

1 **Draft Genome Sequences of Seven Strains of *Dickeya dadantii*, a Quick**

2 **Decline-causing Pathogen in Fruit Trees, Isolated from Japan**

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9 **Running title:** Genome sequences of 7 *D. dadantii* strains in Japan

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12 **Abstract**

13 Plant pathogenic bacterium *Dickeya dadantii* causes quick decline in fruit trees (apple,  
14 Japanese pear, and peach). In this study, we report on the draft genome sequences of  
15 seven strains of *D. dadantii* isolated from fruit trees with typical quick decline  
16 symptoms in Japan.

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18 *Dickeya dadantii* is a plant pathogenic bacterium that causes soft rot disease in various  
19 plants (1). In addition to being a soft-rot pathogen, it causes quick decline (QD) in fruit  
20 trees, such as apple, peach, and Japanese pear trees (2, 3). The symptoms of QD include  
21 red-brown sap leakage from the trunk and/or branches, softening bark, defoliation, leaf  
22 and shoot necrosis, and dieback. However, the lifecycle of *D. dadantii* is not yet well  
23 understood. The genetic characteristics of this bacterium will need to be defined to  
24 develop effective measures for the control of QD. To this end, we performed  
25 whole-genome sequencing (WGS) of seven *D. dadantii* strains isolated from fruit trees  
26 with symptoms of QD.

27 The strains sequenced in this study were obtained from the Institute of Fruit Tree  
28 and Tea Science (NARO) (Table 1), isolated from apple, peach, and Japanese pear trees,  
29 and identified as *D. dadantii* according to our previous studies (2, 3). The strains were  
30 cultivated in YP broth (2) at 27°C for 1 day with agitation at 140 rpm. Then, 1-ml

31 aliquots of each culture were used for DNA extraction with a DNeasy mini kit (Qiagen,  
32 Hilden, Germany). Their genome sequences were determined using WGS, as previously  
33 reported (4, 5, 6, 7). Briefly, the genomic DNA was sequenced using an Ion PGM  
34 sequencer with an Ion PGM Hi-Q View OT2 kit, an Ion PGM Hi-Q View Sequencing  
35 kit, and a 318 Chip kit v2 (Thermo Fisher Scientific, Waltham, MA, USA), according to  
36 the manufacturer's instructions. Default parameters were used unless specified  
37 otherwise. The sequence reads were quality controlled (quality score <20) and the  
38 adapter sequences were removed using the CLC Genomics Workbench (version 10,  
39 except for Kunimi3-1 (ver. 12), and BI1-1 (ver. 20)). Using the resulting reads, contigs  
40 (filtered with a size >500 bp) were assembled *de novo* using the CLC Genomics  
41 Workbench. The draft genomes were annotated using the NCBI Prokaryote Genome  
42 Annotation Pipeline (PGAP).

43 The WGS analysis indicated that the genome size of the seven strains was 4.7-4.9  
44 Mbp with a G+C content of 56.1-56.4% (Table 1). The genome information of this  
45 species was previously published in NCBI as 4.7-5.0 Mbp with a G+C content of  
46 56.3-56.5% (e.g. strains 3937 and DSM 18020; GenBank accession no. CP002038 and  
47 CP023467, respectively), which supports the results of our WGS analysis. Moreover,  
48 PGAP identified 4,311-4,488 genes and multiple rRNA and tRNA genes in these  
49 genomes (Table 1). This information can be used to compare the genomes and gene

50 expression patterns of different strains or species. Therefore, the results of our WGS  
51 analysis may help to elucidate the characteristics of *D. dadantii*, a bacterium related to  
52 the virulence of QD.

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#### 54 **Data availability**

55 All WGS projects in this study have been deposited at DDBJ/ENA/GenBank. The  
56 corresponding read data are available from the Sequence Read Archive (SRA) with the  
57 accession numbers provided in Table 1.

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#### 66 **References**

67 1. Reverchon S, Nasser W. 2013. *Dickeya* ecology, environment sensing and regulation  
68 of virulence programme. Environ Microbiol Rep 5:622–636.

- 69 2. Ota N, Fujikawa T. 2020. Reproduction of bacterial quick decline of fruit trees after  
70 soil inoculation with *Dickeya dadantii*. J Gen Plant Pathol 86:199–204.
- 71 3. Fujikawa T, Ota N, Sasaki M, Nakamura T, Iwanami T. 2019. Emergence of apple  
72 bacterial quick decline caused by *Dickeya dadantii* in Japan. J Gen Plant Pathol  
73 85:314–319.
- 74 4. Fujikawa T, Sawada H. 2016. Genome analysis of the kiwifruit canker pathogen  
75 *Pseudomonas syringae* pv. *actinidiae* biovar 5. Sci Rep 6:21399.
- 76 5. Fujikawa T, Sawada H. 2019. Genome analysis of *Pseudomonas syringae* pv.  
77 *actinidiae* biovar 6, which produces the phytotoxins, phaseolotoxin and coronatine.  
78 Sci Rep 9:3836.
- 79 6. Nakazono-Nagaoka E, Fujikawa T, Shikata A, Tachaapaikoon C, Waeonukul R,  
80 Pason P, Ratanakhanokchai K, Kosugi A. 2019. Draft genome sequence data of  
81 *Clostridium thermocellum* PAL5 possessing high cellulose-degradation ability. Data  
82 in Brief 25:104274.
- 83 7. Sawada H, Fujikawa T, Nishiwaki Y, Horita H. 2020. *Pseudomonas kitaguniensis* sp.  
84 Nov., a pathogen causing bacterial rot of Welsh onion in Japan. Int J Sys Evol  
85 Microbiol. <https://doi.org/10.1099/ijsem.0.004123>

1 **Table 1. Genome data and accession numbers of seven *Dickeya dadantii*.**

Strain	Strain information		Genome information					PGAP <sup>a</sup> annotation			Reads information			
	Isolation host	Isolation area	GenBank accession no.	Genome size (bp)	G+C content (mol%)	No. of contigs	$N_{50}$	Total no. of genes	rRNAs (5S, 16S, 23S)	tRNAs	SRA <sup>b</sup> accession no.	No. of reads	Average length (bp)	Genome coverage (x)
BI1-1	Apple	Japan: Iwate	JABEOZ00000000	4,663,807	56.2	271	39,637	4,311	1, 2, 1	46	SRR11696188	673,373	229.2	33.1
BI3-1	Apple	Japan: Iwate	PHRA00000000	4,771,676	56.3	92	151,183	4,331	4, 1, 1	61	SRR11692751	5,046,907	298.9	316.1
Aka1-1	Peach	Japan: Fukushima	JABEPA00000000	4,714,768	56.4	268	43,532	4,429	5, 1, 1	57	SRR11696187	592,688	258.8	32.5
Yana2-2	Peach	Japan: Fukushima	JABEPB00000000	4,825,596	56.4	204	61,681	4,432	3, 2, 1	48	SRR11696186	1,510,292	255.4	79.9
Kunimi-3	Peach	Japan: Fukushima	SMHE00000000	4,869,298	56.5	141	96,135	4,446	2, 1, 2	62	SRR11730645	1,941,545	287.6	114.7
Kousui1-1	Japanese pear	Japan: Saga	JABEPC00000000	4,881,710	56.1	193	68,290	4,488	5, 1, 3	53	SRR11696185	1,308,248	263.2	70.5
Housui2-1	Japanese pear	Japan: Saga	JABEPD00000000	4,882,493	56.2	197	56,134	4,481	4, 1, 6	49	SRR11696184	1,746,612	258.4	92.4

2 <sup>a</sup>NCBI Pipeline Genome Annotation Pipeline

3 <sup>b</sup>short read archive