

1 **New traps for the capture of *Aedes aegypti* (Linnaeus) and *Aedes albopictus* (Skuse)**

2 **(Diptera: Culicidae) eggs and adults**

3 **Traps for *Aedes aegypti* and *Aedes albopictus***

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28

29 **Abstract**

30 The control of arboviruses carried by *Aedes aegypti* (Linnaeus) and *Aedes albopictus* (Skuse)
31 can be performed with tools that monitor and reduce the circulation of these vectors. Therefore,
32 the efficiency of four types of traps in capturing *A. aegypti* and *A. albopictus* eggs and adults,
33 with the biological product Vectobac WG[®], was evaluated in the field. For this, 20 traps were
34 installed in two locations, which were in the South (Londrina, Paraná) and North (Manaus,
35 Amazonas) Regions of Brazil, from March to April 2017 and January to February 2018,
36 respectively. The UELtrap-E and UELtrap-EA traps captured *A. aegypti* and *A. albopictus* eggs:
37 1703/1866 eggs in Londrina, and 10268/2149 eggs in Manaus, respectively, and presented high
38 ovitraps positivity index (OPI) values (averages: 100%/100% in Londrina, and 100%/96% in
39 Manaus, respectively); and high egg density index (EDI) values (averages: 68/75 in Londrina,
40 and 411/89 in Manaus, respectively), so they had statistically superior efficiency to that of the
41 CRtrap-E and CRtrap-EA traps in both regions, that captured less eggs and adults: 96/69 eggs
42 in Londrina, and 1091/510 eggs in Manaus, respectively. Also presented lower OPI values
43 (averages: 28%/4% in Londrina, and 88%/60% in Manaus, respectively); and lower EDI values
44 (averages: 10.5/9 in Londrina, and 47/30 in Manaus, respectively). The capture ratios of *Aedes*
45 adults in the UELtrap-EA and CRtrap-EA traps in Londrina and Manaus were 53.3%/29.5%
46 and 0%/9.8%, respectively. UELtrap-E and UELtrap-EA can be adopted as efficient tools for
47 *Aedes* monitoring due to their high sensitivity, low cost and ease of use.

48

49 **Keywords:** Arbovirus vectors, dengue, monitoring, entomological surveillance, arboviruses
50 control, public health.

51 **Author summary**

52 *Aedes aegypti* and *Aedes albopictus* are species of mosquitoes responsible for the
53 transmission of several arboviruses that cause infections worldwide. However, there are still no
54 effective and safe vaccines or medications to prevent or treat arboviruses transmitted by these
55 vectors, except for yellow fever. Moreover, current methodologies for monitoring and
56 controlling *A. aegypti* and *A. albopictus* are not fully effective, as evidenced by the increasing
57 cases of the arbovirus transmitted by these mosquitoes or have incompatible costs with the
58 socioeconomic conditions of a large number of people. Thus, the traps tested in this study can
59 be used as more effective and economical tools for monitoring and controlling *A. aegypti* and
60 *A. albopictus*, since they are made with low cost material and they showed high efficiency in
61 the capture of eggs, evidenced by the high values of ovitraps positive index and eggs density
62 index, besides that one of the models captured *Aedes* spp. adults in both regions where they
63 were tested. Therefore, the traps have potential for reducing *Aedes* spp. eggs and adults in the
64 environment and sensibility for determining the local infestation index, which can be reconciled
65 with official government strategies for more accurate vector monitoring and control actions.

66

67 **Introduction**

68 Mosquitoes in the family Culicidae, order Diptera, occur in virtually all regions of the
69 planet. This family is divided into two subfamilies (Anophelinae and Culicinae) in which some
70 species are considered vectors of pathogens of medical importance [1], such as *Aedes*
71 (*Stegomyia*) *aegypti* (Linnaeus, 1762) and *Aedes* (*Stegomyia*) *albopictus* (Skuse, 1894)
72 (Diptera: Culicidae). These species are cosmopolitan and capable of becoming infected with
73 various arboviruses that are responsible for disease and death worldwide [2-5].

74 Although *A. aegypti* is of African origin, its incidence is currently higher in the
75 Americas, Southeast Asia, and the Western Pacific [4, 6, 7]. In Brazil, it is the main vector of

76 the four dengue serotypes (DENV-1, DENV-2, DENV-3, DENV-4) and the urban yellow fever
77 virus, which occurs throughout the Brazilian territory [2, 3, 8]. It also transmits Zika virus
78 (ZIKV) and chikungunya (CHIKV), which are responsible for infections and deaths in over 100
79 countries [3, 9-11].

80 This species has a home habit, with essentially anthropophile and synanthropic behavior
81 [2, 12-14]. Females prefer artificial containers with standing water for laying, such as tires,
82 disposable cups, potted plants and bottles, especially those of dark colors and with rough
83 surfaces [2, 15-17]. In these breeding sites, it is often also possible to find eggs of *A. albopictus*,
84 which originated from Asia, where it is the secondary vector of the dengue virus, which has
85 now spread to Africa, the Americas and Europe [3-5, 18].

86 On the American continent, this species has the potential to carry the same arboviruses
87 as *A. aegypti*, in addition to the ability to carry many other arboviruses in laboratory settings
88 [3-5, 19]. Currently, it has adapted to rural, suburban and urban spaces, with a preference for
89 urban spaces with greater vegetation coverage and near native or secondary forests [5, 20-22].

90 Tropical and subtropical countries, such as Brazil, are favorable for the proliferation of
91 vector mosquitoes, given the high temperatures and abundant rainfall. Economic and social
92 factors, such as the lack of basic sanitation and inadequate water supply in the peripheries of
93 large urban centers, also contribute to the availability of mosquito breeding sites and
94 consequently to the spread of viruses [6, 19, 23, 24].

95 The North Region of Brazil has consistently favorable conditions for *Aedes* spp.
96 proliferation since temperatures remain high throughout the year (annual average of 26 °C),
97 with high precipitation (2000 to 3000 mm annually) [25]. Despite having a mild climate (annual
98 average of approximately 22 °C) and well-defined seasons, South Region of Brazil has a
99 predominance of rains and high temperatures in the summer (average annual rainfall between

100 1500 and 2000 mm) [25], which combined with local structural conditions favor the
101 proliferation of *Aedes* spp.

102 Currently, there are still no safe and effective vaccines or medicines to prevent or treat
103 all the arboviruses carried by these vectors, except for yellow fever [26, 27]. Thus, measures
104 adopted to control these diseases must consist of actions to reduce vector circulation and,
105 consequently, viral circulation [26, 28]. However, the current methodologies for monitoring
106 and controlling *A. aegypti* and *A. albopictus* are not fully effective, as evidenced by the
107 increasing cases of the arbovirus transmitted by these mosquitoes, according to the Brazilian
108 Ministry of Health disclosures [29].

109 In this sense, the use of traps to capture the eggs of *A. aegypti* and *A. albopictus*, which
110 are called ovitrampas in Brazil, may be an important strategy for reducing vector circulation.
111 This tool is capable of promoting both monitoring of vectors as well as allowing the removal
112 of eggs from the environment, providing indices of indirect mosquito abundance and allowing
113 verification of their spatial and temporal distribution through the number of eggs collected [30-
114 34]. In addition, they have been recommended by the Brazilian Ministry of Health to assist in
115 the surveillance and control of *Aedes* spp. [35].

116 Ovitrapas can be optimized by using entomological glue to capture adults, attractive and
117 larvicidal [30, 32, 33, 36]. The grass infusion *Megathyrus maximus* Jacq is used as an effective
118 attractant; it acts as a potentiator for the effectiveness of the adult traps and egg traps [32, 33,
119 37, 38]. *Bacillus thuringiensis israelensis* (Bti) formulations, which is a spore-forming
120 entomopathogen bacterium, are attractive as well as larvicidal because this bacterium
121 synthesizes toxic proteins specific to culicid larvae [39-41].

122 However, the traps available on the market have incompatible costs with the
123 socioeconomic conditions in Brazil, as they are usually coupled with batteries or motors.
124 Therefore, it is necessary to implement traps that are easy to handle and present low cost to

125 public agencies. From this perspective, this study aimed to evaluate the efficiency of different
126 traps for oviposition and capture of *A. aegypti* and *A. albopictus* adults in field conditions in
127 South and North Regions of Brazil to validate new tools that can be effective and economical
128 for vector monitoring.

129

130 **Material and methods**

131 **Study area**

132 The study was conducted in localities situated in the states of Paraná and Amazonas,
133 South and North Regions of Brazil. In Paraná, the traps were installed around five buildings
134 located on the *Campus* (74000 m²) of the Federal Technological University of Paraná (UTFPR)
135 in Londrina city (University Restaurant - 23 ° 18'28.51 "S 51 ° 6'56.52" O; Block A - 23 °
136 18'28.24 "S 51 ° 6'54.04" O; Block B - 23 ° 18'27.40 "S 51 ° 6'54.34" O; Block P - 23 ° 18'27.21
137 "S 51 ° 6'50.77 "O; Block K - 23 ° 18'26.01" S 51 ° 6'48.77 "O). In Amazonas, the traps were
138 distributed at five points located at *Campus* I (255,736.49 m²) of the National Institute for
139 Amazônia Research (INPA) in Manaus city (Point 1 - 3 ° 5'47 "S 59 ° 59'10" O; Point 2 - 3 °
140 5'43 "S 59 ° 59'11" O; Point 3 - 3 ° 5'40 "S 59 ° 59'15" O; Point 4 - 3 ° 5'41 "S 59 ° 59'17 "O;
141 Point 5 - 3 ° 5'42" S 59 ° 59'15 "O).

142 **Trap Characteristics**

143 Four types of traps adapted from the originals [42, 43] were tested: i) UELtrap-E
144 (standard trap) for egg capture (black rounded plastic vase measuring 12 cm length x 11 cm
145 diameter, with a capacity of 750 mL), ii) UELtrap-EA for capture of eggs and adults (12 x 11
146 cm black rounded plastic vase that is 750 ml in volume, with side openings and a lid with a
147 tulle for ventilation on the top and contains a funnel coated with entomological glue), (iii)
148 CRtrap-E for egg capture (clear circular plastic container measuring 8 cm length x 9 cm
149 diameter, with capacity of 500 mL, contains a black cone with a rough part to facilitate

150 oviposition and egg adhesion) and (iv) CRtrap-EA for capturing eggs and adults (8 x 9 cm clear
151 circular plastic container, containing a roughened black outer cone and a lid associated with a
152 funnel coated with entomological glue).

153 The UELtrap-E and UELtrap-EA traps have a Duratree Eucatex® reed that measures
154 13 cm length x 3 cm width, positioned vertically with the rough surface facing upwards to
155 facilitate oviposition and egg adhesion. In contrast, the CRtrap-E and CRtrap-EA consisted of
156 8 cm length x 1 cm width brown plastic reeds with both smooth and rough surfaces that are
157 placed upright with the rough part facing the outside of the opening from the container.

158 **Collection of eggs and adults of *Aedes* under field conditions**

159 Egg and adult collection at the UTFPR *campus* was carried out for five weeks between
160 March and April (autumn season) of 2017. In contrast, at INPA *Campus* I, the collections were
161 carried out from January to February (rainy season) of 2018 for five weeks. In Londrina, the
162 temperature (°C), relative humidity (%) and rainfall values (mm) were obtained through the
163 Instituto Agronômico do Paraná [44], at 11.2 km from the study site, while in Manaus, these
164 data were obtained from the automatic meteorological station installed in Manaus (A101), in
165 the meteorological database for teaching and research (BDMEP) of the National Institute of
166 Meteorology [25].

167 At each collection point, four traps were installed. The sampling design consisted of the
168 installation of one trap of each type at ground level in an area that was sheltered from the sun
169 and rain, had little movement of people and animals, and was at a minimum distance of 20
170 meters from the other traps. Each trap was given 250 ml of water without chlorine and 50 ml
171 of solution containing *M. maximus* (0.11256%) [37] and the biological product Vectobac® WG
172 (0.00083%) (Active ingredient: *B. thuringiensis israelensis*), strain AM65-52, 37.4% w/w; Lot
173 No.: 267-853-PG; Date of manufacture: July 2016; Valent BioSciences Corporation - VBC).

174 The attractant solution (50 mL) used in the traps was obtained from a 50 mg/L dilution of the
175 biological product in 5 L of grass infusion (0.0050% and 0.6754%, respectively).

176 The reeds from UELtrap-E and UELtrap-EA were replaced every seven days and sent
177 to the Medical Entomology Laboratory of Londrina State University, and to the Biological
178 Control and Biotechnology of Malaria and Dengue Laboratory at INPA, where the eggs were
179 quantified with the aid of a 50X stereoscope microscope.

180 The eggs present on the plastic reeds and inside CRtrap-E and CRtrap-EA were
181 quantified *in situ* with the aid of a manual magnifying glass (10x) and double-sided tape for egg
182 removal since the reeds were not replaced. Adults collected with UELtrap-EA and CRtrap-EA
183 were removed with the aid of entomological forceps and stored in glass bottles containing
184 absorbent paper to preserve the integrity of the characteristics. Mosquitoes were counted and
185 identified at the species level using external morphological characters with the aid of
186 stereomicroscopy and the identification keys proposed by [1, 2, 45].

187 Collections in the different study sites were carried out using the Sisbio authorization:
188 23093 license.

189 **Data analysis**

190 After quantification of the collected eggs and adults, the OPI – ovitrap positivity index
191 ($OPI = N^{\circ} \text{ of positive traps} / N^{\circ} \text{ of examined traps} \times 100$ [46] and EDI – egg density index (EDI
192 $= N^{\circ} \text{ of eggs} / N^{\circ} \text{ of positive traps}$) [46] were calculated. The data were submitted to the Lilliefors
193 normality test (K samples) and then compared with the data obtained from the evaluated indices
194 (OPI and EDI). Student's t-test ($p < 0.05$) was used for the data with a normal distribution, and
195 the Mann-Whitney test ($p < 0.05$) was used for data that did not present normality. The BioEstat
196 version 5.3 statistical software for Windows [47] was used to assist in all data analysis.

197 The proportion of female *Aedes* spp. captures in the UELtrap-EA and CRtrap-EA traps
198 were also calculated. This proportion was obtained by calculating the ratio between the total

199 number of eggs and females of *Aedes* spp. collected by the two trap models and considering
200 that each female lays a minimum average of 50 eggs per laying, according to [48] and [49]
201 Thus, let X be the number of females needed to deposit the amount of eggs collected in the
202 traps as follows:

$$X = \frac{\text{number of eggs collected}}{\text{minimum average of eggs per laying}} \quad (1)$$

203 From this, the proportion of female captures of *Aedes* spp. (PC) of the traps is given by
204 the following equation:

$$PC = \frac{\text{number of females caught}}{X} .100 \quad (2)$$

205

206 **Results**

207 **Abiotic data recorded in both sampling regions**

208 At the UTFPR *Campus*, the average temperature was 22.6 °C (14.5-30.6 °C), the
209 average relative humidity was 71.5% (44-96%) and the total precipitation was 113.4 mm (0-
210 47.4 mm) throughout the sampling period. At INPA *Campus* I, the sampling period presented
211 an average temperature of 28.3 °C (20.4-35.7 °C), average relative humidity of 80.1% (57.7-
212 95.7%) and total precipitation of 379.6 mm (0-71.3 mm).

213 **Total eggs and adults of *Aedes* collected at the UTFPR *Campus* in Londrina, Paraná**

214 Considering that the traps were used exclusively for egg capture, it was observed that
215 the UELtrap-E traps were more efficient than the CRtrap-E traps (Fig 1). This result was
216 corroborated when analyzing the average number of eggs obtained for both, since the first
217 obtained an average (341) 18 times higher than the value (19) obtained in the second trap, thus
218 presenting a statistically significant difference between the respective values ($p = 0.0090$)
219 (Table 1).

220 The data obtained from the UELtrap-EA traps presented a higher number of eggs that
 221 from the CRtrap-EA traps (Fig 1). This result was also evident because the average egg number
 222 (373) obtained by the former was observed to be 27 times higher than the average egg number
 223 (14) acquired by the latter; therefore, the differences was statistically significant among the
 224 referenced values ($p = 0.0107$) (Table 1).

225
 226 **Fig 1. The total eggs laid by *Aedes* adults in each trap for five weeks from March to April**
 227 **2017 in Londrina, Paraná, Brazil.**

228
 229
 230 **Table 1. Average, maximum and minimum *Aedes* eggs in the different traps from March**
 231 **to April 2017 in Londrina, Paraná, Brazil.**

Traps	Average (\pm SD)	Maximum	Minimum
UELtrap-E	341 (86.9) ^A	466	222
CRtrap-E	19 (23.7) ^B	60	0
UELtrap-EA	373 (177.4) ^a	612	179
CRtrap-EA	14 (12.5) ^b	33	0

232 Legend: SD = standard deviation. Different letters in the same column indicate a statistically significant
 233 difference ($p < 0.05$) between the average number of eggs obtained for the trap that collects the same
 234 stage (eggs or eggs/adults) using Student's t-test or the Mann-Whitney test.
 235

236 Considering the number of eggs of *Aedes* spp. collected in each week of the field
 237 experiment, no significant difference ($p > 0.05$) was found between the average number of eggs
 238 acquired in the weeks analyzed in all traps tested (Table 2).

239
 240 **Table 2. The average and standard deviation of the *Aedes* obtained in each trap from**
 241 **March to April 2017 in Londrina, Paraná, Brazil.**

Traps	Collection Weeks				
	1°	2°	3°	4°	5°
UELtrap-E	44 \pm 35 ^a	69 \pm 83 ^a	93 \pm 79 ^a	70 \pm 63 ^a	65 \pm 37 ^a

CRtrap-E	2.4 ± 5^a	0^a	3.4 ± 7.6^a	12 ± 12^a	1.4 ± 1.9^a
UELtrap-EA	47 ± 38^a	122 ± 113^a	97 ± 43^a	38 ± 57^a	71 ± 41^a
CRtrap-EA	0^a	3 ± 7^a	7 ± 8^a	1 ± 3^a	3 ± 7^a

242 Legend: The same letters on the same row indicate that there was no statistically significant difference
243 ($p > 0.05$) among the average numbers of eggs obtained each week for all traps tested using Student's t-
244 test or the Mann-Whitney test.
245

246 We obtained an OPI value of 100% with the UELtrap-E traps during the collection
247 weeks (Table 3). These results were higher than the values obtained with the CRtrap-E traps
248 during the five-week period. In the latter trap type, the percentage of positivity varied
249 throughout the sampling period, with no eggs in the second week and a higher percentage in
250 the fourth week (0 to 60%). There was a statistically significant difference between the mean
251 OPI values obtained for each trap ($p = 0.0090$).

252 Considering the EDI values obtained per week in the UELtrap-E traps, lower and higher
253 values were found in the first and third weeks (44 and 93), respectively (Table 3). The EDI data
254 obtained by the CRtrap-E traps also varied throughout the sampling period, with the absence of
255 eggs in the second week and a higher quantity in the fourth week (0 and 20). When analyzing
256 the average of the EDI values obtained in UELtrap-E and CRtrap-E, a statistically significant
257 difference ($p = 0.0002$) was found due to the higher egg density found in the first model (Table
258 3).

259 The OPI values obtained from the UELtrap-EA traps were 100% in all weeks analyzed
260 (Table 3). However, for the CRtrap-EA traps, there was variation in the indices, with the
261 absence of eggs in the first week and a higher percentage in the third week (0 and 60%). A
262 significant difference was observed between the mean OPI values between the two trap types
263 tested ($p = 0.0009$) (Table 3).

264 Regarding the UELtrap-EA EDI values, the results obtained during the collections
 265 showed variations between the indices, with lower and higher values in the fourth and second
 266 weeks (36 and 122), respectively (Table 3). The EDI results obtained in the CRtrap-EA traps
 267 also showed variations throughout the sampling period, with the absence of eggs in the first
 268 week and higher values of eggs in the second and fifth weeks (15), respectively. The egg density
 269 in the UELtrap-EA traps was higher than that obtained in the CRtrap-EA traps, which was
 270 corroborated by the statistically significant difference ($p = 0.0154$) (Table 3).

271

272 **Table 3. Ovitrap positivity index (OPI) and egg density index (EDI) obtained per week**
 273 **in each trap from March to April 2017 in Londrina, Paraná, Brazil.**

Weeks	UELtrap-E		CRtrap-E		UELtrap-EA		CRtrap-EA	
	OPI (%)	EDI	OPI (%)	EDI	OPI (%)	EDI	OPI (%)	EDI
1°	100	44	20	12	100	47	0	0
2°	100	69	0	0	100	122	20	15
3°	100	93	20	17	100	97	60	11
4°	100	70	60	20	100	36	20	6
5°	100	65	40	3.5	100	71	20	15
Average	100 ^A	68 ^a	28 ^B	10.5 ^b	100 ^A	75 ^a	24 ^B	9 ^b

274 Legend: Different letters on the same row indicate a statistically significant difference ($p < 0.05$) between
 275 the mean OPI and EDI values obtained for the trap that collects the same stage (eggs or eggs/adults)
 276 using Student's t-test or Mann-Whitney test.

277

278

279 The UELtrap-EA traps captured 17 specimens; one *A. albopictus*, ten *A. aegypti* and six
 280 *Culex quinquefasciatus* Say, 1823. Regarding the percentage of adults collected from each
 281 species, 6%, 59% and 35% were found for the species *A. albopictus*, *A. aegypti* and *C.*
 282 *quinquefasciatus*, respectively. According to equations 1 and 2, this trap model had a female

283 capture ratio of *Aedes* spp. of 29.50%. This indicated that approximately 29.50% of incoming
 284 females were caught. On the other hand, CRtrap-EA captured only one *C. quinquefasciatus*.

285 **Total *Aedes* eggs and adults collected at INPA Campus I in Manaus, Amazonas**

286 According to the data, UELtrap-E was more efficient at collecting eggs than CRtrap-E
 287 (Figure 2). This efficiency was also verified by comparing the averages of the numbers of eggs
 288 obtained between the two traps, since the former obtained an average (2054) almost 10 times
 289 higher than the value (218) acquired by the latter, with a statistically significant difference ($p =$
 290 0.0183) between the respective values (Table 4).

291 By evaluating the efficiency between UELtrap-EA and CRtrap-EA, a higher quantity of
 292 eggs was verified in the former trap type (Fig 2). This result was also confirmed by observing
 293 that the average number of eggs collected in the former trap type (430) were higher than the
 294 average number of eggs verified in the latter trap type (102), which was evidenced by a
 295 statistically significant difference between the values ($p = 0.0078$) (Table 4).

296
 297 **Fig 2. The total numbers of eggs laid by *Aedes* adults in each trap for five weeks from**
 298 **January to February 2018 in Manaus, Amazonas, Brazil.**
 299

300 **Table 4. Average, maximum and minimum eggs laid by *Aedes* adults in each trap from**
 301 **January to February 2018 in Manaus, Amazonas, Brazil.**

Traps	Average (\pm SD)	Maximum	Minimum
UELtrap-E	2054 (1057) ^A	3773	1068
CRtrap-E	218 (138) ^B	363	10
UELtrap-EA	430 (184) ^a	716	221
CRtrap-EA	102 (82) ^b	190	6

302 Legend: SD = standard deviation. Different letters in the same column indicate a statistically significant
 303 difference ($p < 0.05$) between the average number of eggs obtained for the trap that collects the same
 304 stage (eggs or eggs/adults), Student's t-test or Mann-Whitney test.
 305

306 Regarding the average number of eggs obtained by the UELtrap-E traps during each
 307 week, a statistically significant difference was found between the values obtained in the first
 308 (755 ± 489) and fifth weeks (214 ± 185) (p = 0.0495). This result was different from that
 309 observed in the CRtrap-E trap, where there was no significant difference when comparing the
 310 data obtained in each sampling week (p>0.05) (Table 5).

311 In relation to the average number of eggs obtained in each week of sampling with the
 312 use of the UELtrap-EA traps, a statistically significant difference was observed between the
 313 values obtained in the first (143 ± 77) and fifth weeks (44 ± 18) (p = 0.0488) as well as between
 314 the values obtained for the third (84 ± 46) and fifth weeks (p = 0.0472) (Table 5). Considering
 315 the average number of eggs obtained in the CRtrap-EA traps each week, a difference was
 316 observed between the first (1 ± 3) and third weeks (37 ± 36) (p = 0.0163) (Table 5).

317

318 **Table 5. The average and standard deviation of the *Aedes* obtained from each trap from**
 319 **January to February 2018 in Manaus, Amazonas, Brazil.**

Traps	Collection Weeks				
	1°	2°	3°	4°	5°
UELtrap-E	755±489 ^a	460±129 ^{a,b}	310±159 ^{a,b}	315±132 ^{a,b}	214±185 ^b
CRtrap-E	2±3 ^a	73±71.5 ^a	31±31 ^a	58±58 ^a	55± 55 ^a
UELtrap-EA	143±77 ^a	92±121 ^{a,b}	84±46 ^b	67±51 ^{a,b}	44±18 ^b
CRtrap-EA	1±3 ^b	13±14 ^{a,b}	37±36 ^a	13±25 ^{a,b}	38±61 ^{a,b}

320 Legend: Different letters in the same row indicated a statistically significant difference (p<0.05) between
 321 the average number of eggs obtained each week in all traps tested using Student's t-test or Mann-Whitney
 322 test.

323

324 The OPI values for the UELtrap-E traps demonstrated 100% positive values in all weeks
 325 analyzed in the experiment (Table 6). However, in the CRtrap-E traps, the OPI values varied

326 over the sampling period; however, no significant difference was observed between the average
 327 OPI values obtained by the two types of traps ($p > 0.05$).

328 In reference to the UELtrap-E EDI values, there were variations during different
 329 sampling weeks, with lower and higher values in the fifth and first weeks (214 and 755),
 330 respectively (Table 6). Regarding the EDI values obtained in the CRtrap-E traps, variations
 331 were also observed throughout the sampling period, with lower and higher values being
 332 observed in the first and second weeks (3 and 73), respectively (Table 6). However, when
 333 comparing the average EDI values of the different traps, the results obtained in UELtrap-E were
 334 higher than those obtained in CRtrap-E, which was corroborated by a significant difference
 335 observed ($p = 0.0189$).

336 The OPI values for the UELtrap-EA traps were 100% in four of the five weeks analyzed,
 337 except for the fourth week, when this index decreased to 80%. These values are higher than
 338 those obtained in the CRtrap-EA traps, in which varied in each week of collection, with lower
 339 and higher values in the first and third weeks (20 and 100%), respectively (Table 6), as
 340 evidenced by a significant difference between the average OPI values obtained for the two traps
 341 ($p = 0.0472$).

342 The EDI values obtained in the UELtrap-EA traps varied during the weeks analyzed in
 343 the experiment, showing lower and higher values in the fifth and first weeks (44 and 143),
 344 respectively (Table 6). This result was exactly the opposite in the CRtrap-EA traps. Moreover,
 345 when comparing the average of the EDI values obtained in each trap, a statistically significant
 346 difference was observed ($p = 0.0122$) due to the higher egg density in the UELtrap-EA traps.

347
 348 **Table 6. Ovitrap positivity index (OPI) and egg density index (EDI) obtained per week**
 349 **in each trap from January to February 2018 in Manaus, Amazonas, Brazil.**

Weeks	UELtrap-E		CRtrap-E		UELtrap-EA		CRtrap-EA	
	OPI (%)	EDI	OPI (%)	EDI	OPI (%)	EDI	OPI (%)	EDI

1°	100	755	60	3	100	143	20	6
2°	100	460	100	73	100	92	60	22
3°	100	310	100	31	100	84	100	37
4°	100	315	100	58	80	84	60	21
5°	100	214	80	68	100	44	60	63
Average	100 ^A	411 ^a	88 ^A	47 ^b	96 ^A	89 ^a	60 ^B	30 ^b

350 Legend: Different letters in the same row indicate a statistically significant difference ($p < 0.05$) between
 351 the average index (IPO and IDO) obtained for the traps that collect the same stage (eggs or eggs/adults)
 352 using Student's t-test or Mann-Whitney test.

353

354

355

In the UELtrap-EA traps, 25 mosquito specimens were obtained: 23 *A. albopictus*, one
 356 *Limatus* spp. and one *Limatus durhamii* Theobald, 1901, representing percentages of 92%, 4%
 357 and 4%, respectively. Based on equations 1 and 2, these traps presented a female *Aedes* spp.
 358 capture ratio of 53.51%. This indicated that approximately 53.51% of the females who entered
 359 the traps were caught. On the other hand, in the CRtrap-EA traps, only one *Aedes* spp. was
 360 captured. The capture ratio of *Aedes* spp. female for this trap was 9.80%. Therefore,
 361 approximately 9.80% of the females that entered were captured.

362

363 Discussion

364

When observing the smallest number of eggs and the low EDI and OPI values obtained
 365 by the CRtrap-E and CRtrap-EA traps in both study regions, compared to the values obtained
 366 by the UELtrap-E and UELtrap-EA traps, it can be seen that the configuration of the first group
 367 of traps (smaller blades with less rough surface) may not have provided the ideal conditions for
 368 the *Aedes* spp. females to lay eggs.

369

The light coloration of the traps CRtrap-E and CRtrap-EA may also have influenced egg
 370 laying. According to [2] females of the genus *Aedes* prefer darker places for oviposition. This
 371 fact explains the preference of the females in choosing black traps during egg laying. Therefore,

372 the average numbers of eggs obtained in the UELtrap-E and UELtrap-EA traps and the high
373 values of OPI and EDI observed in both regions indicate that these two trap models (dark color
374 and rough surface) were more inviting to *Aedes* spp. females.

375 However, when comparing the results obtained for each of the trap models between the
376 two sampling regions, it was evident that all models showed higher efficiency in capturing eggs
377 and adults of *Aedes* spp. in the North Region. This can be explained by the climate of the city
378 of Manaus, where temperatures remain high throughout the year (annual average around 26 °
379 C), in addition to having abundant rainfall, mainly between the months of November and June
380 (rainy season) [25, 50, 51], covering the period in that the collections were carried out in
381 Manaus.

382 These climate conditions, combined with precarious socio-environmental and economic
383 conditions, frequent in large urban centers like Manaus, provide an ideal environment for the
384 proliferation of mosquitoes, considering the greater availability of breeding sites in these
385 conditions, in addition to the fact that *Aedes* spp. develops faster in a temperature range of 20
386 to 36 °C, similar to the average in Manaus [6, 19, 23, 51-54].

387 For the *Aedes* species captured, the high abundance of *A. albopictus* obtained from
388 INPA *Campus* I, Manaus, and the low abundance of this species obtained from the UTFPR
389 *Campus*, Londrina, can be explained by the trap installation environment. In Manaus, the area
390 is composed of fragments of forest reserves suitable for the species, which prefer periurban or
391 urban environments with greater vegetation cover, which is characteristic of wild environments
392 [2, 5, 21, 22]. In a study by [55] in Manaus, a high density of *A. albopictus* was observed in
393 both the central and peripheral regions of the city, where it occurred in areas of urban and
394 periurban forest with anthropogenic alterations and a large number of artificial containers,
395 corroborating the present results. More recently, [33] also observed predominance of *A.*
396 *albopictus* in a study carried out in the INPA *Campus* I and II.

397 In contrast, the greater amount of *A. aegypti* caught in Londrina can be explained by the
398 fact that the collection area was fully urbanized, unlike the collection area in Manaus,
399 considering that this species is extremely adapted to the urban environment and highly
400 anthropophile [2, 12, 14]. These results corroborate with the studies of [56] and [34], which
401 monitored *A. aegypti* in the state of Paraná. In these studies, the authors observed a higher
402 frequency of *A. aegypti* in urban areas, whereas in rural areas, *A. albopictus* was predominant.

403 The study of [57] also reported that in the municipality of Londrina, Paraná, Brazil, *A.*
404 *aegypti* populations decreased from urban to rural areas, while the opposite occurred for *A.*
405 *albopictus*. In a more recent study by [22] in São Paulo, Brazil, there was also a relationship
406 between the occurrence of these species and the type of environment, where the highest density
407 of *A. aegypti* was found in areas with lower vegetation cover, while in areas with higher
408 vegetation cover, *A. albopictus* predominated.

409 In general, the efficiency of the traps may have been enhanced by the presence of the
410 grass infusion, as it has proven efficacy in attracting *Aedes* spp. compared with the use of only
411 distilled or piped water [32, 33, 38, 58]. The Vectobac WG[®] (*B. thuringiensis israelensis*)
412 biolarvicide used in the experiment as well as other Bti-based products, in conjunction with
413 traps, also can be an important aid for monitoring in view of the proven efficacy of Bti in control
414 of the larvae of *Aedes* spp. Thus, if the larvae hatch from the eggs laid by the females in the
415 reeds, they will not develop into adult form [32, 33]. In addition, the effect of Bti comes from
416 four major synergistic toxins (Cry4Aa, Cry4Ba, CryIIAa and CytIAa), which may reduce the
417 likelihood of selection of resistant target organisms [41, 59-62], besides not cause damage to
418 other organisms (except Chironomidae and Simuliidae) due to their high specificity for
419 mosquitoes [40, 63].

420 Based on the above, UELtrap-EA has the potential to be used in the monitoring of *A.*
421 *aegypti* and *A. albopictus* since they were the most collected species and only *Aedes* eggs were

422 collected. This model has high sensitivity for determining the local infestation index and can
423 be implemented in public health programs to reduce both eggs and adults of *Aedes* spp. in the
424 environment. UELtrap-E also showed potential for reducing *A. aegypti* and *A. albopictus* eggs
425 in the environment and can be easily transported and used, in addition to having a low cost and
426 high sensitivity for determining the local infestation index, as well as the UELtrap-EA.

427 The results observed for these two models in the two study regions also indicated that
428 both have efficiency in different environments and seasons, with different climates,
429 demonstrating the possibility for use in different locations and periods of the year. Regarding
430 the CRtrap-E and CRtrap-EA traps, although they presented lower efficiency in capturing the
431 eggs and adults of *Aedes*, they can be optimized by using larger reeds with rougher surfaces for
432 fixing eggs as well as by using darker colors.

433 These traps do not inconvenience those in the installation areas or to the health workers
434 who should be charged with monitoring the traps since they do not need to be installed indoors
435 but rather in open areas with a large flow of people, such as outside of universities, institutes
436 and industrial buildings as well as in peridomiciles. These traps are an operationally viable and
437 noninvasive method and may become the most effective, practical and economical way to
438 monitor *A. aegypti* and *A. albopictus* on a local scale, provided that the traps are monitored
439 weekly by technical staff.

440 The entire process can be reconciled with official government strategies for more
441 accurate vector monitoring that can support actions with the population for local surveys and
442 greater efficiency in vector control when necessary.

443

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451

452 **References**

- 453 1. Walter Reed Biosystematics Unit (WRBU). Mosquito Identification Resources. 2020.
454 [Cited 23 April 2020]. Available from: http://www.wrbu.org/VecID_MQ.html.
- 455 2. Forattini OP. Culicidologia Médica - Identificação, biologia e epidemiologia.
456 Culicidologia Médica. 2002.
- 457 3. Pan American Health Organization (PAHO). Folha informativa – Dengue e dengue
458 grave. 2020. [Cited 23 April 2020]. Available from:
459 [https://www.paho.org/bra/index.php?option=com_content&view=article&id=5963:f](https://www.paho.org/bra/index.php?option=com_content&view=article&id=5963:folha-informativa-dengue-e-dengue-grave&Itemid=812)
460 [olha-informativa-dengue-e-dengue-grave&Itemid=812](https://www.paho.org/bra/index.php?option=com_content&view=article&id=5963:folha-informativa-dengue-e-dengue-grave&Itemid=812).
- 461 4. World Health Organization (WHO). Dengue and severe Dengue. 2020a. [Cited 23 April
462 2020]. Available from: [https://www.who.int/en/news-room/fact-](https://www.who.int/en/news-room/fact-sheets/detail/dengue-and-severe-dengue)
463 [sheets/detail/dengue-and-severe-dengue](https://www.who.int/en/news-room/fact-sheets/detail/dengue-and-severe-dengue).
- 464 5. World Health Organization (WHO). The mosquito. 2020b. [Cited 23 April 2020].
465 Available from: <https://www.who.int/denguecontrol/mosquito/en>.
- 466 6. Kraemer MUG, Sinka ME, Duda KA, Mylne AQN, Shearer FM, Barker CM, et al. The
467 global distribution of the arbovirus vectors *Aedes aegypti* and *Ae. Albopictus*.
468 *Elife*,4,e08347. 2015. doi:10.7554/eLife.08347

- 469 7. Ding F, Fu J, Jiang D, Hao M, Lin G. Mapping the spatial distribution of *Aedes aegypti*
470 and *Aedes albopictus*. *Acta Trop.* 2018;178: 155-162.
471 doi:10.1016/j.actatropica.2017.11.020
- 472 8. Carvalho FD, Moreira LA. Why is *Aedes aegypti* Linnaeus so Successful as a Species?
473 *Neotropical Entomology.* 2017; 46(3): 243-255. doi:10.1007/s13744-017-0520-4
- 474 9. Azevedo RSS, Oliveira CS, Vasconcelos PFC. Chikungunya risk for Brazil. *Rev*
475 *Saude Publica.* 2015;49(58). doi:10.1590/S0034-8910.2015049006219
- 476 10. Henriques CMP, Duarte E, Garcia LP. Desafios para o enfrentamento da epidemia de
477 microcefalia. *Epidemiologia e Serviços de Saúde.* 2016; 25(1): 7-10.
478 doi:10.5123/S1679-49742016000100001
- 479 11. Monteiro FJC, Mourão FRP, Ribeiro ESDA, Rêgo MOS, Frances PAC, Souto RNP, et al.
480 Prevalence of dengue, Zika and chikungunya viruses in *Aedes (Stegomyia) aegypti*
481 (Diptera: Culicidae) in a medium-sized city, Amazon, Brazil. *Revista do Instituto de*
482 *Medicina Tropical de São Paulo.* 2020;62:e10. doi:10.1590/s1678-9946202062010
- 483 12. Lima-Camara TN, Honório NA, Lourenço-de-Oliveira R. Freqüência e distribuição
484 espacial de *Aedes aegypti* e *Aedes albopictus* (Diptera, Culicidae) no Rio de Janeiro,
485 Brasil. *Cadernos de saude publica.* 2006;22(10): 2079-2084. doi:10.1590/S0102-
486 311X2006001000013
- 487 13. Zara ALSA, Santos SM, Fernandes-Oliveira ES, Carvalho RG, Coelho GE. Estratégias de
488 controle do *Aedes aegypti*: uma revisão. *Epidemiologia e Serviços de Saúde.*
489 2016;25:391-404.
- 490 14. Fonseca DPD, Serpa LLN, Barbosa GL, Pereira M, Holcman MM., Voltolini JC, et al.
491 Vectors of arboviruses in the state of São Paulo: 30 years of *Aedes aegypti* and *Aedes*

- 492 *albopictus*. Revista de Saúde Pública. 2019;53(84). doi:10.11606/s1518-
- 493 8787.2019053001264
- 494 15. Consoli RA, Oliveira RL. Principais mosquitos de importância sanitária no Brasil, 1ª ed.
- 495 SciELO-Editora FIOCRUZ, Rio de Janeiro; 1994. p 225.
- 496 16. Soares-da-Silva J, Ibiapina SS, Bezerra JMT, Tadei WP, Pinheiro VCS. Variation in
- 497 *Aedes aegypti* (Linnaeus) (Diptera, Culicidae) infestation in artificial containers in
- 498 Caxias, state of Maranhão, Brazil. Revista da Sociedade Brasileira de Medicina
- 499 Tropical. 2012; 45(2): 174-179. doi:10.1590/S0037-86822012000200007
- 500 17. Bezerra JMT, Santana INS, Miranda JP, Tadei WP, Pinheiro VCS. Breeding sites of
- 501 *Aedes aegypti* (Linnaeus) (Diptera, Culicidae): a study about the containers diversity
- 502 in dry and rainy seasons in a dengue-endemic city. Revista de Pesquisa em Saúde.
- 503 2018;18(2): 102-107.
- 504 18. Meena AR, Choudhary NL. Container breeding preference of *Aedes albopictus* in
- 505 urban environment. International Journal of Mosquito Research. 2019;6(5): 44-47.
- 506 19. Vega-Rúa A, Zouache K, Girod R, Failloux AB, Lourenço-de-Oliveira R. High level of
- 507 vector competence of *Aedes aegypti* and *Aedes albopictus* from ten American
- 508 countries as a crucial factor in the spread of Chikungunya virus. Journal of virology.
- 509 2014;88(11): 6294-6306. doi:10.1128/JVI.00370-14
- 510 20. Gomes AC, Souza JMP, Bergamaschi DP, Santos JLF, Andrade VR, Leite OF, et al.
- 511 Atividade antropofílica de *Aedes aegypti* e *Aedes albopictus* em área sob controle e
- 512 vigilância. Revista de Saúde Pública. 2005;39(2): 206-210. doi:10.1590/S0034-
- 513 89102005000200010
- 514 21. Medeiros-Sousa AR, Ceretti-Júnior W, Carvalho GC, Nardi MS, Araujo AB, Vendrami
- 515 DP, et al. Diversity and abundance of mosquitoes (Diptera:Culicidae) in an urban

- 516 park: Larval habitats and temporal variation. *Acta Tropica*. 2015;150: 200-
517 209. doi:10.1016/j.actatropica.2015.08.002
- 518 22. Heinisch MRS, Diaz-Quijano FA, Chiaravalloti-Neto F, Pancetti FGM, Coelho RR,
519 Andrade PS, et al. Seasonal and spatial distribution of *Aedes aegypti* and *Aedes*
520 *albopictus* in a municipal urban park in São Paulo, SP, Brazil. *Acta Tropica*. 2019;189:
521 104-113. doi:10.1016/j.actatropica.2018.09.011
- 522 23. Pinheiro VCS, Tadei WP. Frequency, diversity, and productivity study on the *Aedes*
523 *aegypti* most preferred containers in the city of Manaus, Amazonas, Brazil. *Revista do*
524 *Instituto de Medicina Tropical de São Paulo*. 2002;44(5): 245-250.
525 doi:10.1590/S0036-46652002000500002
- 526 24. Liu-Helmersson J, Brännström Å, Sewe MO, Semenza JC, Rocklöv J. Estimating past,
527 present, and future trends in the global distribution and abundance of the arbovirus
528 vector *Aedes aegypti* under climate change scenarios. *Frontiers in public health*.
529 2019;7: 148. doi:10.3389/fpubh.2019.00148
- 530 25. INMET - National Institute of Meteorology. Meteorological Database for Teaching
531 and Research. 2020. [Cited 23 April 2020]. Available from:
532 <http://www.inmet.gov.br/portal/index.php?r=bdmep/bdmep>.
- 533 26. Silva JVJ, Lopes TRR, Oliveira-Filho EF, Oliveira RAS, Durães-Carvalho R, Gil
534 LHV. Current status, challenges and perspectives in the development of vaccines
535 against yellow fever, dengue, Zika and chikungunya viruses. *Acta Tropica*. 2018a;182:
536 257-263. doi:10.1016/j.actatropica.2018.03.009
- 537 27. Martins MM, Prata-Barbosa A, Cunha AJLA. Arboviroses na infância. *Jornal de*
538 *Pediatria*. 2020;96: 2-11. doi:10.1016/j.jpmed.2019.08.005

- 539 28. Powell JR. Mosquitoes on the move. *Science*. 2016;354(6315): 971-972. doi:
540 10.1126/science.aal1717
- 541 29. Ministério da Saúde (MS). Secretaria de Vigilância em Saúde. Boletim Epidemiológico
542 16. Ministério da Saúde, Secretaria de Vigilância em Saúde – Brasília, DF. 2020.
- 543 30. Facchinelli L, Valerio L, Pombi M, Reiter P, Costantini C, Della Torre A. Development
544 of a novel sticky trap for container-breeding mosquitoes and evaluation of its
545 sampling properties to monitor urban populations of *Aedes albopictus*. *Medical and*
546 *veterinary entomology*. 2007;21(2): 183-195. doi:10.1111/j.1365-2915.2007.00680.x
- 547 31. Regis L, Monteiro AM, Melo-Santos MAVD, Silveira Jr JC, Furtado AF, Acioli RV, et al.
548 Developing new approaches for detecting and preventing *Aedes aegypti* population
549 outbreaks: basis for surveillance, alert and control system. *Memórias do Instituto*
550 *Oswaldo Cruz*. 2008;103(1): 50-59. doi:10.1590/S0074-02762008000100008
- 551 32. Depoli PAC, Zequi JAC, Nascimento KLC, Lopes J. Eficácia de Ovitrapas com
552 Diferentes Atrativos na Vigilância e Controle de *Aedes*. *EntomoBrasilis*. 2016;9(1): 51-
553 55. doi:10.12741/ebrasilis.v9i1.509
- 554 33. Silva WR, Soares-da-Silva J, Ferreira FAS, Rodrigues IB, Tadei WP, Zequi JAC.
555 Oviposition of *Aedes aegypti* Linnaeus, 1762 and *Aedes albopictus* Skuse, 1894
556 (Diptera: Culicidae) under laboratory and field conditions using ovitraps associated to
557 different control agents, Manaus, Amazonas, Brazil. *Revista Brasileira de*
558 *Entomologia*. 2018b;62(4): 304-310. doi:10.1016/j.rbe.2018.08.001
- 559 34. Zequi JAC, Oliveira AA, Santos FP, Lopes J. Monitoramento e controle de *Aedes*
560 *aegypti* (Linnaeus, 1762) e *Aedes albopictus* (Skuse, 1984) com uso de ovitrampas.
561 *Semina: Ciências Biológicas e da Saúde*. 2018;39(2): 93-102. doi:10.5433/1679-
562 0367.2018v39n2p93

- 563 35. Ministério da Saúde (MS). Secretaria de Vigilância em Saúde. Departamento de
564 Vigilância Epidemiológica. Secretaria de Vigilância em Saúde. Departamento de
565 Vigilância Epidemiológica. Diretrizes nacionais para prevenção e controle de
566 epidemias de dengue. Ministério da Saúde, Secretaria de Vigilância em saúde,
567 Departamento de Vigilância Entomológica – Brasília, DF. 2009.
- 568 36. Ritchie SA, Long S, Hart A, Webb CE, Russell RC. An adulticidal sticky ovitrap for
569 sampling container-breeding mosquitoes. *Journal of the American Mosquito Control*
570 *Association*. 2003;19(3): 235-242.
- 571 37. Reiter P, Amador MA, Colon N. Enhancement of the CDC ovitrap with hay infusions
572 for daily monitoring of *Aedes aegypti* populations. *Journal of the American Mosquito*
573 *Control Association*. 1991;7(1): 52-55.
- 574 38. Sant’Ana AL, Roque RA, Eiras AE. Characteristics of grass infusions as oviposition
575 attractants to *Aedes (Stegomyia)* (Diptera: Culicidae). *Journal of medical entomology*.
576 2006; 43(2): 214-220. doi:10.1093/jmedent/43.2.214
- 577 39. Goldberg LJ, Margalit J. A bacterial spore demonstrating rapid larvicidal activity
578 against *Anopheles sergentii*, *Uranotaenia unguiculata*, *Culex univitattus*, *Aedes*
579 *aegypti* and *Culex pipiens*. *Mosquito News*. 1977;37(3): 355-358.
- 580 40. Lee BM, Scott GI. Acute toxicity of temephos, fenoxycarb, diflubenzuron, and
581 methoprene and *Bacillus thuringiensis* var. *israelensis* to the mummichog (*Fundulus*
582 *heteroclitus*). *Bulletin of Environmental Contamination and Toxicology*. 1989;43(6):
583 827-832. doi:10.1007/BF01702051
- 584 41. Boyce R, Lenhart A, Kroeger A, Velayudhan R, Roberts B, Horstick O. *Bacillus*
585 *thuringiensis israelensis* (Bti) for the control of dengue vectors: Systematic literature

- 586 review. *Tropical Medicine & International Health*. 2013;18(5): 564-577.
587 doi:10.1111/tmi.12087
- 588 42. Fay RW, Perry AS. Laboratory studies of ovipositional preferences of *Aedes aegypti*.
589 *Mosquito News*. 1965;25(3): 276-281.
- 590 43. Fay RW, Eliason DA. A preferred oviposition site as a surveillance method for *Aedes*
591 *aegypti*. *Mosquito News* 1966;26(4): 531-535.
- 592 44. IAPAR – Instituto Agronômico do Paraná. 2020. [Cited 23 April 2020]. Available from:
593 <http://www.iapar.br/modules/conteudo/conteudo.php?conteudo=2352>.
- 594 45. Harbach RE. 2020. [Cited 23 April 2020]. Available from: Mosquito Taxonomic
595 Inventory. <http://mosquito-taxonomic-inventory.info/>.
- 596 46. Gomes AC. Medidas dos níveis de infestação urbana para *Aedes (Stegomyia) aegypti*
597 e *Aedes (Stegomyia) albopictus* em programas de Vigilância Entomológica. Informe
598 Epidemiológico do SUS. 1998;7: 49-57.
- 599 47. Ayres M, Ayres JM., Ayres DL, Santos ADA. Aplicações estatísticas nas áreas das
600 ciências bio-médicas. Instituto Mamirauá, Belém. 2007;364.
- 601 48. Bentley MD, Day JF. Chemical ecology and behavioral aspects of mosquito
602 oviposition. *Annual review of entomology*. 1989;34(1): 401-421.
603 doi:10.1146/annurev.en.34.010189.002153
- 604 49. Roque RA. Avaliação de atraentes de oviposição, identificados em infusões de capim
605 colônio (*Panicum maximum*) para fêmeas de *Aedes aegypti* (L. 1762) (Diptera:
606 Culicidae) em condições de semicampo e campo. Masters' dissertation, Universidade
607 Federal de Minas Gerais. 2007. Available from: [http://hdl.handle.net/1843/SAGF-](http://hdl.handle.net/1843/SAGF-765L57)
608 [765L57](http://hdl.handle.net/1843/SAGF-765L57).

- 609 50. Silva D. A função da precipitação no conforto do clima urbano da cidade de
610 Manaus. *Revista Geonorte*. 2012;3(9): 22-40.
- 611 51. Gomes ACS, Santos TS, Coutinho MDL, Silva AR. Estudo biometeorológico entre as
612 condições de tempo e a malária na Amazônia Legal. *Revista de Sustentabilidade e*
613 *Tecnologia Ambiental*. 2014;9(1): 47-58.
- 614 52. Calado DC, Silva MAND. Avaliação da influência da temperatura sobre o
615 desenvolvimento de *Aedes albopictus*. *Revista de Saúde Pública*. 2002;36(2): 173-
616 179. doi:10.1590/S0034-89102002000200009
- 617 53. Beserra, EB, Fernandes, CRM, Silva SAO, Silva LA, Santos JW. Efeitos da
618 temperatura no ciclo de vida, exigências térmicas e estimativas do número de gerações
619 anuais de *Aedes aegypti* (Diptera, Culicidae). *Iheringia Série Zool*. 2009;99(2): 142-
620 148. <https://doi.org/10.1590/s0073-47212009000200004>
- 621 54. Marinho RA, Beserra EB, Bezerra-Gusmão MA, Porto VDS, Olinda RA, Santos CA.
622 Effects of temperature on the life cycle, expansion, and dispersion of *Aedes aegypti*
623 (Diptera: Culicidae) in three cities in Paraíba, Brazil. *Journal of Vector Ecology*.
624 2016;41(1): 1-10. doi:10.1111/jvec.12187
- 625 55. Sá ELR. Estudo das áreas de ocorrência e dos criadouros preferenciais de *Aedes*
626 *albopictus* Skuse, 1894 (Diptera: Culicidae) em Manaus, Amazonas, Brasil. INPA,
627 2004. Available from: <https://bdtd.inpa.gov.br/handle/tede/2903>.
- 628 56. Fantinatti E, Duque JE, Silva AM, Navarro-Silva MA. Abundance and aggregation egg
629 of *Aedes aegypti* L. and *Aedes albopictus* (Skuse) (Diptera: Culicidae) in the north and
630 northwest of the State of Paraná, Brazil. *Neotropical Entomology*. 2007;36(6): 960-
631 965. doi:10.1590/S1519-566X2007000600020

- 632 57. Lopes J, Martins EAC, Oliveira O, Oliveira V, Oliveira Neto BP, Oliveira JE. Dispersion
633 of *Aedes aegypti* (Linnaeus, 1762) and *Aedes albopictus* (Skuse, 1894) in the rural
634 zone of north Paraná State. Brazilian Archives of Biology and Technology. 2004;47(5):
635 739-746. doi:10.1590/S1516-89132004000500009
- 636 58. Nunes LS, Trindade RR, Souto RN. Avaliação da atratividade de ovitrampas a *Aedes*
637 (*Stegomyia*) *aegypti* Linneus (Diptera: Culicidae) no bairro Hospitalidade, Santana,
638 Amapá. Biota Amazônia (Biote Amazonie, Biota Amazonia, Amazonian Biota).
639 2011;1(1): 26-31. doi:10.18561/2179-5746/biotaamazonia.v1n1p26-31
- 640 59. Becker N. Bacterial control of vector-mosquitoes and black flies. Entomopathog Bact
641 from Lab to F Appl. 2000;383-398. doi:10.1007/978-94-017-1429-7_21
- 642 60. Regis L, Silva-Filha MH, Nielsen-LeRoux C, Charles JF. Bacteriological larvicides of
643 dipteran disease vectors. Trends in Parasitology. 2001;17(8): 377-380.
- 644 61. Araújo AP, Araujo Diniz DF, Helvecio E, De Barros RA, De Oliveira CMF, Ayres
645 CFJ, et al. The susceptibility of *Aedes aegypti* populations displaying temephos
646 resistance to *Bacillus thuringiensis israelensis*: A basis for management. Parasites and
647 Vectors. 2013;6(1): 297. doi:10.1186/1756-3305-6-297
- 648 62. Carvalho KS, Crespo MM, Araújo AP, Silva RS, Melo-Santos MAV, Oliveira CMF, et al.
649 Long-term exposure of *Aedes aegypti* to *Bacillus thuringiensis* *svar. israelensis* did not
650 involve altered susceptibility to this microbial larvicide or to other control
651 agents. Parasites & vectors. 2018;11(1): 673. doi:10.1186/s13071-018-3246-1
- 652 63. Lacey LA. *Bacillus thuringiensis* serovariety *israelensis* and *Bacillus sphaericus* for
653 mosquito control. Journal of the American Mosquito Control Association. 2007;23:
654 133-164. doi:10.2987/8756-971X(2007)23[133:BTSIAB]2.0.CO;2

Londrina, Paraná

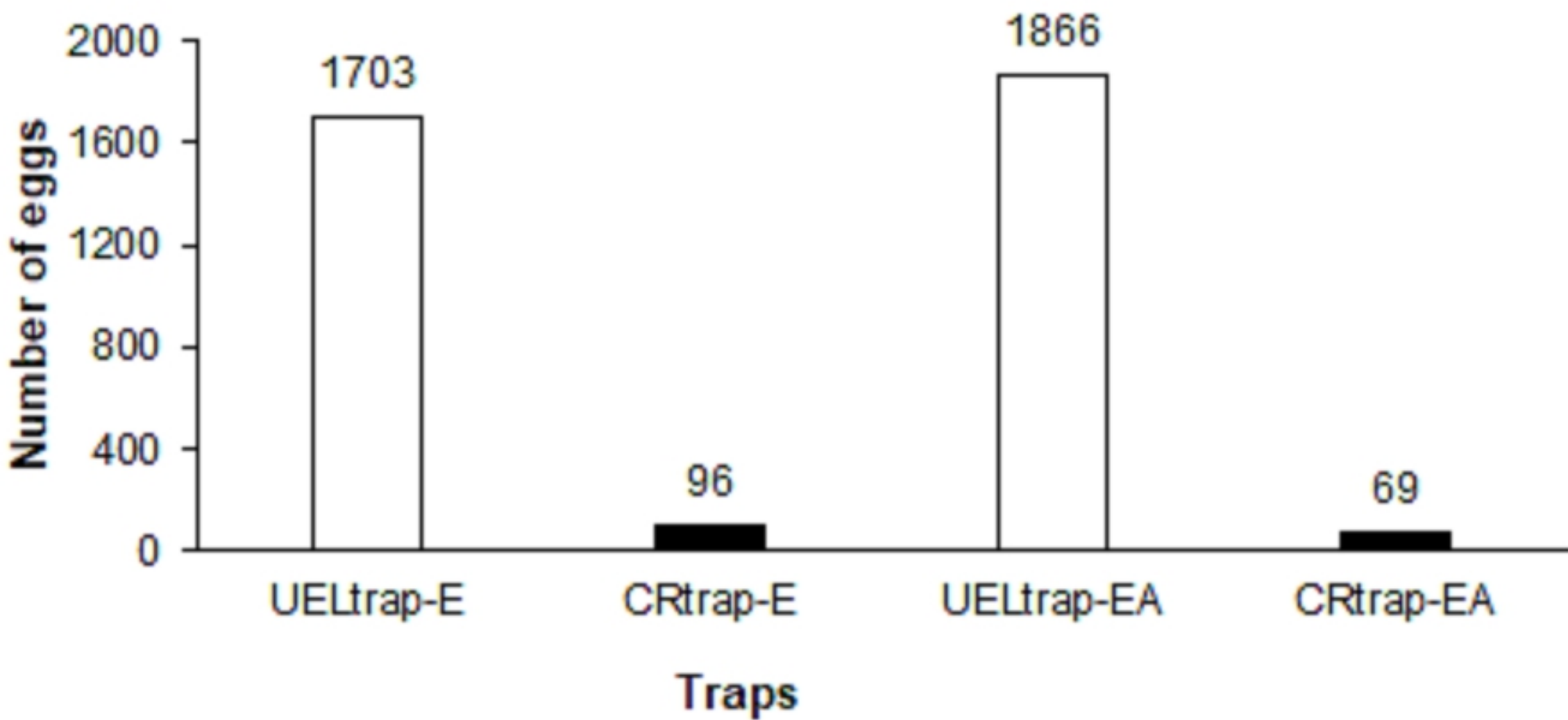


Figure 1

Manaus, Amazonas

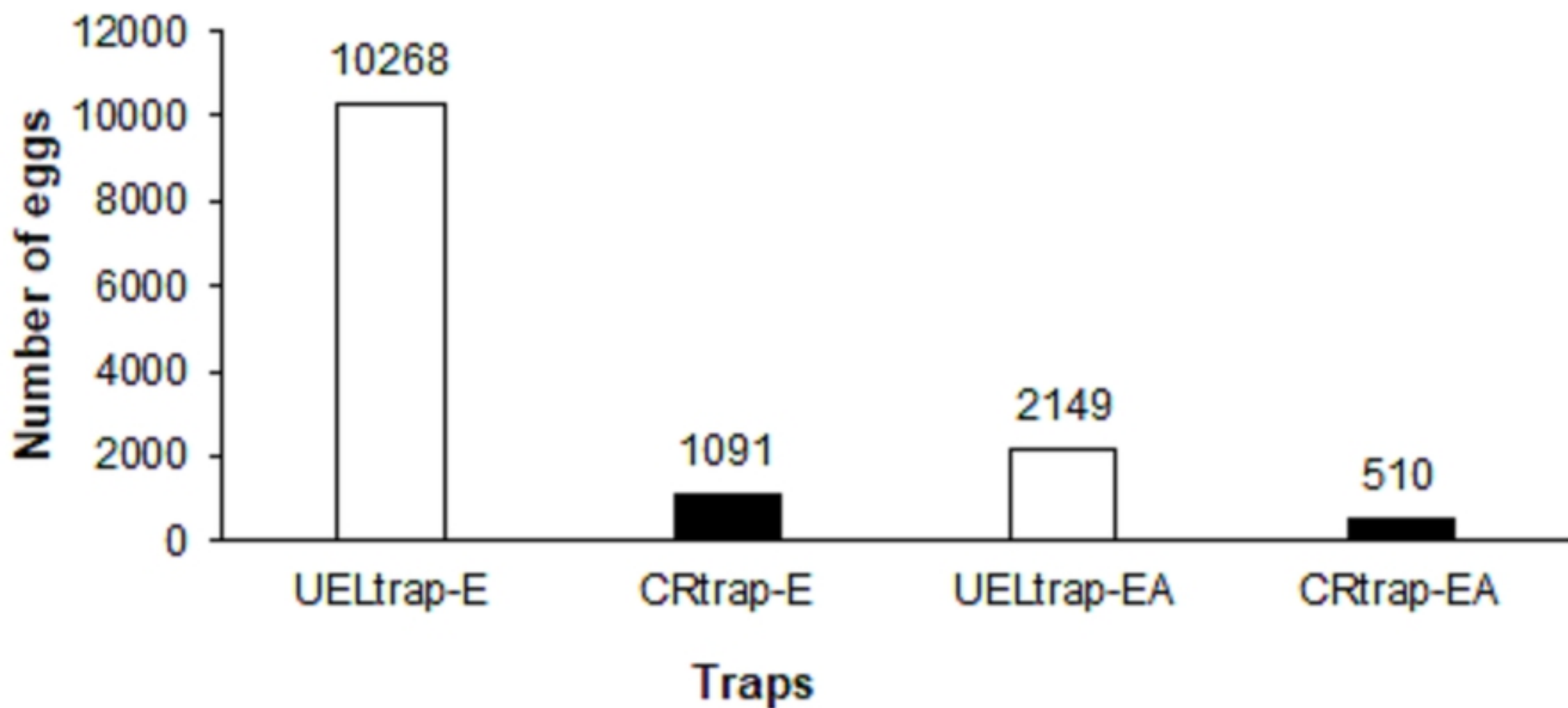


Figure2