

1 WIDE-SCALE COMPARATIVE ANALYSIS OF LONGEVITY GENES AND 2 GENETIC INTERVENTIONS

3 Hagai Yanai¹, Arie Budovsky^{1,2}, Thomer Barzilay¹, Robi Tacutu³, Vadim E. Fraifeld¹

4 ¹ The Shraga Segal Department of Microbiology, Immunology and Genetics, Center
5 for Multidisciplinary Research on Aging, Ben-Gurion University of the Negev, POB
6 653, Beer Sheva 8410501, Israel

7 ² Technological Center, Biotechnology Unit, Beer Sheva 8489101, Israel

8 ³ Computational Biology of Aging Group, Institute of Biochemistry, Romanian
9 Academy, Bucharest 060031, Romania

10
11 Corresponding author:

12 Vadim E. Fraifeld, MD, PhD

13 The Shraga Segal Department of Microbiology, Immunology and Genetics

14 Faculty of Health Sciences

15 Ben-Gurion University of the Negev

16 POB 653, Beer Sheva 8410501, Israel.

17 Phone: +972-8-6477292

18 E-mail: vadim.fraifeld@gmail.com

19
20
21 **Keywords:** longevity genes, evolutionary conservation, comparative analysis, gene
22 enrichment, gene orthology, proteome, public and private mechanisms of
23 aging/longevity

24

1 Abstract

2 Hundreds of genes have been identified as being involved in the control of lifespan in
3 the four common model organisms (yeast, worm, fruit fly and mouse). A major
4 challenge is to determine if longevity-associated genes (LAGs) are model-specific or
5 may play a universal role as longevity regulators across diverse taxa. A wide-scale
6 comparative analysis of the 1,805 known LAGs across 205 species revealed that (i)
7 LAG orthologs are substantially over-represented, from bacteria to mammals,
8 especially noted for essential LAGs; (ii) the effects on lifespan, when manipulating
9 orthologous LAGs in different model organisms, were mostly concordant, despite of a
10 high evolutionary distance between them; (iii) the most conserved LAGs were
11 enriched in translational processes, energy metabolism, development, and DNA
12 repair. The least conserved LAGs were enriched in autophagy (Fungi), G-proteins
13 (Nematodes), and neuroactive ligand-receptor interactions (Chordata). The results
14 also suggest that antagonistic pleiotropy is a conserved principle of aging.

1 Background

2 The role of genetic factors in determination of longevity and aging patterns is an
3 intensively studied issue (Vijg and Suh 2005; Kenyon 2010). Hundreds of genes have
4 thus far been identified as being involved in the control of lifespan in model
5 organisms (Tacutu et al. 2013). These genes (further denoted as longevity-associated
6 genes, LAGs) could be defined as genes whose modulation of function or expression
7 (such as gene knockout, overexpression, partial or full loss-of-function mutations,
8 RNA interference or genetic polymorphisms) results in noticeable changes in
9 longevity or the aging phenotype (Budovsky et al. 2007; Tacutu et al. 2013). We have
10 previously investigated the characteristic features of LAGs and found that (i) they
11 display a marked diversity in their basic function and primary cellular location of the
12 encoded proteins (Budovsky et al. 2007); (ii) LAG-encoded proteins display a high
13 connectivity and interconnectivity. As a result, they form a scale-free protein-protein
14 interaction network (“longevity network”), indicating that LAGs could act in a
15 cooperative manner (Budovsky et al. 2007; Wolfson et al. 2009; Tacutu et al. 2010,
16 2012). (iii) Many LAGs, particularly those that are hubs in the “longevity network”,
17 are involved in age-related diseases (including atherosclerosis, type 2 diabetes,
18 cancer, and Alzheimer’s disease), and in aging-associated conditions (such as
19 oxidative stress, chronic inflammation, and cellular senescence) (Budovsky et al.
20 2007, 2009; Tacutu et al. 2010, 2011). (iv) The majority of LAGs established by that
21 time in yeast, worms, flies, and mice, have human orthologs, indicating their
22 conservation “from yeast to humans” (Budovsky et al. 2007, 2009). This assumption
23 was also supported by studies on specific LAGs or pathways such as Foxo (Martins et
24 al. 2016), insulin/IGF1/mTOR signaling (Tatar et al. 2003; Warner 2005; Piper et al.
25 2008; Ziv and Hu 2011; Gems and Partridge 2013; Zhang and Liu 2014; Pitt and

1 [Kaeberlein 2015](#)), Gadd45 ([Moskalev et al. 2012](#)), and cell–cell and cell–extracellular
2 matrix interaction proteins ([Wolfson et al. 2009](#)). Again, the above studies were
3 limited only to the four model organisms and humans.
4 Now, the existing databases on orthologs, allow for an essential extension of the
5 analysis of LAG orthology, far beyond the traditional model organisms and humans.
6 In particular, the data deposited in the InParanoid database – Eukaryotic Ortholog
7 Groups (<http://inparanoid.sbc.su.se/>, ([Sonnhammer et al. 2015](#))) includes orthologs for
8 the complete proteomes of 273 species. Here, we report the results of an
9 unprecedentedly wide-scale analysis of 1,805 LAGs established in model organisms
10 (available at Human Ageing Genomic Resources (HAGR) – GenAge database.
11 <http://genomics.senescence.info/genes/longevity.html>, ([Tacutu et al. 2013](#))), with
12 regard to their putative relevance to public and private mechanisms of aging.

13
14
15
16
17
18
19
20
21
22
23
24
25
26

1 **Results**

2 **Orthology of longevity-associated genes**

3 Our first question was how LAGs orthologs are distributed across diverse taxonomic
4 groups. For that purpose, we extracted the LAG orthologs for all the species in the
5 InParanoid database, using a software developed in our lab (see Methods). For each
6 gene of interest, the evolutionary conservation was evaluated as the presence or
7 absence of orthologs across 205 proteomes (all species available excluding parasites)
8 for a high InPanaranoid score of 1.0. Parasites were excluded from the analysis
9 because they usually keep the minimal set of genes required for survival in the hosts
10 and thus their inclusion could bias the results into overstating the conservation of
11 these genes and diminish the conservation of others.

12
13 As seen in [Fig. 1](#), for the vast majority of InParanoid species, the fraction of
14 conserved genes was significantly higher for LAGs than for the entire proteome of the
15 same model organism. The few exceptions were fringe cases where the baseline
16 orthology is either very high (phylogenetically very close species, for example, *C.*
17 *elegans* and *C. briggsae*), or very low (phylogenetically very distant species, for
18 example, *M. musculus* and *K. cryptofilum*) ([Suppl. Table 1](#)).

19
20 Remarkably, despite the high diversity of the species under analysis, the ratio between
21 the LAG orthologs and the orthologs of the entire proteome was relatively constant
22 along the evolutionary axis ([Fig. 2](#)). This could indicate that the high conservation of
23 LAGs is relatively independent of evolutionary distance.

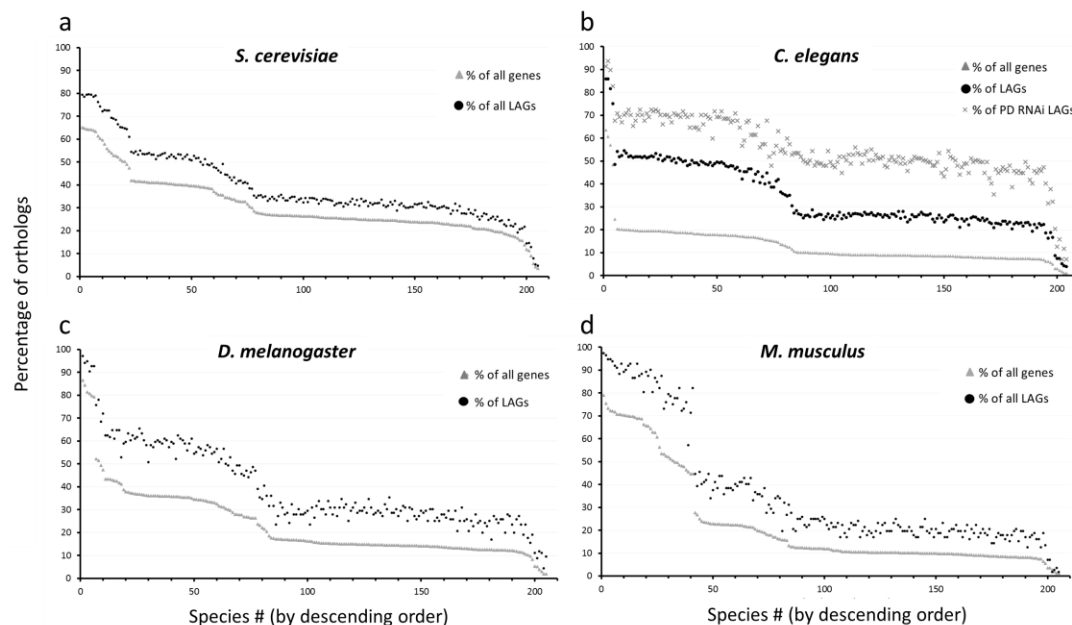


Fig. 1. Percentage of orthologs of longevity-associated genes (LAGs) from the four model organisms across 205 species. Each graph represents one of the four model organisms and the LAGs discovered for that species. Each dot represents the percentage of orthologs between the model species and a single other species (total of 205 species from all Kingdoms, for a full list of species see [Suppl. Table 1](#)). The entire proteome of the model species (extracted from the InParanoid database) was used as control. The species (X-axis) are ordered in descending order of orthology percentage for the entire proteome. Presented are the ortholog percentage of LAGs (black circle), entire proteome (grey triangle), and *C. elegans* essential LAGs discovered by post developmental RNAi (grey x). (a) *S. cerevisiae*, n = 6,590 for control and 824 for LAGs; (b) *C. elegans*, n = 20,325 for control and 733 for LAGs; (c) *D. melanogaster*, n = 13,250 for control and 136 for LAGs; (d) *M. musculus*, n = 21,895 for control and 112 for LAGs. The vast majority of pair-wise differences between LAGs and the entire proteome are significant ($p < 0.05$), with a few exceptions of fringe cases as described in the text.

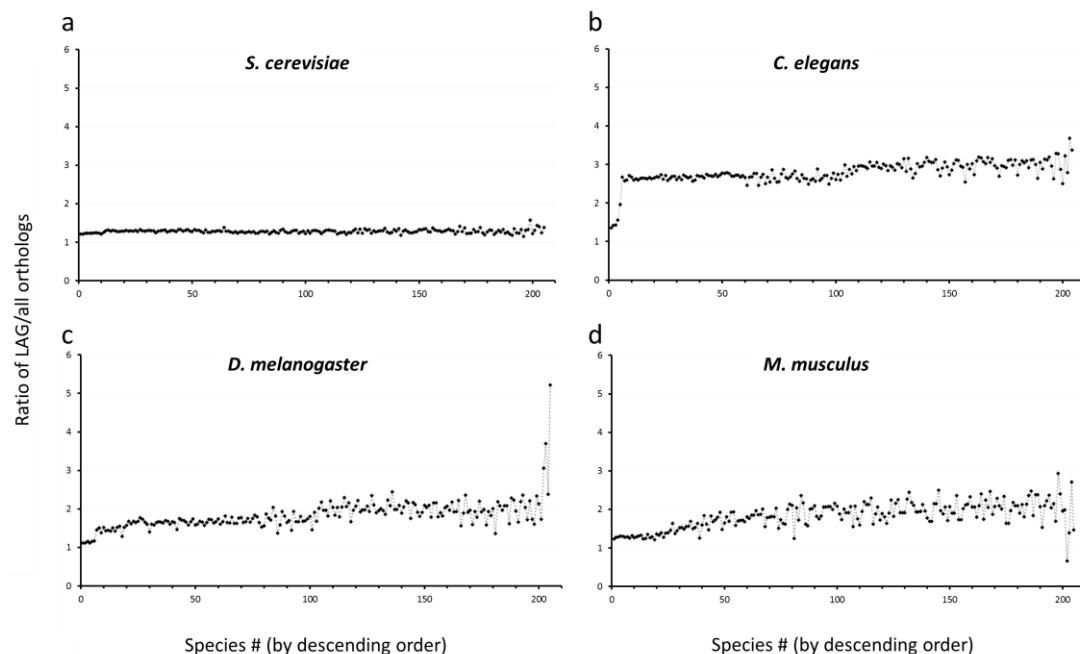


Fig. 2. Ratio of LAGs orthologs to the entire proteome. Each graph represents the LAGs discovered in the indicated model species. Each dot represents the ratio between the orthology for LAGs to the entire proteome, for a single other species (total of 205 species from all Kingdoms, for a full list of species see [Suppl. Table 1](#)). The species (X-axis) are ordered in descending order of orthology percentage for the entire proteome. **(a)** *S. cerevisiae*, n = 6,590 for control and 824 for LAGs; **(b)** *C. elegans*, n = 20,325 for control and 733 for LAGs; **(c)** *D. melanogaster*, n = 13,250 for control and 136 for LAGs; **(d)** *M. musculus*, n = 21,895 for control and 112 for LAGs.

It should be taken into account that genes that *a priori* have orthologs in humans and are involved in basic biological processes or major diseases are more often tested for their potential effect on lifespan. Despite this obvious bias, an important observation is that among the model organisms examined, the highest conservation ratio was observed for *C. elegans* ($p < E-8$ for all comparisons; [Fig. 2a](#)), where the majority of LAGs were identified by means of an unbiased genome-wide RNA interference (RNAi) screens ([Lee et al. 2003](#); [Hamilton et al. 2005](#); [Hansen et al. 2005](#); [Yanos et al. 2012](#)).

1 Using post developmental interventions such as RNAi has allowed for discovering
2 worm longevity regulators that could not be discovered otherwise, because their pre-
3 developmental inactivation causes a lethal phenotype (Tacutu et al. 2012). According
4 to WormBase (<http://www.wormbase.org/> (Howe et al. 2016)), 127 of the 733 known
5 *C. elegans* LAGs are essential for development and growth, meaning an enrichment
6 by approximately 5-fold compared to the entire genome (Tacutu et al. 2012). This is
7 even more pronounced among LAGs that extend worm lifespan by more than 20%
8 when inactivated: they are enriched 15-fold in essential genes. Since essential genes
9 are generally more evolutionary conserved than non-essential ones (Tacutu et al.
10 2011), we compared the evolutionary conservation of the essential worm LAGs to all
11 LAGs and found that these 127 essential LAGs are dramatically more conserved (Fig.
12 1a). As these genes are essential for the early stages of life, but obviously have
13 detrimental effects later in life they, by definition, fit well into Williams's idea of
14 antagonistic pleiotropy (Williams 1957). If so, the results suggest that antagonistic
15 pleiotropy is a conserved principle of aging.

16
17 One of the strong features of InParanoid is that it provides the best balance between
18 sensitivity and specificity (Chen et al. 2007). Yet, the proteomes found in the
19 InParanoid database contain many poorly annotated proteins and predicted transcripts
20 that were not experimentally verified (Sonhammer et al. 2015). These proteins have
21 relatively few orthologs in other species and therefore could influence the results. In
22 contrast, the interactomes from the BioGrid database (<http://www.thebiogrid.org>)
23 almost exclusively include experimentally verified proteins (Chatr-Aryamontri et al.
24 2015). Therefore, the BioGrid data could serve as an additional, high quality control
25 for a more rigorous testing of the evolutionary conservation of LAGs. For this

1 purpose, we used the interactomes of *S. cerevisiae*, *C. elegans*, and *D. melanogaster*.
 2 As seen in [Suppl. Fig. 1](#), the same trend of over-conservation of LAGs was also
 3 observed in comparison to the BioGrid control. Mouse was not included in the
 4 analysis because its BioGrid gene list still contains a relatively small portion of the
 5 entire genome and thus could not provide a reliable control.

6
 7 Altogether, the results clearly show a high evolutionary conservation of LAGs across
 8 distant species. With regard to this, a question arises as to whether this observation is
 9 attributed to an enrichment of specific categories that are known to be strongly
 10 preserved in the course of evolution. From the available data on gene and protein
 11 annotations for the four model species we noted that LAGs are enriched in genes that
 12 belong to categories known to be extraordinarily conserved in evolution, such as the
 13 ribosomal or mitochondrial genes ([Suppl. Tables 2 and 3](#)). However, exclusion of
 14 LAGs belonging to these categories from the analysis had almost no impact ([Suppl.](#)
 15 [Fig. 2](#)). Therefore, we conclude that the high evolutionary conservation of LAGs is
 16 not solely attributed to an enrichment of proteins from exceptionally conserved
 17 categories, but rather reflects a general trend.

18 19 **“Public” and “private” LAG categories**

20 The distinction between public and private mechanisms of aging and longevity is a
 21 fundamental question in comparative studies of biogerontology ([Gems and Partridge](#)
 22 [2013](#)). We attempted to shed some light on this subject based on the evolutionary
 23 conservation of LAGs in different taxa. Yet, it is important to note that if a given
 24 LAG is highly evolutionary conserved, it does not automatically translate to its role in
 25 a public mechanism of aging. In fact, in order to draw conclusions on public or

private mechanisms from the presence or lack of orthologs, one must (i) have a context on the mode of operation of a given protein as its function could differ between species; or (ii) compare groups of proteins belonging to a certain pathway or category, so that generalized assumptions may be made. The data used in this study only allows for the second approach. Thus, we comprised lists of proteins under different conservation criteria, e.g. proteins that have orthologs across at least 12 phyla or have orthologs in a limited number of taxa only (for more details, see [Suppl. Table 4](#)). As shown in [Fig. 3](#), LAGs are generally conserved over more phyla than the entire genome, again indicating their wider evolutionary conservation. Nevertheless, while most LAGs are broadly conserved, a considerable portion of them (around 10-20%) are specific to a relatively small number of phyla ([Fig. 3](#)).

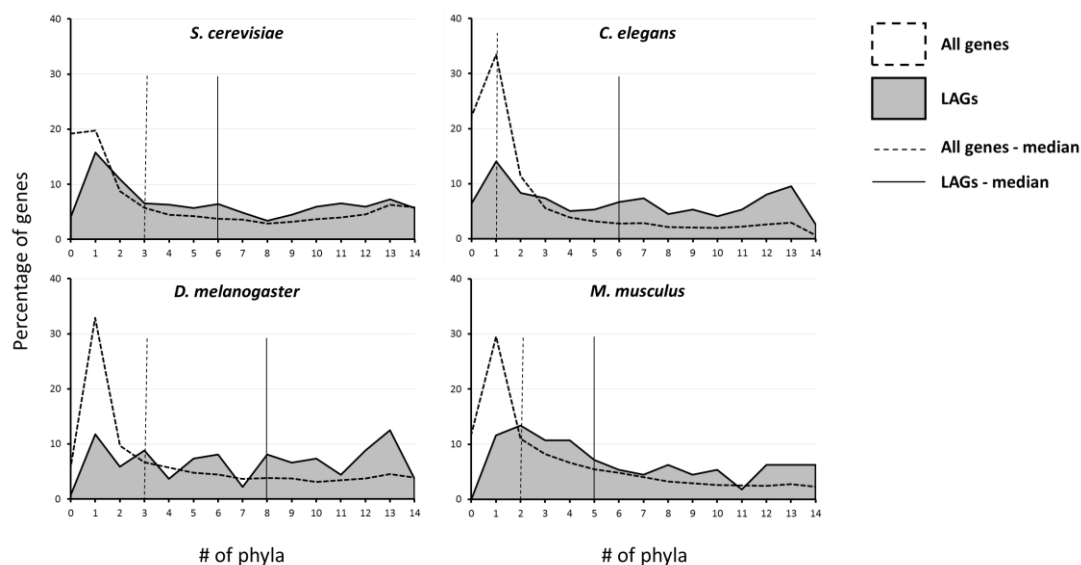


Fig. 3. Distribution of LAGs according to the number of phyla in which LAGs have orthologs. Each graph represents the distribution of LAGs (grey area) discovered in the indicated model species. The entire proteome was used as a control (dotted line). X-axis depicts the number of phyla in which the genes have orthologs. The medians of the distributions are presented as vertical lines: dotted line for all genes and smooth black line for LAGs.

1 To get further insight into the universality of longevity-associated pathways, we
2 carried out an enrichment analysis for LAGs that are conserved across a large number
3 of phyla (“public”) and those that are specific to certain taxonomic group(s)
4 (“private”). For the “private” analysis, we used the phylum of the corresponding
5 model organism (as depicted in the [Suppl. Table 5](#)), since smaller taxonomic groups
6 did not yield statistically conclusive results. Full detailed enrichment analysis is
7 available in [Suppl. Table 6](#).

8

9 Overall, the analysis of the most conserved (public) LAGs revealed that they fall
10 under 4 major categories ([Suppl. Tables 5 and 6](#)): (i) Ribosome and translational
11 processes; (ii) mitochondria and energy metabolism pathways (including the FoxO
12 pathway); (iii) development, and (iv) DNA repair processes. These results provide a
13 strong support for previous studies, in particular those by Smith et al. (2008),
14 McElwee et al. (2007), and Freitas et al. (2011), and highlight the public role of these
15 categories in the control of lifespan. It should however be noted that the number of
16 LAGs was not always sufficient for a robust enrichment analysis, especially for the
17 mouse and fly models (see [Suppl. Table 6](#)); the results from the yeast and worm
18 models were more significant and thus more reliable.

19

20 Due to the high evolutionary conservation of LAGs, those that have orthologs only in
21 the same phylum as the model species in which they were discovered are relatively
22 small in number. Because of that, the enrichment analysis of these genes yielded less
23 significant results ([Suppl. Table 6](#)). Nevertheless, the “private” list for *S. cerevisiae*
24 (i.e. yeast LAGs with orthologs only in Fungi) was found to be enriched with
25 autophagy-related genes ([Suppl. Table 5](#)). For LAGs that have orthologs only in

1 Nematoda, we surprisingly found enrichment in G-protein-related genes. Indeed, both
 2 autophagy and G-protein signaling represent basic and highly conserved processes
 3 which were shown to be involved in aging and longevity in various model organisms
 4 (Lans and Jansen 2007; Hahm et al. 2009; Rubinsztein et al. 2011; Schneider and
 5 Cuervo 2014). Yet, the unusual enrichment of these pathways in yeast and worms
 6 definitely highlights their importance in determination of longevity for these taxa
 7 specifically. For vertebrates, we found a significant enrichment in Neuroactive ligand-
 8 receptor interaction, which could reflect the importance of neuroendocrine regulation
 9 of aging and longevity in higher organisms (Dilman et al. 1986; Frolkis 1988;
 10 Blagosklonny 2013).

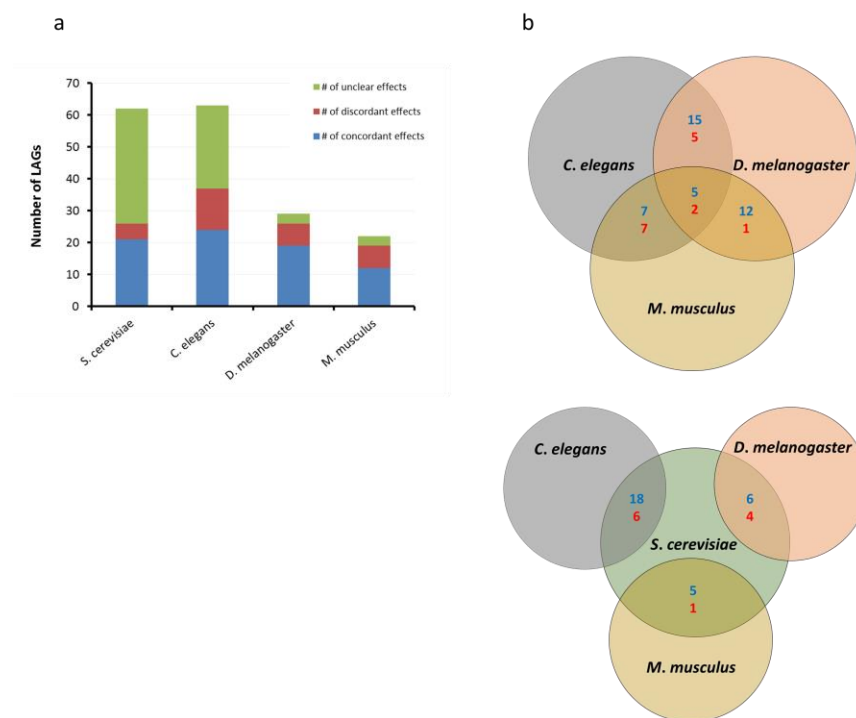
11

12 **Concordancy and discordancy in lifespan-modulating genetic interventions**

13 Considering the conservation of many LAGs over a broad evolutionary distance, a
 14 valid question is whether modulating a given LAG in different species has a similar
 15 impact on longevity, i.e. lifespan extension or reduction. This was previously
 16 addressed for worm and yeast (Smith et al. 2008), where the genetic component of
 17 lifespan determination was found to be significantly conserved. Here, we broadened
 18 the question to *all* available model organisms. Namely, we compared all orthologs
 19 which were shown to have an impact on longevity in more than one species. Overall,
 20 we found that approximately 10% of LAGs' orthologs ($n = 184$) were identified as
 21 such in at least two model organisms; 36 LAGs' orthologs were identified in three and
 22 20 in four model organisms. The number of concordant effects was significantly
 23 higher than the discordant ones ($p < 0.003$). That is, manipulation of LAGs has, more
 24 than often, the same effect in different species (Fig. 4, Suppl. Table 7). A substantial
 25 portion of the genetic interventions in yeast and worms could not be clearly defined as

1 concordant or discordant with other model organisms (Fig. 4a, green), mostly due to a
2 major difference in the methods of evaluation. In particular, in yeast studies, signs of
3 premature aging and a reduction of lifespan could actually be just the results of
4 reduced fitness but not mechanisms of aging *per se* (Tacutu et al. 2013).
5
6 When looking at pairwise comparisons (Fig. 4B), it is evident that the level of
7 concordancy is very high for some pairs of species (for example, *M. musculus* and *D.*
8 *melanogaster*) and lower for others (for example, *M. musculus* and *C. elegans*). In
9 order to discern what could bring about this difference, we calculated a conservation
10 index for each pair of orthologs (as previously described (Huang et al. 2004)) and
11 compared the results to the concordancy/discordancy of the effects. As seen in Suppl.
12 Fig. 3, the observed discordancy could not be explained by sequence dissimilarity.
13 One of the possible explanations for the observed discordancy is that in these cases
14 orthologous LAGs were discovered by interventions which greatly differ from one
15 another (e.g., knockout and overexpression). As such, if a given LAG is knocked-out
16 and as a result the animal ages more rapidly, that LAG is defined as a “pro-longevity”
17 gene; however, an overexpression of the same LAG will not necessarily increase
18 lifespan. For example, a knockout of G protein, alpha subunit (*gpa-9*) in *C. elegans*
19 increases maximum lifespan by up to 50%, but paradoxically, its overexpression also
20 increases maximum lifespan (by 20%) (Schneider and Cuervo 2014). If such a
21 difference occurs in the same species, more so could be expected when testing for
22 effects on lifespan between different model organisms. Indeed, as is evident from
23 Suppl. Fig. 4, the concordancy was significantly higher when a similar intervention
24 was performed. Then, at least some of the discordancy could be explained by a variety
25 in the methods of intervention.

1 Interestingly, only 5 LAGs (*Sod2*, *Sirt1*, *Mtor*, *Fxn*, and *Rps6kb1*; in total, 20
2 orthologs) were tested for their impact on longevity in all 4 model species. The
3 manipulations of these genes showed a predominantly concordant effect on longevity,
4 with the exception of *Fxn* (*Frh-1*) which has an opposite effect only in *C. elegans*
5 (Suppl. Table 7). Altogether, the results indicate a clear trend of concordancy in the
6 effects of LAG manipulations across model species despite a high evolutionary
7 distance between them, and the observed cases of discordancy could mostly be
8 attributed to technical rather than biological issues.



9

10 **Fig. 4. Concordancy in LAG manipulations across model organisms.**

11 Concordancy was determined according to the classification of LAGs as pro- or anti-
12 longevity genes. That is, if a given LAG was determined as a pro- or anti-longevity
13 gene in two or more species, it was termed “concordant”; otherwise, it was termed
14 “discordant”. A detailed table is available in Suppl. Table 7. (a) Summary of the
15 concordancy for LAGs from each model species which have also been tested in two
16 or more species. (b) Venn diagram of the concordancy between species.

17

18

19

1 Conclusions

2 Our wide-scale analysis of longevity-associated genes (LAGs) shows that their
3 orthologs are consistently over-represented across diverse taxa, compared with the
4 orthologs of other genes, and this conservation was relatively independent of
5 evolutionary distance. The high evolutionary conservation was evident for LAGs
6 discovered in all of the four major model organisms (yeast, *S. cerevisiae*; worm, *C.*
7 *elegans*; fly, *D. melanogaster*; mouse, *M. musculus*), but was especially relevant for
8 *C. elegans*, where a large portion of LAGs were identified by genome-wide screens,
9 thus minimizing potential biases. The results from the *C. elegans* analysis also suggest
10 that antagonistic pleiotropy is a highly conserved principle of aging. Another
11 important observation in our study was that the majority of manipulations on LAG
12 orthologs in more than one model animal resulted in concordant effects on longevity.
13 This strengthens the paradigm of “public” longevity pathways and of using model
14 animals to study longevity, even across a large evolutionary distance. This notion is
15 further strengthened when combined with the observation of Smith et al. (2008) who
16 demonstrated that the existence of an ortholog is probably accompanied by a
17 preserved role in longevity. Yet, we also observed LAGs that are highly conserved
18 only in a limited number of taxa, or that displayed discordant effects when tested in
19 more than one species, which could be attributed to “private” mechanisms of aging.
20 Definitely, more comparative studies are warranted to better discriminate between
21 private and public mechanisms, with unified methods of intervention and evaluation
22 in mind. A recent study by Harel et al. (2015) could serve as a step in that direction by
23 offering a new model of short-lived vertebrate species. In perspective, the
24 combination of the existing data on LAGs with the emerging data on their expression

1 throughout lifespan could bring about a deeper understanding of the role of genetic
2 factors in aging and longevity.

3

4

5 **Methods**

6 **Gene lists**

7 *Longevity-associated genes.* The longevity-associated genes (LAGs) are defined as
8 genes whose genetic manipulation in model organisms (*Mus musculus*, *Drosophila*
9 *melanogaster*, *Caenorhabditis elegans* and *Saccharomyces cerevisiae*) was shown to
10 significantly affect their lifespan. The list was obtained from Human Ageing Genomic
11 Resources (HAGR) – GenAge database
12 (<http://genomics.senescence.info/genes/longevity.html>; (Tacutu et al. 2013)).

13

14 *Interactome genes.* Interactome genes were extracted from the BioGrid database
15 (<http://www.thebiogrid.org>; (Chatr-Aryamontri et al. 2015)) and were used as
16 additional, high quality control for a more rigorous testing of the evolutionary
17 conservation of LAGs.

18 *Essential genes.* Genes essential for the development and growth of *C. elegans* were
19 extracted from WormBase (<http://www.wormbase.org/> (Howe et al. 2016)).

20

21 **Determination of orthology**

22 Ortholog determination for each gene was based on the InParanoid database -
23 Eukaryotic Ortholog Groups (<http://inparanoid.sbc.su.se/>, (Sonnhammer et al. 2015)).
24 The analysis was performed for 205 species (all species available excluding parasites;
25 for a full list, see [Suppl. Table 1](#)). The ortholog extraction was performed

1 automatically using software developed in our lab. The taxonomy of the species
2 examined was based on the ITIS database (<http://www.itis.gov/>). The statistical
3 significance of conservation for a group of genes was evaluated with the Chi-squared
4 Goodness of fit test.

5

6 **Gene set enrichment**

7 Enrichment analysis was performed using David Bioinformatics Resources 6.8
8 (<https://david-d.ncifcrf.gov/> (Huang et al. 2009)), and WebGestalt
9 (<http://www.webgestalt.org/>; (Wang et al. 2013)). The enrichment analysis was
10 performed against 3 different backgrounds, including the whole genome, all LAGs,
11 and the genes of the model organism under the same conservation criteria depicted in
12 **Suppl. Table 5.**

13

14

15

16

17

18

19

20

21

22

23

24

1 **Declarations**

2 **Competing interests**

3 The authors declare that they have no competing interests.

4

5 **Funding**

6 This study was supported by the Fund in Memory of Dr. Amir Abramovich (to VEF),
7 the Israel Ministry of Science and Technology (to AB), and the EU funding through
8 the Competitiveness Operational Programme 2014-2020, POC-A.1-A.1.1.4-E-2015
9 (to RT).

10

11 **Authors' contributions**

12 All authors participated in data collection and analysis. In addition, TB wrote all the
13 programs for data extraction from the databases used and, in particular, the ortholog
14 extraction for large sets of genes and species and the calculation of the conservation
15 index. HY wrote the manuscript and prepared the figures and tables. AB participated
16 in writing the manuscript. RT was involved in the computational aspects of analysis.
17 VEF coordinated the study.

18

19 **Acknowledgements**

20 We would like to thank Prof. Marina Wolfson for her helpful suggestions and
21 insightful comments.

22

23

24

References

- 1 Blagosklonny MV (2013) M(o)TOR of aging: MTOR as a universal molecular
2 hypothalamus. *Aging (Albany NY)* 5:490-494. doi: 100580
- 3
4 Budovsky A, Abramovich A, Cohen R, Chalifa-Caspi V, Fraifeld V (2007) Longevity
5 network: construction and implications. *Mech Ageing Dev* 128:117-124. doi: S0047-
6 6374(06)00251-X
- 7 Budovsky A, Tacutu R, Yanai H, Abramovich A, Wolfson M, Fraifeld V (2009)
8 Common gene signature of cancer and longevity. *Mech Ageing Dev* 130:33-39. doi:
9 10.1016/j.mad.2008.04.002
- 10 Chatr-Aryamontri A, Breitkreutz BJ, Oughtred R, Boucher L, Heinicke S, Chen D,
11 Stark C, Breitkreutz A, Kolas N, O'Donnell L, Regulj T, Nixon J, Ramage L, Winter
12 A, Sellam A, Chang C, Hirschman J, Theesfeld C, Rust J, Livstone MS, Dolinski K,
13 Tyers M (2015) The BioGRID interaction database: 2015 update. *Nucleic Acids Res*
14 43:D470-8. doi: 10.1093/nar/gku1204
- 15 Chen F, Mackey AJ, Vermunt JK, Roos DS (2007) Assessing performance of
16 orthology detection strategies applied to eukaryotic genomes. *PLoS One* 2:e383. doi:
17 10.1371/journal.pone.0000383
- 18 Dilman VM, Revskoy SY, Golubev AG (1986) Neuroendocrine-ontogenetic
19 mechanism of aging: toward an integrated theory of aging. *Int Rev Neurobiol* 28:89-
20 156
- 21 Freitas AA, Vasieva O, de Magalhaes JP (2011) A data mining approach for
22 classifying DNA repair genes into ageing-related or non-ageing-related. *BMC*
23 *Genomics* 12:27-2164-12-27. doi: 10.1186/1471-2164-12-27
- 24 Frolkis VV (1988) A hundred questions on neurohumoral mechanisms of aging.
25 *Gerontology* 34:6-13
- 26 Gems D, Partridge L (2013) Genetics of longevity in model organisms: debates and
27 paradigm shifts. *Annu Rev Physiol* 75:621-644. doi: 10.1146/annurev-physiol-
28 030212-183712
- 29 Hahm JH, Kim S, Paik YK (2009) Endogenous cGMP regulates adult longevity via
30 the insulin signaling pathway in *Caenorhabditis elegans*. *Aging Cell* 8:473-483. doi:
31 10.1111/j.1474-9726.2009.00495.x
- 32 Hamilton B, Dong Y, Shindo M, Liu W, Odell I, Ruvkun G, Lee SS (2005) A
33 systematic RNAi screen for longevity genes in *C. elegans*. *Genes Dev* 19:1544-1555.
34 doi: 19/13/1544
- 35 Hansen M, Hsu AL, Dillin A, Kenyon C (2005) New genes tied to endocrine,
36 metabolic, and dietary regulation of lifespan from a *Caenorhabditis elegans* genomic
37 RNAi screen. *PLoS Genet* 1:119-128. doi: 10.1371/journal.pgen.0010017

- 1 Harel I, Benayoun BA, Machado B, Singh PP, Hu CK, Pech MF, Valenzano DR,
2 Zhang E, Sharp SC, Artandi SE, Brunet A (2015) A platform for rapid exploration of
3 aging and diseases in a naturally short-lived vertebrate. *Cell* 160:1013-1026. doi:
4 10.1016/j.cell.2015.01.038
- 5 Howe KL, Bolt BJ, Cain S, Chan J, Chen WJ, Davis P, Done J, Down T, Gao S,
6 Grove C, Harris TW, Kishore R, Lee R, Lomax J, Li Y, Muller HM, Nakamura C,
7 Nuin P, Paulini M, Raciti D, Schindelman G, Stanley E, Tuli MA, Van Auken K,
8 Wang D, Wang X, Williams G, Wright A, Yook K, Berriman M, Kersey P, Schedl T,
9 Stein L, Sternberg PW (2016) WormBase 2016: expanding to enable helminth
10 genomic research. *Nucleic Acids Res* 44:D774-80. doi: 10.1093/nar/gkv1217
- 11 Huang da W, Sherman BT, Lempicki RA (2009) Systematic and integrative analysis
12 of large gene lists using DAVID bioinformatics resources. *Nat Protoc* 4:44-57. doi:
13 10.1038/nprot.2008.211
- 14 Huang H, Winter EE, Wang H, Weinstock KG, Xing H, Goodstadt L, Stenson PD,
15 Cooper DN, Smith D, Alba MM, Ponting CP, Fechtel K (2004) Evolutionary
16 conservation and selection of human disease gene orthologs in the rat and mouse
17 genomes. *Genome Biol* 5:R47. doi: 10.1186/gb-2004-5-7-r47
- 18 Kenyon CJ (2010) The genetics of ageing. *Nature* 464:504-512. doi:
19 10.1038/nature08980
- 20 Lans H, Jansen G (2007) Multiple sensory G proteins in the olfactory, gustatory and
21 nociceptive neurons modulate longevity in *Caenorhabditis elegans*. *Dev Biol* 303:474-
22 482. doi: S0012-1606(06)01381-9
- 23 Lee SS, Lee RY, Fraser AG, Kamath RS, Ahringer J, Ruvkun G (2003) A systematic
24 RNAi screen identifies a critical role for mitochondria in *C. elegans* longevity. *Nat*
25 *Genet* 33:40-48. doi: 10.1038/ng1056
- 26 Martins R, Lithgow GJ, Link W (2016) Long live FOXO: unraveling the role of
27 FOXO proteins in aging and longevity. *Aging Cell* 15:196-207. doi:
28 10.1111/accel.12427
- 29 McElwee JJ, Schuster E, Blanc E, Piper MD, Thomas JH, Patel DS, Selman C,
30 Withers DJ, Thornton JM, Partridge L, Gems D (2007) Evolutionary conservation of
31 regulated longevity assurance mechanisms. *Genome Biol* 8:R132. doi: gb-2007-8-7-
32 r132
- 33 Moskalev AA, Smit-McBride Z, Shaposhnikov MV, Plyusnina EN, Zhavoronkov A,
34 Budovsky A, Tacutu R, Fraifeld VE (2012) Gadd45 proteins: relevance to aging,
35 longevity and age-related pathologies. *Ageing Res Rev* 11:51-66. doi:
36 10.1016/j.arr.2011.09.003
- 37 Piper MD, Selman C, McElwee JJ, Partridge L (2008) Separating cause from effect:
38 how does insulin/IGF signalling control lifespan in worms, flies and mice?. *J Intern*
39 *Med* 263:179-191. doi: 10.1111/j.1365-2796.2007.01906.x

- 1 Pitt JN, Kaeberlein M (2015) Why is aging conserved and what can we do about it?
2 PLoS Biol 13:e1002131. doi: 10.1371/journal.pbio.1002131
- 3 Rubinsztein DC, Marino G, Kroemer G (2011) Autophagy and aging. Cell 146:682-
4 695. doi: 10.1016/j.cell.2011.07.030
- 5 Schneider JL, Cuervo AM (2014) Autophagy and human disease: emerging themes.
6 Curr Opin Genet Dev 26:16-23. doi: 10.1016/j.gde.2014.04.003
- 7 Smith ED, Tsuchiya M, Fox LA, Dang N, Hu D, Kerr EO, Johnston ED, Tchao BN,
8 Pak DN, Welton KL, Promislow DE, Thomas JH, Kaeberlein M, Kennedy BK (2008)
9 Quantitative evidence for conserved longevity pathways between divergent eukaryotic
10 species. Genome Res 18:564-570. doi: 10.1101/gr.074724.107
- 11 Sonnhammer EL, Ostlund G (2015) InParanoid 8: orthology analysis between 273
12 proteomes, mostly eukaryotic. Nucleic Acids Res 43:D234-9. doi:
13 10.1093/nar/gku1203
- 14 Tacutu R, Craig T, Budovsky A, Wuttke D, Lehmann G, Taranukha D, Costa J,
15 Fraifeld VE, de Magalhaes JP (2013) Human Ageing Genomic Resources: integrated
16 databases and tools for the biology and genetics of ageing. Nucleic Acids Res
17 41:D1027-33. doi: 10.1093/nar/gks1155
- 18 Tacutu R, Shore DE, Budovsky A, de Magalhaes JP, Ruvkun G, Fraifeld VE, Curran
19 SP (2012) Prediction of C. elegans longevity genes by human and worm longevity
20 networks. PLoS One 7:e48282. doi: 10.1371/journal.pone.0048282
- 21 Tacutu R, Budovsky A, Yanai H, Fraifeld VE (2011) Molecular links between
22 cellular senescence, longevity and age-related diseases - a systems biology
23 perspective. Aging (Albany NY) 3:1178-1191. doi: 100413
- 24 Tacutu R, Budovsky A, Fraifeld VE (2010) The NetAge database: a compendium of
25 networks for longevity, age-related diseases and associated processes. Biogerontology
26 11:513-522. doi: 10.1007/s10522-010-9265-8
- 27 Tatar M, Bartke A, Antebi A (2003) The endocrine regulation of aging by insulin-like
28 signals. Science 299:1346-1351. doi: 10.1126/science.1081447
- 29 Vijg J, Suh Y (2005) Genetics of longevity and aging. Annu Rev Med 56:193-212.
30 doi: 10.1146/annurev.med.56.082103.104617
- 31 Wang J, Duncan D, Shi Z, Zhang B (2013) WEB-based GEne SeT AnaLysis Toolkit
32 (WebGestalt): update 2013. Nucleic Acids Res 41:W77-83. doi: 10.1093/nar/gkt439
- 33 Warner HR (2005) Longevity genes: from primitive organisms to humans. Mech
34 Ageing Dev 126:235-242. doi: S0047-6374(04)00195-2
- 35 Wolfson M, Budovsky A, Tacutu R, Fraifeld V (2009) The signaling hubs at the
36 crossroad of longevity and age-related disease networks. Int J Biochem Cell Biol
37 41:516-520. doi: 10.1016/j.biocel.2008.08.026

1 Yanos ME, Bennett CF, Kaeberlein M (2012) Genome-Wide RNAi Longevity
2 Screens in *Caenorhabditis elegans*. *Curr Genomics* 13:508-518. doi:
3 10.2174/138920212803251391

4 Zhang J, Liu F (2014) Tissue-specific insulin signaling in the regulation of
5 metabolism and aging. *IUBMB Life* 66:485-495. doi: 10.1002/iub.1293

6 Ziv E, Hu D (2011) Genetic variation in insulin/IGF-1 signaling pathways and
7 longevity. *Ageing Res Rev* 10:201-204. doi: 10.1016/j.arr.2010.09.002

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

1 **Supplementary Figures and Tables**

2

3 **Suppl. Tables 1-4 and 6-7 are provided as additional Excel files.**

4

5 **Suppl. Table 5. “Public” and “private” enriched categories.** The table depicts the

6 most enriched categories for lists of proteins of longevity-associated genes (LAGs)

7 under different evolutionary conservation criteria (defined as the presence of

8 orthologs across a listed number of phyla).

Public/Private	<i>S. cerevisiae</i>	<i>C. elegans</i>	<i>D. melanogaster</i>	<i>M. musculus</i>
Public	LAGs with orthologs across at least 12 phyla	LAGs with orthologs across at least 12 phyla	LAGs with orthologs across at least 10 phyla	LAGs with orthologs across at least 10 phyla
	Ribosome and translation Mitochondria Citrate cycle (TCA cycle)	Ribosome and translation Mitochondria Oxidative phosphorylation NADH activity Development	FoxO signaling Development	DNA repair, especially Nucleotide excision repair
Private (for the indicated taxa)	LAGs conserved only in Fungi/Ascomycota	LAGs conserved only in Nematoda	LAGs conserved only in Arthropoda	LAGs conserved only in Chordata
	Autophagy	G-protein related	No enrichment	Neuroactive ligand-receptor interaction

9

10

11

12

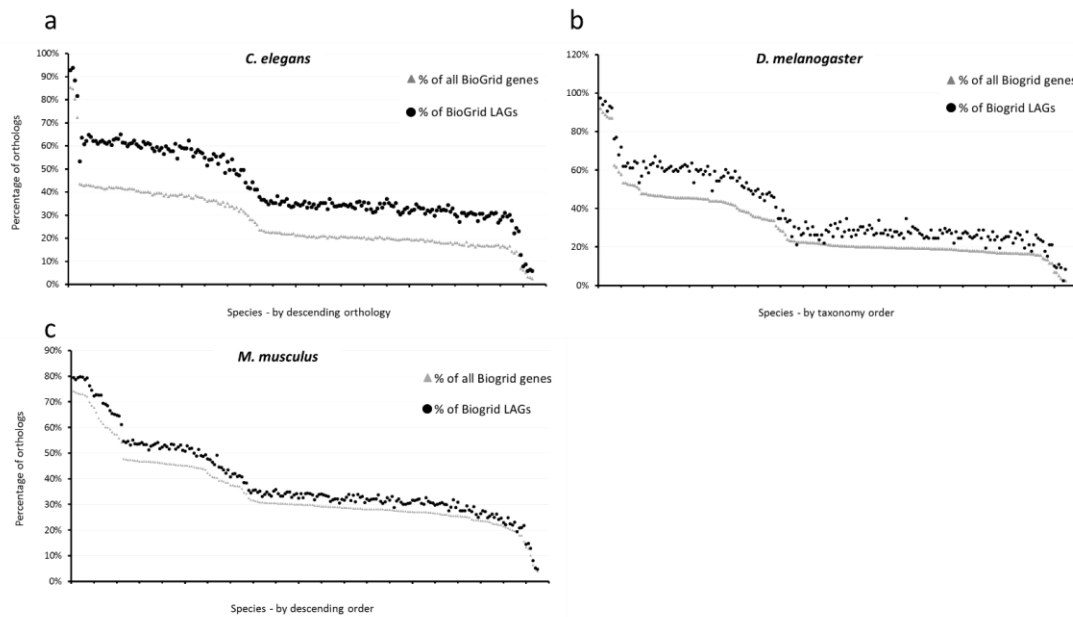
13

14

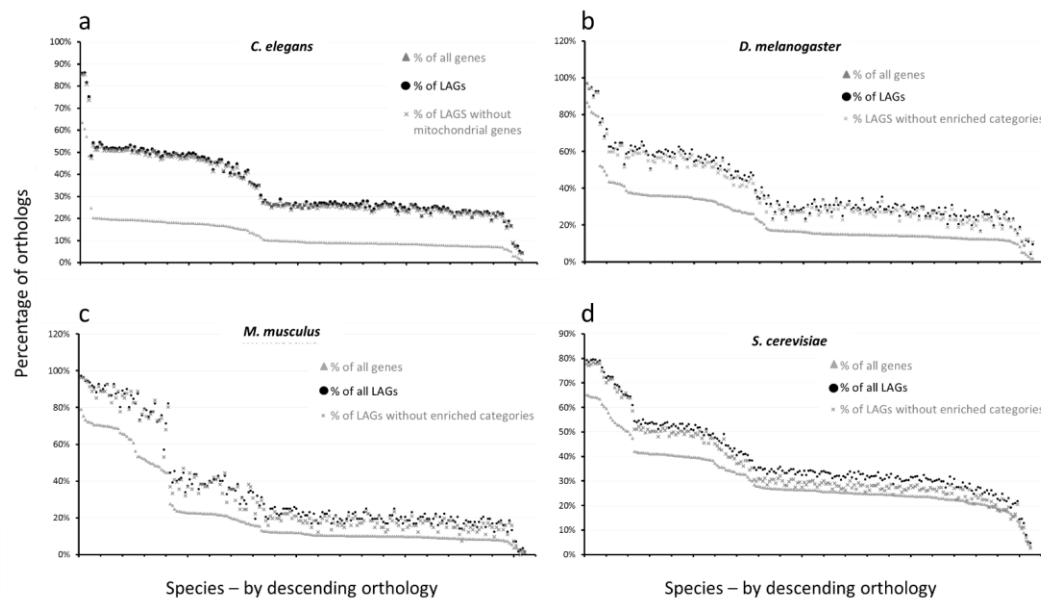
15

16

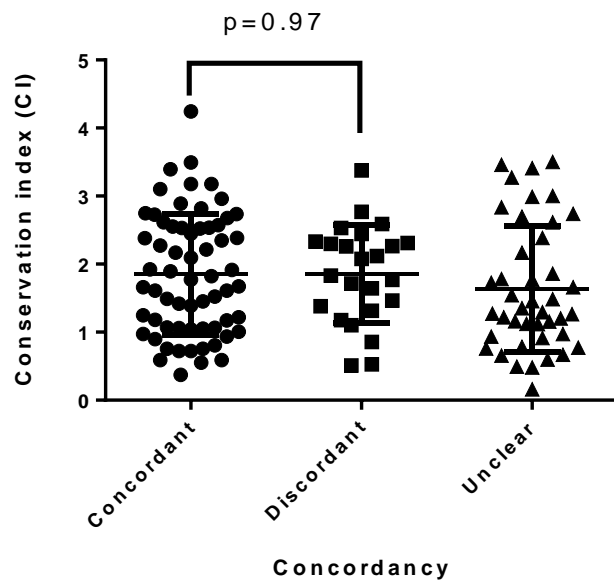
17



Suppl. Fig. 1. Percentage of Interactome LAG orthologs from the four model species. Each graph represents the LAGs discovered in the indicated model species. Each dot represents the percentage of orthologs between the model species and a different target species (total of 205 species from all kingdoms). Target species are ordered in descending order of orthology percentage as determined by the control. LAGs that are listed in BioGrid (black circle), entire proteome in BioGrid (grey triangle). **(a)** *C. elegans*, n = 3,865 for control and 343 for LAGs; **(b)** *D. melanogaster*, n = 8,026 for control and 118 for LAGs; **(c)** *S. cerevisiae*, n = 5,783 for control and 823 for LAGs.

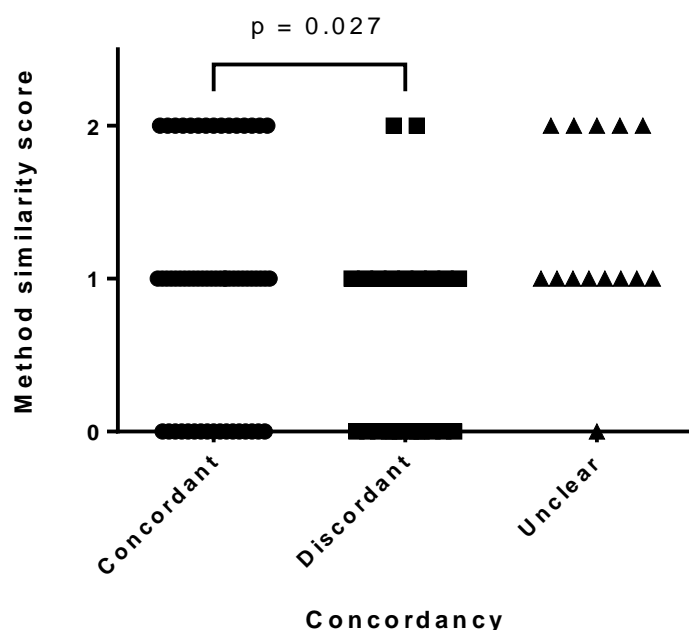


Suppl. Fig. 2. Percentage of LAG orthologs from the four model species after exclusion of proteins from enriched categories. Each graph represents the LAGs discovered in the indicated model species. Each dot represents the percentage of orthologs between the model species and a different target species (total of 205 species from all kingdoms). Target species are ordered in descending order of orthology percentage as determined by the control. LAGs (black circle), entire proteome (grey triangle) and LAGs after exclusion of proteins in enriched categories (grey x). **(a)** *C. elegans*, n = 20,325 for control, 733 for LAGs and 689 LAGs without mitochondrial genes; **(b)** *D. melanogaster*, n = 13,250 for control, 136 for LAGs and 122 for LAGs excluding enriched categories; **(c)** *M. musculus*, n = 21,895 for control, 112 for LAGs and 73 LAGs excluding enriched categories ; **(d)** *S. cerevisiae*, n = 6,590 for control, 824 for LAGs and 551 LAGs excluding enriched categories. All differences presented are highly significant ($p < 10^{-10}$).



1

2 **Suppl. Fig. 3. Conservation index (CI) compared to concordancy of longevity effects.** Each
3 dot represents a pairwise comparison between orthologs of LAGs that were tested for their
4 effect on longevity in more than one model species. The conservation index is the pairwise
5 alignment score normalized to the protein amino acid length.



6

7 **Suppl. Fig. 4. Method similarity score compared to concordancy of longevity effects.** Each
8 dot represents a pairwise comparison between orthologs of LAGs that were tested for their
9 effect on longevity in more than one model species. The method similarity score was
10 determined as: 0 = interventions of opposite directions (e.g. knockout and overexpression);
11 1 = intervention of the same direction but with varied methods (e.g. knockout and RNAi); 2 =
12 interventions that are identical or very close to identical (e.g. knockout and knockout).