

1 **Rapamycin as a potent and selective inhibitor of vascular endothelial growth factor receptor in**
2 **breast carcinoma**

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18 **Abstract**

19 Angiogenesis is the process of new vascular formation, which is derived from various factors.
20 For suppressing cancer cell growth, targeting angiogenesis is one of the therapeutic approaches.
21 Vascular endothelial growth factor family receptors, including Flt-1, Flk-1, and Flt-4, have been
22 found to play an essential role in regulating angiogenesis. In the present study, we evaluated the
23 effects of rapamycin and platelet factor-4 toward breast carcinoma at the proteomic and genomic
24 levels. A total of 60 N-Methyl-N-Nitrosourea-induced rat breast carcinomas were treated with
25 rapamycin, platelet factor-4, and rapamycin+platelet factor-4. The tumors were subsequently
26 subjected to immunohistological protein analysis and polymerase chain reaction gene analysis.
27 Protein analysis was performed using a semi-quantitative scoring method, while the mRNA
28 expression levels were analyzed based on the relative expression ratio. There was a significant
29 difference in the protein and mRNA expression levels for the selected markers. In the
30 rapamycin+platelet factor-4 treated group, the Flt-4 marker was downregulated, whereas there were
31 no differences in the expression levels of other markers, such as Flt-1 and Flk-1. On the other hand,
32 platelet factor-4 did not exhibit a superior angiogenic inhibiting ability in this study. Rapamycin is a
33 potent anti-angiogenic drug; however, platelet factor-4 proved to be a less effective drug of anti-
34 angiogenesis on rat breast carcinoma model.

35 **Keywords:** Breast cancer, N-methyl-N-nitrosourea-induced rat, vascular endothelial growth factor
36 receptor, rapamycin, platelet factor-4

37

39 **Introduction**

40 Angiogenesis, a process by which new vasculature forms, occurs as a result of a variety of
41 factors. Neo-angiogenesis is one of the main processes that allow tumors to grow larger than 2 mm in
42 diameter [1]. Since the discovery of neo-angiogenesis, numerous intensive research studies have
43 been conducted in order to determine the root factors that influence and enhance this process.
44 Targeting angiogenic factors, such as vascular endothelial growth factor (VEGF), platelet-derived
45 growth factor (PDGF), insulin-like growth factor (IGF), fibroblast growth factor (FGF), and their
46 receptors, have potential impact on the development of new strategies to suppress tumor growth,
47 especially at later stages [2].

48 Fms-like tyrosine 1 (Flt-1), Fms-like kinase 1 (Flk-1), and Fms-like tyrosine 4 (Flt-4) are
49 members of the VEGF receptor family and are known as prognostic markers [3]. In addition, each of
50 these VEGF receptors has been found to play a role in the regulation of neo-vascularization [3].
51 Moreover, these receptors co-regulate each other's' expression to trigger angiogenesis. Among these,
52 Flk-1, known as a kinase-derived receptor (KDR), plays a significant role in the regulation of
53 angiogenesis, whereas other molecules in the same family support this process [4]. Flt-1 has also
54 been reported to play a significant role in the branching of new arteries, whereas Flk-1 influences
55 vein formation, and Flt-4 plays a vital role in the regulation of lymphatic vessel formation.

56 Flt-1 is known to be the cognate receptor for VEGF-A. This receptor plays a crucial role in
57 the regulation of neo-angiogenesis alongside Flk-1. Recent studies have revealed that Flt-1 induces
58 angiogenesis along with Flk-1 through several intrinsic pathways, including the PLC- γ , Grb2, and

59 PI3K/Akt, which in turn activate downstream pathways, such as MAPK for proliferation, eNOS for
60 cell permeability, and Caspase 9 for survival [5].

61 Contrarily, Flt-4 plays a significant role in the regulation of lymphangiogenesis, which is a
62 common route of cancer metastases [6]. Most cases of aggressive carcinoma exhibit significant Flt-4
63 expression, which reflects the degree to which several factors, especially Flt-4, play a role in the
64 regulation of neo-lymphangiogenesis [7]. Based on our previous findings in rats with breast
65 carcinoma, the treated groups exhibited a significant downregulation of the Flt-4 receptor; moreover,
66 the cancer cells were significantly less aggressive, and most of the tumor cells appeared localized. In
67 the present study, we examined and analyzed the expression of VEGFRs in rat breast carcinoma
68 following treatment with rapamycin and PF-4.

69

70 **Materials and methods**

71 **Animal procedures**

72 Sixty female Sprague Dawley (SD) rats were obtained from the Animal Research and
73 Services Centre, and ethical clearance was obtained from the Animal Ethics Committee
74 [PPSG/07(A)/044/ (2010) (56)]. The rats were housed at the animal house unit. The caging and
75 handling of the rats conformed to good laboratory practices, as outlined by the Animal Research and
76 Services Centre (ARASC). To avoid discomfort, pain, and stress, the rats were given an analgesic
77 drug (Ketamine) at a dose of 80 mg/kg intraperitoneally during the measurement of the tumor. The
78 rats were maintained in groups of six and fed a standard laboratory diet [8]. They were divided into

79 four groups, and each group was given different interventions/drug(s), as follows: the control
80 (untreated) group 1 [n = 15], rapamycin-treated group 2 [n = 15], platelet factor-4-treated group 3
81 (PF-4) [n = 15], and rapamycin and PF-4-treated group 4 (rapamycin+PF-4) [n = 15]. The rats were
82 sacrificed 5 days after the drug intervention process.

83 **NMU, rapamycin, and PF-4 preparation and tumor induction**

84 NMU (N-nitroso-N-methylurea) was dissolved in freshly prepared 0.9% normal saline prior
85 to the induction process. The NMU solution was injected intraperitoneally into 21-day-old rats at a
86 dose of 70 mg/kg body weight [9], after which each animal was monitored and palpated weekly for
87 mammary tumor lesions. Daily inspection and palpation were also performed to monitor the onset of
88 tumors in the mammary region, and the weight of the rats were measured daily. Rapamycin was
89 mixed with absolute ethanol and diluted in mixtures of 10% polyethylene glycol (PEG)-400, 8%
90 ethanol, and 10% Tween-80 to a final concentration of 20 µg/0.2 mL, whereas PF-4 was dissolved in
91 normal saline with a final concentration of 20 µg/0.2 ml

92 **Study design**

93 This study was conducted *in vivo* and involved an intraperitoneal injection of the NMU
94 carcinogen into 60 female Sprague Dawley rats using a 21G needle to induce breast carcinoma. After
95 the development of breast tumors within 40 days of NMU carcinogen injection, the tumors were then
96 suppressed using an angiogenic inhibitor as either a single treatment (PF-4 or rapamycin) or a
97 combination of these two compounds; for combination treatment, both PF-4 and rapamycin were
98 injected into the tumors. After 5 days of intervention, the rats were terminated, whereas rats in the
99 control group were terminated 45 days after tumor induction. Hematoxylin and eosin staining

100 together with immunohistochemical stains were used to analyze the histology and expression of
101 angiogenic protein markers. All results were then statistically analyzed using the SPSS software
102 version 21.

103 **Immunohistochemistry**

104 Tissue sections on slides were deparaffinized in xylene and then rehydrated in graded alcohol
105 solutions. Antigen retrieval was conducted in a pressure cooker, and the selection of an appropriate
106 buffer depended on the detected marker. Nonspecific background staining was blocked by a 3%
107 hydrogen peroxide solution. Sections were incubated with the primary antibody for 1 h at room
108 temperature, which was followed with several washes and incubation with the secondary antibody
109 using an UltraVision ONE Large Volume Detection System HRP Polymer Kit. The protein reactivity
110 was visualized using a 3,3'-diaminobenzidine (DAB) Plus substrate system (Cat. No. TL-125-PHJ,
111 LabVision, USA). The tissue was counterstained with hematoxylin, which stained the nuclei.

112 **Immunohistochemistry scoring system**

113 Each section was scored in a blinded manner by two observers based on a previously reported
114 semiquantitative scoring method [9]. Each sample was scored based on the percentage of tumor cells
115 stained (0- absence of staining; 1- <30% of tumor cells stained; 2- 30–60% of tumor cells stained;
116 and 3- >60% of tumor cells stained).

117 **RNA expression analysis**

118 The tissue sample extraction was performed using TRIZOL (Invitrogen, USA) to obtain total
119 RNA. All RNA preparation and handling steps were carried out under RNase-free conditions and

120 were performed in a laminar flow hood. Total RNA from each fraction was dissolved in 20 μ L of
121 RNA storage buffer (Ambion, USA) and stored at -80°C until use. RNA concentration was
122 determined by 1% gel electrophoresis and absorbance readings at 260 nm using the Nanodrop-1000
123 spectrophotometer (NanoDrop, Technologies, USA). The reading of RNA samples was within the
124 range of 1.8–2.2, indicating the purity of the sample. This ratio was obtained from the 260/280
125 absorbance ratio. mRNA was isolated from total RNA using the Dynabeads mRNA Purification kit
126 (Invitrogen, USA) according to the manufacturer’s instructions. cDNA synthesis was performed
127 using the SuperScript™ First-Strand Synthesis System (Invitrogen, USA) in a total volume of 20 μ L
128 according to the manufacturer’s instructions.

129 **Quantitative RT- PCR analysis**

130 The expression of 11 genes of interest and three selected reference genes was examined by
131 real-time TaqMan® PCR assay. Expression levels were determined using the exon spanning
132 hydrolysis probes (FAM or MGB dye-labelled), which are commercially available as “Assay on
133 Demand” (Applied Biosystems, Foster City, CA, USA), with optimized primer and probe
134 concentrations as stated in Table 1.

135 **Table 1 List of gene markers by TaqMan® PCR assay used for angiogenesis analysis.**

Gene symbol	Assay ID	Gene name
<i>Flt-4</i>	Rn00570815_m1	fms-related tyrosine kinase 1
<i>Flt-1</i>	Rn00677893_m1	fms-related tyrosine kinase 4

<i>VEGFR-2</i>	Rn00564986_m1	kinase insert domain receptor
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137 Quantification was accomplished with the PikoReal Real-Time qPCR System (Applied
138 Biosystems, USA) using TaqMan® Universal PCR Master Mix and universal thermocycling
139 parameters as recommended by Applied Biosystems. RT-PCR samples were run in duplicates using
140 1 µL cDNA, and the reactions were performed in 96-well plates (Applied Biosystems) in a reaction
141 volume of 20 µL. The gene expression normalization factor for each sample was calculated
142 according to the geometric mean of all three selected reference genes [10]. The Minimum
143 Information for Publication of Quantitative RealTime PCR Experiments (MIQE) guidelines were
144 considered for the performance and interpretations of the qPCR reactions [11]. Gene expression was
145 analyzed according to the relative expression ratio (R-value) obtained from the cycle threshold (CT)
146 value and amplification efficiency (E) value [$E = -1 + 10(-1/\text{slope})$]. The expression of targeted genes
147 for each sample provides evidence that these drugs act at the gene level. The entire gene sequence
148 began to amplify at approximately cycle 25 (Fig. 4).

149 **Statistical analysis**

150 A non-parametric one-way ANOVA test was used to determine the differences in the
151 immunohistochemical expression of Flt-1, Flk-1, and Flt-4 among the experimental groups. Any
152 results where $p < 0.05$ were considered statistically significant. All statistical analyses were
153 conducted using the SPSS software version 21.

155 **Results**

156 One-way ANOVA revealed a significant difference ($p < 0.05$) for all angiogenic markers (Flt-
157 1, Flk-1, and Flt-4) in all experimental groups.

158 **Flt-1/VEGFR1**

159 Rapamycin ($M = 90.1664$, $SD = 7.4487$) exhibited better antiangiogenic effects than PF-4 (M
160 $= 93.7946$, $SD = 7.1303$) and rapamycin+PF-4 ($M = 93.6990$, $SD = 1.8432$) (Fig 1) with respect to
161 Flt-1 marker. This demonstrated that PF-4 and rapamycin+PF-4 exerted fewer effects in the
162 suppression of this angiogenesis marker in tumor cells. Contrarily, rapamycin was effective in
163 blocking angiogenic markers through the mTOR signaling pathway.

164

165 **Fig 1. The expression of VEGFRs signaling protein receptor on rat's mammary carcinoma.** The
166 VEGFRs expressions are highly reflected to the efficacy of treatment given to suppress angiogenesis
167 via VEGFRs signaling blockage. All treatment groups showed reduce VEGFRs expressions with p
168 value is <0.05 .

169

170 **Flk-1/VEGFR2**

171 Rapamycin ($M = 89.9043$, $SD = 7.2542$) again revealed better antiangiogenic effects than PF-
172 4 ($M = 98.9175$, $SD = 2.0487$) and rapamycin+PF-4 ($M = 91.2330$, $SD = 4.0934$) in the suppression
173 of the angiogenic marker Flk-1. This marker has the potential to trigger angiogenesis along with

174 other growth factors. These findings were slightly different from that of the Flt-1 marker, as the
175 rapamycin+PF-4 group exhibited better suppression of this marker. The result still demonstrated a
176 lack of synergistic activity between rapamycin and PF-4. Contrarily, PF-4 did not result in good
177 suppression of this receptor, as mentioned in the literature [12].

178 **Flt-4/VEGFR3**

179 Rapamycin+PF-4 ($M = 87.8700$, $SD = 4.1620$) was very efficacious in suppressing the Flt-4
180 marker compared with either rapamycin or PF-4 alone. Interestingly, the protein was downregulated
181 more in the combination group than in the other groups, suggesting synergistic activity between these
182 drugs. In contrast, the rapamycin group ($M = 90.0236$, $SD = 7.2414$) exhibited similar effects but to a
183 lesser degree than those of the rapamycin+PF-4 group with respect to Flt-4. Both groups showed
184 significant Flt-4 downregulation. However, PF-4 alone ($M = 96.5385$, $SD = 1.8304$) failed in that the
185 protein was slightly downregulated, but the levels were not significantly different from those after no
186 treatment (control).

187 **Protein expression and scoring**

188 VEGFRs were highly expressed in the cytoplasm of tumor cells (breast carcinoma) (Figs 2
189 and 3). The tissues that lacked expression and those without cytoplasmic staining indicated
190 suppressive activity on VEGFRs by the drugs.

191
192 **Fig 2. Photomicrograph of NMU-induced rat's breast tumour tissue.** The stain on tumour cells
193 showed the expressions of Flt-1 marker. Flt-1 immunostaining (a) x100 and (b) x400. Tumour cell
194 (TC), blood vessel (BV)

195 **Fig 3. Photomicrograph of NMU-induced rat's breast tumour tissue.** The stain on tumour cells
196 showed the expressions of Flk-1 marker. Flk-1 immunostaining (a) x100 and (b) x400. Tumour cell
197 (TC), blood vessel (BV)

198
199 A semiquantitative scoring system was used to determine the protein expression and included
200 counts of positive cells and protein expression intensity. The counting was performed on three
201 selected regions of interest based on the region with the highest protein expressions. The control
202 group had a score of 9 points for both the Flt-1 and Flk-1 markers and a score of 7.5 points for Flt-4,
203 indicating that tumor growth was mostly dependent on the formation of new blood vessels to supply
204 the nutrients needed for that growth. Moreover, the high expression of Flt-4 indicated
205 lymphangiogenesis, which is used by tumor cells as a passageway for metastases.

206 **Effects of angiogenesis inhibitors**

207 Rapamycin was found to be a good VEGF inhibitor. Based on our findings (Table 2), the
208 angiogenic protein expressions were either completely absent or were expressed at low levels,
209 indicating that rapamycin can substantially downregulate these proteins. This was supported by the
210 semiquantitative scores, where the protein expression decreased from 9 to 6 points. Interestingly,
211 rapamycin+PF-4 exhibited the best inhibitory effects on the Flt-4 ligand, which had a
212 semiquantitative score of 4.5 out of 9 points. We found that the synergism between the drugs was
213 highest for the Flt-4 marker, which plays a significant role in the regulation of lymphatic vessel
214 formation. However, the effects of rapamycin+PF-4 on another two markers, the Flt-1 and Flk-1
215 ligands, seem less effective, and we found no synergism between the drugs on the suppression of
216 these ligands.

217 **Table 2 Semiquantitative score of VEGFR expression in NMU-induced mammary carcinoma**
218 **in rats. The rapamycin and rapamycin+PF-4 groups exhibited lower scores, whereas the PF-4**
219 **group exhibited less-effective suppressive activity on VEGFR markers.**

Treatment	FLT-1 (%)	Scoring	FLK-1 (%)	Scoring	FLT-4 (%)	Scoring
Control	98.72	9	98.92	9	97.58	7.5
Rapamycin	90.95	7.5	90.45	7.5	90.38	6
PF-4	93.62	9	96.62	7.5	96.54	7.5
Rapamycin+PF-4	93.7	7.5	91.23	7.5	87.87	4.5

220

221 RNA expression

222 The mRNA expression levels of *Flt-1*, *Flk-1*, and *Flt-4* genes from the rapamycin, PF-4, and
223 rapamycin+PF-4 treated groups were significantly downregulated compared with the untreated group
224 (Fig 5), with *p*-values of 0.036, 0.018, and 0.000, respectively. Based on these findings, the
225 expression ratio of genes in all intervention groups were downregulated by a mean factor of 0.591
226 (SE range was 0.441–0.789), 0.458 (SE range was 0.311–0.669), and 0.134 (SE range was 0.078–
227 0.267) with *p*-values < 0.01. Moreover, we found that the drugs used for intervention had suppressed
228 angiogenesis and lymphangiogenesis at the mRNA level with varying intensities. As demonstrated at

229 the protein level, the administration of rapamycin or rapamycin+PF-4 was found to result in
230 significantly lower than the control (no treatment), whereas intervention with PF-4 led to a lack of
231 suppressive activity on tumor growth indicated by high genes expression, specifically with respect to
232 VEGFRs.

233 **Fig 5.** The expression ratio of mRNA angiogenic markers; Flt-1, Flk-1 and Flt-4 on NMU-induced
234 mammary carcinoma under the influence of rapamycin, PF4 and rapamycin+PF4. All VEGFRs
235 marker showed down regulation of gene expression.

236

237 **Discussion**

238 In this study, we elucidate the effects of rapamycin and PF-4 as a single dose and in
239 combination on breast carcinoma *in vivo*. We investigated four intervention groups and a control
240 group (untreated). Rapamycin has been used in several medical applications, including as an
241 antifungal agent, an anti-rejection drug after organ transplants, as a treatment for
242 lymphangioleiomyomatosis (LAM), and as an antiproliferative agent (anti-cancer) [13]. Contrarily,
243 PF-4 is a molecule responsible for inducing blood coagulation and activating platelets during platelet
244 aggregation. PF-4 plays a significant role in wound repair and inflammatory reactions [14]. In this
245 study, we observed the relation of synergistic strength between interventional drugs, which are
246 rapamycin and PF-4 against the proliferative and angiogenic activities in breast carcinoma.

247 Angiogenesis has long been known as fundamental for such diverse physiological processes
248 as embryonic and postnatal development, reproductive functions, and wound repair. The blood
249 vessels provide oxygen and nutrients and carry key regulatory signals to growing tissues [15].

250 Studies on tumorigenesis have shown that the inflammatory response is sometimes accompanied by
251 increased vascular proliferation. This finding led to the proposal that new blood vessels play a crucial
252 pathogenic role in the regulation of inflammatory processes and to the fulfillment of the demands of
253 proliferating cells. Tumors need nutrients to grow, and their only way to obtain sufficient nutrients is
254 through blood vessels [16]. The mechanisms of the development of new vasculature from the
255 existing vessels require the activation of some upstream signaling receptors, which will then trigger
256 some angiogenic pathways, including the mTOR and PI3K/Akt pathways. Based on these findings,
257 we can hypothesize that aggressive tumor cells express more angiogenic proteins than non-
258 aggressive tumors. This indicates that tumors are highly dependent on angiogenesis, which provides
259 them with additional vessels to fulfill the demands for nutrients and oxygen needed by growing
260 tumor cells. In addition, the vessels also function as a metastatic passageway for tumor cells to
261 spread primarily *via* lymphatic vessels.

262 Based on the literature, rapamycin acts on mTOR through several upstream pathways. mTOR
263 can be classified into two distinct types, mTORC1 and mTORC2, which are characterized by the
264 presence of raptor and rictor, respectively. Signaling through mTORC1 involves Grb2 and Ras–Raf–
265 MEK–Erk, whereas signaling through mTORC2 involves two different pathways that involve IRS-1
266 and Grb10 [17]. The VEGF and mTOR pathways are connected by IRS-1 and PI3K/Akt. Inhibition
267 of mTOR pathways will affect the production of VEGF, thus suppressing angiogenesis. In contrast to
268 rapamycin, PF-4 affects the CXCR3 ligand. Activated CXCR3 protein will activate the PI3K/Akt
269 pathway downstream [18]. With the combination of both drugs (rapamycin and PF-4), we
270 hypothesized that they would function synergistically against the PI3K/Akt pathway, which would
271 result in a better prognosis.

272 This study has noted that after the treatment of rapamycin or rapamycin+PF-4, the expression
273 of VEGFRs was significantly suppressed, whereas PF-4 treatment alone resulted in less suppression
274 of targeted markers. Based on these findings, we report that PF-4 and rapamycin do not function
275 synergistically on VEGFRs, but some evidence suggests that they function antagonistically against
276 the ligands. VEGF-A (Flt-1 and Flk-1) was discovered to be a survival factor for endothelial cells,
277 and it exerts its effects through the PI3K/Akt pathway. High VEGF-A expression in tumor cells
278 suggests that VEGF prevents endothelial cell apoptosis induced by serum starvation.

279 The expression of angiogenic markers, primarily Flt-1, Flk-1, and Flt-4, was associated with
280 the size and aggressiveness of tumors. Tumor cells in larger, more aggressive tumors require more
281 nutrients and oxygen for their growth [19]. Based on previous studies, Flt-1 protein was found to be
282 highly associated with the production of Flk-1/KDR proteins, which play a significant role in the
283 regulation of angiogenesis and vasculogenesis. Interestingly, with their roles as signaling receptors,
284 Flt-1, Flk-1, and Flt-4 were reported to be responsible for such disorganization and lethality and were
285 found to be negative regulators of VEGF, at least during the early development. In our rat model, Flt-
286 1 was highly expressed in aggressive tumor cells. This indicates that Flt-1 mediates chemotactic
287 signals and potentially extends the role of the receptor, suggesting that this protein promotes
288 mechanisms other than angiogenesis, such as increasing the survival rate of tumor cells, mitogenesis,
289 and permeability-enhancing effects of VEGF rather than being a non-functional or decoy receptor. In
290 the current study, we found that both receptors (Flt-1 and Flk-1) were potentially elevated in
291 epithelial–mesenchymal transition in breast carcinoma and resulted in a rapid proliferation of tumor
292 cells in this tissue.

293 Angiogenesis highly depends on VEGF protein production, which relies on Flk-1/KDR to
294 activate its pathways [20]. Several previous studies reported that Flk-1 signals act through the
295 PI3K/Akt and PLC- γ /Raf-MEK-ERK signaling cascades, which play a significant role in survival,
296 permeability, migration, and proliferation [21]. Bulk production of these proteins is important to
297 sustain a rapidly growing tumor, which requires additional volumes of oxygen and nutrients. In this
298 study, we found that the expression of Flk-1/KDR was prominent, indicating that angiogenesis was
299 very aggressive. The reduced expression of VEGF markers indicates the suppression of angiogenesis.

300 Flt-4 is the receptor of VEGF-C and VEGF-D and acts as a pivotal regulator of
301 lymphangiogenesis. In this study, we found that the expression of Flt-4 was closely associated with
302 tumor aggressiveness. Previous findings in human breast carcinoma with overexpression of VEGF-C
303 revealed the induction of lymphangiogenesis in and around the tumor. This finding confirms prior
304 evidence that Flt-4 is a vital receptor that regulates the development of new lymphatic vessels, which
305 are an important route for tumor metastases [22]. In this study, we found that Flt-4 was highly
306 expressed in aggressive breast carcinoma but that it was expressed to a lesser degree in the
307 rapamycin- and rapamycin+PF-4-treated groups. These results suggested that both rapamycin and
308 rapamycin+ PF-4 were useful in treating metastatic carcinoma, but that the use of PF-4 alone did not
309 result in good outcomes. Furthermore, we found that the use of antiangiogenic drugs suppressed the
310 development of lymphatic vessels more so than blood vessels. This demonstrates that these drugs
311 have a potent ability to block the interaction between VEGF-C- and Flt-4-soluble VEGFR3 fusion
312 proteins.

313 **Conclusion**

314 In conclusion, rapamycin is a potent antiangiogenic drug acting on breast carcinoma. The
315 downregulation of VEGFR expression at both protein and mRNA levels in the rapamycin-treated
316 group indicates that rapamycin is best used against an angiogenic stimulant. Conversely, PF-4 has
317 been demonstrated to be a less-effective antiangiogenic drug in this breast carcinoma model.

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403 **Authors' contribution**

404 MSMS, HJ, WFWAR, and VG designed experiments, MSMS and TAD performed major
405 experiments and data analysis. MSMS, HJ, and WFWAR performed the statistical analysis. MSMS,
406 HJ, and WFWAR designed the research theme. All authors read and approved the final manuscript.

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409 **Supporting information**

410 Nil

411

412

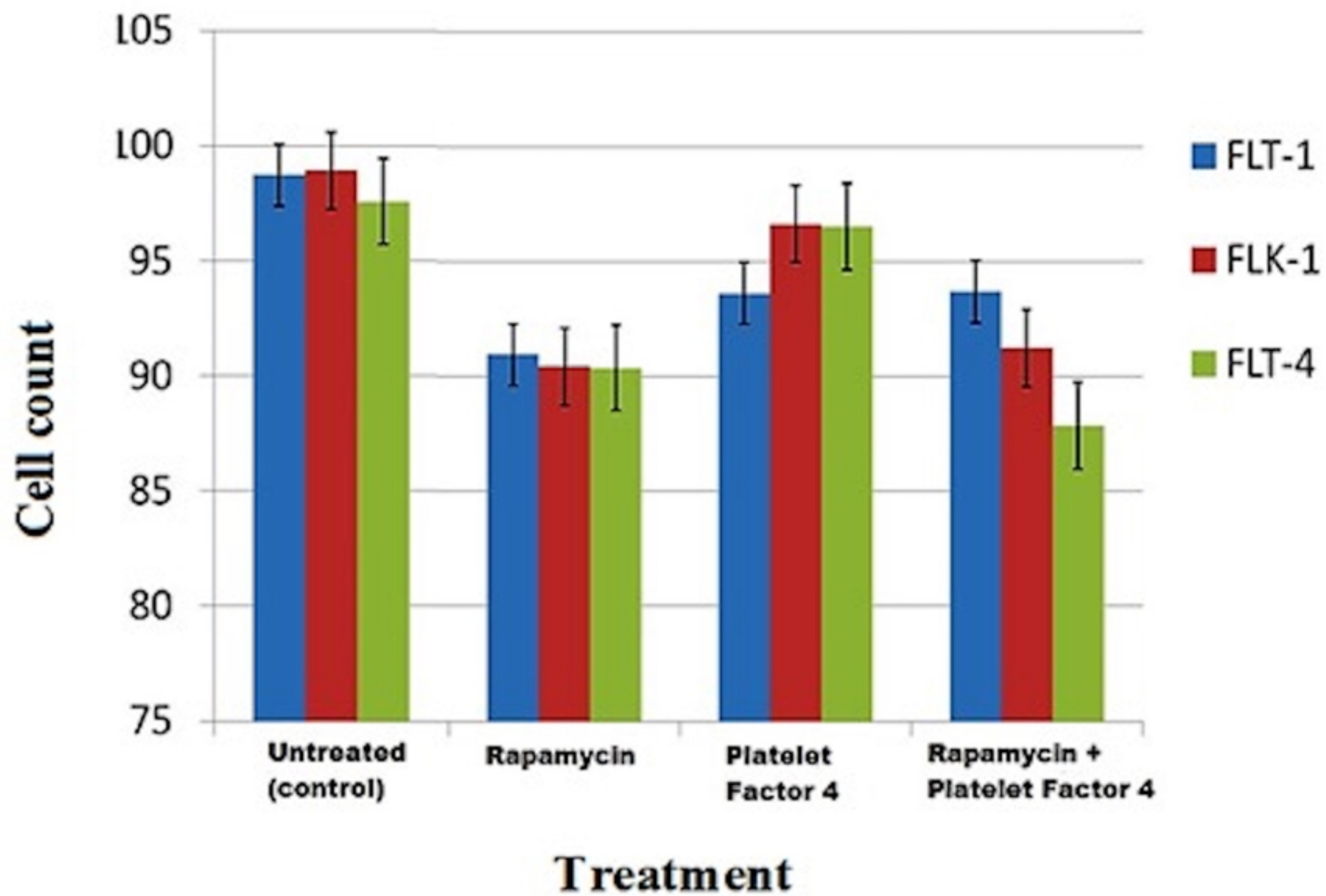
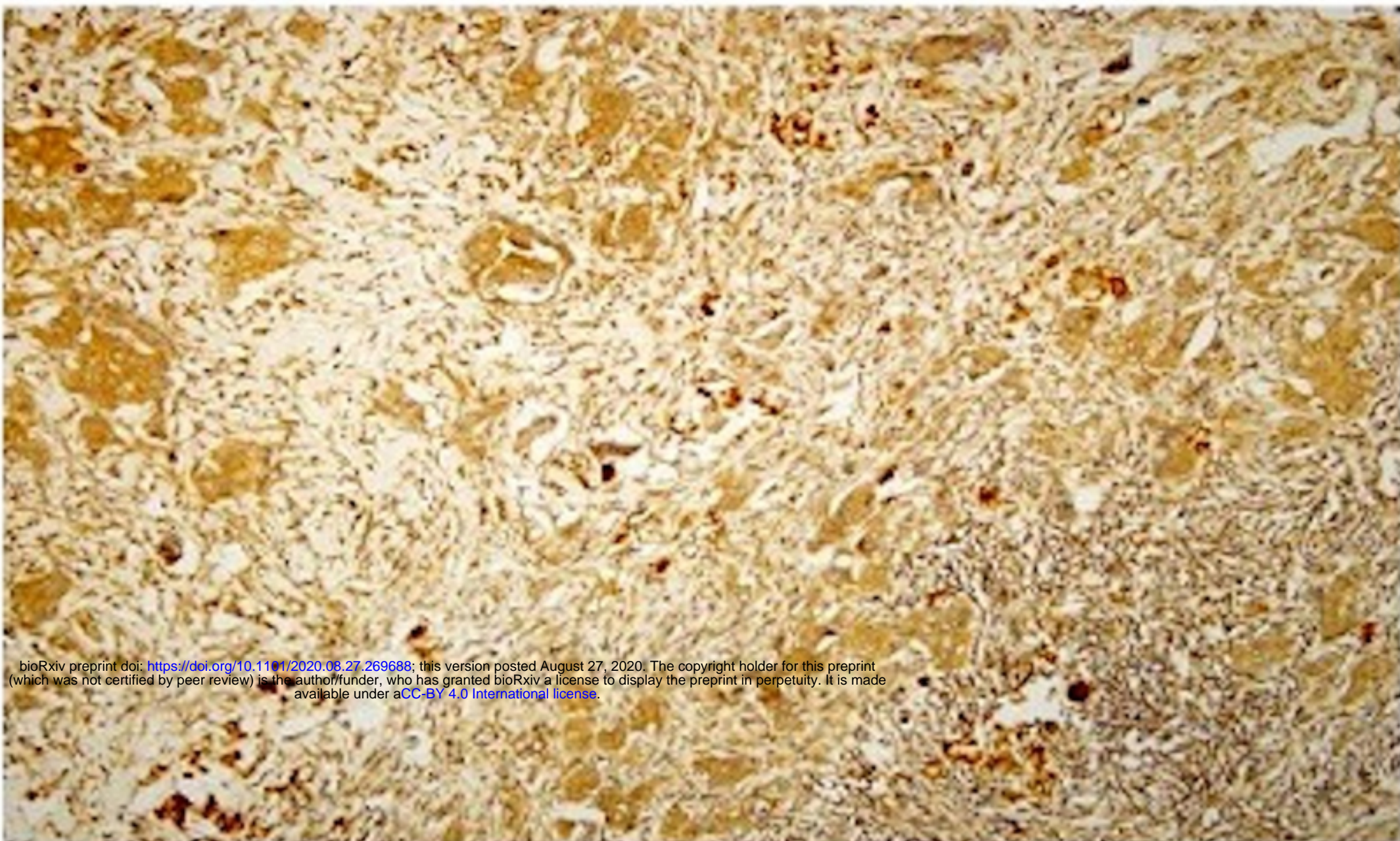
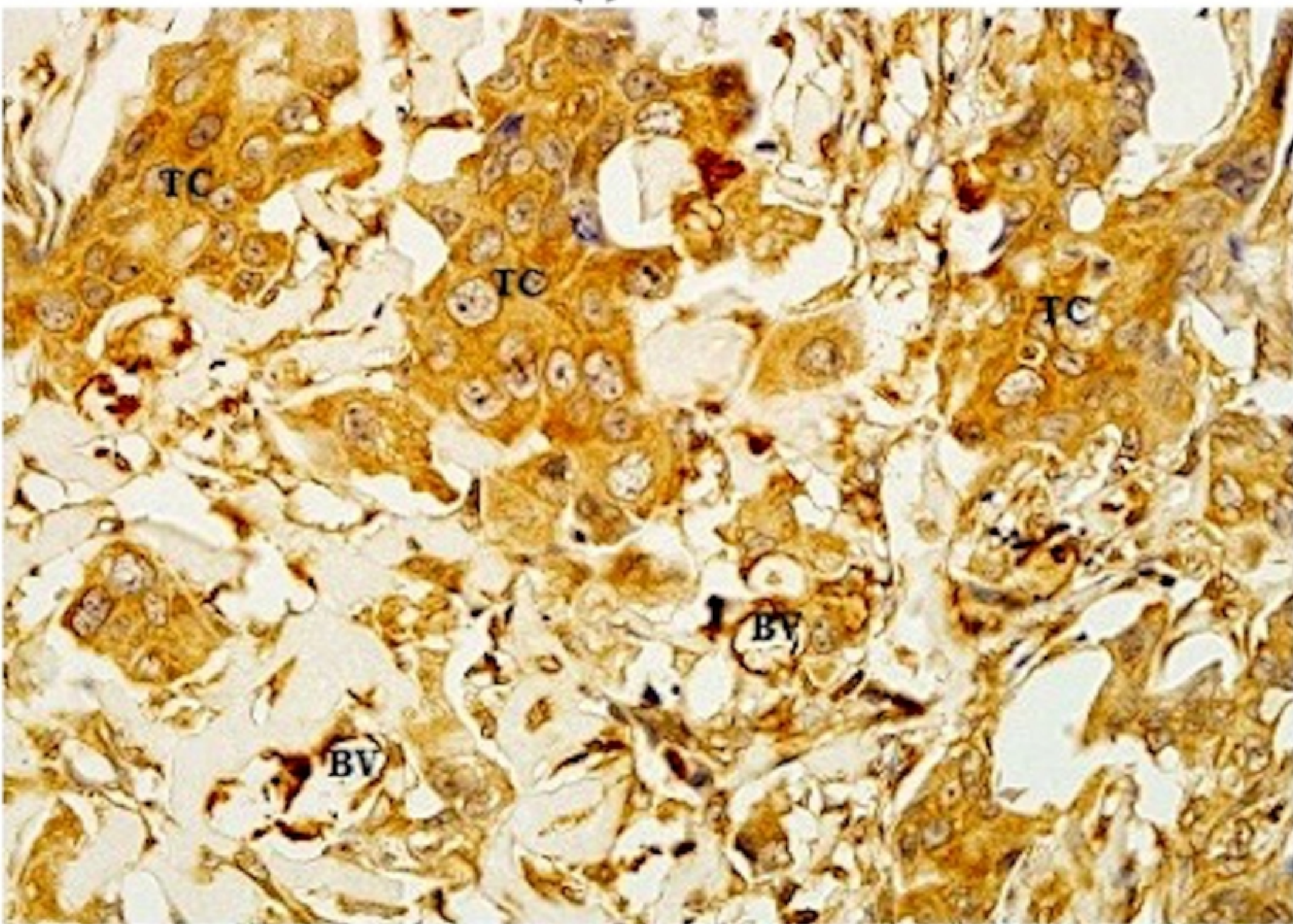


Fig 1

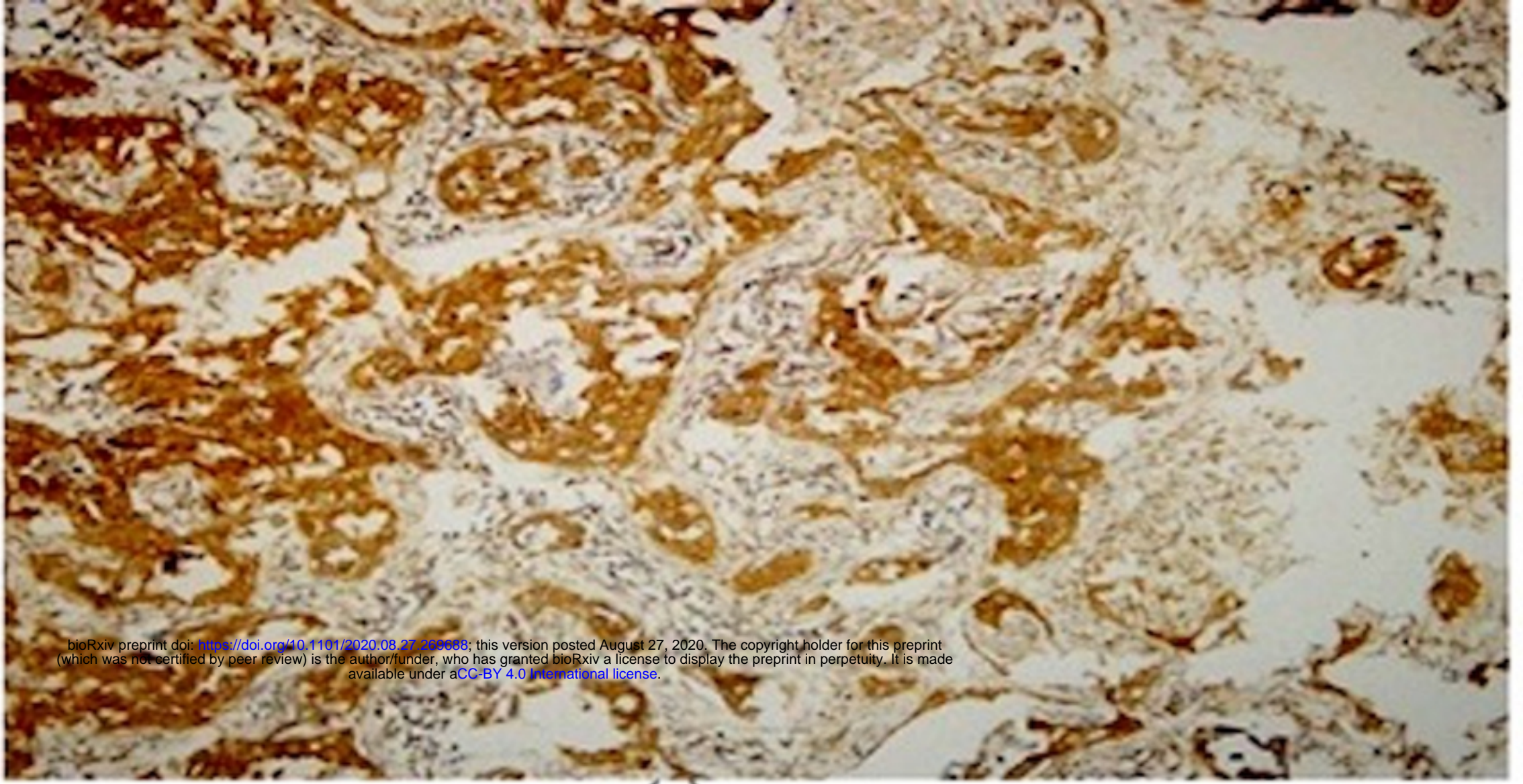


(a)

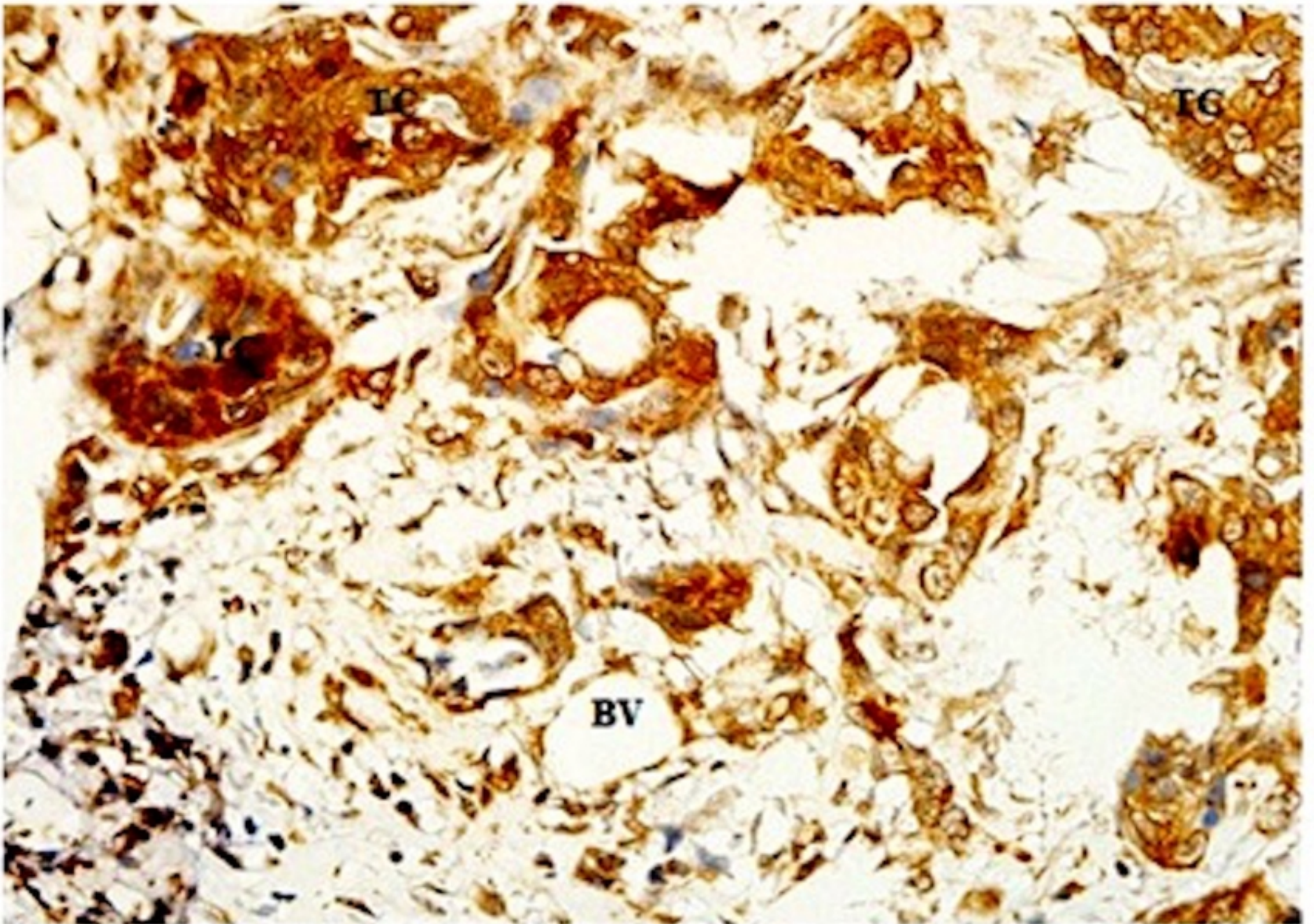


(b)

Fig 2



(a)



(b)

Fig 3

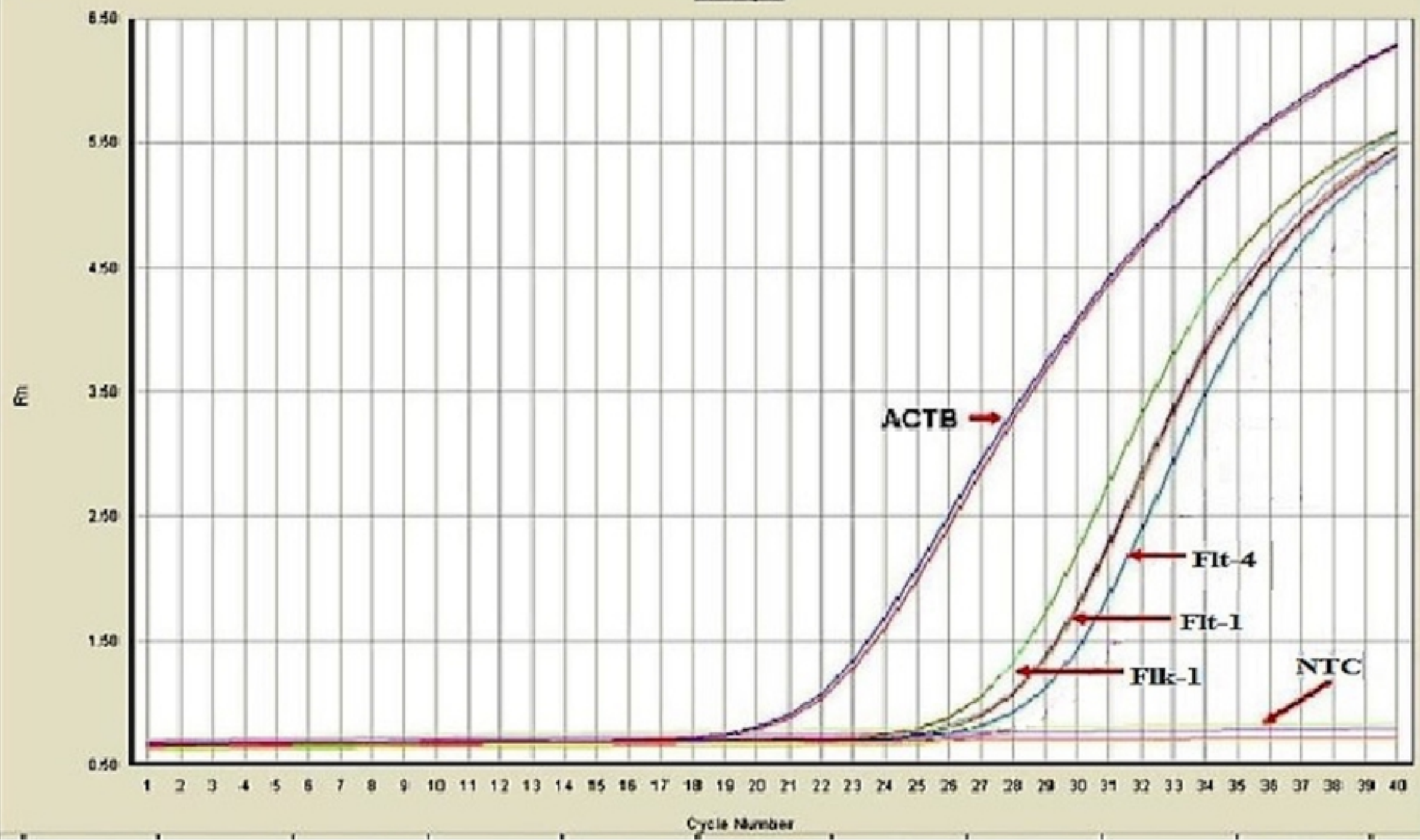


Fig 4

Relative Expression

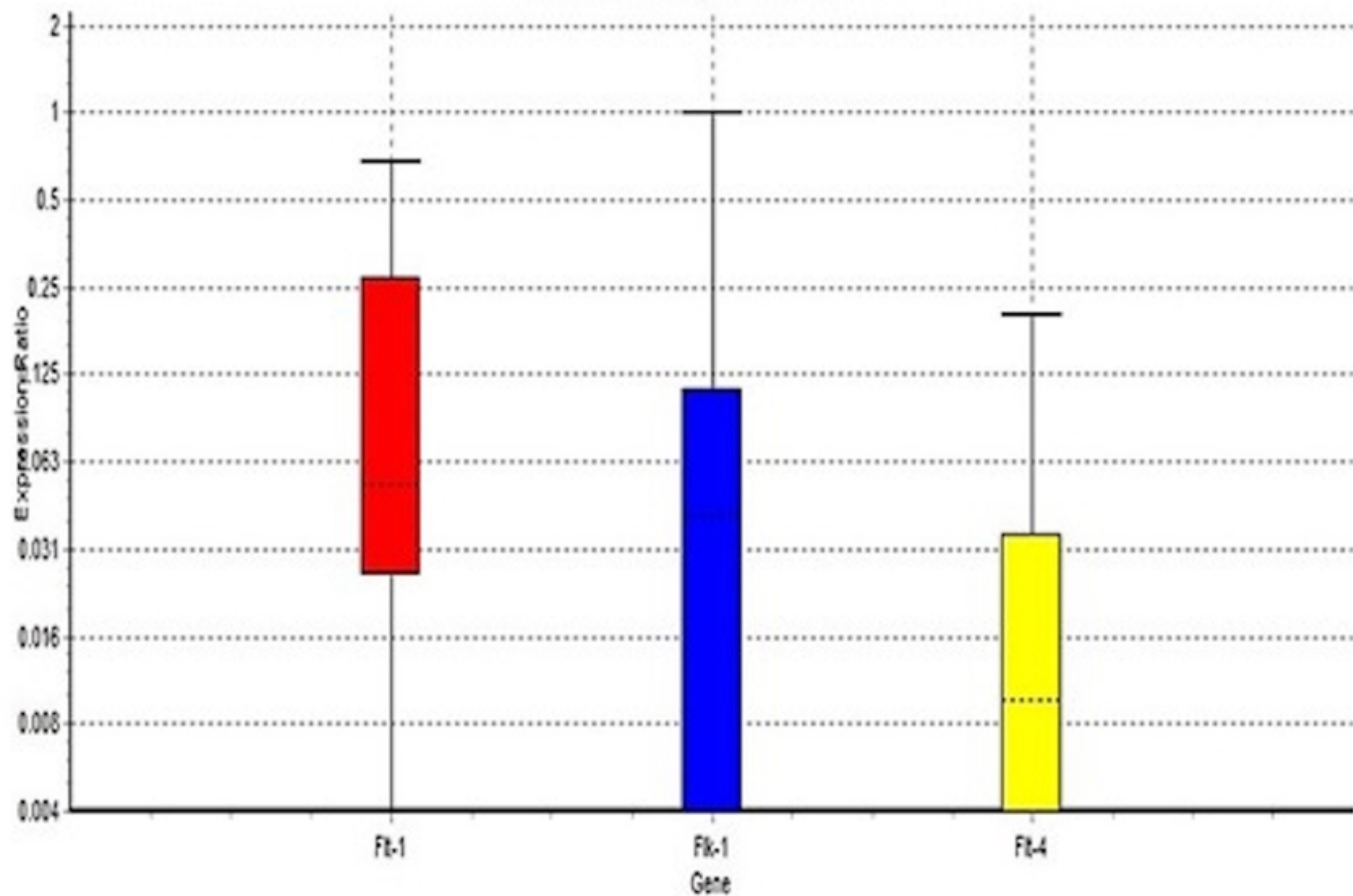


Fig 5