

1 **Application of Pedimap — a pedigree visualization tool — to facilitate the**
2 **decisioning of rice breeding in Sri Lanka**

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15

16 **ABSTRACT**

17 The development of rice cultivars with desirable traits is essential. The decision-making is a
18 crucial step in rice breeding programs. The breeders can make efficient and pragmatic decisions
19 if an organized pedigree visualization platform is available for the material of the rice breeding
20 germplasm. The staple food in Sri Lanka is rice, and there is a great demand for improved
21 varieties with high yield and other promising traits. In the present study, the available data of
22 all the rice varieties released by Rice Research and Development Institute, Sri Lanka, and the
23 related landraces and genotypes were arranged in Pedimap, a pedigree visualization tool.
24 Pedimap can showcase pedigree relationships, phenotypic, and molecular data. The Identity by
25 Descent (IBD) probabilities were calculated using FlexQTL software and included in the
26 Pedimap database. The parentage selection based on the variations of phenotypic traits,
27 selection of marker alleles for molecular breeding, and detection of the founders of genetic
28 effects can be swiftly conducted using Pedimap. However, the power of harnessing the value
29 of Pedimap for making breeding decisions relies on the availability of data for the traits,
30 markers, and genomic sequences. Thus, it is imperative to characterize the breeding
31 germplasms using standard phenomic and genomic characterization procedures before
32 organized into Pedimap. Thereby, the worldwide breeding programs can benefit from each
33 other to produce improved varieties to meet global challenges.

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35 **Keywords:** Breeding database, Breeding decisions, Marker assisted breeding, Pedigree
36 visualization, Planning crosses

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41 INTRODUCTION

42 Rice is one of the major crops in the world, with an annual production over 700 million metric
43 tons [1]. Half of the world population consumes rice as the staple food [2]. Currently, the
44 demand for rice is rapidly increasing due to the growth of the human population [3]. However,
45 the current rice production cannot meet the increasing demand causing severe food security
46 issues. The biotic and abiotic stresses also exert a negative influence on rice production [4].
47 The rice farming is also a way of living for many people, especially in numerous Asian
48 countries [5]. At present, 1.8 million Sri Lankan families engage in rice farming over 870,000
49 hectares [6]. The annual rice production in Sri Lanka is approximately 2.3 million metric tons
50 (MT), which is insufficient to fulfill the domestic rice demand of 3.0 million [7]. Hence, the
51 Sri Lankan government spends about USD 400 million to import rice annually [7,8].

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53 The rice production is mainly affected by drought and irregular rainfall patterns caused by
54 climate change [9,10,11], adverse soil conditions such as salinity [6,12], and pest and disease
55 attacks [13]. The biotic and abiotic stresses in rice farming can be controlled using numerous
56 agronomic practices such as irrigation, drainage, fertilization, and the application of pesticides.
57 However, the rate of success of the controlling methods is limited [13] due to the unpredictable
58 nature of climate change, soil degradation, variations in pest dynamics, and development of
59 pest resistance [14]. Therefore, breeding is considered as the most successful strategy to
60 produce high yielding and stress resilient rice varieties [15]. The improved rice genotypes can
61 also contain the traits for higher consumer preference and organic farming [16]. In the past, the
62 rice varietal improvement was conducted with classical breeding techniques, which are tedious,
63 lengthy, and less feasible in cases such as breeding for pest resistance and submergence
64 tolerance. However, the marker-assisted breeding (MAB) is employed in modern breeding
65 programs to introgress valuable genetic loci from landraces and traditional varieties [17,18]

66 and the desirable haplotypes of Quantitative Trait Loci (QTL) to the improved rice varieties
67 [19-21].

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69 The decision-making process in a breeding program is crucial for successful outcomes. The
70 formulation of decisions before breeding is a multi-step process that consists of the
71 identification of breeding priorities, determination of the genetics of target traits, and
72 employment of pre-breeding methods if required. The economic and technical feasibility,
73 number of parents for crosses, number of selfing and outcrossing cycles, length of the breeding
74 program/cycles, and identification of the selection methods must also be assessed [22]. In the
75 decisioning process, initially, the market trends based on consumer and other stakeholder
76 preferences must be recognized [23]. Subsequently, the novelty and the uniqueness of the
77 breeding objective must be assessed before the execution of the breeding program [22].

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79 The selection of suitable varieties or individual plants as parents and the determination of the
80 selection methods are the two most critical aspects in planning breeding programs [24]. The
81 parental selection depends on the number of prioritized traits for breeding. When multiple
82 characteristics are to be introgressed, the breeders require a prioritized order of parents for
83 stepwise crossing and selection [25,26]. The decision-making process in breeding is entirely
84 based on the available information on phenotypes, genotypes, pedigree, available budget, field
85 and greenhouse space, desired time-to-market etc. Although the data for decision-making for
86 breeding are indispensable, haphazardly collected information would provide less value to the
87 breeders. In many conventional breeding programs, most of the data are recorded in field
88 notebooks and stored in the breeding stations, while very little information is available as
89 computerized databases. If an organized database containing all the essential information for

90 the rice varieties released and the parental genotypes used in breeding, the decisions can be
91 easily made.

92

93 The construction of a database with all the necessary information from varieties and their
94 parents promotes the capacity of data sharing, mining, visualization, and retrieval [27].

95 Pedimap is a pedigree visualization software. The data needed can be imported to Pedimap

96 from FlexQTL, or with some custom script from any other database program. Pedimap is used

97 by many contemporary plant genetics and breeding programs worldwide. As stated in Voorrips

98 et al. (2012) [28], Pedimap can be used to record and utilize breeding history. Pedimap

99 illustrates the available phenotypic and genetic data through pedigrees. All the information,

100 including parentage, qualitative and quantitative data, marker alleles/genotypes, and the

101 calculated identity-by-descent (IBD) probabilities can be presented in Pedimap. Currently,

102 breeders prefer to use pedigree visualization tools like Pedimap since it allows them to access

103 the large pool of genetic and phenotypic data quickly and generate pedigrees that are essential

104 in making breeding decisions.

105

106 In Sri Lanka, Rice Research and Development Institute (RRDI) is the sole organization

107 conducting the rice breeding programs for the national needs. Therefore, in the present study,

108 we report an attempt to organize the information of the released varieties and the parental

109 genotypes of RRDI breeding programs as a Pedimap based database which is a valuable step

110 to take accurate breeding decisions and speed up the process of releasing novel varieties.

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112

113

114 MATERIALS AND METHODS

115 Data Curation

116 The data were collected from RRDI, Sri Lanka and classified under three main categories,
117 namely pedigree history, phenotypic data, and molecular data on rice
118 varieties/landraces/genotypes (herein after collectively referred to as cultivars). The male and
119 female parents and the order of crosses were taken under pedigree history. The average yield
120 of the rice plants, the maturity period in different growing seasons (*Yala* and *Maha* seasons of
121 Sri Lanka [29]), plant height, basal leaf sheath color and additional color patterns,
122 recommended type of the land, level of phosphorus deficiency tolerance, amount of brown rice
123 recovery, milling recovery, head rice recovery, amylose content, gelatinization temperature,
124 weight of 1000 grains, shape of the grain, pericarp color, weight of a kg, color of the buff coat
125 and resistance/susceptibility to pests and diseases; brown planthopper (BPH), bacterial leaf
126 blight and rice blast disease were recorded under phenotypic data (S1 Table). The available
127 DNA marker alleles, marker positions in the linkage map, and allelic scores were entered under
128 molecular data [30-33] (S2 Table).

129

130 Pedimap Procedure

131 A Pedimap input data file is created in MS Excel (2019), and the data file is exported as a tab-
132 delimited text (.txt) file (S3 Table). The input file contains four main subdivisions; header,
133 pedigree, marker data, and IBD probability section (Fig 1). The header consists of five essential
134 elements and one additional element. The name of the population and symbols for unavailable
135 or missing data, null homozygous alleles, and confirmed null alleles are entered to the pedigree
136 section, as shown in Fig 1A. The name of the cultivar must be a string with text or numerical
137 values without spaces.

138 Next to the header, the pedigree section is entered, as shown in Fig 1B. The first column denotes
139 the name of the variety or landrace, and second and third columns are reserved for maternity
140 and paternity information, respectively. The numbers and strings can be included to represent
141 the phenotypes in the first three columns. From the fourth column onwards, any desirable
142 quantitative or qualitative trait values can be entered. All the collected phenotypic data are
143 introduced, as shown in Fig 1B. The third section of the input data file is for marker
144 information. The linkage group of the DNA marker and the marker positions in the linkage
145 map are entered, as shown in Fig 1C. If there are more than one linkage group, all the linkage
146 group maps should be defined successively before entering the allelic scores. The detailed data
147 for each DNA marker can be inserted after revealing the map positions. The respective number
148 of columns, according to the ploidy level, should be incorporated to enter allelic scores. The
149 fourth section is for IBD probability values (Fig 1C). The IBD probabilities cannot be
150 calculated within Pedimap but can be calculated using other software *e.g.* FlexQTL [34], which
151 is a software for QTL analysis. FlexQTL can also generate a complete Pedimap input data file.

152

153 **Demonstration of the Usability of Pedimap**

154 We used the examples 1 and 2 given in Table 1 to show how parental cultivars can be selected
155 for crossing based on diverse breeding objectives and the prioritized traits. The example 3 in
156 Table 1 was used to select parents, indicate the DNA marker allelic representation for MAB,
157 identity by descent calculations, and planning crosses to deduce related details necessary for
158 decision-making for breeding.

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163 **Table 1. The examples used to demonstrate the use of Pedimap in making breeding decisions.**

Example	Trait*					
	Priority# 1	Priority 2	Priority 3	Priority 4	Priority 5	Priority 6
1	White pericarp	Yield \geq 3.5 mt/ha	Resistance or moderate resistance to brown planthopper (BPH)	Maturity period \leq 125 days	Grain shape [§]	-
2	High, high-intermediate, and intermediate amylose content	Yield \geq 3.5 mt/ha	Maturity period \leq 125 days	Resistance or moderate resistance to blast	-	-
3	Phosphorous deficiency tolerance	Yield \geq 5.0 mt/ha	Maturity period 90-105 days	Resistance or moderate resistance to BPH	Resistance or moderate resistance to blast	High, high-intermediate, and intermediate amylose content

164 *The trait classes and records are from RRDI records [35].

165 #The traits are given in the order of priority in making breeding decisions.

166 [§]Variable grain shapes of the intended varieties to be released.

167
168 **Fig 1** The input data file structure of the Pedimap; The input file was created as an MS Excel
169 worksheet, contains four main sections. A: Header, B: Pedigree and phenotypic data, C:
170 Genotypic data. A: In the header section, essential elements are highlighted in blue, which
171 contains the population name, ploidy and codes used in the data. (i): abbreviations for missing
172 data (i.e., unknown), possible null alleles, confirmed null alleles; (ii): NALLELES is only
173 necessary if the IBD probabilities are used, and specifies the total number of founder alleles
174 (i.e. the number of founder times the ploidy). B: The Pedigree section contains the pedigree
175 data of all the individuals, and any phenotypic data of the individuals. The pedigree part is
176 highlighted in purple. (iii): founders (initial parents) are entered with missing values for their
177 parents. Phenotypic data are entered in subsequent columns (iv). C: The Genotypic data section
178 (if present) is divided into three parts: one part for each linkage group the genetic map (v),
179 general information per locus (vi) and positions where IBD probabilities are calculated (vii); a
180 part with the observed alleles per locus per individual (viii), and a part with the Identity-by
181 Descent (IBD) probabilities per position per individual (ix). The final file must be saved as a
182 text (.txt) file.

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189 RESULTS AND DISCUSSION

190 Worldwide plant genetics and breeding programs use Pedimap as the platform for maintaining
191 breeding databases and pedigree visualization. In the RosBREED project [36], the parental and
192 progeny identification, tracing founders, and calculation of allelic representation are conducted
193 using Pedimap. The pedigree display of Pedimap is used to plan crosses in the Rosaceae
194 research community [37, 38], HIDRAS project [39] and visualize of *Arabidopsis thaliana*
195 crosses [40]. Selecting parentage, sketching out crossing schemes, estimating the probability
196 of allelic segregation, and choosing compatible molecular markers for MAB can be achieved
197 using Pedimap [28]. The use of Pedimap as a pedigree visualization tool for the decision-
198 making process in rice breeding is described using three examples (Table 1).

199

200 **Example 1: Selecting parents for higher yield, BPH tolerance, short duration and white** 201 **pericarp with diverse grain shapes**

202 The Pedimap database rice breeding gerplasm in has a total of 224 input cultivars. There are
203 36 intermediate genotypes such as F1 and F2 that were not reported, but we included them to
204 complete the pedigree in Pedimap. Thus, the database has a total of 188 rice cultivars and
205 accessions with known identities with records (S1 Fig, S1 Table). In Example 1, we considered
206 a scheme to select accessions as parents with the parameters given in Table 1 for white pericarp,
207 yield, BPH resistance, maturity period, and the grain shape. These thresholds defined a
208 subpopulation of 26 cultivars (Fig 2). The variation of the yield is given in Fig 2A. According
209 to the color shading given, the breeder can select the required parents for crossing to obtain
210 higher yield levels. However, as shown in Fig 2B, only three cultivars show the complete
211 resistance to BPH. If breeder plans to introgress the complete BPH resistance to the novel
212 varieties, only Bg250, At307, and At306 are available as the sources of resistance. Fig 2C
213 displays the variation for the maturity period. The breeder can choose the parents depending

214 on his objective for the intended maturity period for the novel varieties. Example 1 was
215 exclusively planned to breed for white pericarp. However, the grain shape is also important as
216 a significant quality trait to become a successful variety in the market. Fig 2D shows the
217 variation for grain shapes for the breeder to carry out the selection. If we consider all the traits
218 and selected At307 as a parent based on the pedigree visualization in Pedimap, At307 can
219 provide the genetic basis for high yield, complete resistance to BPH, approximately three
220 months for maturity and intermediate-bold shaped grains. If Bg450 was selected, the yield is
221 still in the higher range with moderate resistance for BPH and short-round grains. However,
222 Bg450 brings the alleles for an extended maturity period (Fig 2).

223

224 **Fig 2** The pedigree visualization for Example 1 (Parents with white pericarp, yield ≥ 3.5 mt/ha,
225 moderate or complete BPH resistance, maturity period ≤ 125 days, and diverse grain shapes).
226 The selected pedigree is colored separately for four traits. A: Yield; B: Degree of resistance to
227 brown planthopper (BPH); C: Maturity period; D: Grain shape. Female and male parentages
228 are indicated by red and purple lines, respectively. The symbol ‘×’ indicates the cross between
229 two parents. The background colors of the cultivar-name boxes indicate the trait values, as
230 shown in the colored legends below.

231

232 **Example 2: Selecting parents for high/high-intermediate amylose content, higher yield,**
233 **short duration, and resistance to blast disease**

234 In Example 2, we considered a scheme to select cultivars/accessions as parents with the
235 parameters given in Table 1 for high/high-intermediate amylose content, higher yield, short
236 duration, and resistance to blast disease. These thresholds defined a subpopulation of 37
237 cultivars/accessions (Fig 3). The breeder can select the high yielding, short-duration, and blast-
238 resistant cultivars as parents from pedigrees visualized in Figs 3A, 3B, and 3C, respectively.
239 The high, high-intermediate, and intermediate amylose contents are depicted in the pedigree
240 given in Fig 3D. Only Bw351, At307, Bg407H, At308, and Bg252 show the complete
241 resistance to blast (Fig 3C). However, At307 is the most promising parent with high yield (Fig
242 3A), short duration (Fig 3B), and high amylose content (Fig 3D) along with complete resistance

243 to blast (Fig 3C). Also, Bg407H is the highest yielding (Fig 3A), blast-resistant (Fig 3C), and
244 high in amylose content (Fig 3D). However, Bg407H is a long duration variety compared to
245 At307. Therefore, the breeder may plan to cross At307 and Bg407H to accomplish the breeding
246 objective of Example 2.

247

248 **Fig 3** The pedigree visualization for Example 2 (parents with high, high-intermediate, and
249 intermediate amylose content, yield ≥ 3.5 mt/ha, moderate or complete resistance to rice blast
250 disease and maturity period ≤ 125 days). The selected pedigree is colored separately for four
251 traits. A: Yield; B: Maturity period; C: Degree of resistance to rice blast disease; D: Amylose
252 content. Female and male parentages are indicated by red and purple lines, respectively. The
253 symbol ‘×’ indicates the cross between two parents, and ‘×’ inside the circle represents selfing.
254 The background colors of the cultivar-name boxes indicate the trait values, as shown in the
255 colored legends below.

256

257 **Example 3: Selecting parents for phosphorus deficiency tolerance, higher yield, short**
258 **duration, resistance to both BPH and blast, and high/intermediate-high amylose content**

259 We selected a set of rice cultivars from the Pedimap database based on the availability of ranked
260 scores for phosphorus deficiency tolerance (PDT). Twenty-four cultivars contain the PDT
261 ranks of high, moderate, and sensitive (Fig 4A). The same set was illustrated using Pedimap
262 for yield (Fig 4B), maturity period (Fig 4C), degree of resistance to BPH (Fig 4D) and blast
263 (Fig 4E), and amylose content (Fig 4F). If At362 is considered as a parent, it can bring
264 resistance to PD, and BPH, moderate resistance to blast, high yield, average maturity period,
265 and intermediate-high amylose content. Similarly, if Bg250 is selected, it can bring moderate
266 resistance to PD and blast, resistance to BPH, moderate yield and shortest maturity period, and
267 high amylose content (Fig 4).

268

269 A sample crossing scheme is shown in Fig 5 to produce a rice variety with high PDT, mean
270 yield ≥ 5.0 mt/ha, maturity period ≤ 105 days, resistant to BPH and blast disease, and higher
271 amylose content. Since there is no reported cultivar for high PDT with complete blast resistance

272 (Fig 4), the illustrated crossing scheme in Fig 5 is proposed with two phases. In the first phase,
273 the crossing of At362 and Bg250 followed by numerous rounds of selfing and selection of the
274 most beneficial lines among the RILs at advanced generations would accomplish the breeding
275 objective only without complete resistance to blast (i.e., a moderate level of blast resistance is
276 possible). In the second phase, the selected RILs from phase 1 can be backcrossed to Bg252 as
277 the donor parent to introgress the complete resistance to blast. The breeder can come up with
278 diverse crossing schemes like the one given in Fig 5 to make effective decisions for breeding
279 and maximize the resource utilization to release varieties in the shortest possible time. The
280 breeder can select any number of parents that are needed to use as sources of resistance and
281 other traits to start crossing. Also, the marker alleles and the IBD probabilities can be checked
282 as illustrated in S2A Fig and S2B Fig respectively.

283

284 **Fig 4** The pedigree visualization for Example 3 (parents ranked for phosphorus deficiency
285 tolerance). The selected pedigree is colored separately for six traits. A: PDT; B: Yield; C:
286 Maturity period; D: Degree of resistance to BPH; E: Degree of resistance to BLAST; F:
287 Amylose content. Female and male parentages are indicated by red and purple lines,
288 respectively. The symbol ‘×’ indicates the cross between two parents. The background colors
289 of the cultivar-name boxes indicate the trait values, as shown in the colored legends below. The
290 cultivars with missing-trait values are indicated by white boxes.

291

292 **Fig 5** The pedigree visualization for planning a crossing scheme. Phase 1: Initial crossing of
293 At362 and Bg250 and pedigree selection to obtain RILs with ≥ 5.0 mt/ha of mean yield, ≤ 105
294 day of maturity period, resistant to BPH, moderately resistant to blast and high level of amylose
295 content. Phase 2: Then backcrossing with Bg252 as the donor parent to introgress the blast
296 resistance.

297

298 The decision-making process in breeding is a tedious task [22]. The breeding germplasm is
299 complex with large numbers of improved varieties, traditional cultivars, landraces, wild
300 germplasm and accessions. Also, there can be large mapping populations and unreleased
301 varieties due to various reasons. The numerous genotypes in breeding germplasm may have
302 extensive records on agronomic data, pest and disease resistance, quality traits, availability of
303 samples, geographic locations, and utilization in diverse breeding programs as parents [41, 42].

304 With the advent of DNA markers and sequencing technologies, a wealth of genomic
305 information is also available [43]. However, one of the recurrent problems in any breeding
306 germplasm in the world is most of the cultivars remain uncharacterized. Thus, they cannot be
307 used directly in breeding activities. Traditionally, breeders keep records in field books. With
308 the development of computer technology, data tabulation is becoming a common practice.
309 However, given the highly complex nature of the datasets in breeding germplasm, data tables
310 have a limited value to the breeders. The tables created with contemporary data managing
311 software cannot graphically display complex pedigrees and variations of qualitative and
312 quantitative traits along with DNA marker information. These database handling platforms do
313 not make use of the pedigree-based capabilities of Pedimap, like selecting related parental
314 varieties/accessions. In this context, Pedimap provides a considerable advantage, as it can
315 visualize pedigree relationships, trait variations, and any other useful information required for
316 decision-making and planning crosses in breeding programs [28]. If all the available details on
317 breeding germplasm are arranged as a database, the breeder can come up with subpopulations
318 based on diverse traits and select the parents for improving multiple traits. However, simple
319 spreadsheets or manually prepared note pages cannot be used to visualize the essential
320 information and complex pedigrees. Breeding programs often suffer a lot when the breeder
321 gets retired or moved to a different position [44-46]. The newly hired breeder cannot practically
322 go through the individual records of the existing breeding germplasm. Thus, there is a strong
323 possibility that valuable breeding germplasm might get lost wasting time, resources, and
324 courage of the retired breeder and his team. However, as a routine practice, if the breeder
325 maintains and updates a Pedimap file for the developing germplasm of breeding materials, the
326 newly hired breeders can go through and identify the value and gaps in the available material
327 for him to plan further. The creation of a Pedimap file is simple, and a novice to informatics
328 can curate and use Pedimap with a little training. Pedimap allows breeders to store data, fetch

329 and visualize genomic information at any time with less effort and complete accuracy [47]. The
330 straightforward accessibility, direct data interpretation, ability to customize the views in
331 multiple fashions, and editable output file formats are the significant features of Pedimap. The
332 graphic files created can be readily imported to image editing software for further visualizations
333 and illustrations. Pedimap is not an opensource software but can be freely obtained by
334 contacting the developers thus even the breeders in developing countries can benefit from
335 Pedimap [28].

336

337 In the current study, we created a Pedimap database for the rice cultivars and accessions
338 prominently used by breeding programs in Sri Lanka. With the available information,
339 significant breeding decisions can be made as we explain in three examples (Figs 2-5).
340 However, it is essential to characterize the cultivars for all the important traits, molecular
341 markers and SNP haplotypes [48], so that breeding decisions can be effectively made [17]. The
342 phenotyping methods must be standard and should follow common procedures across different
343 locations so that the power of the Pedimap database would go up dramatically. Therefore,
344 breeders should always follow the standard, globally acceptable phenomic platforms to
345 characterize the material in breeding germplasm [39, 49].

346

347 **CONCLUSION**

348 The pedigree visualization with variations of phenotypic and molecular data using Pedimap is
349 a user-friendly tool to plan rice breeding programs with higher accuracy and resource
350 optimization. The present study explains the applicability of Pedimap as a decision-making
351 tool to streamline the rice breeding programs in Sri Lanka. However, it is also important to
352 note that accurate characterization of the breeding germplasm for phenotypic and molecular
353 data is the critical prior step to harness the value of Pedimap for breeding.

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489

490 **Supporting information**

491 **S1 Table** Varietal data

492 **S2 Table** Marker data

493 **S3 Table** Pedimap input data file

494 **S1 Fig** Visualization of the entire pedigree of the rice cultivars in the rice breeding germplasm
495 of Sri Lanka. Female and male parentages are indicated by red and purple lines, respectively.
496 The symbol ‘×’ indicates the cross between two parents, and ‘×’ inside the circle represents
497 selfing.

498 **S2 Fig** Visualization of selected marker genotypes and Identity by Descent (IBD). A: Marker
499 alleles. The alleles of the DNA markers K29-N, K41, K48, and K5-N are given in vertical

500 order.; B: IBD probabilities of four Pup1 linked markers. The same pedigree as in the S2A Fig
501 is shown with precalculated IBD data for part of the Pup1 linked region on chromosome 12 at
502 about 55 cM. Since the cultivar linkage maps are not available, we assumed a 0.1 cM gap
503 between adjacent markers for the representation of IBD values. Each color represents a different
504 founder, haplotype. Each rectangle represents one copy of the selected chromosome in an
505 individual. The chromosomal position of the alleles represents the vertical bars, and the width
506 of a color bar indicates the IBD probability of the corresponding founder alleles.

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A

	A	B	C	D	E	F	G	H	I	J	K	L	M
1													
2	POPULATION	=	Sri_Lanka_Rice_Germplasm										
3	UNKNOWN	=	-										
4	NULLHOMOZ	=	\$										
5	CONFIRMEDNULL	=	\$\$										
6	PLOIDY	=	2										
7	NALLELES	=	6										

B

	A	B	C	D	E	F	G	H	I	J	K	L	M
9	PEDIGREE												
10	NAME	PARENT1	PARENT2	Yield	Maturity	Height	Leaf_color	BPH	GM	BL	PDT		
11	Bg94_1	-	-	4.1	105	55	Green	S	S	MS	High		
12	Pokkali	-	-	-	-	-	-	-	-	-	Moderate		
13	Bg300	-	-	5	93	72.2	Green	MR	R	MR	Sensitive		
14	Bg301	-	-	6	93	67	Green	-	-	-	-		
15	At354	Bg94_1	Pokkali	6.5	95	67	Green	MR	MR	S	Sensitive		
16	At401	Bg94_1	Pokkali	5	115	85	Green	MR	R	MS	-		

C

	A	B	C	D	E	F	G	H	I	J	K	L	M
18	LINKAGEGROUP 12												
19	MAP												
20	RM101	48.2											
21	RM277	62											
22													
23	LOCUS	RM101											
24	ALLELENAMES	110	115	120	125								
25	FOUNDERALLELES	110	110	110	110	115	120						
26													
27	LOCUS	RM277											
28	ALLELENAMES	200	225	250	275	300							
29	FOUNDERALLELES	200	200	200	200	300	275						
30	IBDPOSITIONS	48.2	62										
31													
32	ALLELES	RM323											
33	Bg941	140	0	140	0								
34	Pokkali	160	0	180	0								
35													
36	IBDPOSITION	48.6											
37													
38	Bg941	1	0	0	0	0	0	0	0	0	1		
39	Pokkali	0	0	1	0	0	0	0	0	0	0		
40	At354	0	0	0	0	0	0	0	0	1	0		
41	At401	0	0	0	0	0	0	0	0	0	0		
42													
43	IBDPOSITION	62.0											

Define Linkage group

Figure 1

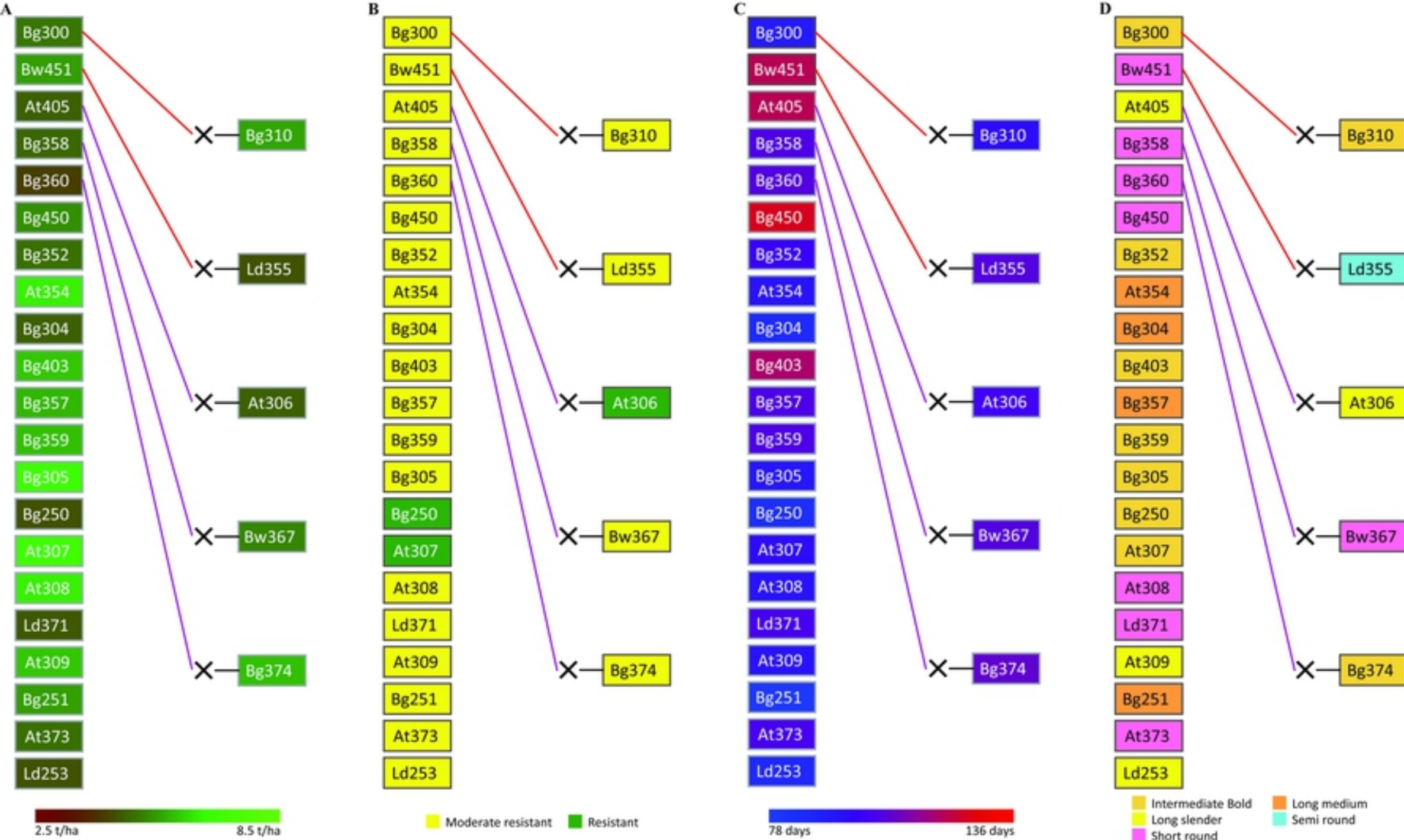


Figure 2

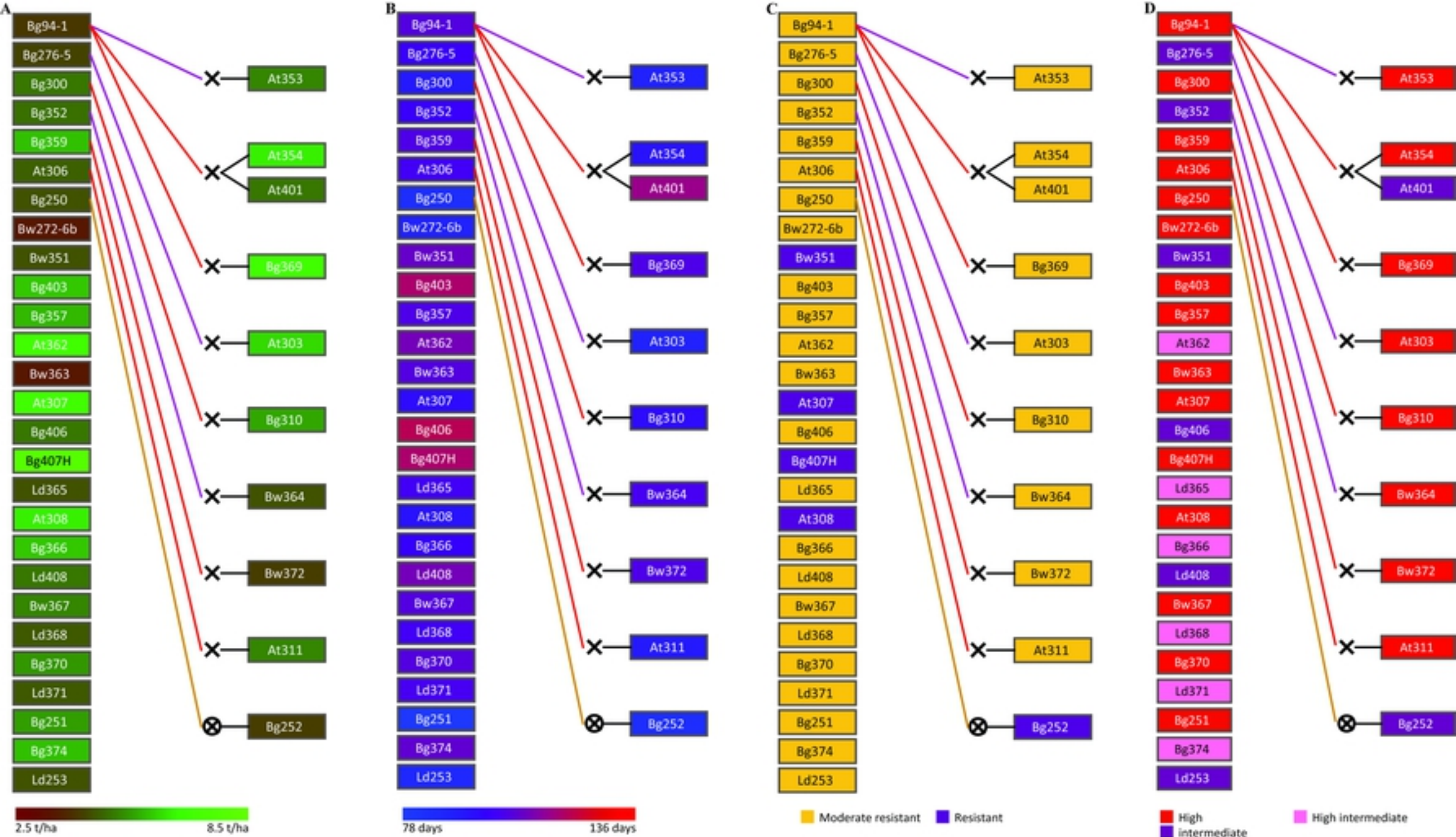
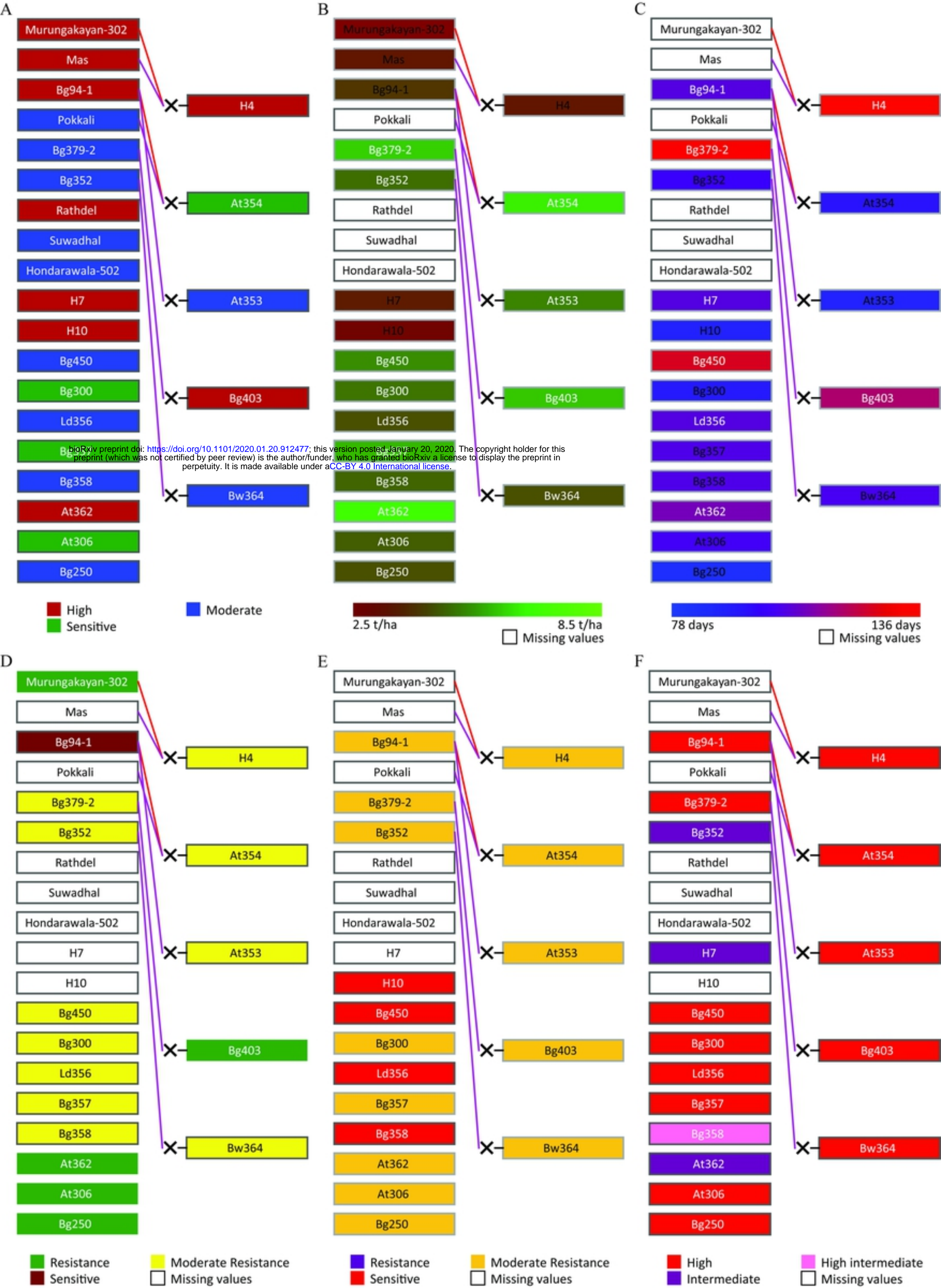


Figure 3



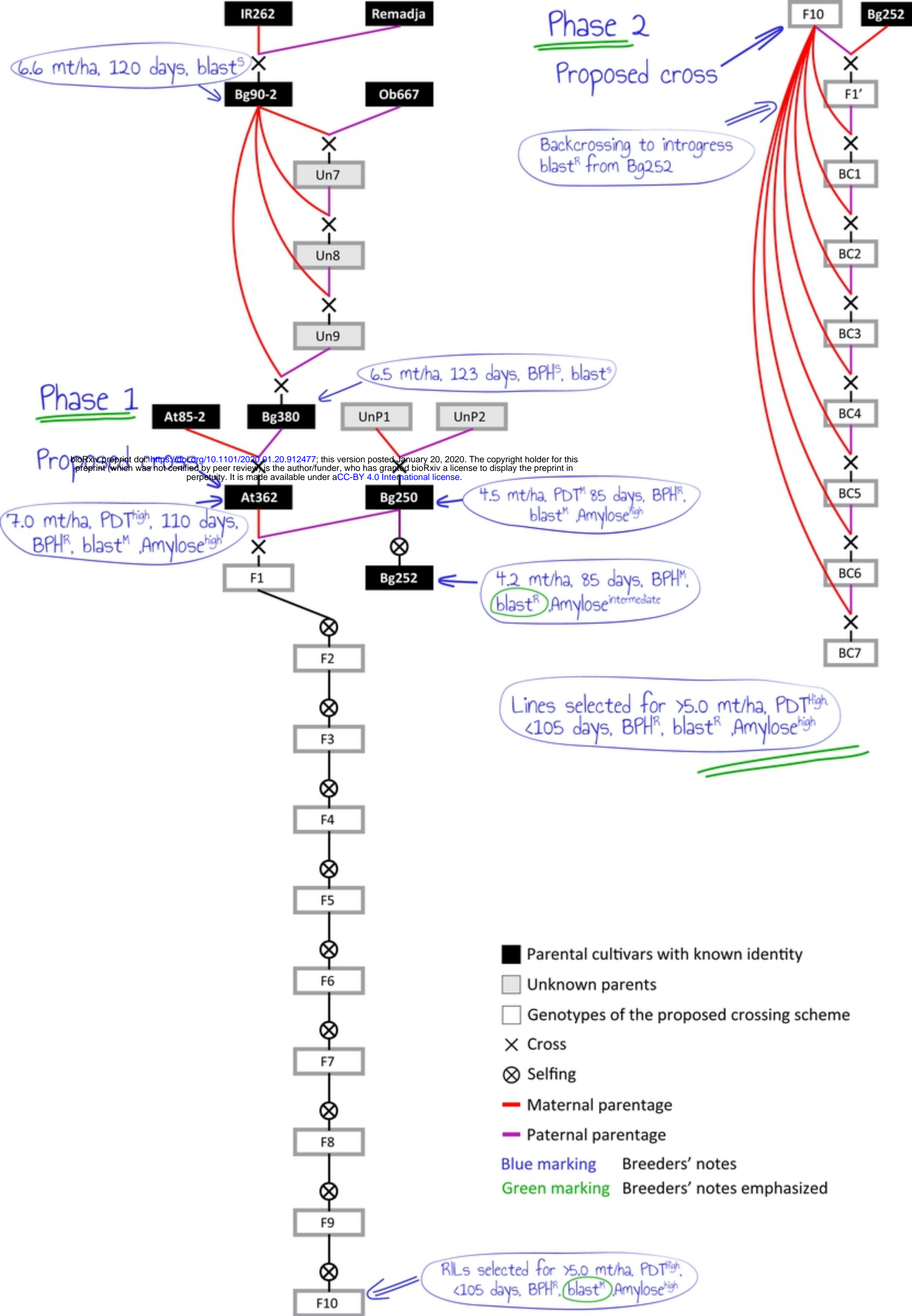


Figure 5