Fine-mapping of genetic loci driving spontaneous clearance of hepatitis C virus infection

Hailiang Huang^{1,2,3}, Priya Duggal⁴, Chloe L. Thio⁵, Rachel Latanich⁵, James J. Goedert⁶, Alessandra Mangia⁷, Andrea L. Cox⁵, Gregory D. Kirk⁵, Shruti Mehta^{4,5}, Joel N. Blankson⁵, Jasneet Aneja³, Laurent Alric⁸, Sharyne M. Donfield⁹, Matthew E. Cramp¹⁰, Salim I. Khakoo¹¹, Leslie H. Tobler¹², Michael Busch¹², Graeme J. Alexander¹³, Hugo R. Rosen¹⁴, Brian R. Edlin¹⁵, Georg M. Lauer³, David L. Thomas⁵, Mark J. Daly^{1,2}, Raymond T. Chung^{3,*}, Arthur Y. Kim^{3,*}

- 10 1. Analytical and Translational Genetics Unit, Massachusetts General Hospital and Harvard
- 11 Medical School, Boston, MA 02114, USA
- 2. Stanley Center for Psychiatric Research, Broad Institute of MIT and Harvard, Cambridge, MA
 02142, USA
- 3. Department of Medicine, Massachusetts General Hospital, Harvard Medical School, Boston,
 MA 02114
- 16 4. Johns Hopkins University Bloomberg School of Public Health, Baltimore, MD 21205
- 17 5. Johns Hopkins University School of Medicine, Baltimore, MD 21205
- 6. Division of Cancer Epidemiology and Genetics, National Cancer Institute, Rockville MD
 20852
- 7. IRCCS Casa Sollievo della Sofferenza Hospital, San Giovanni Rotondo, Italy;
- 21 8. Department of Medicine, Purpan Hospital, University of Toulouse III, Toulouse, France
- 22 9. Rho, Chapel Hill, NC 27517
- 23 10. South West Liver Unit, Plymouth Hospitals NHS Trust, Plymouth, United Kingdom
- 24 11. Henry Welcome Laboratories, Southampton General Hospital, Southampton, UK
- 25 12. University of California, San Francisco and Blood Systems Research Institute, San
- 26 Francisco, CA 94118.

1

2

4

5

- 27 13. Cambridge University Hospitals NHS Foundation Trust and Addenbrooke's Hospital,
- 28 Cambridge, United Kingdom;
- 29 14. University of Colorado, Aurora, Colorado 90045
- 30 15. State University of New York Downstate College of Medicine, Brooklyn, New York
- 32 *Co-corresponding authors

Abstract

Approximately three quarters of acute HCV infections evolve to a chronic state, while one quarter are spontaneously cleared. Genetic predispositions strongly contribute to the development of chronicity. We have conducted a genome-wide association study to identify genomic variants underlying HCV spontaneous clearance using Immunochip in European and African ancestries. We confirmed two previously reported significant associations, in the *IL28B/IFNL4*^{1,2} and MHC regions, with spontaneous clearance in the European population. We further fine-mapped the MHC association to a region of about 50 kilo base pairs, down from 1 mega base pairs in the previous study. Additional analyses suggested that the association in the MHC locus might be significantly stronger for virus subtype 1a than 1b, suggesting that viral subtype may have influenced the genetic mechanism underlying the clearance of HCV.

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

The development of chronic viral infection represents a failure to mount an adequate innate and/or adaptive response to a specific pathogen. Infection with hepatitis C virus (HCV) in humans represents a paradigm of a dichotomous outcome of infection, as approximately three quarters of acute HCV infections evolve to a chronic state, but one quarter are spontaneously cleared³. As such, it is likely that genetic predispositions, especially at loci that regulate the innate and/or adaptive immune response, strongly contribute to the development of chronicity. A prior genome wide association study (GWAS) conducted by our consortium demonstrated striking associations of spontaneous resolution of HCV with polymorphisms near the IFN-L3 locus (IL28B) and in the HLA class II locus (Duggal et al.4). The associations identified in Duggal et al. span large genomic regions, specifically 55,000 base pairs for the Il28B locus and > 1 mega base pairs for the HLA class II locus. Recently advances in genomic technologies allowed a more precise characterization of genetic associations and facilitated resolving these associations to much smaller genomic regions. Firstly, Immunochip⁵, a customized array platform with deeper coverage of loci enriched in autoimmune diseases, provides coverage of additional genomic variants for an opportunity to explore with greater precision the contribution of these loci to the clearance of viral infection. Secondly, additional coverage of the MHC region can be gained using an imputation algorithm that takes into account the long range linkagedisequilibrium in MHC, and a large customized reference panel with improved coverage of the MHC region⁶. Thirdly, fine-mapping algorithms^{7,8}, designed with the goal to resolve known genetic associations to smaller sets of variants, can be used with the high density genomic data to further improve the precision of the genetic associations.

69

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

We therefore conducted an analysis of a large pool of spontaneous resolvers and chronic patients of HCV using the Immunochip platform, the SNP2HLA algorithm with the T1DGC MHC imputation reference panel⁶, and a recently-developed fine-mapping algorithm^{8,9} to (1) more precisely define the susceptible variant within the known associated loci; and (2) identify additional loci associated with clearance. Similar successes have been achieved in other diseases such as inflammatory bowel disease^{8,10}. Additionally, we explored the hypothesis that there are shared mechanisms that define a "brisk" immunity able to confer both susceptibility to autoimmune disease and improved control of pathogens. We also examined the influence of region (North America versus European) upon associations with HLA within the European ancestry, as previous studies have shown variability of results, especially for the class I locus 11-14. **Results** The final dataset after QC has 166,537 variants for 527 cases/828 controls of European ancestry; and 171,161 variants for 75 cases/171 controls of African ancestry (Table 1). For each ancestry, we performed logistic regression under the additive model using the first two principal components as covariates. The QQ plot (Figure 1, using common variants with >2% minor allele frequency) and the genomic control (GC) factors (0.98 for the European ancestry and 0.92 for the African ancestry using designated null variants) indicate the effective control of the population stratification. For European samples, we identified 8 genome-wide significant variants (p-value < 5E-8) in two loci (Figure 2 and Table 2). The variant on chromosome 19, rs8099917, shows the strongest association with spontaneous clearance (p-value=5E-16). Patients

92

93

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

carrying the minor allele, G, are on average 2.5x (odds ratio=0.39) less likely to spontaneously clear the virus compared to those with two copies of the C allele. This variant is roughly 7,000 base pairs upstream of the IL28B gene, and has been previously reported to be associated with HCV spontaneous clearance⁴ and the response to chronic HCV therapy in Asian populations¹⁵. Previous studies have also shown an association between IL28B and interferon-based clearance of HCV16. Because the IL28B/IFLN4 region was not designed as a high-density locus in Immunochip, we could not test other variants in this region for their association with HCV spontaneous clearance, and was unable to provide a better resolution in this locus. The other genome-wide significant locus for the European samples is the major histocompatibility complex (MHC) locus. Genome-wide significant variants in this region are reported in Table 2 (before imputation). We used SNP2HLA⁶ and a customized reference panel from a T1D study to impute missing variants, HLA alleles and amino acid residues for this region. We identified 12 SNPs and 5 amino acids that are genome-wide significant (Figure 3 and Table 3, boldfaced). No secondary signal in this region exceeded the suggestive significance threshold (1x10⁻⁵) after conditioning on the primary signal. Therefore, all variants reported in Table 3 account for the same association signal. Using a fine-mapping algorithm described in another study⁹, we constructed the 99% credible set, which is a set of variants that has 99% probability of having the causal variant in this locus (Table 3, full). Comparing with the previous study⁴ which identified this association to a region of more than 1 mega base pairs, we mapped this association to a much smaller region of 50,562 base pairs.

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

Neither the MHC nor the IL28B locus was genome-wide significant in the African ancestry. Using the heterogeneity test (fixed-effect, implemented in the R metafor package), we found that neither the MHC locus nor the IL28B locus have significantly different effect size (p-values =0.47 and 0.29 respectively) across the two populations. Therefore, the difference in the significance is likely driven by the sample size and/or the allele frequency differences. In addition to the genome-wide significant loci, we examined genes outside the HLA that have been previously associated with HCV spontaneous clearance¹⁷. Only genes IFNG-AS1 (p-value=6E-4) and STAT1 (p-value=3E-4) showed marginal evidence of association (p-value < 1E-3), and this effect was observed only in the European cohort. No genes reached the marginal p-value threshold in the samples of African ancestry. IFNG-ASI is a long noncoding RNA that is expressed in CD4 T cells and promotes Th1 responses¹⁸. STAT1 is one of the key mediators of the type I, II and III interferon responses. Since HCV is particularly diverse, with up to a 30% difference at the amino acid level between major viral genotypes, the strain of infecting virus may influence HLAmediated clearance¹³. Unfortunately, information regarding the virus genotype or subtype was not available in this study, especially in those persons who already cleared the virus, so a direct comparison is therefore not possible. However, an indirect comparison is possible by taking advantage of the observation that North American patients are much more likely to be infected with the 1a virus and European patients are much more likely to be infected by the 1b virus{Pawlotsky:1995eg}. We observed that the association in the class II MHC locus, after accounting for the sample size difference (**Methods**), is

137

138

139

140

141

142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

158

stronger in North American samples than in European samples (Figure 4) with marginal significance (p=0.05), This suggests that viral subtype may have influenced the genetic mechanism underlying the clearance of HCV. Meta-analysis by cohorts confirms this observation (Figure 5). We also interrogated the potentially protective effect of certain SNPs associated with HLA class I alleles previously implicated in spontaneous clearance. No SNP associated with class I was associated with genome-wide significance, including those associated with HLA B*27 subtypes (p-values > 0.05). The strength of association with the SNP most closely linked with *HLA-B*57* and control of HIV-1 (rs2395029) was not genome-wide significant but showed a marked difference by continent (North America p-value=8.6E-4, Europe p-value=0.078, overall p-value=1.0E-4), suggesting that any protective effect of this class I allele differs by region. Autoimmune disorders have been reported to have shared genetic susceptibility loci^{19,20}. For each of 5 major autoimmune diseases, including inflammatory bowel disease, systemic lupus erythematosus, rheumatoid arthritis, celiac disease and multiple sclerosis, we listed all variants that reached p-value<0.001 (or the best variant) in this analysis (Table 4). We found no shared variant after considering multiple testing. This analysis was only performed in the European cohort because the African cohort has limited power due to the sample size, and GWAS results in samples of African ancestry for other autoimmune disorders is more limited. A full exploration of the hypothesis that susceptibility to autoimmunity also confers ability to clear HCV will require a larger sample size. An alternate approach, taken by the International Genetics of Ankylosing Spondylitis Consortium²¹, is to search for the reported associations with other diseases in

loci having suggestive evidence (p-value < 1E-5), *i.e.*, the MHC and the *IL28B* loci in this study. We only performed the search in *IL28B* because MHC has been already implicated in many autoimmune disorders. We searched within 0.5Mb around the lead SNP (rs8099917) in *IL28B* for associations with other diseases that have been reported in the NHGRI GWAS catalog (search performed through UCSC Table Browser on August 24, 2014). Two SNPs were found to be in partial linkage disequilibrium (R²>0.4) with our lead SNP in *IL28B*, including rs12980275 (R²=0.41) associated with lipid levels in hepatitis C treatment²², and rs12979860 (R²=0.42) associated with chronic hepatitis C infection⁴/response to hepatitis C treatment¹⁶, which has been discussed in the previous sections.

Discussion

We have conducted a genome-wide association study to identify genomic variants underlying the HCV spontaneous clearance using immunochip. Consistent with previous reports⁴, two loci were found to be significantly associated with the HCV spontaneous clearance in the European cohort. The immunochip design, the imputation pipeline specifically designed for the MHC region and the novel fine-mapping algorithm facilitated the accurate characterization of classical HLA types and allowed us to achieve a higher resolution in the MHC region. Twelve SNPs and 5 amino acids in the MHC region were found to be significantly associated and no secondary signal remains after conditioning on the best SNP. Fine-mapping mapped this association to a region of about 50 kilo base pairs, down from 1 mega base pairs in the previous study. We found no associated variants in the African cohort, probably due to different genetic background

(in the case of the *IL28B* locus) and limited sample size (in the case of the MHC locus). Additional analyses suggested that the association in the class II MHC locus might be significantly stronger for virus subtype 1a than 1b, suggesting that viral subtype may have influenced the genetic mechanism underlying the clearance of HCV^{13,23}. Additional evidence, such as virus typing, is needed to confirm this finding. Limitations of this study include inability to dissect SNPs near the IL28B/IFLN4 region, as this loci had not been previously implicated in autoimmune GWAS studies. While the Immunochip did include rs8099917 as a surrogate for this region, additional information regarding associations with rs12979860 and ss469415590 is not available {Kumar: 2016ea}. Also, this study was a fine-mapping exercise that narrowed the MHC significantly but was not fully independent due to considerable overlap with the previous GWAS. Previous studies of GWAS data revealed that there are SNPs and loci with evidence of association across multiple immune-mediated diseases¹⁹. We found several variants that have suggestive and plausible evidence of associations with both HCV spontaneous clearance and another autoimmune disorders. Despite the observation that none of these variants are significant after the strict Bonferroni correction, they jointly confirm the concept that shared genetic mechanisms underlie autoimmune disorders and suggest the hypothesis that susceptibility to autoimmunity may also confer ability to clear HCV. Fuller exploration of this hypothesis will require further analyses with larger sample sizes.

Methods

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

223

224

225

226

227

Overview of samples 1,944 samples from 13 cohorts (ALIVE, BBAASH, HGDS, MHCS, Rosen and colleagues, REVELL, BAHSTION, SWAN, Toulouse, Cramp and colleagues, Hencore, Mangia and colleagues, UK Drug Use Cohort) were genotyped in this study, as previously described⁴. Self-clearance of HCV was coded as cases (718 samples) and persistence of HCV was coded as controls (1,180 samples). Samples with unidentified clearance status were not used (46 samples). All samples were genotyped using Illumina's Immunochip, a custom Infinium chip with 196,524 SNPs and small in/dels. A large number of these variants are in 187 high-density regions known to be associated with twelve autoimmune disorders and inflammatory diseases. Variants in these highdensity regions include 289 established associations, variants from 1000 genome project low coverage pilot 1 study²⁴, and variants discovered in re-sequencing²⁵. In addition, roughly 25,000 variants were included as replication of unrelated diseases as part of the WTCCC2 project, with the purpose of serving as null SNPs in analyses. Sample ethnicities To identify the sample ethnicities, we first constructed the principal component axes using Hapmap samples. 988 founders from Hapmap phase 3 (draft release 2)²⁶, including samples from ethnicities ASW, CEU, CHB, CHD, GIH, JPT, LWK, MEX, MKK, TSI and YRI were used. To calculate the principal components, only common variants that are also present in the Immunochip were used, and AT/GC SNPs were excluded to avoid ambiguous strand alignment. We performed LD pruning of the variants, resulting in a total of 15,525 variants used to create the principal components. The study samples were then projected to the principal component axes and assigned the ethnicities based on their

229

230

231

232

233

234

235

236

237

238

239

240

241

242

243

244

245

246

247

248

249

distance to the Hapmap samples. Out of 1,898 samples, 1,416 samples were mapped to European ancestry, 225 samples were mapped to African ancestry and 227 samples were admixtures and were not used in this study. **Quality control** OC was performed separately on samples of European and African ancestries separately. Variants that failed the Hardy–Weinberg equilibrium test in controls (p-value $\leq 1E-5$) or had low call rate (≤95%) were identified, and 24,820 variants were removed in European samples and 20,196 variants were removed in African samples. The remaining variants were used to perform QC in samples. Samples were cleaned for having low call rate (\leq 95%) or having high heterozygosity rate (> 3 standard deviations from the mean). We then created a LD pruned dataset for calculating the identity by state (IBS) matrix and the principal components. We pruned the variants using a sliding window of 50 variants, step size of 5 variants and variance inflation factor threshold of 1.25. There were 20,782 variants in European samples, and 21,778 variants in African samples after the pruning. The IBS matrix was calculated using this LD pruned dataset and checked for sample relatedness. 28 duplicated samples in European cohorts and 9 duplicated samples in cohorts of African ancestry have been identified and removed (pi hat>0.9). The final dataset has 527 cases and 828 controls for European cohorts, and 75 cases and 171 controls for African cohorts. To correct for within European and within African population stratification, we calculated the principal components for samples of European ancestry and African ancestry, respectively. The first two principal components sufficiently control the

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

267

268

269

270

271

population stratification in both ancestries (results not shown) and were use in the association analysis as covariates. **Imputation** Imputation of the MHC region was performed on QC cleaned data using SNP2HLA⁶. This software package takes advantage of the long-range linkage disequilibrium between HLA loci and SNP markers across the MHC region and can perform accurate imputation of classical HLA types starting from SNP genotype data. The reference panel was created using the Type 1 Diabetes Genetics Consortium's high quality HLA reference panel (roughly 5,000 European samples), which includes classical HLA alleles and amino acids at class I (HLA-A, -B, -C) and class II (-DPA1, -DPB1, -DQA1, -DQB1, and -DRB1) loci. **Association test** All association tests were performed in PLINK 1.07 ²⁷ using the logistic regression. We assumed additive models and used the first two principal components as covariates in the regression. HCV spontaneous clearance was coded as case so an odds ratio > 1 indicates the tested allele increases the probability of spontaneous clearance. Test whether an association has the same effect size across two cohorts For a cohort of sample size N, assume the association of interest has p-value of p, and the chi-square statistic, calculated from p using the chi-square distribution, is χ^2 . If this association has the same effect size in the other cohort, the expected chi-square statistic in the other cohort with population M is approximately $\chi^{2\bullet}M/N$. The significance of whether this association has the same effect size can be evaluated using the non-central

- 272 chi-square distribution with one degree of freedom and non-centrality parameter of
- 273 $\chi^{2\bullet}M/N$.

275

276

277

278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

Acknowledgements: This project was funded in whole or in part by the National Institute of Allergy and Infectious Diseases (U19AI088791 and AI082630), the Office of AIDS Research through the Center for Inherited Diseases at Johns Hopkins University, the National Institute on Drug Abuse (R01DA033541, R01DA013324, R01DA12568, and R01DA04334). The MACS is funded by the National Institute of Allergy and Infectious Diseases, with additional supplemental funding from the National Cancer Institute (UO1-AI-35042, UL1-RR025005, UO1-AI-35043, UO1-AI-35039, UO1-AI-35040, and UO1-AI-35041). The REVELL cohort was funded by R01HL076902. Swan cohort was funded by R01-DA16159, R01-DA21550, and UL1-RR024996. The HGDS is funded by the National Institutes of Health, National Institute of Child Health and Human Development, R01-HD-41224. **Author contributions:** Study design: AYK, RTC., DLT, PD, HH; Collecting samples and clinical information: AYK, CLT, RL, JJG, AM, ALC, GDK, SM, JNB, JA, LA, SMD, MEC, SIK, LHT, MB, GJA, HRR, BRE, GML; Performed Quality Control: JA, HH; Statistical analysis: HH, PD; Writing of the manuscript: AYK, RTC, HH. **Data availability:** The data that support the findings of this study are available from the corresponding authors but restrictions apply to the availability of these data, which were used under license for the current study, and so are not publicly available. Data are however available from the authors upon reasonable request. **Author Information:** The study protocols were approved by the institutional review board (IRB) at each center involved with recruitment. Informed consent and permission to share the data were obtained from all subjects, in compliance with the guidelines

specified by the recruiting center's IRB. The authors declare no competing financial interests. Correspondence and requests for materials should be addressed to

AKIM1@mgh.harvard.edu or rtchung@partners.org

Table 1. Variants and samples in this study

300

301 302

	European ancestry	African ancestry	
Variants			
Initial	191,357	191,357	
HWE (p-value <1E-6)	-394	-300	
Missingness (5%)	-24,426	-19,896	
After QC	166,537	171,161	
Samples			
Initial	1,416	227	
Missingness (5%)	-24	-2	
Heterozygosity	-9	-1	
Duplicated	-28	-6	
After QC	527 cases/828 controls	75 cases/171 controls	

Negative numbers indicate numbers of variants or samples removed.

Table 2. Genome-wide significant associations

				European ancestry		African ancestry	
CHR	SNP	POSITION	Tested Allele	OR	p-value	OR	p-value
19	rs8099917	39743165	G	0.385	5.24E-16	0.555	0.2309
6	rs6457620	32663999	G	0.605	1.47E-09	0.758	0.1571
6	rs6457617	32663851	C	0.614	3.43E-09	0.758	0.1573
6	rs9275224	32659878	A	0.615	4.76E-09	0.685	0.0550
6	rs6932517	32678182	C	0.600	1.10E-08	0.557	0.0064
6	rs9357152	32664960	G	1.664	1.20E-08	1.484	0.1075
6	rs9378125	32679732	G	1.657	1.57E-08	1.437	0.1397
6	rs2858324	32660375	A	0.604	2.89E-08	0.590	0.0178

List of variants that have genome-wide significant association with HCV spontaneous clearance (before imputation). The genomic position is in HG18.

Table 3. Associations in the 99% credible set in the MHC region

307

308

309

CHR	SNP	POSITION	Tested	OR	P	Probability
			Allele			
6	rs6457617	32771829	C	0.6172	8.06E-09	0.1197
6	rs6457620	32771977	G	0.6172	8.06E-09	0.1197
6	rs9378125	32787710	G	1.664	1.34E-08	0.0732
6	rs5000632	32771666	G	1.665	1.35E-08	0.0727
6	rs9357152	32772938	G	1.665	1.35E-08	0.0727
6	rs9394113	32773145	G	1.665	1.35E-08	0.0727
6	rs9275224	32767856	A	0.6229	1.60E-08	0.0616
6	AA_DQB1_167_32737787_R	32737787	A	1.717	2.05E-08	0.0483
6	AA_DQB1_13_32740798_G	32740798	A	1.708	3.03E-08	0.0331
6	rs9275516	32782621	A	0.6079	3.60E-08	0.0280
6	SNP_DQB1_32737787	32737787	T	1.7	3.79E-08	0.0266
6	SNP_DQB1_32740798	32740798	G	1.7	3.79E-08	0.0266
6	SNP_DQB1_32740759_T	32740759	P	1.7	3.79E-08	0.0266
6	SNP_DQB1_32740760_A	32740760	P	1.7	3.79E-08	0.0266
6	AA_DQB1_13_32740798_A	32740798	P	1.7	3.79E-08	0.0266
6	AA_DQB1_26_32740759_Y	32740759	P	1.7	3.79E-08	0.0266
6	AA_DQB1_167_32737787_H	32737787	P	1.7	3.79E-08	0.0266
6	rs6932517	32786160	C	0.6123	5.38E-08	0.0190
6	SNP_DQB1_32737837	32737837	A	0.627	8.09E-08	0.0128
6	SNP_DQB1_32737148	32737148	G	1.68	9.42E-08	0.0110
6	AA_DQB1_45_32740702	32740702	E	1.68	9.42E-08	0.0110
6	SNP_DQB1_32740702	32740702	T	1.68	9.42E-08	0.0110
6	SNP_DQB1_32742309_A	32742309	P	1.68	9.42E-08	0.0110
6	HLA_DQB1_0301	32739039	P	1.675	1.14E-07	0.0092
6	rs9469220	32766288	G	0.6393	2.24E-07	0.0048
6	rs2856717	32778286	T	0.6255	2.38E-07	0.0045
6	AA_DQB1_26_32740759_L	32740759	A	1.534	5.32E-07	0.0021
6	SNP_DQB1_32740759_A	32740759	A	1.534	5.32E-07	0.0021
6	SNP_DQB1_32740760_G	32740760	A	1.534	5.32E-07	0.0021
6	rs2858324	32768353	T	0.6397	7.11E - 07	0.0016
6	rs2647012	32772436	A	0.6397	7.11E-07	0.0016

List of variants and HLA alleles in the MHC locus that are in the 99% credible set. Genome-wide significant variants, HLA alleles and amino acid residues are boldfaced.

Table 4. Overlap with other autoimmune disorders

Disease	# variant ^a	Source	Variant	p-value	Direction ^c
IBD^{28}	125/163	Table 1	rs2413583	0.0064	+
SLE^{29}	5/6	Table 2	rs729302	0.2714	-
RA^{30}	31/36	Table 2 and 3	rs11586238	0.0156	+
Celiac ³¹	20/27	Table 2 b	rs2187668	0.0080	-
MS^{32}	90/97	Table 1 and 2	rs9967792	0.0159	+

IBD: inflammatory bowel diseases, including Crohn's disease and ulcerative colitis; SLE: systemic lupus; RA: rheumatoid arthritis; Celiac: celiac disease; MS: multiple sclerosis. ^athe Number of variants in this study and the number of variants reported in the publication. Not all variants are available because of QC and chip design. ^b Only "Previously reported risk variants" and "New loci with genome-wide significant evidence" were included. ^c the '+' direction means the tested allele has the same direction of effect in both diseases, and the '-' direction means the opposite.

Figure 1. QQ plot for cohorts of European and African ancestries. The red line indicates the null distribution. Only variants with minor allele frequency >2% were used in this figure.

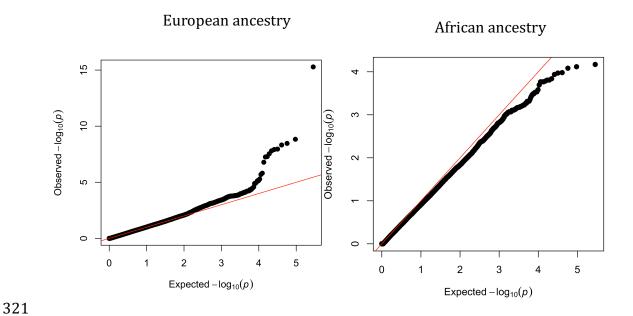


Figure 2. Manhattan plot for cohorts of European and African ancestries. The blue horizontal line indicates the suggestive significance threshold, and the red horizontal line indicates the genome-wide significance threshold.

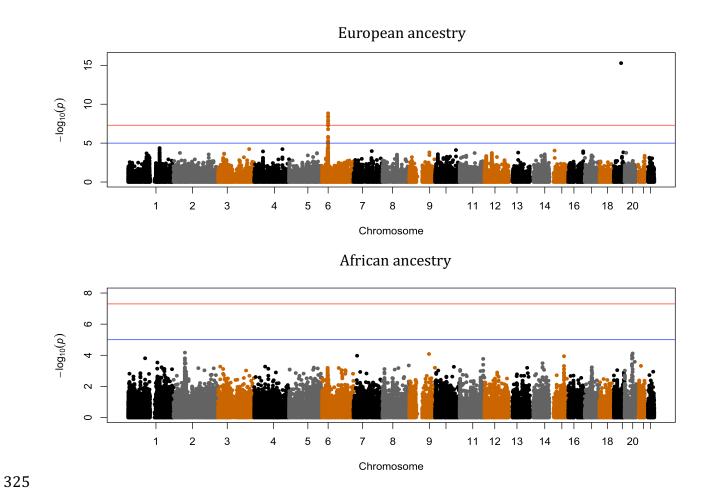


Figure 3. Regional association plot for the MHC class II region. Color indicates the

linkage equilibrium with the top associated variant (rs6457620).

326

327

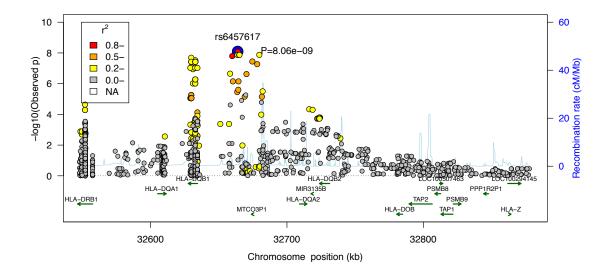
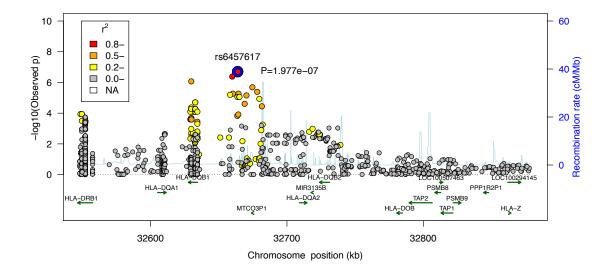


Figure 4. Regional association plot for the MHC class II region in North American samples (top) and European samples (bottom). Color indicates the linkage equilibrium with their respective top associated variant.



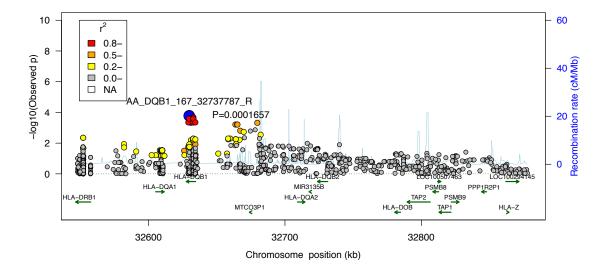
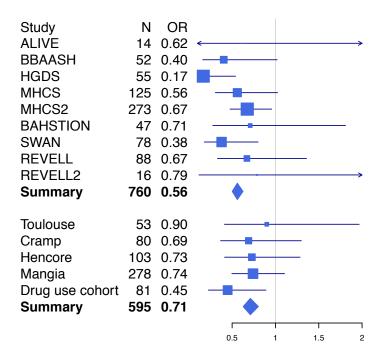


Figure 5. Forest plot for the top MHC class II association (rs6457620). Cohorts have been grouped by the geographical locations of where they were collected: the top panel includes cohorts collected in North America, and the bottom panel includes cohorts collected in Europe.



Reference

- Prokunina-Olsson, L. *et al.* A variant upstream of IFNL3 (IL28B) creating a new interferon gene IFNL4 is associated with impaired clearance of hepatitis C virus.

 Nat Genet 45, 164–171 (2013).
- O'Brien, T. R. *et al.* Comparison of functional variants in IFNL4 and IFNL3 for association with HCV clearance. *Journal of Hepatology* **63**, 1103–1110 (2015).
- 346 3. Shin, E.-C., Sung, P. S. & Park, S.-H. Immune responses and immunopathology in acute and chronic viral hepatitis. *Nat Rev Immunol* **16,** 509–523 (2016).
- Duggal, P. *et al.* Genome-wide association study of spontaneous resolution of hepatitis C virus infection: data from multiple cohorts. *Ann. Intern. Med.* **158**, 235–245 (2013).
- 5. Cortes, A. & Brown, M. A. Promise and pitfalls of the Immunochip. *Arthritis Res Ther* **13**, 101 (2010).
- Jia, X. *et al.* Imputing amino acid polymorphisms in human leukocyte antigens. PLoS ONE **8**, e64683 (2013).
- 355 7. Maller, J. B. *et al.* Bayesian refinement of association signals for 14 loci in 3 common diseases. *Nat Genet* **44,** 1294–1301 (2012).
- Huang, H. et al. Association mapping of inflammatory bowel disease loci to single variant resolution. bioRxiv (2015). doi:10.1101/028688
- Gormley, P. *et al.* Meta-analysis of 375,000 individuals identifies 38 susceptibility loci for migraine. *Nat Genet* **48,** 856–866 (2016).
- Liu, J. Z. et al. Association analyses identify 38 susceptibility loci for inflammatory bowel disease and highlight shared genetic risk across populations.
 Nat Genet (2015). doi:10.1038/ng.3359
- 364 11. Thio, C. L. *et al.* HLA-Cw*04 and hepatitis C virus persistence. *J. Virol.* **76**, 4792–4797 (2002).
- 366 12. McKiernan, S. M. *et al.* Distinct MHC class I and II alleles are associated with hepatitis C viral clearance, originating from a single source. *Hepatology* **40**, 108–368 114 (2004).
- 369 13. Kim, A. Y. *et al.* Spontaneous Control of HCV Is Associated With Expression of HLA-B*57 and Preservation of Targeted Epitopes. *Gastroenterology* **140**, 686–371 696.e1 (2011).
- Kuniholm, M. H. *et al.* Relation of HLA class I and II supertypes with spontaneous clearance of hepatitis C virus. *Genes and Immunity* **14**, 330–335 (2013).
- 374 15. Ochi, H. *et al.* IL-28B predicts response to chronic hepatitis C therapy fine-375 mapping and replication study in Asian populations. *Journal of General Virology* 376 **92,** 1071–1081 (2011).
- 377 16. Ge, D. *et al.* Genetic variation in IL28B predicts hepatitis C treatment-induced viral clearance. *Nature* **461**, 399–401 (2009).
- 379 17. Mosbruger, T. L. *et al.* Large-Scale Candidate Gene Analysis of Spontaneous Clearance of Hepatitis C Virus. *J INFECT DIS* **201,** 1371–1380 (2010).
- 381 18. Peng, H. *et al.* The Long Noncoding RNA IFNG-AS1 Promotes T Helper Type 1 Cells Response in Patients with Hashimoto's Thyroiditis. *Sci. Rep.* **5,** 17702 (2015).
- 384 19. Cotsapas, C. et al. Pervasive sharing of genetic effects in autoimmune disease.

385 *PLoS Genet* 7, e1002254 (2011).

- Richard-Miceli, C. & Criswell, L. A. Emerging patterns of genetic overlap across autoimmune disorders. *Genome Med* **4,** 6 (2012).
- Consortium, I. G. O. A. S. Identification of multiple risk variants for ankylosing spondylitis through high-density genotyping of immune-related loci. *Nat Genet* **45**, 730–738 (2013).
- Clark, P. J. *et al.* Interleukin 28B polymorphisms are the only common genetic variants associated with low-density lipoprotein cholesterol (LDL-C) in genotype-1 chronic hepatitis C and determine the association between LDL-C and treatment response. *J. Viral Hepat.* **19,** 332–340 (2012).
- 395 23. Nitschke, K. *et al.* HLA-B*27 subtype specificity determines targeting and viral evolution of a hepatitis C virus-specific CD8+ T cell epitope. *Journal of Hepatology* **60**, 22–29 (2014).
- 398 24. 1000 Genomes Project Consortium. A map of human genome variation from population-scale sequencing. *Nature* **467**, 1061–1073 (2010).
- Trynka, G. *et al.* Dense genotyping identifies and localizes multiple common and rare variant association signals in celiac disease. *Nat Genet* **43,** 1193–1201 (2011).
- 402 26. International HapMap Consortium. A haplotype map of the human genome. *Nature* **437**, 1299–1320 (2005).
- 404 27. Purcell, S. *et al.* PLINK: A Tool Set for Whole-Genome Association and
 405 Population-Based Linkage Analyses. *The American Journal of Human Genetics* 406 81, 559–575 (2007).
- Jostins, L. *et al.* Host–microbe interactions have shaped the genetic architecture of inflammatory bowel disease. *Nature* **491**, 119–124 (2012).
- 409 29. Lee, Y. H., Bae, S.-C., Choi, S. J., Ji, J. D. & Song, G. G. Genome-wide pathway analysis of genome-wide association studies on systemic lupus erythematosus and rheumatoid arthritis. *Mol Biol Rep* **39**, 10627–10635 (2012).
- 412 30. Stahl, E. A. *et al.* Genome-wide association study meta-analysis identifies seven new rheumatoid arthritis risk loci. *Nat Genet* **42**, 508–514 (2010).
- Dubois, P. C. A. *et al.* Multiple common variants for celiac disease influencing immune gene expression. *Nat Genet* **42**, 295–302 (2010).
- 416 32. International Multiple Sclerosis Genetics Consortium (IMSGC) *et al.* Analysis of
 417 immune-related loci identifies 48 new susceptibility variants for multiple sclerosis.
 418 Nat Genet 45, 1353–1360 (2013).