-07	-8.31	-6.23	5.36E-02	661
C51:2 TAG	1.81E-05	677	2.56E-07	-17.08	-11.22	1.71E-03	661
C45:2 TAG	2.91E-05	677	4.63E-07	-7.89	-4.53	2.51E-02	661
C46:1 TAG	3.06E-05	677	5.40E-07	-7.09	-3.69	3.82E-02	661

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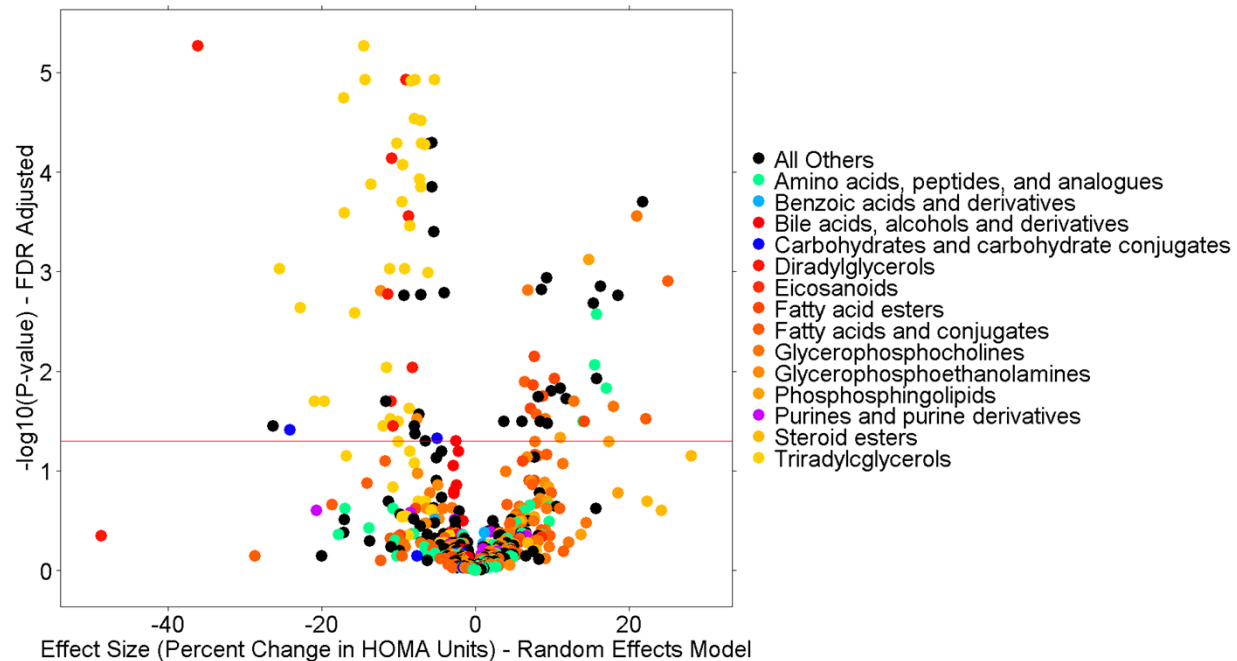
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Comparison of univariate model individual metabolite analysis (the primary analysis) to the full model conditional analyses where traditional clinical measures of age, sex, race, baseline clinical triglycerides, and percent change in weight over the follow-up time period were added to the model. The lack of major changes in effect sizes between the two models indicates the individual metabolites identified in the primary analysis provide additional clinical value over the traditional clinical measures. “Metabolite” is the common name for the metabolite. “N” is the number of individuals in the meta-analysis model. “Pvalue” is the random-effects p-value for the model. “Pvalue_FDR” is the primary p-value after FDR multi-test correction. “Beta” is the random-effects effect size estimate for the model. Pvalues are colored green if < 0.05 and Betas and colored blue to red for being high or low, respectively.

Fig. 2: Meta-analysis of the Univariate Model



250 Effect Size (Percent Change in HOMA Units) - Random Effects Model

251 Volcano plot of the main analysis results from meta-analysis of the three cohorts for the 765

252 known metabolites from all LC-MS methods in this study. The x-axis is the random-effects

253 effect-size estimate in percent change of HOMA-IR units (e.g., “-40” indicates a 40% drop in

254 HOMA-IR over the intervention time period). Each dot is a metabolite colored by the Human

255 Metabolome Database’s (HMDB’s) subclass taxonomic identification. The y-axis is the –

256 $\log_{10}(\text{P-value})$ from the random effects meta-analysis of the three cohorts in this study after

257 FDR correction.

258

259 **Metabolite set analysis to identify enriched biological pathways.** We used metabolite set

260 analysis to determine in an unbiased manner if any particular group of metabolites was

261 overrepresented within the HMDB’s taxonomic sub-classification of metabolites. Figure 3 shows

262 the enrichment plots for the top five sub-classification of metabolites most associated with

263 increased in insulin resistance followed by the top five sub-classification of metabolites most

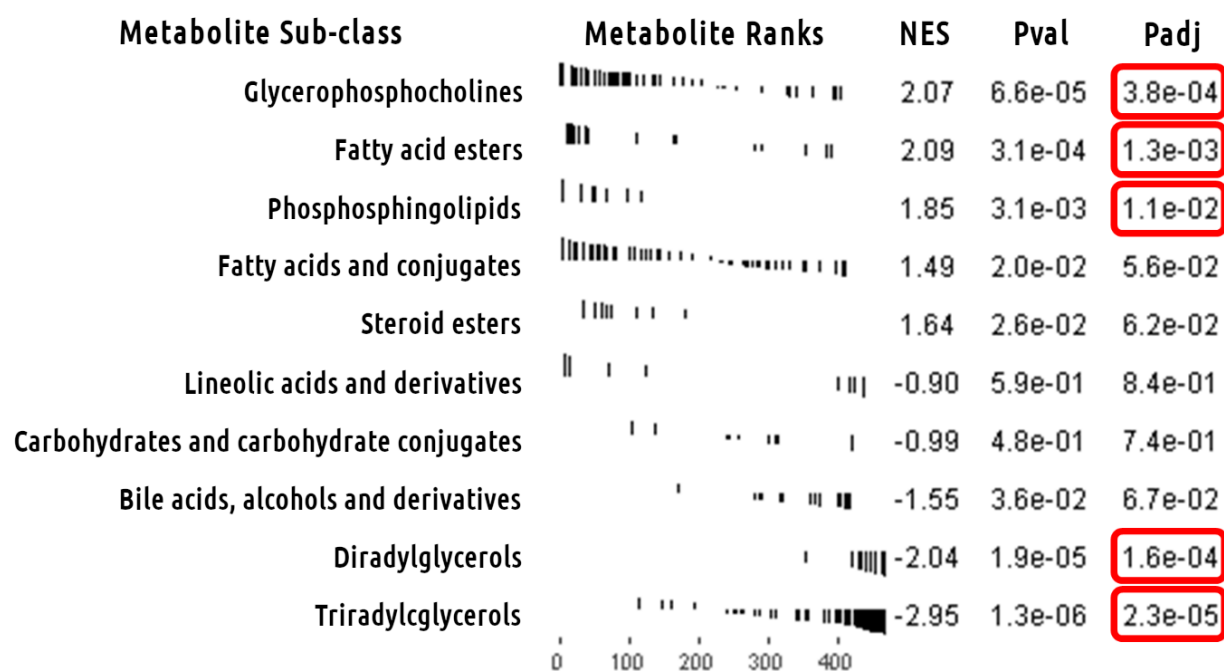
264 associated with reduction in insulin resistance. TAGs (FDR adjusted $p=2.3e-5$) and DAGs (FDR

265 adjusted $p=1.6e-4$) were the most significant sub-classifications associated with reduction in

266 insulin resistance over the intervention period in all 3 cohorts after FDR correction for the 22
 267 sub-classifications tested. In fact, both TAGs and DAGs are associated with reduction in insulin
 268 resistance over the intervention period in this regard. The “Glycerophosphocholines” (FDR
 269 adjusted $p=3.8e-4$), “Fatty acid esters” (FDR adjusted $p=1.3e-3$), and “Phosphosphingolipids”
 270 (FDR adjusted $p=1.1e-2$) sub-classifications were also significantly associated with insulin
 271 resistance after FDR multi-test correction; however, they were associated with increased insulin
 272 resistance over the intervention period. Supplemental Table 3 contains results from all tested
 273 HMDB sub-classifications.

274

275 Fig. 3: Metabolite Set Analysis of Univariate Model Meta-analysis



276

277 Main metabolite set analysis results for Human Metabolome Database’s (HMDB’s) subclass
 278 taxonomic identification of metabolites. Enrichment plots for the top 5 HMDB subclasses
 279 associated with increased insulin resistance followed by the top 5 HMDB subclasses associated
 280 with reduction in insulin resistance. The x-axis is the metabolome sorted by meta-analysis z-
 281 score. The y-axis has a black line for each hit in the *a priori* defined set of metabolites with

282 length equal to the meta-analysis z-score. Triacylglycerols stand out as most significant overall
283 HMDB subclass and most significantly associated with reduction in insulin resistance. “NES” is
284 normalized enrichment score. “Pval” is the p-value for the test. “Padj” is the same p-value after
285 FDR multi-test correction.

286

287 **Metabolites showing heterogeneity of effect between different weight loss interventions.**

288 In analyses designed to determine which metabolites at baseline associate with heterogeneity in
289 percent change in HOMA-IR between different weight loss intervention types, we performed a
290 Cochran q-test of heterogeneity, with a linear correction for the co-variables of age, sex, race,
291 baseline clinical triglycerides, and change in weight over the intervention time period. This type
292 of analysis is designed to find individual metabolites that can be added to a traditional clinical
293 model to aid in determining which of available obesity intervention a patient should be assigned
294 to. These analyses identified 50 metabolites (Supplemental Table 2) with nominally significant
295 p-values ($p < 0.05$) for heterogeneity by weight loss intervention cohort. Table 3 shows results for
296 the top 10 metabolites from these analyses. The top metabolite, for example, was N6,N6-
297 dimethyllysine (meta-analysis $p = 1.14e-4$), which demonstrated an estimated effect size in
298 percent change of HOMA-IR units of 14.63% in the CBD surgical cohort vs -10.57% in the
299 STRRIDE-PD exercise cohort and -2.04% in the WLM behavioral cohort, suggesting that
300 participants with higher baseline levels of this metabolite have a greater improvement of insulin
301 sensitivity in response to exercise than to surgical weight loss. Figure 4 shows a volcano plot of
302 these results. Unlike the findings in the primary analysis, no major HMDB classification
303 dominates the top results.

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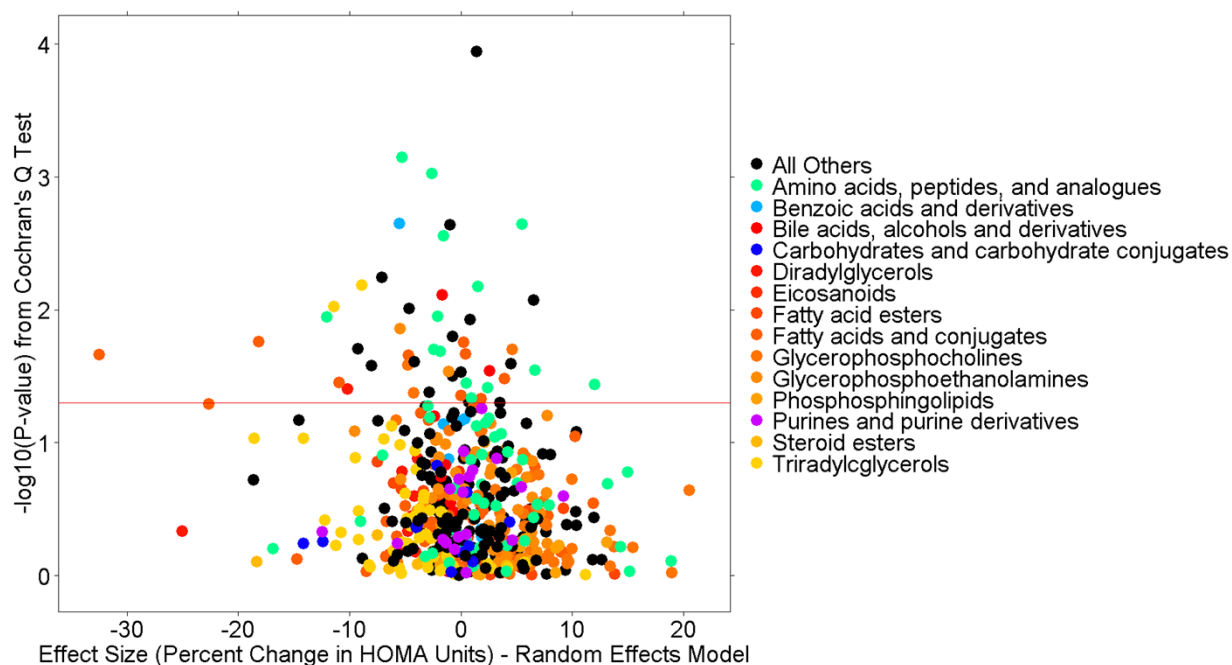
305 Table 3: Top 10 Metabolites Heterogeneous in Effect among Interventions from Full Model

Metabolite	N	Beta	Q_Pvalue	HMDB Sub-class
N6,N6-dimethyllysine	662	1.41	1.14E-04	
N-acetyltryptophan	662	-5.30	7.13E-04	Amino acids, peptides, and analogues
proline	662	-2.57	9.40E-04	Amino acids, peptides, and analogues
gentisate	619	-5.51	2.25E-03	Benzoic acids and derivatives
ADMA	662	5.53	2.28E-03	Amino acids, peptides, and analogues
2-aminoheptanoate	643	-0.96	2.30E-03	
phenylacetylglutamine	662	-1.54	2.79E-03	Amino acids, peptides, and analogues
tryptophan	662	-7.11	5.70E-03	Indolyl carboxylic acids and derivatives
C52:0 TAG	661	-8.87	6.56E-03	Triradylglycerols
NMMA	662	1.56	6.71E-03	Amino acids, peptides, and analogues

306
 307 Top 10 metabolites from the heterogeneity analysis of percent change in HOMA-IR. HOMA-IR is
 308 Homeostatic Model Assessment of Insulin Resistance. Beta is the random-effects effect-size
 309 estimate in percent change of HOMA-IR units (e.g. "-40" indicates a 40% drop in HOMA-IR
 310 over the intervention time period). Q_Pvalue is the p-value from the Cochran Q-test of
 311 heterogeneity. Subclass is the Human Metabolome Database's (HMDB's) subclass taxonomic
 312 identification for the metabolite.

313

314 Fig. 4: Heterogeneity Volcano Plot from Full Model Meta-analysis



315

316 Volcano plot of the heterogeneity analysis results from meta-analysis of the three cohorts for the
317 765 known metabolites from all LC-MS methods in this study. The x-axis is the random-effects
318 effect-size estimate in percent change of HOMA-IR units (e.g., “-40” indicates a 40% drop in
319 HOMA-IR over the intervention time period). Each dot is a metabolite colored by the Human
320 Metabolome Database’s (HMDB’s) subclass taxonomic identification. The y-axis is the –
321 $\log_{10}(\text{P-value})$ from the Cochran Q-test of heterogeneity.

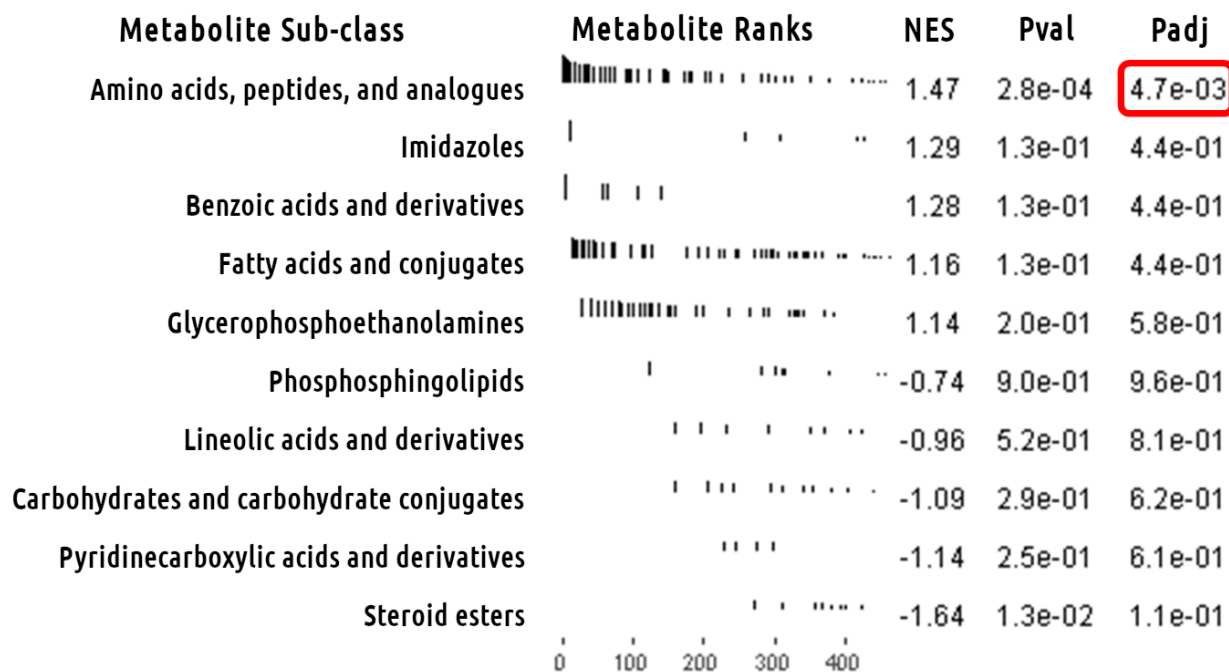
322

323 **Metabolite set analysis for metabolites associated with heterogeneity of effect among**

324 **weight loss interventions.** Metabolite set analysis was again performed, this time for the set of
325 metabolites associated with heterogeneity in response to intervention using the Cochran q-test
326 statistic. Figure 5 shows the enrichment plots for the top five most heterogeneous followed by
327 the five least heterogeneous sub-classification of metabolites. After FDR multi-test correction for
328 the 22 sub-classifications tested, only the metabolite set “Amino acids, peptides, and
329 analogues” was significant (FDR adjusted $p=4.7e-3$). This result is not clear from single
330 metabolite analysis and therefore shows the power of metabolite set analyses like this one.
331 Supplemental Table 4 contains results from all tested HMDB sub-classifications.

332

333 Fig. 5: Heterogeneity Metabolite Set Analysis of Full Model Meta-analysis



334

335 Heterogeneity metabolite set analysis results for Human Metabolome Database's (HMDB's)

336 subclass taxonomic identification of metabolites. Enrichment plots for top 5 most heterogeneous

337 and top 5 least heterogeneous HMDB subclasses between the three cohorts in this study. The

338 x-axis is the metabolome sorted by heterogeneity Q-score. The y-axis has a black line for each

339 hit in the *a priori* defined set of metabolites with length equal to the heterogeneity Q-score. The

340 amino acids, peptides, and analogues subclass stand out as most significantly heterogeneous

341 HMDB subclass. "NES" is normalized enrichment score. "Pval" is the p-value for the test. "Padj"

342 is the same p-value after FDR multi-test correction.

343

344 Discussion

345 In the first-of-its-kind study using a comprehensive metabolomics platform in three large
346 cohorts, we have identified metabolites that, measured at baseline, were associated with
347 improvements in insulin resistance, an important metabolic health measure, across behavioral,
348 exercise and surgical weight loss interventions in individuals with obesity. Specifically, we found
349 that higher baseline levels of complex triglyceride lipid species, triacylglycerols and
350 diacylglycerols, are associated with a more salutatory metabolic response across weight loss
351 interventions. Perhaps more importantly, we identify metabolites that were associated with
352 heterogeneity in improvement in insulin resistance depending on the type of weight loss
353 intervention. Specifically, we found 14 amino acids, peptides, and analogues, measured at
354 baseline, that were associated with differential response to weight loss intervention (N-
355 acetyltryptophan, proline, ADMA, phenylacetylglutamine, NMMA, phenylacetylglutamine,
356 tyrosine, hydroxyproline, N-alpha-acetylarginine, N6-acetyllysine, betaine, histidine, 2-
357 aminooctanoate, and lysine; Supplemental Table 2). These metabolites have great potential in
358 precision medicine for overweight/obese individuals, serving as baseline biomarkers that add to
359 clinical models of metabolic response to weight loss interventions, and to help guide a
360 personalized approach to weight loss intervention.

361
362 Most dietary fat is TAGs, which need to be broken down before absorption in the gut, then
363 reassembled into circulating low and high-density lipoproteins (LDL/HDL). High levels of TAGs
364 have been associated with atherosclerosis and stroke.[38] DAGs are precursors to TAGs that
365 have themselves be associated with immune-independent mechanisms of developing insulin
366 resistance and/or T2DM in muscle and liver tissues.[39,40]

367

368 Previous work has indicated that individual TAGs with lower carbon number (44-52 vs 54-60
369 carbons) and separately lower double-bond content (0-3 vs 4-12 double-bonds) are associated
370 with a higher likelihood of developing T2DM (higher carbon number and higher double-bond
371 content were neutral in effect towards likelihood of developing T2DM).[41] In the current study,
372 we find individual TAGs with carbon numbers between 50 and 55 and double-bond count
373 between two and three were associated with reduction in insulin resistance over an intervention
374 for obesity time period. Other TAG species were neutral in effect toward insulin resistance or
375 slight reduction in insulin resistance (Supplemental Figure 1 and 2). This may mean low carbon
376 number, saturated TAGs that indicated one may develop T2DM in the previous study[41] may
377 also indicate an obesity intervention will be less effective.

378

379 Of note, using a much less comprehensive metabolomic platform, we have previously observed
380 branched chain amino acids (BCAA) to be associated with insulin resistance and that higher
381 baseline levels are associated with a greater decrease in insulin resistance[42,43]. In this study,
382 we find amino acid analogues (including BCAAs) to be heterogeneous among our cohorts. In
383 the case of the individual BCAAs (valine, leucine, isoleucine), they show no association in our
384 CBD surgical cohort, while being positively associated with reduction in insulin resistance in
385 WLM and STRRIDE-PD. The low standard error in the CBD cohort causes the overall inverse-
386 variance weighted meta-analysis of the three cohorts to be non-significant. It is know that gut
387 bacteria can alter the bioavailability of BCAAs.[44] This may indicate that the microbiome is
388 important to consider during exercise/behavioral obesity interventions and less so during
389 surgical interventions perhaps due to antibiotic usage related to having a surgery.

390

391 Another item of note is although our top hit of N6,N6-dimethyllysine is relatively unknown in the
392 literature, our second most significantly heterogeneous amino acid analogues, N-
393 acetyltryptophan, demonstrated the same pattern as the BCAAs; it is also been shown to be

394 associated with host-gut microbiota interactions in both blood and urine bio-specimens.[45–47]
395 Namely that the amino acid tryptophan is modified into N-acetyltryptophan by the gut microbiota
396 and then absorbed into the host human. This adds to our earlier statement that the microbiome
397 is important to consider during exercise/behavioral obesity interventions and less so during
398 surgical interventions. We would point to antibiotic usage related to having surgery “resetting”
399 the microbiome after the baseline sample (used in this study to predict outcomes) has been
400 collected as a plausible reason for this.

401
402 While this is the first study to compare a large number of metabolites measured at baseline
403 across diverse weight loss interventions, the study has a few limitations. The included cohorts
404 are prospective, but are observational and therefore causation of metabolite pathways on insulin
405 resistance cannot be determined, although we note that these associations remained significant
406 after adjustment for amount of weight loss and other important comorbidities. Further, these
407 results highlight important metabolites that might be used in a prospective randomized
408 biomarker-guided clinical trial of assignment to different weight loss interventions. Our study
409 also primarily involves individuals of European and/or African ancestry and therefore has
410 unclear implications for other ancestries.

411
412 We believe this work demonstrates the validity and utility of evaluating the blood metabolome
413 when determining the proper obesity intervention for a patient in a precision medicine context.
414 Our work demonstrates the potential value of measuring individual TAGs and the differentiating
415 ability of amino acid analogues in deciding the best obesity intervention for an individual. Future
416 biomarker-guided intervention studies are necessary to determine clinical utility.

417

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422

423 Author contributions

424 NAB: Data curation, Formal analysis, Investigation, Methodology, Writing

425 LCK: Data curation

426 CBC: Methodology

427 AAD: Methodology

428 REG: Methodology

429 NJP: Data curation

430 BL: Conceptualization, Data curation

431 LPS: Conceptualization, Data curation

432 CBN: Data curation, Methodology

433 WEK: Conceptualization, Data curation

434 SHS: Conceptualization, Funding acquisition, Project administration, Resources, Supervision,
435 Writing

436

437 Conflict of interest statement

438 NAB, LCK, CBC, AAD, REG, BL and LPS have no conflicts. NJP has grants to institution from

439 Amgen and Regeneron/Sanofi; and performs consulting for Esperion. CBN, WEK and SHS

440 have an unlicensed patent on a related research finding (US10317414B2). The other authors

441 have declared that no conflict of interest exists.

442

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- 587
- 588

589 Supporting information captions

590 **Supplemental Table 1: HOMA-IR PC vs logMetabolite - Univariate Model - Analysis**

591 Includes all meta-analysis results for univariate model after filtering (see methods). Metabolite:
592 metabolite name, N_Samples: number of samples, N_Studies: number of cohorts that had data
593 for the metabolite (up to 3), Beta_Fixed: effect size from fixed effects meta-analysis, SE_Fixed:
594 standard error from fixed effects meta-analysis, Zvalue_Fixed: z-score from fixed effects meta-
595 analysis, Pvalue_Fixed: p-value from fixed effects meta-analysis, Beta_Random: effect size
596 from random effects meta-analysis, SE_Random: standard error from random effects meta-
597 analysis, Zvalue_Random: z-score from random effects meta-analysis, Pvalue_Random: p-
598 value from from random effects meta-analysis, Q: Cochran q-test statistic, Q_df: Cochran q-test
599 degrees of freedom, Q_Pvalue: Cochran q-test p-value, Tau2: tau-squared between-study
600 variance, H: H-statistic, I2: I-squared statistic, WLM_GP1_Beta: Effect size within WLM,
601 WLM_GP1_SE: standard error within WLM, WLM_GP1_Num: number of samples within WLM,
602 StrridePD_Beta: effect size within STRRIDE-PD, StrridePD_SE: standard error within
603 STRRIDE-PD, StrridePD_Num: Number of samples within STRRIDE-PD, CBD_Beta: effect size
604 within CBD, CBD_SE: standard error within CBD, CBD_Num: number of samples within CBD,
605 Method: LC-MS method used to measure metabolite, HMDB.ID...representative.ID.: HMDB ID
606 for metabolite, super_class: HMDB metabolite taxonomy super class, class: HMDB metabolite
607 taxonomy class, sub_class: HMDB metabolite taxonomy sub class, missingness: metabolite
608 missingness rate, zeroness: metabolite rate of being zero value, CV: metabolite coefficient of
609 variation, Pvalue_Adj_Fixed: FDR adjusted p-value from fixed effects meta-analysis,
610 Pvalue_Adj_Random: FDR adjusted p-value from random effects meta-analysis.

611

612

613

614 **Supplemental Table 2: HOMA-IR PC vs logMetabolite - Full Model - Analysis**

615 Includes all meta-analysis results for full model after filtering (see methods). Same columns as
616 Supplemental Table 1.

617

618 **Supplemental Table 3: GSEA HOMA-IR PC Taxonomy Subclass Results**

619 Includes all Metabolite Set Analysis of univariate model meta-analysis results for the 22
620 metabolite sets. Pathway: name of metabolite set, pval: p-value from GSEA test, padj: FDR
621 adjusted p-value, ES: enrichment score, NES: normalized enrichment score, nMoreExtreme:
622 number of 1 million permutation that were more extreme than data, size: number of metabolites
623 in metabolite set.

624

625 **Supplemental Table 4: GSEA HOMA-IR PC Heterogeneity Taxonomy Subclass Results**

626 Includes all Heterogeneity Metabolite Set Analysis of full model meta-analysis results for the 22
627 metabolite sets. Same columns as Supplemental Table 3.

628

629 **Supplemental Figure 1: HOMA-IR PC logMetabolite Analysis - Univariate Model -**

630 **Triradylclycerols - Carbon Downsloping Plot - Sig**

631 Plot of TAG metabolite effect size from random effects meta-analysis of univariate model vs
632 number of carbon atom in the TAG.

633

634 **Supplemental Figure 2: HOMA-IR PC logMetabolite Analysis - Univariate Model -**

635 **Triradylclycerols - Bond Downsloping Plot - Sig**

636 Plot of TAG metabolite effect size from random effects meta-analysis of univariate model vs
637 number of double bonds in the TAG.

638