Use of phenylacetonitrile plus acetic acid to monitor *Pandemis pyrusana* (Lepidoptera: Tortricidae) in apple

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ABSTRACT A recent discovery has demonstrated that herbivore induced plant volatile compounds from apple trees infested with larvae were highly attractive to con-specific adult male and female leafrollers. However, this work (conducted in New Zealand and Canada) tested only low doses of kairomone. Our US study here was conducted to assess the attractiveness of higher doses of the six apple volatiles putatively identified from apple trees infested by tortricid larvae to the leafroller, *Pandemis pyrusana* Kearfott. These volatiles included: β-caryophyllene, germacrene D, benzyl alcohol, phenylacetonitrile, (*E*)-nerolidol, and indole. No volatiles were attractive to *P. pyrusana* when used alone. However, traps baited with phenylacetonitrile plus acetic acid caught both sexes of *P. pyrusana*. Traps baited with the other volatiles plus acetic acid caught zero to only incidental numbers of moths (< 1.0). Interestingly, traps baited with phenylacetonitrile plus acetic acid caught significantly more *P. pyrusana* than traps baited with a commercial sex pheromone lure. The evaporation rate of the acetic acid co-lure was an important factor affecting catches of *P. pyrusana* with phenylacetonitrile, and studies are needed to optimize the emission rates of both lure components. Further studies are warranted to develop phenylacetonitrile and possibly other aromatic plant volatiles as bisexual lures for the range of tortricid pests attacking horticultural crops.

KEY WORDS: codling moth, leafroller, host-plant volatile, phenylacetonitrile

INTRODUCTION

Pandemis spp. leafrollers are an important group of tortricid pests attacking pome fruits (apple, *Malus domestica* Borkhausen, and pear, *Pyrus communis* L.), in North America (Chapman 1973, Dombroskie and Sperling 2012) and in Europe and Asia (Dickler 1991). These species are often bivoltine and overwintering larvae in the fall-spring and summer generations feed on the skin of developed fruits adjacent to leaves creating cull fruits (Brunner 1996). Insecticides are most often used to manage these pests; although sex pheromone mating disruption has been developed and implemented on a relatively small acreage in North America (Judd and Gardiner 2008). Sex pheromone-baited traps are used to monitor adult populations to assess both density and phenology, but pest managers feel they are insensitive to local population densities within the orchards (Brunner 1984). Alternative lures, including acetic acid, have been evaluated to monitor both sexes of *Pandemis pyrusana* (Kearfott) (Knight, 2001); however, acetic acid alone was considered to be a weak lure and was not adopted by Washington State growers to monitor *P. pyrusana* (Alway 2003).

Apple seedlings uniquely release several compounds, including acetic acid, acetic anhydride, benzyl alcohol, phenylacetonitrile, indole, 2-phenylethanol, and (E)-nerolidol only when infested by larvae of light brown apple moth (LBAM), *Epiphyas postvittana* (Walker), the obliquebanded leafroller (OBLR), *C. rosaceana* (Harris), and the eye-spotted bud moth (ESBM), *Spilonota ocellana* (Denis & Schiffermüller) (Suckling et al. 2012; El-Sayed et al. 2016). Recently El-Sayed et al. (2016) found that a binary blend of the two herbivore-induced plant volatile compounds (HIPVCs), phenylacetonitrile + acetic acid and 2-phenylethanol + acetic acid, attracted

a significant number of conspecific male and female adult LBAM in New Zealand. Further investigation with other leafrollers (Tortricidae) in Canada, including the ESBM and OBLR, revealed similar systems. The study reported by El-Sayed et al. (2016) targeted mainly ESBM and OBLR using a lower doses of the new kairomone. In this study we have investigated the response of *P. pyrusana* to the compounds reported by El-Sayed et al. (2016) using higher doses of the HIPVCs in Washington State during 2012–2014. Also, studies were conducted to evaluate possible interactions with any of the attractive volatiles when combined with pear ester and acetic acid in binary and ternary blends for moth catches of both *C. pomonella* and *P. pyrusana* in the same trap.

MATERIALS AND METHODS

Chemicals and Lures. Chemical purity of the volatile apple chemicals used in our trials were as follows: benzyl alcohol (99%), (*E*)-nerolidol (85%), phenylacetonitrile (99%), indole (99%), β-caryophyllene (97%), and germacrene D (96%) (Sigma Aldrich, St. Louis, MO). These chemicals were loaded (100 mg) into plastic sachet. Sachet lures consisted of a heat-sealed, semi-permeable polyethylene bag (45 mm x 50 mm, 150 μm wall thickness), with a piece of cellulose acetate filter (15 mm × 45 mm, Moss Packaging Co. Ltd., Wellington, New Zealand) inserted as the carrier substrate. Two acetic acid (glacial acetic acid (99.7%), Sigma Aldrich) co-lures were made by drilling either 1.0 or 3.2 mm holes in the cap of 8 ml Nalgene vials (Nalg-Nunc International, Rochester, NY) and loading each vial with two small cotton balls and 5 ml of glacial acetic acid. A third acetic acid lure, a proprietary round (3.4 cm diameter) plastic membrane cup (Pherocon AA) was provided by Trécé Inc. (Adair, OK). The mean weekly weight loss (evaporation rate) of acetic acid from the three lures was recorded from 30 Aug. to 26 Sept. 2013. Individual lures (N = 10) were placed in red Pherocon VI delta traps (Trécé Inc.) without sticky liners that were hung in the canopy of five linden trees, Tilia cordata Miller, at the Yakima Agricultural Research Laboratory, Wapato, WA. Similarly, the weight loss of sachet lures loaded with phenylacetonitrile (N = 5) was recorded from 28 Aug. to 19

Sept. 2013. In addition, the proprietary lures: Pherocon CM-DA lure loaded with pear ester, the Pherocon CM-DA combo lure loaded with codling moth sex pheromone and pear ester, and the Pherocon PLR lure (#3147) loaded with the *P. pyrusana* sex pheromone were obtained from Trécé Inc.

Field experiment protocols. Studies evaluated single, binary, and ternary blends of selected host-plant volatiles and acetic acid co-lures using a standardized protocol in 2012 and 2013. Studies were conducted in two apple orchards, the USDA research farm situated east of Moxee, WA (46o30'N, 120o10'W), and in a commercial orchard located southwest of Naches, WA (46o43'N, 120o42'W). Orange delta traps with sticky inserts (Pherocon VI, Trécé Inc.) were used in all studies. Five replicates of each lure treatment were randomized and spaced 20–30 m apart. Traps were placed at a 2 m height in the canopy. All lures were placed on the liner. Acetic acid vials with 3.2 mm holes were used in all studies unless specified otherwise. Traps baited with a blank sachet lure were used as a negative control in each study. Studies were repeated a variable number of times and typically each test ran for 5–7 d. When studies were repeated on different dates all traps were collected, new lures were transferred to new liners, and traps were re-randomized in the field. Moths were sexed in all studies.

Comparison of induced volatiles. Plant volatiles, reportedly induced by tortricid larval feeding on apple and attractive to adults (El-Sayed et al., 2016), were evaluated in two consecutive studies in the Moxee orchard during 2013. The first study compared moth catches from 12–19 Aug. in traps baited with phenylacetonitrile, benzyl alcohol, indole, (E)-nerolidol, β-caryophyllene, or germacrene D; and the combination of phenylacetonitrile with an acetic acid co-lure. The second study compared these same six volatiles all in combination with acetic acid co-lures against acetic acid alone from 19–28 Aug.

Combination lures with phenylacetonitrile. Studies conducted in both 2012 and 2013 evaluated phenylacetonitrile alone, and in combination with either pear ester or acetic

acid, pear ester with acetic acid, and phenylacetonitrile plus pear ester and acetic acid in order to determine whether interactions would affect catches of either or both target species. Traps in 2012 were initially placed in the Moxee orchard on 1 Aug. and traps were checked and re-randomized on 8 Aug. This study was terminated on 13 Aug. and traps were moved to the Naches orchard. Traps were checked and re-randomized in this orchard on 22 and 30 Aug., and the study was terminated on 12 Sept. This same study was repeated in 2013 from 20–27 June at both the Moxee and Naches orchards and from 5–12 and 17–27 Aug. in the Moxee orchard. A third study was conducted from 7–21 July 2014 in the Moxee orchard to evaluate the use of phenylacetonitrile with the sex pheromone / pear ester combinational lure and the acetic acid cup lure. A final study was conducted in the Naches orchard from 30 Aug. to 10 Sept. 2014, to compare the magnitude of the total bisexual moth catches of *P. pyrusana* in traps with phenylacetonitrile plus acetic acid with male catches in traps baited with *P. pyrusana* sex pheromone.

The acetic acid evaporation rate was evaluated over three time periods in the Moxee orchard during 2013. Traps baited with phenylacetonitrile sachet lures plus acetic acid vials with either 1.0 or 3.2 mm holes or the Pherocon AA cup lure were compared. New lures were used in each of three separate studies which were conducted from 28 Aug. to 4 Sept., 4–11 Sept., and 12–19 Sept. 2013.

Statistical analyses. Count data were subjected to a square root transformation prior to analysis of variance (ANOVA) to normalize the variances. A randomized, complete block design was used in most studies with date as the blocking variable. Following a significant F-test in the ANOVA, means were separated with Tukey's HSD test with an alpha value of 0.05. Treatments with zero catches or only incidental catch were not included in these analyses.

RESULTS
Comparison of induced volatiles. No *P. pyrusana* were caught with any of the six

plant volatiles during Test 1 in the Moxee orchard (Table 1). In comparison, small numbers of moths were caught when acetic acid was added as a co-lure to traps baited with phenylacetonitrile (Table 1). During the second test comparing these six volatiles in combination with acetic acid, no moths were caught with β-caryophyllene, germacrene D, or acetic acid alone. Incidental moth catches occurred in blank traps and in traps baited with (E)-nerolidol lures. Means > 1 moth per trap were caught with acetic acid plus either benzyl alcohol, indole or phenylacetonitrile. The bisexual total and female moth only catches were significantly greater with acetic acid plus phenylacetonitrile than the other two volatiles plus acetic acid co-lure combinations (Table 1).

Table 1. Comparison of catches of Pandemis leafroller in traps baited with different host- plant volatiles alone (Test 1) or in combination with acetic acid (Test 2), N = 5, Moxee, WA, 2013.

Test	Lures [a]	Mean (SE) catch	
		Total	Females
1	Blank	0.0 (0.0)	0.0 (0.0)
	β-caryophyllene	0.0 (0.0)	0.0 (0.0)
	Germacrene D	0.0 (0.0)	0.0 (0.0)
	(E)-nerolidol	0.0 (0.0)	0.0 (0.0)
	Benzyl alcohol	0.0 (0.0)	0.0 (0.0)
	Indole	0.0 (0.0)	0.0 (0.0)
	Phenylacetonitrile (PA)	0.0 (0.0)	0.0 (0.0)
	PA + acetic acid (AA)	1.6 (0.9)	0.6 (0.4)
2	Blank	0.2 (0.2)	0.2 (0.2)
	AA	0.0 (0.0)	0.0 (0.0)
	β-caryophyllene + AA	0.0 (0.0)	0.0 (0.0)
	Germacrene D + AA	0.0 (0.0)	0.0 (0.0)
	(E)-nerolidol + AA	0.4 (0.4)	0.2 (0.2)
	Benzyl alcohol + AA	1.0 (0.5)b	0.8 (0.6)ab
	Indole + AA	1.0 (0.8)b	0.2 (0.2)b
	PA + AA	8.8 (1.6)a	3.2 (1.1)a
	ANOVA	$F_{1, 12} = 14.29$	$F_{1, 12} = 4.30$
		$P < 0.001$	$P < 0.05$

Column means for test 2 followed by a different letter were significantly different in a one-way ANOVA, Tukey's HSD test, P < 0.05.
[a]Each plant volatile was loaded on a felt pad in plastic sachets in aliquots of 100 μl. Acetic acid (5 ml) was loaded on cotton balls in plastic vials with a 3.1 mm hole.

Combination lures with phenylacetonitrile. Consistent data were collected in both 2012 and 2013 when comparing catches of C. pomonella with phenylacetonitrile alone and with binary and ternary blends including pear ester and/or acetic acid (Tables 2 and 3). Pear ester in combination with acetic acid caught significantly more total and female *C. pomonella* than other lures. Adding phenylacetonitrile to pear ester and acetic acid significantly reduced both total and female moth catches of *C. pomonella*. Catches of *C. pomonella* with either phenylacetonitrile alone and in binary blends with either acetic acid or pear ester were low and did not differ significantly. In addition, traps with phenylacetonitrile alone in the 2013 test caught numbers of *C. pomonella* similar to numbers captured by either acetic acid or pear ester alone. Adding phenylacetonitrile to traps baited with the sex pheromone-pear ester combination lure (Pherocon CM-DA) plus acetic acid did not significantly reduce male, female, or total moth catches in 2014 (Fig. 1).
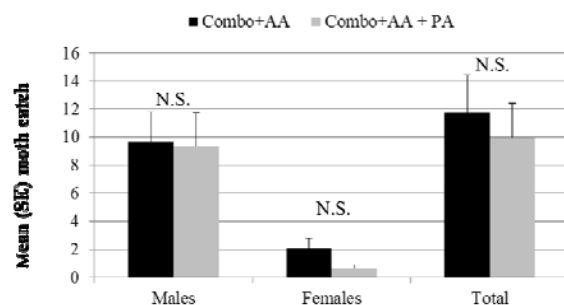

Fig. 1. Mean (SE) number of male, female, and total Cydia pomonella caught in traps with a sex pheromone + pear ester combo lure and an acetic acid co-lure versus these same lures with the addition of a phenylacetonitrile sachet lure, 7–21 July 2014, Moxee WA. 'N.S.' denotes a nonsignificant difference in moth catches between the two types of lures, P > 0.05.

Counts of *P. pyrusana* were more variable in traps than captures of codling moth between both years of the study (Tables 2 and 3). During 2013, traps baited with phenylacetonitrile plus acetic acid or in combination with pear ester caught significantly more total and female moths than all other treatments and did not differ among themselves. However, moth catches in 2012 were lower than in 2013 and no

significant differences among treatments were found for female moth catch. Also, total moth catch in traps baited with pear ester plus acetic acid in 2012 was not significantly different from the four treatments that included phenylacetonitrile.

The average evaporation rate of acetic acid from the co-lures used with phenylacetonitrile was a significant factor affecting moth catches of *P. pyrusana* (Table 4). Evaporation rates ranged 18-fold across the three dispensers tested. Moth catches were significantly lower in traps containing acetic acid lures with the lowest evaporation rate (plastic cup lure), and did not differ between traps baited with plastic vials with the two different hole sizes. Sachet lures loaded with phenylacetonitrile evaporated an average (SE) of 6.3 (0.2) mg d^{-1}.

Table 2. Comparison of catches of codling moth and Pandemis leafroller in orange delta traps unbaited or baited with combinations of phenylacetonitrile, pear ester, and acetic acid, N =10 for codling moth and N= 15 for Pandemis leafroller, Naches, WA 2012.

Lures[a]	Mean (SE) moth catch per trap[b]			
	Codling moth		*Pandemis* leafroller	
	Total	Females	Total	Females
Blank	0.2 (0.1)	0.1 (0.1)	0.0 (0.0)	0.0 (0.0)
Phenylacetonitrile (PA)	0.2 (0.2)c	0.0 (0.0)c	0.9 (0.5)b	0.7 (0.5)
Pear ester (PE) + PA	0.7 (0.3)c	0.6 (0.2)c	0.5 (0.2)b	0.3 (0.2)
PE + acetic acid (AA)	15.7 (3.1)a	8.6 (1.4)a	2.2 (0.8)ab	1.2 (0.5)
PA + AA	0.4 (0.2)c	0.2 (0.1)c	4.9 (1.4)a	1.7 (0.6)
PA + PE + AA	5.1 (0.9)b	2.6 (0.6)b	2.6 (1.5)ab	0.7 (0.5)
ANOVA[c]	$F_{3, 35} = 42.55$	$F_{3, 35} = 34.33$	$F_{4, 68} = 4.92$	$F_{4, 68} = 2.03$
	$P < 0.0001$	$P < 0.0001$	$P < 0.01$	$P = 0.10$

Means followed by a different letter were significantly different in a randomized complete block ANOVA, Tukey's HSD test, P < 0.05.
[a]Phenylacetonitrile was loaded on a felt pad in plastic sachets at 100 μl. Pherocon CM DA lures contained 3.0 mg pear ester. Acetic acid (5 ml) was loaded on cotton balls in plastic vials with a 3.2 mm hole.
[b]Experiments were repeated on two dates in the Moxee orchard: 1–8 and 8–13 Aug.
[c]Data from blank traps for both species and with PA lures for C. pomonella were not included in the analysis.

Moth catches of *P. pyrusana* in traps placed in the Naches orchard were much higher during 2014 than in the two previous years (Fig. 2). Total moth catch was significantly different and nearly 6-fold higher in traps baited with phenylacetonitrile than with the sex pheromone of *P. pyrusana* ($F_{1,8} = 54.64$, $P < 0.001$). Mean catch of both male ($F_{1,8} = 9.30$, $P < 0.05$) and female ($F_{1,8} = 20.01$, $P < 0.01$) moths alone were both significantly higher with this binary lure than the catch of males in the sex pheromone-baited traps
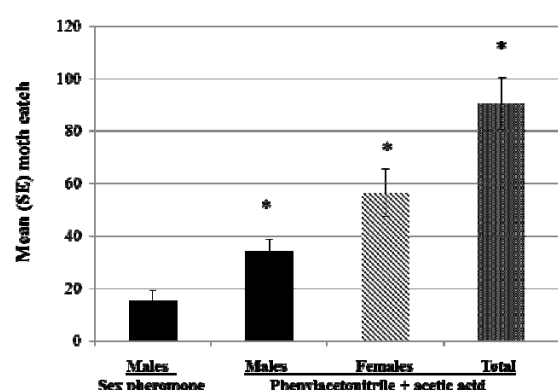

Fig. 2. Mean (SE) number of male, female, and total *Pandemis pyrusana* caught in traps baited with either the sex pheromone lure for *P. pyrusana* (males only) or with phenylacetonitrile plus an acetic acid co-lure (males, females, and total moths). '*' denotes a significant mean difference with male catch in the sex pheromone-baited trap, P < 0.05.

Table 3. Comparison of catches of codling moth and Pandemis leafroller in orange delta traps unbaited or baited with combinations of phenylacetonitrile, pear ester, and acetic acid, N =20, Moxee, WA 2013.

Lures [a]	Mean (SE) moth catch per trap [b]			
	Codling moth		*Pandemis* leafroller	
	Total	Females	Total	Females
Blank	0.1 (0.1)	0.0 (0.0)	0.0 (0.0)	0.0 (0.0)
Acetic acid (AA)	0.6 (0.2)bc	0.5 (0.2)bc	0.6 (0.2)b	0.4 (0.2)b
Pear ester (PE)	0.7 (0.2)bc	0.3 (0.2)bc	0.2 (0.1)b	0.1 (0.1)b
Phenylacetonitrile (PA)	0.4 (0.2)bc	0.1 (0.1)bc	0.2 (0.1)b	0.2 (0.1)b
PE PA	0.6 (0.2)bc	0.1 (0.1)c	0.1 (0.1)b	0.1 (0.1)b
PE – AA	5.2 (1.5)a	2.8 (0.9)a	0.8 (0.2)b	0.4 (0.2)b
PA + AA	0.2 (0.1)c	0.2 (0.1)bc	7.1 (1.6)a	3.8 (1.1)a
PA + PE + AA	1.9 (0.6)b	1.0 (0.3)b	7.8 (1.5)a	4.7 (1.0)a
ANOVA [c]	$F_{6,130} = 11.72$	$F_{6,130} = 9.76$	$F_{6,130} = 39.84$	$F_{6,130} = 27.12$
	$P < 0.0001$	$P < 0.0001$	$P < 0.0001$	$P < 0.0001$

Means followed by a different letter were significantly different in a randomized complete block ANOVA, Tukey's HSD test, P < 0.05.
[a]Phenylacetonitrile was loaded on a felt pad in plastic sachets at 100 µl. Pherocon CM DA lures contained 3.0 mg pear ester. Acetic acid (5 ml) was loaded on cotton balls in plastic vials with a 3.2 mm hole.
[b]Experiments were repeated on four orchard-dates: 20–27 June in both the Moxee and Naches orchards, and from 5–12 and 17–27 Aug. in the Moxee orchard.
[c]Data from blank traps were not included in the analyses.

DISCUSSION

In the studies reported here, phenylacetonitrile and other plant volatiles known to be induced by insect feeding, including benzyl alcohol, indole, and acetic acid provided no intrinsic attraction of *P. pyrusana* when tested alone. However, when any of the three compounds was combined with acetic acid all, and especially phenylacetonitrile, exhibited strong attractiveness to adult male and female *P. pyrusana*. Phenylacetonitrile and acetic acid have been reported as herbivore induced plant volatiles in apple as a result of LBAM larval feeding, while phenylacetonitrile was also identified as herbivore induced plant volatile in apple as results of OBLT and ESBM larval feeding (El-Sayed et al., 2016). It is probable that the effects are caused by fatty acid-amino acid conjugates in caterpillar saliva (Yoshinaga et al. 2010), although this has not been demonstrated directly for Tortricidae as yet. In our previous work, conducted in apple orchards in Canada, combinations of phenylacetonitrile and acetic acid or 2-phenylethanol and acetic acid were attractive to adult male and female OBLR and ESBM. The results obtained in this study combined with earlier results with other tortricids suggest the positive response of adult tortricids to HIPV compounds is a widespread phenomena among these species. Phenylacetonitrile has previously been studied with several tortricids and host-plant material in

laboratory studies. For example, phenylacetonitrile was one of 22 compounds identified from the headspace (< 0.5%) of excised peach shoots, *Prunus persica* L. (Natale et al. 2003), but was not found to be attractive to mated female *Grapholita molesta* (Busck), when used in a blend with three green leaf volatiles plus benzaldehyde (Piñero and Dorn 2007). Apple leaves also released phenylacetonitrile within a blend of volatiles in response to larval feeding by the tortricids *E. postvittana* (Suckling et al. 2012) and *P. heparana* (Giacomuzzi et al. 2013). Yet, neither study measured whether it was attractive to these herbivorous insects.

Phenylacetonitrile is a relatively well known semiochemical involved in a range of insect-insect and insect-plant interactions. These include serving as a male-produced anti-aphrodisiac in the butterfly *Pieris brassicae* L.; facilitating a phoretic dispersal and arrestment by female egg parasitoid Trichogramma brassicae Bezdenko (Fatouros et al. 2005, Huigens et al. 2009); and acting as a male-specific repellant in the locust, *Schistocerca gregaria* (Forskal) in order to manage sperm competition within insect aggregations (Seidelmann and Ferenz 2002). Phenylacetonitrile is also released by male and female flowers of boxelder trees, *Acer negundo* L., and it alone attracts nymphs, and both sexes of the adult box elder bug, *Boisea rubrolineata* (Barber) (Schwarz et al. 2009). Phenylacetonitrile was isolated from oilseed rape, *Brassica napus* L., and shown to attract cabbage seed weevil, *Ceutorhynchus assimilis* Payk (Smart and Blight 1997). Similarly, phenylacetonitrile is induced by feeding of the Japanese beetle, *Popillia japonica* Newman, and the fall webworm, *Hyphantria cunea* Drury, on several plant species, including crabapple, *Malus sp.*, and within a blend of induced

volatiles, serves as an adult aggregation kairomone (Loughrin et al. 1995). Phenylacetonitrile was shown to be released from cabbage, *Brassica spp.*, and nasturtium, *Tropaeolum majus* L., following larval feeding of *P. brassicae* and *P. rapae* L., but not from undamaged plants (Geervliet et al. 1997). In apple, a significant difference in the release of phenylacetonitrile was found among four cultivars, including two transgenic scab resistant cultivars (Vogler et al. 2010).

Table 4. Comparison of catches of Pandemis leafroller in orange delta traps baited with phenylacetonitrile plus one of three different acetic acid co-lures, N = 15, Moxee WA, 2013.

Type of acetic acid lure[a]	Mean (SE) daily weight loss (mg) from lure	Mean (SE) weekly moth catch[b]	
		Total	Females
Plastic cup	4.9 (0.1)c	10.2 (2.2)b	3.3 (0.8)b
1.0-mm vial	16.1 (2.0)b	26.8 (7.2)a	9.1 (2.4)a
3.2-mm vial	88.6 (3.3)a	17.1 (3.5)a	6.8 (1.4)a
ANOVA	$F_{2,27} = 500.8$	$F_{2,40} = 12.56$	$F_{2,40} = 10.42$
	$P < 0.0001$	$P < 0.001$	$P < 0.001$

Means followed by a different letter were significantly different in a randomized complete block ANOVA, Tukey's HSD test, P < 0.05.
[a]Acetic acid lures included the proprietary Pherocon AA (Trécé Inc., Adair, OK) plastic cup and polypropylene vials filled with 5 ml of acetic acid and with either 1.0 or 3.2 mm holes. Phenylacetonitrile sachet lures released an average (SE) of 6.3 (0.2) mg d-1.
[b]Experiments were repeated on three dates: 28 Aug.–4 Sept., 4–11 Sept. and 12–19 Sept. 2013 in the Moxee orchard.

Previous studies with plant-feeding Lepidoptera (primarily noctuids and pyralids) have correlated the presence of larval feeding on a host plant with the repulsion of the con-specific adults (Landolt 1993, De Moraes et al. 2001, Signoretti et al. 2012, Reisenman et al. 2013, Zakir et al. 2013, Hatano et al. 2015). No previous studies on tortricid moths have reported whether adults cue to host-plant volatiles induced by larval feeding. When presented alone the induced plant volatiles examined in this study exhibited no attraction to adult *P.*

pyrusana based on the absence of moths in traps. However, we did not study whether these induced volatiles were behaviorally repellent or interfered with volatile-mediated host selection and/or sex pheromone communication (Hatano et al. 2015).

Our findings suggest that acetic acid is an important synergist of herbivore-induced plant volatiles for adult *P. pyrusana*. Interestingly, acetic acid has rarely been reported in the analysis of volatile collections from damaged apple foliage; trace levels were found in samples of cut flowering apple branches (Bengtsson et al. 2001), from apple leaves infested with phytophagous mites (Llusia and Peñuelas 2001), and from drought-stressed trees (Ebel et al. 1995). Interestingly, an earlier paper found phenylacetonitrile was released by spider mite damage on apples (Takabayashi et al. 1994). Acetic acid is a primary metabolite of plant metabolism under stress conditions and is regulated by the production and catabolism of ethanol and acetaldehyde (Oikawa and Lerdau 2013). However, acetic acid could also be produced by the growth of yeast and bacteria prevalent in the phyllosphere of apple leaves, or on surfaces of senescent flowers which are relatively rich in sugars and other carbohydrates (Atlas and Bartha 1981, Kinkel 1997). A potential role for microorganisms in the herbivore induction of plant volatiles has been suggested previously (Dicke and Hilker 2003, Hilker and Meiners 2006); yet, few studies have tried to measure this interaction. One exception was the purported role of regurgitated endosymbionts in the induction of plant defenses (Spiteller et al. 2000).

The interplay of microbial and plant volatiles with an adult tortricid pest has perhaps best been studied with *Lobesia botrana* (Denis and Schiffermüller) on grapes, *Vitis vinifera* L., (Tasin et al. 2005, 2011). In this system, neither acetic acid nor the induced volatile 2-phenylethanol was collected from undamaged flowers and fruit (Tasin et al. 2005). However, both compounds were abundant in volatiles from fruit infested with yeasts or acetic bacteria, and both female oviposition and net reproductive rate were significantly increased as a result of microbial infestations of fruit (Tasin et al. 2011). In contrast, fruit infected by Botrytis cinerea (Persoon), did not release acetic acid and was repulsive to females and reduced the overall fitness of the insect. Inoculation of fruit with acetic bacteria produced > 100-fold higher levels of acetic acid than with the yeasts, and oviposition was reduced compared with clean fruit, suggesting an overdose of acetic acid can occur. In our studies, the higher emission rates of acetic acid tested were more attractive, but neither the emission rates of phenylacetonitrile nor acetic acid have been optimized for attraction of *P. pyrusana*.

Our studies can likely impact the pest management of *P. pyrusana*. Surprisingly, traps baited with phenylacetonitrile and acetic acid caught more moths than sex pheromone traps in an orchard not treated with leafroller sex pheromones. These data may suggest that the commercial lures for *P. pyrusana* are not optimized or that other species in the *Pandemis limitata* (Robinson) group with different sex pheromone blends are more dominant in the two orchards studied (Dombroskie and Sperling 2012). The respective sex pheromones of P. limitata and *P. pyrusana* have been identified as 91:9 and 94:6 blends of (Z)-11-tetradecenyl acetate and (Z)-9-tetradecenyl acetate (Roelofs et al. 1976, 1977). At least one commercial lure has been reported to be relatively ineffective for *P. pyrusana* due to contamination

with < 1.0% of (Z)-9-dodecenyl acetate (Brunner and Fisher 1998).

We found that adding pear ester to traps baited with phenylacetonitrile and acetic acid did not reduce the catch of *P. pyrusana*. However, adding phenylacetonitrile to traps baited with pear ester and acetic acid significantly reduced catches of C. pomonella. Thus ternary blends in a single trap would be useful in monitoring *P. pyrusana*, but may compromise catches of *C. pomonella*. However, a significant drop in catch of C. pomonella did not occur when phenylacetonitrile and acetic acid were added to the combination sex pheromone plus pear ester lure. The potential benefit of improved monitoring of *P. pyrusana*, especially of female moths, with phenylacetonitrile would have to outweigh the information lost from a drop in catches of female *C. pomonella* in these combination traps (summarized in Knight et al. 2014). Hypothetically, this could occur in certified-organic orchards if the pest pressure from *C. pomonella* is low, efficacious tools to manage leafrollers are limited and, especially, when sex pheromone-based mating disruption for leafrollers is used and the standard sex pheromone-baited traps are ineffective. The ability to monitor multiple species could fuel the adoption of labor-saving remote electronic traps (Guarnieri et al. 2011, Kim et al. 2011).

Our positive results with enhancing the bisexual moth catches of *P. pyrusana* suggest that studies should continue to identify and test other plant volatiles induced by insect feeding on pome fruits (Boevé et al. 1996, Suckling et al. 2012, Giacomuzzi et al. 2013, El-Sayed et al. 2016). The use of more complex and ratio-specific blends of induced volatiles should be evaluated, perhaps both alone and in the context of the common volatiles released by uninfested and infested plants (Bruce and Pickett 2011). For example, green leaf volatiles have been found to enhance catches with sex pheromones in field trials with some tortricid pests (Light et al. 1993, Varela et al. 2011, Yu et al. 2015). The release rates of both (Z)-3-hexenyl acetate and (Z)-3-hexenyl benzoate were increased with herbivory (Suckling et al. 2012), and in a previous study combining (Z)-3-hexenyl acetate with acetic acid nearly doubled the catch of *P. pyrusana* (Knight et al. 2014). Polyphagous tortricid pests such as *P. pyrusana* attack a number of horticultural crops other than pome fruits. Future studies should more broadly characterize the induced plant volatiles in these crops to identify other possible attractants, including green leaf volatiles, aromatics, alcohols, and terpenes, and test these in combination with acetic acid as attractive lures. It is likely that systematic screening can now improve these lures for pest management of several target species.

REFERENCES

Alway, T. 2003. Exploring alternatives to organophosphate insecticides with apple and pear: the Areawide II Project in Washington State. Compact Fruit Tree 36: 91–93.

Atlas, R. M. and R. Bartha. 1981. Microbial ecology: fundamentals and applications. CAB, London, United Kingdom.

Bengtsson, M., A.-C. Ba ckman, I. Liblikas, M. I. Ramirez, A.-K. Borg-Karlson, L. Ansebo, P. Anderson, J. Lo fqvist, and P. Witzgall. 2001. Plant odor analysis of apple: antennal response of codling moth females to apple volatiles during phenological development. J. Agric. Food Chem. 49: 3736–3741.

Boevé, J. L., U. Lengwiler, L. Tollsten, S. Dorn, and C. J. Turlings. 1996. Volatiles emitted by apple fruitlets infested by larvae of the European apple sawfly. Phytochem. 42: 373–381.

Bruce, T. J. A. and J. A. Pickett. 2011. Perception of plant volatile blends by herbivorous insects — finding the right mix. Phytochem. 72: 1605–1611.

Brunner J. F. 1984. Pandemis leafroller, a pest of increasing concern to Washington fruit growers. Proc. WA State Hort. Assoc. 79: 119–125.

Brunner, J. F. 1996. Management of leafrollers in Washington apple orchards. Proc. WA State Hort. Assoc. 91: 257–263.

Brunner, J. F. and J. Fisher. 1998. Inhibition of leafroller captures in pheromone traps. Proceedings of the 79th Western Orchard Pest and Disease Management Conference, 7–9 January 1998, Portland, OR.

Chapman, P. J. 1973. Bionomics of the apple-feeding Tortricidae. Annu. Rev. Entomol. 18: 73–96.

De-Moraes, C. M., M. C. Mescher, and J. H. Tumlinson. 2001. Caterpillar-induced nocturnal plant volatiles repel conspecific females. Nature 410: 577–580.

Dicke, M. and M. Hilker. 2003. Induced plant defenses: from molecular biology to evolutionary ecology. Basic Appl. Ecol. 4: 3–14.

Dickler, E. 1991. Tortricid pests of pome and stone fruits, Eurasian species, pp. 435-452. In L. P. S. van der Geest and H. H. Evenhuis (eds), Tortricoid pests. Elsevier, Amsterdam.

Dombroskie, J. J. and F. A. H. Sperling. 2012. Phylogeny of Nearctic Pandemis (Lepidoptera: Tortricidae), with focus on species boundaries in the *P. limitata* group. Ann. Entomol. Soc. Am. 105: 768–780.

Ebel, R. C., J. P. Mattheis, and D. A. Buchanan. 1995. Drought stress of apple trees alters leaf emissions of volatile compounds. Physiol. Plant. 93: 709–712.

El-Sayed, A. M., A. L. Knight, J. A. Byers, G. J. R. Judd, and D. M. Suckling. 2016.

Caterpillar-induced plant volatiles attract conspecific adults in nature. Sci Reports 6: 37555.

Fatouros, N. E., M. E. Huigens, J. J. A. Van Loon, M. Dicke, and M. Hilker. 2005. Chemical communication — butterfly anti-aphrodisiac lures parasitic wasps. Nature 433: 704.

Geervliet, J. B. F., M. A. Posthumus, L. E. M. Vet, and M. Dicke. 1997. Comparative analysis of headspace volatiles from different caterpillar-infested or uninfested food plants of Pieris species. J. Chem. Ecol. 23: 2935–2954.

Giacomuzzi, V. J. Abraham, and S. Angeli. 2013. Feeding damage of *Pandemis heparana* induces the release of specific volatile compounds from apple plants. Proc Conf. Intern. Res. Food Security, Univ. Hohenheim. Germany, 17–19 September 2013.

Guarnieri, A., S. Maini, G. Molari, and V. Rondelli. 2011. Automatic trap for moth detection in integrated pest management. Bull. Insect. 64: 247–251.

Hatano, E., A. M. Saveer, F. Borrero-Echeverry, M. Strauch, A. Zakir, M. Bengtsson, R. Ingell, P. Anderson, P. G. Becher, P. Witzgall, ad T. Dekker. 2015. A herbivore-induced plant volatile interferes with host plant and mate location in moths through suppression of olfactory signaling pathways. BMC Biology 13: DOI 10.1186/s12915-015-0188-3.

Hilker, M. and T. Meiners. 2006. Early herbivore alert: insect eggs induce plant defense. J. Chem. Ecol. 32: 1379–1397.

Huigens, M. E., F. G. Pashalidou, M.-H. Qian, T. Bukovinszky, H. M. Smid, J. J. A. Van Loon, M. Dicke, and N. E. Fatouros. 2009. Hitch-hiking parasitic wasp learns to exploit butterfly antiaphrodisiac. Proc. Natl. Acad. Sci. USA 106: 820–825.

Judd, G.J.R. and Gardiner, M.G.T. 2008. Efficacy of Isomate-CM/LR for managing codling moth and leafrollers by mating disruption in organic apple orchards in

western Canada. J. Entomol. Soc. Brit. Columbia 105: 45–60.

Kim, Y., S. Jung, Y. Kim, and Y. Lee. 2011. Real-time monitoring of oriental fruit moth, *Grapholita molesta*, populations using a remote sensing pheromone trap in apple orchards. J. Asia-Pac. Entomol. 14: 259–262.

Kinkel, L. L. 1997. Microbial population dynamics on leaves. Annu. Rev. Phytopathol. 35: 327–347.

Knight, A. L. 2001. Monitoring the seasonal population density of Pandemis pyrusana (Lepidoptera: Tortricidae) within a diverse fruit crop production area in the Yakima Valley, WA. J. Entomol. Soc. British Columbia 98: 217–225.

Knight, A. L. and J. E. Turner. 1999. Mating disruption of *Pandemis spp.* (Lepidoptera: Tortricidae). Environ. Entomol. 28: 81–87.

Knight, A. L., R. Hilton, E. Basoalto, and L. L. Stelinski. 2014. Use of glacial acetic acid to enhance bisexual monitoring of tortricid pests with kairomone lures in pome fruits. Environ. Entomol. 43: 1628–1640.

Landolt P. J. 1993. Effects of host plant leaf damage on cabbage looper moth attraction and oviposition. Entomol. Exp. Appl. 67: 79–85.

Light, D. M., R. A. Flath, R. G. Buttery, F. G. Zalom, R. E. Rice, J. C. Dickens, and E. B. Jang. 1993. Host plant green-leaf volatiles synergize the synthetic sex pheromones of the corn earworm and codling moth (Lepidoptera) Chemoecol. 4: 145–152.

Llusia, J. And J. Peñuelas. 2001. Emission of volatile organic compounds by apple trees under spider mite attack and attraction of predatory mites. Exper. Appl. Acar. 25: 65–77.

Loughrin, J. H. D. A. Potter, and T. R. Hamilton-Kemp. 1995. Volatile compounds induced by herbivory act as aggregation kariomones for the Japanese beetle (Popillia japonica Newman). J. Chem. Ecol. 21: 1457–1467.

Natale D., L. Mattiacc, A. Hern, E. Pasqualini, and S. Dorn S, 2003. Response of female *Cydia molesta* (Lepidoptera: Tortricidae) to plant derived volatiles. Bull. Entomol. Res. 93, 335–342.

Oikawa, P. Y. and M. T. Lerdau. 2013. Catabolism of volatile organic compounds influences plant survival. Trends Plant Sci. 18: 695–703.

Piñero, J. C. and S. Dorn. 2007. Synergism between aromatic compounds and green leaf volatiles derived from the host plant underlies female attraction in the oriental fruit moth. Entomol. Exper. Appl.125, 185–194.

Reisenman, C. E., J. A. Riffell, K. Duffy, A. Pesque, D. Mikles, and B. Goodwin. 2013. Species-specific effects of herbivory on the oviposition behavior of the moth *Manduca sexta*. J. Chem. Ecol. 39: 76–89.

Roelofs, W. L., A. Carde, A. Hill, and R. T. Carde. 1976. Sex pheromone of the threelined leafroller, *Pandemis limitata*. Environ. Entomol. 5: 649–652.

Roelofs, W. L., R. F. Lagier, and S. C. Hoyt. 1977. Sex pheromone of the moth, *Pandemis pyrusana*. Environ. Entomol. 6: 353–354.

Schwarz, J., R. Gries, K. Hillier, N. Vickers, and G. Gries. 2009. Phenology of semiochemical-mediated host foraging by the western boxelder bug, *Boisea rubrolineata*, an aposematic seed predator. J. Chem. Ecol. 35: 58–70.

Seidelmann, K. and H.-J. Ferenz. 2002. Courtship inhibition pheromone in desert locusts, *Schistocera gregaria*. J. Insect Physiol. 48: 991–996.

Signoretti, A. G., M. F. Penaflor, and J. M. Bento. 2012. Fall armyworm, *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae), female moths respond to herbivore-induced corn volatiles. Neotrop. Entomol. 41: 22–26.

Smart, L. E. and M. M. Blight. 1997. Field discrimination of oilseed rape, Brassica

napus volatiles by cabbage seed weevil, *Ceutorhynchus assimilis*. J. Chem. Ecol. 23: 2555–2567.

Spiteller, D., K. Dettner, and W. Boland. 2000. Gut bacteria may be involved in interactions between plants, herbivores and their predators: microbial biosynthesis of N-acylglutamine surfactants as elicitors of plant volatiles. Biol. Chem. 381: 755–762.

Suckling, D. M., A. M. Twidle, A. R. Gibb, L. M. Manning, V. J. Mitchell, T. E. S. Sullivan, S. L. Wee, and A. M. El-Sayed. 2012. Volatiles from apple trees infested with light brown apple moth larvae attract the parasitoid *Dolichogenida tasmanica*. J. Agric. Food Chem. 60: 9562–9566.

Takabayashi, J., M. Dicke, and M. A. Posthumus. 1994. Volatile herbivore-induced terpenoids in plant-mite interactions: variation caused by biotic and abiotic factors. J. Chem. Ecol. 20: 1329–1354.

Tasin, M., G. Anfora, C. Ioriatti, S. Carlin, A. De Cristofaro, S. Schimidt, M. Bengtsson, G. Versini, and P. Witzgall. 2005. Antennal and behavioral responses of grapevine moth *Lobesia botrana* females to volatiles from grapevine. J. Chem. Ecol. 31: 77–87.

Tasin, M., E. Betta, S. Carlin, F. Gasperi, F. Mattivi, and I. Pertot. 2011. Volatiles that encode host-plant quality in the grapevine moth. Phytochem. 72: 1999–2005.

Varela, N., J. Avilla, S. Anton, and C. Gemeno. 2011. Synergism of pheromone and host-plant volatile blends in the attraction of *Grapholita molesta* males. Entomol. Exper. Appl. 141: 114–122.

Vogler U., A. S. Rott, C. Gessler, and S. Dorn. 2010. Comparison between volatile emissions from transgenic apples and from two representative classically bred apple cultivars. Transgenic Res. 19: 77–89.

Yoshinaga, N., H. T. Alborn, T. Nakanishi, D. M. Suckling, R. Nishida, J. H. Tumlinson, and N. Mori. 2010. Fatty acid-amino acid conjugates diversification in lepidopteran caterpillars. J. Chem. Ecol. 36: 319–325.

Yu, H., J. Feng, Q. Zhang, and H. Xu. 2015. (Z)-3-hexenyl acetate and 1-undecanol increase male attraction to sex pheromone trap in *Grapholita molesta* (Busck) (Lepidoptera: Tortricidae). Inter. J. Pest Manag. 61: 30–35.

Zakir, A., M. Bengtsson, M. M. Sadek, B. S. Hansson, P. Witzgall, and P. Anderson. 2013. Specific response to herbivore-induced de novo synthesized plant volatiles provides reliable information for host plant selection. J. Exper. Biol. 216: 3257–3263.