TITLE: Divergent selection on behavioral and chemical traits contributes to isolation between populations of *Drosophila melanogaster*

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Abstract

Speciation is driven by traits that can act to prohibit mating between nascent lineages, including male courtship and female preference for male traits. Mating barriers involving these traits evolve quickly because there is strong selection on males and females to maximize reproductive success, and the tight co-evolution of mating interactions can lead to rapid diversification of sexual behavior. Using lineages of *D. melanogaster* that show strong asymmetrical reproductive isolation, we ask two key questions: which specific male traits are females selecting, and are these traits under divergent sexual selection? These questions have proven extremely challenging to answer, because even in closely related lineages males often differ in multiple traits related to mating behavior. We address these questions by estimating selection gradients for male courtship and cuticular hydrocarbons for two different female genotypes. We identify both specific behaviors and several cuticular hydrocarbons that are under divergent sexual selection and likely contribute to reproductive isolation. Additionally a subset of these traits are plastic; males adjust these traits based on the identity of the female genotype they interact with. These results suggest that even when male courtship is not fixed between lineages, ongoing selection can contribute to reproductive isolation.

Introduction

Sexual selection is a powerful force that drives courtship behavior evolution in populations, and can eventually lead to reproductive isolation (KIRKPATRICK 1982; ANDERSSON 1994; BOUGHMAN 2002; COYNE AND ORR 2004; RITCHIE 2007). This observation is supported by comparative studies that quantify rapid trait evolution across widely diverged clades (BARRACLOUGH et al. 1995; MOLLER AND CUERVO 1998) as well as by studies that estimate selection gradients on sexual traits in focal populations within species (HILL 1991; BROOKS AND ENDLER 2001; REBARM et al. 2009; CALLANDER et al. 2012; OH AND SHAW 2013; STEIGER AND STOKL 2014). Sexual selection can vary between populations, which could be a prerequisite for divergent selection and speciation (CHENOWETH et al. 2010; WATTS et al. 2019), but documenting ongoing divergent selection in the context of lineage divergence is more difficult (HIGGIE et al. 2000; MAAN et al. 2004; SVENSSON et al. 2006; MAAN et al. 2010; PAUERS AND MCKINNON 2012; LANGERHANS AND MAKOWICZ 2013; SELZ et al. 2016; WILKINS et al. 2016). As a result, it has been questioned whether one can make inferences about the speciation process by looking at within-population sexual selection (PANHUIS et al. 2001; VAN DOORN et al. 2004; RITCHIE 2007). The inability to make connections between ongoing sexual selection and reproductive isolation may reflect that the traits that are most intensely studied for premating reproductive isolation are traits that are conspicuous fixed differences between well-established species (MCKINNON AND RUNDLE 2002; COYNE AND ORR 2004; QVARNSTROM et al. 2010). This contrasts with recently diverged species that may not show fixed differences in key mating-related traits, but nevertheless show strong reproductive isolation (MALLET 2008; HENDRY et al. 2009; MEROT et al. 2017). By estimating selection gradients in lineages that are recently diverged or diverging and show strong isolation, one can determine which traits contribute to

reproductive isolation and whether ongoing selection reflects divergence in traits that contribute to reproductive isolation.

One reason why ongoing (i.e. contemporary) sexual selection might not provide information about how fixed species differences have evolved is because the targets or intensity of sexual selection have changed over the course of speciation (SCHLUTER AND PRICE 1993; PRICE 1998), especially if mating signals are environmentally dependent (reviewed in CANDOLIN 2019). When species differences are fixed, ongoing sexual selection can continue to act, but may affect traits not directly associated with reproductive isolation (RYAN AND RAND 1993; BOAKE *et al.* 1997). In nascent lineages, however, one is much more likely to identify divergent selection on traits that contribute directly to reproductive isolation. One challenge for studying nascent lineages is that suites of traits show correlated divergence (HOHENLOHE AND ARNOLD 2010; OH AND SHAW 2013). When multiple correlated traits are divergent it can be difficult to disentangle which traits are important for female mate choice (HOHENLOHE AND ARNOLD 2010). This can be especially true when there are overlapping trait values between lineages, or if males have the potential to change their courtship based on female identity (PFENNIG *et al.* 2010; BERDAN *et al.* 2019; FOX *et al.* 2019).

In models of speciation by sexual selection, it is often assumed that there are single optimal male traits that are selected by a uniform female preference (LANDE 1981; KIRKPATRICK 1982; KIRKPATRICK AND RAVIGNE 2002). However, variation in female preference often occurs, where one male phenotype is not uniformly preferred (JENNIONS AND PETRIE 1997; REBAR AND RODRIGUEZ 2013; MENDELSON *et al.* 2014; KELLEY 2018). The maintenance of variation in male courtship traits and female preferences can lead to frequency-dependent selection (OTTO *et al.* 2008), which in turn can lead to rapid evolution of reproductive isolation if populations become

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geographically isolated (OTTO *et al.* 2008; MENDELSON *et al.* 2014; CASTILLO AND DELPH 2016). Variation in female mate preference could also maintain selection for male courtship plasticity, if males maximize fitness by tailoring courtship to match female preference. Males can alter their copulatory behavior, usually in intensity of courtship or size of ejaculate, based on the presence of rivals or based on the mating status of females (reviewed in BRETMAN *et al.* 2011; KELLEY AND JENNIONS 2011). Males can also alter chemical signals, such as cuticular hydrocarbons (CHCs), in response to being exposed to females or to different social environments (PETFIELD *et al.* 2005; OTTE *et al.* 2018).

In this study we use incompletely isolated lineages of *D. melanogaster* that show strong asymmetrical reproductive isolation to determine which traits are under divergent selection. *D. melanogaster* originated in southern Africa and migrated out of Africa in the past 10,000-15,000 years (LI AND STEPHAN 2006; POOL *et al.* 2012; KAPOPOULOU *et al.* 2018). This resulted in lineages that remained in the ancestral range, called Z-type, and cosmopolitan lineages, M-type, that spread worldwide (WU *et al.* 1995; HOLLOCHER *et al.* 1997). Reproductive isolation between these lineages is asymmetric; M-type females mate indiscriminately when given a choice between males of each type, while Z-type females typically show strong preference for Z-type males (WU *et al.* 1995; HOLLOCHER *et al.* 1997). Given that female preference has evolved rapidly, it is assumed that male courtship has also rapidly evolved; however, divergence in male courtship has not been thoroughly tested, aside from the role of a few cuticular hydrocarbons (Grillet et al. 2012).

We estimated selection gradients on courtship behaviors and cuticular hydrocarbons for both M-type and Z-type females to determine if divergent selection is acting on specific traits. From previous work using common M-type lab strains, we had *a priori* predictions that the time

spent producing vibratory courtship song, and the cuticular hydrocarbons 7-tricosene (7-C23) and cis-vaccenyl acetate (cVA), would be under positive selection in M-type lineages (WU et al. 1995; TALYN AND DOWSE 2004; GRILLET et al. 2006; KURTOVIC et al. 2007; SCOTT et al. 2011). We predicted that these traits might be used for Z-type females to discriminate against M-type males (GRILLET et al. 2012), similar to a species recognition trait. But to make connections between divergent selection and reproductive isolation, we needed to determine which traits are under selection specifically in the Z-type lineage. We found several behavior and cuticular hydrocarbons under divergent selection for both linear selection gradients and for the probability that copulation occurred. Several traits were also plastic and female-genotype dependent. Overall we identified traits that likely contribute to reproductive isolation through contemporary divergent selection in *D. melanogaster* populations.

Methods

Fly Strains

All stocks were reared on standard yeast-glucose media prepared at Cornell University, and were kept at room temperature (~22°C). Information for each strain is provided in the Supplemental Information. Whether a strain is Z-type or M-type has historically been classified by female preference behavior using choice tests, where females are presented with a choice between a M-type and Z-type male. Z-type females choose Z-type males preferentially (WU et al. 1995; HOLLOCHER et al. 1997). There is some correlation within a genotype between Z-type female behavior and male courtship that facilitates choice by another Z-type female genotype (HOLLOCHER et al. 1997), suggesting that these strains could be classified as having Z-type male behavior. For all mating experiments and cuticular hydrocarbon extractions, virgin males and

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females were collected and aged for one week. Each male fly was kept in an individual vial since male mating behaviors can be affected by their interactions with other males (DIXON *et al.* 2003).

Video Recording and analysis of Courtship Behavior

To record multiple courtship interactions simultaneously we used a chamber design that allowed for easy loading of individuals with a mechanism to keep individuals separated until we could start recording (KOEMANS *et al.* 2017). All mating experiments were carried out within two hours of lights on (between 9:00AM and 11:00AM local time) as this is the time we could observe the maximum number of copulations. In pilot recordings we found no differences in courtship intensity or copulation rate for recordings that were done early or late in our recording window. The recording chamber was placed into a lightbox with LED strips (Konseen 16" Square Mini Dimmable Photo Light Box) to control light intensity across trials. We recorded courtship for 30 minutes with a camera (RoHS 0.3MP B&W Firefly MV USB 2.0 Camera). We used two courtship chambers to allow each chamber to be empty between recordings so any volatile cues from the flies would be dissipated.

To quantify the time spent performing specific behaviors we used the software BORIS (FRIARD AND GAMBA 2016). We quantified the behavior of all males from pairs that copulated successfully, as well as several unsuccessful trials selected for specific comparisons (see below). Behaviors were scored manually because we were interested in potentially identifying novel behaviors. While BORIS can categorize a behavior as a state behavior (a single instance) or duration (time behavior takes place with distinct stop/start) we chose to record all behaviors at state behaviors at 10 second intervals (similar to GAERTNER *et al.* 2015). This might underestimate the time spent on any given behavior, but still captures large differences in behavioral sequence since on average, interactions were 10.5 minutes long.

During the first stage of observations, we watched all movies to determine the total number of behaviors that were occurring. Most of these are well described elements of *Drosophila* courtship and include separate (no courtship), singing, chasing, licking, attempted-copulation, copulating, and scissoring (COBB *et al.* 1985; COBB AND JALLON 1990). We noticed that singing behavior occurred at two positions relative to the female, so we initially designated a separate singing (singing-2 is from mostly in front of the female, head-to-head, compared to "normal" singing which occurs with the male behind or slightly to the side of the female). We also noticed several "circling" behaviors where the male moved in an arc around the female while wing displaying (singing or scissoring; Supplemental Video 1). After watching many videos we determined that given the setup of our courtship chamber, we could not reliably score licking and tapping, so we combined these with chasing and following behaviors into "engaging". The frequency of circling behaviors across this initial dataset indicated that only one circling behavior was frequent enough to reliably identify, so all circling behaviors were combined into a single metric. This left us with 8 categories: separate, engaging, singing, singing-2, scissoring, circling, attempted-copulation, and copulating.

When scoring the male behaviors, the video was run through twice the actual speed and paused every 10 seconds to record a behavior. The observer recorded observations blindly; they did not know the identity of the genotypes in the specific well/date they were watching. The genotype identity of each replicate was recorded separately from the video file and only attached to the observation data for statistical analysis. The scoring for trials that successfully copulated ended with a single "copulation" observation as soon as the flies started copulation. There were several trials that were unsuccessful in mating that we watched to make specific comparisons

(see below in Statistical analysis). The unsuccessful trials were quantified for a time equal to the average copulation latency of the successful trials for the same genotype combinations.

Cataloging potential Z-type behaviors

During the initial recordings, our goal was to catalog behaviors for one standard Z-type strain (Z53), one standard M-type strain (DGRP882), and one *D. simulans* strain (SA22). Each mating included a single virgin male and virgin female. Though the courtship sequence of *D. melanogaster* is well known, this information comes exclusively from M-type strains (COBB *et al.* 1985) so it was critical to establish what, if any, additional behaviors may be occurring in the Z-type strains. The *D. simulans* strain served as a standard for the scissoring behavior as it is a frequent in their courtship display (COBB *et al.* 1985; Supplemental Video 2) and there is evidence that some genotypes of *D. melanogaster* with African ancestry may engage in this behavior (YUKILEVICH AND TRUE 2008). We recorded six pairs of flies during every trial block to increase efficiency but also documented behaviors that required increased magnification. We were not able to accurately estimate tapping or licking behaviors with this setup.

Selection acting on courtship behaviors

Our second stage of recording was aimed at identifying behaviors under selection by correlating mating behaviors with copulation success and copulation latency, which was our proxy for mating success and fitness (HOFFMANN 1999; TAYLOR *et al.* 2007). We conducted mating between Z53 females or DGRP882 females and the males from our panel of M and Z-type genotypes. During each block, ten pairs of flies were video recorded simultaneously with five pairs courting Z53 females and the remaining five courting DGRP882 females. We used five male genotypes per block such that the Z53 and DGRP882 females were paired with the same genotypes within a block. These five male genotypes for each block were determined from a

randomized sequence that was clustered into groups of five. The order of the sequence was rotated after every three blocks of recording, so that each strain had equal probability of being in an early vs late block across the course of the experiments. This randomization scheme limited the effects on courtship intensity both due to time of day and blocks occurring across multiple weeks. We repeated this scheme until all genotype combinations had five successful matings, which resulted in some combinations having more successful matings than others.

Cuticular hydrocarbon collection and quantification

We collected cuticular hydrocarbons (CHCs) from three males and three females for all of the genotypes used in the behavioral studies, except for the *D. simulans* strain. CHCs are influenced by both temperature and humidity so we wanted to ensure that all collections were done in parallel to the behavioral observations (NOORMAN AND DEN OTTER 2002; SAVARIT AND FERVEUR 2002). Male and female virgins were collected and stored individually and aged 7 days until CHCs were extracted. CHCs were extracted in HPLC grade hexane (99% purity), by washing flies in 8 dram glass vials with 100µl hexane for 1 min on an orbital shaker. The solvent was then transferred into an autosampler vial with a small volume insert. These samples were left in a fume hood for four hours for the solvent to evaporate. After the solvent had evaporated vials were sealed and placed at -20°C until analysis. For analysis CHCs were re-eluted in 50µl hexane with an internal standard (hexacosane C27). This was chosen as an internal standard based on (DEMBECK *et al.* 2015).

To measure plasticity in male CHCs as a product of interacting with different female genotypes, we carried out an experiment where we exposed Z53 and DGRP882 males to females from their own and opposite genotypes, we also kept some males virgin with no contact. After allowing courtship for 10 min we separated the pairs and allowed males to remain in individual

vials for 30 min. Within the 10 minutes all of the flies had copulated except DGRP882 males paired with Z53 males. We had initially tried to avoid copulation, but most pairs copulated rapidly. We ensured however, that we did not see increases in known "female" CHCs confirming that any changes were not the result of female CHCs rubbing off of males, but changes due to male activity. After 30 minutes all individuals were immediately CHC extracted.

To quantify CHCs we ran our samples on a Shimadzu GC 2014 with flame ionization detector. We used an Agilent DB-5 column that was 20m long with 0.18mm inner diameter, and 0.18um film thickness. We injected 2µl og sample into a preheated injection port at 290°C using the AOC-20i autoinjector. After injection the sample was equilibrated at 50°C for 1 min. The oven temperature was increased to 150°C at a rate of 20°C/minute, and then ramped to 300°C at 5°C/min. There was a final hold at 300°C for 10 minutes. All samples were run in splitless mode, with a sampling time of 1 min and split ratio of 20. The carrier and makeup gas was helium and we used a constant linear velocity of 35cm/sec. The purge flow was 3ml/min split. The detector temperature was 340°C and sampling rate was 40cm/sec. This resulted in a chromatogram for each sample where the quantity of each compound is represented as an area under the corresponding peak. The compounds were identified based on Kovats retention index, which compares retention time of known and unknown compounds, and compared to previous reports for CHCs in *D. melanogaster* that captured presumptive Z-type compounds (DEMBECK *et al.* 2015). The area of each peak was divided by the area of the hexacosane (C27) internal standard, so quantities could be compared between samples.

Statistical analyses

Summarizing courtship behavior

To determine copulation latency and the time spent (or percentage of time spent) on each behavior we needed to estimate courtship initiation from the series of behavioral observations. Copulation latency is defined as the time from courtship initiation until copulation so determining courtship initiation was critical for our analyses. We estimated courtship initiation in two ways: the time when the first non "separate" behavior took place or the time after three consecutive non "separate" behaviors took place. The rationale behind the second method was to account for observations where males and females were close and scored as "engaging" but courtship was broken off and there was a long gap before courtship was reinitiated. For the majority of the cases the difference between courtship initiation based on the two methods was minimal (mean=7.4 seconds) and did not influence the overall estimate of copulation latency. For a few observations there was up to 4 minutes difference (max=262.318 seconds) and re-evaluating these outliers it was clear that the first "engaging" was not courtship initiation. Given this comparison, we used the second method as it was more accurate. The time spent in a particular behavior was calculated as the number of timepoints scored as this behavior, divided by the total number of timepoints recorded until copulation occurred. Since we were recorded a behavior at every timepoint every 10 seconds, this made the number of timepoints per minute uniform across trials. Because the length of each trial was unequal and depending on the time to copulation, to compare across trials we analyzed the percentage of time spent in a specific behavior.

Comparison of courtship between recording periods

To ensure that the same behaviors we observed in our initial cataloging (Fall 2018) were consistent with behaviors when we wanted to determine selection (Spring 2019) we compared common genotype combinations between these two collection periods and analyzed Z53 female x Z53 female and DGRP882 female x DGRP882 male interactions (Supplemental Methods).

Quantification of courtship effort (courtship index) in trials that failed to copulate

We focused our efforts on quantifying behavior for interactions that ended in copulation because these were the only useful trials to make correlations between phenotypes and copulation latency (fitness). However, we wanted to ensure that we were justified in not using trials where no copulation occurred as representative of the behavioral sequence. Additionally because no copulations occurred between Z53 females and DGRP882 males we needed to ensure that these males were actively courting to be able to include them in our analyses (Supplemental Methods).

To determine if males that failed to copulate were courting or not in other genotype combinations we compared successful and non-successful matings between Z29 males and either DGRP882 females or Z53 females. The reason we chose this male genotype is that there was an equal proportion of males that did and did not successfully copulate with both female genotypes (Supplemental Methods).

Mating rejection in long-term experiments

After analyzing videos we observed that there were several genotype combinations that failed to copulate. We expected Z53 to reject known M-type males and consistent with this prediction we did not observe copulations between Z53 females and DGRP882 or Canton S males. There were several genotype combinations that we did not a priori expect to fail including

Z53 females x LZ21 males and DGRP882 females with Z30 or LZ21 males. To determine if this was a function of our short recording time we set up single-pair matings between these genotypes and measured the time until progeny were first seen over a 16 day period. Because virgin females can lay unfertilized eggs we do not focus on egg production but note that in intrastrain control crosses, Z53 females generally start laying eggs later than DGRP882 females. This could indicate that Z53 takes longer to mate in general (Supplemental Information).

Plasticity analysis

To determine if male trait values for both behavioral traits and CHCs changed with respect to which female genotype a male was courting, we looked for evidence of consistent phenotypic plasticity across genotypes. We evaluated models that included male and female genotype and their interaction on each trait. The male genotype effect would capture differences between males. The female genotype effect would indicate plasticity, if males, on average exhibited different trait values when interacting with different females. This pattern would graphically appear as parallel reaction norms (ROFF 1997). We included the interaction effect into our models because this would demonstrate that males differed in their effects and did not have consistent plasticity.

To test these effects we used aligned ranks transformation (ART) ANOVAs using the ARtools package in R (KAY AND WOBBROCK 2019). These models are non-parametric which was well suited for our analysis since the behavioral variables were bound by 0-1, did not produce normally distributed residuals in standard ANOVAs, and had large differences in variance between groups (heteroskedasticity). All three characteristics would have made standard ANOVA inappropriate (FOX 2016). We favored this non-parametric approach instead of transforming the data since not all transformations maintain additivity and constant variance

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(MCULLAGH AND NELDER 1989; FENG *et al.* 2014) and other modelling approaches, such as quantile regression, are robust to these issues (KOENKER 2005).

We also tested whether there was plasticity in how the courtship behavioral sequence was carried out by males when interacting with different females. We quantified this difference using transitions between behavioral states. Behavioral sequences can be viewed as Markov chains, and analyzing transition matrices in this Markov chain framework have been useful to determine differences in a behavior sequence in courtship behavior (MARKOW 1987; GAERTNER *et al.* 2015). First we constructed a transition matrix by summing all of the transitions across all trials for a specific female genotype x male genotype combination. Then we tested whether the transition matrix for a given male genotype differed when interacting with the different female genotypes, using a homogeneity test from the markov package in R (SPEDICATO 2017). If these matrices were not homogeneous we could infer that the overall transition rates were different but not which particular transitions were responsible for this pattern. To determine if there were consistent transition differences we looked for plasticity of individual transitions after a variable reduction process using the VSURF package in R (GENUER *et al.* 2019).

For the CHC plasticity analysis we first needed to carry out dimension reduction to avoid multiple testing issues. While principal components (PC) are a convenient way to summarize this multivariate data, and PC scores can be used directly in analyses, this strategy suffers from the inability to examine/interpret the effects of individual variables. Instead we used the same random forest procedure that we used for the transition data. For this classification problem CHCs could be explained by a categorical variable that captured the interaction of male x female genotype (6 levels for this experiment, see methods above). From the remaining variables we carried out ART ANOVA testing for male, female and male x female genotype effects.

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Selection acting on male traits

To determine if directional selection was acting on particular male traits and whether it was acting in the same direction across the two female genotypes, we used quantile regression to determine relationships between the male trait and copulation latency (our proxy for mating success and fitness). We used quantile regression to condition our regression model on the median rather than the mean, which is useful for non-normally distributed data (KOENKER 2005). We used the package quantreg in R (KOENKER 2019) to fit models that included effects of female genotype, the trait of interest, and the interaction of genotype and trait on copulation latency. The effect of female genotype often captured the fact that the distribution of traits was not identical between female genotypes. One reason was plasticity (see results) the other was that not all genotypes successfully copulated with both females. As a result the same male genotypes were not present for both female genotypes. The effect of the continuous trait on copulation latency was a slope and can be interpreted as the selection gradient. The interaction term is the change in slope/selection gradient that occurs when interacting with the second female genotype. Because we did not measure CHCs for each male used in the courtship trials we used the mean to represent the CHC of a given male genotype. To be consistent across all of our male traits we correspondingly used the mean for behavioral traits for each male genotype as well. This was also necessary given the differences in sample sizes between male genotypes for the number of successful trials.

For behavioral traits we used quantile regression on engaging, circling, and attempted-copulation. Because singing-2 was infrequent and correlated with time spent singing, we combined these into a single metric. We did not estimate regressions for the separate behavior, as this is simply the remaining proportion after accounting for the other courtship behaviors. The

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courtship behaviors were not strongly correlated within a male genotype, or between female genotypes so we decided to analyze each trait independently (Supplemental Table 1) . For the analysis of CHCs on copulation latency we used random forest to reduce the number of variables before completing regression. The VSURF package only allows for a single effect that is either continuous or categorical, which precluded variable selection on our full model of interest. To circumvent this issue, we examined predictive relationships between the copulation latency and the amount of a specific CHC compound using the complete dataset (copulation latency across all genotype combinations), and reduced datasets that represented the individual female genotypes. In this way we expected to identify CHCs that might be important to one, but not the other female genotype. We then estimated the selection gradients on the selected variables and reported confidence intervals for each parameter, to demonstrate how/if the selection gradient changes between female genotypes. We also included the compounds cis-vaccenyl acetate (cVA) and 7-tricosene (7-C23) because of their reported importance to mating interactions in *D. melanogaster* (GRILLET *et al.* 2012)*,* but these were not identified as important to selection from the random forest variable reduction.

For a subset of behavioral traits and CHCs we also wanted to determine if selection was acting on the probability of mating occurring, rather than on the speed by which it occurred. For this analysis we used binomial regression with the number of successful and unsuccessful copulations per female genotype x male genotype combination as the dependent variable. For behavioral traits we focused on the scissoring and circling behaviors since they did not show a linear response for the Z-type female genotype (see results), but appear to be Z-type specific behaviors. We also included singing since this is thought to dominate M-type courtship (COBB *et al.* 1985; TALYN AND DOWSE 2004). For CHCs we focused on 7-C23 and 5-tricosene (5-C32),

because previous studies have shown they are involved in mating probability (GRILLET *et al.* 2012) as well as cVA.

Correlations Between Courtship Traits and Cuticular Hydrocarbons

Females that strongly choose Z-type over M-type males often have the specific CHC profiles that are potentially correlated with their genotype at the *desat2* locus (TAKAHASHI *et al.* 2001; FANG *et al.* 2002; CHERTEMPS *et al.* 2006; COYNE AND ELWYN 2006a). Z-type female preference and male acceptance by a Z-type female are expected to be correlated (HOLLOCHER *et al.* 1997). We wanted to determine if there were traits that were correlated and might be diagnostic of Z-type males. First we estimated the correlation between phenotypic distance of CHCs for male and female genotypes. We assumed the amount of the Z specific female compound 5,9-heptacosadiene (5,9-HD) would be diagnostic of Z-type female. We then used principal component analysis (PCA) on the combined male courtship and behavior data to see which traits were strongly correlated in the principal components.

Results

Courtship of Z-type and M-type males is significantly different

We quantified the behavior of a standard *D. melanogaster* M-type strain (DGRP882), a typical Z-type strain (Z53), and one strain of *D. simulans* (SA22) to determine similarities and differences among these lineages. To our knowledge, all previous studies of male courtship behavior in *D. melanogaster* used only M-type strains. We identified several behaviors in the Z-type strain that did not occur in the M-type strain, but do occur in other species of the *melanogaster* subgroup. This included scissoring, where males open and close their wings rapidly (Supplemental Video 2), similar to scissoring found in *D. simulans* (as defined in COBB

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et al. 1985). We also observed circling, where males move around the female in an arc while vibrating or scissoring at the same time (COBB *et al.* 1985; Supplemental Video 1). In addition, the overall time spent on behaviors common between the two *D. melanogaster* genotypes was significantly different. Specifically, the Z-type strain spent less time singing compared to the M-type strain, but more time singing than *D. simulans* (Supplemental Fig. 3). For circling, we did not observe this behavior in the M-type strains but there was ample variation across Z-type strains.

Z-type and M-type strains are reproductively isolated

We next wanted to assess whether discrimination by Z-females is due to differences in courtship intensity of M-type and Z-type males. We observed that there were no differences in male willingness to court females of either genotype, measured as courtship index, that could explain reproductive isolation between Z-type females x M-type males. Despite active male courting, Z53 females strongly rejected both DGRP882 and Canton S M-type males, consistent with previous reports of asymmetric reproductive isolation in this system (WU *et al.* 1995). This rejection persisted over longer experimental timeframes than the limited time we video recorded. In order to expand our ability to assay matings between strains, we used production of progeny as a proxy for mating success. When we compared control matings between Z53 females x Z53 males with matings between Z53 females and several other genotypes, we saw significant differences in the number of days until we first observed eggs and larvae (Supplemental Fig. 4A). In crosses with Z53 females, larvae appeared in vials from Z53 males and LZ21 males starting on the same day, which we would not have predicted from the video recording trials where no LZ21 males mated with Z53 females. Larvae took significantly longer to appear in crosses between Z53 females and the M-type males DGRP882 and Canton S as well as for ZH33

males. Due to the small sample sizes and crossing survival curves, we could not determine if these male genotypes were significantly different, but qualitatively fewer larvae were produced when M-type males (DGRP882 and Canton S) mated with Z53 females over the course of the experiment (Supplemental Figure 4A).

Based on the literature we would not have expected reproductive isolation between M-type females and Z-type males (WU *et al.* 1995; COYNE AND ELWYN 2006). Nevertheless, there were some genotypes that did not mate within the 30 minute video recordings, so we again measured reproductive isolation using progeny production. For crosses with DGRP882 females we did not perform a within strain control, but instead used ZH33 males as our baseline because they quickly produced both eggs and larvae indicating that they mated within the first 24 hours (Supplemental Fig. 4B). Compared to ZH33 males, LZ21 and Z30 males took significantly longer to produce larvae and in fact very few matings were successful over the course of the 16 day experiment (Supplemental Fig. 4B). This level of reproductive isolation was similar in magnitude to the Z53 female x M-type male crosses.

Male courtship behavior is plastic

Having found significant differences between the courtship of Z53 and DGRP882 males, we next tested whether male courtship is a static or plastic trait. Using a panel of Z-type and M-type males we quantified the differences in behaviors when males interacted with either a Z- or M-type female. For the majority of the male genotypes, the time spent carrying out specific behavioral traits was female-genotype dependent. The two traits that showed consistent plasticity were attempted-copulation and singing (Supplemental Table 2). The strongest signal was for singing, where males consistently spent more time singing when courting the M-type female

(Figure 1). Engaging and circling also had significant genotype effects but there was not a consistent pattern of plasticity across genotypes (Supplemental Table 1).

Courtship plasticity changes the behavioral sequence

Given that the time spent on behaviors was plastic, this could affect the behavioral sequence and transitions between behaviors. We therefore compared transition matrices for male genotypes interacting separately with the two female genotypes. Seven out of the twelve male genotypes had significantly different transition matrices (Supplemental Table 3). The transitions that contributed to this plasticity were singing-attempted-copulation and engaging-singing (Supplemental Table 4). When interacting with Z-type females the male genotypes transitioned more from singing to attempted-copulation and transitioned less from engaging to singing (Supplemental Fig. 5). This reflects that males were singing less in this interaction. The transition between singing-scissoring had significant differences between male genotypes, but was not female-genotype dependent (Supplemental Table 4).

Male cuticular hydrocarbons are plastic

We next tested the hypothesis that males change their CHCs upon exposure to females by quantifying CHCs in Z53 and DGRP882 males that had either been kept virgin or were exposed to Z53 and DGRP882 females. After variable reduction we were left with five compounds from the original twenty-one that we had identified, four of which had significant genotype and treatment effects (Supplemental Table 5). The compounds 5-C25 and 9-C25 showed similar patterns of plasticity, and the effect was mainly seen for Z53 (Supplemental Table 5). Z53 males had reductions in these compounds when exposed to either female strain. The other two compounds had changes that depended on both the male and female genotype. Males reduced their amount of the compound 7-C23 when they were exposed to females of the opposite type.

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For 2-Me-C30, DGRP882 males increased their amount of this compound with DGRP882 females, but Z53 males decreased this compound with DGRP882 females.

Divergent selection acts on courtship traits

To test for directional selection on courtship traits within a lineage, and divergent selection between lineages, we used quantile regression to determine the relationships between the six behavioral traits and copulation latency. We assumed shorter courtship latency was a proxy for increased mating success. A negative regression coefficient indicates a positive selection gradient and a trait that is favored by a specific female genotype. Attempted-copulation was the only trait that had divergent selection gradients for the two female genotypes. There was a significant relationship between increased attempted-copulation and shorter copulation latency (positive selection gradient) for Z53 females, and the opposite pattern for DGRP882 females (Figure 2A; Supplemental Table 6). For singing we only observed selection in one female background. There was no relationship for DGRP882 females, but there was a significant relationship between more singing and longer copulation latency (negative selection gradient) for Z53 females (Figure 2B; Supplemental Table 6). Lastly, for both genotypes there was a relationship between more scissoring and longer copulation latency. This relationship was not significantly different between the female genotypes (Fig. 2C; Supplemental Table 6).

In addition to testing for linear relationships we also wanted to determine if courtship traits predicted whether a female would copulate (successes vs failures) using binomial regression. We analyzed the effects of singing, scissoring, and circling on the probability of copulation. This type of selection can occur if there is a threshold for acceptance of mating. For singing, the pattern for Z53 females was similar to the linear selection analysis, where more singing decreased the probability of copulation (Supplemental Table 7). In this analysis we did

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observe divergent selection for DGRP882 females, where more singing increased the probability of mating (Supplemental Table 7). The time spent scissoring was not related to the probability of copulation for DGRP882 females, but did increase the probability of copulation for Z53 females (Supplemental Table 7). The time spent circling did not have significant effects on the probability of copulation for either female genotype.

Divergent selection acts on cuticular hydrocarbons

We estimated directional selection of cuticular hydrocarbons (CHCs) to test which compounds contributed to mating success, and if there were differences between the female genotypes. After variable reduction we reduced the number of CHCs we examined from twenty-one to five compounds. We included two additional compounds cis-vaccenyl acetate (cVA) and 7-tricosene (7-C23) based on *a priori* expectations of their impact on mating success. Four compounds had divergent selection gradients that were female-genotype dependent. For tetracosane (n-C24), 9-pentacosene (9-C25), and 2-methyl-hexacosane (2-Me-C26) there was a significant relationship between greater quantities of these compounds and decreased copulation latency for Z53 females and an increased copulation latency for DGRP882 females (Figure 3; Supplemental Table 8). For cVA the opposite pattern was true, greater quantities reduced copulation latency for DGRP882 females and increased copulation latency for Z53 females (Figure 3D; Supplemental Table 8). The remaining two compounds did not have significant relationships with copulation latency.

Previous studies indicated that some CHCs, including 7-C23 and 5-tricosene (5-C23), predict the probability of successful copulation (GRILLET *et al.* 2012), so we used binomial regression to test the hypothesis that the amounts of 7-C23, 5-C23, or tricosane (n-C23) predict probability of copulation. These compounds had no effect on copulation probability for DGRP882 females, but 7-C23 and n-C23 had significant effects on decreasing copulation

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probability for Z53 females (Supplemental Table 9). When we analyzed cVA in binomial regression the results were consistent with the linear regression. Greater quantities of cVA increased mating probability with DGRP882 females and reduced mating probability with Z53 females (Supplemental Table 9).

Variation in courtship traits results in discrete clusters of Z-type male genotypes

Given the behavioral trait differences between the male genotypes and their impacts on mating success, we wanted to determine if there was a suite of traits and CHCs that together could be diagnostic of a Z-type male. Most of our information on Z-type flies comes from female choice and CHC experiments, so we first determined the distributions of the C27 isomers 7,11-heptacosadiene (7,11-HD) and 5,9-heptacosadiene (5,9-HD) in our strains. The distributions of these compounds had large variation separating the two known M-type strains (DGRP882 and Canton S) and ZH42 from the other Z-type strains (Supplemental Fig. 6). The M-type strains had high quantities of 7,11-HD and low quantities of 5,9-HD. Most of the suspected Z-type genotypes produced less 7,11-HD than the M-type strains, but there was a large range in the quantity of 5,9-HD produced by these genotypes (Supplemental Fig. 6). We determined if there was a phenotypic correlation between CHCs of males and females using Euclidean distances with respect to the Z53 strain (r=0.561, P=0.044). This captures the overall CHC differences in genotypes without relying on a single compound. Z53 males and females both had the largest phenotypic distance from DGRP882, and across all genotypes the CHC phenotypic distances were greater for female compounds than male compounds. The correlation was not as strong as might have been expected though, because of genotypes that are divergent from both DGRP882 and Z53 in phenotypic space (Supplemental Fig. 7).

Given this variation in CHC phenotypes we determined using principal component (PC) analysis the relationships between male CHCs and male behavioral traits when courting Z53 females. While genotypes were spread across the PC space, there was clustering based on the loadings of specific combinations of behaviors and CHCs (Figure 4). For example, one cluster included ZH42 and DGRP882 and the loadings that contributed were Singing, cVA and 7-C23. In the opposite direction in PC space, Z53 and a few other Z-type strains clustered with contributing loadings including Attempted-Copulation and C25 isomers. The last main cluster included Z30, LS4-7, and other strains that were mostly separated by the Scissoring behavior and additional CHC compounds. Overall, the different clusters suggest there is not a single courtship and CHC combination that can diagnose all Z-type males.

Discussion

While sexual selection appears to be an important driver of speciation based on macroevolutionary patterns, connecting contemporary divergent sexual selection to reproductive isolation has been challenging. We leveraged lineages of *D. melanogaster* with a panel of strains to quantify selection on relevant male courtship traits for females from two diverging lineages. We comprehensively documented differences in male behavior and cuticular hydrocarbons (CHCs) between the M-type and Z-type lineages that allowed us to test for selection on these suites of traits. We confirmed that the time spent singing and the amount of the hydrocarbon cVA are under positive selection in M-type lineages. We then showed that these traits are under negative selection in Z-type lineages, where increasing time spent singing or increased amounts of cVA increased copulation latency. This example demonstrates the role of these traits, and others we document, as potential contributors to reproductive isolation. Interestingly, even

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though M-type strains are thought to mate indiscriminately, which suggests that few traits could be under negative selection in this lineage, we found several traits that were favored by the Z-type strain and disfavored by the M-type strain. Together our results demonstrate that in early divergent lineages, there are connections between sexual selection and reproductive isolation, and that variation in courtship traits can be maintained by both male plasticity and variation in female preference.

Female mate choice is often correlated with the presence of a conspicuous fixed difference between species (MCKINNON AND RUNDLE 2002; COYNE AND ORR 2004; QVARNSTROM *et al.* 2010), but in young divergent lineages, trait variation may segregate between populations (MALLET 2008; HENDRY *et al.* 2009; MEROT *et al.* 2017). As a result, both quantitative differences and the presence or absence of traits could shape reproductive isolation. Most of the male genotypes we used were collected from Africa but we did not expect them to have uniform phenotypes. Given the recent and historical geneflow of European ancestry back into Africa (POOL *et al.* 2012) one might expect several of these strains to be more M-type or intermediate in phenotype. Indeed, we saw this with ZH42, which was phenotypically very similar in both behavior and CHCs to the M-type strains (Fig. 4). There was large phenotypic variation among the remaining strains, and for both courtship behavior and CHCs, we identified examples of traits where we could quantify divergent selection between lineages or selection within a single lineage. Interestingly this did not necessarily correspond to the presence/absence of the trait between lineages.

We observed that Z-type strains carry out two behaviors, scissoring and circling, that we did not see in M-type strains. Circling was not under selection in either lineage, but scissoring had a more interesting pattern. Scissoring has previously been observed in *D. melanogaster*

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strains from the Bahamas and Caribbean that contain African ancestry, but it was a minor part of their courtship (YUKILEVICH AND TRUE 2008). Analyzing our panel of presumably Z-type strains we uncovered genotypes that display the scissoring behavior at a higher frequency. Increased time spent scissoring increased the probability of copulation with Z-type females, but for linear selection increased time spent scissoring was under negative selection for both M-type and Z-type females. In fact, we saw very strong reproductive isolation between the M-type female and males from the Z-type strain that displayed the most scissoring. This level of reproductive isolation was as strong as the isolation that is typical between Z-type females and M-type males (Supplemental Fig. 4). Scissoring thus provides an interesting example where a lineage-specific trait may contribute to reproductive isolation and shape female mate preference, but is not under divergent linear selection.

Both sexual selection and environmental selection can shape patterns of CHC divergence (HIGGIE et al. 2000; GREENBERG et al. 2003; CHUNG et al. 2014), and these traits are examples of magic traits, contributing to both ecological divergence and reproductive isolation (SERVEDIO et al. 2011). We identified several CHCs that were under divergent sexual selection between lineages. We achieved this by focusing on African and Z-type strains and used data reduction techniques that do not rely on a priori candidate compounds. Previous studies did not determine which CHCs are under positive selection in Z-type lineages, which could be due to the phenotypic distribution of the genotypes that were chosen and the focus on compounds that are typically in high quantities in M-type genotypes, specifically isomers of tricosene (GRILLET et al. 2012). For the intensively studied compound 7-tricosene (7-C23) we recapitulated previous reports that it is under negative selection by Z-type strains (GRILLET et al. 2012), but could not detect positive selection for M-type strains. Since we focused on Z-type strains, they may not

contain high levels of 7-C23 typically observed in M-type lineages. Alternatively, it may be that all strains had the minimum amount of 7-C23 that would ensure acceptance by M-type females. If there is a threshold for mating acceptance based on this compound, it would be consistent with the idea that M-type females mate more indiscriminately (WU *et al.* 1995).

Another mechanism that could contribute to the asymmetry of reproductive isolation in this system is that male courtship, and in part CHCs, are female-genotype dependent. Male courtship is described in seemingly contradictory terms, sometimes as a stereotypical sequence (COBB *et al.* 1985; GAERTNER *et al.* 2015), while other times as plastic (DUKAS AND DUKAS 2012; ARBUTHNOTT *et al.* 2017; FILICE *et al.* 2020). This discrepancy might come from the fact that plasticity is often quantified in terms of courtship effort and intensity, rather than the behavioral sequence. We identified behaviors that were consistently plastic across genotypes and resulted in changes to the fundamental behavioral sequence. We also detected significant plasticity for some CHCs in *D. melanogaster,* which can be further explored in future studies. Plasticity that is female-genotype dependent has the potential to impact how we view sexual selection, by creating scenarios where males can maintain fitness in the presence of variation in female mate preference. Selection can maintain plasticity in recently diverged lineages where males come into contact with multiple female genotypes and gain fitness by adjusting their courtship displays. Asymmetrical reproductive isolation, which is especially common in *Drosophila* (YUKILEVICH 2012) and other systems where both females and males contribute to mate choice, might also reflect plasticity. For example, even though M-type males, such as DGRP882, reduced the time spent singing when they courted Z-type females, they still spent significantly more time singing than the Z-type genotypes and ultimately failed to mate with Z-type females. Female-genotype dependent courtship plasticity suggests that males require

lineage-specific female cues for successful courtship (BARKER 1967; COBB AND JALLON 1990; SEEHOLZER *et al.* 2018). CHCs can both stimulate and inhibit mating (FERVEUR 2005; GRILLET *et al.* 2006; EJIMA *et al.* 2007), and a strong difference exists in the female CHCs of Z-type and M-type females (COYNE *et al.* 1999; DALLERAC *et al.* 2000). The ability for males to change their courtship plastically can alter their courtship in subsequent mating interactions, which are typically not considered in the study of speciation.

The interaction between plasticity and learning can have important impacts on the strength of reproductive isolation and outcomes for speciation (KUJTAN AND DUKAS 2009; VERZIJDEN *et al.* 2012). Successful mating can create positive associative learning (EJIMA *et al.* 2007; GRIFFITH AND EJIMA 2009; KOEMANS *et al.* 2017; ZER-KRISPIL *et al.* 2018). If males adopt a successful courting strategy as a result of this conditioned learning they may show less plasticity in subsequent mating bouts (DUKAS AND DUKAS 2012). If instead, plasticity is maintained across subsequent mating encounters, then gene flow would continue and reproductive isolation would not evolve (KIRKPATRICK AND NUISMER 2004; PFENNIG *et al.* 2010). Plasticity within a species could therefore maintain variation in male courtship and female preference would persist in populations. This segregating variation could potentially facilitate rapid speciation, but only after geographic separation (MENDELSON *et al.* 2014; CASTILLO AND DELPH 2016). We identified large differences in male behaviors and CHCs among Z-type strains, such that we could not define a single behavioral trait and CHC combination that classifies Z-type vs M-type males. This intriguing result suggests that there are segregating preferences in these populations. Variation, similar to what we currently observe, could have facilitated the rapid divergence of M-type strains from Z-type ancestors when these lineages migrated out of

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Africa. This has now created strong asymmetric reproductive isolation that is maintained by ongoing divergent sexual selection.

Acknowledgments

Literature Cited

Andersson, M., 1994 *Sexual Selection*. Princeton University Press, Princeton.

Arbuthnott, D., T. Y. Fedina, S. D. Pletcher and D. E. L. Promislow, 2017 Mate choice in fruit flies is rational and adaptive. Nature Communications 8**:** 13953.

Barker, J., 1967 Factors affecting sexual isolation between *Drosophila melanogaster* and *Drosophila simulans*. American Naturalist 101**:** 277-287.

Barraclough, T. G., P. H. Harvey and S. Nee, 1995 Sexual selection and taxonomic diversity in passerine birds. Proceedings of the Royal Society B-Biological Sciences 259**:** 211-215.

Berdan, E., S. Enge, G. M. Nylund, M. Wellenreuther, G. A. Martens *et al.*, 2019 Genetic divergence and phenotypic plasticity contribute to variation in cuticular hydrocarbons in the seaweed fly Coelopa frigida. Ecology and Evolution**:** 12156-12170.

Boake, C. R. B., M. P. DeAngelis and D. K. Andreadis, 1997 Is sexual selection and species recognition a continuum? Mating behavior of the stalk-eyed fly *Drosophila heteroneura*. Proceedings of the National Academy of Sciences of the United States of America 94**:** 12442-12445.

Boughman, J. W., 2002 How sensory drive can promote speciation. Trends in Ecology & Evolution 17**:** 571-577.

Bretman, A., G. M.J.G. and T. Chapman, 2011 Quick-change artists: male plastic and behavioural responses to rivals. Trends in Ecology & Evolution 26**:** 467-473.

Brooks, R., and J. A. Endler, 2001 Direct and indirect sexual selection and quantitative genetics of male traits in guppies (*Poecilia reticulata*). Evolution 55**:** 1002-1015.

Callander, S., M. D. Jennions and P. R. Y. Backwell, 2012 The effect of claw size and wave rate on female choice in a fiddler crab. Journal of Ethology 30**:** 151-155.

Candolin, U., 2019 Mate choice in a changing world. Biological Reviews 94**:** 1246-1260.

Castillo, D. M., and L. F. Delph, 2016 Male-female genotype interations maintain variaiton in traits important for sexual interactions and reproductive isolation. Evolution 70**:** 1667-1673.

Chenoweth, S. F., H. D. Rundle and M. W. Blows, 2010 The contribution of selection and genetic constraints to phenotypic divergence. American Naturalist 175: 186-196.

Chertemps, T., L. Duportets, C. Labeur, M. Ueyama and C. Wicker-Thomas, 2006 A female-specific desaturase gene responsible for diene hydrocarbon biosynthesis and courtship behavior in *Drosophila melanogaster*. Insect Molecular Biology 15: 465-473.

Chung, H., D. W. Loehlin, H. D. Dufour, K. Vaccaro, J. G. Millar *et al.*, 2014 A Single Gene Affects Both Ecological Divergence and Mate Choice in Drosophila. Science 343: 1148-1151.

Cobb, M., K. Connolly and B. Burnet, 1985 Courtship behavior in the melanogaster sub-group of *Drosophila*. Behavior 95: 203-231.

Cobb, M., and J. M. Jallon, 1990 Pheremones, mate recognition and courtship stimulation in the *Drosophila melanogaster* species subgroup. Animal Behaviour 39: 1058-1067.

Coyne, J. A., and S. Elwyn, 2006a Desaturase-2, environmental adaptation, and sexual isolation in Drosophila melanogaster. Evolution 60: 626-627.

Coyne, J. A., and S. Elwyn, 2006b Does the desaturase-2 locus in Drosophila melanogaster cause adaptation and sexual isolation? Evolution 60: 279-291.

Coyne, J. A., and H. A. Orr, 2004 *Speciation*. Sinauer Associates, Inc.

Coyne, J. A., C. Wicker-Thomas and J. M. Jallon, 1999 A gene responsible for a cuticular hydrocarbon polymorphism in Drosophila melanogaster. Genetical Research 73: 189-203.

Dallerac, R., C. Labeur, J. M. Jallon, D. C. Knipple, W. L. Roelofs *et al.*, 2000 A Delta 9 desaturase gene with a different substrate specificity is responsible for the cuticular diene hydrocarbon polymorphism in Drosophila melanogaster. Proceedings of the National Academy of Sciences of the United States of America 97: 9449-9454.

Dembeck, L., K. Boroczky, W. Huang, C. Schal, R. Anholt *et al.*, 2015 Genetic architechture of natural variation in cuticular hydrocarbon composition in *Drosophila melanogaster*. eLife 4: e09861.

Dixon, S. M., J. A. Coyne and M. A. F. Noor, 2003 The evolution of conspecific sperm precedence in *Drosophila*. Molecular Ecology 12: 1179-1184.

Dukas, R., and L. Dukas, 2012 Learning about propsective mates in male fruit flies: effects of acceptance and rejection. Animal Behaviour 84: 1427-1434.

Ejima, A., B. P. C. Smith, C. Lucas, W. V. D. G. van Naters, C. J. Miller *et al.*, 2007 Generalization of courtship learning in *Drosophila* is mediated by cis-vaccenyl acetate. Current Biology 17: 599-605.

Fang, S., A. Takahashi and C. I. Wu, 2002 A mutation in the promoter of desaturase 2 is correlated with sexual isolation between drosophila behavioral races. Genetics 162: 781-784.

Feng, C., H. L. Wang, N., T. Chen, H. He, Y. Lu *et al.*, 2014 Log transformation and its implications for data analysis. Shanghai Archives of Psychiatry 26: 105-109.

Ferveur, J. F., 2005 Cuticular hydrocarbons: their evolution and roles in *Drosophila* pheromonal communication. Behavior Genetics 35: 279-295.

Filice, D. C., R. Bhargava and R. Dukas, 2020 Plasticity in male mating behavior modulates female life history in fruit flies. Evolution 74: 365-376.

Fox, J., 2016 *Applied regression analysis and generalized linear models*. Sage Publications.

Fox, R. J., L. Fromhage and M. D. Jennions, 2019 Sexual selection, phenotypic plasticity, and female reproductive output. Philosophical Transactions of the Royal Society B 374: 20180184.

Friard, O., and M. Gamba, 2016 BORIS: a free, versatile open-source event-logging software for video/audio coding and live observations. Methods in Ecology and Evolution 7: 1325-1330.

Gaertner, B. E., E. A. Ruedi, L. J. McCoy, J. M. Moore, M. F. Wolfner *et al.*, 2015 Heritable variation in courtship patterns in *Drosophila melanogaster*. G3-Genes Genomes Genetics 5: 531-539.

Genuer, R., J. M. Poggi and C. Tuleau-Malot, 2019 VSURF: Variable Selection Using Random Forests. R package version 1.1.0. https://CRAN.R-project.org/package=VSURF.

Greenberg, A. J., J. Moran, J. A. Coyne and C. I. Wu, 2003 Ecological adaptation during incipient speciation revealed by precise gene replacement. Science 302: 1754-1757.

Griffith, L. C., and A. Ejima, 2009 Courtship learning in *Drosophila melanogaster*: diverse plasticity of a reproductive behavior. Learning and Memory 16: 743-750.

Grillet, M., L. Dartevelle and J. F. Ferveur, 2006 A Drosophila male pheromone affects female sexual receptivity. Proceedings of the Royal Society B-Biological Sciences 273: 315-323.

Grillet, M., C. Everaerts, B. Houot, M. G. Ritchie, M. Cobb *et al.*, 2012 Incipient speciaiton in *Drosophila melanogaster* involves chemical signals. Scienctific Reports 2: 224.

Hendry, A. P., D. I. Bolnick, D. Berner and C. L. Peichel, 2009 Along the speciation continuum in sticklebacks. Journal of Fish Biology 75: 2000-2036.

Higgie, M., S. F. Chenoweth and M. W. Blows, 2000 Natiral selection and the reinforcement of mate recognition. Science 290: 519-521.

Hill, G. E., 1991 Plumage coloration is a sexually selected indicator of male quality. Nature 350: 337-339.

Hoffmann, A. A., 1999 Is the heritability for courtship and mating speed in *Drosophila* (fruit fly) low? Heredity 82: 158-162.

Hohenlohe, P. A., and S. J. Arnold, 2010 Dimensionality of mate choice, sexual isolation, and speciation. Proceedings of the National Academy of Sciences of the United States of America 107: 16583-16588.

Hollocher, H., C. T. Ting, F. Pollack and C. I. Wu, 1997 Incipient speciation by sexual isolation in Drosophila melanogaster: Variation in mating preference and correlation between sexes. Evolution 51: 1175-1181.

Jennions, M. D., and M. Petrie, 1997 Variation in mate choice and mating preferences: a review of causes and consequences. Biological Reviews 72: 283-327.

Kapopoulou, A., S. P. Pfeifer, J. D. Jensen and S. Laurent, 2018 The demographic history of African *Drosophila melanogaster*. Genome Biology and Evolution 10: 2338-2342.

Kay, M., and J. Wobbrock, 2019 ARTool: Aligned Rank Transform for Nonparametric Factorial ANOVAs R package version 0.10.6. https://github.com/mjskay/ARTool.

Kelley, C. D., 2018 The causes and evolutionary consequences of variation in female mate choice in insects: the effects of individual state, genotypes and environments. Current Opinion in Insect Science 27: 1-8.

Kelley, C. D., and M. D. Jennions, 2011 Sexual selection and sperm quantity: meta-analyses of strategic ejaculation. Biology Reviews 86: 863-884.

Kirkpatrick, M., 1982 Sexual selection and the evolution of female choice. Evolution 36: 1-12.

Kirkpatrick, M., and S. L. Nuismer, 2004 Sexual selection can constrain sympatric speciation. Proceedings of the Royal Society B-Biological Sciences 271**:** 687-693.

Kirkpatrick, M., and V. Ravigne, 2002 Speciation by natural and sexual selection: models and experiments. The American Naturalist 159**:** S3.

Koemans, T. S., C. Oppitz, R. A. T. Donders, H. van Bokhoven, A. Schenck *et al.*, 2017 Drosophila Courtship Conditioning As a Measure of Learning and Memory. Journal of Visualized Experiments 124**:** e55808.

Koenker, R., 2005 *Quantile Regression*. Cambridge University Press, Cambridge.

Koenker, R., 2019 quantreg: Quantile Regression. R package version 5.54. https://CRAN.R-project.org/package=quantreg.

Kujtan, L., and R. Dukas, 2009 Learning magnifies individual variation in heterospecific mating propsensity. Animal Behaviour 78**:** 549-554.

Kurtovic, A., A. Widmer and B. J. Dickson, 2007 A single class of olfactory neurons mediates behavioral responses to a *Drosophila* sex pheromone. Nature 446**:** 542-546.

Lande, R., 1981 Models of speciation by sexual selection on polygenic traits. Proceedings of the National Academy of Sciences of the United States of America-Biological Sciences 78**:** 3721-3725.

Langerhans, R. B., and A. M. Makowicz, 2013 Sexual selection paves the road to sexual isolation during ecological speciaiton. Evolutionary Ecology Research 15**:** 633-651.

Li, H., and W. Stephan, 2006 Inferring the demographic history and rate of adaptive substitution in Drosophila. Plos Genetics 2**:** e166.

Maan, M. E., O. Seehausen, L. Soderberg, L. Johnson, E. A. P. Ripmeester *et al.*, 2004 Intraspecific sexual selection on a speciation trai, male coloration, in the Lake Victoria cichlid *Pundamilia nyererei*. Proceedings of the Royal Society B-Biological Sciences 271**:** 2445-2452.

Maan, M. E., O. Seehausen and J. J. M. van Alphen, 2010 Female mating preference and male coloration covary with water transparency in a Lake Victoria cichlid fish. Biological Journal of the Linnean Soceity 99**:** 398-406.

Mallet, J., 2008 Hybridization, ecological races and the nature of species: empirical evidence for the ease of speciation. Philosophical Transactions of the Royal Society B 363**:** 2971-2986.

Markow, T. A., 1987 Behavioral and sensory basis of courtship in *Drosophila melanogaster*. Proceedings of the National Academy of Sciences of the United States of America 84**:** 6200-6204.

McKinnon, J. S., and H. D. Rundle, 2002 Speciation in nature: the thresspine stickleback model systems. Trends in Ecology & Evolution 17**:** 480-488.

MCullagh, P., and J. A. Nelder, 1989 *Generalized linear models*. Chapman & Hall, Boca Raton, Florida.

Mendelson, T. C., M. D. Martin and S. M. Flaxman, 2014 Mutation-order divergence by sexual selection: diversification of sexual signals in similar environments as a first step in speciation. Ecology Letters 17**:** 1053-1066.

Merot, C., C. Salazar, R. M. Merrill, C. D. Jiggins and M. Joron, 2017 What shapes the continuum of reproductive isolation? Lessons from *Heliconius* butterflies. Proceedings of the Royal Society B-Biological Sciences 284**:** 20170335.

Moller, A. P., and J. J. Cuervo, 1998 Speciation and feather ornamentation in birds. Evolution 52**:** 859-869.

Noorman, N., and C. J. Den Otter, 2002 Effects of relative humidity, temperature, and population density on production of cuticular hydrocarbons in housefly *Musca domestica*. Journal of Chemical Ecology 28**:** 1819-1829.

Oh, K. P., and K. L. Shaw, 2013 Multivariate sexual selection in a rapidly evolving speciation phenotype. Proceedings of the Royal Society B-Biological Sciences 280**:** 20130482.

Otte, T., M. Hilker and S. Geiselhardt, 2018 Phenotypic plasticity of cuticular hydrocarbon profiles in insects. Journal of Chemical Ecology 44**:** 235-247.

Otto, S. P., M. R. Servedio and S. L. Nuismer, 2008 Frequency-dependent selection and the evolution of assortative mating. Genetics 179**:** 2091-2112.

Panhuis, T. M., R. Butlin, M. Zuk and T. Tregenza, 2001 Sexual selection and speciation. Trends in Ecology & Evolution 16**:** 364-371.

Pauers, M. J., and J. S. McKinnon, 2012 SExual selection on color and behavior within and between cichlid populations: implications for speciation. Current Zoology 58**:** 475-483.

Petfield, D., S. F. Chenoweth, H. D. Rundle and M. W. Blows, 2005 Genetic variance in female condition predicts indirect genetic variance in male sexual display traits. Proceedings of the National Academy of Sciences of the United States of America 102**:** 6045-6050.

Pfennig, D. W., M. A. Wund, E. C. Snell-Rood, T. Cruickshank, S. C. Schlichting *et al.*, 2010 Phenotypic plasticity's impacts on diversification and speciation. Trends in Ecology & Evolution 25**:** 459-467.

Pool, J. E., R. B. Corbett-Detig, R. P. Sugino, K. A. Stevens, C. M. Cardeno *et al.*, 2012 Population Genomics of sub-saharan *Drosophila melanogaster*: African diversity and non-African admixture. Plos Genetics 8**:** e1003080.

Price, T., 1998 Sexual selection and natural selection in bird speciation. Philosophical Transactions of the Royal Society B 353**:** 251-260.

Qvarnstrom, A., A. M. Rice and H. Ellegran, 2010 Speciation in *Ficedula* flycathers. Philosophical Transactions of the Royal Society B 365**:** 1841-1852.

Rebar, D., and R. L. Rodriguez, 2013 Genetic variation in social influence on mate preferences. Proceedings of the Royal Society B-Biological Sciences 280**:** 20130803.

Rebarm, D., N. W. Bailey and M. Zuk, 2009 Courtship song's role during female mate choice in the filed cricket *Teleogryllus oceanicus*. Behavioral Ecology 20**:** 1307-1314.

Ritchie, M. G., 2007 Sexual selection and speciation. Annual Review of Ecology, Evolution, and Systematics 38**:** 79-102.

Roff, D. A., 1997 Phenotypic plasticity and reaction norms in *Evolutionary Quantitative Genetics*. Springer, Boston, MA.

Ryan, M. J., and A. S. Rand, 1993 Species recognition and sexual selection as a unitary problem in animal communication. Evolution 47**:** 647-657.

Savarit, F., and J. F. Ferveur, 2002 Temperature affects the ontogeny of sexually dimorphic cuticular hydrocarbons in *Drosophila melanogaster*. Journal of Experimental Biology 205**:** 3241-3249.

Schluter, D., and T. Price, 1993 Honest, perception and population divergence in sexually selected traits. Proceedings of the Royal Society B-Biological Sciences 253**:** 117-122.

Scott, D., A. Shields, M. Straker, H. Dalrymple, P. K. Dhillon *et al.*, 2011 Variation in the male pheromones and mating success in wild caught *Drosophila melanogaster*. Plos ONE 6**:** e23645.

Seeholzer, L. F., M. Seppo, D. L. Stern and V. Rutta, 2018 Evoluton of a central neural circuit underlies *Drosophila* mate preference. Nature 559**:** 564-569.

Selz, O. M., R. Thommen, M. E. R. Pierotti, J. M. Anaya-Rojas and O. Seehausen, 2016 Differences in male coloration are predicted by divergent sexual selection between populations of a cichlid fish. Proceedings of the Royal Society B-Biological Sciences 283**:** 20160172.

Servedio, M. R., G. S. van Doorn, M. Kopp, A. M. Frame and P. Nosil, 2011 Magic traits in speciation: 'magic' but not rare? Trends in Ecology & Evolution 26**:** 389-397.

Spedicato, G., 2017 Discrete time markov chains with R. The R Journal 9**:** 84--104.

Steiger, S., and J. Stokl, 2014 The role of sexual selection in the evolution of chemical signals in insecrts. Insects 5**:** 423-438.

Svensson, E. I., F. Eroukhmanoff and M. Friberg, 2006 Effects of natural and sexual selection on adaptive population divergence and premating isolation in a damselfly. Evolution 60**:** 1242-1253.

Takahashi, A., S. C. Tsaur, J. A. Coyne and C. I. Wu, 2001 The nucleotide changes governing cuticular hydrocarbon variation and their evolution in Drosophila melanogaster. Proceedings of the National Academy of Sciences of the United States of America 98**:** 3920-3925.

Talyn, B. C., and H. B. Dowse, 2004 The role of coursthip song in sexual selection and species recognition by female *Drosophila melanogaster*. Animal Behaviour 68**:** 1165-1180.

Taylor, M. L., N. Wedell and D. J. Hosken, 2007 The heritability of attractiveness. Current Biology 17**:** R959-960.

van Doorn, G. S., U. Dieckmann and F. J. Weissing, 2004 Sympatric pseciation by sexual selection: a critical reevaluation. American Naturalist 163**:** 709-725s.

Verzijden, M. N., C. Ten Cate, M. R. Servedio, G. M. Kozak, J. W. Boughman *et al.*, 2012 The impact of learning on sexual selection and speciation. Trends in Ecology & Evolution 27**:** 511-519.

Watts, J. C., A. Flynn, B. Tenhumberg and E. A. Hebets, 2019 Contemporary sexual selection does not explain variaiton in male display traits among populations. Evolution 73**:** 1927-1940.

Wilkins, M. R., H. Karaardic, Y. Vortman, T. L. Parchman, T. Albrecht *et al.*, 2016 Phenotypic differentiation is associated with divergent sexual selection among closely related barn swallow populations. Journal of Evolutionary Biology 29**:** 2410-2421.

Wu, C. I., H. Hollocher, D. J. Begun, C. F. Aquadro, Y. J. Xu *et al.*, 1995 Sexual isolation in *Drosophila melanogaster* - a possible case of incipeint speciation. Proceedings of the National Academy of Sciences of the United States of America 92**:** 2519-2523.

Yukilevich, R., 2012 Asymmetrical patterns of speciation uniquely support reinforcement in *Drosophila*. Evolution 66**:** 1430-1446.

Yukilevich, R., and J. R. True, 2008 African morphology, behavior and pheromones underlie incipient sexual isolation between US and Caribbean *Drosophila melanogaster*. Evolution 62**:** 2807-2828.

Zer-Krispil, S., H. Zak, L. Shao, S. Ben-Shaanan, L. Tordjman *et al.*, 2018 Ejaculation induced by the activation of Crz neurons is rewarding to *Drosophila* males. Current Biology 28**:** 1445-1452.
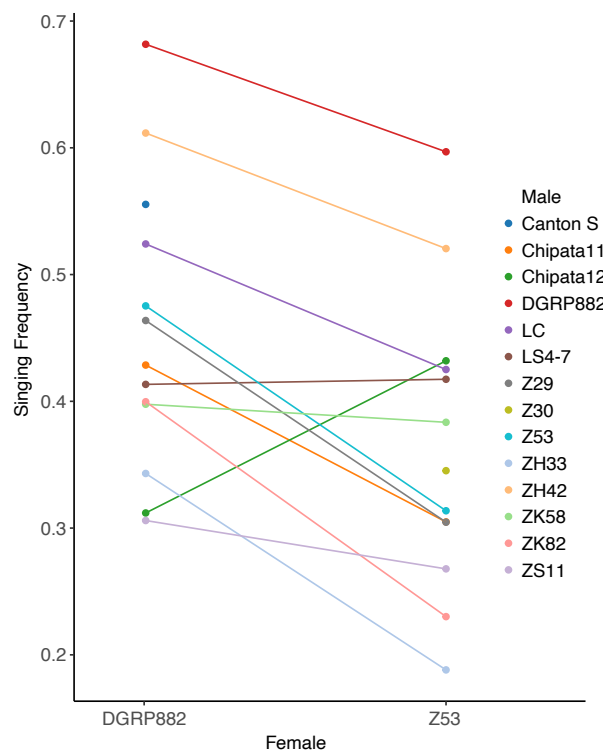
Figure 1. Male courtship singing frequency is female-genotype dependent. There was a consistent decrease in singing frequencies for most strains when males of each genotype were presented to a Z53 female, compared with being presented to a DGRP-882 female. Each point represents the average singing frequency of the given male genotype (indicated by color) when presented to a female of either genotype. Points that occur only in Z53 or DGRP882 female backgrounds did not copulate with the other genotype. This includes Canton S males only copulating with DGRP882 females and Z30 males only copulating with Z53 females. DGRP882 male behavior was analyzed when courting Z53 females even though they did not copulate.
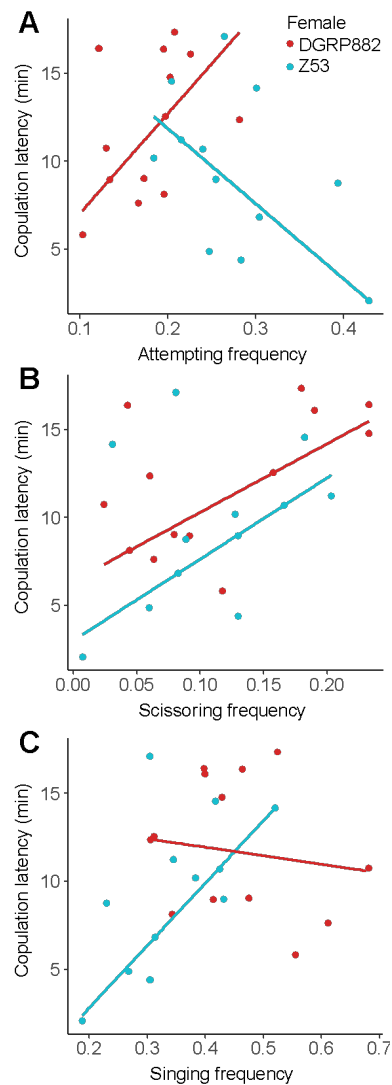
Figure 2. Directional selection on male courtship behavior is female-genotype-dependent. A) There was a negative relationship between attempted-copulation frequency and copulation latency for males interacting with Z53 females and the opposite pattern for males interacting with DGRP-882 females. B) There was a positive relationship between Scissoring frequency and copulation latency for DGRP-882 females but no relationship for Z53 females. C) There was a positive relationship between singing frequency and copulation latency for Z53 females but no relationship for DGRP-882 females. Shorter copulation latency is a proxy for increased mating success. Each point in the figures represents the average behavioral frequency and copulation latency of a male genotype when presented to a female of the given genotype. The regression lines were produced using quantile regression.
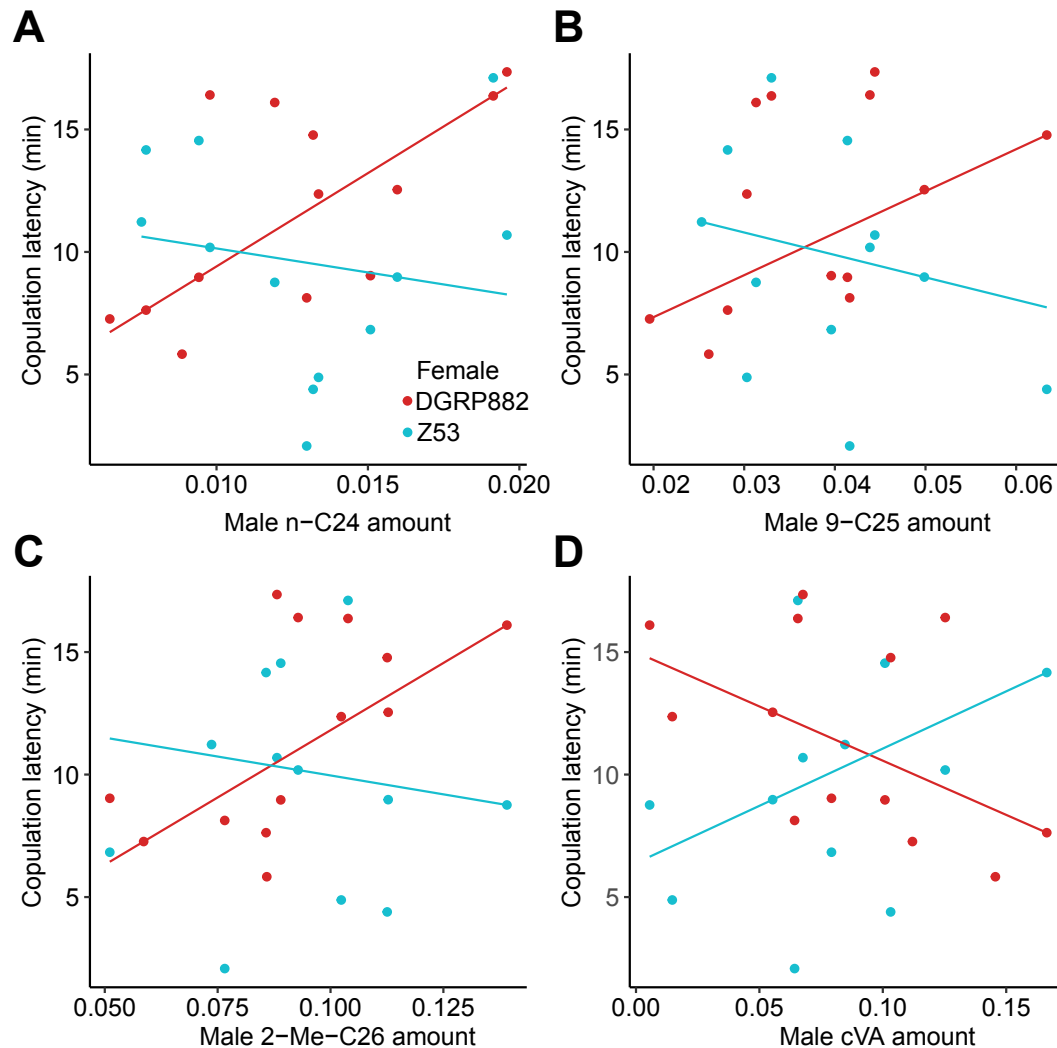
Figure 3. Directional selection on male cuticular hydrocarbons (CHCs) is female-genotype-dependent. There were significant positive relationships for DGRP-882 females and negative relationships for Z53 females for copulation latency and the amounts of three CHCs in males including A) tetracosane, n-C24, B) 9-pentacosene, 9-C25 and, C) 2-methyl-hexacosane, 2-Me-C26. D) The opposite pattern occurred for cVA with a negative relationship between the amount of cVA and copulation latency for males with DGRP-882 females and a positive relationship with Z53 females. Shorter copulation latency is a proxy of increased mating success. Each point in the figures represents the average relative abundance of the corresponding CHC and copulation latency of a specific male genotype, when presented to the indicated female genotype. The regression lines were produced using quantile regression.
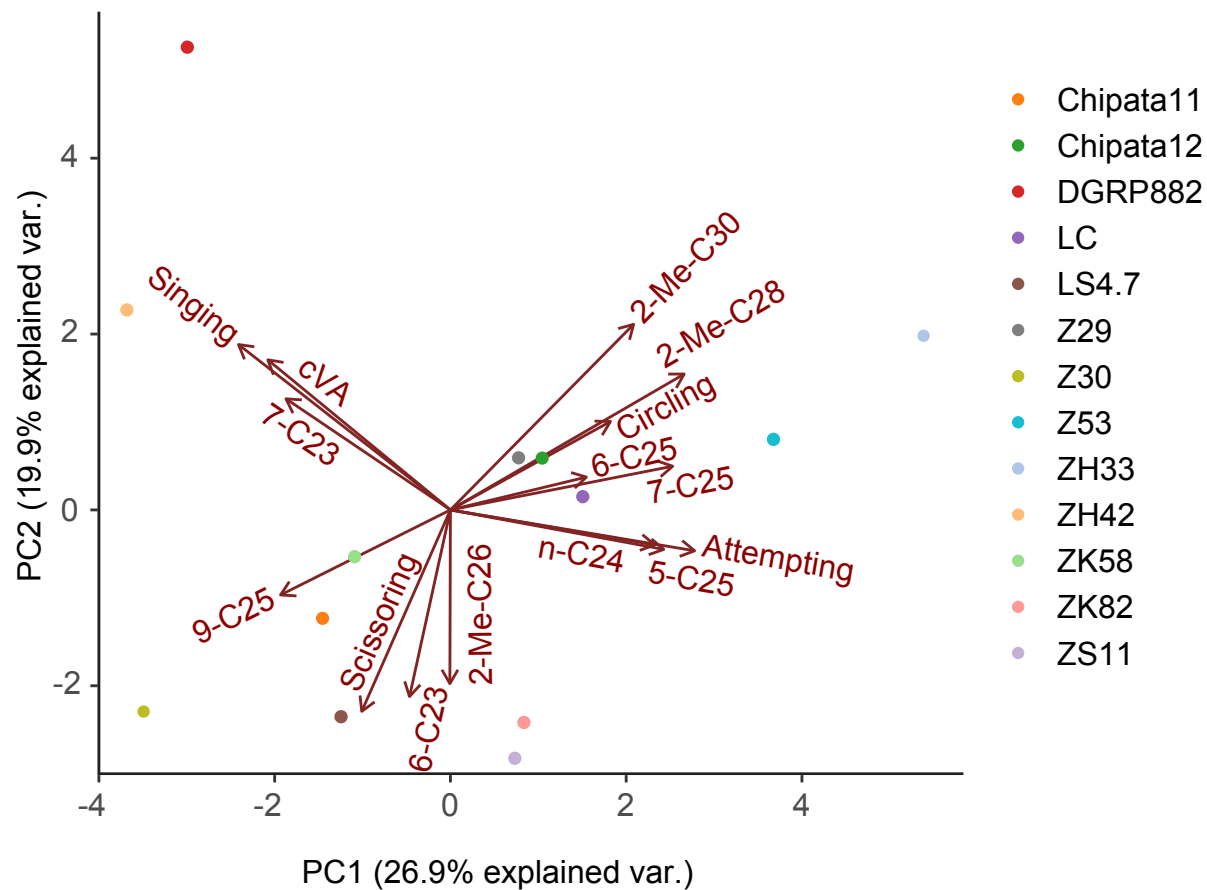
Figure 4. The combination of male cuticular hydrocarbons and behaviors that characterize distinct male courtship phenotypes. The combination of the singing behavior and the cuticular hydrocarbons cVA and 7-tricosene (7-C23) defines the known M-type genotypes. The presumably Z-type males fall into two broad clusters defined by unique behavior and CHC combinations. Each point represents the average value for a given male genotype with behavioral data coming from interactions with Z53 females. Each arrow represents one main variable used to construct the principal components, the length of the arrow represents the loading on a particular principal component axis.