

Stochastic microbial community assembly decreases biogeochemical function

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Abstract: Much research has focused on improving ecosystem models by incorporating microbial regulation of biogeochemistry. However, models still struggle to predict biogeochemical function in future scenarios linked to accelerating global environmental change. Ecological mechanisms may influence the relationship between microbial communities and biogeochemistry, and here, we show that stochastic dispersal processes (e.g., wind-driven or hydrologic transport) can suppress biogeochemical function. Microbial communities are assembled by deterministic (e.g., selection) and stochastic (e.g., dispersal) processes, and the balance of these two processes is hypothesized to influence how microbial communities correspond to biogeochemical function. We explore the theoretical basis for this hypothesis and use ecological simulation models to demonstrate potential influences of assembly processes on ecosystem function. We assemble ‘receiving’ communities under different levels of dispersal from a source community (selection-only, moderate dispersal, and homogenizing dispersal). We then calculate the degree to which assembled individuals are adapted to their environment and relate the level of adaptation to biogeochemical function. We also use ecological null models to further link assembly the level of deterministic assembly to function. We find that dispersal can decrease biogeochemical function by increasing the proportion of maladapted taxa, outweighing selection. The niche breadth of taxa is also a key determinant of biogeochemical function, suggesting a tradeoff between the function of generalist and specialist species. Together, our results highlight the importance of considering ecological assembly processes to reduce uncertainty in predictions of biogeochemical cycles under future environmental scenarios.

Keywords: stochasticity, determinism, microbial ecology, biogeochemistry, null model, ecosystem function

1. Introduction

Environmental systems are undergoing rapid changes in the wake of heightened anthropogenic activity. Predictive ecosystem models have been developed and refined since the 1980s to forecast how these changes may influence future ecosystem health [e.g., 1,2]. While such models originally focused on inferring biogeochemical process rates from substrate pool sizes, recent efforts have improved forecasts by including properties of the microbial communities that mediate these cycles, such as microbial biomass [3-6], dormancy [7], carbon use efficiency [8], and community structure or functional traits [3,9,10]. Yet, these models still struggle to accurately represent biogeochemical cycling in dynamic natural ecosystems, and scientists have called for a better understanding of microbial ecology to increase predictive ability [11-13]. As models begin to consider microbial ecology, there is a need to decipher linkages among spatiotemporal microbial processes and ecosystem-level biogeochemical function. Understanding these relationships will help us reduce uncertainty in future ecosystem functioning [11,12,14-17].

Recent attempts to link microbial communities and ecosystem-level processes have yielded mixed results [12,13,18-21]. A common theme of this research is that microbial community structure reflects the aggregation of historical processes, and such processes may in turn influence the function of a community [13,14,22-25]. For instance, Evans and Wallenstein [22] demonstrate that precipitation history constrains microbial biogeochemical responses to drying and rewetting cycles. Likewise, Fukami *et al.* [26] show legacy effects of microbial colonization history on decomposer communities, and historical dispersal events may be a driver of community function in certain circumstances [27,28].

Several researchers have proposed that the inclusion of community assembly mechanisms, such as dispersal and selection, may enhance our predictions of microbially-mediated biogeochemical cycling [13,29-31]. These processes operate through space and time to determine microbial community composition [12,23,29,32]. They fall into two predominate categories that can be summarized as correlated (i.e., deterministic) or uncorrelated (i.e., stochastic) to environmental conditions. Stochastic processes can be further classified into dispersal, evolutionary diversification, and ecological drift, while determinism is largely dictated by selection [29,33]. Experimental research has shown unpredictable relationships between microbial diversity and ecosystem function, leading to the hypothesis that differences in community assembly history—and thus the relative contributions of stochastic and deterministic processes—drives relationships between microbial community structure and biogeochemical function [30,31].

In particular, many systems have high rates of dispersal that may decrease community-level biogeochemical function. Aquatic systems such as streams, rivers, and marine communities experience hydrologic mixing that disperses microorganisms into new environments [34-36]. In the terrestrial biosphere, snowmelt and rainfall events facilitate hydrologic connectivity in soils [37,38] that can transport organisms, and wind can play a large role in dispersing spore-forming microorganisms [39-41]. High rates of dispersal can add organisms to a microbial community that are not well-suited to local environmental conditions, decreasing the community's overall efficiency of biogeochemical cycling. For example, pH [42] and salinity [43,44] are widely considered strong environmental filters on microbial community structure. If microorganisms disperse from a moderate pH or salinity environment to a more extreme environment, they may be maladapted and have to expend energy to express traits that maintain neutral internal pH (e.g., H⁺ pumps) or maintain cellular water content (e.g., osmotic stress factors). These cell maintenance activities detract from the energy available to transform biogeochemical constituents and may suppress overall community rates of biogeochemical function. In contrast, locally adapted species would putatively have more efficient mechanisms for cell maintenance in the local environment and possess more expendable energy for catalyzing biogeochemical reactions.

The cumulative impacts of ecological processes through time and how they relate to ecosystem-level processes is an emerging research frontier in ecosystem science [13,14,22,24]. Hawkes and Keitt [14] demonstrate through simplified ecological models that historical conditions can constrain the response of a microbial community to an environmental perturbation. They lay a solid theoretical framework in which community-level microbial functions are the accretion of individual life-histories that determine population growth, composition, and fitness. However, they acknowledge their exclusion of dispersal processes from their models and do not explicitly consider stochasticity in their analysis. Hawkes and Keitt [14] therefore provide a foundation for future research and call for a holistic understanding of historical processes on microbial function, with a particular emphasis on the underlying mechanisms generating these trends.

Building on the work of Hawkes and Keitt [14], we hypothesize that stochastic assembly processes strongly influence biogeochemical function. Our aim here is to formalize this hypothesis and provide a simulation-based demonstration of how stochastic assembly can influence function. To do so, we employ ecological simulation models to explicitly represent dispersal and selection-based processes, and we link the resulting communities to biogeochemical function through organismal adaptation.

2. Materials and Methods

All simulations, null models, statistical analyses, and graphics were completed in *R* software (<https://cran.r-project.org/>). The simulation models consisted of two parts and were followed by statistical analysis. One hundred replicates were run for each parameter combination in the simulation models.

First, a regional species was constructed following the protocol outlined in Stegen *et al.* [32]. Regional species pools were constructed by simulating diversification in which new species arise through mutations in the environmental optima of ancestral organisms. Environmental optima evolve along an arbitrary continuum from 0 to 1, following a Brownian process. Regional species pools reach equilibria according to the constraints described by Stegen *et al.* [32] and Hurlbert and Stegen [45] and summarized here: (1) we define a maximum number of total individuals in the pool ($2E+06$) such that the population size of a given species declines with an increasing number of species, and (2) the probability of extinction for a given species increases as its population size decreases according to a negative exponential function [population extinction probability $\propto \exp(-0.001 \cdot \text{population size})$].

The evolution of a regional species pool was initiated from a single ancestor with randomly chosen environmental optimum (initially comprising all $2E+06$ individuals in the population). Mutation probability was set as $1.00E-05$. A descendant's environmental optimum deviated from its ancestor by a quantity selected from a Gaussian distribution with mean 0 and SD 0.2. Following mutation, populations sizes were reduced evenly such that the total number of individuals remained 2 million. The simulation was run for 250 time steps, which was sufficient to reach equilibrium species richness.

The model's second component assembled 4 local communities from the regional species pool according to scenarios conceptualized to test our hypotheses. Species were drawn from the regional species pool to generate a source community under weak selection and three receiving communities with no dispersal, moderate dispersal, and high dispersal in which the strength of selection/niche breadth and environmental conditions were allowed to vary across simulations. All communities had 100 species and 10,000 individuals, drawn probabilistically from the regional species pool. Selection probabilities of species from the regional pool were set by a Gaussian function with standard deviation set as the 'niche breadth' and the deviation of each species optimum from the defined environment per the following equation:

$$SP = e^{-\frac{d^2 \sqrt{2n\pi}}{2n}}, \quad (1)$$

where SP is the selection probability, d is the deviation from the environmental optima, and n is the niche breadth.

In the source community, we used one niche breadth for all simulations, which was the maximum value used for receiving communities (0.175). This value represents weak selection, which allows for assembly of a broader range of species than under strong selection. The environmental conditions in the source community was also set to a single value using the following procedure: We generated 10 regional species pools and combined species abundances and environmental optima from these pools to generate one aggregate pool representative of the probable distributions of environmental optima yielded by our simulations. We set the environmental optimum of the source community to one end of this spectrum (5th percentile) to allow for comparisons with receiving communities that had the same or larger environmental values. This maximized the range of environmental differences between the source and receiving communities.

For receiving communities, we allowed the environmental optimum and niche breadth to vary across simulations. Environmental optima ranged from 0.05 to 0.95 by intervals of 0.04736842 to yield 20 conditions. Niche breadth ranged from 0.0075 to 0.175 by 0.008815789 to yield 20 conditions. Receiving communities were assembled under all possible combinations of environmental optima and niche breadths. Communities for our selection-only case were assembled based only on the selection

probabilities as defined by equation 1, using the same approach as for the assembly of the source community. For moderate and homogenizing dispersal, we modified selection probabilities to incorporate species dispersing from the source community as defined by the following equation:

$$SP_{\text{disp}} = SP + 0.05(S_{\text{source}}^D), \quad (2)$$

where SP_{disp} is the selection probability accounting for dispersal, S_{source} is species abundances in the source community, and D a parameter reflecting dispersal rate. Parameter D was set to 1 for moderate dispersal and 2 for homogenizing dispersal. All possible communities were simulated with 100 replicate regional species pools such that all possible combination of parameters were used once with each regional species pool. Biogeochemical function was calculated for all communities as the sum of species abundances multiplied by their selection probabilities.

Following the assembly of communities, the role of selection in structuring communities was determined via null models developed by Stegen *et al.* [32,46]. Each receiving community was compared to an associated source community that was assembled from the same regional species pool. The approach uses pairwise phylogenetic turnover between communities, calculated using the mean-nearest-taxon-distance (βMNTD) metric [47,48], to infer the relative influence of selection. Communities were evaluated for significantly less turnover than expected ($\beta\text{NTI} < -2$, homogeneous selection) or more turnover than expected ($\beta\text{NTI} > 2$, variable selection) by comparing observed βMNTD values to the mean of a null distribution of βMNTD values – and normalizing by its standard deviation – to yield the β -nearest taxon index (βNTI) [49]. Significance levels for βNTI are based on standard deviations: $|\beta\text{NTI}| = 2$ denotes two standard deviations from the mean of the null distribution. Inferences from βNTI have previously been shown to be robust [13,23,32,50].

Finally, we analyzed differences in model outputs using standard statistical approaches. We used pairwise Kolmogorov-Smirnov tests to compare distributions of species optima between simulations. To compare biogeochemical function of the three dispersal cases, we used ANOVA followed by post-hoc Tukey's HSD tests. We also analyzed changes in biogeochemical function across changes in the environmental parameter using quadratic regressions. Finally, correlations between βNTI and function were assessed with linear regression.

3. Results and Discussion

As ecosystem process models become more sophisticated [e.g., 7,9,11], there is a need to improve these models by better understanding the linkages among spatiotemporal microbial processes and ecosystem-level functions. Here, we use ecological simulation models to demonstrate the importance of stochastic microbial assembly in predicting future biogeochemical functioning. We propose that (1) stochastically assembled communities are composed of species that are not well adapted to their environment and, in turn, that (2) stochastic assembly processes decrease biogeochemical function (Figure 1).

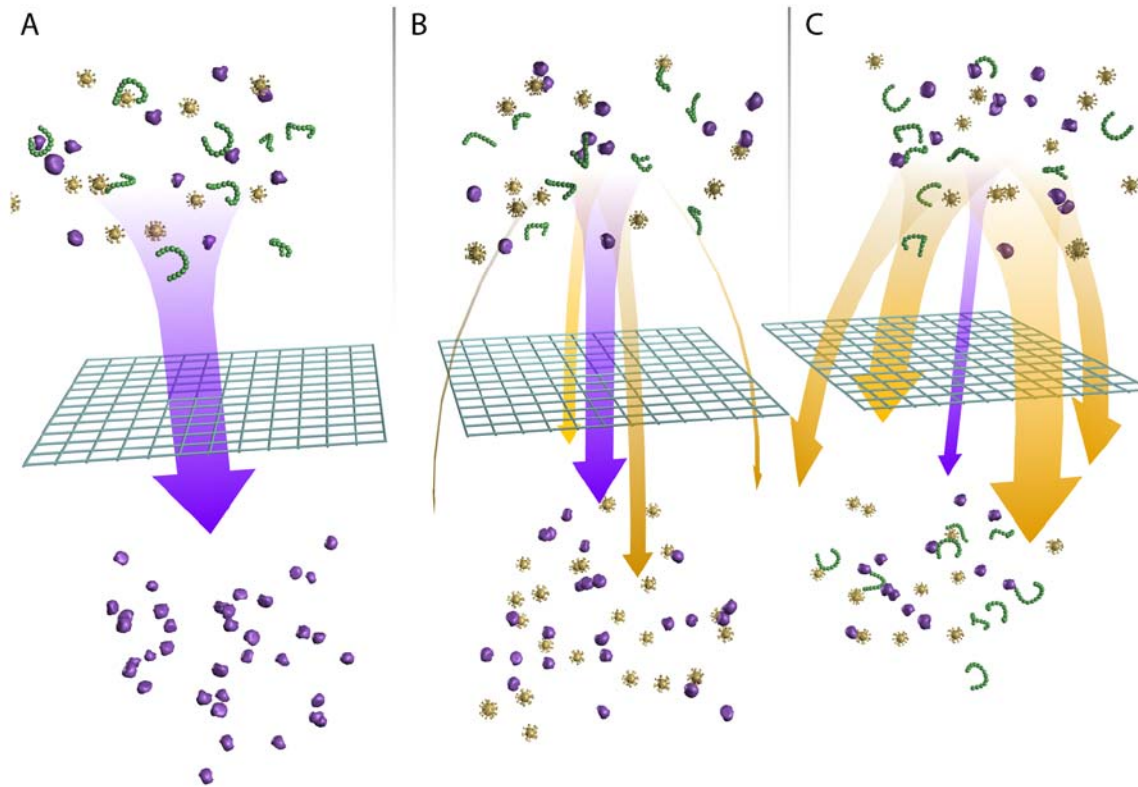


Figure 1. We propose a conceptual model in which stochastic assembly processes decrease biogeochemical function. (A) In a community structured entirely by determinism, environmental filtering restricts community composition to species that are well-adapted to prevailing conditions, resulting in enhanced biogeochemical function. (B) In communities with moderate stochasticity (conceptualized here as dispersal), environmental filters are weaker, and stochasticity increases the abundance of maladapted organisms in the community. In turn, the community is less efficient and exhibits lower biogeochemical function. (C) Under high levels of stochasticity, environmental filters are minimal, allowing for the assembly of a heterogeneous community. The community is unsuited to the environment and mediates low rates of biogeochemical function.

We found that microbial community assembly history impacted biogeochemistry by altering the fitness of organisms to a given environment. The niche optima of taxa in selection-only communities more closely matched their simulated environmental conditions compared to communities assembled with dispersal (Figure 2, $p < 0.001$). These selection-only communities also experienced the highest rates of biogeochemical function on average (Figure 3a, $p < 0.0001$).

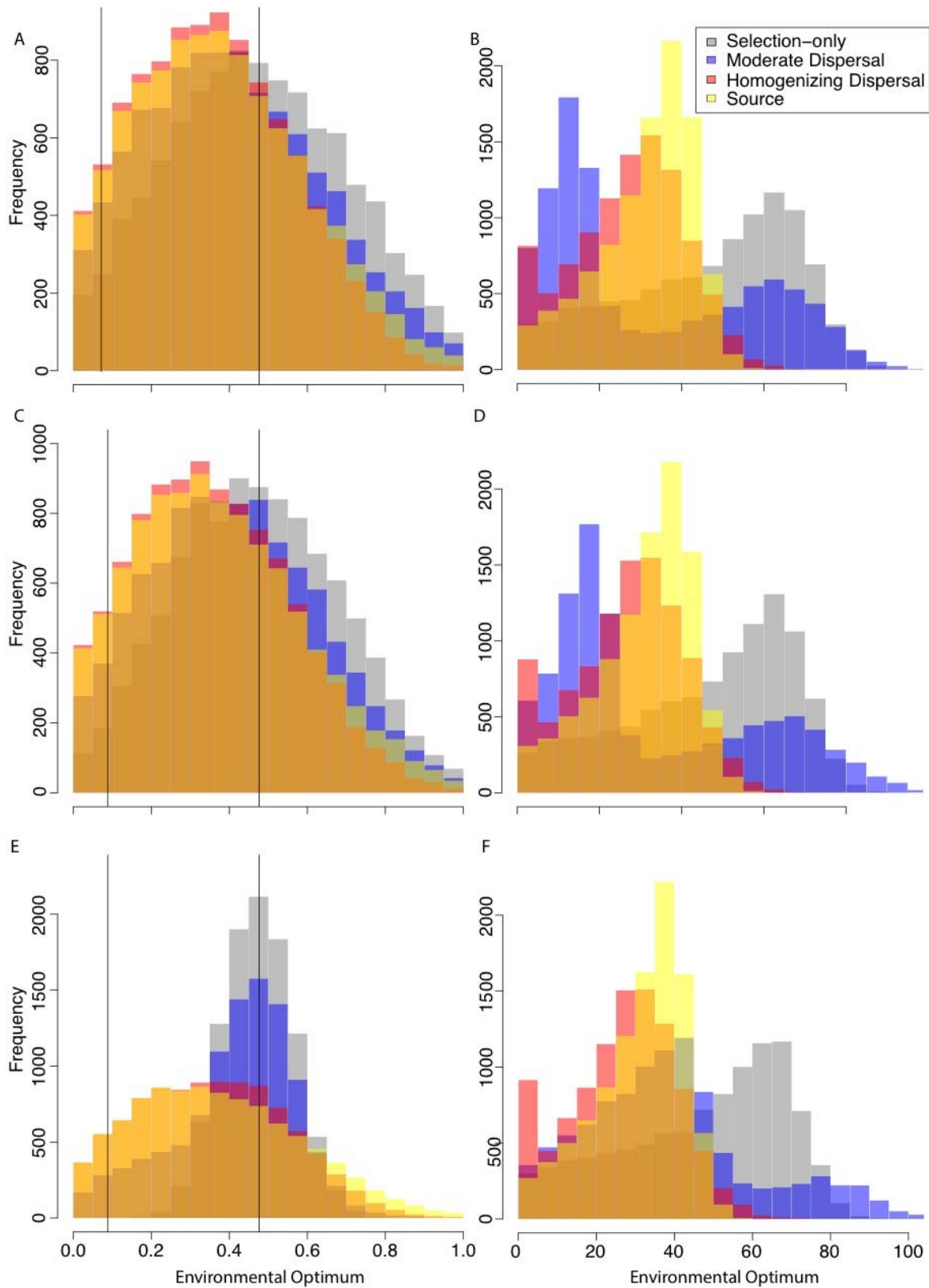


Figure 2. Histograms of species optima are shown under narrow (A-B, niche breadth = 0.0075), moderate (C-D, niche breadth = 0.086842105), and wide (E-F, niche breadth = 0.175) niches in an example

environment (0.476315789). Column 1 displays optima without accounting for species abundances and column 2 displays abundance-weighted values. The source community and its environment is displayed in yellow. Regardless of niche breadth, selection-only scenarios (gray) displayed the tightest distribution around the defined environment (vertical black line), followed by moderate (blue) and homogenizing (red) dispersal. Moderate dispersal scenarios had bimodal abundance-weighted distributions, signifying contributions from both the source and local communities. Further, narrow niche breadth supported tighter distributions in all dispersal cases.

In natural systems, microbial community composition differences can be due to competitive dynamics that select for organisms in different niches (determinism) [51,52] and to immigration of new taxa from the regional species pool (stochasticity) [29,49,53]. Strong local selective pressures can lead to more fit species and enhanced biogeochemistry [29]. Due to the lack of immigrating maladapted species in the selection-only simulations, biogeochemical rates were maintained regardless of the difference between source and receiving community environments. This indicates that maintenance of biogeochemical function can be due solely to species adaptation to local conditions (as opposed to kinetic or resource limitations that are not included in our models). Indeed, a plethora of literature demonstrates that environmental features such as pH [42], nutrients [54], and salinity [43,44] impact microbial community structure and biogeochemical function, and our results support linkages among these ecosystem properties via microbial adaptation.

We also indicate that microorganisms from dissimilar environments, such as those transported from longer distances or across steep physical or geochemical gradients, are more likely to suppress biogeochemical function than those from more similar environments. When we included dispersal from a source community, greater differences between the source and receiving communities led to decreases in biogeochemical function in the receiving communities (Figure 3b, $p < 0.0001$), and this effect became more pronounced as dispersal increased. Natural systems are influenced by some combination of dispersal and selection and our results indicate that function is maximized when dispersal is minimized and selection is maximized.

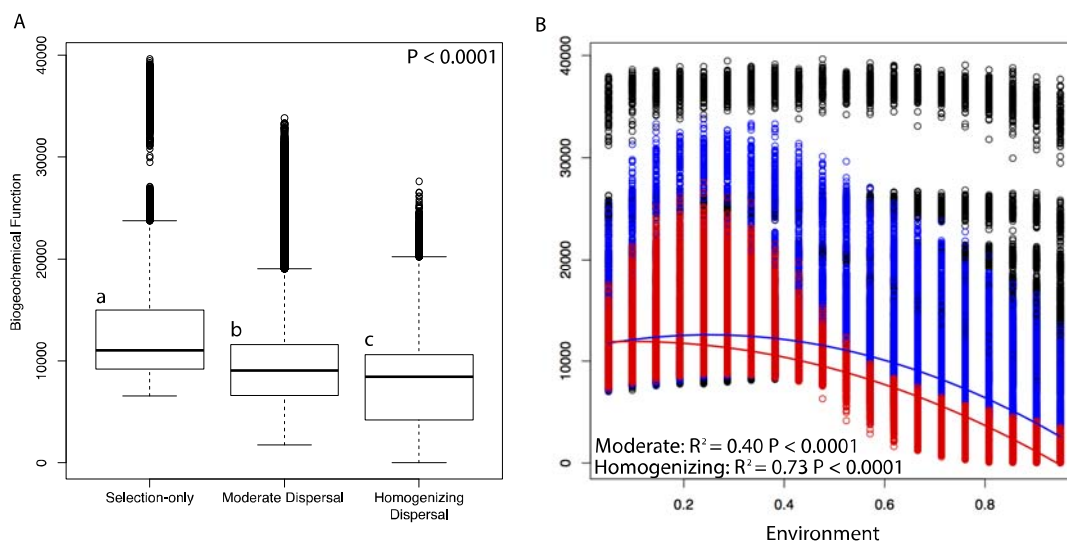


Figure 3. (a) Biogeochemical function varied across dispersal cases, with the lowest function in the case of homogenizing dispersal. Upper and lower hinges of the box plot represent the 75th and 25th percentiles and whiskers represent 1.5 times the 75th and 25th percentiles, respectively. Letters denote statistical significance. **(b)** In the selection-only case (black), biogeochemical function did not vary with changes in

the environment. With moderate (blue) and homogenizing (red) dispersal, biogeochemical maxima were consistent with environmental optima of species in the source community (Figure 3). Beyond these maxima, function declined as the environment became more dissimilar to the source community.

We also observed that niche breadth of taxa in the receiving community was a key parameter in dictating biogeochemical function when selective pressures differed between source and receiving communities. In cases without dispersal, biogeochemical function was dictated entirely by niche breadth regardless of differences in selective pressures between source and receiving communities, as inferred from null models (Figure 4a, 4d). Selective pressures in the selection-only receiving communities were most dissimilar to the source community ($\beta\text{NTI} > 2$) in simulations with both narrow niche breadth and environmental conditions that were very different from the source community (Figure 4d). This relationship was also apparent (but weaker) in simulations with an intermediate amount of dispersal (Figure 4b). In receiving communities with high rates of dispersal, stochasticity ($|\beta\text{NTI}| < 2$) was the dominant process regardless of the niche breadth or environmental condition (Figure 4f). When environmental conditions were dissimilar to the source community, biogeochemical function increased with the relative influence of deterministic assembly, as inferred by βNTI (Figure 5). This indicates that βNTI may be an effective tool for predicting biogeochemical function across environments.

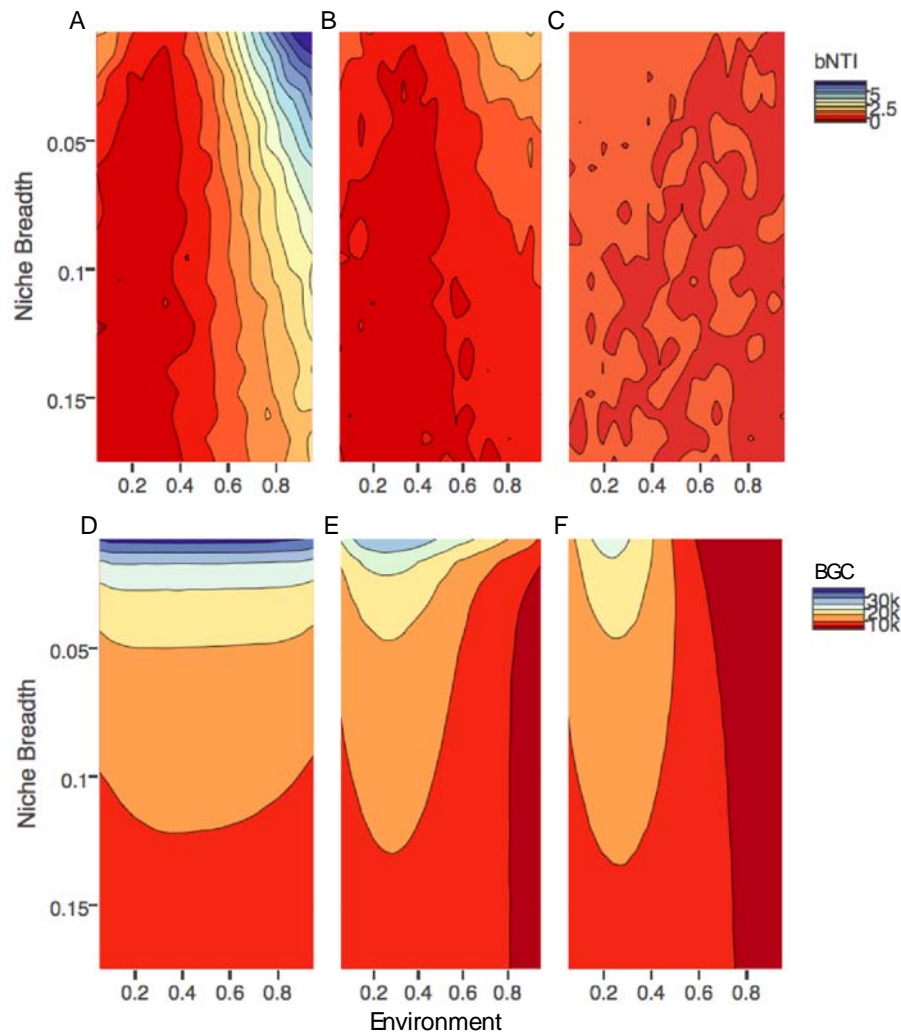


Figure 4. Average biogeochemical function (**a–c**) and selective environments (**d–f**) are displayed for each dispersal case across all parameter combinations. Biogeochemical function was entirely explained by niche breadth in selection-only scenarios (**a**) vs. a combination of niche breadth and environment in dispersal cases (**b–c**). In dispersal cases (**b–c**), biogeochemical maxima were observed when environmental conditions matched the source community and niche breadth was narrow. Selective pressures were primarily stochastic ($|\beta_{NTII}| = 2$) in both dispersal cases (**e–f**). Without dispersal (**d**), the selective environment was most dissimilar to the source community when niche breadth was smallest and the environment was most dissimilar to the source community.

Further, broader niches and thus weaker selection led to lower rates of biogeochemical function, supporting a tradeoff between communities comprised of specialist vs. generalist species [14,55,56]. Previous work in microbial systems has posited life-history tradeoffs between specialist vs. generalist species, whereby specialists expend more energy to establish their niches but function at higher levels once established [57]. Specialist species have also been found to be more sensitive to changes in the environment due to strong adaptation to their local environment, with generalists being more resilient but functioning at lower rates [58]. While we do not address temporal dynamics in our model, the separation of biogeochemical function based on niche breadth indicates a central role for the balance of specialist vs. generalist microorganisms within a community in determining function, regardless of prevailing environmental conditions.

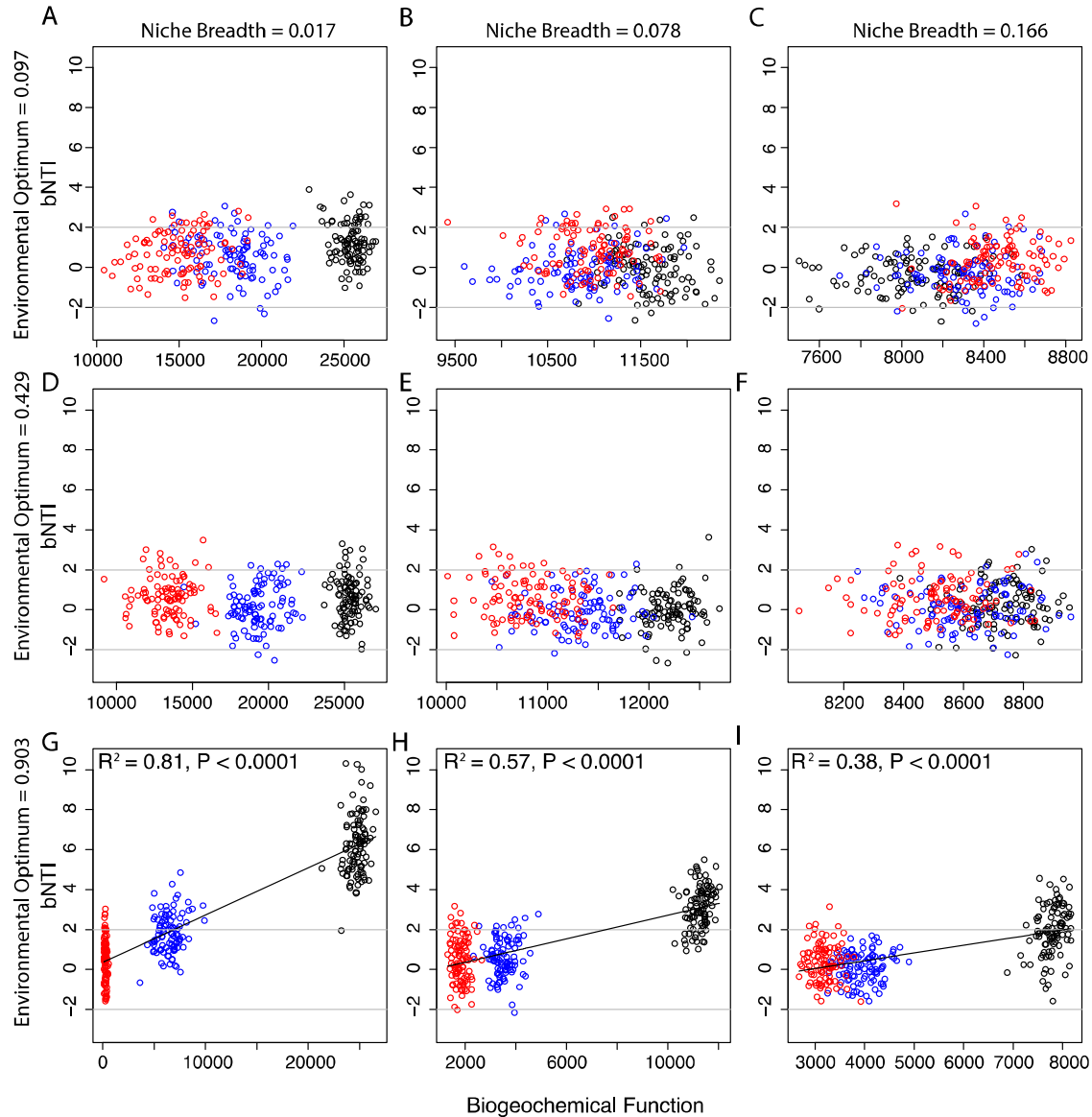


Figure 5. The relationship between β NTI and biogeochemical function varied across dispersal cases and parameter space. We show nine representative relationships in (a–i) to depict the range of variability in each parameter. Dispersal case is shown with color — selection-only is black, moderate dispersal is blue, and homogenizing dispersal is red. Niche breadth varies by column from small (1st) to large (3rd), and environment varies by row from more (top) to less (bottom) similar to the source community. β NTI values are mostly stochastic under similar and moderately different environmental conditions to the source community and do not correlate with biogeochemical function in these scenarios (a–f). However, β NTI correlated with function when the environment was most dissimilar to the source community (g–i), a relationship that declined with increasing niche breadth.

While our results suggest that maladapted immigrating organisms decrease biogeochemical function, it is important to note that stochasticity may offer buffering capacity that maintains or increases biogeochemical function relative to well-adapted deterministic communities in the context of future environmental perturbations not simulated here [58]. Stochastic spatial processes, such as dispersal, may

lead to coexistence of species with different environmental optima resulting in a community that can rapidly adapt to changing environment conditions maintain biogeochemical function in the face of perturbation. Researchers have long demonstrated positive relationships between biodiversity and ecosystem function in both macrobial [59,60] and microbial [61-63] systems, and new work has highlighted the role of stochasticity in maintaining this connection [64]. Conversely, a lack of stochasticity may result in species so well-optimized to their environment that they are vulnerable to environmental change [58]. While these communities would putatively exhibit high rates of biogeochemical function at steady state, their function would plummet in response to perturbation, akin to observations of a tradeoff between function and vulnerability in plant communities [65,66].

Taken together, we reveal how stochastic community assembly can decrease adaptation to local environments and, in turn, decrease biogeochemical function. Our modelling approach demonstrates plausible outcomes of microbial assembly processes on ecosystem functioning, and integrating this knowledge with factors such as historical abiotic conditions, competitive dynamics, and life-history traits could substantially improve ecosystem model predictions. Microbially-explicit models (e.g., MIMICS, MEND) are rapidly becoming more sophisticated and are readily amenable to modules that represent ecological assembly processes [3,4]. We propose that new microbially-explicit models should go beyond microbial mechanisms at a given point in time or space, and building upon the foundation laid by Hawkes and Keitt (2015), incorporate ecological dynamics that operate across longer time scales to influence biogeochemical function.

Some models already offer the capability to incorporate ecological dynamics with only slight adjustments to their model structures. For instance, hydrologists have used PFLOTRAN to predict patterns of ground-water surface water mixing in river corridors [67], and this model has been recently extended to incorporate aerobic respiration [68]. Since the extended version of PFLOTRAN includes transport and biogeochemistry, additional structural elements could be included that represent the influence of dispersal (e.g., through advective transport) on biogeochemical rates. Several diffusion-based model frameworks also exist for terrestrial modelling efforts [e.g., 69] and could be integrated into larger scale community land models (CLM's) to adjust rate parameters. Alternatively, one could add a minimal amount of complexity to address dispersal processes in mechanistic ecological models such as Hawkes and Keitt's (2015) model by allowing organisms to be added from source communities. Resulting information could then be used either to offset rate calculations in biogeochemical models or to create simplified ecological modules to insert into larger scale ecosystem models.

Although there are many available avenues to merge modelling efforts in microbial ecology and ecosystem science, there is little debate that integrated models will increase the accuracy of predictions in novel environments. Future research should focus on identifying the full suite of ecological processes that are critical to biogeochemical cycling and incorporating these mechanisms into emerging model frameworks.

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