

Infectious Reactivation of Cytomegalovirus Explaining Age- and Sex-Specific Patterns of Seroprevalence

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1 **Abstract**

2 Human cytomegalovirus is a herpes virus with poorly understood transmission
3 dynamics. We here provide quantitative estimates of the transmissibility of primary
4 infection, reactivation, and re-infection using age- and sex-specific antibody response
5 data. The data are optimally described by three distributions of antibody
6 measurements, i.e. uninfected, infected, and infected after reactivation/re-infection.
7 Estimates of seroprevalence increase gradually with age, such that at 80 years 73%
8 (95%CrI: 64%-78%) of females and 62% (95%CrI: 55%-68%) of males is infected, while
9 57% (95%CrI: 47%-67%) of females and 37% (95%CrI: 28%-46%) of males has
10 experienced a reactivation or re-infection episode. Merging the statistical analyses with
11 transmission models, we find that infectious reactivation is key to provide a good fit fit
12 to the data. Estimated reactivation rates increase from low values in children to 2%-6%

13 per year older women. The results advance a hypothesis in which adult-to-adult
14 transmission after infectious reactivation is the main driver of infection.

15 **Introduction**

16 Human cytomegalovirus (CMV) is a highly prevalent herpesvirus that infects between
17 30% and 100% of persons in populations throughout the world [1]. Usually thought to
18 be a relatively benign persistent infection, CMV is able to cause serious disease in the
19 immunocompromised and offspring of pregnant women with an active infection [2–5].
20 CMV also has been implicated in a variety of diseases in healthy persons [4, 6], and may
21 play a role in aging of the immune system [7–10], perhaps thereby reducing the
22 effectiveness of influenza vaccination in older persons [11–13].

23 Although the importance of CMV to public health is acknowledged, and even though
24 the development and registration of a vaccine has been declared a priority [14, 15], little
25 quantitative information is available on the transmission dynamics of CMV. At present,
26 the only population-level data derive from serological studies, aiming to uncover which
27 part of the population is infected at what age. These studies show that i) a sizable
28 fraction of infants is infected perinatally (before 6 month of age), ii) seroprevalence
29 increases gradually with age and is usually higher in females than in males, and iii) the
30 probability of seropositivity is associated with both ethnicity and socioeconomic status,
31 with non-western ethnicity and lower socioeconomic status being associated with higher
32 rates of seropositivity [1, 16–19].

33 Person-to-person transmission of CMV to an uninfected person can occur from a
34 primary infected person, or from a person who is experiencing a reactivation episode or
35 has been reinfected [4]. Here, we analyze data from a large-scale serological study to
36 obtain quantitative estimates of the relative importance of these transmission
37 routes [19]. We fit mixture models linked to age- and sex-specific transmission models
38 to the data to study the ability of different hypotheses explaining the serological data.
39 Specifically, we quantify the incidence and transmissibility of primary infection,
40 re-infection, and reactivation. Throughout, our premise is that measurements of
41 antibody concentrations provide information on whether or not a person has been
42 infected, and whether or not re-infection or reactivation have occurred. Persons with

43 low measurements are considered uninfected (susceptible), while persons with
44 intermediate and high antibody concentrations are infected and infected with
45 subsequent re-infection or reactivation, respectively.

46 We find that approximately 20% of infants is infected at the age of six months in the
47 Netherlands, that seroprevalence (i.e. the fraction of the population that is infected)
48 increases gradually with age to 60% and 70% in 80-year-old males and females, and that
49 the fraction of the infected population with high antibody concentration increases from
50 low values in children to 35% and 55% in elderly male and female persons, respectively.
51 Reactivation rates are found to be substantially higher in females than in males. The
52 analyses show that significant infectious reactivation in adults is necessary to explain
53 the serological data, supporting the notion that infectious reactivation is an important
54 driver of transmission.

55 The results have implications for control of CMV by vaccination, but also in a
56 broader context, as increased antibody concentrations are an indicator of T cell immune
57 memory inflation, impaired viral control, accelerated immunosenescence, and vascular
58 pathologies (see [10, 20] and references therein).

59 **Results**

60 **Prevalence estimation**

61 Fig 1 presents the data stratified by sex and age, with fits of the statistical model (Fig 7
62 for overview). The data and model fit show peaks at low antibody measurements (-2.9
63 U/ml and ≈ -2 U/ml), corresponding to uninfected persons (denoted by S, for
64 susceptible; Methods). In both sexes, there is a third peak at higher measurements ($1-3$
65 U/ml) that shifts to higher values with increasing age. This peak is composed of
66 persons who are infected (denoted by L, for latently infected) and persons who are
67 infected with high antibody concentrations (denoted by B, for boosted antibodies).
68 Using the estimated distributions for these classes, we find that classification of persons
69 as uninfected versus infected is near perfect (Youden index: 0.97), while classification of
70 persons with high antibody concentrations is good (Youden index: 0.71; Fig 2, Fig 8).
71 These results correspond well with the threshold for infection of the supplier by the

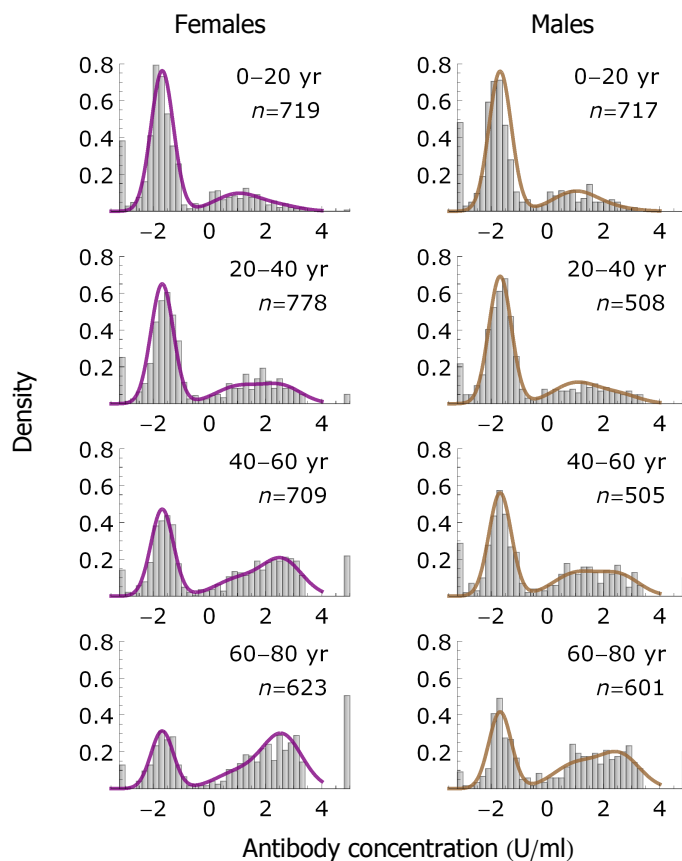


Fig 1. Data and model fit. Data (histograms) and model fit (lines) of antibody concentration measurements by age group and sex. Left- and right-hand panels show results for females (purple) and males (brown), respectively. The leftmost bars at -2.9 contain samples that are assumed uninfected, and the rightmost bars at 4.5 contain samples that are right censored (with concentration >3.41; Methods). Insets show the age group and number of samples.

72 assay (-0.8 U/ml), and show that our classification with three components is supported
73 by the data (i.e. has high probability of yielding an informed decision).

74 Fig 3 shows the estimated prevalences in females and males as a function of age.
75 The susceptible prevalence decreases gradually with age, from approximately 0.80 in
76 infants (females: 0.81, 95%CrI: 0.77-0.85; males: 0.80, 95%CrI: 0.76-0.84) to 0.27
77 (95%CrI: 0.22-0.34) and 0.38 (95%CrI: 0.32-0.45) at 80 years in females and males,
78 respectively. In both females and males the latently infected prevalence remains
79 approximately constant, ranging from 0.15 to 0.20 in females and from 0.18 to 0.28 in
80 males. In contrast, the prevalence of persons with increased antibodies increases
81 strongly with age, especially in females. In fact, the prevalence of persons with

82 increased antibodies increases from 0.09 (95%CrI: 0.06-0.13) at 20 years to 0.57
83 (95%CrI: 0.47-0.67) at 80 years in females, and from 0.04 (95%CrI: 0.03-0.07) to 0.37
84 (95%CrI: 0.28-0.46) in males. Hence, in older persons the prevalence of persons with
85 increased antibodies is 54% (or 20 per cent points) higher in females than in males.

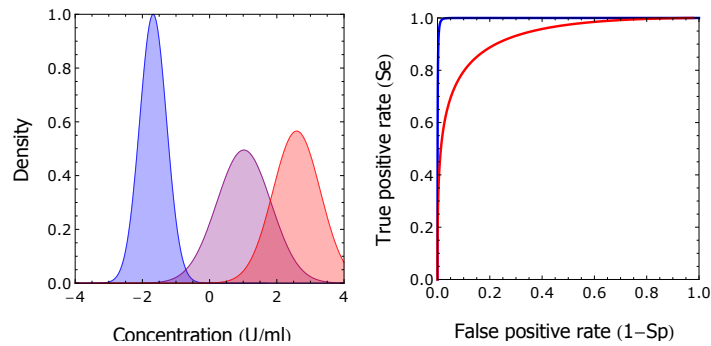


Fig 2. Classification of samples. Shown are the estimated mixture distributions using the parameter medians of the posterior distribution (left-hand panel; blue: susceptible; purple: infected); red: infected with increased antibody concentration), and receiver operating characteristic of binary classifications taking the estimated distributions as ground truth (right-hand panel). Maximal sum of sensitivity and specificity (Se+Sp) for classification of uninfected versus infected persons is 1.97 at antibody concentration -0.70 U/ml, with sensitivity 0.99 and specificity 0.98 (Figure S8). This value corresponds well with the threshold for infection of -0.8 U/ml suggested by the supplier of the assay. Maximal Se+Sp for classification of persons with increased antibody concentration is 1.70 at antibody concentration 1.81 U/ml, with sensitivity 0.84 and specificity 0.87 (Fig 8).

86 Of particular interest is the prevalence of infection in females of childbearing age, as
87 this group is at risk of transmission to the fetus or newborn. Using the above analyses,
88 we find that the prevalence of infection (i.e. the combined prevalence in the L and B
89 classes) is 0.30 (95%CrI: 0.27-0.33) in 20-year-old females and 0.42 (95%CrI: 0.39-0.46)
90 in 40-year-old females. If we combine these figures with the observation that
91 approximately 20% of children infected at six months of age, and that less than 5% of
92 children in the Netherlands in 2007 had a mother under 20 years or over 40 years, we
93 deduce that the probability of perinatal transmission could be between $0.20/0.42=0.48$
94 and $0.20/0.30=0.67$, with the exact figure depending on the distribution of ages at
95 which mothers give birth. In addition, one could envisage that the highest risk of
96 (severe) infection of the fetus or newborn is when mothers are infected or experience a
97 reactivation episode. The estimated rates at which susceptible females of 20 and 40

98 years are infected are 0.0055 per year (95%CrI: 0.0036-0.0077) and 0.0092 per year
99 (95%CrI: 0.0069-0.011) per year, respectively. The rates at which latently infected
100 females of 20 and 40 years are re-infected or experience a reactivation episode are of
101 similar magnitude, and are estimated at 0.0059 per year (95%CrI: 0.0038-0.0086) and
102 0.0093 per year (95%CrI: 0.0064-0.012), respectively. The overall rates of infection,
103 reactivation, and re-infection in 20 and 40 year-old females are given by the sum of the
104 above estimates, and are approximately 1% and 2% per year, respectively.

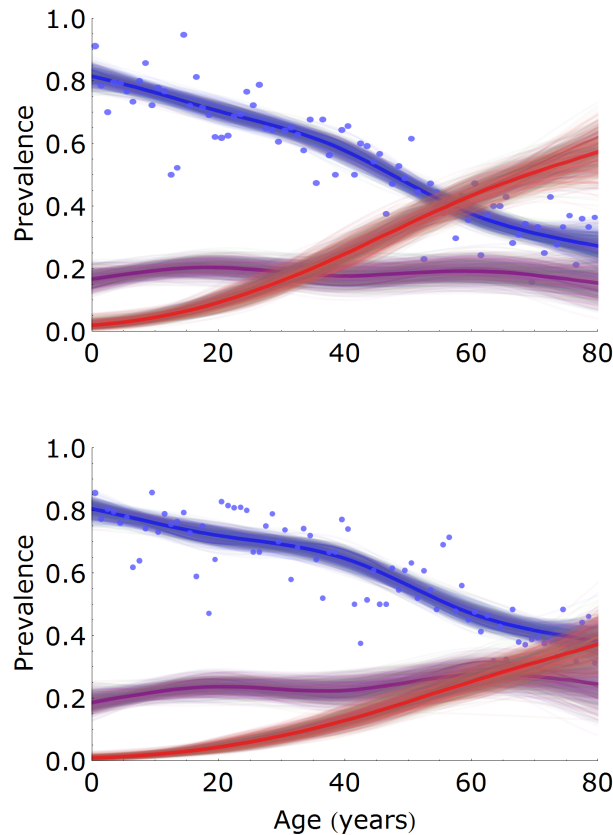


Fig 3. Estimation of age- and sex-specific prevalence. Prevalence estimates are presented for females (top) and males (bottom), and for classes of low (susceptible, blue), medium (latently infected, purple), and high (latently infected with increased antibodies, red) antibody measurements. Shown are 1,000 samples from the posterior distribution (thin lines) with posterior medians (bold lines). Dots indicate the fraction of samples in each year-class that would be classified as uninfected with the cut-off specified by the supplier of the assay. The number of samples per year-class is approximately 35 and 30 for females and males, respectively.

Table 1. Model selection of transmission scenarios.

Scenario	n	ℓ_{max}	ΔAIC
Full model	9	-7195.2	2.0
No re-infection	8	-7195.2	0
No reactivation	3	-7413.3	426.3
Reactivation/re-infection not infectious	8	-7233.8	75.2

For each of four model scenarios the number of parameters, maximal log-likelihood ℓ_{max} , and AIC difference with the best fitting model are given.

105 Estimation of reactivation and re-infection rates

106 Naive estimates of the reactivation and re-infection rates can be obtained by
107 transformation of the spline prevalence estimates (Methods), indicating that infectious
108 reactivation is required to explain the serological data (Fig 9). Estimation with the
109 transmission model confirms these findings, and shows that a model without
110 reactivation fits the data worse ($\Delta AIC=426.3$) than either a model with reactivation
111 and re-infection (full model), or a model with reactivation but no re-infection (Table 1;
112 Methods). A fourth model with reactivation and re-infection that is not infectious
113 ($\beta_2 \equiv 0$) performs better than the model without reactivation, but worse than either the
114 full model or the model with no re-infection ($\Delta AIC=75.2$).

115 The prevalences estimated with the mixture model and the corresponding maximum
116 likelihood estimates of the transmission models are given in Fig 10, and show that a
117 transmission model without reactivation overestimates the prevalence of infection,
118 especially in children. Overall, models without infectious reactivation have low empirical
119 support, while there is no decisive statistical evidence in favor of either a model with
120 infectious reactivation and infectious re-infection ($\Delta AIC= 2.0$) or a model with
121 infectious reactivation only (highest AIC; [21]). These results are supplemented and
122 supported in a Bayesian analysis of the latter two models (Methods). Using the
123 Watanabe Akaike information criterion (WAIC) that better gauges out-of-sample
124 predictive performance than AIC we find similar performance of both models
125 (WAIC=14,410.4 for the full model versus WAIC=14,409.1 for the model without
126 re-infection; [22, 23]).

127 Inspection of the parameter posterior medians of the two best-performing models
128 shows that both yield similar estimates of the transmissibility of primary infection
129 ($\widehat{\beta}_1=0.0015$ per year (95%CrI: $6.2 \cdot 10^{-5}$ -0.0072) for the full model and $\widehat{\beta}_1=0.0014$ per

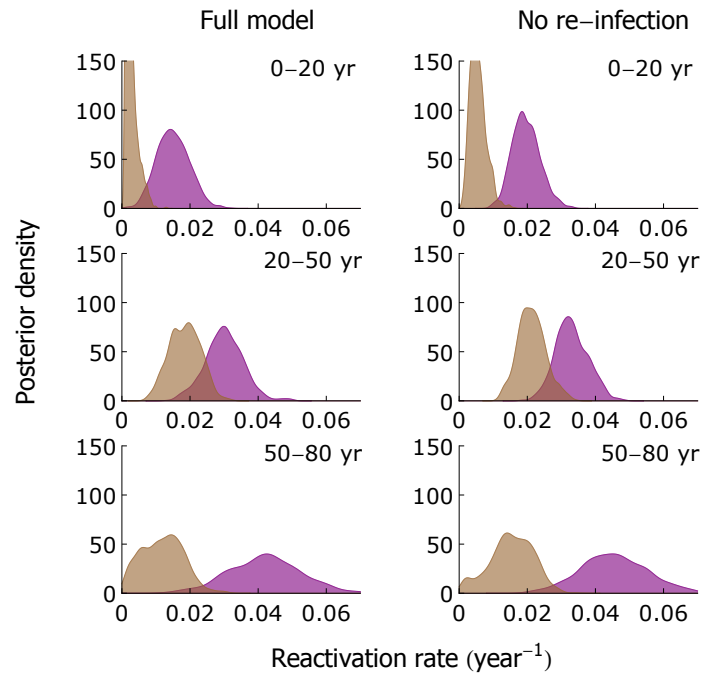


Fig 4. Estimation of age-specific reactivation rates. Shown are kernel-smoothed estimates of the reactivation rates in persons aged 0-20 years (top), 20-50 years (middle), and 50-80 years (bottom) in the full model (left-hand panels) and model without re-infection (right-hand panels; Table 1). Estimates for females and males are shown in purple and brown, respectively.

130 year (95%CrI: $4.9 \cdot 10^{-5}$ -0.0071) for the model without re-infection), and nearly identical
 131 estimates of the transmissibility of reactivation and re-infection ($\widehat{\beta}_2=0.052$ per year
 132 (95%CrI: 0.045-0.057) in both scenarios). In the model without re-infection the
 133 proportionality parameter governing the re-infection rate (the probability of re-infection
 134 in a contact that would lead to infection if the contacted person were uninfected) is zero
 135 by definition, and in the full model the proportionality parameter is dominated by the
 136 prior distribution $U(0, 1)$, indicating that it cannot be estimated with meaningful
 137 precision from the data ($\widehat{z}=0.33$; 95%CrI: 0.035-0.78).

138 Estimates of the reactivation rates are quantitatively close in models with
 139 reactivation (Fig 4). Reactivation rates generally increase with increasing age, and are
 140 substantially higher in females than in males. In the full model, the estimated
 141 reactivation rate is 0.015 per year (95%CrI: 0.0064-0.025) in 0-20 year-old females,
 142 which doubles to 0.030 per year (95%CrI: 0.019-0.041) in 20-50 year-old females, and
 143 then increases further to 0.042 per year (95%CrI: 0.021-0.062) in 50+-year-old females.

144 The corresponding reactivation rates in males are 0.0022 per year (95%CrI:
145 0.0010-0.0074), 0.019 per year (95%CrI: 0.0098-0.029), and 0.012 per year (95%CrI:
146 0.0015-0.023). In the model without re-infection these estimates are 0.019 per year
147 (95%CrI: 0.012-0.028), 0.033 per year (95%CrI: 0.024-0.043), and 0.045 per year
148 (95%CrI: 0.026-0.064) in females, and 0.0052 per year (95%CrI: 0.0019-0.010), 0.021 per
149 year (95%CrI: 0.014-0.030), and 0.015 per year (95%CrI: 0.0016-0.027) in males. Hence,
150 estimates of reactivation rates are higher and slightly more precise in the model without
151 re-infection than in the model with re-infection.

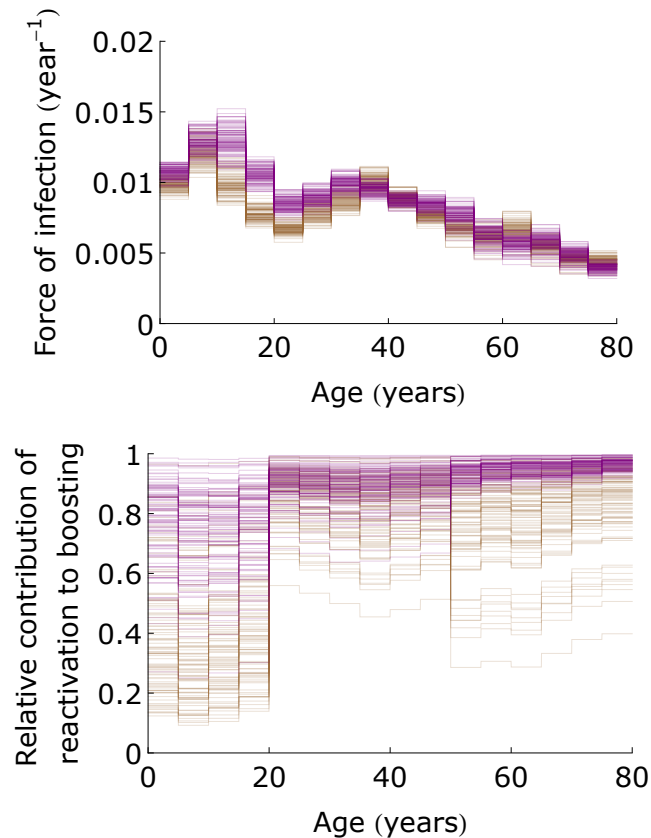


Fig 5. The force of infection and contribution of reactivation to antibody boosting. Shown are estimates of the forces of infection in females and males (top; purple: females; brown: males) and relative contributions of reactivation to antibody boosting ($\rho^i(a) / (\rho^i(a) + z\lambda^i(a))$ with $i \in \{\varphi, \sigma\}$) (bottom) in the full model as a function of age. Both panels show 100 samples from the posterior distribution.

152 In both models with reactivation, estimates of the force of infection increase from
153 approximately 0.01 per year in the youngest age group to 0.012-0.014 per year in 10-15
154 year-old girls (Fig 5). Owing to the slightly higher contact rates in females than in men,

155 the estimated force of infection is usually a couple percentage points higher in females
156 than in males in the age groups 10-25 years [24]. In older age groups, estimates of the
157 forces of infection decrease to lower values (0.005-0.01 per year). Noteworthy, the
158 extreme age-specific differences in the force of infection usually observed for directly
159 transmitted infectious diseases, with high infection rates in children and much lower
160 rates in adults, are much less pronounced here due to infectious reactivation combined
161 with age-assortative mixing (Fig 10) [24–26].

162 Further, estimates of re-infection rate ($z\lambda(a)$) are considerably smaller than
163 estimates of the reactivation rates ($\rho^{\varnothing}(a)$ and $\rho^{\sigma}(a)$) because the estimated force of
164 infection ($\lambda(a)$) is small in all age groups (Fig 5). Hence, re-infection contributes little
165 to boosting of the antibody concentrations in those age groups where most of the
166 boosting occurs (>20 years; Fig 3).

167 Discussion

168 Our study of population-wide serological data shows that IgG antibody concentrations
169 contain a wealth of information on the transmission dynamics of CMV. Specifically, the
170 analyses reveal that (i) the prevalence of CMV increases gradually with age such that at
171 old age the majority of persons in the Netherlands are infected; (ii) except for the very
172 young, prevalence of CMV is systematically higher in females than in males. This is
173 mainly due to a higher incidence of infection in adult women than in adult men of
174 similar age; (iii) antibody concentrations in seropositive (i.e. infected) persons increase
175 monotonically with age, especially in women; (iv) the above findings (i)-(iii) cannot be
176 explained by simple transmission models in which only primary infection is infectious.
177 This is caused by the fact that transmissibility of primary infection determines the rate
178 at which age-specific prevalence increases; if transmissibility of primary infection would
179 be high then a high prevalence of infection is expected in children (Fig 10). In other
180 words, the fact that seroprevalence increases gradually with age puts an upper bound on
181 the force of infection, and this in turn constrains the transmissibility of primary
182 infection to low values.

183 While aforementioned findings (i)-(iii) have been noticed before in other settings ([1]
184 and references therein, [19]), our analyses are the first to provide precise estimates using

185 a large population sample. Moreover, the analyses advance a transmission hypothesis
186 based on the notion that reactivation contributes significantly to the transmission
187 dynamics of CMV. Since prevalence of infection has been shown to increase in a gradual
188 manner in several other studies [1], this explanation may not be restricted to the Dutch
189 situation, but hold in general. Underpinning this hypothesis, next to the well-known
190 observations of shedding of CMV in breast milk and cervical material in the third
191 trimester of pregnancy [27–29], detectable virus also has been found in some healthy
192 persons under 70 years and in all persons over 70 years in one study [31], while in
193 another study CMV DNA has been detected in urine of the majority of older
194 persons [32]. These findings add to our belief that infectious reactivation may be a
195 plausible explanation for the patterns of infection observed in serological data.

196 The main implication is that the majority of CMV infections may not be caused by
197 transmission among children after primary infection, even though levels of shedding can
198 be high in infants [28,33], but rather by older persons who go through a reactivation
199 episode. As a result, infectious reactivation is expected to be an important driver of
200 CMV transmission in the population. This contrasts with common childhood diseases
201 such as measles, mumps, rubella, and pertussis, for which infection in unvaccinated
202 populations generally occurs at a young age and children are the drivers of transmission.
203 It also contrasts with other herpes viruses such as varicella zoster virus and Epstein-Bar
204 virus for which well over 50% of the population is infected at the age of 10 years [25]. It
205 may be comparable with other herpes viruses such as HSV1 and HSV2, which show a
206 slowly increasing age-specific seroprevalence [34]. A corollary is that primary CMV
207 infection alone is unlikely to be able to maintain sustained transmission in the
208 population. In fact, further analyses based on the current parameter estimates suggest
209 that primary infection coupled with infectious reactivation would not be sufficient to
210 ensure persistence, as the estimated basic reproduction number is smaller than the
211 threshold value 1 (Methods; Fig 12). This, in turn, indicates that perinatal transmission
212 is required for persistence.

213 With infectious reactivation and perinatal infection being putative drivers of
214 transmission, it is to be expected that elimination by vaccination may prove more
215 difficult than for directly transmitted pathogens, as it will require the pool of latently
216 infected persons to dwindle to zero by demographic turnover. This can take up to the

217 lifetime of one generation, and perhaps more if vaccination cannot prevent perinatal
218 transmission to infants who are too young for vaccination. Thus, a question is whether
219 vaccination formulations and strategies exist that minimize the probability of
220 transmission to young infants. This is all the more of importance as a main source of
221 morbidity is by congenital infection, and the timescale on which reductions in congenital
222 disease are expected determines the projected health impact of vaccination [35]. In this
223 context, next to the ability of a vaccine to prevent infection it may perhaps be equally
224 important that a vaccine is able to reduce the probability of reactivation. Such
225 reductions are likely mediated by T-cell responses of the host, and several (but not all)
226 vaccines under development are expected to induce boosting of T-cell immune
227 responses [36–38].

228 A number of limitations and assumptions deserve scrutiny. First, the transmission
229 model analyses assume that the population is in endemic equilibrium. For a single
230 cross-sectional data set such as considered in the present study this assumption is
231 unavoidable if one does not want to introduce additional parameters that cannot be
232 estimated by the data. Reassuringly, the patterns of infection present in the serological
233 data have been found in several serological studies carried out in high-income countries
234 over the past decades [1]. Also, no systematic patterns of increasing or decreasing
235 seroprevalence over time have been found, and this is further reason to believe that
236 there have not been major changes in the epidemiology of CMV over time [1]. Second,
237 we assume that antibody measurements not only give information on CMV infection
238 status, but also whether or not reactivation/re-infection have taken place.
239 Unfortunately, there is no direct empirical evidence available confirming or falsifying
240 this assumption, and this is an area where in-depth comparison of the infection and
241 immune status of persons with low and high antibody concentrations is urgently needed.
242 Third, the analyses assume that person-to-person transmission is proportional to
243 observed human contact patterns (Fig 11; [24,39]). Although this assumption is
244 commonly made and has met with considerable success (e.g., [39–42]), it is conceivable
245 that transmission of CMV does not abide by the social contact hypothesis, and that a
246 more complex contact structure would be able to explain the patterns of seroprevalence
247 in a simple transmission model. To investigate the impact of the contact structure, we
248 have re-analyzed all transmission model with a uniform contact structure, and found

249 that a model with infectious reactivation provides the best fit to the data (not shown).
250 By extension, it also remains a possibility that the patterns of infection are generated
251 by a complex pattern of susceptibility increasing with age. Again, evidence for or
252 against this possibility is lacking.

253 Throughout, the analyses are based on the observation that, at the population level,
254 CMV IgG antibody concentrations are well-described by three distributions. We have
255 exploited this observation to build a transmission model with three classes, pertaining
256 to persons who are uninfected, infected, or infected after reactivation or re-infection. In
257 this model, a person experiences at most one reactivation or re-infection event during its
258 lifetime. In reality, it is more likely that such reactivation and/or re-infection events
259 may occur more often. Unfortunately, it is not possible to statistically identify the
260 parameters in models that would include multiple reactivations/re-infection events. As
261 a result, transitions from the infected class to the infected class with increased
262 antibodies may in fact be the result of multiple underlying reactivation or re-infection
263 events, and the infectiousness parameter of reactivation and re-infection should be
264 interpreted as a compound parameter describing the overall impact of multiple
265 reactivations and re-infections. This may help explain why estimated infectiousness of
266 primary infection is much lower than estimated infectiousness after reactivation or
267 re-infection, even though it is known that prolonged and high-level virus shedding can
268 occur in bodily fluids after primary infection in children [28,29].

269 **Materials and Methods**

270 **Study design**

271 The analyses make use of sera from the PIENTER2 project, a cross-sectional
272 population-based study carried out in the Netherlands in 2006-2007. Details have been
273 published elsewhere [19]. Briefly, 40 municipalities distributed over five geographic
274 regions of the Netherlands were randomly selected with probabilities proportional to
275 their population size, and an age-stratified sample was drawn from the population
276 register. A total of 19,781 persons were invited to complete a questionnaire and donate
277 a blood sample. Serum samples and questionnaires were obtained from 6,382

278 participants. To exclude the interference of maternal antibodies, we restrict analyses to
279 sera from persons older than 6 months (6,215 samples). We further select Dutch persons
280 and migrants of Western ethnicity to preclude confounding by ethnicity (5,179 samples)
281 and stratify the data by sex [19], yielding 2,842 and 2,337 samples from female and male
282 participants, respectively. The data are available at GitHub
283 (github.com/mvboven/cmV-serology).

284 **Ethics**

285 The study was approved by the Medical Ethics Testing Committee of the foundation of
286 therapeutic evaluation of medicines (METC-STEG) in Almere, the Netherlands (clinical
287 trial number: ISRCTN 20164309). All participants or their legal representatives had
288 given written informed consent. Details of the study are given elsewhere [30].

289 **Antibody assay**

290 We use the ETI-CYTOK-G PLUS (DiaSorin, Saluggia, Italy) Elisa to detect
291 CMV-specific IgG antibodies. The assay yields continuous measurements (henceforth
292 called 'antibody concentration') and has an upper limit of 10 units/ml in our test
293 setting. A small number of samples is right-censored at this limit (140 persons).
294 According to the provider of the assay, samples with measurement lower than 0.4
295 units/ml should be classified as uninfected, while samples with measurement ≥ 0.4
296 units/ml should be classified as infected.

297 **Mixture model**

298 The data are analyzed using a mixture model with sex- and age-specific mixing
299 functions. A Box-Cox transformation with parameter $\psi=0.3$ yields a distribution of
300 antibody measurements of samples with low antibody concentration that is
301 approximately Normal, and we henceforth base all analyses on the transformed
302 measurements (denoted by U/ml). The estimated prevalences are robust to this
303 transformation, which merely serves to ensure that the data are well-described by
304 Normal mixture distributions (not shown). We distinguish three distributions,
305 describing samples with low (susceptible, S), intermediate (latently infected, L), and

306 high (increased antibodies, B) antibody concentrations. The L and B distributions are
307 modeled by a Normal distribution with means and standard deviations independent of
308 age and sex, and the S distribution is modeled by a mixture of a spike and a normal
309 distribution (see below).

310 Right-censoring is applied to the 140 samples above the upper limit of 3.41 U/ml (10
311 units/ml on the original scale). As there appears a spike at -2.91 U/ml (0.0001 units/ml
312 on the original scale) in the data (263 persons), we model the S component by a
313 mixture of a spike at this value and a Normal distribution (i.e. an inflated normal
314 distribution). In this way, samples with concentration at the spike belong to the
315 susceptible component with probability 1.

316 We model the probability of each of the three outcomes in terms of log-odds, taking
317 the probability of being in the S component as reference. This allows us to write the
318 log-odds of being in component L or B as linear functions of age and sex. The design
319 matrix of the resulting multinomial logistic model consists of natural cubic splines with
320 interior knots at 20, 40 and 60 years and boundary knots at 0 and 80 years. Hence, the
321 mixing functions (prevalences) have flexible shape, which allows these to be optimally
322 informed by the data. In the results, sex is put in the model as main effect, as analyses
323 show no noticeable improvement in fit when including age by sex interaction (not
324 shown).

325 We estimate parameters in a Bayesian framework using R and JAGS [43, 44].
326 Non-informative Normal prior distributions are set on the means of the three component
327 distributions ($\mathcal{N}(0, 0.001)$). Label switching is prevented by prior ordering of the means.
328 The precisions of the components are given flat Gamma prior distributions
329 ($\Gamma(0.5, 0.005)$). The spline parameters are also given non-informative Normal prior
330 distributions ($\mathcal{N}(0, 0.001)$). We apply a QR-decomposition to the design matrix to
331 improve mixing and run 10 MCMC chains in parallel, yielding a total of 10,000 samples.
332 Although mixing of the unthinned samples is already satisfactory, we apply an
333 additional 1/10 thinning, giving a total of 1,000 samples from the posterior distribution.

334 **Classification**

335 The distributions characterizing the subpopulations with low (susceptible, S),
336 intermediate (latently infected, L), and high (increased antibodies, B) antibody
337 concentrations allow assignment of probabilities to individual samples. For any single
338 observed sample we can calculate the probability that the corresponding person is
339 susceptible, latently infected, or latently infected with increased antibodies. In detection
340 theory, the specificity (the probability of correctly classifying a negative subject) and
341 sensitivity (the probability of correctly classifying a positive subject) are used to
342 characterize the power of a detection procedure. The relation between sensitivity and
343 specificity may be graphed with antibody concentration specifying a cut-off for binary
344 classification as a parameter in a receiver operating characteristic (ROC)
345 graph [42, 45, 46]. Here, we investigate the ability of the mixture analyses to distinguish
346 uninfected (S) from infected (L+B) samples, and to distinguish samples with increased
347 antibody concentration (B) from uninfected and latently infected samples (S+L). The
348 results are laid down in Fig 2 and Fig 8.

349 **Transmission model**

350 Next to the statistical analysis using mixture models, we analyze the data with
351 transmission models. The transmission models take the estimated mixture distributions
352 or, more precisely, the medians of the posterior distribution as input. In line with the
353 above, we focus on a sex- and age-structured model in which persons are
354 probabilistically classified in one of three classes, viz. uninfected (S), latently infected
355 (L), and latently infected after reactivation or re-infection (B). As the infectious period
356 is short relative to the lifespan of the host (weeks versus decades) we do not explicitly
357 model the infectious periods, and assume that the transitions from the S to the L and
358 from the L to the B classes occur instantaneous (the short-disease approximation, [47]).
359 Further, we focus on the endemic equilibrium of the transmission model so that all
360 variables are time-independent [47, 48]. Fig 6 shows a schematic of the model. For sexes

361 $i \in \{\varphi, \sigma\}$, the age-dependent differential equations are given by

$$\begin{aligned} \frac{dS^i(a)}{da} &= -\lambda^i(a)S^i(a) \\ \frac{dL^i(a)}{da} &= \lambda^i(a)S^i(a) - (\rho^i(a) + z\lambda^i(a))L^i(a) \\ \frac{dB^i(a)}{da} &= (\rho^i(a) + z\lambda^i(a))L^i(a), \end{aligned} \quad (1)$$

362 with forces of infection

$$\lambda^i(a) = \sum_{j \in \{\varphi, \sigma\}} \int_0^M c^{ij}(a, a') (\beta_1 \lambda^j(a') S^j(a') + \beta_2 (\rho^j(a') + z\lambda^j(a')) L^j(a')) da'. \quad (2)$$

363 In Eqs (1)-(2), $z\lambda^j(a)$ and Eq $\rho^j(a)$ are the age-specific re-infection and reactivation
 364 rates, z is the susceptibility to re-infection of latently infected persons relative to the
 365 susceptibility of uninfected persons ($0 \leq z \leq 1$), $c^{ij}(a, a')$ represents the contact rate
 366 between persons of age a' and sex j , and those of age a and sex i [24,39], β_1 and β_2 are
 367 proportionality parameters determining the transmissibility of primary infection and
 368 reactivation/re-infection, and M is the maximum age. As the data do not extend
 369 beyond 80 years we take $M = 80$ years. Notice that $\lambda^j(a)S^j(a)$ and
 370 $(\rho^j(a) + z\lambda^j(a))L^j(a)$ are the incidence of primary infection and the incidence of
 371 reactivation and re-infection, so that $\beta_1 \lambda^j(a)S^j(a)$ and $\beta_2 (\rho^j(a) + z\lambda^j(a))L^j(a)$ are
 372 the infectious output generated by primary infection and reactivation/re-infection,
 373 respectively [47].

374 As in earlier studies, contact rates (contact intensities in the notation of [24] are
 375 hard-wired into the model using data on social contact patterns, thereby adopting the
 376 social contact hypothesis [24,25,39]. Here we use the mixing matrix of an analysis of
 377 the data from the Netherlands gathered in 2006/2007 with demographic composition of
 378 the Dutch population in 2007 [24]. See Fig 11 and GitHub
 379 (github.com/mvboven/cm-v-serology) for the contact matrix and demographic data.

380 The differential equations can be solved in terms of the forces of infection using the
 381 variation of constants method. Here we assume, based on results of the mixture model,
 382 that a non-negligible fraction of infants is infected in the first six months of life and the
 383 fraction infected is equal in female and male infants [19]. Hence, we have
 384 $S^\varphi(0) = S^\sigma(0) = S_0$, $L^\varphi(0) = L^\sigma(0) = 1 - S_0$, and $B^\varphi(0) = B^\sigma(0) = 0$ as initial

$$S^i(a) \xrightarrow[\text{infection}]{\lambda^i(a)} L^i(a) \xrightarrow[\text{reactivation/re-infection}]{\rho^i(a) + z \lambda^i(a)} B^i(a)$$

Fig 6. Schematic of the model. $S^i(a)$ denotes the age-specific proportion of uninfected persons of sex i ($i \in \{\varphi, \sigma\}$), $L^i(a)$ represents the age-specific proportion of latently infected persons, and $B^i(a)$ is the age-specific proportion of infected persons with increased antibody concentration. The infection and re-infection rates are given by $\lambda^i(a)$ and $z\lambda^i(a)$, and the reactivation rates are given by $\rho^i(a)$.

385 conditions, and the solution of (1) is given by

$$\begin{aligned} S^i(a) &= S_0^i e^{-\int_0^a \lambda(\tau) d\tau} \\ L^i(a) &= (1 - S_0^i) e^{-\int_0^a \rho(\tau) + z\lambda(\tau) d\tau} + \\ &S_0^i \int_0^a \lambda(\tau) e^{-\int_0^\tau \lambda(\tau') d\tau' - \int_\tau^a \rho(\tau') + z\lambda(\tau') d\tau'} d\tau . \end{aligned} \quad (3)$$

386 Insertion of Eq (3) in Eq (2) yields two integral equations for the age-specific forces of
 387 infection in females and males [25, 26, 49, 50]. These equations cannot be solved
 388 explicitly in general. It is possible, however, to solve the equations for specific functions.
 389 Here we assume that transition rates and contact rates are constant on predefined
 390 age-intervals, and insert the explicit solution of Eq (3) in terms of force of infection in
 391 Equation (2) (see below).

392 Naive estimation

393 Before turning attention to estimation based on the full transmission model, we would
 394 like to remark that naive estimates of the force of infection and reactivation rate that do
 395 not take the infection feedback (i.e. Eq 2) into account can be obtained by
 396 transformation of the spline prevalence estimates. Specifically, rearranging the model
 397 equations (Eq (1)) yields the following expressions for the force of infection and
 398 reactivation rate

$$\begin{aligned} \lambda(a) &= -\frac{d}{da} \log(S(a)) \\ \rho(a) &= \frac{dB(a)}{da} \frac{1}{L(a)} - z\lambda(a) , \end{aligned} \quad (4)$$

399 where superscripts denoting sex have been dropped for notational convenience. Since
 400 the prevalences $S(a)$, $L(a)$, and $B(a)$ are known (the spline estimates) and can be

401 differentiated, this immediately yields an estimate of the force of infection $\lambda(a)$ and
 402 subsequently also an estimate of the reactivation rate $\rho(a)$ (if z is assumed known).
 403 Notice that these estimates are not necessarily compatible with the transmission model.
 404 In particular, the rates so estimated are not necessarily positive (Fig 9).

405 Assuming that re-infection is negligible ($z=0$), we find that estimates of the forces of
 406 infection are low up to the age of 40 years in both females and males (generally <0.01
 407 per year), increase to higher values in adults aged 40-60 years (0.01-0.03 per year), and
 408 are variable in older adults (Fig 9). Estimates of the reactivation rates increase from low
 409 values in children to 0.03-0.08 per year in 40-year-old females, and to approximately
 410 0.02 per year in 40-year-old males. At older ages, estimates of the reactivation rates are
 411 variable, and have the tendency to drop to slightly lower values. If we take the
 412 alternative extreme that primary infection and re-infection occur at identical rates
 413 ($z = 1$), estimates of the reactivation rates are lower but the general pattern of
 414 substantial reactivation in adults remains (Fig 9).

415 Numerical solution of the forces of infection

416 For statistical analysis based on the full transmission model (Eqs 1)-(2) we assume that
 417 reactivation and contact rates are constant in certain predefined age-intervals. From
 418 Eq (2), it then follows that the force of infection is piecewise constant as well.
 419 Throughout, we consider age intervals of fixed size $\Delta a = 5$ years, so that the limits of
 420 the $n = M/\Delta a = 16$ age classes are defined by the vector $\mathbf{a} = (0, \Delta a, 2\Delta a, \dots, n\Delta a)$.
 421 Hence, the j -th class ($j = 1, \dots, n$) contains all persons with age in the interval
 422 $[a_{[j]}, a_{[j+1]})$, where $a_{[j]}$ denotes the j -th element of \mathbf{a} . Subsequently, the forces of
 423 infection $\lambda^i(a)$ and reactivation rates $\rho^i(a)$ are replaced by their counterparts λ_j^i and ρ_j^i .
 424 Similarly, $S^i(a)$, $L^i(a)$, and $B^i(a)$ are replaced by S_j^i , L_j^i , and B_j^i . Insertion in Eq (3)
 425 and integrating over the (constant) rates yields

$$\begin{aligned}
 S_j^i &= S_0 e^{-\Delta a \sum_{k=1}^j \lambda_k^i} \\
 L_j^i &= (1 - S_0) e^{-\Delta a \sum_{k=1}^j \rho_k^i + z \lambda_k^i} + \\
 & S_0 \sum_{k=1}^j \lambda_k^i \frac{e^{-\Delta a (\rho_k^i + z \lambda_k^i)} - e^{-\Delta a \lambda_k^i}}{(1 - z) \lambda_k^i - \rho_k^i} \times \\
 & e^{-\Delta a (\sum_{l=1}^{k-1} \lambda_l^i - \sum_{l=k+1}^j \rho_l^i + z \lambda_l^i)},
 \end{aligned} \tag{5}$$

426 where $i \in \{\varphi, \sigma\}$ and $B_j^i = 1 - S_j^i - L_j^i$. Insertion of Eq (5) in Eq (2) and making use of
427 the fact that the cumulative incidences of infection and reactivation/re-infection in age
428 class j are given by $\int_{a_{[j]}}^{a_{[j+1]}} \lambda^i(a) S^i(a) da = S^i(a_{[j]}) - S^i(a_{[j+1]})$ and
429 $B^i(a_{[j+1]}) - B^i(a_{[j]})$, yields $2n = 32$ equations ($n = 16$ per sex) for the forces of
430 infection. These equations can be solved numerically. Here we use a Quasi-Newton
431 (secant) method to solve the equations.

432 Likelihood and estimation

433 With transition rates and forces of infection at hand, we calculate the log-likelihood of
434 the data. Here, the log-likelihood of each observation is given by a mixture distribution.
435 For instance, the log-likelihood contribution of a sample with antibody measurement c
436 in a person of sex i and age a is given by $\log(S^i(a)f_S(c) + L^i(a)f_I(c) + B^i(a)f_B(c))$,
437 where $S^i(a)$, $L^i(a)$, and $B^i(a)$ are the age specific prevalences in sex i , and $f_S(c)$, $f_L(c)$,
438 and $f_B(c)$ are the densities of the mixture distributions at antibody concentration c .

439 To reduce computation times and enable better comparison of the prevalence
440 estimates of the statistical and mechanistic models, we take the posterior medians of the
441 component distributions of the logistic model as inputs in the transmission model.
442 Hence, the transmission model takes the component distributions as given and provides
443 estimate the prevalences via the transmission and reactivation rates. Here, based on
444 preliminary analyses, reactivation rates are modeled by piecewise constant functions
445 with steps at 20 and 50 years, i.e. with constant rates on the intervals $[0, 20)$, $[20, 50)$,
446 and $[50, 80)$ years. Hence, the reactivation rates are characterized by three parameters
447 in each sex, viz. $\rho_{[0,20)}^i$, $\rho_{[20,50)}^i$, and $\rho_{[50,80)}^i$ ($i \in \{\varphi, \sigma\}$).

448 We employ a Markov chain Monte Carlo (MCMC) method to obtain maximum
449 likelihood (ML) estimates in a pre-screening of models as well as Bayesian estimates for
450 a subset of models that perform well in the pre-screening. For this purpose, we take
451 non-informative (improper) uniform prior distributions for all parameters. Steps in the
452 estimation procedure are as follows. First, for new parameters β_1 , β_2 , z , or ρ_x^i we solve
453 the $2n = 32$ equations for the forces of infection λ_k^i . Second, we use the forces of
454 infection to calculate the age-specific prevalences $S^i(a)$, $L^i(a)$, and $B^i(a)$. Third, the
455 age-specific prevalences are used to calculate the log-likelihood. Fourth, the new

456 parameters are accepted or rejected, and the above steps are repeated. Throughout,
457 updating of parameters is based on a single component random-walk Metropolis
458 algorithm using Normal proposal distributions with the current value as mean and
459 standard deviations tuned to achieve acceptance ratios in the 20%-50% range. In the
460 Bayesian analyses, output is generated for 5,000 cycles after a burn-in of 1,000, and a
461 thinned sample of 1,000 is used for analysis. Convergence of chains is assessed visually.
462 Parameter estimates are represented by posterior medians, and bounds of 95% credible
463 intervals are given by 2.5 and 97.5 percentiles. For model selection, we report the
464 Watanabe Akaike Information Criterion (WAIC) (using p_{WAIC2} , see [22,23]). We have
465 performed a pre-screening of models using ML since the Bayesian analysis does not
466 always yield properly converged chains for poorly fitting models. Here, we use a
467 simulated annealing method in which the Metropolis acceptance probability p is
468 replaced by $p^{1/T}$, where $T = 1/\sqrt{t}$ is the temperature at iteration t . In this manner,
469 proper ML estimates can be obtained within hours (~ 500 iterations). Comparison of
470 models in the pre-screening is based on the Akaike Information Criterion (AIC). The
471 analyses are performed with Mathematica 10.0.

472 In the main text we consider a suite of simplifications of the full model specified by
473 Equations (1)-(2). In the simplification we assume that (i) there is no re-infection
474 ($z = 0$), (ii) there is no reactivation ($\rho_x^i = 0$ for all i and x), or (iii) reactivation and
475 re-infection are not infectious ($\beta_2 = 0$).

476 **Reproduction numbers**

477 With estimates of key parameters at hand we can further our understanding of the
478 transmission dynamics of CMV, e.g., by calculation of the basic and type reproduction
479 numbers [47]. These reproduction number give insight in the drivers of transmission,
480 and also in the control effort required for elimination or eradication. Full calculation is
481 not straightforward, as CMV can be transmitted after primary infection, after
482 reactivation, and perinatally from mother to offspring. While our analyses have yielded
483 estimates of the former two, formal estimates of perinatal transmission are still lacking.
484 Furthermore, calculation of reproduction numbers implicitly assume a stable host
485 demography and known female fertility distribution function [51]. To obtain partial

486 insight of the impact of the transmission by sex, age and transmission route, we assume
487 type 1 demography (everybody lives exactly to the age of $M = 80$ years), and calculate
488 reproduction numbers in the presence of direct transmission and transmission after
489 reactivation but without perinatal transmission. Central in the calculations are the
490 so-called kernels $k^{ij}(a, \tau)$ of the next-generation operator, which determine the number
491 of cases of sex i and age a generated by an infected person of sex j and age τ . In our
492 case, a straightforward calculation [47] shows that $k^{ij}(a, \tau)$ is given by

$$k^{ij}(a, \tau) = \beta_1 c^{ij}(a, \tau) + \beta_2 \int_{\tau}^M c^{ij}(a, \tau') \rho^j(\tau') e^{-\int_{\tau'}^{\tau} \rho^j(v) dv} d\tau'$$

493 Discretizing the above kernel while making the simplifying assumption that only a
494 single event can happen in any one age group, we obtain approximate estimates of the
495 sex- and age-specific reproduction numbers, collected in the so-called next-generation
496 matrix (Fig 12). Subsequently, we derive estimates of the basic reproduction number R_0
497 and the reproduction numbers between the sexes R^{ij} as the dominant eigenvalues of the
498 next-generation matrix and the sex-specific next-generation matrices, respectively.
499 These calculations give the following results: R_0 is estimated (by its posterior median)
500 at 0.83 (95%CrI: 0.78-0.93), and the type reproduction numbers are estimated at 0.57
501 (95%CrI: 0.53-0.62) for transmission from female to female, 0.45 (95%CrI: 0.42-0.50) for
502 transmission from female to male, 0.30 (95%CrI: 0.25-0.36) for transmission from male
503 to female, and 0.31 (95%CrI: 0.28-0.40) for transmission from male to male. Hence, the
504 above estimates indicate that females contribute more to overall transmission than
505 males because of the higher estimated rates of reactivation in females than in males,
506 while CMV would be unable to persist in the population in the absence of perinatal
507 transmission (as $R_0 < 1$).

508 Fig 12 further shows that males older than 50 years do not infect many persons
509 because they have a low reactivation rates and low contact intensities. Infected females,
510 especially young females, are expected to infect significantly more persons, the reason
511 being that their reactivation rates and contact intensities are higher. The observation
512 that both males and females generally infect persons that are older than themselves can
513 be explained by the fact that reactivation generally occurs years after primary infection,
514 and that such reactivation is expected to produce infection in persons who have similar

515 age at the moment of reactivation, due to highly age-assortative mixing patterns
516 (Fig 11).

517 The above analyses can be generalized and made more precise by using full vital
518 statistics of the population (i.e. by using historical age-specific birth rates and
519 anticipated future trends, and historical and anticipated age- and sex-specific mortality
520 rates), but this is beyond the scope of the current study. The above partial analyses
521 provided here do provide evidence that CMV needs both perinatal transmission and
522 transmission after reactivation to be able to persist in the population.

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526 **Competing interests**

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672 **Supplementary Figures**

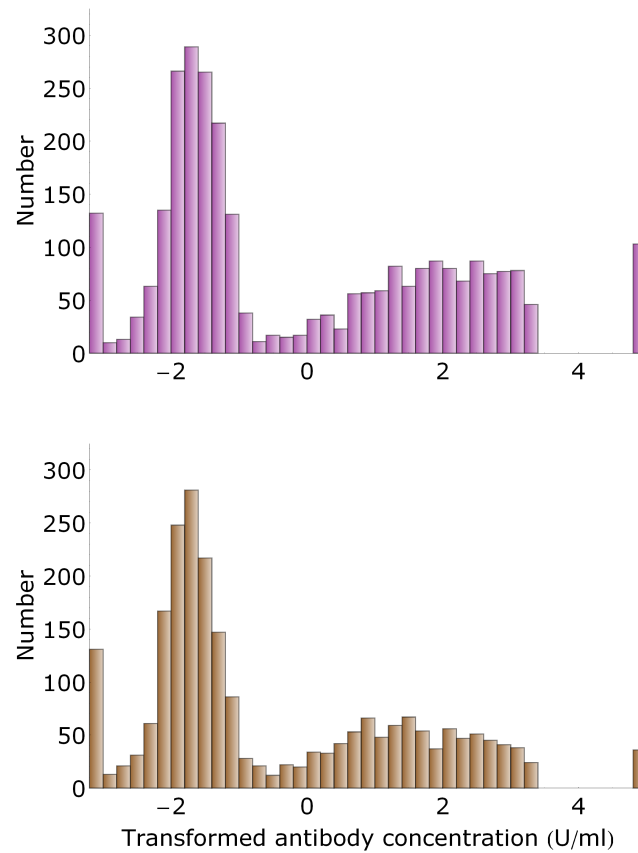


Fig 7. Numbers of samples as a function of antibody concentration.

Histograms shows the number of samples by antibody concentration class (top, purple: females; bottom, brown: males). Samples in the bars at 5 U/ml are right-censored at concentration 3.41 U/ml. Total number of female and males samples is 2,842 and 2,337, respectively.

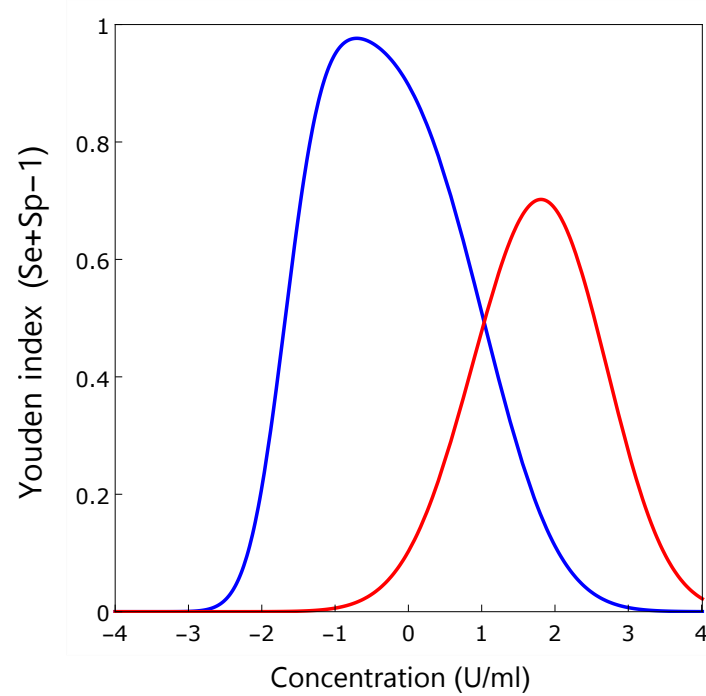


Fig 8. Youden index as a function of antibody concentration. Shown are the Youden indices ($Se+Sp-1$) for binary classifications, i.e. when using the antibody concentration on the x-axis as cut-off. Classifications are uninfected (S) versus infected (L+B; blue line) and uninfected plus infected with intermediate antibody concentration (S+L) versus infected with increased antibody concentration (B; red line).

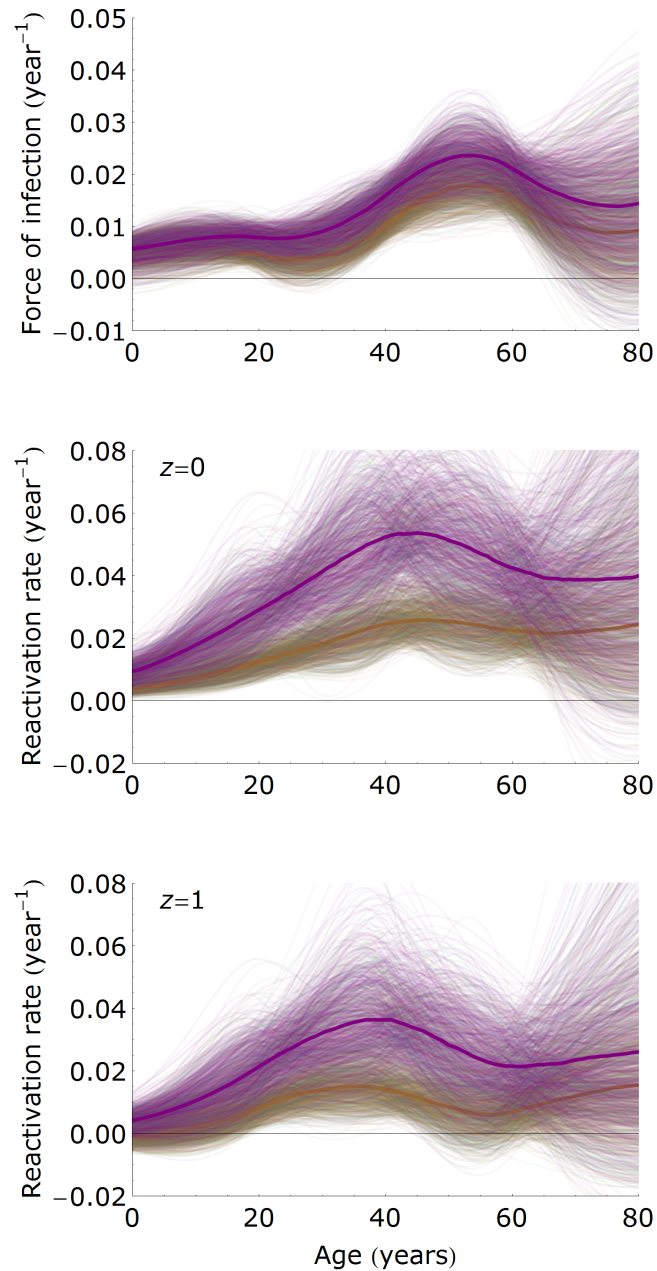


Fig 9. Estimates of transmission parameters with the mixture model.

Shown are estimates of the forces of infection (top panel) and reactivation rates (middle and bottom panel) in females (purple) and males (brown) based on spline estimates of the age specific prevalence in the mixture model and Equation (3). The middle panel shows results in the absence of re-infection ($z = 0$), and the bottom panel if re-infection occurs at the same rate as primary infection ($z = 1$). Shown are 1,000 samples from the posterior distribution (thin lines) with the posterior medians (bold lines). Notice that the force of infection is higher in females than in males, that the reactivation rate is substantially higher in females than in males, and that both the reactivation rates and forces of infection are highest in 30 – 50 year old persons.

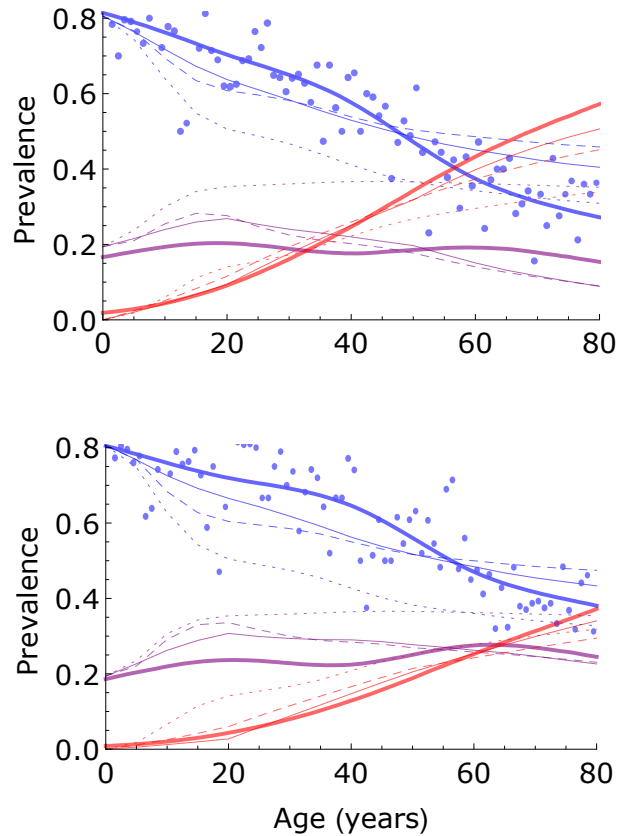


Fig 10. Comparison of fits of the statistical and transmission models. Shown are prevalence estimates for females (top) and males (bottom), and for classes of low (blue), medium (purple), and high (red) antibody measurements. The data and fit (posterior medians) of the statistical model are indicated by dots and thick lines, respectively (cf. Fig 3). Thin solid lines show the fit (using the ML parameter estimates) of the transmission model with infectious reactivation and re-infection. Dotted and dashed lines show the fits of the transmission model with no reactivation and with reactivation and re-infection not being infectious, respectively. The fit of the transmission model with infectious reactivation but no re-infection is very close to the fit of the full model with infectious reactivation and infectious re-infection, and is not shown.

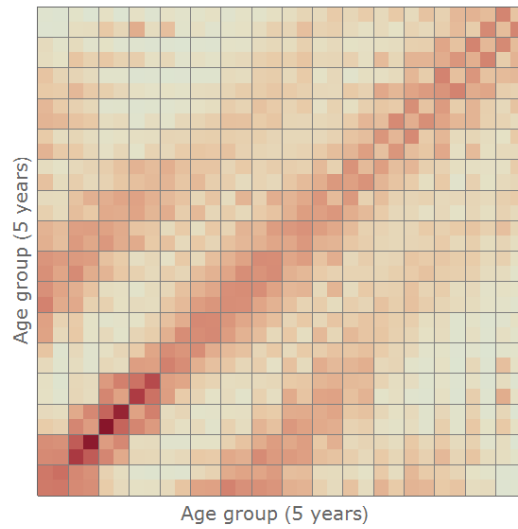
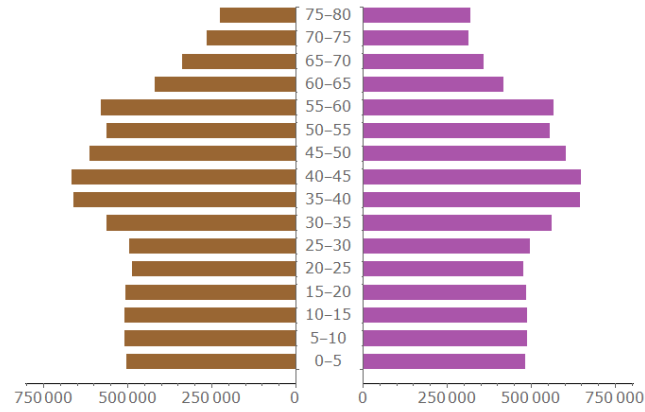


Fig 11. Demography and contact patterns. Shown are the demographic composition of the Dutch population in 2007 and estimated contact rates using data from 2006-2007. The main stratification of contact rates in 5-year age groups, and the secondary stratification by sex (2 by 2 blocks). See [24] for details.

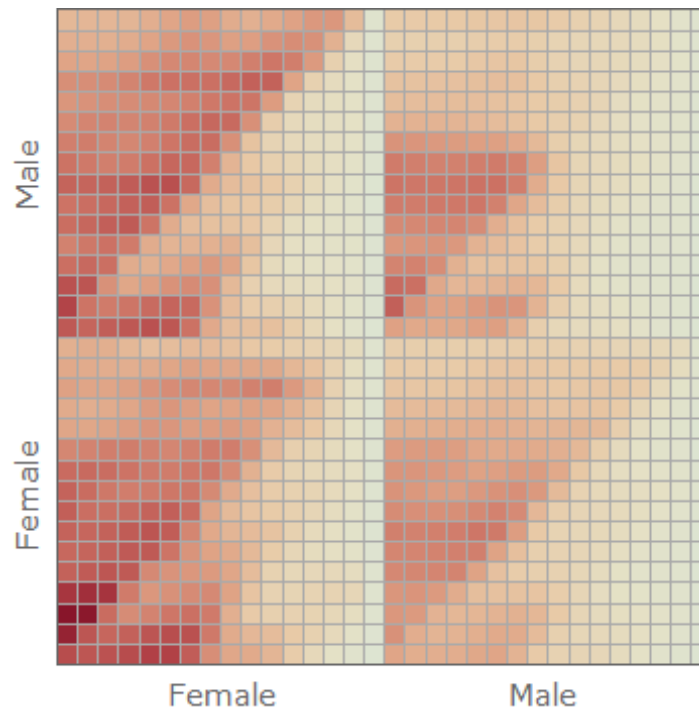


Fig 12. Graphical representation of the next-generation matrix. Shown is the next-generation matrix, stratified by sex and age (16 age groups of 5 years). The top left-hand block shows $k^{\sigma\varphi}$, representing the female-to-male transmission ($R^{\sigma\varphi} = 0.45$). Top right-hand block represents male-to-male transmission ($R^{\sigma\sigma} = 0.31$). The bottom blocks depict the female-to-female and male-to-female transmission matrices ($R^{\varphi\varphi} = 0.57$ and $R^{\varphi\sigma} = 0.30$). The basic reproduction number is $R_0 = 0.83$. Orange and blue represent high and low transmission intensity, respectively.