- 1 MESMERISED: Super-accelerating T₁ relaxometry and diffusion MRI with
- 2 STEAM at 7 T for quantitative multi-contrast and diffusion imaging F.J. Fritz a,b, B.A. Poser a, A. Roebroeck a
 - ^a Department of Cognitive Neuroscience, Faculty of Psychology and Neuroscience, Maastricht University, Maastricht, The Netherlands, ^b Institut für Systemische Neurowissenschaften, Zentrum für Experimentelle Medizin, Universitätklinikum Hamburg-Eppendorf (UKE), Hamburg, Deutschland.

Abstract

There is an increasing interest in quantitative imaging of T₁, T₂ and diffusion contrast in the brain due to greater robustness against bias fields and artifacts, as well as better biophysical interpretability in terms of microstructure. However, acquisition time constraints are a challenge, particularly when multiple quantitative contrasts are desired and when extensive sampling of diffusion directions, high b-values or long diffusion times are needed for multi-compartment microstructure modeling. Although ultra-high fields of 7 T and above have desirable properties for many MR modalities, the shortening T₂ and the high specific absorption rate (SAR) of inversion and refocusing pulses bring great challenges to quantitative T₁, T₂ and diffusion imaging. Here, we present the MESMERISED sequence (Multiplexed Echo Shifted Multiband Excited and Recalled Imaging of STEAM Encoded Diffusion). MESMERISED removes the dead time in Stimulated Echo Acquisition Mode (STEAM) imaging by an echo-shifting mechanism. The echo-shift (ES) factor is independent of multiband (MB) acceleration and allows for very high multiplicative (ESxMB) acceleration factors, particularly under moderate and long mixing times. This results in super-acceleration and high time efficiency at 7 T for quantitative T₁ and diffusion imaging, while also retaining the capacity to perform quantitative T₂ and B₁ mapping. We demonstrate the super-acceleration of MESMERISED for whole-brain T₁ relaxometry with total acceleration factors up to 36 at 1.8 mm isotropic resolution, and up to 54 at 1.25 mm resolution qT₁ imaging, corresponding to a 6x and 9x speedup, respectively, compared to MB-only accelerated acquisitions. We then demonstrate highly efficient diffusion MRI with high b-values and long diffusion times in two separate cases. First, we show that super-accelerated multi-shell diffusion acquisitions with 370 whole-brain diffusion volumes over 8 b-value shells up to b = 7000 s/mm² can be generated at 2 mm isotropic in under 8 minutes, a data rate of almost a volume per second, or at 1.8 mm isotropic in under 11 minutes, achieving up to 3.4x speedup compared to MB-only. A comparison of b = 7000 s/mm² MESMERISED against standard MB pulsed gradient spin echo (PGSE) diffusion imaging shows 70% higher SNR efficiency and greater effectiveness in supporting complex diffusion signal modeling. Second, we demonstrate time-efficient sampling of different diffusion times with 1.8 mm isotropic diffusion data acquired at four diffusion times up to 290 ms, which supports both Diffusion Tensor Imaging (DTI) and Diffusion Kurtosis Imaging (DKI) at each diffusion time. Finally, we demonstrate how adding quantitative T₂ and B₁⁺ mapping to super-accelerated qT₁ and diffusion imaging enables efficient quantitative multi-contrast mapping with the same MESMERISED sequence and the same readout train. MESMERISED extends possibilities to efficiently probe T₁, T₂ and diffusion contrast for multi-component modeling of tissue microstructure.

Keywords

Quantitative MRI, Diffusion MRI, Multi-contrast MRI, 7 T, Accelerated Imaging, Microstructure

Introduction

3

4 5

6 7

8

9

10

11

12 13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

Magnetic resonance imaging (MRI) derives most of its versatility as an imaging technique from being able to weight images towards a multitude of contrasts, prominently among which are T₁, T₂/T₂* and diffusion contrasts. Recently, there is an increasing interest in quantitative imaging of these contrasts for two main reasons. First, quantitative T₁, T₂ and diffusion images possess greater robustness against bias fields and artifacts, since the effects of spatially slowly varying radio frequency (RF) transmit and receive fields are mostly removed. Second, quantitative images have better biophysical interpretability, since they have interpretable physical units. This makes them a better proxy for biological microstructure and phenomena, such as myelin (e.g. Callaghan et al., 2015; Lutti et al., 2014; Weiskopf et al., 2013), g-ratio (e.g. Berman et al., 2019; Campbell et al., 2018), iron (e.g. Fukunaga et al., 2010; Stuber et al., 2014; Weiskopf et al., 2013), axonal density (e.g. Assaf and Basser, 2005; Zhang et al., 2012) as well as mesoscopic structure due to random axonal packing (e.g. Novikov et al., 2014). However, since quantitative MRI (qMRI) requires the acquisition of many volumes, from which the quantitative image is then derived, long acquisition times are a challenge, particularly when multiple quantitative contrasts are desired.

Acquisition time constraints are particularly pressing for diffusion MRI (dMRI). The signal varies with the orientation, strength and diffusion time encoded by diffusion gradients, making diffusion contrast inherently multi-dimensional. Moreover, recent developments in biophysical microstructure quantification with dMRI require sampling more of the multi-dimensional contrast space than with older methods. The conventional method for the analysis of white matter in dMRI imaging is the tensor model in Diffusion Tensor Imaging (DTI; Basser et al., 1994; Pierpaoli et al., 1996). Although a DTI derived index variable such as Fractional Anisotropy (FA) is sensitive, it is also unspecific, since differences in FA can reflect different axonal properties such as axon density, diameter distribution and myelination (Assaf et al., 2004; Assaf and Pasternak, 2008; Beaulieu, 2002; De Santis et al., 2014; Jones et al., 2013). Therefore, advances in diffusion microstructure modeling have aimed at greater specificity than DTI in relating the dMRI signal to the underlying cellular microstructure (Alexander et al., 2010; Assaf et al., 2013; Assaf and Basser, 2005; Assaf et al., 2008; Assaf et al., 2004; De Santis et al., 2014; Fieremans et al., 2013; Jelescu et al., 2015; Panagiotaki et al., 2012; Santis et al., 2014; Zhang et al., 2012). Models such as the Neurite Orientation Dispersion and Density Imaging model (NODDI; Zhang et al., 2012), the Combined Hindered And Restricted Model of Diffusion (CHARMED; Assaf and Basser, 2005), and the White Matter Tract Integrity model (WMTI; Fieremans et al., 2013) can provide measures specific to fiber density and orientation dispersion. These require the acquisition of a large number of diffusion directions over multiple b-values, sometimes extending to b-values as high as 6000 - 8000 s/mm², easily leading to acquisition times of 10 - 20 minutes or longer. In addition, sensitivity can be gained to axonal diameter distributions (Alexander et al., 2010; Assaf et al., 2008; De Santis et al., 2016a; Zhang et al., 2011), water-exchange over cell membranes (Li et al., 2016; Nilsson et al., 2013) and mesoscopic structural effects due to random axonal packing (Burcaw et al., 2015; Novikov et al., 2014), but this requires the acquisition of a large number of bvalues and diffusion times extending to 200 - 800 ms, often leading to acquisition times of 30 - 60 min.

Developments in echo-planar imaging (EPI) acquisition have allowed significant speed-up in the acquisition of volume time series weighted for T_1 , T_2/T_2* or diffusion. Prominently among these is multiband (MB) or simultaneous multi-slice (SMS) acquisition (Feinberg et al., 2010; Larkman et al., 2001; Moeller et al., 2010; Setsompop et al., 2012), which allows 2- to 4-fold speed-up for high flip

angle applications at the expense of a higher specific absorption rate (SAR). However, even with MB acceleration, quantitative multi-contrast studies that require extensive diffusion MRI, as well as mapping of T_1 and/or T_2 , are facing prohibitively long acquisition times.

In the domain of high spatial resolution imaging, ultra-high fields (UHF) of 7 Tesla (7 T) and above have enabled efficient high spatial resolution mapping of quantitative T₁ (e.g. Marques et al., 2010; O'Brien et al., 2014), T₂* (e.g. Fukunaga et al., 2010) and diffusion (e.g. Vu et al., 2015). In this work, our main aim was to establish whether the high signal-to-noise ratio (SNR) available at 7 T could be leveraged for highly time-efficient quantitative multi-contrast acquisition at moderate resolutions, particularly when including dMRI at high b-values or long diffusion times. Although UHF has desirable properties for many MR modalities, the shortening of T₂ and the high SAR load of inversion and refocusing pulses bring challenges to efficient quantitative T₁, T₂ and diffusion imaging. Diffusion MRI sequences such as pulsed-gradient spin-echo (PGSE; e.g. Vu et al., 2015) and steady-state free precession (SSFP; e.g. Lu et al., 2012) have been optimized at 7 T to mitigate some of these limitations. However, they remain limited to MB acceleration factors of 2 to 4 (Eichner et al., 2014; Vu et al., 2015; Wu et al., 2013), with SAR as the main limiting factor.

Therefore, we investigated the possibilities of the Stimulated Echo Acquisition Mode (STEAM) sequence (Frahm et al., 1985). The STEAM sequence has some attractive advantages for multicontrast MRI at UHF. First, it is highly multi-contrast capable, being able to provide quantitative T₁ (qT_1) contrast (by relaxometry on the mixing time, TM), quantitative T_2 (qT_2) contrast (by relaxometry on the echo time, TE), transmit (B₁⁺) field mapping, and quantitative diffusion contrast. Furthermore, both b-value and diffusion time parameters in dMRI with STEAM can be probed at the high values desirable for many diffusion microstructure applications. Second, both a spin echo (SE) and stimulated echo (STE) signal can be independently acquired after a single excitation pulse, enhancing the capacity for efficient multi-contrast acquisition. Third, STEAM is highly efficient in SAR given its preferred 90°-90°-90° flip angle scheme. It has ~60% of the SAR of a comparable 90°-180° spin-echo sequence and ~33% of the SAR of a comparable 180°-90°-180° inversion-prepared spin-echo sequence (assuming similar slice-selection and a quadratic SAR/flip angle relationship). This also means that applying multiband pulses for SMS imaging is easier to apply for high MB factors in STEAM since SAR increases linearly with MB factor. However, the STEAM sequence also has inherent disadvantages. Both the SE and STE signals have only 50% of the signal level and signal to noise ratio (SNR), compared to the SE from a 90°-180° spin-echo sequence. Moreover, there is a considerable dead time in STEAM (Figure 1A), especially when moderate to long mixing times (useful for T₁weighting and probing of long diffusion times) are required, which leads to a low SNR per unit time efficiency.

Here, we present the MESMERISED sequence (Multiplexed Echo Shifted Multiband Excited and Recalled Imaging of STEAM Encoded Diffusion). MESMERISED removes the dead time in STEAM imaging by an echo-shifting mechanism. The echo-shift (ES) factor is independent of MB acceleration and allows for very high multiplicative ESxMB acceleration factors, particularly under moderate and long TM's. This results in high duty cycle and high time efficiency at 7 T, particularly for quantitative T₁ and diffusion imaging. In related work, Feinberg et al. (2010) introduced a different echo-shifting mechanism, termed simultaneous image refocusing (SIR) for gradient-echo EPI and pulsed-gradient spin echo (PGSE) EPI diffusion sequences. It also introduced the combined use of multiplexing SIR with MB acceleration to achieve very high multiplicative acceleration factors, closely related to the multiplexing of ES and MB in MESMERISED. Echo-shifting has been most applied in gradient-echo or

steady-state free precession sequences to achieve echo times longer than their repetition time (Ehses et al., 2015; Gibson et al., 2006). Shifted slice orderings to make more effective use of dead time have been introduced early on in inversion recovery EPI for quantitative T₁ mapping (Ordidge et al., 1990) and EPI based mapping has long been recognized for its high SNR per unit time (Mansfield et al., 1986). More recently, a multi-slice, inversion recovery EPI sequence (MS-IR-EPI) was proposed to enable T₁ mapping with high spatial resolution and fidelity, making effective use of shifted slice orderings and simultaneous multi-slice acquisition (Panchuelo et al., 2021). The possibility of increasing the duty cycle of STEAM by exciting other slices or bands during the mixing time has recently been suggested (Zhang et al., 2019). To our knowledge, we here present the first demonstration of the echo-shifting mechanism in STEAM for quantitative T₁ and diffusion mapping, and of the block reordering property (Figure 1C) which allows high acceleration and full acquisition duty cycle for multiple mixing times or diffusion times at the same TR.

We demonstrate the super-acceleration of MESMERISED for whole-brain T_1 relaxometry with total acceleration factors up to 36 for 1.8 mm resolution and up to 54 for 1.25 mm resolution qT_1 imaging. We then investigate highly efficient diffusion MRI with high b-values and long diffusion times. We show super-accelerated multi-shell diffusion acquisitions with 370 whole-brain diffusion volumes over 8 b-value shells up to $b = 7000 \text{ s/mm}^2$ at isotropic resolutions of 1.8 mm and 2.0 mm. We compare MESMERISED and standard MB PGSE for their SNR efficiency and their effectiveness in supporting complex diffusion signal modeling for the case of b-value = 7000 s/mm^2 dMRI at 7 T. Subsequently, we demonstrate time-efficient sampling of different diffusion times with 1.8 mm isotropic diffusion data acquired at four diffusion times up to 290 ms. Finally, we demonstrate that quantitative T_2 and B_1^+ mapping without echo-shifting acceleration can be added to achieve comprehensive multi-contrast mapping with the same MESMERISED sequence.

Theory

The MESMERISED sequence is based on the standard (diffusion-weighted) STEAM sequence (Figure 1A). The STEAM pulse sequence uses three radiofrequency (RF) pulses: excitation (Exc), storing (Sto) and recalling (Rec) pulses, which can create several spin echoes (SEs) and a stimulated echo (STE), which was first recognized by Erwin Hahn (Hahn, 1950) and later introduced to MRI by Frahm and colleagues (Frahm et al., 1985). In general terms, the excitation pulse places the magnetization into the transverse plane where it undergoes T₂* decay. Subsequently, the storing pulse places ('stores') part of the total magnetization along the longitudinal axis where it is subject to T₁ decay during the mixing time (TM). The remaining part of the magnetization forms the primary SE signal, also known as an "eight-ball echo" (Hahn, 1950) or Hahn echo, since it is the lesser known form of a spin echo formed by a 90°-90° pulse pair. Finally, the recalling pulse places ('recalls') the stored magnetization to the transverse plane where it once again undergoes T2* decay until the STE is formed. This sequence of pulses can create three more spin-echo signals (the second, third and fourth spin-echo) after the STE, which are the consequence of the free induction decay (FID) signal generated by the storing and recalling pulses. We focus on the first or primary SE signal in the remainder of this work and refer to it simply as 'the SE' hereafter. Whereas the SE undergoes only T2 decay, the STE signal undergoes both T₁ and T₂ decay, because the signal is obtained from spins stored along the longitudinal axis for the duration of the TM, during which there is T₁ relaxation. Furthermore, both echoes (SE and STE) share the transverse magnetization created by the excitation pulse and therefore split the total excited signal, which becomes evenly distributed between SE and STE when the storing pulse has a flip angle of 90°.

136137

138139

140

141142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

158159

160

161

162

163

164

165

166167

168

169170

171172

173174

175

MESMERISED removes the dead time in STEAM by shifting Exc-Sto pulse pairs and SE readout for one or more MB slice groups into the dead time of earlier slice groups (Figure 1B). We refer to this as echo-shifting, by analogy to sequences that shift sequence elements to achieve echo times longer than their repetition time (TR) in gradient-echo or steady-state free precession sequences (c.f. Ehses et al., 2015; Gibson et al., 2006). However, we note the shifting mechanism here is fundamentally different and does not achieve TE > TR. The echo-shifting in MESMERISED temporally interleaves and multiplexes the contrast evolution of several slices or MB slice-groups. The stored longitudinal magnetization of earlier Exc-Sto and SE readout sequence blocks (SE_i blocks) evolves while later SE_{i+n} blocks are executed and SEs are acquired. Subsequently, STEs are recalled and read-out in a sequence of STE_i blocks (Figure 1B), placed at time TM to their corresponding SE_i blocks. We refer to the number of echo-shifted slice groups as the echo-shift factor (ES factor). The echo-shifting is independent of MB acceleration and allows for very high multiplicative ESxMB acceleration factors, particularly under moderate and long TM's. This results in full duty cycle and high time efficiency, particularly for quantitative T₁ and diffusion imaging.

For instance, T_1 relaxometry with STEAM requires a range of TM's acquired under the same TR and TE. Whereas total acquisition time of T_1 relaxometry with MB-STEAM would be determined by the TR of the *longest* TM, acquisition time of T_1 relaxometry with MESMERISED is limited by the TR of *shortest* possible TM. Here, MESMERISED makes use of its *block reordering property* (Figure 1C): a different choice of ES factor determines a simple reordering of the same sequence blocks and this enables a different mixing time to be acquired at the same minimized TR. Therefore, in MESMERISED all TM's or diffusion times required, for instance to accurately fit an exponential decay model, can be acquired at full acquisition duty cycle and the same short TR.

For diffusion imaging, mono-polar diffusion gradient pairs are inserted symmetrically around the storing and recalling pulses. Although only the first (before the storing pulse) and fourth (after the recalling pulse) gradients are needed for diffusion weighting of the STE signal, there are several advantages to a double time-symmetric placement of four gradients. First, these also serve as crusher gradients at low amplitude and duration in non-diffusion-weighted imaging, helping to remove unwanted echo pathways. Second, the first pair of gradients create a pulsed-gradient spin echo (PGSE) acquisition in the SE_i block, with the corresponding PGSE diffusion contrast in the SE. Third, the second and third gradient act as spoilers during the mixing time for unwanted (secondary and tertiary) spin echo pathways (as shown in a phase graph, see Supplementary Figure 1). This spoiler action is enhanced in MESMERISED, since the gradients of echo-shifted blocks add to the total spoiler-moment during TM. Additionally, 'unbalancing' the third gradient with respect to the fourth by modifying its moment provides a further degree of adjustment to avoid unwanted echo pathways (cf. Supplementary Figure 1). MESMERISED can acquire diffusion-weighted signals at long diffusion times and high b-values in its STE signal, at relatively low duty cycle demands on the gradients. In addition, it allows highly accelerated probing of multiple diffusion times at the same TR and TE using block reordering property (Figure 1C) in the same way as in T₁ relaxometry. Finally, as a STEAM variant with both a SE and STE readout, MESMERISED can acquire both a low b-value and short diffusion time (in its diffusion-weighted SEs) and a high b-value and long diffusion time (in its diffusion-weighted STEs) in a single TR.

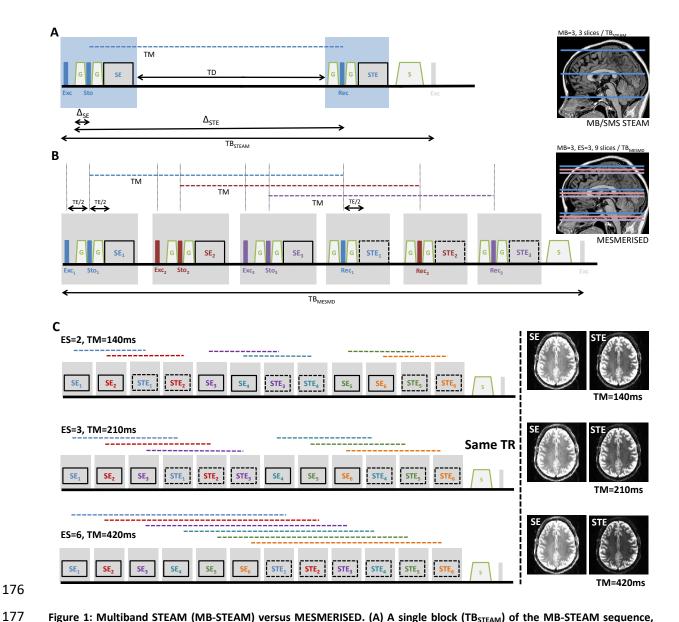


Figure 1: Multiband STEAM (MB-STEAM) versus MESMERISED. (A) A single block (TB_{STEAM}) of the MB-STEAM sequence, consisting of a spin-echo (SE) and stimulated-echo (STE) readout, will acquire MB slices per block, leaving considerable dead time (TD), mostly determined by the desired mixing time (TM). (B) The MESMERISED sequence utilizes this dead time by echo-shifting one or more SE blocks (here ES factor 3), resulting in full duty cycle and high time efficiency. The stored longitudinal magnetization (horizontal dashed lines) of earlier SE blocks, evolves while later STE blocks are executed and SE_i's are acquired. Subsequently, STE_i's are read-out in the STE blocks. MESMERISED will acquire ESxMB slices per TB_{MSMD}, which is slightly longer than TB_{STEAM}. Exc/Sto/Rec: spatially selective Excitation/Storing/Recalling pulse; TE: echo time; G: diffusion/crusher gradient; S: spoiler gradient. (C) The block reordering property: MESMERISED enables a multitude of mixing times at the same TR with full duty cycle. Here SE and STE echoes are acquired for 6 slice groups, each containing a number of slices determined by the employed MB factor. SE and STE readout for the same slice groups, as well as the time evolution of longitudinally stored magnetization during the mixing time (dashed horizontal lines), are encoded by the same color. In MESMERISED each echo-shift factor (ES factor) engenders a simple reordering of the same sequence blocks to enable different mixing times at the same TR. This TR is the same for all allowable TM's and is minimized by employing full acquisition duty cycle. The illustrated examples are for sequence block length of 70 ms, enabling TM = 140 ms for ES factor 2 (top), TM = 210 ms for ES = 3 (middle) and TM = 420 ms for ES = 6 (bottom).

Methods

192

228

229

230

231

232

233

234

235

236

237

238

239240

241

242

243

244

245

246

247

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265266

267

268269

MESMERISED sequence implementation

The MESMERISED sequence was implemented on a Siemens 7 T MAGNETOM platform (Siemens Healthineers Erlangen, Germany). A sequence unit (or building block) was created consisting of a single multiband selective pulse surrounded by two gradient pulses and followed by an echo-planar imaging (EPI) readout. This unit serves both as an STE block and, when including a matched multiband pulse at the outset, as an SE block (Figure 1B). Blipped-CAIPIRINHA simultaneous multislice reconstruction (Setsompop et al., 2012) with the split-slice GRAPPA leak-block approach (Cauley et al., 2014) was performed by using the online reconstruction that is part of the MGH SMS-EPI release for Siemens platforms (https://www.nmr.mgh.harvard.edu/software/c2p/sms). In-plane acceleration was reconstructed with GRAPPA (Griswold et al., 2002) using FLEET autocalibration scans as described in (Blazejewska et al., 2017). The MESMERISED implementation allows for usercontrollable MB, GRAPPA and ES acceleration factors, along with the common parameters (TE, TR, number of slices, and resolution parameters: slice thickness, in-plane resolution, readout bandwidth). For diffusion imaging, duration δ , amplitude |G| and time spacing Δ of the diffusion gradient pairs are user-controllable to determine b-value and diffusion time for the SE (b_{SE} and Δ _{SE}= Δ) and the STE (b_{STE} and Δ _{STE}= Δ +TM). Diffusion gradient direction tables were implemented as multiples of 12 directions (24, 36, 48, 60) distributed around a full sphere by an electrostatic repulsion algorithm (Jones, 2004), interspersed with b₀ volumes every 12 directions. Multi-shell tables were implemented by combining tables and suitable modulation of |G| by scaling the length of the diffusion direction vector. To remove the need for SAR-intensive fat saturation (FatSat) pulses, "fat-unfocusing" as described in (Ivanov et al., 2010) was achieved by the appropriate setting of the pulse lengths and bandwidth-time product (BWTP).

MRI data acquisition

Whole-brain acquisitions were performed on a whole-body research 7 T Siemens MAGNETOM (Siemens Healthcare, Erlangen, Germany) system with gradients delivering 70 mT/m maximum strength at a maximum slew rate of 200 T/m/s per physical axis. For RF transmission and reception, a single-channel-transmit/32-channel-receive head coil (Nova Medical, Wilmington, MA, USA) was used. Written informed consent was obtained from all the subjects before imaging and all procedures were approved by the Ethics Review Committee Psychology and Neuroscience (ERCPN) of the faculty of Psychology and Neuroscience, Maastricht University. MESMERISED was tested on six healthy subjects (4 males and 2 females, average weight 75 \pm +- 5 kg and age 29 \pm +- 3 years). Dielectric pads (Teeuwisse et al., 2012) were used to improve B₁⁺ field homogeneity by positioning two pads at the ears/temples. All sessions started with a localizer scan in three orthogonal directions. Subsequently, a B₀ field map was acquired (Cusack and Papadakis, 2002) and used to optimized B₀ shim currents. An absolute B₁⁺ map (using a pre-saturation turbo-flash (PreSat-TFL) sequence; Chung et al., 2010) was then acquired and used to calibrate the global transmit flip angle.

Subsequently, MESMERISED acquisitions were performed with full brain coverage, with slices in a transverse orientation. All diffusion and crusher gradients were run in full symmetric format, i.e. unbalancing of the third gradient was not needed to avoid echo pathway contamination in the experiments reported here. EPI readout was executed along the anterior-posterior (AP) phase-encoded direction, and undersampled with GRAPPA acceleration factor 2 or 3 and using a partial Fourier factor of 6/8. In-plane reference (ACS) lines for GRAPPA (24 lines for factor 2, 36 lines for

270 factor 3) were acquired using FLEET at a flip angle 12° with 2 to 5 dummy pre-scans. For blipped CAIPI

MB (Breuer et al., 2006; Setsompop et al., 2012), a CAIPI-shift (CS) factor was employed (MB: 3, CS: 2;

and MB: 4, CS: 3). Every acquisition was accompanied by 5 volumes with a reversed-phase encoding

direction (posterior-anterior) only for the contrast with highest signal (i.e. shortest TM or TE, flip

angles 90°-90°, and/or b₀ acquisition), used for correction of off-resonance distortions. For all the

acquisitions, the slices were acquired in an interleaved ordering without slice gap.

MESMERISED protocols were created for imaging of quantitative T_1 (qT_1), quantitative T_2 (qT_2), transmit field (B_1^+) mapping and diffusion. In MESMERISED the number of slices must be an integer multiple of ESxMB for all employed ES and MB factors. For qT_2 , B_1^+ and diffusion protocols with a single mixing time or diffusion time this provides a single constraint. For qT_1 or diffusion protocols with multiple TM or diffusion times (Δ), this provides a set of constraints, one for each ES/TM combination. These constraints are easily solved by choosing slice numbers N with a large number of integer divisors (highly composite numbers), such as 60, 72, 90 and 108, which can be used for isotropic imaging resolutions of 2 mm, 1.8 mm, 1.5 mm, and 1.25 mm, respectively. Each integer divisor of slice number divided by MB factor (i.e. N/MB), then determines an allowable ES factor under the same TR (because it determines a single temporal reordering of sequence blocks, see Figure 1C). For instance, at 72 slices with MB = 3, N/MB = 24 the allowable ES factors are [1, 2, 3, 4, 6, 8, 12, 24]. At full duty cycle, each such ES is associated with a single minimum TM, determined by the length of RF pulses, gradient pulses, and EPI train length. Again due to MESMERISED's temporal block reordering property, the minimum TM for each allowable ES is (very close to) ES*TM_{ES=1}, i.e. an

- A proof of principle test was performed to demonstrate that MESMERISED's echo-shifting does not affect the resulting SE and STE images, and therefore the same signal can be acquired in shorter total acquisition time. Sets of MESMERISED SE and STE images were acquired at 1.8 mm isotropic resolution at different ES but fixed TM = 300 ms, TR = 11.88 s and TE = 47.5 ms. ESxMB combinations of 1x3, 2x3 and 4x3 were acquired with transverse slices, GRAPPA 2, FoV 230 x 230 mm² and partial Fourier 6/8. The achievable (but unused) minimum TR for ESxMB= 2x3 and 4x3 were recorded to quantify the acceleration potential of MESMERISED.
- qT_1 acquisition

integer multiple of the minimum TM for ES = 1.

- Quantitative T₁ imaging was performed at 1.8 mm isotropic and 1.25 mm isotropic, by performing relaxometry over a range of allowable ES/TM combinations at the same TR and TE. Five repetitions of each TM were acquired to assess retrospectively what level of data averaging (1, 3 or 5 repetitions) supports robust exponential T₁-decay model fitting. Additionally, T₁ relaxometry was performed both at the minimum possible TR, at full duty cycle and acceleration, and with a doubling of TR and all TM's. This was done to assess the range of TM's necessary for robust exponential T₁-decay model fitting. Full acquisition parameters are given in Supplementary Table 1.
 - Diffusion acquisition
- Multi-shell diffusion imaging was performed at 1.8 mm and 2.0 mm isotropic for different maximum b-values. Diffusion acquisitions were designed to explore the strengths of MESMERISED in achieving a high number of volumes per unit time, and either high b-values or a large range of Δ 's. A set of multi-shell diffusion acquisitions with maximum b-value 7000 s/mm², 185 directions over 4 shells (a total of 370 SE and STE volumes), were performed at 1.8 mm isotropic and different ES/TM, realizing

different diffusion times between 80 ms and 430 ms in the STE. A 4-shell acquisition at 2 mm

isotropic with maximum b-value 7000 s/mm² was performed to explore the effect of a very short TR (2.36 sec) and a very high number of volumes per unit time.

MESMERISED and standard MB PGSE were compared for their signal-to-noise efficiency in b-value = 7000 s/mm² dMRI at 7 T and their effectiveness in supporting complex diffusion signal modeling. In order to achieve an objective comparison under the same RF pulse, EPI readout and image reconstruction implementations, the same MESMERISED-capable sequence was run in PGSE mode by omitting the STE block and modifying the storing pulse flip angle (180°) and phase. Whole brain (72 slices) 1.8 mm isotropic b = 7000 s/mm² diffusion-weighted MESMERISED with 4 echo-shift factors (ESxMB: 2x3, 3x3, 4x3 and 6x3; TE: 49.9, 46.2, 43.8, 40.9 ms; TM: 140, 210, 280 and 420 ms; SE b-val: 752, 483, 350, 223 s/mm², respectively) and PGSE (MB:2; TR: 4.21 s.; TE: 97.8 ms) were optimized to run just within SAR and gradient duty cycle limits. For each, 60 directions at b = 7000 s/mm² with 6 b₀s (for diffusion microstructure modelling) as well as 27 matched b₀s (for temporal SNR and SNR efficiency calculation) were acquired in one healthy subject (male, 79 kg). MESMERISED at MB = 3 and PGSE at MB = 2 were selected, since under basic assumptions (SAR grows linearly with MB factor and quadratic with flip angle) these settings would have a similar SAR load (within 10% at the same TR with MESMERISED-MB3 10% more efficient than PGSE-MB2). Pulse lengths (4.48 ms for excitation, 8.96 ms for refocusing, storing and recalling) were matched between MESMERISED and PGSE and chosen to satisfy "fat-unfocusing" constraints as described in (Ivanov et al., 2010), as well as balancing a reasonable length (compared to white matter T2*) with SAR constraints. A potential TE shortening advantage for MESMERISED by instead running the excitation pulse as the long pulse and storing and recalling as the short pulse was avoided to keep the RF-pulse regime as similar as possible. Flip angle calibration was performed as previously described for 7 T diffusion acquisition (Vu et al., 2015), which lead to a reference voltage of 235 V for the 79 kg subject and ~15-20% overflipping in the center of the brain. SAR constraints alone forced PGSE to run at a TR of close to 4 s, far higher than the full acquisition duty cycle TR of ~2.5 s. Moreover, gradient duty cycle constraints forced further TR compromises for PGSE as established by running the protocol with the evenly sphere-sampled 60 direction table at the desired b = 7000 s/mm² while adjusting diffusion gradient amplitude and duration (and by extension TE and TR) until the full protocol ran without gradient power amplifier overloads. Gradient limits forced b = 7000 s/mm² PGSE to run at |G|: 49 mT/m, lengthening TR and TE (to 4.21 s. and 97.8 ms, respectively), whereas for MESMERISED this allowed full diffusion gradient amplitude (|G|: 70 mT/m) and full acquisition duty cycle TR (3.4 s). EPI readout (GRAPPA-2 with FLEET ACS, FoV 218 x 218 mm² and PF 6/8) was matched for echo-spacing (0.57 ms) and for readout bandwidth within 5% (MESMERISED: 2164 Hz/pix; PGSE: 2066 Hz/pix). Forcing a full EPI bandwidth match would increase PGSE TE and TR further through a peripheral nerve stimulation limit that did not occur in MESMERISED at their respective TRs. All four MESMERISED protocols and the PGSE protocol ran between 90% and 100% of the 6-minute SAR limit.

- Finally, to assess efficient exploration of diffusion-time dependent effects on mean diffusivity and mean kurtosis with MESMERISED, an acquisition at 1.8 mm isotropic at four different diffusion times and two different b-values (1000 and 2500 s/mm²) was performed. Full acquisition parameters for all diffusion protocols are given in Supplementary Table 3.
- 353 *Quantitative multi-contrast acquisition*

313

314

315

316

317

318

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

340

341342

343

344

345

346347

- Quantitative multi-contrast mapping was performed by adding quantitative T_2 and B_1^+ mapping to
- multi-shell diffusion and quantitative T₁ mapping for one subject, all at the same 1.8 mm resolution.
- 356 Quantitative T₂ and B₁⁺ mapping were performed at 1.8 mm by relaxometry on a range of TE's (from

28.3 to 88.3 ms in steps of 20.0 ms), and by cosine model fitting of Exc-Sto-Rec = α° - $2\alpha^{\circ}$ - α° for a range of multipliers α (from 30° to 110° in steps of 20°; Lutti et al., 2012), respectively. Although neither qT_2 nor B_1^+ mapping benefits in terms of acquisition speed from MESMERISED's echo-shifting or block reordering mechanisms, T_2 relaxometry was performed at both ES = 1 and at ES = 3, to assess whether robust qT_2 mapping can be performed under echo-shifting. Full acquisition parameters for qT_2 and B_1^+ mapping are given in Supplementary Table 2.

Image processing and analysis

363

364

365

366367

368

369

370

371

372

373

374375

376

377

378

379380

381

392

393

394

Each MESMERISED dataset (all repetitions for all TM's (qT_1) , TE's (qT_2) , flip angles (B_1^+) , or directions and b-values for diffusion) was individually denoised, separately for the SE volumes and STE volumes, except for the proof-of-principle signal comparison acquisitions (reported in Figure 2) and the MESMERISED – PGSE high-b diffusion comparison acquisitions (reported in Figure 9 and 10, as well as Supplementary Figures 2, 3 and 4). Denoising was performed before any other step following the approach by Veraart et al. (2016), using the mrdenoise tool in MRtrix 3.0 (Tournier et al., 2019) with a kernel size of 5 or 7. Subsequently, all data were corrected for movement, off-resonance and eddycurrent artifacts using the Topup and Eddy tools of FSL v6.0.1 (Jenkinson et al., 2012). Given that the TE, EPI readout-train, and crusher/diffusion gradients are exactly matched between SE and STE within each acquisition, B₀ field related distortion estimation can be performed on either SE or STE. Therefore, topup was performed on the highest signal SE volumes (i.e. shortest TE, flip angles 90° -180° - 90°, and/or b₀ acquisition). Since (particularly long time constant) eddy currents can differ between SE and STE, FSL Eddy was then performed separately on SE and STE volumes with this Topup input. Since data was acquired with shortest-TM volumes first, the first acquired SE and STE volumes are already in close spatial alignment (having been acquired ~ 100 - 300 ms apart in time), which ensures good alignment of all SE and STE volumes. Brain masks for each subject and each MR contrast, needed for the preprocessing and analysis steps, were obtained using the BET tool of FSL v6.0.1. After correction, the following analyses were performed.

382 qT_1 analysis

For qT₁ analysis, two distinct relaxometry approaches were used which fit volumes at varying TM with the same TR and TE. In the first approach, only the STE volumes are used. The STE signal as a function of TR, TM, TE and flip angles can be written:

386
$$S_{STE}(TR, TM, TE, \alpha_i) = \frac{1}{2}S_0\left(1 - \exp\left(-\frac{TR - TM}{T_1}\right)\right)\sin(\alpha_1)\sin(\alpha_2)\sin(\alpha_3)\exp\left(-\frac{TM}{T_1}\right)\exp\left(-\frac{TE}{T_2}\right)$$
 (1)

387 Where S_0 contains all parameters not varying with TR, TM, TE and flip angles, such as proton density 388 and Tx/Rx spatial sensitivity profiles, and α_1 , α_2 , and α_3 are the flip angles (in radians) of excitation, 389 storing, and recalling pulse, respectively. Absorbing all terms that do not vary with TR or TM into the 390 S_0 term, we get:

391
$$S_{STE}(S_0, TM, TR, T_1) = S_0(\alpha_i, TE, T_2) \left(1 - \exp\left(-\frac{TR - TM}{T_1}\right)\right) \exp\left(-\frac{TM}{T_1}\right)$$
 (2)

In the first approach, equation 2 was fitted to STE volumes at varying TM and maps for S_0 and T_1 were obtained. In this approach, the S_0 absorbs effects of flip angle and T_2 decay (as well as RF receive coil sensitivities and proton density), and the fitted curve is a product of exponentials.

- In the second approach, both SE and STE volumes are used. The SE signal and STE/SE signal ratio as a
- function of TR, TM, TE and flip angles can be written:

397
$$S_{SE}(TR, TM, TE, \alpha_i) = S_0 \left(1 - \exp\left(-\frac{TR - TM}{T_1} \right) \right) \sin(\alpha_1) \sin^2\left(\frac{\alpha_2}{2} \right) \exp\left(-\frac{TE}{T_2} \right)$$
(3)

398
$$\frac{S_{STE}}{S_{SE}} = \frac{\sin(\alpha_2)\sin(\alpha_3)}{1-\cos(\alpha_2)} \exp\left(-\frac{TM}{T_1}\right)$$
 (4)

Absorbing all terms that do not vary with TR or TM for the ratio into an S_0^* term, we get:

$$400 \qquad \frac{S_{STE}}{S_{SE}} = S_0^*(\alpha_i) \exp\left(-\frac{TM}{T_1}\right) \tag{5}$$

- In the second approach equation 5 was fitted to STE/SE ratio volumes at varying TM and maps for S_0^*
- and T_1 were obtained. In this approach, the S_0^* absorbs effects of flip angle, and the fitted curve is a
- simple exponential. Therefore, the second approach uses the available SE images in MESMERISED to
- 404 establish a potentially lower complexity fit (simple exponential vs. product of exponentials in the first
- approach). Approach 1 and 2 were both evaluated for the estimation of qT₁ maps and compared for
- 406 the precision of their estimates by evaluating Fisher Information Matrix (FIM) local uncertainty
- 407 (Harms et al., 2019). In addition, the uncertainty of qT₁ estimates was evaluated for an increasing
- 408 number (1, 3, 5) of repetitions per TM volume.
- 409 Diffusion analysis
- The diffusion-weighting in MESMERISED is the same as for the underlying diffusion-weighted STEAM
- 411 sequence: whereas the SE signal is diffusion-weighted by gradients before and after the storing
- pulse, separated by the diffusion time Δ_{SE} (S_{dSE}), the STE signal is weighted by the diffusion gradient
- 413 before the storing pulse and the gradient after the recalling pulse, separated by the diffusion time
- 414 Δ_{STE} (S_{dSTE}; see Figure 1). The latter diffusion time is equal to Δ_{SE} + TM. The corresponding diffusion-
- 415 weighted signal equations for both echoes are defined as:

416
$$S_{dSE}(TR, TM, TE, \alpha_i, b) = S_{SE}(TR, TM, TE, \alpha_i) \exp\left(-(\gamma G\delta)^2 \left(\Delta_{SE} - \frac{\delta}{3}\right)D\right)$$
 (7)

417
$$S_{dSTE}(TR, TM, TE, T_1, \alpha_i, b) = S_{STE}(TR, TM, TE, T_1, \alpha_i) \exp\left(-(\gamma G\delta)^2 \left(\Delta_{SE} + TM - \frac{\delta}{3}\right)D\right)$$
(8)

- Where, S_{SE} and S_{STE} are the SE and STE signals defined in equations 1 and 3, respectively, y is the
- gyromagnetic ratio (in Hz/mT), δ is the diffusion gradient duration (in s), G is the diffusion gradient
- amplitude (equal for the gradients-pair in the storing pulse and the gradient after the recalling pulse,
- 421 in mT/m) and D is the (scalar) diffusivity (in m²/s). For diffusion MRI modeling the exponential
- 422 diffusion terms in eq 7 and 8 were replaced with the desired diffusion model, with the corresponding
- b value and diffusion times. For the 4-shell (max b = 7000 s/mm²) single-TM acquisitions, the
- diffusion data from the STE signal was analyzed by DTI on the 1st shell, Diffusion Tensor Kurtosis
- 425 (Jensen et al., 2005) on the 1st and 2nd shell, Ball&Stick models (Behrens et al., 2007; Behrens et al.,
- 426 2003) with one (B&S_{r1}), two (B&S_{r2}) and three (B&S_{r3}) sticks on all shells, NODDI model (Zhang et al.,
- 427 2012) on the 1st and 2nd shell, CHARMED model (Assaf and Basser, 2005) with one (CHARMED_{r1}) and
- 428 two (CHARMED_{r2}) cylinder compartments on all shells. For the SE volumes from the same

acquisitions, the highest b-value shell was analyzed with DTI. For the 2-shell multi-TM acquisition, both DTI (1st shell) and DKI (1st and 2nd shell) analysis were applied to the STE signal.

For the MESMERISED-PGSE b = 7000 s/mm² diffusion imaging comparison the SE and STE volumes of each MESMERISED run (ES = 2, 3, 4, and 6) and the PGSE volumes were each separately preprocessed for distortions and movements as described above, without denoising. Subsequently, the Ball&Stick model with two sticks (B&S₁₂) was fitted to the b = 7000 s/mm² MESMERISED STE and PGSE volumes (60 directions + 6 b₀s) and the Diffusion Tensor model was fitted to the MESMERISED SE volumes, evaluating voxel-wise uncertainty as described below. For consistency and completeness, the Tensor model was fitted to all SE volume series, despite the fact that the b-values (752, 483, 350, and 223 s/mm² for ES factor 2, 3, 4 and 6, respectively) of the higher ES factors is likely too low for robust fitting of the Tensor model. The fraction-of-sticks (FS) map and fractional anisotropy (FA) maps, and their uncertainty, were used for assessment of the fit quality of the B&S_{r2} and Tensor models, respectively. Voxel wise temporal signal-to-noise ratio (tSNR) maps were calculated from the b₀ only acquisitions, discarding the first 2 and last 5 of 27 volumes, computing tSNR = mean/std over the remaining 20 volumes, and lightly smoothing the resulting map with a Gaussian smoothing kernel with standard deviation of 0.5 voxel. Voxel wise tSNR efficiency (tSNR-eff) was computed as tSNR/sqrt(T_{vol-acq}) and tSNR per unit time as tSNR/T_{vol-acq}, where T_{vol-acq} is the single volume acquisition time, which is TR/2 for STE and SE in MESMERISED and equal to TR for PGSE. White matter (WM) summary statistics of tSNR-eff, tSNR per unit time and diffusion modeling uncertainty were computed over the same deep WM mask for all acquisitions, obtained from a thresholded FS map processed with a 3D image erosion and median filtering step to avoid effects of tissue boundaries or volume misalignment. The mean and standard deviation over the WM mask was computed for tSNReff, tSNR per unit time and diffusion modeling uncertainty, excluding voxel with FS or FA values lower than 0.15, since uncertainty estimation can be less reliable close to hard parameter value limits (Harms et al., 2019).

454 B_1^+ and qT_2 analysis

429

430

431

432

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

451

452 453

464

For B_1^+ mapping, the approach used by (Lutti et al., 2012) was used on the acquired data at multiple flip angles. It consisted in taking the ratio between STE and SE signals, where the flip angle ratio between the three STEAM pulses is α° - $2\alpha^\circ$ - α° (e.g. for $\alpha = 90^\circ$: 90° - 180° - 90° degrees for excitation, storing and recalling pulses respectively). The STE/SE ratio is proportional to the cosine of the excitation flip angle:

$$460 \qquad \frac{S_{STE}}{S_{SE}}(\alpha) = S_0 \frac{\sin(2\alpha)\sin(\alpha)}{\sin(\alpha)^2} \exp\left(-\frac{TM_0}{T_1}\right) \to S_0^*(TM_0, b_0)\cos(\alpha) \tag{6}$$

- Therefore, the relative B_1^+ map can be estimated from a sufficient number of ratio volumes with varying α . For qT₂ mapping only the SE signal was used for both acquisitions (ES = 1 and 3) separately,
- 463 fitting a simple exponential decay of TE's to estimate a qT_2 map.

Model optimization and uncertainty

All modeling analyses and parameter uncertainty quantifications above were performed in the Microstructure Diffusion Toolbox (MDT; https://mdt-toolbox.readthedocs.io/en/latest_release/) with Powell optimizer at patience 10, and using a cascaded optimization approach (Harms et al., 2017). For diffusion modeling, the cascade consisted of: S₀ estimation – B&S_{r1} estimation – desired

model; for qT_1 , qT_2 and B_1^+ analysis, the cascade consisted of: S_0 estimation – desired model. The precision or uncertainty of estimates was evaluated by computing standard deviations derived from the Fisher Information Matrix (FIM), tightly related to the Cramer Rao Lower Bound (CRLB). Whereas the CRLB amounts to the FIM evaluated around a ground truth value, the FIM can be evaluated around a maximum likelihood point estimate and this allows for estimating the standard deviation, or uncertainty, of the point estimate, both for linear (Andersson, 2008) and non-linear models (Alexander, 2008; Harms et al., 2019). FIM uncertainty was calculated as described in Harms et al. (2019) using numerical evaluation of the FIM, and analytic uncertainty propagation for the derived FA parameter for the Tensor diffusion model, as implemented in MDT.

Results

Figure 2 shows a comparison between images obtained using MESMERISED without ES (ESxMB = 1x3, equivalent to MB-STEAM, cf. Figure 1A), ESxMB = 2x3 and 4x3 for the SE and STE images. Qualitatively, Figure 2 shows that images retain the same contrast and characteristics when dead time is filled up through echo-shifting. At the same time, the minimum TR (minTR) illustrates that MESMERISED allows these images to be acquired at higher acceleration in a shorter time, going from a 11.88 s minTR with ESxMB = 1x3, to 4.83 s minTR with ESxMB = 4x3. Here we should note that, if the shorter TR is actually used, the contrast and SNR might change compared to the long TR images shown here due to magnetization and steady-state effects. The minTR also illustrates that although the additional acceleration compared to multiband STEAM is significant (e.g. 2.46x for ESxMB = 4x3 compared to ESxMB = 1x3), and multiplicative with MB factor (e.g. 7.38x for ESxMB = 4x3 compared to ESxMB = 1x1), the temporal acceleration factor is generally not linear with ES, due to the time added by the STE_i (i> 1) blocks (c.f. Figure 1B). The bottom row of Figure 2 shows a more direct comparison in signal line plots. MESMERISED accelerated signals (2x3 and 4x3) follow the MB STEAM signal (1x3) closely.

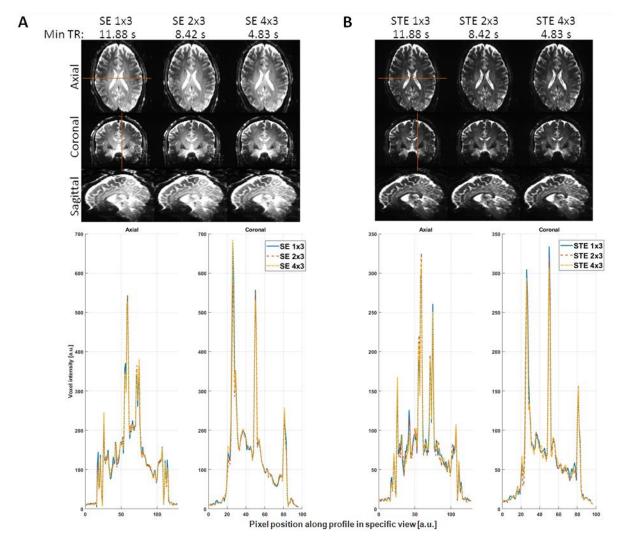


Figure 2: Comparison of MESMERISED images at 1.8 mm isotropic, acquired with MB = 3 at different echo-shift factors of ES = 1, ES = 2 and ES = 4 but otherwise identical parameters (TR = 11.88 s; TE = 47.5 ms; TM = 300 ms) for spin-echo (A, top) and stimulated echo (B, top) The minimal achievable TR (min TR) is indicated to illustrate increased acceleration

capability with ES > 1. Bottom row: corresponding signal line plots for SE (A) and STE (B) for the three echo-shifts for the red line in the axial view (left line plot) and coronal view (right line plot).

qT₁ mapping

499

500

501

502

503

504

505

506507

508509

510511

512

513

514

515516

517518

519520

521

Figure 3A shows MESMERISED T₁ relaxometry data at 1.8 mm isotropic. The relaxometry on TM is performed at constant TE (28 ms) and TR (6.9 s) by increasing the ES factor proportionally to TM, for TM's from 140 ms to 1680 ms (see Supplementary Table 1). This results in total acceleration factors (ESxMB) from 3 to 36, highlighting the super-acceleration capability of MESMERISED for T₁ relaxometry. For MB-STEAM and STEAM (without MB) the TR for relaxometry would be limited by the largest TM and would equal 41.51 and 124.60 s, respectively, which means that relative temporal speed-ups of 6.02x and 18.06x are achieved compared to MB-STEAM and STEAM, respectively. Figure 3B shows the whole-brain isotropic resolution S₀ and qT₁ maps resulting from analysis of STE data (i.e. using Equation 2). The S_0 map estimated in this way retains strong T_2 weighting (along with weighting for proton density and Tx/Rx fields), which is apparent in the contrast between white matter (WM) and gray matter (GM) structures. The qT₁ map also shows a high contrast level due to the quantitative differences in T₁ between WM and GM structures. For comparison, Figure 4A shows $S_0^{\,*}$ and qT_1 maps resulting from analysis of the STE/SE data from the same acquisition (i.e. using equation 5). The resulting qT₁ map is qualitatively highly similar. However, the theoretical expectation (described above) that the S_0^* map is more weighted for the transmit field (i.e. local flip angle) and less for T2 compared to the S0 map is borne out in a decreased WM/GM contrast and an increased central darkening strongly (anti-)correlated with the transmit field (described below).

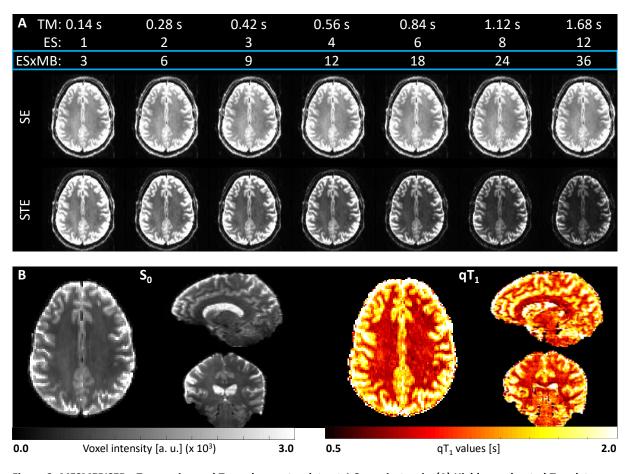


Figure 2: MESMERISED qT_1 mapping and T_1w relaxometry data at 1.8 mm isotropic. (A) Highly accelerated T_1w data was acquired by increasing ES factor in tandem with TM to keep the TR constant and short TM, ES and total acceleration

factor EsxMB are given. The acquired SE and STE signals are shown with the STE signal visibly attenuating with TM. (B) The corresponding S_0 (left) and qT_1 (right) maps estimated from the STE signal only (Equation 2).

522

523

524

525

526

527528

529

530 531

532

533

534535

536

537

538539

540

541

542

543544

545

546

547

548

549

550

551552

553

554

Figure 4B shows MESMERISED S_0^* and qT_1 maps at 1.25 mm isotropic, from data at constant TE (31.7 ms) and TR (8.8 s), resulting in total acceleration factors (ESxMB) from 3 to 54 over 8 different TM's (Supplementary Table 1). For MB-STEAM and STEAM (without MB) the TR for relaxometry would be limited by the largest TM and would equal 79.79 and 239.40 s, respectively, which means that relative temporal speed-ups of 9.07x and 27.20x are achieved compared to MB-STEAM and STEAM, respectively.

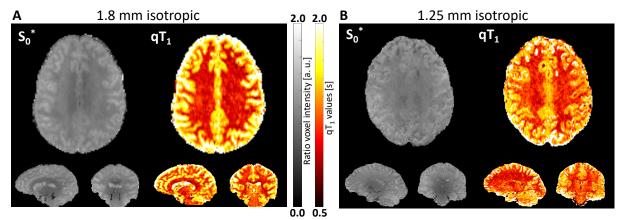


Figure 4: MESMERISED qT_1 mapping at 1.8 mm (A) and 1.25 mm (B) isotropic using the STE/SE ratio T_1 -relaxometry model (Equation 5) on data acquired using 3 repetitions (TA: 2:07 and 4:01 min, respectively).

A more detailed comparison of the uncertainty of qT₁ maps for the more SNR challenged 1.25 mm isotropic data is shown in Figure 5. This compares a full duty cycle acquisition (cf. Figure 1C) with a maximum TM of 1080 ms (short TR, TM; top) with a half duty cycle acquisition were TR and all TMs are doubled (long TR, TM; bottom; maximum TM of 2160 ms). This shows that uncertainty in qT₁ estimates is considerably lower for all long TR,TM acquisitions compared to short TR,TM acquisitions. Furthermore, an upward bias in qT₁ estimates is visible for full duty cycle short TR,TM acquisitions. This is easily explainable by the TM range, implied by the block reordering property under full duty cycle (i.e. short TR,TM), that is insufficient (from 70ms to 1080ms) for the expected T1's at 7 T (1 - 1.5 s). When TR and all TMs are doubled, and hence acquisition duty cycle is decreased to 50%, this range (140ms to 2160ms) is much more suitable and results in good estimates and uncertainty. Therefore, focusing on the bottom row, it is observed that uncertainty in qT_1 estimates is considerably lower 1) with an increasing number of repetitions and 2) with STE/SE ratio analysis compared to STE-only analysis. Although 1) is expected given the averaging effect of repetitions, the benefit of STE/SE ratio analysis is more interesting, especially in comparison with the effect of repetitions. With the STE/SE ratio analysis the local maxima in uncertainty for the long TR, TM data is at a similar level with a single repetition (acquisition time 1:40) as for the STE-only analysis with 5 repetitions (acquisition time 6:22); the STE/SE ratio analysis with three repetitions (acquisition time 4:01) has lower uncertainty everywhere (Figure 5, bottom row). Additionally, the uncertainty for STE/SE ratio analysis is more homogenous over the brain. Therefore, the 1.25 mm isotropic wholebrain qT₁ map in Figure 4 can be achieved in 2-4 minutes, and the 1.8 mm isotropic qT₁ map in 1-2 minutes, depending on the desired precision level and STE/SE ratio analysis can provide higher precision, comparable to that achieved by averaging over multiple repetitions.

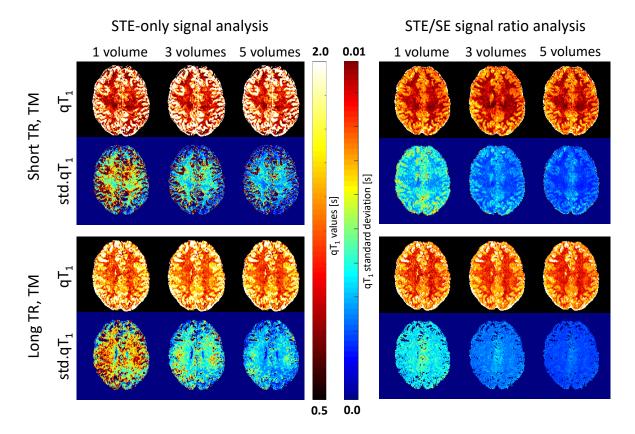


Figure 5. MESMERISED 1.25 mm isotropic qT_1 mapping and its uncertainty compared between: short vs. long TR and TM (top vs. bottom), STE-only vs STE/SE ratio analysis (left vs. right) and number of repetitions (1, 3 and 5). Short TR is 4.4 s with a maximum TM of 1080 ms, in which case 1/3/5 repetitions have an acquisition time of 1:05 / 2:16 / 3:26 respectively; long TR is 8.8 s with a maximum TM of 2160 ms, in which case 1/3/5 repetitions have an acquisition time of 1:40 / 4:01 / 6:22 respectively. STE-only qT1 analysis is performed with Equation 2, whereas STE/SE ratio analysis is performed with Equation 5. Uncertainty or precision of the point-estimates is mapped as standard deviations derived from the Fisher Information Matrix.

Diffusion MRI

Figure 6 illustrates the two-dimensional parameter space of diffusion times (vertical) and b-value (horizontal) with MESMERISED diffusion data at 1.8 mm isotropic (where the further 2 dimensions of diffusion direction on the sphere are left implicit). Here, different diffusion times were realized by varying TM and utilizing the maximum echo-shift factor allowed at each TM, individually minimizing TE and TR. Both TR's are close to 3.4 s (see Supplementary Table 3), showing a capacity to explore high b-values, up to 7000 s/mm², and different diffusion times, at high acceleration and data rate. As before, the relative temporal speedup achieved with MESMERISED will increase with ES factor and here speedups are achieved compared to MB-STEAM and STEAM of 1.47x and 4.40x for ES = 2, and 1.95x and 5.85x for ES = 3, respectively. In the following, we first focus on the possibilities afforded by multi-shell acquisitions at a single diffusion time and then demonstrate the possibilities of acquisitions at varying diffusion times.

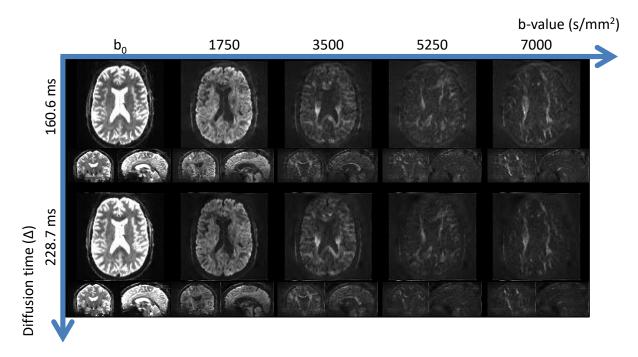


Figure 6: MESMERISED diffusion data at 1.8 mm isotropic spanning the space of diffusion time (vertical) and b-value (horizontal). Whole-brain STE images are shown for two different diffusion times, each corresponding to a distinct ES factor (2 for 160.6 ms and 3 for 228.7 ms) and individually optimized TE and TR, with different b-values from the same multi-shell acquisition. Simultaneously acquired SE images are not shown.

Multi-shell diffusion MRI

577578

579

580

581

582

583

584 585

586

587

588 589

590

591

592593

594

595

596

597 598

599

Figure 7 shows example slices out of a 4-shell MESMERISED acquisition and analysis results for the simultaneously acquired STE images (at a diffusion time of 160.6 ms and b-values of 1750, 3500, 5250, and 7000 s/mm²) and SE images (at a diffusion time of 20.6 ms and b-values of 188, 376, 564, and 752 s/mm²). This makes explicit how the acquisition of both diffusion-weighted SE volumes and STE volumes provides 8 useful shells of diffusion data in a 4-shell acquisition, even when analyzed separately. Whereas the low-b SE volumes support analysis with Gaussian diffusion models such as the DTI model, the high-b STE volumes are suitable for non-Gaussian diffusion models, such as the DKI, NODDI and CHARMED models. Here, the Tensor model was applied in both the lowest b-value STE shell (Figure 7A) and the highest b-value SE shell (Figure 7B), showing some qualitative differences, likely related to the large difference in b-value (752 vs. 1750 s/mm²) at the limits of applicability of the Tensor model. This data was acquired at ES = 2 at a TR of 3.45 s and a TE of 49.8 ms, which tunes the diffusion time of the STE volumes to the shorter end of the achievable range and the b-values of the SE volumes to the higher end, which is accompanied with a higher TE than would be the case for higher echo-shift factors. When a longer diffusion time and higher echo-shift factor is chosen for the same STE volume maximum b-value (e.g. 7000 s/mm² here), diffusion gradient duration δ naturally decreases, along with the SE maximum b-value (see Supplementary Table 4).

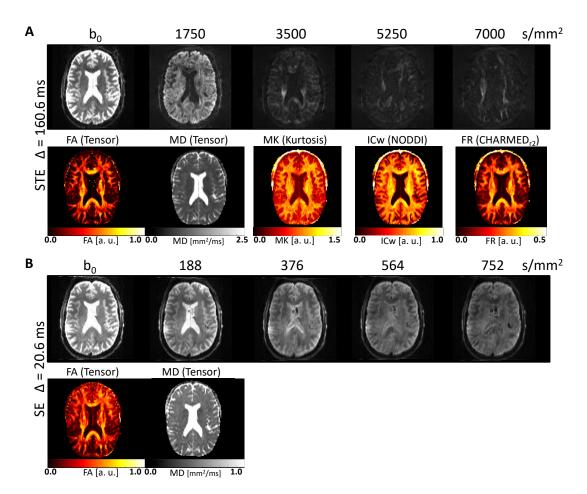


Figure 7: MESMERISED data and diffusion modeling results for simultaneously acquired STE volumes (at high b-values and diffusion time 160.6 ms) and SE volumes (at low b-values and diffusion time 20.6 ms) at 1.8 mm isotropic (ES = 2, TR = 3.45 s, TA = 11:08). (A) Example transverse slices through STE volumes at each b-value in the 4-shell (168 direction) acquisition (top) and their analysis with Tensor, Kurtosis, NODDI and CHARMED models (applied to suitable combinations of shells, bottom). (B) Example transverse slices through SE volumes at each b-value in the 4-shell (168 direction) acquisition (top) and their analysis with the Tensor model (bottom).

 Figure 8 shows two 4-shell (max-b = 7000 s/mm^2) acquisitions which realize full duty cycle at similar mixing and diffusion time at 2 mm isotropic (ES = 5, TM = 290 ms; top) and 1.8 mm (ES = 4, TM = 280 ms; bottom). The relative temporal speedup achieved compared to MB-STEAM is 2.95x for the 2 mm dataset and 3.43x for the 1.8 mm dataset. The 2 mm acquisition achieves a shorter TR (2.36 s vs. 3.37 s) and therefore enables a higher data rate of \sim 1.2 s per volume (for SE and STE together). This generates the full 8-shell 370 volume dataset in an acquisition time of 7 min and 47 s (vs. 10 min and 53 s for 1.8 mm), showing that very short acquisition times are possible for extensive multi-shell data, particularly at resolutions of 2 mm and above, at which high b-value diffusion acquisitions are mostly performed.

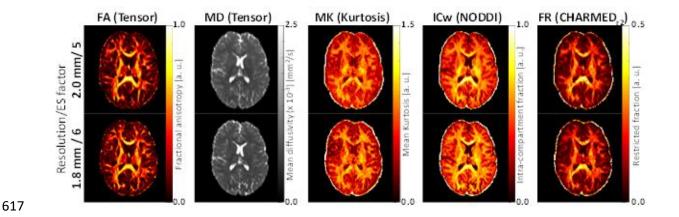


Figure 8: MESMERISED diffusion modeling results for 185 direction multi-shell MESMERISED diffusion acquisitions at different resolutions. Analysis results for Tensor, Kurtosis, NODDI and CHARMED models (applied to suitable combinations of shells) for 2.0mm isotropic (ES = 5, TR = 2.36 s, TA = 7:47) and 1.8 mm isotropic (ES = 6, TR = 3.37 s, TA = 10:53) in the same subject.

		Acq. time/vol	b0 tSNR	b0 tSNR eff	b0 tSNR/sec	FS/FA uncert
		(s.)	mean (std)	mean (std) (s ^{-1/2})	mean (std) (s ⁻¹)	mean (std)
MSMD	STE	1.705	13.41 (3.75)	10.28 (2.89)	7.89 (2.21)	0.058 (0.042)
MB3-ES2	SE	1.705	15.05 (4.29)	11.54 (3.29)	8.85 (2.52)	0.069 (0.032)
MSMD	STE	1.685	13.89 (4.01)	10.70 (3.09)	8.24 (2.38)	0.057 (0.043)
MB3-ES3	SE	1.685	17.82 (5.10)	13.73 (3.93)	10.58 (3.03)	0.073 (0.034)
MSMD	STE	1.680	12.97 (3.69)	10.01 (2.85)	7.72 (2.20)	0.060 (0.079)
MB3-ES4	SE	1.680	17.69 (5.12)	13.65 (3.95)	10.53 (3.05)	0.084 (0.038)
MSMD	STE	1.675	12.66 (3.73)	9.78 (2.89)	7.56 (2.23)	0.065 (0.044)
MB3-ES6	SE	1.675	19.91 (5.40)	15.38 (4.17)	11.88 (3.23)	0.110 (0.051)
PGSE MB2		4.210	12.81 (4.04)	6.24 (1.97)	3.04 (0.96)	0.118 (3.741)

Table 1. Quantitative results for the comparison of MESMERISED with PGSE for b = 7000 s/mm2 diffusion imaging. Results for MESMERISED with MB = 3 and ES = 2, 3, 4, and 6, for each of the STE (b = 7000s/mm2) and corresponding SE volumes and for PGSE (b = 7000s/mm2) with MB = 2. Acq. time/vol: acquisition time per volume, TR/2 for MESMERISED STE and SE, TR for PGSE, as the basis for reported SNR efficiency and SNR per unit time calculations. b_0 tSNR: temporal SNR calculated from b_0 volumes, mean and standard deviation over white matter. b_0 tSNReff: temporal SNR per unit time (in units of $s^{-1/2}$) calculated from b_0 volumes, mean and standard deviation over white matter. b_0 tSNR/sec: temporal SNR per unit time (in units of s^{-1}) calculated from b_0 volumes, mean and standard deviation over white matter. FS/FA uncert: the uncertainty of fraction-of-sticks (FS for MESMERISED STE and PGSE volumes) and DTI fractional anisotropy (FA for MESMERISED SE volumes), mean and standard deviation over white matter.

Comparison with PGSE for 7 T high-b multi-shell diffusion MRI

We compared MESMERISED's tSNR efficiency for high b-value 7 T multi-shell acquisitions with MB PGSE, focusing on the highest b-value shell (b = 7000 s/mm^2) which is the most SNR limiting. Figure 9 shows results for MESMERISED for four different ES factors and at MB = 3 (Fig. 9A-D) and PGSE at MB = 2 (Fig. 9E), optimized for efficiency given SAR and gradient system limits. It can be seen that MESMERISED provides two useful volumes of multi-shell data (STE and SE) at two b-values in a shorter TR (\sim 3.4 s) than the TR for a single volume (4.21 s) for PGSE. In MESMERISED, tSNR is higher for SE than STE, as expected from the additional T₁ decay in STE, and tSNR maps for MESMERISED STE show higher values in both deep and cortical gray matter than PGSE. However, since high b-value

diffusion acquisitions are generally aimed at white matter, we quantitatively compared tSNR efficiency (tSNR-eff) in white matter. White matter tSNR-eff is markedly higher for both STE and SE volumes of all MESMERISED acquisitions than for PGSE (Fig. 9F, Table 1). MESMERISED STE tSNR-eff is at a stable level for all ES factors (2,3,4,6) while MESMERISED SE tSNR-eff increases with ES and decreasing TE. MESMERISED's main efficiency advantage is through the acquisition of both STE and SE volumes in a shorter TR, providing a far shorter acquisition time per volume (1.7 s versus 4.21 s for PGSE). Thus, although MESMERISED STE white matter tSNR, without regard of acquisition time, is 8.4% higher than for PGSE (e.g. for ES = 3: 13.89 vs. 12.81), tSNR per unit time is 171% higher for MESMERISED (8.24 s⁻¹ vs 3.04 s⁻¹), and tSNR efficiency is 71% higher for MESMERISED (10.70 s^{-1/2} vs 6.24 s^{-1/2}; Table 1).

The standard deviations of 30 - 40% of the mean indicate that there is quite some variation of tSNR over white matter in all cases. A comparison of tSNR with a B_1^+ field map (Figure 10A, B) shows that B_1^+ inhomogeneity is likely a factor in this: the spatial tSNR variation follows similar low frequency patterns in STE, SE and PGSE, which co-vary with B_1^+ inhomogeneity. For instance, the region of low flip angle deviation at the green arrow on the subject's left side (Figure 10A) is associated with high tSNR, whereas a region of high flip angle deviation, at a similar distance to the RF receiver coils (red arrow) shows low tSNR. With about 20% overflipping in the center of the brain, underflipping in regions of low B_1^+ reaches close to 50% (particular in right inferotemporal and inferior cerebellar regions; Supplementary Figure 2A,B), which are also the regions of lowest tSNR, up to 50% lower than highest observed values in each of MESMERISED STE, SE and PGSE (Supplementary Figure 2D, 5D and D6, respectively).

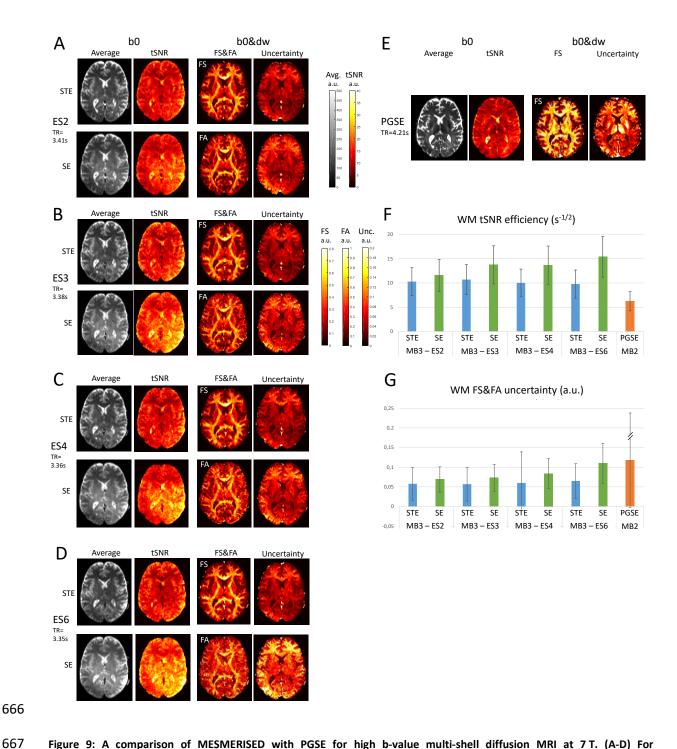


Figure 9: A comparison of MESMERISED with PGSE for high b-value multi-shell diffusion MRI at 7 T. (A-D) For MESMERISED with each of ESxMB = 2x3 (ES2 in A), ESxMB = 3x3 (ES3 in B), ESxMB = 4x3 (ES4 in C), ESxMB = 6x3 (ES6 in D): a transverse slice of whole brain 1.8mm isotropic results for MESMERISED with both STE (top) and SE (bottom) results. The left two panels show results derived from the b0 only acquisition with the average b0 volume and tSNR. The right two panels show results derived from the 60dw + 6b0 acquisition with FS (for STE) and FA (for SE) and their uncertainty. (E) PGSE results with MB = 2 as in A-D for its single spin-echo. (F) Mean tSNR efficiency over white matter only for all MESMERISED (MSMD) acquisitions (both STE and SE volumes) and PGSE. (G) Mean uncertainty over white matter only, for all MESMERISED (MSMD) acquisitions (both STE and SE volumes) and PGSE. Error bars: standard deviation. //: uncertainty error bar for PGSE downscaled 30x. All map scaling equal, and according to the color scale bars.

680

681

682

683

684

685 686

687

688

689

690

691 692

693

694

695

696 697

698

699

700

701

703

704

705

706

707

708

709

710

711

712

713

Beyond tSNR and tSNR efficiency, we assessed the effectiveness of MESMERISED and PGSE to support fiber orientation and microstructure modeling (Figure 9A-E, right columns, and used the uncertainty of FS and FA as a measure of modeling precision supported by the data. Fraction-of-stick (FS) maps for the high-b MESMERSED STE volumes have excellent WM-GM contrast for all ES factors. In clear distinction, FR for PGSE lacks WM-GM contrast, particularly in deep GM nuclei, and appears to have an overall upward bias. Fractional anisotropy (FA) maps for low-b MESMERISED SE volumes also show good delineation of WM and GM for ES = 2, 3 and 4. Overall, FS and FA uncertainty and tSNR appear negatively associated and driven by B₁⁺ inhomogeneity within each volume, a pattern which is evident over the whole brain (Supplementary Figures 2-4). FS uncertainty for STE appears lowest of all maps and relatively constant over all ES factors. FA uncertainty for SE increases with ES factor, despite increasing tSNR, likely reflecting the decreasing b-values with ES (cf. supplementary table 4). In particular, the high ES = 6 FA uncertainty values likely reflect the low b = 223 s/mm2 associated with these SE volumes, which is very low for DTI modeling, although low uncertainty values are still observed in regions of high tSNR. Compared over white matter, FS uncertainty over white matter is markedly lower for MESMERISED STE volumes than for PGSE, and white matter FA uncertainty is relatively low for MESMERISED SE volumes for ES = 2, 3, and 4 (Figure 9G). A significant factor beyond tSNR in the upward bias and higher uncertainty of FS for PGSE could be Gibbs ringing. Indeed, line plots of PGSE signal at the border of the lateral ventricle show considerable ringing which is much decreased in MESMERISED under the same image reconstruction (Figure 10C), corresponding to water signal that is roughly twice as high in PGSE with roughly similar WM signal (Supplementary Figure 2C vs 4C). The PGSE signal ringing clearly translates to strong ringing the PGSE FS map, whereas ringing in the MESMERISED FS map is strongly decreased in comparison (Figure 10D).

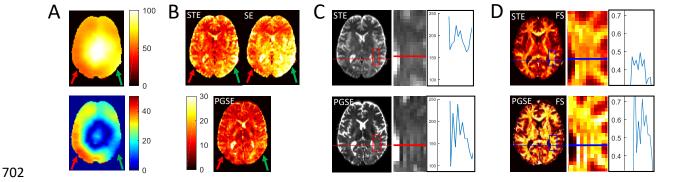


Figure 10: 7 T B_1^+ field inhomogeneity and Gibbs ringing in high-b diffusion imaging with MESMERISED and PGSE. (A) A transverse slice through an absolute B_1^+ map (in units of degrees; top) and the deviation of the local flip angle from the nominal 90 degrees (bottom) with indication of regions of low deviation (green arrow on the subject's left side; radiological display convention) and of high deviation (red arrow for the subject's right side). (B) tSNR for MESMERISED with ESxMB = 3x3 for both STE and SE b0 volumes and PGSE. Regions of low and high flip angle deviation are marked as in A. (C) The mean b_0 image for MESMERISED (ESxMB = 3x3) STE volumes and PGSE. Magnified portion at the edge of the lateral ventricle (middle; position indicated by the red box) and line-plot of single voxel intensities (right; position indicated by the edge of the lateral ventricle (middle; position indicated by the blue box) and line-plot of single voxel intensities at the edge (right; position indicated by the blue line).

Varying diffusion time

714715

716

717718

719

720

721

722

723

724 725

726

727

728

729

730

731

732 733

734

735

736

737

738

739

Analogous to keeping TR constant in MESMERISED T₁ relaxometry (by increasing ES factor proportionally to TM), different diffusion times for the STE volumes at the same TR can be realized by increasing ES factor with increasing diffusion time. When TE, b-value and diffusion gradient duration are also kept constant, this opens the possibility to explore the behavior of diffusion phenomena over different diffusion times. Figure 10 illustrates this with 2-shell diffusion data acquired at four diffusion times with otherwise matched parameters (i.e. TR, TE, δ and b-value are equal throughout, see Supplementary Table 3). Each 1.8 mm isotropic dataset had 24 directions at b = 1000 s/mm², 48 directions at $b = 2500 \text{ s/mm}^2$ and 8 b_0 's, for a total of 70 STE volumes (and 70 SE volumes, not shown here, with two-shell b-values for ES factor 1, 2, 3 and 4, respectively: [52, 130], [68 and 171], [99, 247], and [180, 450] s/mm²). Because of the same-TR requirement the relative temporal speedup for ES = 4 applies to the entire dataset and the speedup achieved compared to MB-STEAM is 2.38x, which means the < 17-minute acquisition time for MESMERISED would increase to over 40 minutes with MB-STEAM. The multi-shell data is sufficient for whole-brain DTI analysis (on the b = 1000s/mm² shell, Figure 10B) and DKI analysis (on both shells) at each diffusion time (Figure 10C), and to assess how diffusion measures such as mean diffusivity (MD) and mean kurtosis (MK) change with diffusion time (Figure 10D).

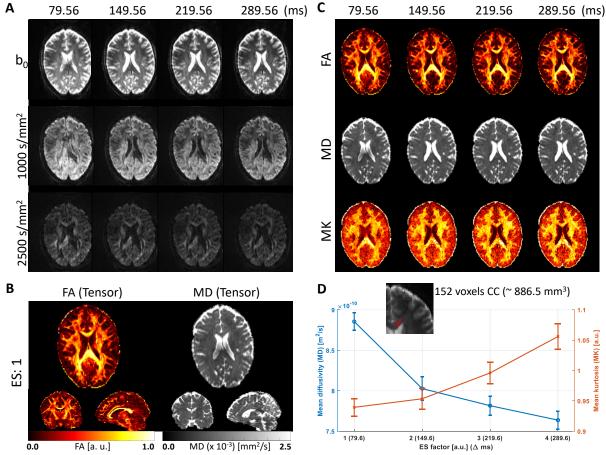


Figure 11: MESMERISED data and diffusion modeling results for STE volumes over several diffusion times at two b-values with matched TE and TR (TR = 3.50 s, TE = 45 ms) at 1.8 mm isotropic. (A) Example transverse slices through STE volumes at the two b-values and four diffusion times. (B) Whole-brain Tensor modeling resulting in FA and MD maps for the ES = 1 data (b = 1000 s/mm² shell only). (C) Tensor and Kurtosis modeling results (applied to suitable combinations of shells) for the different diffusion times. (D) Mean diffusivity (MD, blue) and Mean Kurtosis (MK, red) with their corresponding standard error plotted over ES factors/diffusion times for an ROI (red in the inset, with a total of 152 voxels along 10 slices) in the genu of the corpus callosum.

Quantitative multi-contrast mapping

740

741

742

743744

745

746

747

748

749

750

751

752

753

754

755

756

757

758

759

760

761

STEAM's ability to perform qT₂ and B₁⁺ mapping in addition to qT₁ and diffusion imaging creates an opportunity for fast multi-contrast mapping. Therefore, we explored MESMERISED's capacity for fast mapping of qT_1 , qT_2 , B_1 + and high-b diffusion with the same pulse sequence, given that qT_1 and diffusion are greatly accelerated. Here it is important to note that qT₂ and B₁⁺ mapping, per se, do not benefit from the acceleration by echo-shifting in MESMERISED. However, both these STEAM mapping methods do not require long TMs and are intrinsically already fast (and full acquisition duty cycle) techniques. Figure 12 shows whole brain quantitative multi-contrast mapping at 1.8 mm isotropic, achieving qT_1 , qT_2 , B_1^+ mapping as well as extensive diffusion mapping in about 17 minutes. Crucially, the use of the same sequence and readout EPI provides for intrinsically aligned maps. Supplementary Figure 5 shows how qT2 maps are derived from relaxometry on TE (both with and without echo-shifting), showing that qT2 mapping with the SE volumes can be performed independent of echo-shifting and the TM of the STE. Supplementary Figure 6 shows how MESMERISED B_1^+ mapping is derived from both the SE and STE volumes for different α_{TX} multipliers in an α_{TX} - $2\alpha_{TX}$ - α_{TX} flip angle scheme for Exc – Sto – Rec pulses. The result in Figure 12 is for 3 repetitions of all volumes in the qT₁, qT₂ and B₁⁺ mapping as well as a total of 8 b-shells over 370 volumes for the diffusion. This delivers high precision (low uncertainty, cf. Figure 5) in qT_1 , qT_2 and B₁⁺ mapping, each in roughly 2 minutes, and diffusion mapping in 11 minutes. Alternatively, acquiring a single repetition for the qT_1 , qT_2 and B_1^+ maps, each in roughly 1 minute, and acquiring 200 volumes for the diffusion dataset (cf. Supplementary Tables 1-3), would lower the total multi-contrast acquisition time to under 10 minutes (i.e. qT₁ mapping in 1:18, qT₂ mapping 0:56, B₁⁺ mapping 1:05 and 200 volume multi-shell diffusion in 6:15).

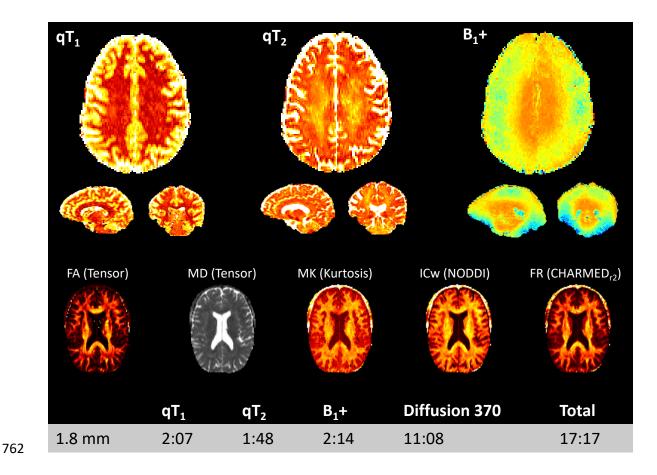


Figure 12: Whole brain quantitative multi-contrast mapping with MESMERISED at 1.8 mm isotropic in 17 minutes 17 seconds. This high precision protocol consists of 3 repetitions of all volumes in the qT₁, qT₂ and B₁⁺ as well as 185 high-b multi-shell STE volumes and 185 low-b multi-shell SE diffusion weighted volumes for a total of 8 b-shells over 370 volumes.

Discussion

763

764

765

766

767

768 769

770

771

772

773774

775

776

777

778

779

780

781

782

783

784

785 786

787

788 789

790

791

792 793

794

795

796

797

798

799 800

801

802

803

804

805

806

MESMERISED achieves super-accelerated 7 T STEAM imaging by combining echo-shifting and multiband/simultaneous multi-slice acceleration, leading to very high multiplicative acceleration factors and time efficiency for qT_1 and diffusion imaging. Furthermore, qT_2 and B_1^+ mapping can be performed with the same pulse sequence. MESMERISED inherits from STEAM the capacity to simultaneous generate both SE images and STE images and the possibility of performing qT_1 , qT_2 , B_1^+ mapping as well as diffusion-weighted imaging. It also inherits the property of 50% lower SNR and 40% lower SAR compared to a similar spin-echo sequence, and 66% lower SAR compared to an inversion-prepared spin-echo sequence. In essence, MESMERISED capitalizes on the low SAR of the STEAM sequence to considerably increase its SNR per unit time through the multiplicative ESxMB acceleration. This enables fast qT_1 , diffusion and quantitative multi-contrast mapping over the wholebrain, which we will discuss in turn.

qT₁ mapping

MESMERISED T₁ relaxometry can be performed at a low constant TR through the block reordering property (Figure 1): increasing the ES factor proportionally to TM results in increasing total acceleration factors (e.g. from 3 to 36 in 1.8 mm data and from 3 to 54 in 1.25 mm data in Figure 4), highlighting the super-acceleration capability of MESMERISED for qT_1 imaging. Although the temporal speed-up of the multiplicative ESxMB acceleration is not linear with ES, the speed-up compared to multiband only accelerated STEAM can be very large (e.g. 6.02x faster than MB = 3 for 1.8 mm and 9.07x faster than MB = 3 for 1.25 mm). In addition, MESMERISED simultaneously provides spin-echo images in the same acquisition time which, we show here, can be used to either improve the precision of the qT₁ maps in the same acquisition time or achieve even shorter acquisition times for the same precision level. For instance, uncertainty in 1.25 mm isotropic qT₁ maps at a single repetition (acquisition time 1:40) with STE/SE ratio analysis are at similar levels as that of three repetitions (acquisition time 4:01) or even 5 repetitions (acquisition time 6:22) for STE-only analysis (Figure 5, bottom row), which amounts to an effective speedup of ~4x purely through the use of both SE and STE in qT₁ mapping. Furthermore, the precision tends to be more homogenous over the brain. The efficiency gain for high precision could be explained by the lower complexity of model fit performed in STE/SE ratio analysis, and the better homogeneity by a decreased confounding effect of T₂, RF receive coil sensitivities and proton density on the fit. The efficiency gain is particularly interesting in the context of the theoretical efficiency of MESMERISED compared to e.g. IR-EPI. The use of a STE halves the signal intensity and SNR, which in principle requires an acquisition time factor of 4 to overcome and accelerating STEAM for qT₁ mapping would therefore only seem to provide useful SNR efficiency benefits beyond a 4x speedup. However, these results indicate that effectively using both the half signal SE and half signal STE in STEAM can compensate for the 50% signal decrease, at least in terms of the precision of the resulting qT₁ maps. Here it should be noted that good results for qT₁ mapping are obtained at less than 100% duty cycle, with a doubling of the minimum achievable TR and TM (Figure 5, bottom row vs. top row), providing a better spacing and range of TM's for T_1 relaxometry. Potentially, in the future, this space can be used for simultaneous sampling of other contrasts, such as a multi-echo EPI readout (Poser and Norris, 2009) for qT_2/qT_2^* mapping for further efficiency.

When compared to the MP2RAGE sequence (Marques et al., 2010), MESMERISED's single-shot EPI readout strategy lends itself to sampling considerably more points on the full T₁ decay curve (here 7 or 8 TMs vs. 2 inversion times for MP2RAGE) in less time, allowing a more straightforward relaxometry approach, but does so at a considerably lower resolution (here 1.8 to 1.25 mm vs. 1 to 0.6 mm for MP2RAGE). Similarly, the variable flip angle approach in DESPOT1 (Deoni, 2007) with spoiled gradient recalled echo (SPGR) images lends itself well for 1 mm resolution qT₁ maps at 3 T, but contends with considerably sensitivity to B₁⁺ inhomogeneities. In that sense MESMERISED's T₁ relaxation sampling strategy is closer to MS-IR-EPI (Panchuelo et al., 2021) and it could benefit from the multi-shot readout applied there to enable qT₁ mapping with higher spatial resolution than presented here. The denser sampling of the decay curve in MESMERISED and MS-IR-EPI may lend itself to better characterisation of multi-component or multi-compartment qT₁ mapping than MP2RAGE and DESPOT, particularly in the context of quantitative multi-parameter mapping and when spatially aligned diffusion MRI is needed.

Diffusion MRI

807

808 809

810

811

812

813

814

815

816

817

818

819

820

821

822

823

824

825

826 827

828

829

830

831

832

833

834

835

836

837

838

839

840

841

842

843

844

845

846

847848

849

850

For diffusion imaging, MESMERISED inherits from STEAM the capacity to achieve long diffusion times and high b-values, as well as the possibility to simultaneously generate diffusion-weighted SE images and diffusion-weighted STE images. MESMERISED adds to this a very high data rate to explore a large range of b-values, diffusion directions, or diffusion times in a short time. For instance, 370 wholebrain diffusion volumes over 8 b-value shells up to b = 7000 s/mm² can be generated at 2 mm isotropic in 7 min and 47 s (ESxMB = 5x3, 2.95x speedup compared to MB-STEAM), a data rate of almost a volume per second, or at 1.8 mm isotropic in 10 min and 53 s (ESxMB = 6x3, 3.43x speedup compared to MB-STEAM), a data rate of more than a volume per 2 seconds. In addition, this is achieved with a lower diffusion gradient duty cycle (as well as less gradient coil heating and accompanying cooling requirements) than PGSE, since the longer diffusion times in STEAM dictate shorter diffusion gradient durations for the same maximum b-value. The shells consist of one half SE volumes in low-b shells which can support analysis with Gaussian diffusion models as in DTI, and one half STE volumes in the same number high-b shells which are suitable for non-Gaussian diffusion models, such as DKI, NODDI and CHARMED models. This provides a wealth of diffusion information from several different models in short acquisition time, which could be further extended in future work. For instance, instead of collecting a relatively large number of directions in 4 shells for each SE and STE, fewer (e.g. 6-12) directions could be acquired at each of many (e.g. 14-28) b-values. When carefully chosen, this would continue to support fitting of the aforementioned models on high-b STE volumes, while creating an opportunity to additionally fit the intra-voxel incoherent motion (IVIM) model (LeBihan, 1990) to the richly sampled low b-value domain in the SE volumes. Further tuning potential for the b-values of SE volumes (for a given STE max-b) lies in the choice of mixing time (and therefore diffusion time) of the STE volumes because, for the same TR and STE b-value, the SE bvalue varies with different choices of TM and δ (see Supplementary Table 4).

Comparison with MB PGSE for 7 T high-b multi-shell diffusion MRI

In a direct comparison for high-b multi-shell diffusion imaging at 7 T, MESMERISED displays markedly higher tSNR efficiency in white matter than MB PGSE at 7 T, when matched for RF-pulse regime, readout EPI train and image reconstruction. MESMERISED tSNR per unit time is ~170% higher, and its

tSNR efficiency is ~70% higher in white matter than that of MB PGSE. The gains in gray matter are potentially even higher, but the comparison was aimed at white matter, since this is currently the main application domain of high-b diffusion MRI. To illustrate, this efficiency allows MESMERISED to acquire a 370 volume 8-shell diffusion dataset at 1.8 mm isotropic in ~11 min, which would take MB PGSE ~26.5 min to acquire at lower tSNR. The main factors in this are MESMERISED's acquisition of both a high-b STE and low-b SE volume in a TR, and PGSE's limitations of SAR and gradient duty cycle, which forces it to run at increased TR and TE. Of note here is that SAR and gradient limits each have a similar individual effect on PGSE TR: in the investigated case, SAR limits forced its TR upwards to 4 s and gradient limits forced it up a further ~5% to 4.21 s. Therefore, avoiding only one of the limits would not shorten TR much to improve PGSE efficiency. MESMERISED avoids these limitations through a lower SAR and gradient load allowing full acquisition duty cycle. Combined with its effective use of both the diffusion weighted SE and STE, this allows it to overcome the 50% signal disadvantage (and the theoretical factor 4x acquisition time disadvantage) and outperform MB PGSE for 7 T high-b diffusion imaging.

B₁⁺ maps at the employed reference voltage show that at 20% overflipping in the center of the brain, up to 50% underflipping occurred in other parts of the brain, despite the use of dielectric pads. This suggest that higher reference voltages could perhaps provide a higher overall level of tSNR over the brain, at the cost of lower tSNR in the center, higher SAR levels, and consequently increase in (SAR limited) TRs. This might affect PGSE more than MESMERISED since PGSE at MB = 2 has roughly 10% higher SAR per TR than MESMERISED at MB = 3. The SAR load for PGSE could be improved through parallel transmission (pTX) techniques and optimized MB pulses such as PINS (Norris et al., 2014; Norris et al., 2011), multi-PINS (Eichner et al., 2014) and pTx-MB (Poser et al., 2014; Wu et al., 2013) but the improved SAR would equally allow higher MB factors and higher efficiency in MESMERISED. The fact that B₁⁺ inhomogeneity is one of the main drivers of spatial variations in tSNR and diffusion modeling uncertainty strongly suggests that methods for improved B₁⁺ homogeneity, such as pTX or improved dielectric pad strategies, are desirable for both PGSE and MESMERISED.

Beyond efficiency, MESMERISED's effectiveness in fiber orientation and microstructure modeling was qualitatively and quantitatively higher than that of PGSE. Diffusion modeling parameter maps (FS and FA) for both MESMERSED STE and SE volumes have excellent WM-GM contrast for several different echo-shift factors, whereas those for PGSE lack WM-GM contrast, particularly in deep GM nuclei, and appear to have an overall upward bias. Modeling precision quantified by uncertainty is markedly better for both MESMERISED STE and SE volumes than for PGSE, excluding DTI modeling at very low b-values. A significant factor beyond tSNR in the upward bias and higher uncertainty of FS for PGSE is the strong observed Gibbs ringing caused by the high water signal, likely through the long TE and strong resulting T2-weighting. The assessment of efficiency and effectiveness on 7T multi-shell diffusion MRI was performed at b = 7000 s/mm² with a high performing 70 mT/m gradient. The chosen b-value is non-arbitrary, as it has been shown that this is the minimum diffusion weighting necessary to fully suppress the contribution of extra-axonal water to the signal and obtain a high specificity to the intra-axonal compartment (more generally: the restricted compartment internal to thin axonal, glial or dendritic processes; Veraart et al., 2019), after the need for high b-values to accurately distinguish intra- and extra-cellular diffusion had already been made clear (Assaf and Basser, 2005; Assaf et al., 2004). Various types of 7 T PGSE-EPI acquisitions at moderate b-values (between ~1000 and ~2000 s/mm²) have yielded high quality data at very high spatial resolutions, highly suitable for axon orientation modeling and tractography (Eichner et al., 2014; Vu et al., 2015), including those with in-plane segmentation (Heidemann et al., 2010), and slice-dithered throughplane resolution enhancement (Setsompop et al., 2018). However, such acquisitions lack the intraaxonal signal specificity at high b-values, important for accurate diffusion modeling of axonal microstructure and integrity, which we targeted here. Therefore, we believe that while the domain of moderate-b high-resolution 7 T diffusion imaging may be best served by PGSE-EPI variants, MESMERISED is superior for high-b moderate resolution 7 T diffusion imaging. In the future, the limits imposed by gradient duty cycle for high-b PGSE could be improved by further gradient hardware developments beyond the employed generation of high performing 70-80 mT/m gradients. For PGSE, higher amplitudes, higher duty cycle, or both simultaneously, could strongly reduce its TE, which is dominated by diffusion gradient duration, while the already short TEs in MESMERISED might benefit less. On the other hand, low TE readout strategies, such as center-out spiral sampling, could strongly reduce TE in MESMERISED (which is dominated by readout train length), and provide proportionally more benefit than for PGSE.

Varying diffusion time

896

897

898

899

900

901

902

903

904

905

906

907

908

909 910

911 912

913

914

915

916

917

918

919

920

921 922

923

924

925

926

927

928

929

930

931

932

933

934

935

936

937

938

The combination of high data rate and a wide range of achievable diffusion times makes MESMERISED an excellent tool to investigate the behavior of diffusion phenomena at long and varying diffusion times. Several studies have shown that processes such as exchange or mesoscopic structure due to random axonal packing leave their mark in the change of diffusion MRI measures with increasing diffusion time (Burcaw et al., 2015; Nilsson et al., 2013; Novikov et al., 2014). Diffusion time-resolved dMRI can even be used to achieve some sensitivity to axonal diameter distributions (Alexander et al., 2010; Assaf et al., 2008; De Santis et al., 2016b). Most of these applications require quite extensive sampling of diffusion times, b-values and, sometimes, directions, often leading to long (e.g. 30 - 60 min) acquisitions with current techniques. MESMERISED can achieve time-efficient sampling of different diffusion times in the STE volumes at otherwise matched parameters by increasing ES factor with increasing diffusion time. We demonstrate this with 1.8 mm isotropic diffusion data acquired at four diffusion times from ~ 80 ms to ~ 290 ms, which supports both DTI and DKI at each diffusion time (Figure 12). The analysis of the resulting STE data with DTI and DKI captures a dependence of mean diffusivity and mean kurtosis with increasing diffusion time in white matter (Figure 10D), in agreement with earlier reports (Fieremans et al., 2016; Jespersen et al., 2018). In the future, MESMERISED could be a useful tool in further investigating such micro- and meso-structural phenomena. In principle, less than the number of directions acquired here per bvalue (24 for b = 1000 s/mm², 48 for b = 2500 s/mm² at a total acquisition time of ~ 17 min for all diffusion times) might be sufficient for DKI and thus a shorter MESMERISED acquisition may be possible for this purpose. Furthermore, sampling higher diffusion times is a possibility, since correspondingly higher ES factors allow these to be acquired at the same TR (e.g. 430 ms at ES = 6 and 560 ms at ES = 8), and the performed PGSE comparison experiment shows that high tSNR is maintained with MESMERISED even at b-values up to b = 7000 s/mm² and a diffusion time up to 430ms.

Quantitative multi-contrast mapping

MESMERISED enables fast multi-contrast mapping over the whole-brain, for instance at 1.8 mm isotropic, with qT_1 mapping, qT_2 mapping, B_1^+ mapping and 370 volume multi-shell diffusion, in a little over 17 minutes. Accepting a lower precision level in the quantitative maps (i.e. working with a

single, rather than three repetitions), and 200 volumes in the diffusion dataset (cf. Supplementary Tables 1-3), would lower the total multi-contrast acquisition time to under 10 minutes (i.e. qT1 mapping in 1:18, qT2 mapping 0:56, B₁⁺ mapping 1:05 and 200 volume multi-shell diffusion in 6:15). Here, each modality includes ~30 s of GRAPPA and MB reference scans, which in the future might be shared, providing another 1:30 time gain. Here we note again that qT₂ and B₁⁺ mapping, per se, do not benefit from the accelerating echo-shifting capacity in MESMERISED. They derive from basic properties of the STEAM sequence, i.e. relaxometry on SE or STE echo times and the α - 2α - α technique, respectively, which can be performed independently of any echo-shifting. However, the ability to perform qT2 and B1+ mapping with the same pulse sequence that provides highly accelerated qT₁ and diffusion imaging has considerable advantages for fast multi-contrast mapping. First, all quantitative contrasts are generated by the same sequence and readout train, leading to similar image distortions which aid alignment of the different maps where needed. Second, for the same reason, the images from all contrasts share the same image noise distribution, which aids denoising with methods that take all images into account (Cordero-Grande et al., 2019; Moeller et al., 2021; Veraart et al., 2016). Third, well-aligned B_1^+ maps can help in correcting transmit field related biases in quantitative maps, for instance in qT_1 maps (Marques et al., 2010). As also mentioned above, in comparison to quantitative multi-contrast mapping techniques based on a SPGR readout, such as DESPOT1/DESPOT2 (Deoni et al., 2005), MP2RAGEME (Caan et al., 2019) and MPM (Weiskopf et al., 2013), MESMERISED with EPI readout will more easily sample many more points on T₁ and T₂ decay curves, allowing a more straightforward relaxometry approach, but at a lower resolution. In its capacity for sampling qT_1 , qT_2 and diffusion contrasts efficiently, MESMERISED is similar to the ZEBRA technique (Hutter et al., 2018), which is instead based on IR-prepared spin-echo EPI, and therefore more SAR intensive. This is less of a hindrance at 3T, and extensive sampling of the multi-dimensional [T1, T2*, b] contrast space could be achieved efficiently with ZEBRA, though at considerably lower spatial resolution (2.5 vs. 1.8 mm isotropic) and lower b-values (2600 vs. 7000 s/mm²) than those reported here. We believe the efficient sampling of the $[T_1, T_2, B_1, b, \Delta]$ contrast space in MESMERISED is ideal for characterisation of multiple microstructural components or compartments, particularly when spatially aligned diffusion MRI with high b-values and/or varying diffusion times is needed.

Outlook

939

940 941

942

943

944

945

946

947

948

949

950

951

952

953

954

955

956957

958

959

960

961

962 963

964

965

966

967

968

969

970

971

972973

974

975

976

977

978

979

980

981

982

983

As mentioned above, a useful extension will be a multi-echo EPI readout, which would enable further efficiency in mapping both qT_1 and qT_2/qT_2^* since unused duty cycle in optimal-TM sampling for T_1 relaxometry can be used to sample multiple TE's per shot for T_2/T_2^* relaxometry. This could potentially also make use of efficient sparse sampling trajectories of the temporal relaxation curve, such as in echo-planar time-resolved imaging (EPTI; Wang et al., 2019). A multi-echo readout would also enable further speed-up of mixed or dependent contrast sampling, such as mixed T_2/T_2^* relaxometry and diffusion imaging (Tax, 2017; Veraart et al., 2018), and help characterise tissue compartments with separate relaxation constants and diffusion behavior (De Santis et al., 2016a; Hutter et al., 2018; Wu, 2017). Future combination with SAR efficient MB pulses, such as PINS (Norris et al., 2014; Norris et al., 2011), multi-PINS (Eichner et al., 2014) and pTx-MB (Poser et al., 2014; Wu et al., 2013) could allow high data quality at potentially even higher MB factors. For the fastest acquisitions, acquisition of the reference data for GRAPPA and MB reconstruction takes a considerable proportion of the running time (about 30 s in the current implementation), which might be sped-up further with more efficient strategies. Additionally, a flexible table-based sequence

looping implementation in which all parameters (TM, ES, TE, flip angles, diffusion direction and gradient amplitude) can be controlled independently, could allow running qT_1 , qT_2 , B_1^+ and diffusion mapping in a single sequence run, akin to magnetic resonance fingerprinting (Ma et al., 2013) in its Cartesian form (Benjamin et al., 2019). When EPI readouts are employed, shared reference scans and reversed phase-encode direction scans, would further decrease overhead time. Finally, the SAR efficiency of a STEAM-based sequence makes it very attractive for high duty cycle and multiband acceleration at 7 T, but many of the advantages could translate well to 3 T, particularly when MB factors are increased even further, as allowed by modern 32 to 64 channel head or head/neck RF-coils.

Conclusion

MESMERISED achieves super-accelerated 7 T STEAM imaging by combining echo-shifting and multiband/simultaneous multi-slice acceleration. This leads to very high multiplicative acceleration factors and time efficiency for qT₁ and diffusion imaging. MESMERISED can probe combined T₁, T₂ and diffusion contrast with high time efficiency for fast multi-contrast mapping and characterization of multi-component relaxometry, diffusion, and exchange.

Acknowledgements

AR and FJ were supported by an ERC Starting Grant (MULTICONNECT, #639938). AR was further supported by a Dutch science foundation (NWO) VIDI Grant (#14637). FJ was partially supported by the German Research Foundation (DFG Emmy Noether Program: MO 2397/4-1). BP is partially funded by NWO VIDI Grant 016.Vidi.178.052. and NIH R01 MH111444/MH/NIMH.

Declaration of interest

AR, BP an FJ are co-inventors on a patent application related to the technique described in this manuscript.

References

1011

1031

1033

1040

1041

1042

1043

- 1012 Alexander, D.C., 2008. A general framework for experiment design in diffusion MRI and its 1013 application in measuring direct tissue-microstructure features. Magn Reson Med 60, 439-448.
- 1014 Alexander, D.C., Hubbard, P.L., Hall, M.G., Moore, E.A., Ptito, M., Parker, G.J., Dyrby, T.B., 2010. 1015 Orientationally invariant indices of axon diameter and density from diffusion MRI. Neuroimage 1016 52, 1374-1389.
- 1017 Andersson, J.L., 2008. Maximum a posteriori estimation of diffusion tensor parameters using a Rician noise model: why, how and but. Neuroimage 42, 1340-1356. 1018
- 1019 Assaf, Y., Alexander, D.C., Jones, D.K., Bizzi, A., Behrens, T.E., Clark, C.A., Cohen, Y., Dyrby, T.B., Huppi, 1020 P.S., Knoesche, T.R., Lebihan, D., Parker, G.J., Poupon, C., consortium, C., Anaby, D., Anwander, 1021 A., Bar, L., Barazany, D., Blumenfeld-Katzir, T., De-Santis, S., Duclap, D., Figini, M., Fischi, E., 1022 Guevara, P., Hubbard, P., Hofstetter, S., Jbabdi, S., Kunz, N., Lazeyras, F., Lebois, A., Liptrot, M.G., 1023 Lundell, H., Mangin, J.F., Dominguez, D.M., Morozov, D., Schreiber, J., Seunarine, K., Nava, S., 1024 Poupon, C., Riffert, T., Sasson, E., Schmitt, B., Shemesh, N., Sotiropoulos, S.N., Tavor, I., Zhang, 1025 H.G., Zhou, F.L., 2013. The CONNECT project: Combining macro- and micro-structure. 1026 Neuroimage 80, 273-282.
- 1027 Assaf, Y., Basser, P.J., 2005. Composite hindered and restricted model of diffusion (CHARMED) MR 1028 imaging of the human brain. Neuroimage 27, 48-58.
- 1029 Assaf, Y., Blumenfeld-Katzir, T., Yovel, Y., Basser, P.J., 2008. AxCaliber: a method for measuring axon 1030 diameter distribution from diffusion MRI. Magn Reson Med 59, 1347-1354.
- Assaf, Y., Freidlin, R.Z., Rohde, G.K., Basser, P.J., 2004. New modeling and experimental framework to characterize hindered and restricted water diffusion in brain white matter. Magn Reson Med 52, 1032 965-978.
- 1034 Assaf, Y., Pasternak, O., 2008. Diffusion tensor imaging (DTI)-based white matter mapping in brain 1035 research: a review. J Mol Neurosci 34, 51-61.
- 1036 Basser, P.J., Mattiello, J., LeBihan, D., 1994. MR diffusion tensor spectroscopy and imaging. Biophys J 1037 66, 259-267.
- 1038 Beaulieu, C., 2002. The basis of anisotropic water diffusion in the nervous system - a technical 1039 review. NMR Biomed 15, 435-455.
 - Behrens, T.E., Berg, H.J., Jbabdi, S., Rushworth, M.F., Woolrich, M.W., 2007. Probabilistic diffusion tractography with multiple fibre orientations: What can we gain? Neuroimage 34, 144-155.
 - Behrens, T.E., Woolrich, M.W., Jenkinson, M., Johansen-Berg, H., Nunes, R.G., Clare, S., Matthews, P.M., Brady, J.M., Smith, S.M., 2003. Characterization and propagation of uncertainty in diffusion-weighted MR imaging. Magn Reson Med 50, 1077-1088.
- 1045 Benjamin, A.J.V., Gomez, P.A., Golbabaee, M., Mahbub, Z.B., Sprenger, T., Menzel, M.I., Davies, M., 1046 Marshall, I., 2019. Multi-shot Echo Planar Imaging for accelerated Cartesian MR Fingerprinting: 1047 An alternative to conventional spiral MR Fingerprinting. Magn Reson Imaging 61, 20-32.
- 1048 Berman, S., Filo, S., Mezer, A.A., 2019. Modeling conduction delays in the corpus callosum using MRI-1049 measured g-ratio. Neuroimage 195, 128-139.
- 1050 Blazejewska, A.I., Bhat, H., Wald, L.L., Polimeni, J.R., 2017. Reduction of across-run variability of 1051 temporal SNR in accelerated EPI time-series data through FLEET-based robust autocalibration. 1052 Neuroimage 152, 348-359.
- 1053 Breuer, F.A., Blaimer, M., Mueller, M.F., Seiberlich, N., Heidemann, R.M., Griswold, M.A., Jakob, P.M., 1054 2006. Controlled aliasing in volumetric parallel imaging (2D CAIPIRINHA). Magn Reson Med 55, 1055 549-556.
- 1056 Burcaw, L.M., Fieremans, E., Novikov, D.S., 2015. Mesoscopic structure of neuronal tracts from time-1057 dependent diffusion. Neuroimage 114, 18-37.
- 1058 Caan, M.W.A., Bazin, P.L., Marques, J.P., de Hollander, G., Dumoulin, S.O., van der Zwaag, W., 2019. 1059 MP2RAGEME: T1, T2 (*), and QSM mapping in one sequence at 7 tesla. Hum Brain Mapp 40, 1060 1786-1798.
- 1061 Callaghan, M.F., Helms, G., Lutti, A., Mohammadi, S., Weiskopf, N., 2015. A general linear 1062 relaxometry model of R1 using imaging data. Magn Reson Med 73, 1309-1314.

- 1063 Campbell, J.S.W., Leppert, I.R., Narayanan, S., Boudreau, M., Duval, T., Cohen-Adad, J., Pike, G.B., 1064 Stikov, N., 2018. Promise and pitfalls of g-ratio estimation with MRI. Neuroimage 182, 80-96.
- 1065 Cauley, S.F., Polimeni, J.R., Bhat, H., Wald, L.L., Setsompop, K., 2014. Interslice leakage artifact 1066 reduction technique for simultaneous multislice acquisitions. Magn Reson Med 72, 93-102.
- 1067 Chung, S., Kim, D., Breton, E., Axel, L., 2010. Rapid B1+ mapping using a preconditioning RF pulse with 1068 TurboFLASH readout. Magn Reson Med 64, 439-446.
- 1069 Cordero-Grande, L., Christiaens, D., Hutter, J., Price, A.N., Hajnal, J.V., 2019. Complex diffusion-1070 weighted image estimation via matrix recovery under general noise models. Neuroimage 200, 1071 391-404.
- 1072 Cusack, R., Papadakis, N., 2002. New robust 3-D phase unwrapping algorithms: application to 1073 magnetic field mapping and undistorting echoplanar images. Neuroimage 16, 754-764.
 - De Santis, S., Assaf, Y., Jeurissen, B., Jones, D.K., Roebroeck, A., 2016a. T1 relaxometry of crossing fibres in the human brain. Neuroimage 141, 133-142.
 - De Santis, S., Drakesmith, M., Bells, S., Assaf, Y., Jones, D.K., 2014. Why diffusion tensor MRI does well only some of the time: variance and covariance of white matter tissue microstructure attributes in the living human brain. Neuroimage 89, 35-44.
- 1079 De Santis, S., Jones, D.K., Roebroeck, A., 2016b. Including diffusion time dependence in the extra-1080 axonal space improves in vivo estimates of axonal diameter and density in human white matter. Neuroimage 130, 91-103.
 - Deoni, S.C., 2007. High-resolution T1 mapping of the brain at 3T with driven equilibrium single pulse observation of T1 with high-speed incorporation of RF field inhomogeneities (DESPOT1-HIFI). J Magn Reson Imaging 26, 1106-1111.
 - Deoni, S.C., Peters, T.M., Rutt, B.K., 2005. High-resolution T1 and T2 mapping of the brain in a clinically acceptable time with DESPOT1 and DESPOT2. Magn Reson Med 53, 237-241.
 - Ehses, P., Bause, J., Shajan, G., Scheffler, K., 2015. Efficient generation of T2*-weighted contrast by interslice echo-shifting for human functional and anatomical imaging at 9.4 Tesla. Magn Reson Med 74, 1698-1704.
- 1090 Eichner, C., Wald, L.L., Setsompop, K., 2014. A low power radiofrequency pulse for simultaneous 1091 multislice excitation and refocusing. Magn Reson Med 72, 949-958.
 - Feinberg, D.A., Moeller, S., Smith, S.M., Auerbach, E., Ramanna, S., Gunther, M., Glasser, M.F., Miller, K.L., Ugurbil, K., Yacoub, E., 2010. Multiplexed echo planar imaging for sub-second whole brain FMRI and fast diffusion imaging. PLoS One 5, e15710.
 - Fieremans, E., Benitez, A., Jensen, J.H., Falangola, M.F., Tabesh, A., Deardorff, R.L., Spampinato, M.V., Babb, J.S., Novikov, D.S., Ferris, S.H., Helpern, J.A., 2013. Novel white matter tract integrity metrics sensitive to Alzheimer disease progression. AJNR Am J Neuroradiol 34, 2105-2112.
- 1098 Fieremans, E., Burcaw, L.M., Lee, H.H., Lemberskiy, G., Veraart, J., Novikov, D.S., 2016. In vivo 1099 observation and biophysical interpretation of time-dependent diffusion in human white matter. 1100 Neuroimage 129, 414-427.
- 1101 Frahm, J., Merboldt, K.D., Hänicke, W., Haase, A., 1985. Stimulated echo imaging. Journal of Magnetic 1102 Resonance 64, 81-93.
- 1103 Fukunaga, M., Li, T.Q., van Gelderen, P., de Zwart, J.A., Shmueli, K., Yao, B., Lee, J., Maric, D., 1104 Aronova, M.A., Zhang, G., Leapman, R.D., Schenck, J.F., Merkle, H., Duyn, J.H., 2010. Layer-1105 specific variation of iron content in cerebral cortex as a source of MRI contrast. Proc Natl Acad 1106 Sci U S A 107, 3834-3839.
- 1107 Gibson, A., Peters, A.M., Bowtell, R., 2006. Echo-shifted multislice EPI for high-speed fMRI. Magn 1108 Reson Imaging 24, 433-442.
- Griswold, M.A., Jakob, P.M., Heidemann, R.M., Nittka, M., Jellus, V., Wang, J., Kiefer, B., Haase, A., 1109 1110 2002. Generalized autocalibrating partially parallel acquisitions (GRAPPA). Magn Reson Med 47, 1111 1202-1210.
- Hahn, E.L., 1950. Spin Echoes. Physical Review 40. 1112
- 1113 Harms, R.L., Fritz, F.J., Schoenmakers, S., Roebroeck, A., 2019. Fast quantification of uncertainty in
- 1114 non-linear diffusion MRI models for artifact detection and more power in group studies. bioRxiv,
- 1115 651547.

1075

1076 1077

1078

1081

1082 1083

1084

1085

1086

1087

1088

1089

1092

1093

1094

1095

1096

- Harms, R.L., Fritz, F.J., Tobisch, A., Goebel, R., Roebroeck, A., 2017. Robust and fast nonlinear
 optimization of diffusion MRI microstructure models. Neuroimage 155, 82-96.
- Heidemann, R.M., Porter, D.A., Anwander, A., Feiweier, T., Heberlein, K., Knosche, T.R., Turner, R.,
 2010. Diffusion imaging in humans at 7T using readout-segmented EPI and GRAPPA. Magn
 Reson Med 64, 9-14.
- Hutter, J., Slator, P.J., Christiaens, D., Teixeira, R., Roberts, T., Jackson, L., Price, A.N., Malik, S., Hajnal, J.V., 2018. Integrated and efficient diffusion-relaxometry using ZEBRA. Sci Rep 8, 15138.
- 1123 Ivanov, D., Schafer, A., Streicher, M.N., Heidemann, R.M., Trampel, R., Turner, R., 2010. A simple low-1124 SAR technique for chemical-shift selection with high-field spin-echo imaging. Magn Reson Med 1125 64, 319-326.
- Jelescu, I.O., Veraart, J., Adisetiyo, V., Milla, S.S., Novikov, D.S., Fieremans, E., 2015. One diffusion acquisition and different white matter models: how does microstructure change in human early development based on WMTI and NODDI? Neuroimage 107, 242-256.
- Jenkinson, M., Beckmann, C.F., Behrens, T.E., Woolrich, M.W., Smith, S.M., 2012. Fsl. Neuroimage 62, 1130 782-790.
- Jensen, J.H., Helpern, J.A., Ramani, A., Lu, H., Kaczynski, K., 2005. Diffusional kurtosis imaging: the quantification of non-gaussian water diffusion by means of magnetic resonance imaging. Magn Reson Med 53, 1432-1440.
- Jespersen, S.N., Olesen, J.L., Hansen, B., Shemesh, N., 2018. Diffusion time dependence of microstructural parameters in fixed spinal cord. Neuroimage 182, 329-342.
- Jones, D.K., 2004. The effect of gradient sampling schemes on measures derived from diffusion tensor MRI: a Monte Carlo study. Magn Reson Med 51, 807-815.
- Jones, D.K., Knosche, T.R., Turner, R., 2013. White matter integrity, fiber count, and other fallacies: the do's and don'ts of diffusion MRI. Neuroimage 73, 239-254.
- Larkman, D.J., Hajnal, J.V., Herlihy, A.H., Coutts, G.A., Young, I.R., Ehnholm, G., 2001. Use of multicoil arrays for separation of signal from multiple slices simultaneously excited. J Magn Reson Imaging 13, 313-317.
- LeBihan, D., 1990. IVIM method measures diffusion and perfusion. Diagn Imaging (San Franc) 12, 133, 136.
- Li, K., Li, H., Zhang, X.Y., Stokes, A.M., Jiang, X., Kang, H., Quarles, C.C., Zu, Z., Gochberg, D.F., Gore,
 J.C., Xu, J., 2016. Influence of water compartmentation and heterogeneous relaxation on
 quantitative magnetization transfer imaging in rodent brain tumors. Magn Reson Med 76, 635 644.
- Lu, L., Erokwu, B., Lee, G., Gulani, V., Griswold, M.A., Dell, K.M., Flask, C.A., 2012. Diffusion-prepared
 fast imaging with steady-state free precession (DP-FISP): a rapid diffusion MRI technique at 7 T.
 Magn Reson Med 68, 868-873.
- Lutti, A., Dick, F., Sereno, M.I., Weiskopf, N., 2014. Using high-resolution quantitative mapping of R1 as an index of cortical myelination. Neuroimage 93 Pt 2, 176-188.
- Lutti, A., Stadler, J., Josephs, O., Windischberger, C., Speck, O., Bernarding, J., Hutton, C., Weiskopf,
 N., 2012. Robust and fast whole brain mapping of the RF transmit field B1 at 7T. PLoS One 7,
 e32379.
- 1157 Ma, D., Gulani, V., Seiberlich, N., Liu, K., Sunshine, J.L., Duerk, J.L., Griswold, M.A., 2013. Magnetic 1158 resonance fingerprinting. Nature 495, 187-192.
- Mansfield, P., Guilfoyle, D.N., Ordidge, R.J., Coupland, R.E., 1986. Measurement of T1 by echo-planar imaging and the construction of computer-generated images. Phys Med Biol 31, 113-124.
- Marques, J.P., Kober, T., Krueger, G., van der Zwaag, W., Van de Moortele, P.F., Gruetter, R., 2010.

 MP2RAGE, a self bias-field corrected sequence for improved segmentation and T1-mapping at high field. Neuroimage 49, 1271-1281.
- Moeller, S., Pisharady, P.K., Ramanna, S., Lenglet, C., Wu, X., Dowdle, L., Yacoub, E., Ugurbil, K.,
 Akcakaya, M., 2021. Noise reduction with Distribution Corrected (NORDIC) PCA in dMRI with
 complex-valued parameter-free locally low-rank processing. Neuroimage 226, 117539.

- Moeller, S., Yacoub, E., Olman, C.A., Auerbach, E., Strupp, J., Harel, N., Ugurbil, K., 2010. Multiband multislice GE-EPI at 7 tesla, with 16-fold acceleration using partial parallel imaging with application to high spatial and temporal whole-brain fMRI. Magn Reson Med 63, 1144-1153.
- Nilsson, M., van Westen, D., Stahlberg, F., Sundgren, P.C., Latt, J., 2013. The role of tissue microstructure and water exchange in biophysical modelling of diffusion in white matter. MAGMA 26, 345-370.
- Norris, D.G., Boyacioglu, R., Schulz, J., Barth, M., Koopmans, P.J., 2014. Application of PINS radiofrequency pulses to reduce power deposition in RARE/turbo spin echo imaging of the human head. Magn Reson Med 71, 44-49.
- 1176 Norris, D.G., Koopmans, P.J., Boyacioglu, R., Barth, M., 2011. Power Independent of Number of Slices 1177 (PINS) radiofrequency pulses for low-power simultaneous multislice excitation. Magn Reson 1178 Med 66, 1234-1240.
- Novikov, D.S., Jensen, J.H., Helpern, J.A., Fieremans, E., 2014. Revealing mesoscopic structural universality with diffusion. Proc Natl Acad Sci U S A 111, 5088-5093.
- O'Brien, K.R., Kober, T., Hagmann, P., Maeder, P., Marques, J., Lazeyras, F., Krueger, G., Roche, A., 2014. Robust T1-weighted structural brain imaging and morphometry at 7T using MP2RAGE. PLoS One 9, e99676.
- Ordidge, R.J., Gibbs, P., Chapman, B., Stehling, M.K., Mansfield, P., 1990. High-speed multislice T1 mapping using inversion-recovery echo-planar imaging. Magn Reson Med 16, 238-245.
 - Panagiotaki, E., Schneider, T., Siow, B., Hall, M.G., Lythgoe, M.F., Alexander, D.C., 2012. Compartment models of the diffusion MR signal in brain white matter: a taxonomy and comparison. Neuroimage 59, 2241-2254.

1187

- Panchuelo, R.M.S., Mougin, O., Turner, R., Francis, S.T., 2021. Quantitative T1 mapping using multislice multi-shot inversion recovery EPI. Neuroimage, 117976.
- Pierpaoli, C., Jezzard, P., Basser, P.J., Barnett, A., Di Chiro, G., 1996. Diffusion tensor MR imaging of the human brain. Radiology 201, 637-648.
- Poser, B.A., Anderson, R.J., Guerin, B., Setsompop, K., Deng, W., Mareyam, A., Serano, P., Wald, L.L., Stenger, V.A., 2014. Simultaneous multislice excitation by parallel transmission. Magn Reson Med 71, 1416-1427.
- Poser, B.A., Norris, D.G., 2009. Investigating the benefits of multi-echo EPI for fMRI at 7 T. Neuroimage 45, 1162-1172.
- Santis, S., Assaf, Y., Evans, C.J., Jones, D.K., 2014. Improved precision in CHARMED assessment of white matter through sampling scheme optimization and model parsimony testing. Magn Reson Med 71, 661-671.
- Setsompop, K., Fan, Q., Stockmann, J., Bilgic, B., Huang, S., Cauley, S.F., Nummenmaa, A., Wang, F.,
 Rathi, Y., Witzel, T., Wald, L.L., 2018. High-resolution in vivo diffusion imaging of the human
 brain with generalized slice dithered enhanced resolution: Simultaneous multislice (gSliderSMS). Magn Reson Med 79, 141-151.
- Setsompop, K., Gagoski, B.A., Polimeni, J.R., Witzel, T., Wedeen, V.J., Wald, L.L., 2012. Blippedcontrolled aliasing in parallel imaging for simultaneous multislice echo planar imaging with reduced g-factor penalty. Magn Reson Med 67, 1210-1224.
- Stuber, C., Morawski, M., Schafer, A., Labadie, C., Wahnert, M., Leuze, C., Streicher, M., Barapatre,
 N., Reimann, K., Geyer, S., Spemann, D., Turner, R., 2014. Myelin and iron concentration in the
 human brain: a quantitative study of MRI contrast. Neuroimage 93 Pt 1, 95-106.
- Tax, C.M.W.R., U.S.; Witzel, T.; Jones, D.K., 2017. Disentangling in two dimensions in the living human brain: Feasbilty of relaxometry-diffusometry using ultra-strong gradients. ISMRM 25th Annual Meeting & Exhibition, Honolulu, HI, USA.
- Teeuwisse, W.M., Brink, W.M., Webb, A.G., 2012. Quantitative assessment of the effects of highpermittivity pads in 7 Tesla MRI of the brain. Magn Reson Med 67, 1285-1293.
- Tournier, J.D., Smith, R., Raffelt, D., Tabbara, R., Dhollander, T., Pietsch, M., Christiaens, D., Jeurissen,
- B., Yeh, C.H., Connelly, A., 2019. MRtrix3: A fast, flexible and open software framework for medical image processing and visualisation. Neuroimage 202, 116137.

- 1219 Veraart, J., Fieremans, E., Novikov, D.S., 2019. On the scaling behavior of water diffusion in human 1220 brain white matter. Neuroimage 185, 379-387.
- 1221 Veraart, J., Novikov, D.S., Christiaens, D., Ades-Aron, B., Sijbers, J., Fieremans, E., 2016. Denoising of diffusion MRI using random matrix theory. Neuroimage 142, 394-406.
- 1223 Veraart, J., Novikov, D.S., Fieremans, E., 2018. TE dependent Diffusion Imaging (TEdDI) distinguishes 1224 between compartmental T2 relaxation times. Neuroimage 182, 360-369.
- Vu, A.T., Auerbach, E., Lenglet, C., Moeller, S., Sotiropoulos, S.N., Jbabdi, S., Andersson, J., Yacoub, E.,
 Ugurbil, K., 2015. High resolution whole brain diffusion imaging at 7T for the Human
 Connectome Project. Neuroimage 122, 318-331.
- Wang, F., Dong, Z., Reese, T.G., Bilgic, B., Katherine Manhard, M., Chen, J., Polimeni, J.R., Wald, L.L.,
 Setsompop, K., 2019. Echo planar time-resolved imaging (EPTI). Magn Reson Med 81, 3599 3615.
- Weiskopf, N., Suckling, J., Williams, G., Correia, M.M., Inkster, B., Tait, R., Ooi, C., Bullmore, E.T., Lutti,
 A., 2013. Quantitative multi-parameter mapping of R1, PD(*), MT, and R2(*) at 3T: a multi center validation. Frontiers in neuroscience 7, 95.
- Wu, H.T., Q.; Poetter, C.; Zhu, K.; Middione, M.; Kerr, A.B.; McNab, J.A.; Dougherty, R., 2017. Whole
 Brain Inversion Recovery Diffusion Weighted Imaging Using Slice-Shuffled Acquisition. ISMRM
 25th Annual Meeting & Exhibition, Honolulu, HI, USA.
- Wu, X., Schmitter, S., Auerbach, E.J., Moeller, S., Ugurbil, K., Van de Moortele, P.F., 2013.
 Simultaneous multislice multiband parallel radiofrequency excitation with independent slice-specific transmit B1 homogenization. Magn Reson Med 70, 630-638.
- Zhang, H., Hubbard, P.L., Parker, G.J., Alexander, D.C., 2011. Axon diameter mapping in the presence
 of orientation dispersion with diffusion MRI. Neuroimage 56, 1301-1315.
- Zhang, H., Schneider, T., Wheeler-Kingshott, C.A., Alexander, D.C., 2012. NODDI: practical in vivo
 neurite orientation dispersion and density imaging of the human brain. Neuroimage 61, 1000 1016.
- Zhang, Y., Wells, S.A., Hernando, D., 2019. Stimulated echo based mapping (STEM) of T1, T2, and
 apparent diffusion coefficient: validation and protocol optimization. Magn Reson Med 81, 167 181.