

1 **Expansion and accelerated evolution of 9-exon odorant receptors** 2 **in *Polistes* paper wasps (Hymenoptera: Vespidae)**

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7 **Abstract**

8 Independent origins of sociality in bees and ants are associated with independent expansions of
9 the odorant receptor (OR) gene family. In ants, one clade within the OR gene family, the 9-exon
10 subfamily, has dramatically expanded. These receptors detect cuticular hydrocarbons (CHCs), key
11 social signaling molecules in insects. It is unclear to what extent 9-exon OR subfamily expansion
12 is associated with the independent evolution of sociality across Hymenoptera, warranting studies
13 of taxa with independently derived social behavior. Here we describe OR gene family evolution
14 in the northern paper wasp, *Polistes fuscatus*, and compare it to four additional paper wasp species
15 spanning ~40 million years of divergence. We find 200 functional OR genes in *P. fuscatus*
16 matching predictions from neuroanatomy, and more than half of these are in the 9-exon subfamily.
17 Lineage-specific expansions of 9-exon subfamily ORs are tandemly arrayed in *Polistes* genomes
18 and exhibit a breakdown in microsynteny relative to tandem arrays in other OR subfamilies. There
19 is evidence of episodic positive diversifying selection shaping ORs in expanded subfamilies,
20 including 9-exon, E, H, and L, but 9-exon ORs do not stand out as selectively diversified among
21 *Polistes* species. Accelerated evolution has resulted in lower amino acid similarity and higher d_N/d_S
22 among 9-exon ORs compared to other OR subfamilies. Patterns of OR evolution within *Polistes*
23 are consistent with 9-exon OR function in CHC perception by combinatorial coding, with both
24 selection and drift contributing to interspecies differences in copy number and sequence.

25 **Keywords:** odorant receptor, paper wasp, birth-and-death evolution, comparative genomics,
26 tandem array, antennal lobe glomeruli

27 **Introduction**

28 Chemosensation is an organism's ability to sense chemicals in the environment, whether by
29 olfaction (smell) or gustation (taste). Insects rely on chemosensation when identifying a
30 compatible mate, avoiding toxic microbes, escaping predators, and finding food - whether a patch
31 of nectar-rich flowers or a nutritious caterpillar to predate or parasitize (Wicher 2015; Fleischer et
32 al. 2018; Yan et al. 2020). Social insects further use chemicals to communicate with each other
33 about nest membership, caste, food locations, and alarms (Leonhardt et al. 2016). The evolution
34 of the remarkable chemical communication abilities in social bees and ants is associated with
35 expansion of the odorant receptor (OR) gene family, and the 9-exon OR subfamily in particular
36 has experienced increased gene turnover and sequence evolution relative to other OR subfamilies
37 (Engsontia et al. 2015; Kapheim et al. 2015; Zhou et al. 2015; Karpe et al. 2016, 2017; McKenzie
38 et al. 2016). Comparative studies across species with different evolutionary histories and variable
39 social behaviors can shed light on the molecular evolutionary processes underlying chemical signal
40 perception (Tsutsui et al. 2013; Yan et al. 2020). The independent origins of sociality in wasps
41 provide an opportunity to compare patterns of OR gene family evolution to those that have been
42 observed within social bees and ants (Hines et al. 2007). At the same time, studies of closely related
43 species with similar ecologies and life histories can reveal the dynamics of receptor evolution at
44 finer timescales (Guo & Kim 2007; Brand et al. 2015; Karpe et al. 2016; Brand & Ramirez 2017;
45 Miller CH et al. 2020). A better understanding of the short-term mechanisms of OR evolution
46 provides additional insights into the molecular evolutionary dynamics shaping receptor diversity
47 across more distantly related taxa.

48 The molecular evolution of the OR gene family is best described as a birth-and-death
49 process, in which genes are duplicated and deleted over evolutionary time (Nei 2007; Nozawa &
50 Nei 2007; Eirín-López et al. 2012). The combined forces of random drift and natural selection
51 determine the extent of gene copy number variation and the rate of sequence evolution (Nei 2007;
52 Nozawa & Nei 2007). The behavioral and molecular functions of ORs shape their molecular
53 evolution. Organisms use odorant receptor proteins to detect stimuli and provide input to neural
54 circuits that determine decision-making, here termed behavioral function (Yapici et al. 2014). At
55 the molecular level, the specificity with which an OR binds chemical compounds or families of
56 compounds (ligands) determines the tuning specificity of the olfactory receptor neuron (ORN) in
57 which it is expressed (Hallem et al. 2004). ORN tuning specificity describes the relationship

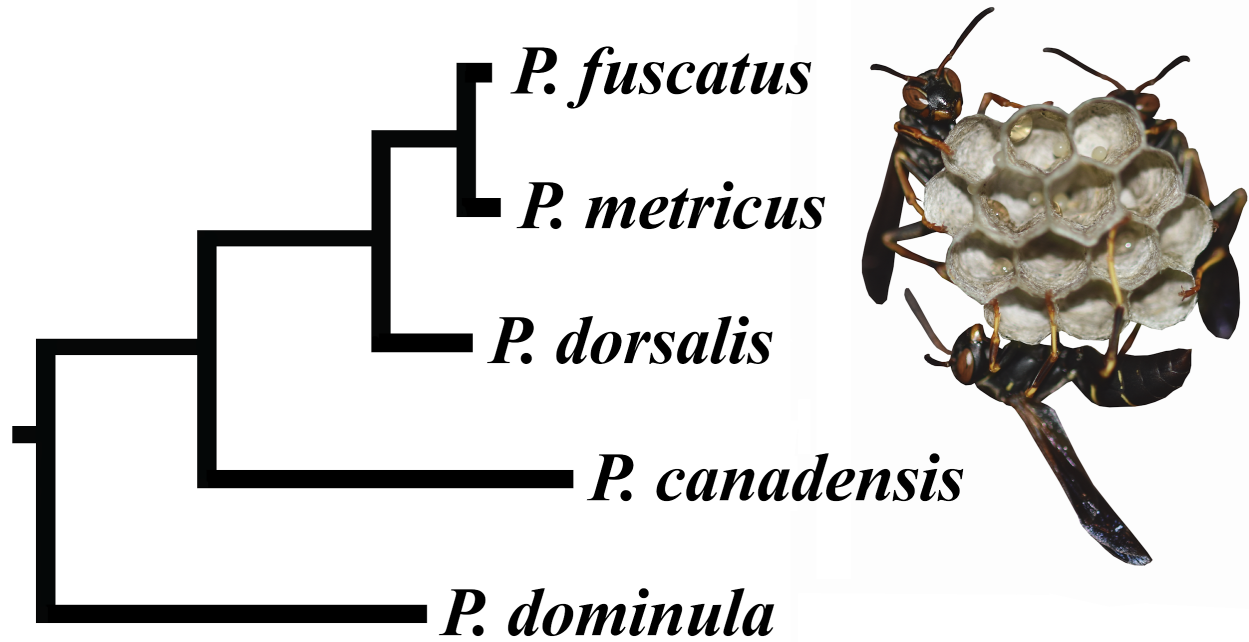
58 between the change in the frequency of action potentials fired by the ORN across a range of
59 different chemical stimuli. An OR's molecular function can be categorized as specialist if ligand
60 binding is highly specific, i.e. one chemical, or generalist if ligand binding allows for detection of
61 multiple chemicals or chemical types. Narrowly tuned ORNs express specialist ORs that bind a
62 specific ligand, creating a dedicated channel of olfaction, while broadly tuned ORNs express
63 generalist ORs that bind a larger spectrum of ligands (Touhara & Vosshall 2009; Andersson et al.
64 2015). Generalist ORs are important for combinatorial coding: the process of combining input
65 from multiple ORs that bind an overlapping set of ligands in order to discriminate a large number
66 of ligands (Malnic et al. 1999). In nature, ORs exist on a spectrum from specialist to generalist,
67 and ORN responses are dependent upon a complex interaction of OR, ligand concentration, and
68 odorant binding proteins (Vogt et al. 1991; Hallem et al. 2004; Hallem & Carlson 2006; Stensmyr
69 et al. 2012; Mathew et al. 2013; Ebrahim et al. 2015; Dweck et al. 2015; Münch & Galizia 2016).

70 There are distinct sets of predictions for the molecular evolution of ORs depending on their
71 behavioral and molecular functions. Negative selection (purifying selection) is expected to
72 conserve specialist ORs, which are not resilient to mutations due to the tight link between receptor
73 and ligand in a dedicated olfactory channel (Andersson et al. 2015). For example, a volatile emitted
74 by toxic microbes triggers avoidance behavior in *Drosophila melanogaster* by binding a specialist
75 OR that is conserved across the genus (Stensmyr et al. 2012). In contrast to dedicated olfactory
76 channels, combinatorial coding is thought to involve ORs with overlapping responses to ligands
77 (Andersson et al. 2015). Mutations that slightly alter the response profiles of functionally
78 redundant ORs may not be eliminated by negative selection, since other ORs can help compensate
79 (Fishilevich et al. 2005; Keller & Vosshall 2007). Copy number variation and relaxed selection
80 allow ORs to gain mutations that might endow them with an adaptive behavioral function.
81 Divergent chemosensory landscapes between species lead to increased copy number variation as
82 ORs with new behavioral functions are gained and ORs that detect irrelevant ligands are lost
83 (Ramdya & Benton 2010; Goldman-Huertas et al. 2015).

84 The molecular evolution of the OR gene family is dynamic among the Hymenoptera, with
85 prevalent lineage-specific gene expansions and losses, especially in the 9-exon OR subfamily
86 (Engsontia et al. 2015; Zhou et al. 2015; McKenzie & Kronauer 2018). The 9-exon ORs constitute
87 about one third of all ant ORs, and have evolved rapidly in ants, frequently under positive selection,
88 leading researchers to propose that 9-exon ORs facilitate recognition of cuticular hydrocarbons

89 (CHCs) (Smith CR, Smith CD et al. 2011; Smith CD, Zimin et al. 2011; Zhou et al. 2012;
90 Engson et al. 2015; Zhou et al. 2015; McKenzie et al. 2016). CHCs are used by insects to
91 waterproof the cuticle and to communicate with conspecifics (Blomquist & Bagnères 2010). While
92 less pronounced than in ants, dynamic evolution is also characteristic of 9-exon OR evolution in
93 social bees, which rely on CHCs in communication (Sadd et al. 2015; Karpe et al. 2016, 2017).
94 Functional studies in which ORs were transfected into an empty *D. melanogaster* ORN have
95 verified that at least some 9-exon ORs of the ant *Harpegnathos saltator* overlap in their responses
96 to ligands, with multiple 9-exon ORs responding to the same CHC molecule and unique 9-exon
97 ORs responding to multiple different CHC molecules (Pask et al. 2017; Slone et al. 2017).
98 Functional ORs are necessary for normal nesting behavior and for nestmate recognition in ants, a
99 process which involves detecting variation in the CHCs on the cuticles of conspecifics (Lavine et
100 al. 1990; van Zweden & d’Ettorre 2010; Sturgis & Gordon 2012; Tribble et al. 2017; Yan et al.
101 2017; Ferguson et al. 2020). Together these studies suggest that 9-exon ORs function in
102 combinatorial coding of CHC perception.

103 Like other Hymenoptera, vespid wasps, including the genus *Polistes*, use CHCs in
104 complex social behaviors (Gamboa et al. 1986, 1996; Dani & Turillazzi 2018). *Polistes* use
105 chemicals as signals and cues in a variety of behaviors, including during mate attraction, mate
106 compatibility recognition, queen recognition, dominance/fertility signaling, and nestmate
107 recognition (Reed & Landolt 1990; Post & Jeanne 1984; Dapporto et al. 2007; Jandt et al. 2014;
108 Sledge et al. 2001a, 2001b, 2004; Oi et al. 2019; Espelie et al. 1994). Recent efforts to sequence
109 *Polistes* genomes provide an opportunity to resolve patterns of OR evolution among closely related
110 species as an independent test of 9-exon OR gene subfamily expansion during social evolution and
111 (Patalano et al. 2015; Standage et al. 2016; Miller SE et al. 2020). We annotated the OR repertoires
112 of five *Polistes* species representing ~40 million years of evolution: *P. fuscatus*, *P. metricus*, *P.*
113 *dorsalis*, *P. canadensis*, and *P. dominula* (Figure 1). Combining neuroanatomy, manual gene
114 annotation, and molecular evolution, we examined the evolution of odorant receptors in light of
115 the patterns predicted for different behavioral and molecular functions. We discover that social
116 wasps, like ants, have an expanded subfamily of 9-exon ORs. Between *Polistes* species, 9-exon
117 ORs exhibit dynamic evolution relative to ORs in other subfamilies, which are highly conserved.
118 Expansion and birth-and-death evolution of the 9-exon OR subfamily in social wasps is consistent
119 with a unique function in combinatorial coding perception of CHCs.



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Fig. 1: Phylogeny of five *Polistes* species considered in this study: *P. fuscatus*, *P. metricus*, *P. dorsalis*, *P. canadensis*, and *P. dominula*. The photo to the right of the phylogeny shows multiple *P. fuscatus* foundresses on a nest. Phylogenetic tree built from 16S ribosomal RNA and cytochrome oxidase subunit I tree in Supplemental Fig 2 in Sheehan et al. 2015.

125 Results and Discussion

126 Antennal Lobe Neuroanatomy and Manual Gene Annotation Predict 200 ORs in *P. fuscatus*

127 In order to predict the OR repertoires of *P. fuscatus* and four other *Polistes* species, we combined
128 fluorescent confocal microscopy of the *P. fuscatus* antennal lobe with manual genome annotation
129 informed by antennal RNAseq. We found 229 glomeruli in the antennal lobe of an adult gyne
130 (female reproductive) (Figure S1). Across a sample of insects, the number of intact OR genes in
131 the genome correlates with the number of glomeruli in the antennal lobe, predicting 229 ORs in
132 the *P. fuscatus* genome (Figure 2). Here we focus on the *P. fuscatus* genome because it has nearly
133 chromosome level scaffolds and is the best assembled *Polistes* genome (Table S1; Patalano et al.
134 2015; Standage et al. 2016; Miller SE et al. 2020). Automated annotation using the MAKER
135 pipeline (Holt & Yandell 2011) without guidance from antennal mRNA predicted 115 OR gene
136 models in the *P. fuscatus* genome. A combined *P. fuscatus* male and gyne (reproductive female)
137 antennal transcriptome generated using Trinity (Haas et al. 2013) yielded 89 OR genes greater than
138 900 nucleotides in length. Some long Trinity genes contain multiple 7-transmembrane domains
139 and likely represent concatenated OR genes. The small fraction of the *P. fuscatus* OR repertoire
140 predicted by transcriptome assembly is consistent with previous observations that annotation of
141 OR repertoires using only transcriptome data typically fails to recover all ORs (Karpe et al. 2016,
142 2017, 2020).

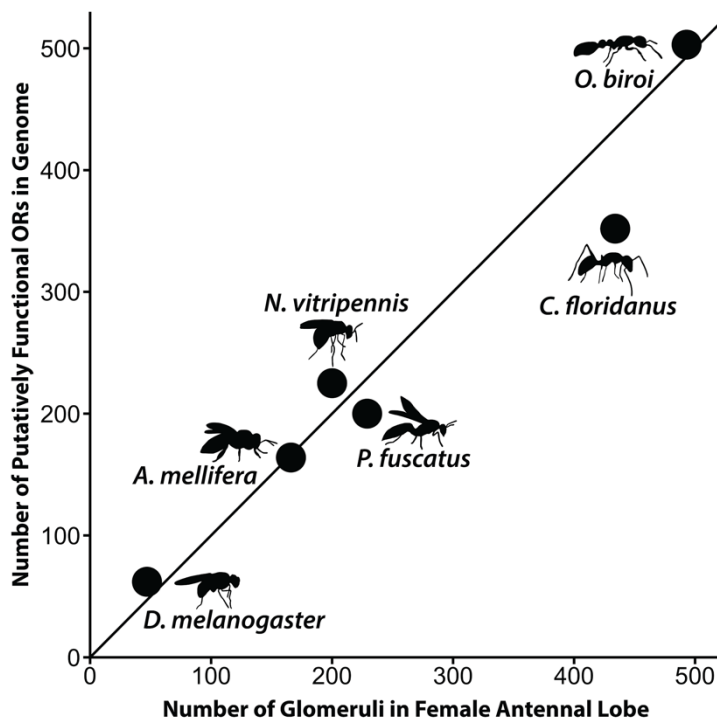


Fig. 2: The number of functional ORs is correlated with the number of antennal lobe glomeruli across insect species (50 glomeruli and 62 ORs in the genome of the common fruit fly *D. melanogaster*, Fishilevich & Vosshall 2005; 166 glomeruli in the worker, 103 glomeruli in the drone, and 163 functional ORs in the genome of the honey bee *A. mellifera*, Arnold et al. 1985, Robertson & Wanner 2006; ~200 glomeruli in females and 225 intact ORs in the genome of the parasitic wasp *N. vitripennis*, Groothuis et al. 2019, Robertson & Wanner 2006; ~434 glomeruli in the worker, 258 in the male, and 352 functional ORs in the genome of the ant *C. floridanus*, Zube & Rössler 2008, Zhou et al. 2012; 493 glomeruli in the worker, 119 glomeruli in the male, and 503 intact ORs in the genome of the ant *O. biroi*, McKenzie et al. 2016; McKenzie & Kronauer 2018; 229 glomeruli in a gyne and 200 putatively functional ORs in the genome of *P. fuscatus*). The diagonal line represents a line of equality with slope of 1. Insect silhouette sizes are not to scale.

164 Manual gene annotation of *P. fuscatus* ORs recovered 231 gene models across 28 scaffolds (Figure
 165 S2), of which 28 are pseudogenes and 10 are incomplete gene models (7 missing N termini, 2
 166 missing C termini, and one missing both N and C termini). Since functional insect ORs are
 167 typically composed of 400 amino acids, we defined gene models as putatively functional if they
 168 coded for proteins greater than or equal to 300 amino acids in length. In *P. fuscatus*, the 200
 169 putatively functional gene models encode protein sequences with an average length of 395 ± 15
 170 (SD) amino acids, and 198 of these gene models encode protein sequences greater than 350 amino
 171 acids in length (Table 1). Odorant receptor proteins possess seven transmembrane domains. The
 172 putatively functional *P. fuscatus* OR proteins possess on average 5.95 ± 0.91 (SD) transmembrane
 173 domains as predicted by TMHMM version 2.0c (Sonnhammer et al. 1998) and 6.43 ± 1.13 (SD)
 174 as predicted by Phobius version 1.01 (Käll et al. 2004). For comparison, transmembrane domain
 175 prediction in 61 *D. melanogaster* ORs coding for proteins greater than 375 amino acids in length
 176 found on average 5.77 ± 1.12 (SD) transmembrane domains as predicted by TMHMM version 2.0c
 177 and 6.18 ± 1.09 (SD) as predicted by Phobius version 1.01 (sequences from Hopf et al. 2015
 178 Supplemental Data 1). The close match between the number of ORs predicted by neuroanatomy
 179 and the number recovered from manual annotation suggests that we have identified nearly all of
 180 the OR genes in *P. fuscatus*. The number of transmembrane domains predicted are comparable to
 181 annotations of *D. melanogaster* and approach the 7 transmembrane domains expected for insect
 182 ORs. Manual OR gene annotation in *P. fuscatus* and four other *Polistes* genomes is summarized
 183 in Table 1:

Species	Putatively Functional ORs (>300 aa)	Mean Length in Amino Acids (\pm SD)	Mean Trans-membrane domains by TMHMM	Mean Trans-membrane domains by Phobius	Total Gene Models	PSE	Incomplete OR gene models
<i>P. fuscatus</i>	200	395 ± 15	5.95 ± 0.91	6.43 ± 1.13	231	28	10
<i>P. metricus</i>	204	396 ± 13	5.96 ± 0.85	6.45 ± 1.17	217	12	9
<i>P. dorsalis</i>	177	393 ± 20	5.90 ± 0.90	6.40 ± 1.21	203	16	24
<i>P. canadensis</i>	188	394 ± 17	5.95 ± 0.91	6.48 ± 1.20	235	13	59
<i>P. dominula</i>	180	392 ± 19	5.99 ± 0.88	6.59 ± 1.33	202	7	33

184 9-exon OR Subfamily Expanded During the Evolution of Social Wasps

185 We conducted a Hymenoptera-wide analysis of OR evolution to test the prediction that 9-exon
186 subfamily ORs were independently expanded during the evolution of eusociality in vespid wasps.
187 By comparing the *P. fuscatus* OR repertoire to other Hymenopterans, our findings reinforce
188 previous results showing that across Hymenopteran families, ORs evolve with lineage-specific
189 expansions of multiple OR subfamilies (Figure 3). Gene gain and loss events were predicted using
190 NOTUNG (Chen et al. 2000) and mapped onto a species cladogram of 14 Hymenopterans (Figure
191 4). NOTUNG estimated an ancestral Apocritan repertoire of 56 ORs, which has expanded
192 independently during the evolution of braconid wasps, ants, bees, and paper wasps (Figure 4). The
193 9-exon subfamily is commonly expanded across Hymenoptera (~90 genes on average), and
194 comprises ~36% of social insect OR repertoires. The largest lineage-specific expansions of
195 Hymenopteran 9-exon ORs have occurred independently during the evolution of ants and social
196 wasps. In *P. fuscatus*, this clade has expanded to 105 genes, comprising 53% of the OR gene set
197 (Figure 4). Given the well-documented use of CHCs as signal molecules in *Polistes* (Singer 1998;
198 Dani et al. 2001; Dani 2009), it is not surprising to find expansions in the CHC-detecting 9-exon
199 subfamily in this genus. Subfamilies L, T, H, E, and V have also expanded in *Polistes*, but not to
200 the extent of the 9-exon OR subfamily.

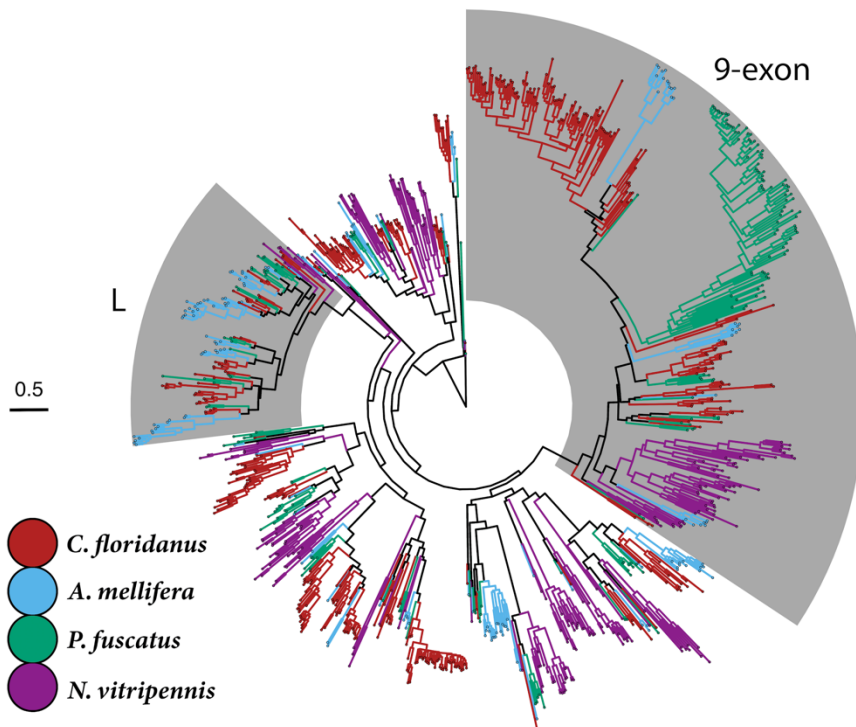
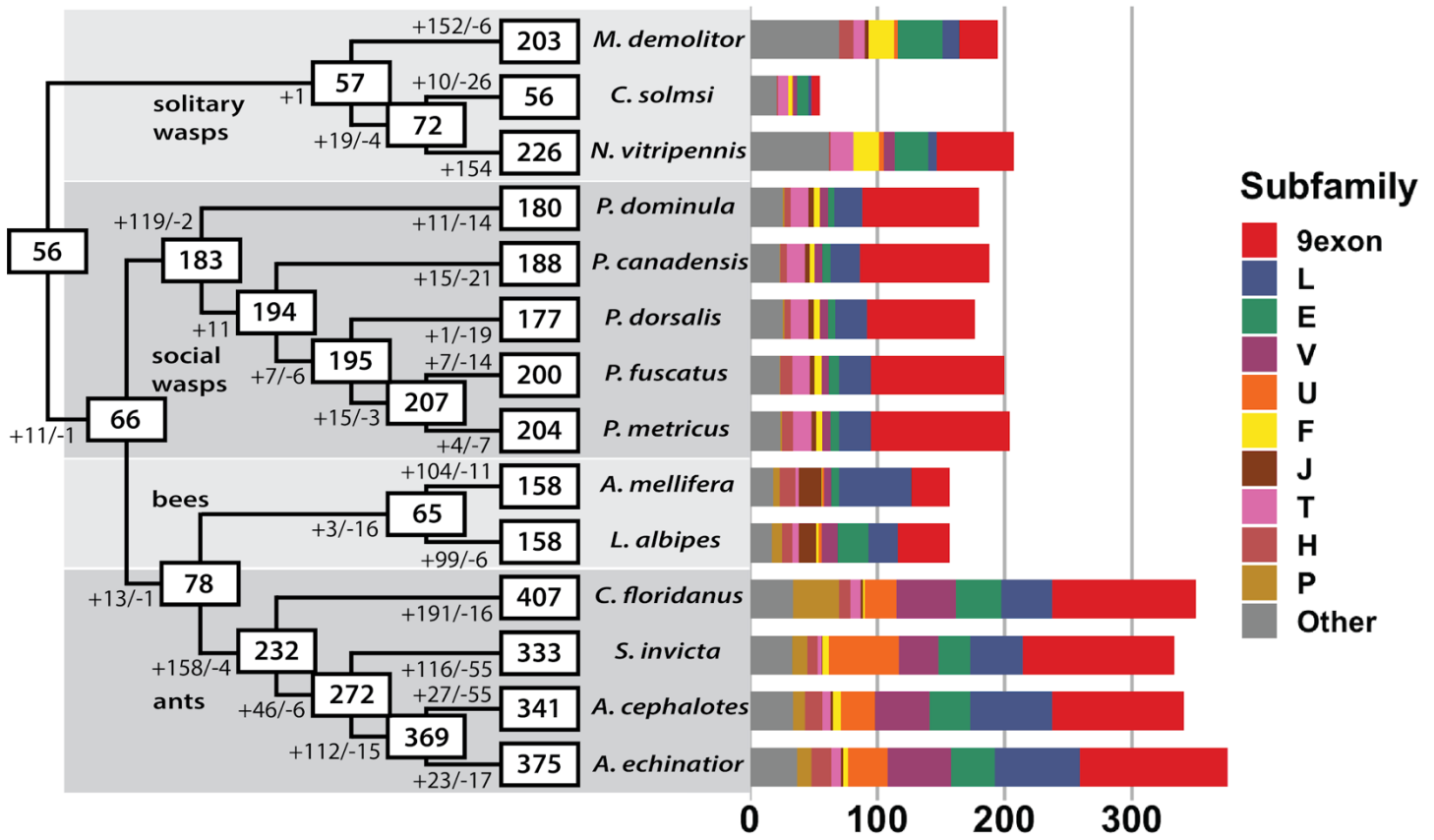


Fig. 3: Maximum likelihood OR protein tree constructed using data from four Hymenopterans (*Apis mellifera*, Robertson & Wanner 2006; *Camponotus floridanus*, Zhou et al. 2012; *Nasonia vitripennis*, Robertson et al. 2010). Branches are colored by species (Red: *Acromyrmex echinator*; Light blue: *A. mellifera*; Green: *P. fuscatus*; Purple: *N. vitripennis*). The L and 9-exon subfamilies are highlighted. Scale bar represents mean substitutions per site.

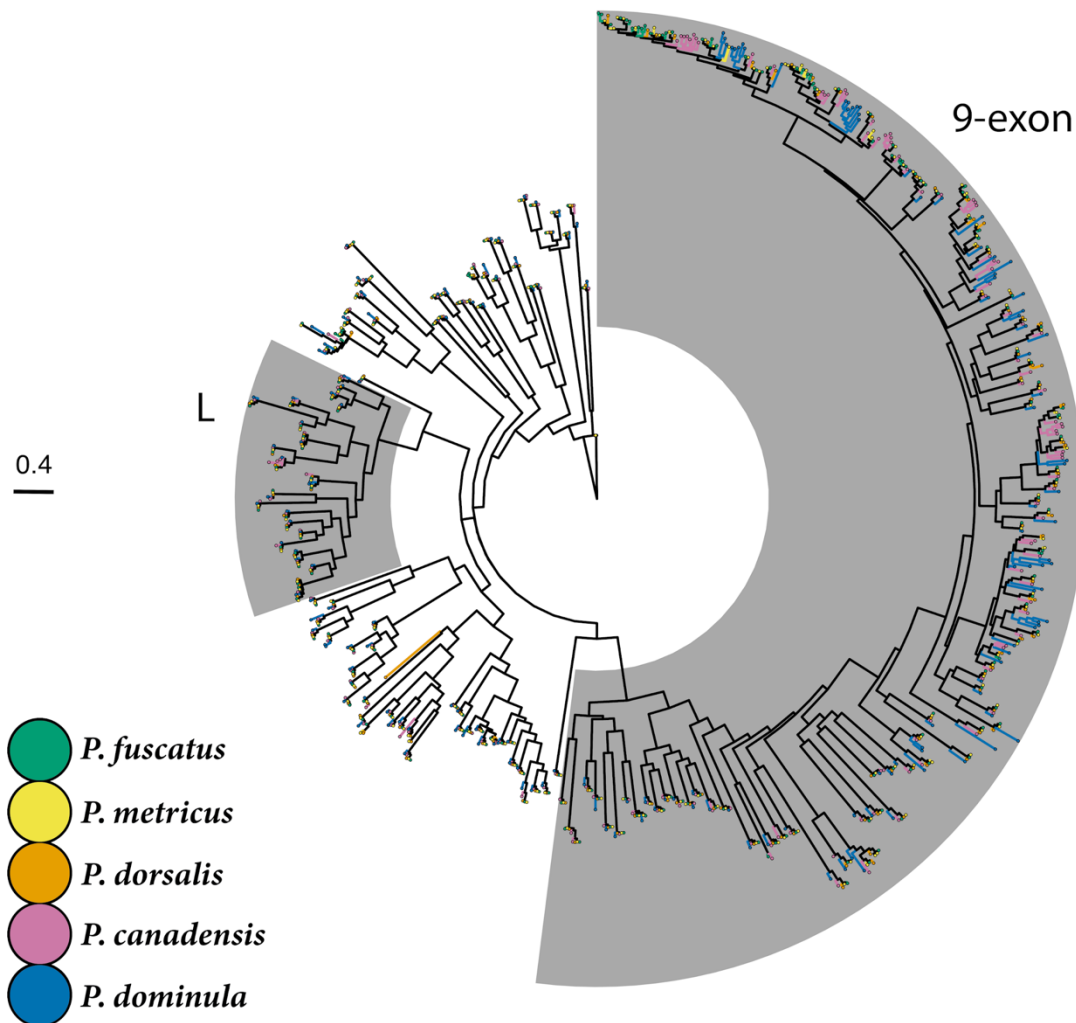


219 **Fig. 4:** Cladogram of Hymenoptera species showing estimated number of OR gene gain and loss events
 220 along branches and estimated size of ancestral and extant species OR repertoires in boxes. To the right is
 221 a bar chart showing numbers of ORs broken down by subfamily. Non-Polistine OR data are from
 222 Robertson et al. 2010 and Zhou et al. 2012, 2015. The set of intact ORs that are greater than 300 amino
 223 acids was used for gene gain and loss estimates. For *C. floridanus*, only ORs considered putatively
 224 functional by Zhou et al. 2012 were used to generate the bar chart showing numbers of ORs broken down
 225 by subfamily.

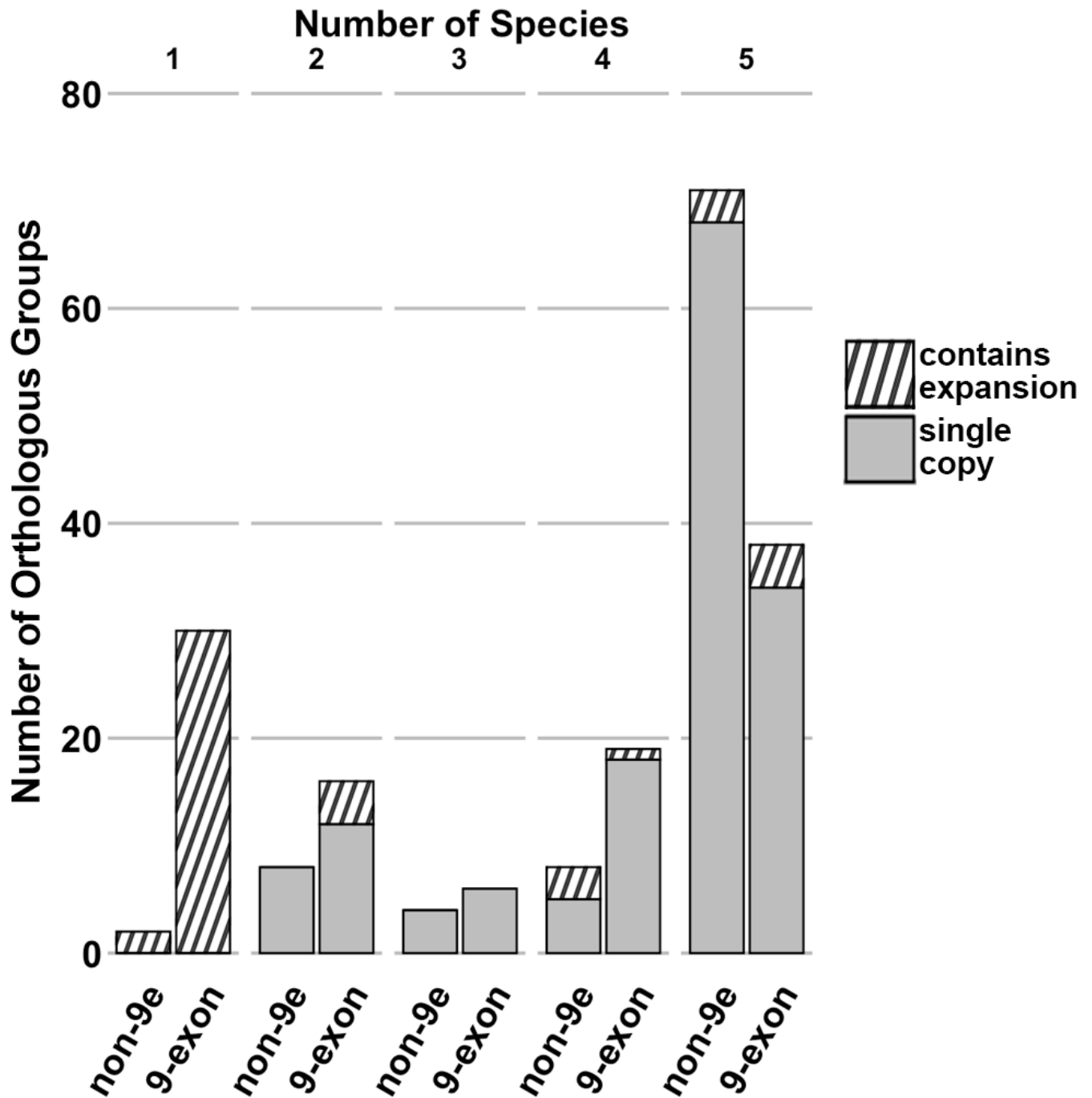
226 The 9-exon Subfamily Shows a Distinct Pattern of Orthology within *Polistes*

227 We next examined the evolutionary history of OR genes among the five *Polistes* species to reveal
 228 patterns of orthology and paralogy within subfamilies. Across the *Polistes* genus, most OR
 229 subfamilies are highly conserved (Figure 5). About 70% of non-9-exon family *P. fuscatus* ORs are
 230 in 1:1 orthology with all other *Polistes* species sampled as predicted by OrthoFinder (Emms &
 231 Kelly 2015; Table S3). The remaining orthologous groups contain an expansion in one or more
 232 species (Figure 6). Considering non-9-exon ORs, most ORs are shared by all five *Polistes* species
 233 examined, and most expansions are shared across all five species. Given that the species examined
 234 here span ~40 million years of divergence (Peters et al. 2017), the conservation of most of the OR
 235 repertoire is notable and may be related to the similarity of ecological and social niches found

236 among *Polistes* wasps. While a common evolutionary history has led to large 9-exon OR
237 complements in all *Polistes* species examined, lineage-specific gains and losses of 9-exon ORs
238 account for most of the variation in OR repertoires size across *Polistes* species (Figure 4). In
239 contrast with the other OR subfamilies, the 9-exon OR subfamily shows more lineage specificity
240 with only 32% of *P. fuscatus* 9-exon ORs showing simple 1:1 orthology across all five *Polistes*
241 examined (Table S3). Most 9-exon subfamily orthologous groups contain gene copies from four
242 or fewer species, and lineage-specific expansions are more common in 9-exon OR orthologous
243 groups (Figure 6). Patterns of orthology (or rather the relative lack thereof) among 9-exon OR
244 genes compared to the rest of the OR gene subfamilies suggest unique evolutionary processes
245 shaping 9-exon ORs.



277 **Fig. 5:** Maximum likelihood OR protein tree with branches colored by species (Green: *P. fuscatus*;
278 Yellow: *P. metricus*; Orange: *P. dorsalis*; Magenta: *P. canadensis*; Blue: *P. dominula*). The L and 9-exon
279 subfamilies are highlighted. Scale bar represents mean substitutions per site.

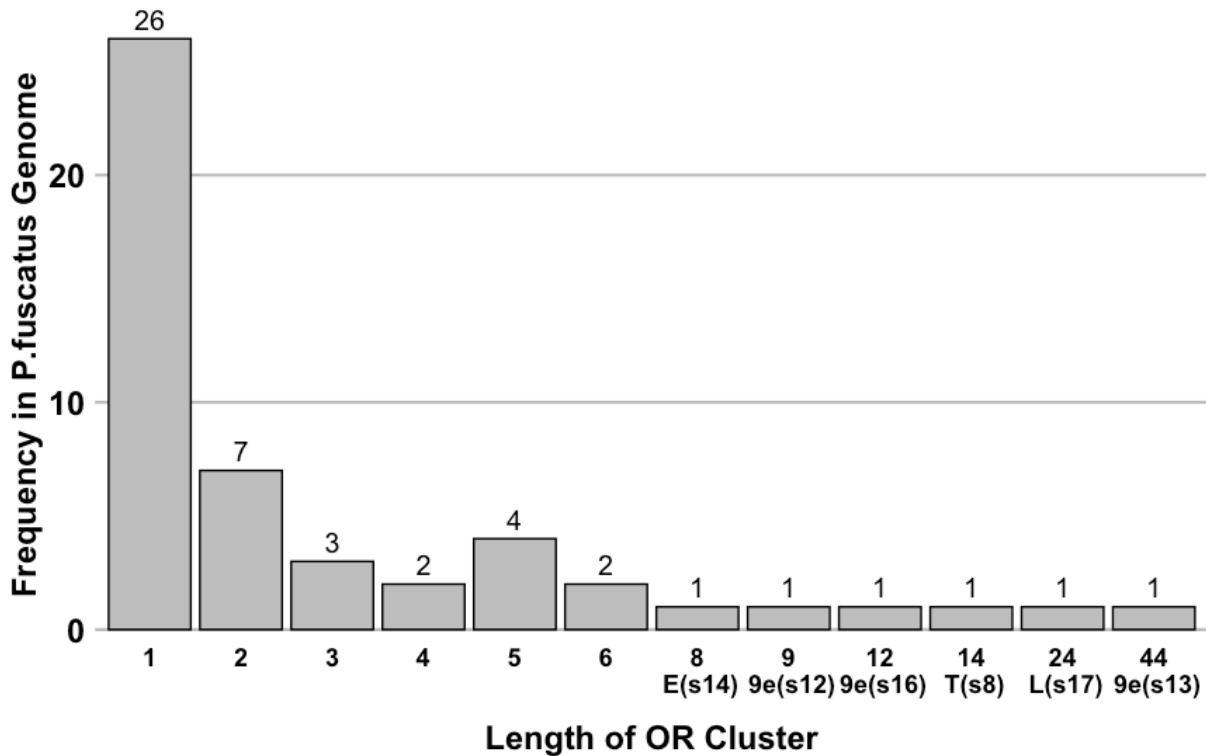


280 **Fig. 6:** Stacked bar chart showing the number of species (x-axis) represented in each orthologous group
281 (y-axis), and whether or not each orthologous group is single copy (shaded bottom portion of bar) or
282 contains an expansion in at least one species (top striped portion of bar). Orthologous groups are split into
283 two categories: non-9-exon orthologous groups (left bar) and 9-exon orthologous groups (right bar).

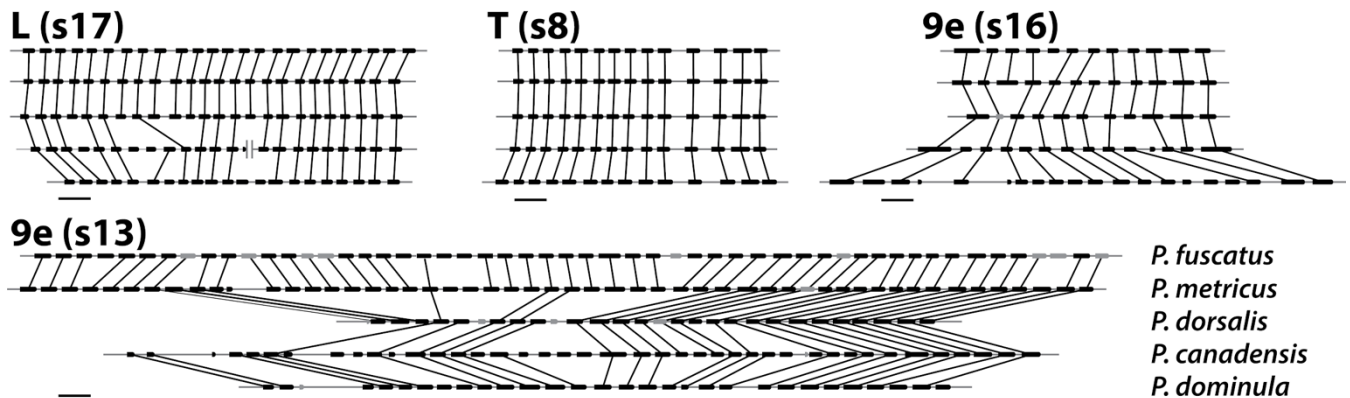
284 Microsynteny Reveals Recent Birth-and-Death Events in *Polistes* 9-exon Subfamily

285 Expanded gene families often occur as tandem arrays, a genomic architecture that can contribute
286 to increased rates of gene birth and death, increasing copy number variation among species (Ohno
287 1970). Therefore, we examined how genomic organization varies between OR subfamilies in
288 *Polistes* species to generate insights into the molecular evolutionary mechanisms shaping OR
289 subfamily function. Genomic organization of ORs across *Polistes* is consistent with a model of
290 birth-and-death evolution shaping OR repertoires. As in bees, gene gain and loss at a small number
291 of loci containing tandem arrays is responsible for most copy number variation in the OR family
292 across closely-related species (Brand & Ramírez 2017). In *P. fuscatus*, 62% of ORs occur in
293 tandem arrays of 6 or more genes (Figure 7A). The frequency of tandem arrays and the tail-to-
294 head orientations of neighboring genes point to tandem duplication as the primary mechanism of
295 OR expansion, likely caused by non-allelic homologous recombination (Lynch 2007; Ramdya &
296 Benton 2010). We examined microsynteny among genes and pseudogenes in the four longest
297 tandem arrays of ORs in *Polistes* genomes (Figure 7B). There is marked decrease in OR synteny
298 among genes in orthologous 9-exon OR arrays compared to tandem arrays of L and T subfamily
299 ORs in the *Polistes* genus. The longest OR gene tandem array in *P. fuscatus* is comprised of 44
300 genes in the 9-exon subfamily on scaffold 13 (s13), which corresponds to homologous arrays of
301 50 genes *P. metricus*, 25 genes in *P. dorsalis*, 33 genes in *P. canadensis*, and 29 genes in *P.*
302 *dominula*. Only 34% of *P. fuscatus* ORs in this array have orthologs across all *Polistes* species
303 sampled (Figure 7B). The second longest OR gene tandem array in *P. fuscatus* contains 24 ORs in
304 the L subfamily on scaffold 17 (s17), and these ORs show 1:1 orthology across *P. fuscatus*, *P.*
305 *metricus*, and *P. dorsalis*, while *P. canadensis* possesses an array of ~23 genes split across two
306 scaffolds, and *P. dominula* possesses an array of 21 ORs at this locus (Figure 7B). This tandem
307 array, widely expanded across Hymenoptera, has been expanded and conserved across *Polistes*.
308 The T subfamily, located on scaffold 8 (s8) of the *P. fuscatus* genome, is composed of 14 tandemly
309 arrayed genes that show 1:1 orthology across five *Polistes* (Figure 7B). Differences in the extent
310 of microsynteny among tandem arrays belonging to different OR subfamilies highlight the unique
311 evolutionary processes shaping 9-exon OR evolution in paper wasps. At the same time, the

312 extreme conservation of L and T subfamily tandem arrays across species highlights the strong
 313 conservation of the OR repertoire outside of the 9-exon subfamily among *Polistes* species.

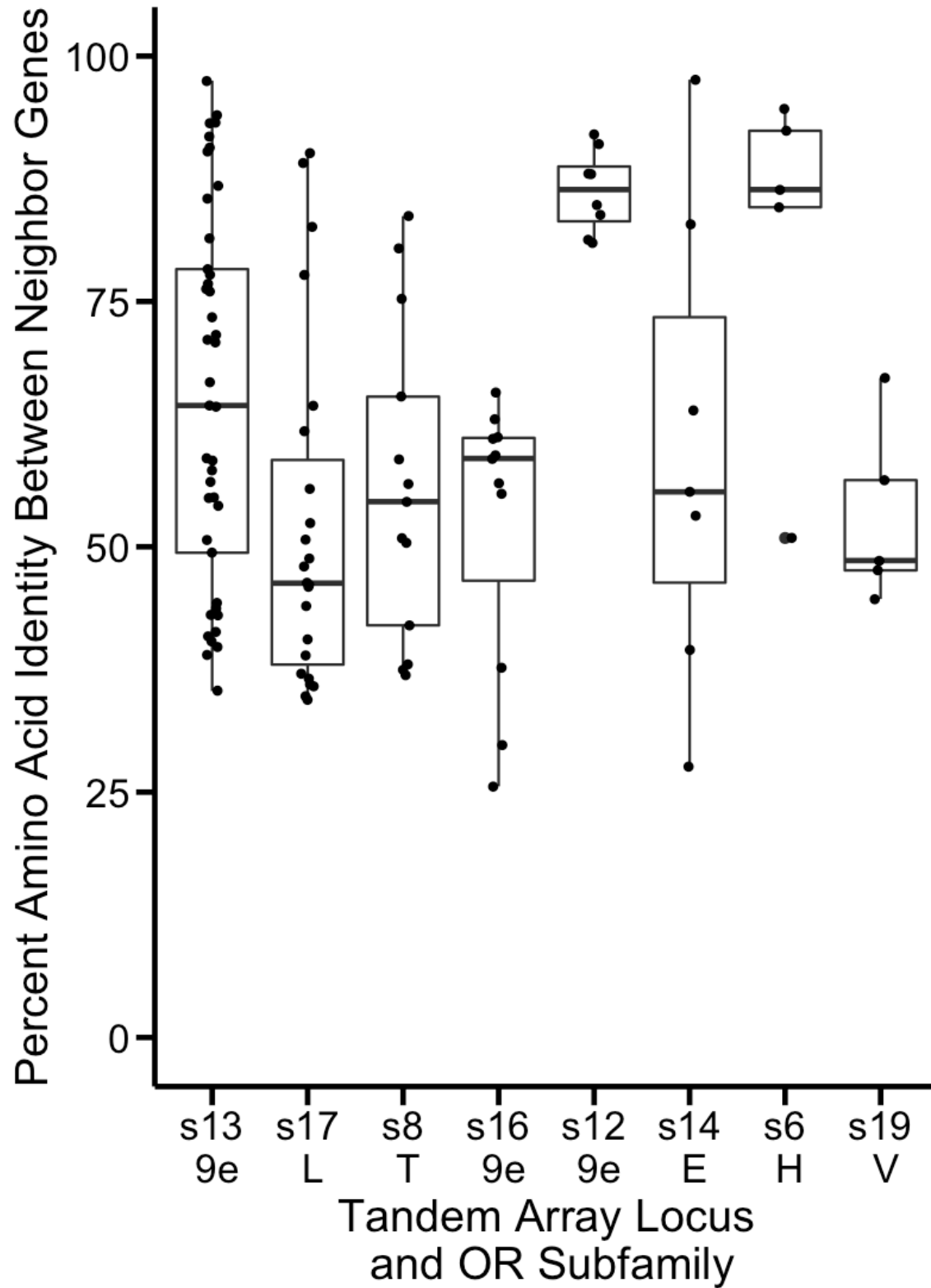


314 **Fig. 7:** (A) Frequency of OR gene singletons and tandem arrays in the *P. fuscatus* genome. 62% of ORs
 315 in *P. fuscatus* occur in tandem arrays of 6 or more genes. The longest tandem array is a 44 gene cluster on
 316 scaffold 13 (s13) containing 9-exon subfamily ORs. The first row of x-axis labels is the number of OR
 317 genes in a tandem array cluster, and the second row labels the OR subfamily and scaffold number
 318 (abbreviated s# in parentheses) of the six longest tandem arrays.



319 (B) Genome alignments of four loci containing tandem arrays of OR genes in all *Polistes* species
 320 examined. Each alignment is labeled with the corresponding subfamily and *P. fuscatus* scaffold number
 321 (abbreviated s# in parentheses). Black boxes represent functional genes and gray boxes represent
 322 pseudogenes. Directionality of genes is denoted by curved corners at their 3' (tail) end. Black lines
 323 connect orthologous genes between species. Genomic scaffolds are represented by horizontal, gray lines,
 324 and scaffold ends are represented by vertical gray lines. The black scale bars represent 5kb.

325 Microsynteny analysis suggests a process of ongoing gene turnover in 9-exon arrays but stasis in
326 most other expanded subfamilies. More recent turnover should be associated with higher pairwise
327 amino acid identity between neighboring genes in an array if they are the result of recent
328 duplication events (Ohno 1970; Bohbot et al. 2007). To explore the relationship between amino
329 acid divergence and tandem array locus, we compared the mean percent amino acid identity among
330 neighboring genes within an array between the eight loci containing the longest tandem arrays of
331 ORs in the *P. fuscatus* genome using one-way ANOVA (Figure 8). Mean percent amino acid
332 identity of neighboring genes was significantly separated by OR array identity (DF = 7; F = 5.39;
333 P = 2.67e-05). Differences between particular OR tandem arrays were identified using Tukey HSD
334 post hoc tests. The mean percent amino acid identity among neighboring genes within one tandem
335 array of nine 9-exon ORs on scaffold 12 (s12) of the *P. fuscatus* genome is higher than in the s13
336 9-exon array (P Adj = 0.04586), the s17 L array (P Adj = 0.00013), the s8 T array (P Adj =
337 0.00458), the s16 9-exon array (P Adj = 0.00124), and the s19 V array (P Adj = 0.02247). The s12
338 9-exon OR array is composed of a larger proportion of pseudogenes (5 PSE, 10 intact gene models)
339 than the other two 9-exon arrays (s13: 9 PSE, 44 intact gene models; s16: 0 PSE, 12 intact gene
340 models). ORs in the s12 9-exon array lack clear orthologous relationships with ORs in species
341 other than *P. metricus*. Taken together, the high within array sequence similarity, high frequency
342 of pseudogenes, and low orthology exhibited by this array indicate that it is the result of one or
343 more recent gene duplication events since the divergence of *P. fuscatus* and *P. metricus*, the most
344 closely related species analyzed here. The s19 H subfamily array also shows higher amino acid
345 sequence identity among neighboring genes than the s17 L subfamily array (P Adj = 0.01625) and
346 the s16 9-exon array (P Adj = 0.04038). Increased amino acid similarity may also occur within
347 older tandem arrays as a result of gene conversion (Nagawa et al. 2002). However, we searched
348 for gene conversion using GENECONV (Sawyer 1989) and did not detect gene conversion events
349 within the s12 9-exon array or in the s19 H array after Bonferroni correction. Patterns of genomic
350 organization of OR genes in *Polistes* genomes lead to the conclusion that gene gain and loss in the
351 9-exon OR subfamily is an ongoing process within this genus, in contrast to the stable and
352 conserved tandem arrays in most other OR subfamilies.



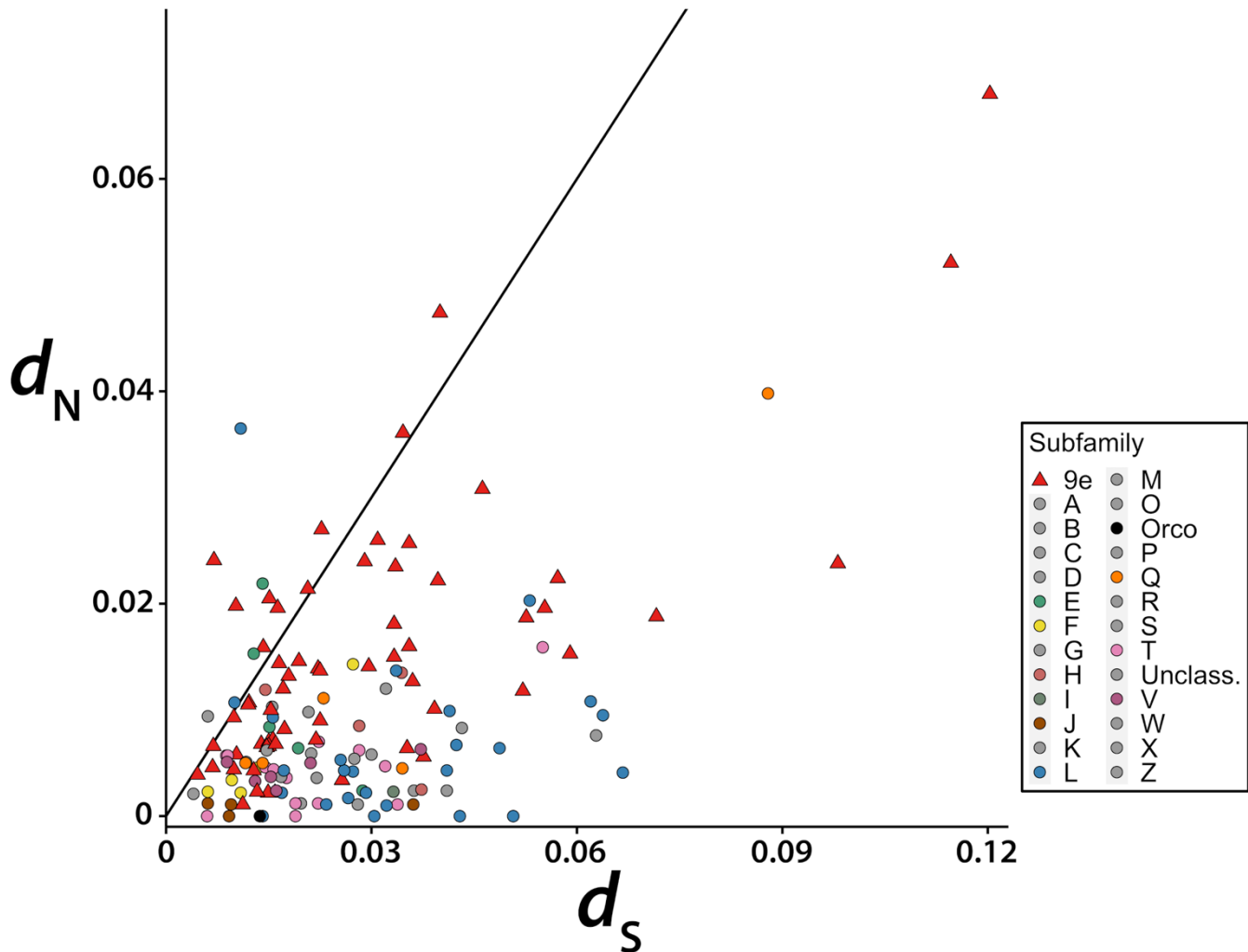
400 **Fig. 8:** Percent amino acid identity between neighboring genes at eight loci containing the longest OR
401 gene tandem arrays in the *P. fuscatus* genome. Arrays are ordered by length in gene number, from longest
402 (44 9-exon ORs) to shortest (6 H ORs and 6 V ORs).

403 Accelerated Evolution of 9-exon ORs and Positive Selection in Expanded OR Subfamilies

404 We examined patterns of OR evolution among five *Polistes* species to test the prediction that OR
405 subfamilies exhibit signatures of positive selection. We were especially interested in whether the
406 recent dynamism in 9-exon OR gene copy number has been accompanied by episodes of positive
407 selection in *Polistes*. HyPhy aBSREL (Smith MD et al. 2015) analyses of *Polistes* OR subfamilies
408 detected eight branches under episodic positive diversifying selection, all in OR subfamilies with
409 expansions: three branches in the 9-exon subfamily (0.33% of 918 9-exon subfamily branches;
410 Figure S4); three branches in the L subfamily (1.28% of 234 L subfamily branches; Figure S5);
411 one branch in the E subfamily (1.67% of 60 E subfamily branches; Figure S6); and one branch in
412 the H subfamily (1.54% of 65 H subfamily branches; Figure S7). This supports the hypothesis that
413 gene duplication releases duplicate genes from selective constraints, allowing duplicate sequences
414 to evolve towards other evolutionary optima (Ohno 1970). While the 9-exon OR subfamily is not
415 unique among expanded OR subfamilies in its instances of episodic positive selection as measured
416 by HyPhy aBSREL, the rate of amino acid divergence is higher among the 9-exon OR subfamily
417 as a whole. The 91% mean amino acid identity among 1:1 9-exon subfamily orthologs in *Polistes*
418 is significantly lower than the 95% mean amino acid identity among 1:1 orthologs in all other OR
419 subfamilies (Figure S8; Welch Two Sample t-test: $P = 7.051e-05$).

420 To further evaluate the patterns of nucleotide substitution driving accelerated amino acid
421 evolution of 9-exon ORs, we computed the values of d_N and d_S for pairwise alignments of 150
422 single copy orthologs between *P. fuscatus* and *P. dorsalis* (Figure 9) using model yn00 of PAML
423 (Yang 2007). Values of d_N are significantly higher in 9-exon (mean $d_N = 0.015$) compared to other
424 OR ortholog pairs (mean $d_N = 0.006$) (Welch Two Sample t-test, $P = 5.317e-07$). Values of d_S are
425 not significantly elevated among 9-exon ortholog pairs compared to other OR subfamilies (mean
426 $d_S = 0.029$ in 9-exon ORs and 0.025 in non-9exon ORs, $P = 0.343$). Omega values (d_N/d_S) greater
427 than 1 are often considered evidence of positive selection, while $d_N/d_S = 1$ corresponds to neutral
428 drift, and $d_N/d_S < 1$ is evidence of negative selection. The omega value (d_N/d_S) for all genes is less
429 than one, suggesting negative selection. However, omega is significantly higher in 9-exon ORs
430 than in non-9-exon ORs, indicating that negative selection is weaker on 9-exon ORs (Welch Two
431 Sample t-test: mean omega = 0.32 in non-9-exon ORs and 0.644 in 9-exon ORs, $P = 8.027e-05$).
432 In general, negative selection conserves ORs shared by *P. fuscatus* and *P. dorsalis* (mean omega
433 = 0.454), but an elevated rate of non-synonymous substitutions in 9-exon ortholog pairs imply

434 relaxed negative selection and more drift responsible for sequence evolution in the 9-exon relative
435 to other OR subfamilies.



436 **Fig. 9:** The values of d_S (x-axis) and d_N (y-axis) from pairwise alignments of *P. fuscatus* and *P. dorsalis*
437 orthologs are elevated in the 9-exon OR subfamily relative to other OR subfamilies. The diagonal line
438 represents a line of equality with slope of 1. Omega values of $d_N/d_S > 1$ may be evidence of positive
439 selection, while $d_N/d_S = 1$ may result from neutral drift, and $d_N/d_S < 1$ may result from negative selection.

440 Conclusions

441 By manually annotating the OR repertoires of five social wasp species spanning ~40 million years
442 of divergence in the *Polistes* genus, this study adds a higher resolution lens to our view of the
443 evolution of social insect odorant receptors. During the diversification of *Polistes*, evolutionary
444 patterns show genus-wide conservation of the ~200 OR repertoire except for the 9-exon genes,
445 which show elevated turnover and lower sequence conservation. The 9-exon OR subfamily has
446 dramatically expanded in paper wasps, and now makes up over half of the *Polistes* OR gene set.
447 Social and ecological niches are relatively conserved within *Polistes*, though there is considerable
448 variation in social behavior and ecological niches among vespid wasps (Ross & Matthews 1991;
449 O'Neill 2001). For example, an analysis of three new, high-quality hornet genomes suggested that
450 the highly eusocial hornets have larger OR repertoires compared to the primitively eusocial
451 *Polistes* (Harrop et al. 2020). That analysis recovered less than half of the ORs reported here for
452 *Polistes*, likely due to a lack of antennal transcriptome data, suggesting that hornets may have even
453 larger OR repertoires than has been reported. Evidence from the hornet *Vespa velutina*, including
454 the discovery of 264 antennal lobe glomeruli, indicates that the hornet OR repertoire has expanded
455 (Couto et al. 2016, 2017). Future analysis of additional high-quality genomes and antennal
456 transcriptomes of diverse social and solitary wasps within Vespidae will allow for further
457 examination of the relationship between social behavior and 9-exon as well as other OR subfamily
458 expansions.

459 Social insect species differ in their level of sociality and extent of olfactory recognition
460 abilities (d'Ettorre & Moore 2008; Rehan & Toth 2015). Some social aspects of the *Polistes* colony
461 cycle vary across species. For example, the average number of cooperative foundresses varies from
462 1 to ~6, and average sizes of mature nests may vary ~60 to ~490 cells (Reeve 1991; Sheehan et al.
463 2015; Miller SE et al. 2018). Increased 9-exon OR copy number may facilitate complex olfactory
464 recognition in species with larger colony sizes, higher cooperative nest-founding rates, and greater
465 sympatry with related species. However, expansions of 9-exon ORs are not exclusive to social
466 wasps, suggesting that the specific chemical ecology of an insect is a more influential factor
467 shaping OR evolution than level of sociality (Karpe et al. 2017). Furthermore, a meta-analysis
468 found that the complexity of CHC phenotypes does not differ between social and solitary
469 Hymenopteran species (Kather & Martin 2015). The CHC profile of *Nasonia vitripennis* includes
470 at least 52 CHC compounds, and detection of CHCs on prey items may help *Microplitis* identify

471 prey (Lewis et al. 1988; Niehuis et al. 2011). The need for solitary wasps to perceive CHCs could
472 explain why *N. vitripennis* and *M. demolitor* exhibit expansions in the 9-exon OR subfamily.

473 Electrophysiological deorphanization studies of 9-exon ORs in the ant *Harpegnathos*
474 *saltator* offer key insights into how 9-exon OR coding might relate to gene expansion. Through
475 combinatorial coding, 9-exon ORs can detect a large variety of structurally diverse CHCs. Pask et
476 al. (2017) examined 22 *H. saltator* 9-exon ORs, a subset of the 118 annotated 9-exon ORs in this
477 species, and found that 9-exon ORs were responsive to CHCs, and overlapped in their responses
478 to multiple CHC compounds. The combined responses of these 22 ORs to CHC extracts from
479 different castes were sufficient to map the CHC profiles of males, workers, and reproductive
480 females (gamergates) to separate regions of a 22-dimensional receptor space (Pask et al. 2017).
481 This highlights the ability of 9-exon ORs to facilitate social recognition by combinatorial coding.
482 In social insect colonies, CHC variation holds information at multiple levels of conspecific
483 recognition, from inter-colony nestmate recognition to within colony individual recognition
484 (d'Ettorre & Moore 2008). Expansion of the 9-exon OR subfamily might result from selection for
485 more combinations of ORs that together can discriminate between subtle qualitative and
486 quantitative variations in CHC blends of conspecifics. Nest-specific quantitative variation in CHCs
487 has been documented across *Polistes* species (Espelie et al. 1990; Singer et al. 1992; Espelie et al.
488 1994; Layton et al. 1994), but the molecular mechanisms underlying nestmate recognition in
489 *Polistes* are still obscure. Increased copy number of 9-exon ORs may not only expand the
490 qualitative range of compounds perceived by paper wasps, but also the quantitative olfactory
491 space, since wasps may be able to discern unique concentration differences between CHC blends
492 as a result of the combined action of 9-exon ORs with various response thresholds. Gene
493 duplication can also promote regulatory diversification (Kucharski et al. 2016; Dyson &
494 Goodisman 2020). Between castes and across stages of the colony cycle, CHCs vary in *P. metricus*
495 (Toth et al. 2014). Regulatory subfunctionalization of duplicate ORs could be responsible for
496 caste- and colony phase-specific expression of ORs involved in detecting caste-specific and
497 seasonally variable CHCs.

498 Most expanded OR subfamilies are highly conserved in copy number across five *Polistes*
499 species, with the exception of the 9-exon OR subfamily. In particular, one portion of the 9-exon
500 subfamily arranged in a single tandem array (*P. fuscatus* 9e s13) has experienced dynamic
501 evolution. What might drive rapid gain and loss of 9-exon ORs? Divergent social chemical

502 landscapes between species may cause gene turnover as 9-exon OR evolution tracks evolutionarily
503 labile chemical signals. *P. fuscatus* and *P. metricus* are closely related, and both species possess
504 CHC profiles consisting of linear and methyl-branched alkanes (Espelie et al. 1990; Espelie et al.
505 1994). However, the *P. fuscatus* CHC profile includes a higher proportion of alkenes than *P.*
506 *metricus* or *P. dominulus*, and the position of the methylated carbon of methyl-branched alkanes
507 is sometimes shifted between species (Espelie et al. 1990; Singer et al. 1992; Espelie et al. 1994;
508 Layton et al. 1994). Ant 9-exon ORs respond differently to subtle variations in CHC structure
509 (Pask et al. 2017). Between closely related *Polistes* species, structural isomers of methyl-branched
510 alkanes probably activate different ensembles of ORs.

511 If a chemical develops new behavioral relevance in a lineage, gene duplication would allow
512 the olfactory system to explore chemical space in the direction of this compound. HyPhy aBSREL
513 analyses identified eight branches in expanded OR subfamilies, including the 9-exon subfamily,
514 that have undergone positive selection during the last ~40 million years, consistent with
515 neofunctionalization or subfunctionalization of duplicated genes. Signatures of positive selection
516 on OR genes may reflect directional selection to perceive species-specific chemical signals.
517 Perception of species-specific CHCs might be important in mate compatibility recognition. In
518 *Polistes*, mating occurs at territories defended by males and often frequented by multiple species
519 (Post & Jeanne 1983; Reed & Landolt 1990). However, the frequency of interspecific mating is
520 low, suggesting *Polistes* use vision and/or olfaction to inform their mating decisions (Miller SE et
521 al. 2019). Duplication and deletion of ORs would facilitate evolution of species-specific chemical
522 signaling systems that could contribute to reproductive isolation of sympatric species. If a chemical
523 signal is lost in a species, the corresponding ORs may become obsolete, and would be expected to
524 pseudogenize and be purged from the genome. Duplication and deletion of ORs could also lead to
525 species-specific chemical signaling in the absence of evolutionary change in chemical signals
526 (Cande et al. 2013). However, OR evolution is not strictly necessary for such a difference to evolve
527 between species, and circuit-level changes can prescribe new valence to chemical signals that are
528 shared between species and perceived by common peripheral receptors (Seeholzer et al. 2018).
529 Neutral processes also contribute to birth-and-death events of ORs. There may be an advantage
530 for a large copy number up to a point, followed by random gene duplication and deletion around
531 this optimal copy number. This random genomic drift has been proposed to shape vertebrate

532 olfactory receptor evolution and copy number variation in all large multigene families (Nei 2007;
533 but see Hayden et al. 2010).

534 Aside from the 9-exon OR subfamily, gene expansions have occurred in subfamilies L, T,
535 H, E, and V (Figure 4). A larger variety of ORs relaying information through ORNs to a larger
536 number of antennal lobe glomeruli will increase sensory acuity in any olfactory discrimination
537 task, social or otherwise. An ancient locus of tandemly duplicated L subfamily ORs observed
538 across social insects has expanded in *Polistes*, although to a lesser extent than in other social insects
539 (~50 L subfamily ORs in honeybee and ants, 25 L subfamily ORs in a tandem array on *P. fuscatus*
540 scaffold 17). Odorant receptors in the L subfamily are thought to detect queen pheromone
541 components and fatty acids in bees as well as CHCs in ants (Wanner et al. 2007; Karpe et al. 2016;
542 Pask et al. 2017). The T subfamily has expanded to a greater degree in *P. fuscatus* (14 genes) than
543 in ants (~7 genes) and the honeybee (2 genes), but no ORs in this clade have been functionally
544 characterized. *P. fuscatus* has 9 H subfamily ORs, which are putative floral odorant detectors in
545 bees, and which also respond to CHCs and other general odorants in ants (Claudianos et al. 2014;
546 Slone et al. 2017). Fatty acids and volatile organic compounds are produced by flowers that wasps
547 rely on as a source of carbohydrates (Raguso 2008). Expansions in several OR subfamilies may
548 increase olfactory discrimination of chemicals with diverse behavioral relevance. *Polistes* species
549 are distributed globally in temperate and tropical regions, occupying similar social and ecological
550 niches as generalist predators and floral foragers that form primitively eusocial societies (Reeve
551 1991; Richter 2000). A conserved set of ORs may perform common functions in conserved
552 behaviors across paper wasp species. High levels of OR conservation are also consistent with a
553 specialist molecular function of an OR in a dedicated channel of olfaction. Patterns of molecular
554 evolution suggest conserved behavioral and molecular functions of most non-9-exon *Polistes* OR
555 subfamilies.

556 The differences between the conserved OR repertoires in *Drosophila* and the more
557 dynamic evolution of vertebrate OR gene families have given rise to speculation about the
558 relationship between OR function and evolution (Nozawa & Nei 2007; Andersson et al. 2015).
559 There is prevalent negative selection conserving odorant receptors across *Drosophila* species, and
560 the majority of *D. melanogaster* ORs form simple orthologous relationships across the genus
561 (Clark et al. 2007; Guo & Kim 2007; McBride & Arguello 2007; Nozawa & Nei 2007; Sánchez-
562 Gracia et al. 2009; Mansourian & Stensmyr 2015). In paper wasps we report both highly conserved

563 OR expansions similar to those seen in *Drosophila* as well as elevated gene turnover and drift
564 among the 9-exon ORs, reminiscent of a more vertebrate-like evolutionary pattern. If the highly
565 dynamic clades of 9-exon ORs of social wasps are involved in more combinatorial coding
566 compared to other more conserved 9-exon or non-9-exon ORs, that would indicate a link between
567 molecular evolution of receptors and neural coding. Further investigations into the relative tuning
568 of 9-exon as well as more conserved ORs in social wasps and other social insects provide a
569 promising research direction to investigate the links between molecular evolutionary patterns,
570 receptor tuning, and neural coding.

571 **Materials and Methods**

572 Antennal lobe imaging

573 Immunocytochemistry: In the fall of 2018, 20 adult *P. fuscatus* wasps (10 male and 10
574 gyne) were collected in Ithaca, NY. Wasps were immobilized by cooling at 4°C, heads were
575 removed, and brains were dissected from the head capsule and placed in ~1 ml of 4%
576 paraformaldehyde in 0.01M phosphate buffered saline (PFA) overnight at 4°C. Mouse monoclonal
577 anti-synapsin antibodies (SYNORF1, 3C11; Data Bank Hybridoma) were used as primary
578 antibodies to label synaptically rich antennal lobe glomeruli. Phalloidin conjugated with TRITC
579 (Tetramethylrhodamine Isothiocyanate; Invitrogen) was used to distinguish glomeruli borders and
580 label antennal nerve tracts. F(ab')₂ fragments of donkey anti-mouse antibodies conjugated to
581 Alexa 488 were used as secondary antibodies to visualize anti-synapsin (Jackson ImmunoResearch
582 Laboratories). After overnight fixation, brains were washed in 0.01M phosphate buffered saline
583 with Triton-X 100 (PBS-TX) 6 times for 20 min. Brains were incubated for three nights at room
584 temperature with anti-synapsin at 1:800 in PBS-TX solution. Brains were then washed again in
585 PBS-TX (6 times for 20 min) and incubated for three nights at RT with donkey anti-mouse
586 antibodies conjugated with Alexa 488 at 1:270, and then with Phalloidin-TRITC at 1:160 in PBS-
587 TX. After staining with secondary antibodies, brains were post-fixed with 4% PFA for 10 min and
588 dehydrated using increasing concentrations of methanol. Following full dehydration and two
589 washes in 100% methanol, brains were cleared in methyl salicylate overnight. They were then
590 mounted on slides in methyl salicylate.

591 Imaging: Images were collected using an inverted 880 confocal laser scanning microscope
592 (Zeiss LSM880 Indimo, AxioObserver) with a C-Apochromat10x/0.45 water-immersion objective
593 and a 1.8x digital zoom using the appropriate filter and laser setting for each fluorescent molecule
594 and 8x line averaging. Image stacks were collected using 10-15 µm optical sections for both males
595 and gynes. To individually label the glomeruli from confocal image stacks we used AVIZO and
596 Amira software (Thermo Scientific, FEI) to make 3D reconstructions of the antennal lobe from
597 whole mount preparations. Confocal stack files were imported into AVIZO software, and the voxel
598 dimension outputs of each image stack were imported to provide correct dimensions (Thermo
599 Scientific, FEI). Using the image segmentation function, we then identified glomeruli by hand
600 throughout the entire image stack.

601 Antennal RNAseq

602 *P. fuscatus* mRNA library preparation and sequencing: In the fall of 2017, 12 adult *P.*
603 *fuscatus* wasps (6 males and 6 gynes) from Ithaca, NY and Watkins Glen, NY were freeze-killed
604 on dry ice and stored at -80°C. Each pair of antennae was cut at the base of the scape and weighed
605 (paired gyne antennae = 1.27 mg ± 0.26 SD; paired male antennae = 1.57 mg ± 0.44 SD). RNA
606 was isolated from 2 pooled antennae of each individual using the PureLink RNA Micro Kit
607 (Ambion). Libraries were prepared using the NEBNext Ultra II RNA Library Prep Kit for Illumina
608 (NEB #E7770L) with the NEBNext Poly(A) mRNA Magnetic Isolation Module (NEB #E7490)
609 and NEBNext Multiplex Oligos for Illumina (NEB #E7600S). Libraries were analyzed on a
610 fragment analyzer for quality control and sequenced on two lanes of an Illumina HiSeq (Illumina,
611 San Diego, CA, USA).

612 Mapping mRNA reads of *P. fuscatus* and *P. dominula*: Raw reads from *P. fuscatus*
613 antennae were trimmed using Trimmomatic v.0.39 to (1) remove adapter sequences, (2) cut
614 leading and trailing bases below quality of 3, (3) cut 4-base wide windows when the average
615 quality per base dropped below 20, and (4) remove reads below 35 base pairs (bp) long (Bolger et
616 al. 2014). After trimming and filtering, more than 154 million paired end reads remained
617 (75,846,826 male; 78,453,620 gyne) with an average length of 132 bp. Processed paired end reads
618 were mapped, first to the *P. fuscatus* genome and subsequently to the genomes of 2 other *Polistes*
619 species in the fuscopolistes clade (*P. metricus* and *P. dorsalis*), using STAR v2.6 to generate BAM
620 files sorted by genome coordinate (Dobin et al. 2013). Each read was allowed to map to no more
621 than one locus. Mapped reads from each sex were combined and assembled using Trinity genome-
622 guided assembly (Haas et al. 2013). Summary statistics for the assembly are shown in Table S2.
623 Raw reads from the whole-body transcriptome of a single individual *P. dominula* were
624 downloaded from the Sequence Read Archive (Lopez-Osorio et al. 2017). To aid in manual
625 annotation of ORs, reads were aligned to the *P. dominula* genome with STAR v2.7, using the same
626 parameters as described above for *P. fuscatus*.

627 Gene annotation

628 OR annotation in *P. fuscatus*: The *Polistes fuscatus* genome was assembled as described
629 in Miller SE et al. 2020. Genome scaffolds containing putative ORs were identified based on
630 sequence similarity with published insect ORs from a range of species: *Atta cephalotes*,
631 *Acromyrmex echinaior*, *Apis mellifera*, *Camponotus floridanus*, *Cardiocondyla obscurior*,

632 *Ceratosolen solmsi*, *Drosophila melanogaster*, *Eulaema bombiformis*, *Euglossa dilemma*,
633 *Euglossa flammea*, *Euglossa imperialis*, *Eulaema meriana*, *Eufriesea mexicana*, *Lasioglossum*
634 *albipes*, *Microplitis demolitor*, *Monomorium pharaonis*, *Melipona quadrifasciata*, *Nasonia*
635 *vitripennis*, *Solenopsis invicta* (Robertson et al. 2003, Robertson et al. 2010, Zhou et al. 2012,
636 Zhou et al. 2015, Brand & Ramirez 2017, McKenzie & Kronauer 2018). The *P. fuscatus* genome
637 was queried with amino acid sequences from each of the listed species using the TBLASTN
638 algorithm with an e-value cutoff of 1e-5 (Altschul et al. 1997). Putative coding regions were
639 identified by examining TBLASTN match regions and chaining the match regions according to
640 criteria similar to those used by Zhou et al. (2012): neighboring match regions were included in
641 the same coding region if the match regions were in close proximity and if the upstream amino
642 acid query sequence was N-terminal to that of the downstream match region. Coding sequence
643 (CDS) was determined by manually curating exon-intron boundaries in Geneious v11.1.5 using
644 evidence from MAKER automated predictions, TBLASTN homology, Trinity predicted
645 transcripts (aligned to genome using BLAT) and mapped RNAseq reads. We consulted version 24
646 of the Havana annotation guidelines to inform manual annotation of exon-intron splice sites
647 (March 2016 guidelines by the Human and Vertebrate Analysis and Annotation group at WTSI).
648 “AG” was the only permitted acceptor site. “GT” and “GC” were both acceptable donor sites,
649 although “GC” was only present in a small number of genes. Amino acid sequences of ORs
650 generated by this pipeline were used as queries in an iterative round of TBLASTN with scoring
651 matrix PAM30 (Pearson 2013), to search for species-specific expansions, especially in the highly
652 divergent 9-exon receptor clade. The majority of apparently functional ORs that were not detected
653 by the automated annotation and required extensive manual curation were 9-exon subfamily
654 receptors (e.g. Figure S9).

655 OR annotation in 4 other *Polistes*: The *Polistes dorsalis* and *Polistes metricus* genomes
656 were assembled and annotated as described in Miller SE et al. 2020. The *Polistes canadensis* and
657 *Polistes dominula* genomes and annotations were accessed through NCBI (Patalano et al. 2015;
658 Standage et al. 2016). Putative ORs were identified by using TBLASTN with the same sample of
659 Hymenopteran protein query sequences used during manual annotation of *P. fuscatus* ORs, with
660 the addition of *P. fuscatus* OR protein sequences. Uncertain gene models were aligned using
661 Muscle version 3.8.425 with maximum 4 iterations and gene models were manually adjusted based
662 on homology with functional ORs in *P. fuscatus* (Edgar 2004). Gene models were called

663 pseudogenes if they exhibited frame-shift mutation, premature stop codons, or unacceptable 5'
664 donor or 3' receptor splice sites.

665 Quality assessment of putatively functional ORs: All *Polistes* OR protein sequences greater
666 than 300 amino acids in length were analyzed using InterProScan version 5.17-56.0 to check for
667 the InterPro olfactory receptor annotation IPR004117, the 7tm odorant receptor Pfam accession
668 PF02949, and the odorant receptor PANTHER accession PTHR21137 (Jones P et al. 2014). For
669 each of 192 orthologous groups predicted by OrthoFinder, the orthologs were aligned using
670 Muscle version 3.8.425 with maximum 4 iterations, and alignments were inspected for anomalies
671 that could be the result of annotation error. Adjustments were made to the gene models in 29
672 orthologous groups. The majority of these adjustments were changes in the length of already
673 annotated exons with few or no mapped mRNA reads. Transmembrane helices of all putatively
674 functional (>300 aa) ORs were predicted using TMHMM version 2.0c (Sonnhammer et al. 1998)
675 and Phobius version 1.01 (Käll et al. 2004).

676 Naming ORs: All ORs were named with a species-specific prefix using the first letter of
677 the genus followed by the first three letters of the species epithet (e.g. PfusOR#). Orco was not
678 numbered (e.g. PfusOrco), and no OR was labeled OR1, since some previous studies refer to Orco
679 as OR1. All tuning ORs were numbered in the order in which they appeared in the genome,
680 beginning with OR2. Therefore, OR names with the same number do not necessarily indicate
681 orthology among *Polistes*. Incomplete gene models were named with a suffix to indicate which
682 features of the gene model are missing, following Oxley et al. (2014): “NTE” missing N-terminus,
683 “INT” missing interior sequence, “CTE” missing C terminus, “NI” missing N terminus and interior
684 sequence, “NC” missing N and C termini. All pseudogenes were given the suffix “PSE.”

685 Search for ORs in unassembled genomic reads: Genome misassembly is a potential cause
686 of overestimation of pseudogenes, and is more common in repetitive regions of the genome. For
687 example, many ORs that were previously annotated as pseudogenes in the clonal raider ant were
688 not identified in a new, chromosome arm level assembly of the genome, which also yielded many
689 more functional OR gene models (McKenzie & Kronauer 2018). Given the completeness of the
690 *Polistes* genomes (Table S1) and the close correspondence between functional *P. fuscatus* ORs
691 and number of *P. fuscatus* antennal lobe glomeruli, this type of error likely accounts for minimal
692 annotation errors in this species. We searched for sequence similarity to ORs in the unassembled
693 genomic reads of *P. dorsalis*, the species in which we initially found the fewest functional ORs.

694 The 10x Genomics Supernova Assembler version 2.1.1 failed to assemble unmapped *P. dorsalis*
695 genomic reads greater than or equal to 150 nucleotides (~21.4 million paired-end reads). Therefore,
696 the reads themselves were formatted as a blast database and queried using TBLASTN at an e-value
697 cutoff of 1e-5. The same sample of Hymenopteran protein sequences used during manual
698 annotation, with the addition of the putatively functional OR protein sequences of all five *Polistes*
699 species, were used to search the unaligned *P. dorsalis* genomic reads. This resulted in significant
700 hits to 120 paired-end reads. Given that on average 159 ± 555 paired-end reads map to a given OR
701 gene in *P. fuscatus*, it is unlikely that many ORs were lost in unassembled genomic reads.
702 However, it cannot be ruled out that genome misassembly may account for overestimation of
703 pseudogenes in some cases.

704 Phylogenetic reconstruction of gene families

705 Hymenoptera-wide analysis: We used published OR sequences from *Apis mellifera*,
706 *Nasonia vitripennis* (Robertson & Wanner 2006; Robertson et al. 2010), and *Camponotus*
707 *floridanus* (Zhou et al. 2012), along with OR sequences of *P. fuscatus* (this study) to reconstruct
708 the phylogeny of the OR gene family. All amino acid (aa) sequences of chemoreceptors at least
709 300 aa in length were aligned using MAFFT version 7.453 with the high accuracy "E-INS-i"
710 method ("genafpair" option combined with 1000 cycles of iterative refinement ("maxiterate"))
711 (Kato and Standley 2013). The alignment was trimmed using trimAl v3 with the "automated1"
712 option to automatically choose the parameters that best fit the qualities of the alignment (Capella-
713 Gutierrez et al. 2009), generating a phylip file with 1066 sequences of 231 aa length. Gene trees
714 were generated with RAxML version 8.2.12 using the Maximum Likelihood optimality criterion
715 with 100 rapid bootstrap inferences, CAT model of rate heterogeneity across sites (Stamatakis
716 2006), JTT amino acid replacement matrix, F option to use empirical base frequencies from the
717 input alignment ("-m PROTCATJTT"), "-p 12345" to set initial random number for parsimony
718 search, *N. vitripennis* Orco as outgroup, "-f a" to combine bootstrap and maximum likelihood
719 searches in one run, "-x 12345" to set initial random number for bootstrap search, "-N 100" to
720 perform a bootstrap analysis informed by 100 unique maximum likelihood trees, and "-k" to add
721 branch lengths to trees (Stamatakis 2014, Jones DT et al. 1992). The tree was rerooted in FigTree
722 v1.4.4 (<https://github.com/rambaut/figtree/releases/tag/v1.4.4>) to visually clarify the position of
723 the Orco gene clade. The tree was visualized in R version 3.6.1 using ggtree (Figure 4A; Yu et al.
724 2017; R Core Team 2019).

725 Estimation of gene gain and loss events: Gene duplication and loss events were
726 reconstructed by reconciling a gene tree with a species tree in NOTUNG version 2.9.1.3 (Durand
727 et al. 2006; Vernot et al. 2008). A gene tree of putatively functional OR proteins at least 300 amino
728 acids in length was generated using published protein sequences for 4 ants (*A. echinator*, *A.*
729 *cephalotes*, *S. invicta*, *C. floridanus*; Zhou et al. 2012, 2015), 2 bees (*A. mellifera*, *L. albipes*;
730 Robertson et al. 2010, Zhou et al. 2015), 3 nonsocial wasps (*N. vitripennis*, *M. demolitor*, *C. solmsi*;
731 Robertson et al. 2010, Zhou et al. 2015), and 5 social wasps (*P. fuscatus*, *P. metricus*, *P. dorsalis*,
732 *P. canadensis*, *P. dominula*; annotated in this study). The gene tree was generated as described
733 above for the Hymenoptera OR tree, without rerooting. Following Zhou et al. 2012, all internal
734 branches with bootstrap support values below 70 were collapsed into polytomies in R using ggtree.
735 Notung was run in rearrangement mode using an edge weight (bootstrap value) threshold of 90,
736 with cost of “1.5” for a duplication and cost of “1” for a loss.

737 Analysis of *Polistes* ORs: The OR sequences of *P. fuscatus*, *P. metricus*, *P. dorsalis*, *P.*
738 *canadensis*, and *P. dominula* were filtered to include only ORs of at least 300 amino acids in
739 length. These were aligned using MAFFT ("E-INS-i" method) and trimmed in trimal using the
740 parameters described above for the Hymenoptera OR tree. The tree was generated using RAxML,
741 rerooted in FigTree, and visualized in R using ggtree as described above. Trees in the
742 supplementary materials were visualized using Interactive Tree Of Life (iTOL) version 5.6.2
743 (Letunic & Bork 2019). Orthology of *Polistes* OR amino acid sequences was inferred using
744 sequence similarity and gene collinearity in the genome. Orthologs were called based on similarity
745 of amino acid sequences with OrthoFinder version 2.2.7 using DIAMOND for sequence similarity
746 searches with an e-value cutoff of 0.001 (Emms & Kelly 2015). OrthoFinder predicted 135
747 orthologous groups that included gene copies from both *P. fuscatus* and *P. dorsalis*. For the
748 purpose of pairwise d_N/d_S analysis, this set of orthologs was expanded by inspecting the gene tree
749 (Figure 5; Figure S3) and Muscle alignments (see "Genomic organization" below) of syntenic
750 regions. Gene copies within a clade supported by a bootstrap value of at least 70 and showing
751 synteny were considered orthologs in the expanded ortholog set of 150 single copy orthologs
752 between *P. fuscatus* and *P. dorsalis*.

753 Genomic organization

754 OR genes and pseudogenes were considered to be in a tandem array if they were
755 uninterrupted by non-OR genes and were within 5kb of each other. The lengths of OR clusters

756 reflect the number of putatively functional ORs - gene models coding for proteins at least 300
757 amino acids in length - and exclude the pseudogenes contained within the array. The pairwise
758 percent amino acid identity between neighbors in an array was calculated using only putatively
759 functional ORs that neighbored another putatively functional OR within 5kb. Two functional ORs
760 separated by a pseudogenized OR were considered neighbors if their coding sequences were within
761 5kb of each other. Neighboring OR protein sequences were aligned using Muscle v3.8.425 with
762 maximum 4 iterations to determine percent amino acid identity. One-way ANOVA comparing
763 percent amino acid identity of neighbors across eight tandem arrays was performed using the "aov"
764 function, and Tukey HSD test with the "TukeyHSD" function, both of which are in the base R
765 package stats version 3.6.1 (R Core Team 2019).

766 Sequence analyses

767 Tests for positive selection: All putatively functional *Polistes* ORs greater than 350 amino
768 acids in length were used in analyses of episodic diversifying selection within OR subfamilies
769 using aBSREL in HyPhy version 2.5.15 (Smith MD et al. 2015). Subfamily-specific alignments
770 were generated using MAFFT version 7.453 ("E-INS-i") and trimmed in trimal with the
771 "gappyout" option. Trees were generated using RAxML as above, except the
772 "PROTGAMMAAUTO" model was used to automatically choose the best protein model given
773 likelihood. CDS alignments were generated using trimal with the "backtranslate" option added to
774 the above parameters. The significance threshold for each aBSREL analysis was Holm-Bonferroni
775 corrected $p=0.05$. Values of pairwise d_N/d_S for orthologs shared by *P. fuscatus* and *P. dorsalis*
776 were calculated using PAML version 4.9 (Yang 2007) program yn00. The values of d_N and d_S
777 reported were calculated using the Yang & Nielsen (2000) method. Values of d_N , d_S , and omega
778 were compared between 9-exon and non-9-exon orthologous pairs using the function "t.test" in the
779 base R package stats version 3.6.1 (R Core Team 2019).

780 Test for gene conversion: To search for gene conversion events, all *Polistes* OR subfamily
781 codon alignments were analyzed using GENECONV version 1.81a with options "/lp" to list all
782 pairwise significant fragments, "/w123" to set initial random number, and "-nolog" to avoid a
783 segmentation fault in UNIX (Sawyer 1989). Bonferroni-corrected KA p-values below 0.05 were
784 considered significant.

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794 **Author Contributions**

795 AWL and MJS conceptualized the study. SEM and MJS sequenced and assembled the genomes.
796 CMJ dissected and imaged antennal lobes. AWL sequenced mRNA and manually annotated
797 ORs. AWL and MFF labeled antennal lobe glomeruli. AWL conducted all downstream analyses
798 and wrote the paper. CMJ wrote the methods section on antennal lobe imaging. AWL, SEM,
799 CMJ, and MJS edited the final manuscript.

800 **Supplementary Materials**

801 GFF files of OR models for 5 *Polistes* species
802 Fasta files of OR amino acid sequences and CDS for 5 *Polistes* species
803 Supplementary excel file with meta-data associated with ORs of 5 *Polistes* species
804 Table S1
805 Table S2
806 Table S3
807 Figure S1
808 Figure S2
809 Figure S3
810 Figure S4
811 Figure S5
812 Figure S6
813 Figure S7
814 Figure S8
815 Figure S9

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