

1 **TITLE PAGE**

2 ***In vitro* trypanocidal effect of sera from *Erythrocebus patas* (Red Patas monkey) and**
3 ***Chlorocebus tantalus* (Tantalus monkey) on *Trypanosoma brucei brucei* Plimmer &**
4 **Bradford, 1899 and *Trypanosoma congolense* Broden, 1894**

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16

17 **ABSTRACT**

18 *Anti-Trypanosoma brucei brucei* and anti-*Trypanosoma congolense* activities of sera from two
19 species of uninfected zoo-primates, *Erythrocebus patas* (red patas monkey) and *Chlorocebus*
20 *tantalus* (tantalus monkey) were investigated. The sera were screened using thick films and
21 haematocrit centrifugation technique (HCT), to ensure that the donor primates were not infected
22 with trypanosomes. *Trypanosoma brucei brucei* (Federe strain) and *Trypanosoma congolense*
23 were suspended in supplemented RPMI (Rossvelt Park Memorial Institute) 1640 medium and the
24 motility of the parasite was used as index of viability after the addition of each test serum. The
25 selected primate sera exhibited some degree of anti-*Trypanosoma brucei brucei* activities *in vitro*.
26 Red patas monkey serum had an inhibition index of 0.27, while that of Tantalus monkey was 0.34,
27 against *Trypanosoma brucei brucei*, with mean survival times of 22.00±1.73 hours for red patas
28 monkey serum and 19.67±0.58 hours for tantalus monkey serum, which are significantly lower
29 (P<0.05) than that of the control (30.00±0.00 hours). The selected primate sera had pronounced
30 inhibitory activities against *Trypanosoma congolense*. Sera from the two species of primate had
31 very high anti-*Trypanosoma congolense* activity showing an inhibition index of 0.91 for Red patas
32 monkey serum and 0.90 for Tantalus monkey serum, with marked and significant reduction
33 (P<0.05) in survival time of 7.00±1.73 hours in Red patas monkey serum and 7.67±0.58 hours in
34 Tantalus monkey serum, compared with the control (74.00±1.00 hours). The *in vitro* anti-
35 trypanosomal activity of the serum samples was shown to be cidal in nature. The activity was not
36 associated with xanthine oxidase. This study revealed that sera from red patas monkey and tantalus
37 monkey had a moderate anti-*Trypanosoma brucei brucei* activity and a very high anti-
38 *Trypanosoma congolense* activity *in vitro* suggesting the presence of some non-specific materials.

39 **Keywords:** Zoo-primates, anti-*Trypanosoma brucei brucei* activity, anti-*Trypanosoma*
40 *congolense* activity, survival time, inhibition index, xanthine oxidase, non-specific materials.

41 **Authors' Summary**

42 The mechanisms that allow trypanosomiasis-resistant animals to control blood trypanosomes are
43 being investigated, to identify non-specific factors that kill trypanosomes or limit their
44 proliferation, contributing to host resistance. For instance, xanthine oxidase has been isolated and

45 identified as the protein that kills trypanosomes in Cape buffalo. Humans and several other
46 primates are also known to be resistant to infection by several animal-specific trypanosome
47 species. In this study, sera from some zoo primates, red patas monkey and tantalus monkey, tested
48 on *Trypanosoma brucei brucei* and *Trypanosoma congolense in vitro*, showed a slight anti-
49 *Trypanosoma brucei brucei* activity and a very high anti-*Trypanosoma congolense* activity. These
50 activities were shown to be cidal in nature and not associated with the protein xanthine oxidase.
51 The authors suggest that non-specific factors other than the enzyme xanthine oxidase might have
52 accounted for the sera anti-trypanosomal activities.

53 **Introduction**

54 African animal trypanosomiasis constitutes a major impediment to efficient and profitable
55 livestock production in Africa generally and particularly in Nigeria. It causes about 3 million
56 deaths annually in cattle and production losses of about US\$ 1.2 billion [1]. Although several
57 measures have been used to attempt the control of the disease, the main control measure remains
58 the use of trypanocidal drugs since the antigenic variation in the parasite has render the production
59 of vaccine (using antibody) a very difficult task. Thus trypanosomiasis is still a great challenge to
60 scientists. New control approaches are based on nonspecific factors. Some wild animals are known
61 to be naturally trypanosomiasis-resistant. Natural resistance is attributable to natural specific
62 antibodies and nonspecific serum factors capable of killing trypanosomes or limiting their rate of
63 population growth, thereby contributing to the trypanotolerance/trypanoresistance of some
64 animals. For instance, 3 distinct constitutive defense mechanisms against *T. brucei* have been
65 detected by Black *et al.* [2], in sera from sub-Saharan mammals other than primates that show a
66 high level of resistance to African trypanosomes, and they reported that these mechanisms act
67 together with parasite-specific antibody responses to control the severity of infection arising in the

68 reservoir hosts [2]. In the same regard, some studies have also been carried out on some zoo
69 primates [3], [4], [5]. The present study was targeting at investigating the presence of nonspecific
70 factors against *T. brucei brucei* and *T. congolense*, in the sera of two species of Zoo Primates,
71 *Erythrocebus patas* (Red patas monkey) and *Chlorocebus tantalus* (Tantalus monkey).

72 **Materials and methods**

73 *Ethical statement*

74 The ethical approval for this research was obtained from the Committee on Animal Use and Care,
75 Directorate of Academic Planning & Monitoring, Ahmadu Bello University, Zaria, with the
76 Approval No: ABUCAUC/2017/007. Local approval was given by the Director of Kano
77 Zoological Garden. Blood samples were collected in accordance with best practice guidelines to
78 minimise stress on donor primates. The laboratory animals used were kept in a clean aerated room
79 and were given necessary care.

80 *Donnor primates and sampling sites*

81 Blood was collected from red patas monkey and tantalus monkey reared in an area free of
82 trypanosomes, the Kano Zoological Garden, Kano State, Nigeria. Kano is 481 metres (1,578 feet)
83 above sea level. The city lies to the north of the Jos Plateau, in the Sudanian Savanna region that
84 stretches across the south of the Sahel. The city lies near where the Kano and Challawa rivers
85 flowing from the southwest converge to form the Hadejia River, which eventually flows into Lake
86 Chad to the east [6].

87 *Sample collection and trypanosome screening test*

88 This was done with the assistance of some experts. Each blood sample was aseptically collected
89 from the donor animal through the femoral vein and was divided into two. One part was kept as
90 whole using EDTA (ethyl-diamine-tetra-acetic acid) while the other one was dispensed into sterile
91 plain glass test tube, allowed to clot for 2-3 hours at room temperature and serum was collected
92 after centrifugation at 1,000 g [2] and stored in a freezer at -20°C after screening, for further
93 analysis. The screening was done on each of the blood sample with EDTA, using two of the
94 standard detection techniques (thick films) and haematocrit centrifugation technique (HCT) [7].

95

96 *In vitro detection of anti-trypanosomal activity of the test sera*

97 This was done using microtitre plates as follows:

- 98 - Trypanosomes from the buffy coat were suspended in RPMI 1640 (Rossvelt Park
99 Memorial Institute 1640) medium supplemented with 2% glucose, 2mM sodium pyruvate,
100 10% heat-inactivated (56 C, 30 min) fetal bovine serum [8], sodium bicarbonate and
101 sodium pyruvate and antibiotics (streptomycin 100µg/ml, penicillin 100U/ml).
- 102 - 50µl of each monkey serum sample was introduced into one of the 96 wells of a microtitre
103 plate and 50µl of *T. brucei brucei* or *T. congolense* suspension 8 per field (31.62×10^6 / ml
104 of blood) was added to it, rocked gently to mix and incubated at room temperature.
- 105 - A drop of about 5µl of each mixture was examined microscopically, hourly using wet film
106 method. Cessation in motility of parasites was taken as indication of serum activity against
107 the parasites [9].

108 - The motility of *T. brucei brucei* or *T. congolense* in each well was compared with the
109 motility of the same parasites in the control well without test serum.

110 A formula was derived [10] to determine the Anti-trypanosomal Activity Index (ATI) of each
111 serum sample.

$$112 \quad \text{ATI} = \frac{T^c - T_s}{T^c}$$

113 Where: T_c is the survival time of the parasites in the control medium

114 T_s is the survival time of the parasites in the sample

115 *Infectivity assessment*

116 100µl of the mixture of the *in vitro* affected parasite with effective serum was inoculated into mice
117 and monitored by microscopy for trypanosomes on daily basis, 10 and 30 days for *T. brucei brucei*
118 and *T. congolense* respectively, to determine if the observed anti-trypanosomal activity was
119 inhibitory or cidal.

120 *Detection of the xanthine oxidase content of the selected sera*

121 The technique of Black *et al.* [2] was adopted to detect the xanthine oxidase content of the test
122 sera. Accordingly, aliquots (100 µl) of each serum was added to 900 µl of H₂O₂-assay buffer (0.5
123 mM xanthine and 1 mM 2, 4, 6 tribromo-3-hydroxybenzoic acid in 0.1 mM 4-amino-antipyrine
124 with a final concentration of 8 units horse-radish peroxidase per ml). The mixture was incubated
125 at 25°C for 30 min, immediately chilled in an ice bath, and absorbance read at 512 nm wave length
126 was recorded after zeroing the spectrophotometer with an equivalent mixture lacking horseradish
127 peroxidase. A serial dilution of the commercial cow's milk xanthine oxidase was done; absorbance

128 was also recorded at 512 nm wave length and used to plot a standard curve. It has been established
129 that detection of H₂O₂ produced in serum by this assay is not affected by other enzymes in serum,
130 including catalase [11]. The xanthine oxidase content of serum was determined by reading the
131 serum value against the cow's milk xanthine oxidase standard curve.

132 *Data analysis*

133 One-way Analysis of variance (ANOVA) was used for sera's inter-species comparison of the
134 parasites survival time. Student's t-test was used to compare the mean survival time of the two
135 species of parasites, *T. brucei brucei* and *T. congolense* in the test sera. The significant difference
136 was at the level of probability 0.5. All data were expressed as means \pm Standard Error.

137 **Results**

138 *Trypanosome Infection Status of Sera from Selected Primates*

139 Blood samples from selected monkeys were negative for trypanosomes by thick blood film and
140 haematocrit centrifugation technique (HCT).

141

142 *The effect of sera from Red patas monkey and Tantalus monkey on T. brucei brucei*

143 There was no significant difference in the trypanosomal activity of the sera of both primates
144 (22.00 \pm 1.73 and 19.67 \pm 0.58 hours respectively); meanwhile, the mean survival times of the
145 parasites in these sera were significantly lower (P<0.05) than that of the control, the RPMI 1640
146 (30.00 \pm 0.00 hours), showing a slight anti-*Trypanosoma brucei brucei* activity with ATI values of
147 0.27 and 0.34 respectively (Table 1).

148 **Table 1:** Effect of sera of selected Zoo Primates on *T. brucei brucei* *in vitro*

Source of serum	Survival time (hrs)	ATI
<i>Erythrocebus patas</i> (Red Pattas monkey)	22.00±1.73 ^a	0.27
<i>Chlorocebus tantalus</i> (Tantalus monkey)	19.67±0.58 ^a	0.34
RPMI 1640	30.00±0.00 ^b	0

149 Values are mean ± standard deviation. Values with different superscripts down the column are
150 significantly different (P<0.05). ATI: Anti-trypanosomal activity Index.

151

152 *The effect of sera from Red patas monkey and Tantalus monkey on T. congolense*

153 Sera from the selected zoo primates exhibited very high anti-*Trypanosoma congolense* activity *in*
154 *vitro* with very high index, close to one (ATI of 0.91 for Red pattas monkey and 0.90 for Tantalus
155 monkey). This was shown by a very reduced survival time of *T. congolense* 7.00±1.73 hours in
156 Red pattas monkey serum and 7.67±0.58 hours in Tantalus monkey (Table 2). These two values
157 are not statistically different (P>0.05) from each other, but both are significantly (P<0.05) lower
158 than the control, the RPMI 1640 (74.00±1.00 hours).

159 **Table 2:** Effect of sera of selected Zoo Primates on *T. congolense*

Source of serum	Survival time (hrs)	ATI
<i>Erythrocebus patas</i> (Red Pattas monkey)	7.00±1.73 ^a	0.91
<i>Chlorocebus tantalus</i> (Tantalus monkey)	7.67±0.58 ^a	0.90

RPMI 1640	74.67±1.00 ^b	0
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160
 161 Values are mean ± standard deviation. Values with different superscripts down the column are
 162 significantly different (P<0.05). ATI: Anti-trypanosomal activity Index.

163
 164 *The compared effect of sera from Red patas monkey and Tantalus monkey on T. bucei brucei and*
 165 *T. congolense in vitro*

166 Each of these sera had a very high anti-*Trypanosoma congolense* activity Index (ATI), 0.91 for
 167 Red pattas monkey and 0.90 for Tantalus monkey. This was very high compared to their activity
 168 index against *T. brucei brucei* (0.27 for Red patas monkey and 0.34 for Tantalus monkey). The
 169 survival times of *T. congolense* in these sera are significantly lower (P<0.05) than that of *T. brucei*
 170 *brucei* in the same sera (Table 3).

171 **Table 3:** Compared effect of sera from selected Zoo primates on *T. brucei brucei* and *T.*
 172 *congolense in vitro*

Source of serum	Survival time of <i>T. brucei</i>	ATI on <i>T. brucei</i>	Survival time of <i>T. congolense</i>	ATI of the serum on <i>T. congolense</i>
Red Pattas monkey (<i>Erythrocebus patas</i>)	22.00±1.73 ^b	0.27	7.00±1.73 ^a	0.91
<i>Chlorocebus tantalus</i> (Tantalus monkey)	19.67±0.58 ^b	0.34	7.67±0.58 ^a	0.90
RPMI 1640	30.00±0.00 ^a	0	74.00±1.00 ^b	0

173 Values are mean \pm standard deviation. Values with different superscripts across the rows are significantly
174 different ($P < 0.05$). ATI: Anti-trypanosomal activity Index.

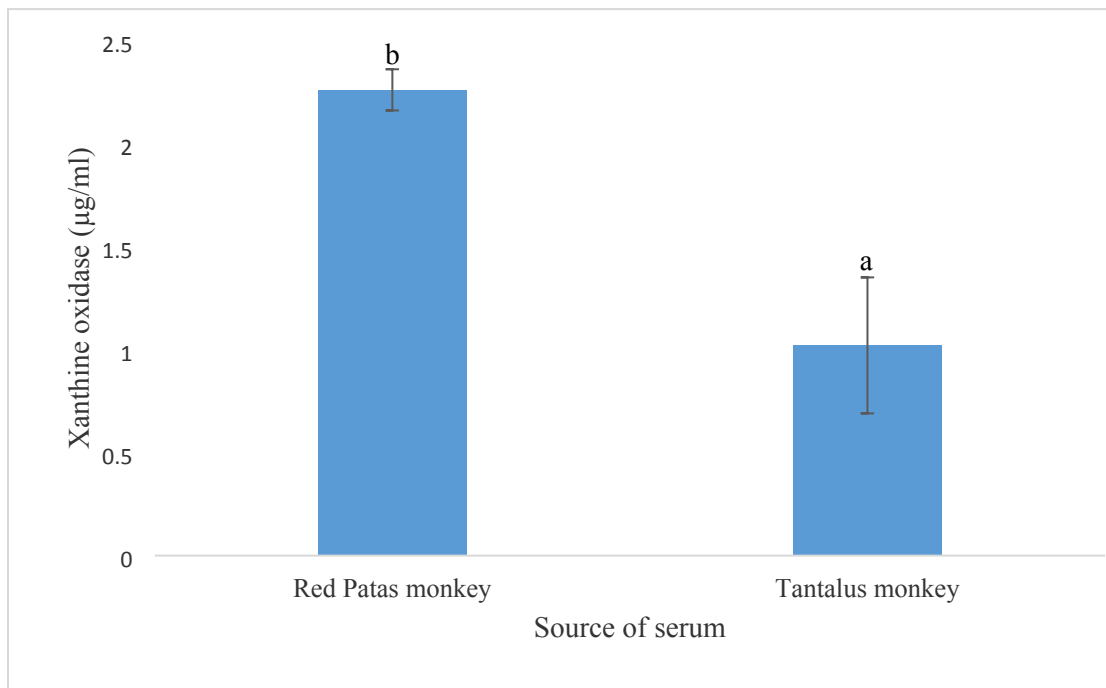
175 *Infectivity*

176 None of the parasites affected by the test sera *in vitro* was able to cause infection to the mice.

177 *Xanthine oxidase content of sera of selected zoo primates*

178 The xanthine oxidase content of sera of the selected zoo primates is presented in Figure 1. It was
179 observed that concentration of xanthine oxidase in the serum of red patas monkey was significantly
180 higher ($P < 0.05$) than that of Tantalus monkey (2.26 ± 0.10 and 1.02 ± 0.33 $\mu\text{g/ml}$ respective.

181



182

183 **Figure 1:** Xanthine oxidase content of sera of selected zoo primates

184 Values are mean concentrations. Values with different superscripts are significantly different
185 ($P < 0.05$).

186 Discussion

187 A simple technique was employed using the motility of trypanosomes as indicator of parasites
188 viability [9] as it has long been established that parasites motility could be a measure of viability
189 among most zooflagellate parasites [12]. Also, Atawodi *et al.* [13] reported that the technique
190 correlated well with other *in vitro* methods.

191 The fact that sera from the two zoo primates, Red patas monkey and Tantalus monkey showed a
192 slight anti-*Trypanosoma brucei brucei* activity Index (ATI of 0.27 and 0.34 respectively) and a
193 very high anti-*Trypanosoma congolense* activity Index *in vitro* (ATI of 0.91 and 0.90 respectively)
194 with a highly reduced mean survival time of the parasites of about 7.00 ± 1.73 hours for red patas
195 monkey and 7.67 ± 0.58 hours for tantalus monkey, demonstrate an innate immunity of these
196 primates to *T. congolense*, and to some lesser extend to *T. brucei brucei*. Human and several other
197 primates are known to be resistant to infection by several animal-specific trypanosome species
198 including *T. brucei brucei*, *T. congolense*, *T. vivax* and *T. evansi* [3]. The results obtained suggest
199 that Red pattas monkey serum as well as Tantalus monkey serum might possess some trypanolytic
200 factors against *T. congolense*, and to some lesser extend against *T. brucei brucei*. This claim can
201 be supported by the fact that the innate immunity of human and several other primates to the
202 trypanosomes is due to the expression of a unique subset of high-density lipoproteins (HDLs)
203 referred to as the trypanosome lytic factors (TLFs) in their blood [3]. Being that the trypanosome
204 lytic factor is primarily composed of Apolipoprotein L1 (APOL1) and a haptoglobin-related
205 protein [14], and considering the findings of Jirku *et al.*, [4] who demonstrated that mandrill serum
206 was able to efficiently lyse *T. brucei brucei* and *T. brucei rhodesiense*, and to some extent *T. bucei*
207 *gambiense*, while the chimpanzee serum failed to lyse any of these subspecies because of the
208 secondary loss of the APOL1 gene, the anti-trypanosomal activity of red pattas monkey and

209 tantalus monkey sera observed in this study could be attributable to Apolipoprotein L1 (APOL1).
210 These findings also correlate with some previous ones among which the identification of an
211 Apolipoprotein L1 (APOL1) in a subset of Old World monkeys [15], [16], [17], [18], the
212 demonstration of the *in vitro* lytic ability of serum and purified recombinant protein of an
213 Apolipoprotein L1 (APOL1) ortholog from the West African Guinea baboon (*Papio papio*), which
214 is able to lyse all subspecies of *T. brucei* including *T. brucei gambiense* [5].

215 The parasites were inactivated by the monkeys' sera *in vitro* and unable to cause infection to mice
216 suggesting that the anti-trypanosomal activity observed could be cidal. Considering the fact that
217 the sera were obtained from primates that had not previously been exposed to trypanosome
218 infection, these results are also suggestive of the nonspecific nature of the anti-trypanosomal
219 materials present in the sera. The difference in the xanthine oxidase content of sera from red patas
220 monkey and Tantalus monkey indicates that the similar anti-trypanosomal activity observed is not
221 associated to the activity of this enzyme. This finding gives more support to our suggestion on the
222 possible trypanolytic factor previously mentioned.

223

224 **Conclusion**

225 This study revealed anti-*Trypanosoma brucei brucei* and anti-*Trypanosoma congolense* properties
226 of the sera of the two zoo primates, Red patas monkey and Tantalus monkey *in vitro*. Both sera
227 had slight anti-*Trypanosoma brucei brucei* activity and a very high anti-*Trypanosoma congolense*
228 activity *in vitro*. These activities, showing some innate immunity is attributable to some
229 nonspecific factors (trypanolytic factors) present in the sera of the Red patas monkey and the
230 Tantalus monkey.

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237

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