

1 **The impact of allometry on vomer shape and its implications for the taxonomy and**  
2 **cranial kinesis of crown-group birds**

3

4 Olivia Plateau<sup>1\*</sup>, Christian Foth<sup>1</sup>

5 <sup>1</sup>Department of Geosciences, University of Fribourg, Chemin du Musée 6, CH-1700

6 Fribourg, Switzerland

7 \*e-mail: [olivia.plateau@unifr.ch](mailto:olivia.plateau@unifr.ch)

8

9 O.P. (ORCID: 0000-0002-8321-2687)

10 C.F. (ORCID: 0000-0002-9410-4569)

11

12 **Abstract**

13 Crown birds are subdivided into two main groups, Palaeognathae and Neognathae, that can  
14 be distinguished, among others, by the organization of the bones in their pterygoid-palatine  
15 complex (PPC). Shape variation to the vomer, which is the most anterior part of the PPC, was  
16 recently analysed by Hu et al. (2019) with help of geometric morphometrics to discover  
17 morphological differences between palaeognath and neognath birds. Based on this study, the  
18 vomer was identified as sufficient to distinguish the two main groups (and even more  
19 inclusive neognath groups) and their cranial kinetic system. As there are notable size  
20 differences between the skulls of palaeognaths and neognaths, we here investigate the impact  
21 of allometry on vomeral shape and its implication for taxonomic classification by re-  
22 analysing the data of the previous study. Different types of multivariate statistical analyses  
23 reveal that taxonomic identification based on vomeral shape is strongly impaired by  
24 allometry, as the error of correct identification is high when shape data is corrected for size.  
25 This finding is evident by a great overlap between palaeognath and neognath subclades in

26 morphospace. The correct identification is further influenced by the convergent presence of a  
27 flattened vomeral morphotype in multiple neognath subclades. As the evolution of cranial  
28 kinesis has been linked to vomeral shape in the original study, the existing correlation  
29 between shape and size of the vomer across different bird groups found in the present study  
30 questions this conclusion. In fact, cranial kinesis in crown birds results from the loss of the  
31 jugal-postorbital bar in the temporal region and ectopterygoid in the PPC and the  
32 combination of a mobilized quadrate-zygomatic arch complex and a flexible PPC. Therefore,  
33 we can conclude that the vomer itself is not a suitable proxy for exploring the evolution of  
34 cranial kinesis in crown birds and their ancestors.

35

## 36 **Introduction**

37 The pteryoid-palatine complex (PPC) of crown birds is mainly formed by five bones: the  
38 unpaired vomer that results from the fusion of the originally paired vomer elements and the  
39 paired pterygoids and palatines. Its general morphology was first studied by Huxley (1867),  
40 who distinguished the clade Palaeognathae from all other birds on the basis of palatal  
41 morphology. Although the PPC of palaeognaths is quite variable (McDowell 1948), it is  
42 characterized by a large vomer that is only partly fused. The pterygoids and palatines are  
43 highly connected, forming a rigid unit that articulates with the braincase via well-developed  
44 basiptyergoid processes, while a contact with the parasphenoid is not present (see Bellairs &  
45 Jenkin 1960; Zusi 1993; Gussekloo et al. 2001, Mayr 2017; Fig. 1A). In contrast, neognath  
46 birds possess a movable joint between pterygoid and palatine, which plays an important role  
47 in the kinematic movement of the upper jaw. Here, the pterygoid articulates with the  
48 parasphenoid, while the basiptyergoid processes are often reduced. The vomer is highly  
49 variable in size and shape and often has no connection with the upper jaw beyond an  
50 association with the nasal septum and the palatine. In some neognaths, the vomer is greatly

51 reduced or even absent (see Bellairs & Jenkin 1960; Bock 1964; Zusi 1993; Mayr 2017, Fig.  
52 1A).

53 In a recent paper, Hu et al. (2019) investigated palate evolution in crown birds and  
54 their stem, focusing on the morphology of the vomer. Using 3D geometric morphometrics,  
55 the study found that the vomeral shape of neognaths is clearly distinguishable from  
56 palaeognaths, in that the latter group has a stronger similarity with their non-avian ancestors.  
57 Linking vomer shape with the kinetic abilities of the skull, the authors concluded that cranial  
58 kinesis represents an innovation of Neognathae. Furthermore, the authors concluded that  
59 vomeral morphology allows for a taxonomic differentiation between the major groups of  
60 neognaths, namely Aequorlithornithes, Galloanseres, Gruiformes, and Inopinaves. However,  
61 according to their PCA results, all groups strongly overlap each other within PC1, while a  
62 taxonomic differentiation is only noticeable within PC2 (other principal components are not  
63 shown). Taking the great size variation of the vomer of neognath birds into account (Zusi  
64 1993), we wonder if the reported taxonomic differentiation between palaeognaths and the  
65 neognath subclades could alternatively be related to allometry, i.e. the dependence of shape  
66 on size (Klingenberg 1998), rather than pure shape variation. In order to test this hypothesis,  
67 we re-analysed the dataset of Hu et al. (2019), comparing allometric shape data with non-  
68 allometric residuals, and re-evaluating the role of the vomer in the evolution of cranial kinesis  
69 in crown birds.

70

## 71 **Materials and Methods**

72 The published 3D models and landmarks data of 41 specimens including 36 species were  
73 downloaded from Hu et al. 2019 (<https://doi.org/10.6084/m9.figshare.7769279.v2>). This  
74 dataset contains five extinct species (two stem line representatives: the troodontid  
75 *Sinovenator changii*, the Avialae *Sapeornis chaoyangensis*; and three fossil palaeognath

76 crown birds from the clade Dinornithiformes: *Pachyornis australis*, *Megalapteryx didinus*  
77 and *Diornis robustus*), five extant Paleognathae and 27 extant Neognathae representing the  
78 two major clades of crown birds.

79 The original landmarks data (Dataset A) is composed of five anatomical landmarks  
80 and 43 semi-landmarks (see Hu et al. 2019). The landmark data were imported into the  
81 software *R* v.3.5.2 (R Core Team, 2018). Using the *plotAllSpecimens* function of *Geomorph*  
82 v.3.2.1 (Adams et al. 2013) in *R*, we notice great variability for each anatomical landmark,  
83 resulting from two main shapes in the vomer. First, the majority of bird possesses a fused  
84 vomer that is bilaterally symmetric and roof-shaped in transection, having a horizontal  
85 orientation within the pterygoid-palatine complex (Fig. 1B). And second, some members of  
86 Aequorlornithes (e.g., *Podiceps nigricollis*, and *Podilymbus podiceps*), Galloanseres (e.g.,  
87 *Anas crecca*, *Anseranas semipalmata*, and *Cairina moschata*) and Inopinaves (e.g., *Aquila*  
88 *audax*, *Falco cenchroides*, and *Haliastur sphenurus*) have a fused vomer that is completely  
89 mediolaterally flattened in transection and vertically orientated within the pterygoid-palatine  
90 complex (Fig. 1B). Therefore, we created a second dataset (Dataset B), where species with  
91 flat vomer morphology were excluded. Furthermore, the palaeognath birds *Struthio camelus*  
92 and *Dromaius novaehollandiae* of the original Dataset A were represented by both juvenile  
93 and adult specimens. Because ontogenetic variation could, however, potentially impair size  
94 and position of the palaeognath morphospace, we removed the juvenile and subadult  
95 specimens of *S. camelus* and *D. novaehollandiae* in order to rerun the analysis just with adult  
96 semaphoronts (Dataset C). Finally, we created a fourth dataset, where both juvenile/subadult  
97 specimens and species with flat vomers were removed from the sample (Dataset D).

98 For superposition of the 3D landmark data, we followed Hu et al. (2019) by  
99 performing a Generalized Procrustes analysis (*GPA*). The *GPA* was done with the help of the  
100 *gpagen* function in *Geomorph*. Afterward, we performed a principal component analysis

101 (*PCA*) in order to visualize the shape variability of the vomer and the variance of  
102 morphospace for two groupings: (1) Paleognathae versus Neognathae and (2) Paleognathae,  
103 Inopinaves, Galloanserae, Gruiformes and Aequorlitorornithes. This was done with the  
104 *plotTangentSpace* function from *Geomorph*.

105         Because the vomer showed great variation in centroid size after superimposition,  
106 ranging from 14.60 (*Manorina melanocephala*) to 168.318 (*Dromaieus novehollandia*), we  
107 tested if there is a significant correlation between Procrustes coordinates and log-transformed  
108 centroid size (Goodall 1991) using the function *procD.lm* in *Geomorph*. This function  
109 performs a multivariate regression between the shape and size with a permutation of 10,000  
110 iterations. A significant relationship between both parameters indicates that the superimposed  
111 shape still contains an allometric signal. Based on this correlation we estimated non-  
112 allometric residuals of the Procrustes coordinates and repeated the *PCA*. In addition, we  
113 tested each of the first eleven PCs that together describe more than 95 of total variation for  
114 allometric signals.

115         To test for potential overlap in morphospace of vomer shapes in different clades of  
116 crown bird (see grouping 1 and 2) and their relation to the stem line representatives *S. changii*  
117 and *S. chaoyangensis*, we applied three different multivariate statistical methods, using the  
118 first eleven PCs as input data. We first applied a nonparametric multivariate analysis of  
119 variance (*perMANOVA*). This method evaluates the potential overlapping of groups in  
120 morphospace by testing the significance of their distribution on the basis of permutation  
121 (10,000 replications) and Euclidean distance (as one of several possible distance measures),  
122 not requiring normal distribution of the data (Anderson, 2001; Hammer & Harper, 2006). The  
123 spatial relationship of groups relative to each other is expressed by an *F* value and *p* value.  
124 For the five-group comparison, the *p* values were Bonferroni-corrected by multiplying the  
125 value with the number of comparisons. Next, we ran a discriminant analysis (*DA*), which

126 reduces a multivariate data set to a few dimensions by maximizing the separation between  
127 two or more groups using Mahalanobis distances. This distance measure is estimated from  
128 the pooled within-group covariance matrix, resulting in a linear discriminant classifier and an  
129 estimated group assignment for each species. The results were cross-validated using  
130 Jackknife resampling (Hammer & Harper, 2006; Hammer 2020). Both multivariate tests were  
131 done with the program *PAST v.4.03* (Hammer et al. 2001). Finally, we performed a  
132 phylogenetic flexible discriminant analysis (*pFDA*) (Schmitz & Motani, 2011; Motani &  
133 Schmitz, 2011) in *R*. This method removes the phylogenetic bias from the categorical  
134 variables before the actual discriminant analysis is undertaken by estimating Pagel's lambda,  
135 which tests how the grouping correlates with phylogeny. This was done for all allometric and  
136 non-allometric datasets.

137 For phylogeny, a set of 1,000 relaxed-clock molecular trees, which follow the  
138 topology of Hackett et al. (2008) and summarize the range of uncertainties in terms of time  
139 calibration of ancestral nodes, were downloaded from birdtree (<http://birdtree.org>) (Jetz et al.  
140 2012, 2014) including all extant bird species in the dataset (Supplementary Data S1). Due to  
141 uncertainties in the taxonomic identification of *Aquila sp.*, this specimen was removed from  
142 the sample as we could not include it in the phylogeny. Because the specimen occupies  
143 almost the same position as *Aquila audax*, we consider this deletion to have a negligible  
144 effect on the outcome of the analyses. Furthermore, the species *Sterna bergii* and *Grus*  
145 *rubicunda* used in the analysis from Hu et al. (2019) are junior synonyms of *Thalasseus*  
146 *bergii* (Bridge et al. 2005) and *Antigone rubicunda* (Krajewski et al. 2010). Using the  
147 function *consensus.edges* in the *R* package *phytools v.0.7-20*, we computed a temporal  
148 consensus. The extinct dinornithiform species were placed as sister-group to Tinamidae  
149 following Mitchell et al. (2014). Because of their recent extinction (Holdaway & Jacomb  
150 2000; Turvey & Holdaway 2005), the age was set to zero, similar to the other crown birds.

151 The stem line representatives *S. changii* and *S. chaoyangensis* were added following the time-  
152 calibrated phylogeny of Rauhut et al. (2019). Because of the presence of juvenile specimens  
153 in dataset A and B, we added the juvenile specimens by splitting OTU of *S. camelus* and *D.*  
154 *novehollandia* into a polytomy with each OUT having a branch length of one year (this value  
155 had to be standardized, as *pFDA* requires an isometric tree).

156 The error of correct identification from the resulting confusion matrices was  
157 compared between allometric and non-allometric data. For these comparisons, we used non-  
158 parametric *Mann-Whitney U* and *Kruskal-Wallis* tests, which both estimates, whether or not  
159 two univariate samples were taken from populations with equal medians, being more robust  
160 against small sample sizes and data without normal distribution (Hammer & Harper 2006).  
161 Both tests were run with *PAST*.

162 Finally, we applied for 19 species an ordinary least square regression analysis to test  
163 the correlation between log-transformed vomer and the skull size using a log-transformed box  
164 volume (Height x Width x Length). The measurements of the skull box volume were taken  
165 from skullsite (<https://skullsite.com>).

166

## 167 **Results**

168 Based on the *PCA* of the original dataset, the first two PCs explain over 52% (Fig. 2A) of  
169 total shape variation (PC1: 27.5%; PC2:25.1%). The morphospace of palaeognaths and  
170 neognaths is almost equal in size. Taking the small sample size of palaeognaths into account,  
171 the size of their morphospace indicates great shape variation. Both groups show a strong  
172 overlap along PC1 and a partial overlap along PC2. When comparing neognath subclades,  
173 aequorlornithines show strong overlap along both PCs with the palaeognath morphospace.  
174 Gruiforms lie in the overlapping area of both groups. The morphospace of inopinaves and

175 galloanserinae overlap with each other in both axes, but are separated from palaeognaths,  
176 aequorlitorrnithines and gruiforms along PC2.

177 Allometry summarizes about 6.4% of total shape variation. Using non-allometric  
178 residuals PC1 explains 29.3% and PC2 22.9% (Fig 2.B). While the general distribution of the  
179 single bird clades does not change along PC1, the groups are less separated along PC2, which  
180 contains the major allometric signal within the principal components (slope: -0.523;  $R^2$ :  
181 0.185;  $p$ : 0.005; predicted variation: 19.5%), which is 4.9% of total shape variation in the  
182 original dataset. Here, the palaeognath morphospace overlaps fully with aequorlitorrnithines  
183 and gruiforms, partly with inopinaves and marginally with galloanserinae. For the three other  
184 datasets, we observe more or less similar general trends before and after size correction,  
185 although the single morphospaces are partly shrunk. In all cases, the two stem line  
186 representatives *Sinovenator changii* and *Sapeornis chaoyangensis* lie within the marginal  
187 area of the palaeognaths/aequorlitorrnithines morphospace. Here, vomer morphology of the  
188 troodontid *S. changii* is more bird-like than that of the avialian *S. chaoyangensis*  
189 (Supplementary Data S2-S3).

190 In all studied datasets, the *perMANOVA* found a significant separation between  
191 palaeognath and neognath birds, showing no impact of allometry. For the five-group  
192 comparison of the original dataset (A), the overall results still indicate significant separation  
193 between clades for both allometric and non-allometric data. However, group-by-group  
194 comparison of allometric data indicates an overlap in morphospace of gruiformes with  
195 aequorlitorrnithines, inopinaves, galloanserinae and palaeognaths. These overlaps of  
196 gruiforms with other bird clades persist when allometry is removed from shape, but in  
197 addition, aequorlitorrnithines cannot be distinguished from palaeognaths anymore, as  
198 indicated by the *PCA* results (Fig. 2 A,B). The overlap between clades increases with the  
199 exclusion of species with flat vomers and non-adult semaphoronts.



200 For the original dataset (A) with allometry included, the *DA* identifies all species  
201 correctly as palaeognaths or neognaths. The error of false identification increases to 2.6% if  
202 the data are jack-knifed. When allometry is removed, the error increases to 13.2% before and  
203 36.8% after jack-knife resampling. In the former case, the misidentifications are restricted to  
204 neognath birds, which are wrongly classified as palaeognaths, while jack-knifing leads to  
205 identification errors in both groups. For the five-group comparison, all species of dataset (A)  
206 are correctly identified, when allometry is still present. The error is 18.4% after jack-knife  
207 resampling, showing minor mismatches in all clades. Excluding allometry, the error increases  
208 to 10.5% before and 47.4% after jack-knifing. While in the former case, a few  
209 aequorlornithine (2) and inopinave (1) species are wrongly identified as palaeognaths (Fig.  
210 2 C,D), palaeognaths cannot be separated from the neognath subclades anymore after  
211 resampling. The exclusion of species with flat vomers and non-adult semaphoronts leads to  
212 an increase of error.

213 The *pFDA* found 15.8% of wrong identifications when palaeognaths are compared  
214 with neognaths in the original dataset (A). This error increases to 31.6% if shape is corrected  
215 for allometry. In both cases, error is primarily based on the wrong identification of  
216 palaeognath specimens as neognaths. When palaeognaths are compared with neognath  
217 subclades, the error of correct identification is 10.5% before and 26.3% after allometry is  
218 removed from the data. For the allometric data, the misidentification result from the overlap  
219 between paleognaths, aequorlornithines and gruiforms. The misidentifications between  
220 these three groups are increased when shape is corrected for allometry, while inopinaves are  
221 in part also wrongly identified as palaeognaths. The exclusion of species with a flat vomer  
222 and/or non-adult semaphoronts usually causes a decrease of false identifications. However,  
223 the non-allometric dataset shows an increase in error for the two-group comparison, when  
224 species with flat vomers are excluded, and for the five-group comparison, when only adult

225 semaphoronts are taken into account (Fig. 2 E,F). Nevertheless, for all four datasets, the error  
226 of correct identification is significantly higher for non-allometric vomer shapes (Fig 3A,  
227 Supplementary Data S4).

228

## 229 **Discussion**

230 The skull of crown birds possesses a complex kinetic system that includes a mobilized  
231 quadrate, the zygomatic arch (= jugal bar) and the pterygoid-palatine complex (PPC) that  
232 allows for the simultaneous, but restricted motion of both jaws (Bock 1964; Zusi 1984).  
233 According to Zusi (1984), the kinetic system can be differentiated into three main types. (1)  
234 Prokinesis describes the rotation of the whole beak around the nasal-frontal hinge. (2)  
235 Amphikinesis is derived from prokinesis, including the rotation of the beak around the nasal-  
236 frontal hinge plus an additional flexion of the anterior portion of the beak. (3) In contrast,  
237 rhynchokinesis includes a simple flexion of the beak around one or several bending zones  
238 rostral to the nasal-frontal suture, lacking a true hinge. Depending on the position of the  
239 bending zones, rhynchokinesis can be further differentiated into five subtypes. Most  
240 palaeognath birds possess central rhynchokinesis, while neognaths have realized all types of  
241 cranial kinesis (Zusi 1984), including some taxa with akinetic skulls (Reid 1835; Sims 1955;  
242 Degrange et al. 2010). In the past, several authors (Hofer 1954; Simonetta 1960; Bock 1963)  
243 suggested a close relationship between the morphology of the PPC and type of cranial  
244 kinesis. However, Gussekloo et al. (2001) demonstrated that all types of kinesis present in  
245 crown birds have similar movements of the quadrate, zygomatic arch and PPC. Palaeognaths  
246 and neognaths only differ in the magnitude of kinetic movements in that palaeognaths have  
247 slightly more restricted movement due to their rigid palate missing a movable joint between  
248 the pterygoid and palatine (Gussekloo et al. 2005).

249           Thus, although the geometric morphometric results of the vomer shape by Hu et al.  
250 (2019) implicate at first view a distinct separation between palaeognaths and neognaths, this  
251 separation does not necessarily reflect their conclusions regarding the evolution of cranial  
252 kinesis in crown birds. As indicated by the *PCA*, palaeognaths occupy an enormous vomeral  
253 morphospace (Hu et al. 2019), which mirrors their generally large palatal disparity (see  
254 McDowell 1948) and partly overlaps with gruiforms and aequorlitorinithines. In all cases  
255 tested, however, the exclusion of allometric shape variation generally increases the error of  
256 misidentification between all groups, indicating that the taxonomic distinctions of shape  
257 found by Hu et al. (2019) are at least partly an artefact of size. This primarily concerns PC2,  
258 which according to Hu et al. (2019) separates palaeognaths from neognaths, but also contains  
259 the major allometric information. According to shape variation explained by PC2, larger birds  
260 tend to evolve vomers that are more dorsoventrally compressed. Only members of the  
261 galloanserinae could be still identified with a high amount of certainty when allometry is  
262 excluded.

263           Thus, our finding supports previous studies that demonstrated a relevant impact of  
264 allometry on skull shape evolution in birds (Klingenberg & Marugán-Lobón 2013; Bright et  
265 al. 2016; Linde-Medina 2016; Tokita et al. 2016, Bright et al. 2019). By modifying the  
266 dataset, it becomes further clear that both the homoplastic presence of flat vomers in  
267 aequorlitorinithines, inopinaves, galloanserinae (Dataset B) and ontogenetic variation (Dataset  
268 C) affects the accuracy of taxonomic identification. In addition, palaeognaths and neognaths  
269 do not differ in vomer size when compared to the head size (Fig. 3B). Consequently, vomer  
270 shape is not practical for taxonomic identification and should not be used as a proxy to infer  
271 the presence or absence of cranial kinesis in crown birds or their stem. In fact, *DA* and *pFDA*  
272 frequently identified the troodontid *Sinovenator changii* and avialan *Sapeornis*  
273 *chaoyangensis* as neognaths or neognath subclades when allometry is excluded, while the

274 original dataset implied a referral to palaeognaths (see also Hu et al. 2019). However, the  
275 skull anatomy of both species indicates no cranial kinesis (Xu et al. 2002; Wang et al. 2017;  
276 Yin et al. 2018; Hu et al. 2020).

277 The origin and evolution of cranial kinesis in the stem line of birds is still not well  
278 understood due to the rarity of complete three-dimensional skulls. However, skull material  
279 from the ornithurines *Ichthyornis dispar* and *Hesperornis regalis* indicates a certain degree  
280 of rhynchokinesis (Bühler et al. 1988; Field et al. 2018) that might be comparable to that of  
281 extant palaeognaths or some aequorlornithines, but further shows that this functional  
282 character was already present before the origin of the crown. Their kinesis is indicated by the  
283 loss of the jugal-postorbital bar and the ectopterygoid (resulting in a loss of contact in the  
284 jugal with the skull roof and the palate), the presence of a mobile bicondylar quadrate and a  
285 mobile joint between quadrate and quadratojugal. Recently, Plateau & Foth (2020) speculated  
286 that the peramorphic bone fusion in the braincase could be also related to cranial kinesis, in  
287 which the fusion-induced immobility constrains a controlled kinetic dorsoventral flexion of  
288 the avian beak during biting/picking. Based on these characters, most Mesozoic Avialae  
289 (including *Sapeornis chaoyangensis*) still had akinetic skulls, although some Enantiornithes  
290 possessing a reduced jugal-postorbital bar might have evolved primitive kinesis convergently  
291 to Ornithurae (O'Connor & Chiappe 2011).

292

### 293 **Acknowledgements**

294 We thank Walter Joyce for comments on the manuscript and the Swiss National Science  
295 Foundation (PZ00P2\_174040 to C.F.) for financial support.

296

### 297 **Additional information**

### 298 **Funding**

299 This study was funded by the Swiss National Science Foundation (PZ00P2\_174040).

300

### 301 **Competing interests**

302 The authors declare no competing interests.

303

### 304 **Author Approvals**

305 All authors have seen and approved the manuscript. The manuscript has not been accepted or  
306 published elsewhere.

307

### 308 **Author contributions**

309 O.P. and C.F. designed the research project and analysed the data; and O.P. and C.F. wrote  
310 the paper and prepared all figures.

311

### 312 **Data availability**

313 The 3D models and landmarks data of Hu et al. (2019) are available at Figshare (DOI:  
314 <https://doi.org/10.6084/m9.figshare.7769279.v2>).

315

### 316 **Supplementary information**

- 317 • Data S1: Phylogenetic trees used for *pFDA*.
- 318 • Data S2: PCA results of all dataset before and after correction for allometry.
- 319 • Data S3: PCA plots of Dataset B-D before and after correction for allometry.
- 320 • Data S4: Results of npMANOVA, DA and pFDA.
- 321 • Data S5: R Code including all statistical analyses.

322

### 323 **References**

- 324 Adams DC, Otárola-Castillo E. 2013. *geomorph*: an R package for the collection and analysis  
325 of geometric morphometric shape data. *Methods in Ecology and Evolution* 4:393–399.
- 326 Anderson MJ. 2001. A new method for non-parametric multivariate analysis of variance.  
327 *Austral Ecology* 26:32–46.
- 328 Bellairs ADA, Jenkin CR. 1960. The skeleton of birds. In: Marshall AJ ed. *Biology and*  
329 *Comparative Physiology of Birds, Vol. 1*. New York: Academic Press, 241–300.
- 330 Bock WJ. 1963. The cranial evidence for ratite affinities. *Proceedings of the 13th*  
331 *International Ornithological Congress* 1:39–54.
- 332 Bock WJ. 1964. Kinetics of the avian skull. *Journal of Morphology* 114:1–42.
- 333 Bridge ES, Jones AW, Baker AJ. 2005. A phylogenetic framework for the terns (Sternini)  
334 inferred from mtDNA sequences: implications for taxonomy and plumage evolution.  
335 *Molecular Phylogenetics and Evolution* 35:459–469.
- 336 Bright JA, Marugán-Lobón J, Cobb SN, Rayfield J. 2016. The shapes of bird beaks are highly  
337 controlled by nondietary factors. *Proceedings of the National Academy of Sciences,*  
338 *U.S.A.* 113:5352–5357.
- 339 Bright JA, Marugán-Lobón J, Rayfield J, Cobb SN. 2019. The multifactorial nature of beak  
340 and skull shape evolution in parrots and cockatoos (Psittaciformes). *BMC Evolutionary*  
341 *Biology* 19:1–9.
- 342 Bühler P, Martin LD, Witmer LM. 1988. Cranial kinesis in the Late Cretaceous birds  
343 *Hesperornis* and *Parahesperornis*. *The Auk* 105:111–122.
- 344 Degrange F, Tambussi C, Moreno K, Witmer L, Wroe S. 2010. Mechanical analysis of  
345 feeding behavior in the extinct “Terror Bird” *Andalgalornis steulleti* (Gruiformes:  
346 Phorusrhacidae). *PLoS ONE* 5:e11856.
- 347 Field DJ, Hanson M, Burnham DA, Wilson LE, Super K, Ehret D, Ebersole JA, Bhullar B-  
348 AS. 2018. Complete *Ichthyornis* skull illuminates mosaic assembly of the avian head.

- 349 *Nature* 557:96–100.
- 350 Goodall C. 1991. Procrustes methods in the statistical analysis of shape. *Journal of the Royal*  
351 *Statistical Society: Series B (Methodological)* 53:285–321.
- 352 Gussekloo SWS, Bout RG. 2005. The kinematics of feeding and drinking in palaeognathous  
353 birds in relation to cranial morphology. *The Journal of Experimental Biology* 208:3395–  
354 3407.
- 355 Gussekloo SWS, Vosselman MG, Bout RG. 2001. Three-dimensional kinematics of skeletal  
356 elements in avian prokinetic and rhynchokinetic skull determined by roentgen  
357 stereophotogrammetrics. *The Journal of Experimental Biology* 204:1735–1744.
- 358 Hackett SJ, Kimball RT, Reddy S, Browie RCK, Braun EL, Chojnowski JL, Cox A, Han K,  
359 Harshman J, Huddleston CJ, Marks BD, Miglia KJ, Moore WS, Sheldon FH, Steadman  
360 DW, Witt CC, Yuri T. 2008. A phylogenomic study of birds reveals their evolutionary  
361 history. *Science* 320:1763–1768.
- 362 Hammer O, Harper DAT. 2006. *Paleontological Data Analysis*. Malden: Blackwell  
363 Publishing.
- 364 Hammer O, Harper DAT, Ryan PD. 2001. PAST: paleontological statistics software package  
365 for education and data analysis. *Palaeontologia Electronica* 4:1–9.
- 366 Hammer O. 2020. *PAST Paleontological Statistics v.4.03. Reference Manual*. Oslo:  
367 University of Oslo.
- 368 Hofer H. 1954. Neuere Untersuchungen zur Kopfmorphologie der Vögel. *Acta XI Congressus*  
369 *Internationalis Ornithologici*:104–137.
- 370 Holdaway RN, Jacomb C. 2000. Rapid extinction of the moas (Aves: Dinornithiformes):  
371 model, test, and implications. *Science* 287:2250–2254.
- 372 Hu H, O'Connor J, McDonald P, Wroe S. 2020. Cranial osteology of the Early Cretaceous  
373 *Sapeornis chaoyangensis* (Aves: Pygostylia). *Cretaceous Research* 113:104496.

- 374 Hu H, Sansalone G, Wroe S, McDonald PG, O'Connor JK, Li Z, Xu X, Zhou Z. 2019.  
375 Evolution of the vomer and its implications for cranial kinesis in Paraves. *Proceedings*  
376 *of the National Academy of Sciences, U.S.A.* 116:19571–19578.
- 377 Huxley TH. 1867. On the classification of birds; and on the taxonomic value of the  
378 modifications of certain cranial bones observable in the class. *Proceedings of the*  
379 *Zoological Society of London* 27:415–472.
- 380 Jetz W, Thomas GH, Joy JB, Hartmann K, Mooers AO. 2012. The global diversity of birds in  
381 space and time. *Nature* 491:444–448.
- 382 Jetz W, Thomas GH, Joy JB, Redding DW, Hartmann K, Mooers AO. 2014. Global  
383 distribution and conservation of evolutionary distinctness in birds. *Current Biology*  
384 24:1–12.
- 385 Klingenberg CP. 1998. Heterochrony and allometry: the analysis of evolutionary change in  
386 ontogeny. *Biological Reviews* 73:79–123.
- 387 Klingenberg CP, Marugán-Lobón J. 2013. Evolutionary covariation in geometric  
388 morphometric data: analyzing integration, modularity, and allometry in phylogenetic  
389 context. *Systematic Biology* 62:591–610.
- 390 Krajewski C, Sipiorski JT, Anderson FE. 2010. Complete mitochondrial genome sequences  
391 and the phylogeny of cranes (Gruiformes: Gruidae). *The Auk* 127:440–452.
- 392 Linde-Medina M. 2016. Testing the cranial evolutionary allometric “rule” in Galliformes.  
393 *Journal of Evolutionary Biology* 29:1873–1878.
- 394 Mayr G. 2017. *Avian evolution*. Chichester: John Wiley.
- 395 McDowell S. 1948. The bony palate of birds. Part I. The Palaeognathae. *The Auk* 65:520–  
396 549.
- 397 Mitchell K, Llamas B, Soubrier J, Rawlence N, Worthy T, Wood J, Lee M, Cooper A. 2014.  
398 Ancient DNA reveals elephant birds and kiwi are sister taxa and clarifies ratite bird



- 399 evolution. *Science* 344:898–900.
- 400 Motani R, Schmitz L. 2011. Phylogenetic versus functional signals in the evolution of form-  
401 function relationships in terrestrial vision. *Evolution* 65:2245–2257.
- 402 O’Connor JK, Chiappe LM. 2011. A revision of enantiornithine (Aves: Ornithothoraces)  
403 skull morphology. *Journal of Systematic Palaeontology* 9:135–157.
- 404 Plateau O, Foth C. 2020. Birds have peramorphic skulls, too: anatomical network analyses  
405 reveal oppositional heterochronies in avian skull evolution. *Communications Biology* 3.
- 406 R Development Core Team. 2011. *R: a language and environment for statistical computing*.  
407 <http://www.r-project.org>.
- 408 Rauhut OWM, Tischlinger H, Foth C. 2019. A non-archaeopterygid avialan theropod from  
409 the Late Jurassic of southern Germany. *eLife* 8:e43789.
- 410 Reid J. 1835. Anatomical description of the Patagonian penguin. *Proceedings of the*  
411 *Zoological Society of London* 3:132–148.
- 412 Revell LJ. 2012. *phytools*: an R package for phylogenetic comparative biology (and other  
413 things). *Methods in Ecology and Evolution* 3:217–223.
- 414 Schmitz L, Motani R. 2011. Nocturnality in dinosaurs inferred from scleral ring and orbit  
415 morphology. *Science* 332:705–708.
- 416 Simonetta A. 1960. On the mechanical implications of the avian skull and their bearing on  
417 the evolution and classification of birds. *The Quarterly Review of Biology* 35:206–220.
- 418 Sims S. 1955. The morphology of the head of the hawfinch (*Coccothraustes coccothraustes*).  
419 *Bulletin of the British Museum* 2:371–393.
- 420 Tokita M, Yano W, James H, Abzhanov A. 2016. Cranial shape evolution in adaptive  
421 radiations of birds: comparative morphometrics of Darwin’s finches and Hawaiian  
422 honeycreepers. *Philosophical Transactions of the Royal Society B* 372:20150481.
- 423 Turvey ST, Holdaway RN. 2005. Postnatal ontogeny, population structure, and extinction of

- 424 the giant moa *Dinornis*. *Journal of Morphology* 265:70–86.
- 425 Wang Y, Hu H, O'Connor JK, Wang M, Xu X, Zhou Z, Wang X, Zheng X. 2017. A  
426 previously undescribed specimen reveals new information on the dentition of *Sapeornis*  
427 *chaoyangensis*. *Cretaceous Research* 74:1–10
- 428 Xu X, Norell MA, Wang X, Makovicky PJ, Wu X. 2002. A basal troodontid from the Early  
429 Cretaceous of China. *Nature* 415:780–784.
- 430 Yin Y, Pei R, Zhou C. 2018. Cranial morphology of *Sinovenator changii* (Theropoda:  
431 Troodontidae) on the new material from the Yixian Formation of western Liaoning,  
432 China. *PeerJ* 6:e4977. DOI: 10.7717/peerj.4977.
- 433 Zusi RL. 1984. A functional and evolutionary analysis of rynchokinesis in birds.  
434 *Smithsonian Contributions to Zoology* 395:1-40.
- 435 Zusi RL. 1993. Patterns of diversity in the avian skull. In: Hanken J, Hall BK eds. *The skull*.  
436 *Vol. 2. Patterns of structural and systematic diversity*. Chicago: University of Chicago  
437 Press, 391–437.

438

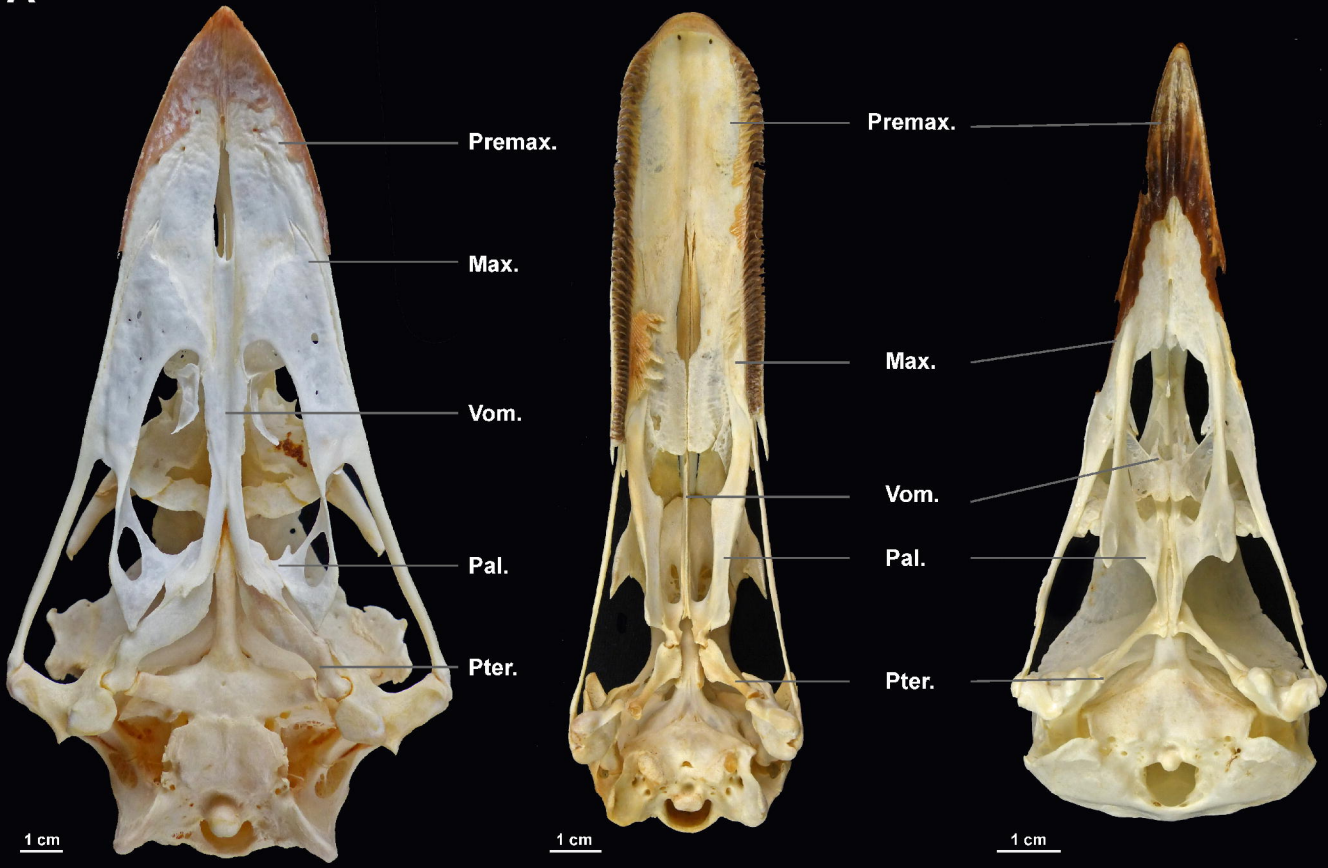
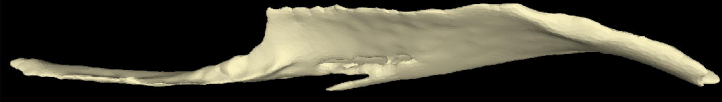
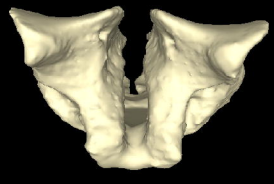
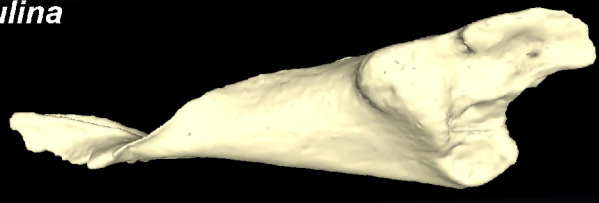
### 439 **Figure legends**

440 **Fig. 1. Anatomical organization of the pterygoid-palatine complex (PPC) and shape**  
441 **variability of the vomer in palaeognath and neognath birds. (A)** Palates of *Dromaius*  
442 *novaehollandia* (left), *Cygnus olor* (middle) and *Corvus corax* (right) in ventral view (all  
443 specimens form the Natural History Museum of Fribourg/University of Fribourg). **(B)** 3D  
444 models of the vomer of *Dinornis robustus*, *Anas crecca* and *Corvus sp.* in lateral view (left)  
445 and anterior view (right) from (not at scale) (3D models from Hu et al. 2019). Max,  
446 Maxillary; Pal, Palatine; Premax, Premaxillary; Pter, Pterygoid; Vom, Vomer.

447

448 **Fig. 2. Differences between allometric and non-allometric morphospaces of the vomer**  
449 **(Dataset A) in palaeognath and neognath birds. (A) PCA** results of allometric data. **(B)**  
450 *PCA* results of non-allometric data. **(C) DA** results of allometric data. **(D) DA** results of non-  
451 allometric data. **(E) pFDA** results of allometric data. **(F) pFDA** results of non-allometric data.  
452

453 **Fig. 3. Errors of correct taxonomic identification for all comparisons of Dataset A-D.**  
454 **(A)** Two-group identification (Palaeognathae and Neognathae) before (red) and after (green)  
455 correction for allometry. *DA*, Discriminant analysis; *DA\*JK*, Discriminant Analysis with  
456 jackknife resampling; *pFDA*, phylogenetic Flexible Discriminant Analysis. **(B)** Five-group  
457 identification (Palaeognathae, Aequorlitorhithes, Galloanserae, Gruiformes and Inopinaves).  
458 **(C)** *OLS* regression (black line) between log-transformed skull box volume and log-  
459 transformed centroid size of the vomer. Grey shadow mark the area of the 95% confidence  
460 interval.

**A***D. novaehollandiae**C. olor**C. corax***B***D. robustus**A. crecca**S. graculina*

Dorsal  
Anterior

Dorsal  
Lateral

