Mapping Autosomal Recessive Intellectual Disability: Combined Microarray and Exome Sequencing Identifies 26 Novel Candidate Genes in 192 Consanguineous Families

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Approximately 1% of the population worldwide are affected by intellectual disability (ID), the vast majority of whom currently receive no molecular diagnosis. Previous studies indicate high levels of genetic heterogeneity, with estimates of over 2500 autosomal ID genes, with the majority being autosomal recessive (AR). Here, we combined microarray genotyping, homozygosity-by-descent (HBD) mapping, copy number variation (CNV) analysis, and whole exome sequencing (WES) to identify disease genes/mutations for 192 multiplex Pakistani and Iranian consanguineous families ascertained for non-syndromic ID. We identified definite or candidate mutations (or CNVs) in 51% of families, in 72 different genes, including 26 not previously reported for ARID. The new ARID genes include nine with loss-of-function mutations (*ABI2*, *MAPK8*, *MPDZ, PIDD1*, *SLAIN1*, *TBC1D23*, *TRAPPC6B*, *UBA7*, and *USP44*), and missense mutations include the first reports of variants in *BDNF* or *TET1* associated with ID. The genes identified also showed overlap with *de novo* gene-sets for other neuropsychiatric disorders. Transcriptional studies show prominent expression in the prenatal brain. The high yield of AR mutations for ID signals that this approach has excellent clinical potential, will inform clinical diagnostics, including clinical whole exome and genome sequencing, for populations where consanguinity is common. As with other AR disorders, the relevance will also impact outbred populations.

Introduction

About 1% of the population worldwide is affected by intellectual disability (ID)[1] which can have a devastating impact on the lives of the affected individuals and their families, and is a major challenge at the clinical level. Genetic factors are involved in the etiology of 25-50% of ID cases[2]. The clinical presentation and etiology of ID are complex and highly heterogeneous, leading to a poor rate of molecular diagnosis, resulting in inadequate clinical management and counselling.

ID can be divided into two groups: syndromic (S) ID, where comorbid illness or physical features are present, and nonsyndromic (NS) ID, where no such comorbidities are present. Of ~700 known ID genes (S and NS), fewer than 50 genes are mutated in NS autosomal recessive ID (NS-ARID)[2]. X-linked ID may account for only 10-12 % of ID cases[3]. Dominant autosomal variants occurring *de novo* may contribute to a large proportion of sporadic cases, particularly in outbred populations[4-6]. Autosomal recessive (AR) variants also play a significant role in ID because recessive variants can remain in the population in a heterozygous form. Estimates suggest that there may be more than 2500 autosomal ID genes in total— the majority being recessive[7]. In populations with high levels of consanguinity most ID-causing mutations are recessive[7]. Even in outbred populations estimates are that 13-24% of ID cases are likely to be due to autosomal recessive causes[7].

Despite an increased diagnostic yield among those who receive testing[2] the vast majority with ID currently receive no molecular diagnosis[8]—a shortcoming that can impact health and lifespan. Recent studies indicate that the median age of death for males with ID is 13

years younger, and for females 20 years younger, than in the general population[9]. Although advances in genotyping and sequencing technology have accelerated the rate of gene discovery for ID[10], the majority of ID genes remain undetected. However, large scale ID family studies are making significant inroads[11]. Homozygosity mapping has been proven to be an effective method for gene identification in consanguineous populations[11-17]. Consanguineous marriages lead to a marked increase in frequency of severe recessive disorders[18]. Collectively, countries with levels of consanguinity higher than 10%, mainly from Africa, the Middle East and South Asia[19, 20], represent a population of ~1.3 billion. In Pakistan, ~62.7% of the population engage in consanguineous marriages, of which ~80.4% are first cousin marriages[21] . The rate of consanguineous marriages in Iran is estimated at 40%[22]. Here we present a study of multiplex ID families from Pakistan (N=176) and Iran (N=16), using microarray-based genotyping to identify autozygous regions (HBD shared by affected family members), coupled with whole exome sequencing (WES) to identify causal variants (see Figure 1 for workflow). In total, we have identified single candidate genes/variants for 88 families (50% were loss-of-function (LoF) mutations), and ten candidate pathogenic genomic variants (CNVs) in nine families.

Methods

Family recruitment

Institutional research ethics board consent was given for the study through the Centre for Addiction & Mental Health, Toronto, also through institutes at the recruiting sites (details in the Supplementary Methods). Families were recruited on the basis of diagnosis of ID in more than one individual (or ID and/or learning disability for N=13 families; or both ID and psychosis

within the family, N=5), but with no obvious dysmorphic features or comorbidities, and with parental consanguinity. Typically marriages were first or second cousin marriages, however in a number of families the exact relationship between the parents could not be established, but the marriage was within the same clan or caste. Written informed consent was obtained for all participants. Blood was drawn, and genomic DNA was extracted by standard methods. Cases of fragile X (tested using established methods[23, 24]), Down's syndrome and other clearly recognizable syndromes were excluded. Summary statistics for the families is given in Table 1.

Autozygosity Mapping

Autozygosity mapping was performed using microarray data from the Illumina Human CoreExome, Affymetrix Mapping 500K *Nsp*I, Affymetrix CytoScan HD and Affymetrix SNP 5.0 or 6.0. All genotyping was performed as per manufacturer's protocol, and data processed using either the Affymetrix Genotyping Console (500K and 5.0), Affymetrix ChAS Software Suite (CytoScan HD and SNP 6.0)or the Illumina GenomeStudio platform (Illumina CoreExome). All data was exported to a PLINK format for analysis using HomozygosityMapper[25] and FSuite[26]. Details are provided in the Supplementary Methods.

CNV Analysis

CNV analysis was performed to identify homozygous CNVs in HBD regions, or heterozygous CNVs that could indicate cases of intra-familial genetic heterogeneity, that should then be excluded from the HBD/WES analyses (details in the Supplementary Methods).

Whole Exome Sequencing (WES)

WES was performed for one or more affected member from each family, using sequencing facilities at CAMH, Toronto. Three different next generation sequencing platforms were used across the study, taking advantage of newer platforms as they became available for research at CAMH (SOLiD 5500 platform (Life Technologies) for 49 families (51 individuals), using a protocol reported previously[27]; Ion Proton platform/Ion Ampliseq™ Exome kit (Life Technologies) for 49 individuals from 30 families; Illumina HiSeq2500 platform, using ThruPLEX DNA-seq 96D kit (Rubicon, R400407) and SureSelect XT2 Target Enrichment (Agilent Technologies) system for 150 families).

Sequencing Alignment and Variant Calling

We used an in-house pipeline to map and call variants on the different types of sequencing data for this study. The pipeline is summarized in Supplementary Figure 1 and in Supplementary Methods.

Once putative variants are identified, Sanger sequencing was used to validate variants and determine if the variant segregates with the disease by testing parents, and other affected and unaffected individuals within the family.

Gene list construction

We combined the genes listed in Supplementary Table 3 with other genes for X linked or autosomal recessive intellectual disability mental retardation in OMIM. This full list is available as Supplementary Table 7. This gene list was used in the pathway and gene expression analyses (details in the Supplementary Methods).

Database searches across different neuropsychiatric and neurodevelopmental disorders, and functional and animal models

We have compared our variants to several databases including EpilepsyGene, Gene2Phenotype, Gene2Cognition, Schizophrenia Genebook, published disease-specific gene-sets[6], HGMD[28], OMIM[29], Zfin[30], targets of FMRP through high-throughput sequencing of RNAs by cross-linking immune-precipitation (HITS-CLIP) [31], and Mouseportal (http://www.sanger.ac.uk/science/collaboration/mouse-resource-portal, accessed May 2016). Searches were performed to identify overlapping genes/variants with our dataset

Results

Our study has identified single candidate genes/variants for 88 (81 autosomal recessive, 6 X-linked, and 1 *de novo* (heterozygous)) of the 192 families (Table 2 & 3, & Supplementary Table 3a). Twenty six of the genes identified in this cohort have not previously been reported for NS-ARID. An additional eleven genes were first reported previously from this cohort (*CC2D2A*[32]; *TCTN2*[33]; *TRAPPC9*[12]; *MAN1B1*[13]; *FBXO31*[27]; *METTL23*[14]; *FMN2*[15]; *DCPS*[16]; *HMNT*[17]; *NSUN2*[34]; *MBOAT7*[35l]). Most of these genes have since been reported in multiple ID families, including in outbred populations (e.g. *TRAPPC9*[36]; *MAN1B1*[37]). Likely pathogenic CNVs were identified in at least nine families (not including the VPS13B and TUSC3 deletions in AN51 and AN21 respectively).

Homozygous Loss-of-Function Mutations

Homozygous truncating LoF mutations were found as single candidate variants in 43 families, including nine genes that are previously unpublished in relation to NS-ARID, namely *ABI2*, *MAPK8*, *MPDZ, PIDD1*, *SLAIN1*, *TBC1D23*, *TRAPPC6B*, *UBA7*, and *USP44.* None of these genes have been noted as tolerant to loss of function through WES of 3,222 British adults of Pakistani origin[38].

We identified the same nonsense mutation in *PIDD1* in two unrelated Pakistani families (ASMR105 and ASMR110; Figure 2). *PIDD1* encodes p53-Induced Death Domain Protein (PIDD; MIM 605247). Gln863* disrupts the death domain (DD), through which PIDD1 interacts with other DD proteins such as RIP1 or CRADD/RAIDD. Truncating mutations in *CRADD* have previously been reported for NS-ARID (MRT34[39]), and thus our findings add support to the involvement of PIDD-related pathways, in the etiology of intellectual disability.

Homozygous truncating mutations in *TRAPPC9* have been reported for NS-ARID [12, 36, 40-43], thus it is of much interest that we have identified a nonsense mutation in a gene encoding a second member of the same protein trafficking particle complex, namely *TRAPPC6B*.

A nonsense mutation was identified in *SLAIN1* segregating fully in family PK68 (Figure 2). *SLAIN1* and *SLAIN2* encode microtubule plus-end tracking proteins that have been shown to be crucial to normal axonal growth in developing hippocampal neurons[44].

We report here a homozygous nonsense mutation in the gene *UBA7* for family PK34. *UBA7* encodes ubiquitin-activating enzyme 7, believed to be involved in the ubiquitin conjugation pathway[45]. Family PK34 is one of three families in the study that did not meet criteria for ID, and instead were reported as having a learning disorder, and are considered

relatively high functioning. We also report that this variant is present at a relatively high frequency in the South Asian population (MAF=0.0054; ExAC database), and two homozygotes for this variant were among the control group. Thus, this variant and gene could be a risk factor for a much milder form of cognitive disability, and thus potentially present in the control South Asian population (N>8,000) used in the ExAC database.

We also identified a LoF mutation in *TMEM135*, previously been reported in the large Iranian NS-ARID cohort by Najmabadi et al, 2011 (in which a missense mutation, Cys228Ser was reported)[11], and in Wright et al, 2014 (missense)[46], also as a quantitative trait locus for intelligence[47].

Missense mutations

We identified homozygous missense variants as single candidate variants in 43 families, including 16 for which the genes identified have not previously been reported for NS-ARID (*AFF3*, *BDNF*, *CAPS*, *DMBT1*, *DUOX2*, *EXTL3*, *FBXO47*, *LAMC1*, *MAP3K7*, *SDK2*, *SPATA13*, *SUMF2*, *SYNRG*, *TET1*, *VPS35*, and *ZBTB11*). Although LoF mutations are frequently more convincing than missense, a number of the homozygous missense changes we report here are or particular interest, due to the known nature or function of the protein or the likely effect of the amino acid substitution on protein function. For instance, here we report a homozygous missense change Met122Thr in the gene for the brain-derived neurotrophic factor, *BDNF*, which has been implicated in many studies of neuropsychiatric disorders[48], and is a known gene target of *MECP2*, the Rett syndrome gene[49, 50] This variant replaces a methionine- a large, non-polar residue with an S-methyl thioether side chain- that is fully conserved across the vertebrate

lineage, with threonine- a small, polar residue with a hydroxyl side group. This variant is reported with a frequency of 0.0001647 in ExAC (in South Asians MAF=0.001211), with no homozygotes, and has not previously been reported in any publications. The BDNF protein is important for the survival, differentiation and development of neurons within the central nervous system (CNS). Hence, the identification of Met122Thr in connection with a cognitive disability will likely be of great interest, and functional studies of the effects of Met122Thr on BDNF function are indicated.

We report a homozygous Lys2056Asn change in *TET1* for family PK70 that is not present in any variant or mutation databases. Methylcytosine dioxygenases TET proteins play a role in the DNA methylation process and gene activation, and TET1 is an important regulator of neuronal differentiation. TET1 is implicated in SCZ and BD by down-regulating *GAD1*, *RELN*, and *BDNF* genes through epigenetic mechanisms in the prefrontal cortex, and in the cerebellum in autism[51, 52]. Significant upregulation of *TET1* mRNA in the parietal cortex of psychosis patients has also been reported[53]. *Tet1* knockout shows impaired hippocampal neurogenesis and cognitive deficits in mouse [54], and abnormal brain morphology in zebrafish[55]. The large basic, charged, hydrophobic lysine residue at this position is conserved in mammals (or glutamic acid in non-mammalian vertebrates) and is replaced here by a small polar asparagine residue.

The homozygous His880Gln change identified in the zinc finger/BTB domain gene *ZBTB11* in family AN50 disrupts a canonical Zn^{2+}-binding residue in one of the zinc fingers, and is likely to result in an alteration in the specificity of DNA-binding/gene regulation. *AFF3* is an autosomal homolog of the X-linked ID gene, *AFF2* (MRX-FRAXE; MIM 309547). As with *AFF2*,

AFF3 is also associated with a fragile site (FRA2A) for which expansion of a CGG trinucleotide repeat triggers hypermethylation and gene silencing in association with neurodevelopmental disorders[56]. The Gly1215Val variant identified here is situated within a five-amino acid C-terminal motif (Gln-Gly-Leu-His-Trp) that is highly conserved across the vertebrate lineage.

For family AS70, a missense mutation was identified in exon 12 of the gene *MAP3K7*. Although missense mutations in this gene have recently been reported for frontometaphyseal dysplasia[57] (FMD) and cardiospondylocarpofacial syndrome (CSCF syndrome[58], family members of AS70 have no skeletal dysplasia or obvious dysmorphic features, suggesting even greater pleiotropy at this gene. We note that, while the Arg410Gln mutation we identified is located in exon 12, which is alternatively spliced and present in both transcripts B and C, but neither A nor D, all *MAP3K7* mutations so far reported for FMD or CSCF syndrome are located in canonical exons (i.e. in all four transcript variants). Furthermore, analysis of mouse RNAseq data from the ENCODE UW project (through UCSC Genome Browser) suggest that while exon 12 is expressed in many adult tissues, including brain, it is absent from skeletal muscle.

Families with multiple variants

For eleven additional families between two and four putative damaging variants were identified after filtration that fulfilled criteria and segregated in the families, but without functional evidence in support of pathogenicity it is not yet possible to narrow these variants down to a single candidate (Supplementary Table 3B).

X-linked variants

A number of the families were compatible for both AR and X-linkage, and several variants or CNVs on the X chromosome have been identified, including two variants in *ATRX*, and a 6.7Mb interstitial duplication on Xp22.31-p22.2.

We report a missense mutation, Ser57Pro, in the X-chromosomal gene *MAGEA11*, segregating in family PK87. A Gln4Arg variant in MAGEA11, was recently reported among a cohort of X-linked ID families[59]. MAGEA11 shows protein interaction with TRMT1 — also reported here for NS-ARID— by yeast-two-hybrid[60-62].

In another large multiplex and multi-branch family the missense mutation Arg190His of the X-linked *MECP2* gene is present in a hemizygous form in a single male with mild ID, as well as in a heterozygous form in several females with mild ID or mild ID with psychosis or depression. Interestingly, for the female heterozygotes with ID, all appeared cognitively normal until age ~9 years, when cognitive regression started, and for the single male hemizygote, cognitive regression began much earlier, <5 years of age. The family also has several males with schizophrenia, with onset ~18 years but without cognitive regression amounting to intellectual disability, who are wild type for this variant (see Supplementary Figure 2). The Arg190 residue is a critical DNA-binding amino acid within an AT-hook domain, and previously a *de novo* mutation at this residue, Arg190Cys, has been reported in a SCZ patient[63].

Genes with 'hits' in multiple families:

Confidence in gene discovery relies on the identification of multiple affected families with mutations in the same gene, and/or replication in further studies. However, due to the anticipated high degree of genetic heterogeneity for NS-ARID, large sample sizes are typically

required in order to find multiple families with mutations in the same gene. Comparison across different studies thus becomes vital. Of the ARID genes reported here for the first time, several are conspicuous by the presence of mutations in multiple families. In our study, the same nonsense mutation in *PIDD1* occurs in two apparently unrelated Pakistani families. We also provide confirmatory families/mutations for recently reported ARID genes such as *C12ORF4*, *CCDC82*, *MBOAT7*, *IMPA1*, *TMEM135*, *PGAP2*, *GPT2* (2 families), *TDP2* (2 families), and *GMPPA* (Table 3, and Supplementary Table 3). Also, mutations in the gene for the ID-associated brain malformation polymicrogyria, *GPR56*, are present in three families in this study, and could thus represent a relatively large proportion of ID families in these populations.

Families with mutations in previously identified genes for metabolic or hormonal disorders:

We report here three mutations in thyroid dyshormonogenesis (TDH) genes *TPO* (thyroid peroxidase), *TG* (thyroglobulin)[64]), also *DUOX2* (thyroid oxidase 2). With adequate clinical resources available in most developed countries, many of these would likely have been diagnosed, and in some cases, for example, those with mutations in N-acetylglutamate synthase (*NAGS*), *TPO*, *TG* and *DUOX2*, early treatment would have prevented ID development. Given the prevalence of mutations in these genes in a relatively modest number of families, it is likely that these disorders are relatively common causes of ID in populations where consanguinity is common but access to clinical diagnostics is poor. Mutations in *GNE* have previously been linked with sialuria (dominant; MIM 269921) — an extremely rare metabolic disorder— and Nonaka myopathy (recessive; MIM 605820), whereas here we report a homozygous missense mutation in a family with just ID and no myopathy. Although we were

unable to get biochemical analysis on this family, we anticipate that this discovery represents a previously unreported recessive form of sialuria.

Copy Number Variation (CNV) analysis:

In addition to HBD analysis, microarray data was used for CNV analysis, firstly to look for possible intra-familial genetic heterogeneity with large, probably pathogenic heterozygous loss/gain CNVs, and secondly to look for homozygous genic CNVs within mapped HBD regions. For the former, several candidate pathogenic CNVs were identified (Supplementary Table 4), including an 8.4Mb deletion of 2q14.1-q14.3 (chr2:116583565-124954598) in one of the two affected individuals in family PK117. Deletions within this region have previously been reported for autism spectrum disorder[65], and in a patient with mild holoprosencephaly spectrum phenotype[66]. An overlapping, but slightly proximal deletion (~chr2: 114188161-119321989) has been reported to be compatible with normal phenotype[67]. In one family, PK28, all three affected males in one branch were shown to have a large (6.7Mb) interstitial duplication spanning cytobands Xp22.31-p22.2.

For homozygous CNVs in HBD regions, we have identified homozygous deletions in known ARID genes in several families (Table 3, and Supplementary Table 3), including a 170 kb homozygous deletion spanning 9 out of 10 exons of NS-ARID gene, *TUSC3*, in family AN21[68], also a 51 kb deletion spanning exons 37 to 40 of the Cohen syndrome gene, *VPS13B*, in family AN51[69]. Homozygous disrupting loss CNVs include a 50 kb deletion spanning exons 2 and 3 of *RAB8B* in PK95-7, however, this was not in a shared HBD region, and was thus considered as a potential case of intra-familial genetic heterogeneity.

Databases for neuropsychiatric disorders:

It has been noted that genes relevant for ID are frequently also identified in individuals with autism spectrum disorders (ASD), and epilepsy (which both frequently also present with ID). In addition there is growing support for overlap of ID genes with other neuropsychiatric disorders[70]. For example, genes such as *NRXN1* and *ANK3* have been linked with SCZ, BD and ASD by CNV and/or genome-wide association studies are also known ARID genes (*NRXN1*: PTHLS2, MIM 614325; *ANK3*: MRT37, MIM 615494). For this reason, we attempted to evaluate the genes identified here in variant databases or among gene lists for such disorders. We have screened various epilepsy or neuropsychiatric disease-specific databases or published datasets for the presence of variants in these genes. In the SCZ/control exome sequence database Genebook[71, 72], none of the variants we have identified in Table 2 were present. However, for several of the genes, there is a reported increased burden of rare variants in SCZ cases versus controls ($p<0.05$ for *VPS35*, *SYNRG*, *DMBT1*, *ALPI*, and *NEU4*; $p<0.002$ for *SLC13A5*), although when corrected for multiple testing, no individual gene-based tests achieved statistical significance.

In epilepsy databases, we identified *ATRX*, *MECP2*, *SLC13A5*, and *ST3GAL3*, all of which have been reported in cases of epilepsy as well as ID. These four genes are either inherited as autosomal recessive or X-linked recessive. *ATRX* and *MECP2* interact with each other and are involved in chromatin binding and gene regulation[73], and mutations in either gene can have diverse effects on DNA methylation patterns and brain development.

Protein interaction analysis

We used BioGrid (http://thebiogrid.org) to identify protein interactors for each of the 67 different ARID genes identified among our population (Supplementary Table 3A) as well as *GRIN2B* (Supplementary Table 3C). Interacting proteins are listed, along with gene ontology processes, functions, and cellular compartments, in Supplementary Table 5.

Pathway analyses

In agreement with past findings of genetic heterogeneity, we observed limited overlap with the gene ontology gene sets. Of the 14,312 sets tested, 16 survive multiple test correction (q-value < 0.05) and a few top-ranked terms are of interest (Supplementary Table 6). Protein glycosylation which is known to be associated with intellectual disability was ranked 6th with 9 overlapping genes (corrected $p < 0.05$). As mentioned above, *TPO*, *DUOX2*, and *TG* are known to be involved in thyroid hormone generation (rank: 14, corrected $p < 0.05$). The smoothened signaling pathway is ranked 32nd with 4 overlapping genes (*NDST1*, *CC2D2A*, *TCTN2*, and *BBS7*).

Anatomical expression analyses

With the exception of the spinal cord, all neural tissues were enriched in the expression of the ID genes (corrected $p < 0.0001$, Supplementary Table 8). The frontal cortex was the most enriched with 1.4 times the expected number of expressed genes. While the brain does express a majority of all genes, it appears that the ID genes show specificity. Of the 17 fetal tissues tested, the female gonad and testis have the highest number of expressed genes but do not show enriched expression of the ID list (corrected $p > 0.12$).

Developmental brain expression analyses

Across the developmental transcriptome, the ID genes are expressed at higher levels in the normal prenatal brain (Figure 3). The amygdaloid complex shows the most consistent

enrichment (5 specimens with significantly higher expression of ID genes). Fetal brain samples from 21 and 24 post-conception weeks show the highest number of regions with significant expression (> 4 or more). In contrast, enriched expression in the postnatal brain samples is not observed. This prenatal pattern found in exon microarray expression profiles is also observed in RNA sequencing measurements in a largely overlapping set of samples from the same resource (BrainSpan, Supplementary Figure 3,). The RNA sequencing measurements show donor specific patterns with many enriched regions. These global patterns are not consistent across donors of the same age, suggesting the RNA sequencing data may have normalization artefacts that are not seen in the microarray measurements. Grouping the ID gene list into genes associated with glycosylation or hormones and metabolism shows above average prenatal expression for all groupings (Figure 3). Genes with metabolic or hormonal associated function have the most stable trajectory. We also observe higher expression for these groups in the prenatal brain in the BrainCloud[74] resource that assayed the prefrontal cortex across human development (Supplementary Figure 4).

Discussion

This study describes a cohort of 192 multiplex ID families from Pakistan and Iran. With combined microarray genotyping, homozygosity-by-descent (HBD) mapping, copy number variation (CNV) analysis, and whole exome sequencing (WES) we identified definite or candidate mutations (or likely pathogenic CNVs) in 51% of families, in 72 different genes, including 26 not previously reported for ARID.

We note that 50% of the variants we report as single probable mutations are LoF changes. This compares well with the earlier study reported by Najamabadi et al, 2011[11], in which 50 new genes for NS-ARID were reported, of which 40% had LoF mutations. In general, LoF mutations provide a higher degree of confidence of disease association over missense mutations.

There are a number of likely reasons that genes/mutations were not found for some families: 1. Intra-familial etiologic heterogeneity; 2. poor WES depth of coverage at the etiologic gene/mutation; 3. causative mutations may be intronic or intergenic, and thus not picked up by WES. Whole genome sequencing may address this issue; 4. Common variants have been reported in association with many complex diseases including traits like intelligence or cognitive ability[75, 76]. It is plausible that a proportion of ID cases with familial aggregation are caused by variants, possibly interacting, that are common in the general population but with low penetrance. These variants working together could potentially cause ID, however, this hypothesis has not been explored in ID. 5. non-genetic factors may be prevalent in some families, e.g. prenatal or perinatal insult[77].

Given that there may be an overlap in genetic etiology of neurodevelopmental and neuropsychiatric disorders, we cross-referenced our ID gene list with those from studies of other neuropsychiatric/neurodevelopmental disorders. We see multiple instances of genes identified for ARID that are implicated across disorders (see Figure 4). For instance, of the newly identified genes , *SLAIN1* is listed as one of the top-ranked genes for the burden of variants among a large cohort with SCZ (N=1,392)[78]. Data from the Autism Sequencing Consortium[79]

reports *de novo* LoF mutations in *SPATA13* and *TBC1D23*, as well as *de novo* missense mutations in *ABI2*, *TET1*, and *SYNRG*. Li et al (2016)[80] report variants in both *MPDZ* and *SPATA13* in ASD cohorts, and a *de novo* missense variant in *MAPK8* in SCZ[72]. Also, whereas heterozygous (typically *de novo*) mutations in *SCN1A* have been linked with ASD, ID, and epileptic encephalopathy, the missense *SCN1A* variant we report here is homozygous. A number of ARID genes (e.g. *NRXN1*, *CNTNTAP2*, *ANK3*) have already been implicated in neuropsychiatric disorders by GWAS or CNV studies (as heterozygous). We speculate that, in addition to pleiotropy with other neuropsychiatric/neurodevelopmental disorders, the mode of inheritance may also be variable, (e.g. *SPATA13*: homozygous missense variant in ID (this study) and *de novo* LoF variant in ASD[80]; *HNMT*: homozygous missense variant in ID[17], and heterozygous splice mutation in SCZ (Genebook)). ID may be conceptualized as a much severer form of neurodevelopmental disorder, compared to ASD and SCZ.

Transcriptional analysis of our gene set combined with known ARID genes shows greater levels of transcription in the prenatal brain than the postnatal or adult brain (Figure 3A), and in particular higher levels in frontal cortex, hippocampus and amygdala (Figure 3B). ID is primarily a disorder of brain development, and thus it is reassuring to observe the ID genes showing relevant patterns of spatiotemporal expression. Pathway analysis of the gene set showed a few significant pathways; thyroid metabolism was prominent, as was protein glycosylation and the Smoothened signaling pathway. So far this and other studies have predicted involvement of numerous different pathways in ID, which is probably a reflection of the huge genetic heterogeneity in ID.

Our findings and those of other groups studying the genetics of NS-ARID will be invaluable for the development of targeted sequencing gene panels for diagnostic screening, and for the development of clinical whole exome and whole genome sequencing. In this regard, both the replication of previous findings and new discoveries in this study are important. With replication the confidence in the validity of the findings is enhanced, and new discoveries open up the opportunities for further exploration of the functions and biological pathways of the newly associated genes. Cumulatively, such studies are also mapping genes across the human genome for which LoF mutations are viable, and their roles in human development are thus likely to be amenable for further study and comparison with similar mutations in model organisms. Furthermore, a more complete picture is being assembled of the molecular components and mechanisms that are important for the development of a fully functioning central nervous system, as well as documenting points in the mechanisms that are most vulnerable to genetic mutation. Through comparison with similar studies, we also note that there are many more such discoveries to be made to complete the picture. The ultimate challenge, to devise targeted therapeutic strategies for ID patients, is thus a step closer.

The discovery of disease causing mutations in consanguineous families immediately creates opportunities for carrier screening among relatives and prevention of ID. This is a direct benefit to the families' communities and to public health. In addition, finding genetic causes for ID makes it possible to subgroup ID individuals by gene or pathway. This can lead to the development of cohorts which can then be studied prospectively for the natural course of the disease, health complications, and also can be targeted for therapeutic strategies. This is a step towards personalized medicine in this important clinical population.

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Conflict of Interest

The authors report no conflicts of interest.

Supplementary information is available at *Molecular Psychiatry*'s website.

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Figure Legends:

Figure 1. Microarray and exome sequencing work-flow

Figure 2. Pedigrees and HomozygosityMapper output for eight of the families. The locus harboring the mutation is indicated in the HomozygosityMapper plots with yellow shading. HBD regions for each family, confirmed as autozygous and haploidentical, are provided in Supplementary Table 2. The gene name and mutation (at the protein level) is indicated), and genotypes for available family members. Additional pedigrees and HBD plots are provided in Supplementary Figure 2.

Figure 3. Spatiotemporal expression of ID genes in human brain development: a) Scaled expression trajectories of the ID Genes, averaged across brain regions. Solid lines are average expression for a group of genes. The lighter and thinner lines show trajectories of individual genes. Average expression for each gene group is depicted with points (green: glycosylation associated (14 genes), blue: hormone or metabolic (20), purple: remainder of the ID Genes (66) and red: remaining genes assayed). Local regression was used to smooth the z-scored expression values for individual genes and gene group averages (LOESS). A vertical dashed line marks birth. **b)** Heatmap showing brain samples that are enriched for specific expression of ID genes. For each brain region and brain combination, z-scored expression values for the ID genes was compared against all other genes (Wilcoxon one sided test, FDR corrected p-values). Significance levels are indicated by black (non-significant result), red ($p < 0.05$), orange ($p < 0.005$) and yellow ($p < 0.0005$). Missing values are shown in grey. Brain regions include: ventrolateral prefrontal cortex (VFC), dorsolateral prefrontal cortex (DFC), orbital frontal cortex (OFC), anterior (rostral) cingulate (medial prefrontal) cortex (MFC), posterior (caudal) superior

temporal cortex (area 22c) (STC), inferolateral temporal cortex (area TEv, area 20) (ITC), posteroventral (inferior) parietal cortex (IPC), primary motor cortex (area M1, area 4) (M1C), primary somatosensory cortex (area S1, areas 3,1,2) (S1C), primary auditory cortex (core) (A1C), primary visual cortex (striate cortex, area V1/17) (V1C), hippocampus (hippocampal formation) (HIP), amygdaloid complex (AMY), mediodorsal nucleus of thalamus (MD), striatum (STR) and the cerebellar cortex (CBC).

Figure 4. Cross-disorder overlap: The Venn diagrams shown indicate genes for which **a.** variants have been reported in other neuropsychiatric or neurodevelopmental disorders, either homozygous, compound heterozygous, or *de novo*, through searches of published gene-list including databases such as EpilepsyGene, Gene2Phenotype, Gene2Cognition, Schizophrenia Genebook, published disease-specific gene-sets[6], HGMD[28], OMIM[29]; **b.** functional or animal models with relevant phenotypes have been reported including resource such as Zfin[30] , targets of FMRP[31] and Mouseportal (http://www.sanger.ac.uk/science/collaboration/mouse-resource-portal, accessed May 2016). Genes newly reported here are in red text.

Table 1: Summary statistics for Iranian and Pakistani family cohorts.

Source Country	Total Families	Total DNA Samples	Total Affected	Mean Number of ID Affected/Family	*F* Coefficient of inbreeding	Number of Families with Gene/Mutation Identified: ARID (XLID; *de novo*)	Number of Families with Pathogenic CNV Identified
Iran	16	193	58	4.375	0.058034056	6 (0;0)	0
Pakistan	176	1379	627	3.5625	0.084862578	81 (6; 1)	9[a]

[a]In family PK73 two different likely pathogenic CNV losses were identified, but only counted here as a single family with pathogenic CNV.

Table 2: Novel candidate genes and sequence variants identified for the Pakistani and Iranian ID families. A list of all variants identified including known syndromic ARID or XLID genes identified in the cohort is give in Supplementary Table 3a, and 3b for families where between 2 and 4 variants were identified.. See Supplementary Table 3 for *in silico* predictions of effects of variants.

	Family	Gene	Genome (ExAC MAF/S.Asian MAF)	cDNA (Protein)
1	PJ7	*LAMC1*	Chr1:183083732A>T (0/0)	NM_002293.3:c.1088A>T (p.His363Leu)
2	PK113	*AFF3*	Chr2:100167973C>A (0/0)	NM_002285.2:c.3644G>T (p.Gly1215Val)
3[a]	AS61	*ABI2*	Chr2:204245039C>T (0/0)	NM_005759.4:c.394C>T (p.Arg132*)
4[b]	PK34	*UBA7*	Chr3:49848458C>A (8.01E-04/5.45E-03)	NM_003335.2:c.1189G>T (p.Glu397*)
5	PJ9	*TBC1D23*	Chr3:100037953delA (0/0)	NM_018309.4:c.1683delA (p.Glu562Argfs*3)
6	AN50	*ZBTB11*	Chr3:101371344A>C (0/0)	NM_014415.3:c.2640T>G (p.His880Gln)
7	AS70	*MAP3K7*	Chr6:91254333C>T (1.65E-05/1.21E-04)	NM_145331.2:c.1229G>A (p.Arg410Gln)
8	PJ2	*SUMF2*	Chr7:56146063A>G (8.70E-06/7.73E-5)	NM_015411.2:c.740A>G (p. Tyr247Cys)
9	AS20	*EXTL3*	Chr8:28,575,513C>G (0/0)	NM_001440.3:c.1937C>G (p. Ser646Cys)
10[c]	AS66	*MPDZ*	Chr9:13109992_13109995delGAAA (0/0)	NM_003829.4:c.5811_5814delTTTC (p.Phe1938Leufs*3)
11	AS22	*MAPK8*	Chr10:49628265_49628265delC (0/0)	NM_002750.3:c.518delC (p.Arg174Glyfs*8)
12	PK70	*TET1*	Chr10:70451328G>T (0/0)	NM_030625.2:c.6168G>T (p.Lys2056Asn)
13	IDSG19	DMBT1	Chr10:124356559C>G (0/na)	NM_007329.2:c.2906C>G (p.Thr969Arg)
14[d]	AS105 & 110	*PIDD1*	Chr11:799453G>A (0/0)	NM_145886.3:c.2587C>T (p.Gln863*)
15	PK135	*BDNF*	Chr11:27679747A>G (1.65E-04/1.21E-03)	NM_001709.4:c.365T>C (p.Met122Thr)
16	PK94	*USP44*	Chr12:95927147_95927160delinsA (0/0)	NM_032147.3:c.873_886delinsT (p.Leu291Phefs*8)
17	AS23	*SPATA13*	Chr13:24797332C>T (4.71E-05/0)	NM_001166271.1:c.265C>T (p.Arg89Trp)
18	PK68	*SLAIN1*	Chr13:78320722_78320722delA (0/0)	NM_144595.3:c.135delA (p.Thr49Hisfs*96)
19	IDH3	*TRAPPC6B*	Chr14:39628712G>A (1.65E-05/na)	NM_177452.3:c.124C>T (p.Arg42*)
20	PK79	*VPS35*	Chr16:46705735T>G (0/0)	NM_018206.4:c.1406A>C (p.Gln469Pro)
21	PK91	*SYNRG*	Chr17:35896199C>T (8.66E-06/0)	NM_007247.4:c.3548G>A (p.Arg1183His)
22	IDSG29	*FBXO47*	Chr17:37107906G>C (0/na)	NM_001008777.2:c.544C>G (p.Arg182Gly)
23	AN53	*SDK2*	Chr17:71431658C>T (5.82E-05/0)	NM_001144952.1:c.1126G>A (p.Gly376Ser)
24	AN48	*CAPS*	Chr19:5914970G>A (2.17E-04/1.27E-04)	NM_004058.3:c.281G>A (p.Arg94Gln)
25	PJ6	*GPR64*	ChrX:19055718T>C (8.45E-05/0)	NM_005756.3:c.191A>G (p.Asn64Ser)
26	PK87	*MAGEA11*	ChrX:148796213C>T (0/0)	NM_005366.4:c.169T>C (p.Ser57Pro)

a. The homozygous nonsense mutation reported for family AS61 is located within a large HBD region shared between just two of the four affected individuals. The other two are different phenotypically distinct, with ID and myopathy. Support for this gene comes from functional studies, including with *Abi2* null mice[96].

b. Two homozygotes for this variant are indicated in the ExAC South Asian population (N=8,251 individuals). N.B. affected individuals in PK34 have learning disability rather than ID. The South Asian cohort used in ExAC were part of the Pakistan Risk of Myocardial Infarction Study (PROMIS), and would not have excluded subjects in this category.

c. The *MPDZ* mutation reported here is not within a significant region of HBD, and is present as homozygous in the affected mother and the first-born affected son. The affected brother and sister are both heterozygous, but the father is wild type homozygous, suggesting non paternity for the first child.

Families AS105 and AS110 both carry the same mutation in *PIDD1*, and share a common haplotype, however, these families are apparently not closely related. Using Prest-plus[81] to estimate IBD using Maximum Likelihood Estimation, it was confirmed that the two families are separated by at least four generations, which is comparable to

relatedness for the background population, and thus AS105 and AS110 are considered unrelated. na=not applicable, as Iranian exome data is not available in ExAC database.

Table 3: Candidate genes and sequence variants identified for the Pakistani and Iranian ID families in support of previously reported non-syndromic ID genes. Asterisks indicate genes/variants already reported by us in this cohort For subjects where the phenotype is clearly NS-ARID, despite prior association of the gene with S-ARID (e.g. *LRP2* and *MECP2*), we have included the variant in this list. See Table S3 for *in silico* predictions of effects of variants.

	Family	Gene	Disease; MIM #	Genome (ExAC MAF/S.Asian MAF)	cDNA (Protein)	Ref
1	AN49	*FMN2**	MRT47; 616193	Chr1:240370627_240370628delinsG (0/0)	NM_020066.4:c.2515_2517delinsCG (p.Thr839Argfs*48)	[16]
2[a]	AS114	*FMN2*	MRT47; 616193	Chr1:240370641_240370642delCT (0/0)	NM_020066.4: c.2529_2530delCT (p.Ser844Cysfs*6)	
3	PJ12	*LRP2**	222448	Chr2:170027106C>T (2.47E-04/9.08E-04)	NM_004525.2: c.11335G>A (p.Asp3779Asn)	[90]
4	IDH10	*HNMT**	MRT51; 616739	Chr2:138727776G>A (1.66E-05/na)	NM_006895.2:c.179G>A (p.Glu60Asp)	[18]
5[b]	PK11	*TUSC3*	MRT7; 611093	Chr8:15480694C>T (0/0)	NM_006765.3:c.244C>T (p.Arg82*)	[91]
6	AN21	*TUSC3**	MRT7: 611093	Chr8:15,521,688_15,692,362del (N/A)	NM_178234 del exons 5-10	[78]
7	PK54	*IMPA1*		Chr8:82583195_82583196delinsC (0/0)	NM_005536.3:c.544_545delinsG (p.Leu182Valfs*54)	[45]
8	AS19	*TRAPPC9**	MRT13; 613192	Chr8:141407724G>A (8.24E-06/6.06E-05)	NM_031466.7:c.1423C>T (p.Arg475*)	[13]
9	AS102	*PGAP2*	MRT17; 614207	Chr11:3846254G>C (4.95E-05/3.64E-04)	NM_014489.3:c.713G>C (p.Arg238Pro)	[42]
10	PK62	*TMEM135*		Chr11:87032300delTT (0/0)	NM_022918.3:c.1304_1305delTT (p.Phe435Serfs*31)	[12]
11	AS17	*CCDC82*		Chr11:96117377G>A (1.65E-05/6.05E-05)	NM_024725.3:c.535C>T (p.Arg179*)	[82]
12	ZA5	*C12ORF4*		Chr12:4599717delA (0/0)	NM_020374.2:c.1537delT (p.Ser513Leufs*7)	[83, 84]
13	PJ5	*DCPS**	ARS; 616459	Chr11:126208295G>A (8.25E-06/0)	NM_014026.3: c.636+1G>A (15 amino acid insertion)	[17]
14[c]	PK12	*GPT2*	MRT49; 616281	Chr16:46918865C>G (0/0)	NM_133443.2:c.238C>G (p.Gln80Glu)	[92]
15	PK85	*GPT2*	MRT49; 616281	Chr16:46956326C>T (3.33E-05/1.89E-04)	NM_133443.3:c.1210C>T (p.Arg404*)	[92]
16	AS72	*FBXO31**	MRT45; 615979	Chr16:87369054_87369059delinsT (0/0)	NM_024735.3:c.847_852delinsA (p.Cys283Asnfs*81)	[28]
17	PK33	*NAGS*	NAGSD; 237310	Chr17:42085083C>T (7.74E-05/4.54E-04)	NM_153006.2:c.1393C>T (p.Arg465Trp)	[93]
18	PK31	*METTL23**	MRT44; 615942	Chr17:74729449C>T (0/0)	NM_001080510:c.397C>T (p.Gln133*)	[15]
19	IAID3	*METTL23**	MRT44; 615942	Chr17: 74729211_74729215del (0/0)	NM_001080510.3:c.236_240del (p.Thr80Glyfs*20)	[15]
20	AN37	*TRMT1*		Chr19:13223779_13223810del (0/0)	NM_017722.3:c.657_688del (p.Gln219Hisfs*22)	[12]
21	PJ3	*MBOAT7*		Chr19:54684518_54684524del (0/0)	NM_024298.3:c.820_826del (p. Gly274Profs*47)	[35]
22	AS101	*ARHGEF6*	MRX46; 300436	ChrX:135829705A>G (1.14E-05/9.91E-05)	NM_004840.2:c.296T>C (p.Val99Ala)	[94]
23	PK55	*MECP2*	MRXS13; 300055	ChrX:153296710C>T (0/0)	NM_004992.3:c.569G>A (p.Arg190His)	[95]

a. Affected individual in AS114 with *FMN2* mutation is a genocopy, and genes/mutations for the other four affected individuals in this pedigree have yet to be established.

b. Homozygous nonsense mutation in *TUSC3* is not in shared HBD region, and does not segregate with other affected members of family PK11.

c. Missense change in *GPT2* is also predicted to affect a splice donor site (BDGP (www.fruitfly.org) splice predictor score: WT: 0.78; mut: 0.58).

na=not applicable, as Iranian exome data is not available in ExAC database.

Supplementary Information:

Supplementary Methods

Supplementary Figure 1: Bioinformatic pipeline for whole exome sequence (WES) analysis.

Supplementary Figure 2: Pedigrees, HomozygosityMapper output and FSuite circo-plots for families with single variants identified, in addition to those shown in Figure 2.

Supplementary Figure 3: Spatiotemporal expression of ID genes in human development using RNA sequencing data.

Supplementary Figure 4: Developmental expression pattern of ID genes in the human prefrontal cortex.

Supplementary Table 1: Family statistics

Supplementary Table 2: Homozygosity-by-descent/autozygosity shared regions, as defined using HomozygosityMapper, cross-referenced with FSuite.

Supplementary Table 3: Mutations identified per family. A. Single homozygous variant identified. B. Two to four variants identified. C. Dominant/de novo mutation identified.

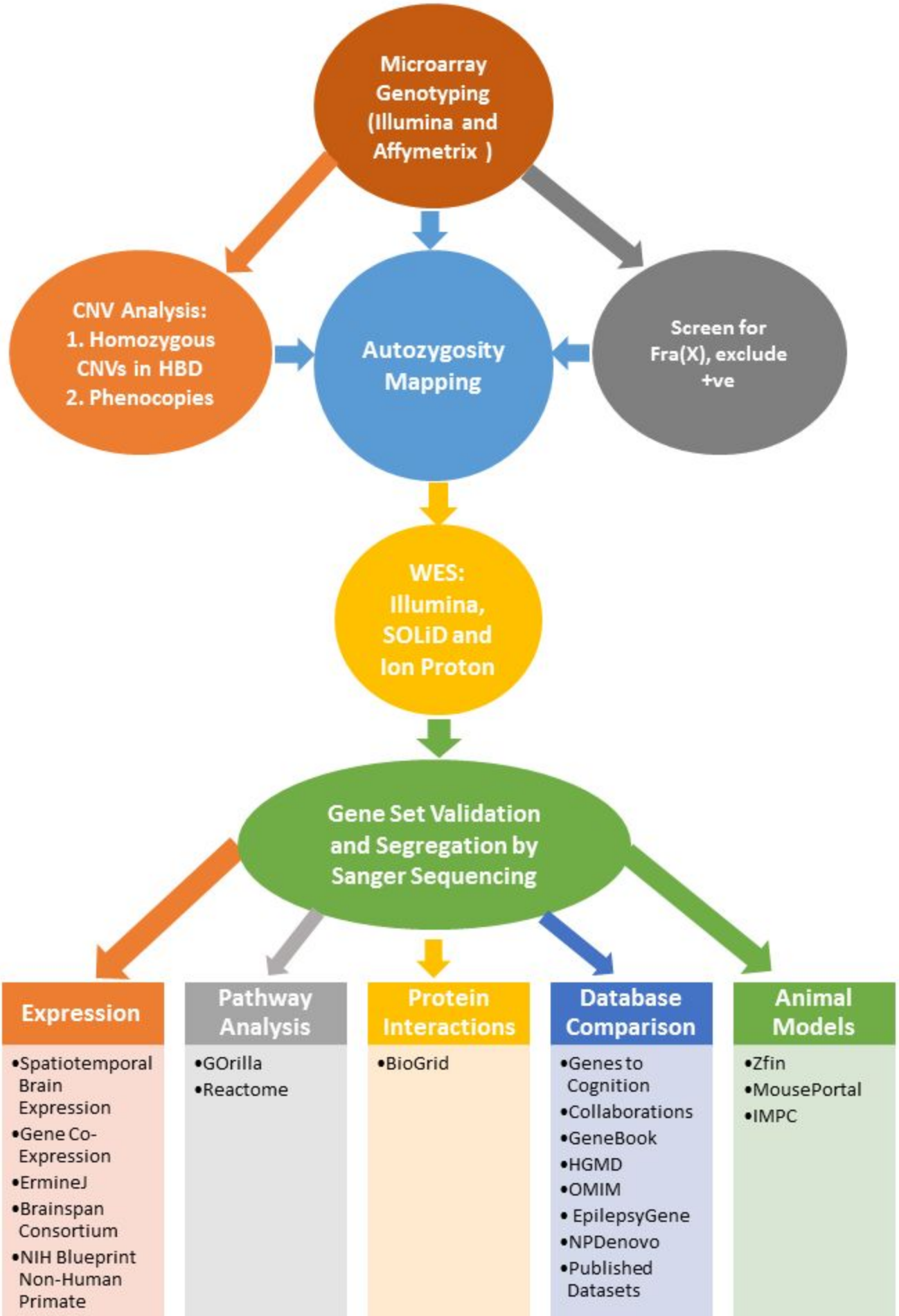
Supplementary Table 4: Pathogenic CNVs and variants of unknown significance identified by microarray analysis.

Supplementary Table 5: BioGRID protein interaction analysis.

Supplementary Table 6: Gene Ontology Pathway analysis

Supplementary Table 7: Gene List for anatomic/temporal transcription analyses.

Supplementary Table 8: Top anatomical regions for ID gene expression.


Microarray Genotyping (Illumina and Affymetrix)
CNV Analysis: 1. Homozygous CNVs in HBD 2. Phenocopies
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Gene Set Validation and Segregation by Sanger Sequencing
Expression
•Spatiotemporal Brain Expression
•Gene Co-Expression
•ErmineJ
•Brainspan Consortium
•NIH Blueprint Non-Human Primate
Pathway Analysis
•GOrilla
•Reactome
Protein Interactions
•BioGrid
Database Comparison
•Genes to Cognition
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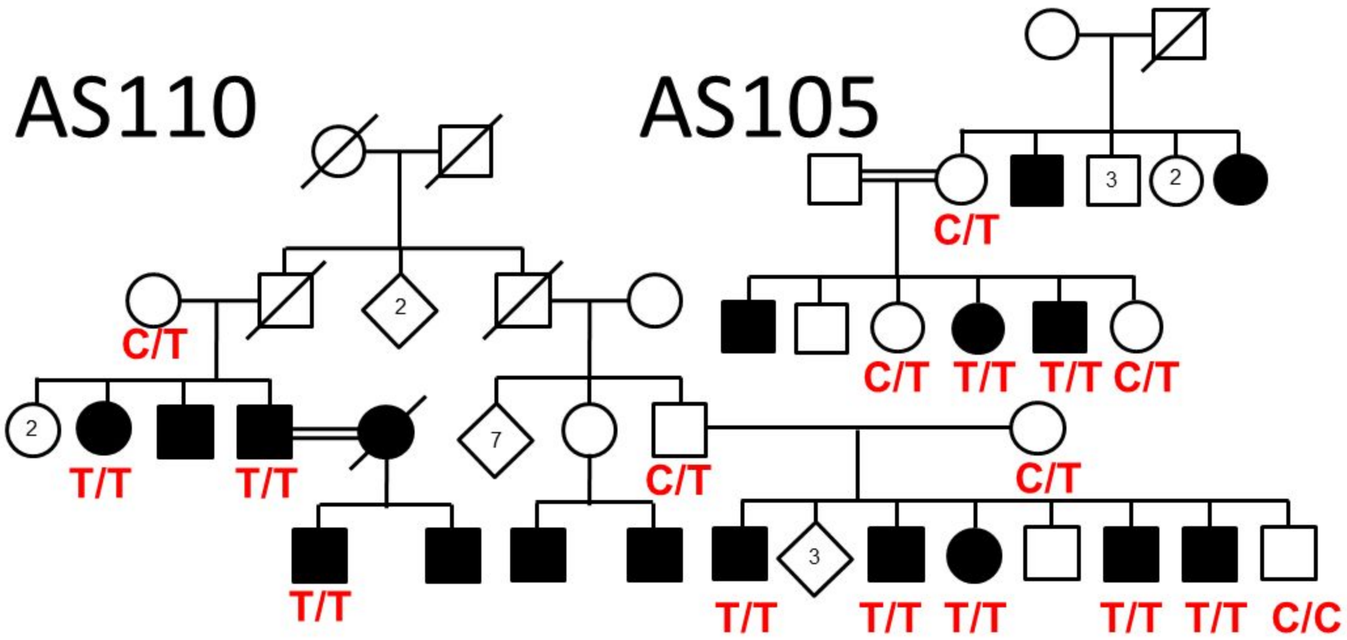

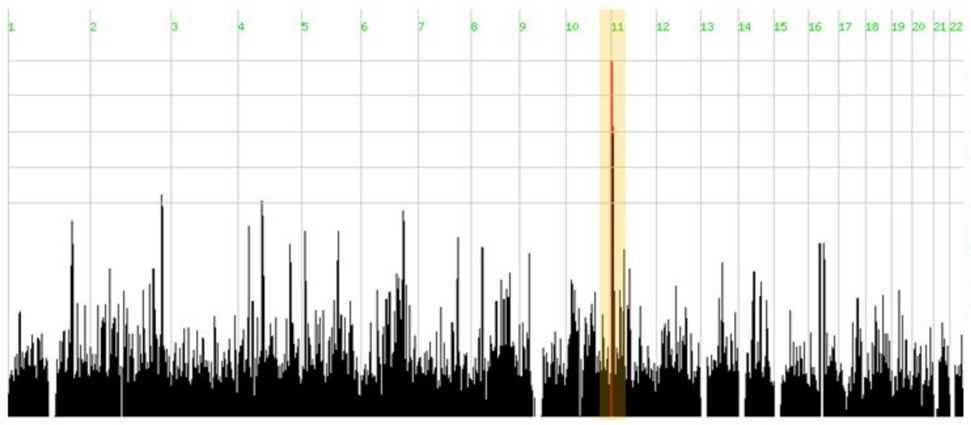

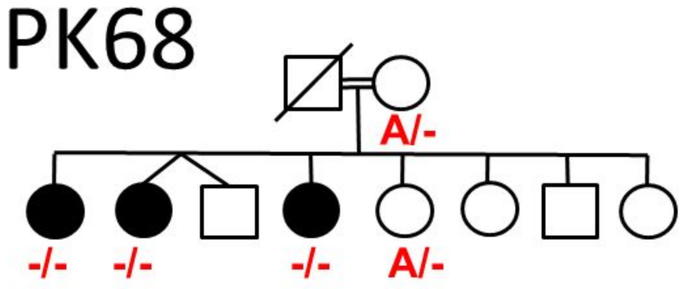

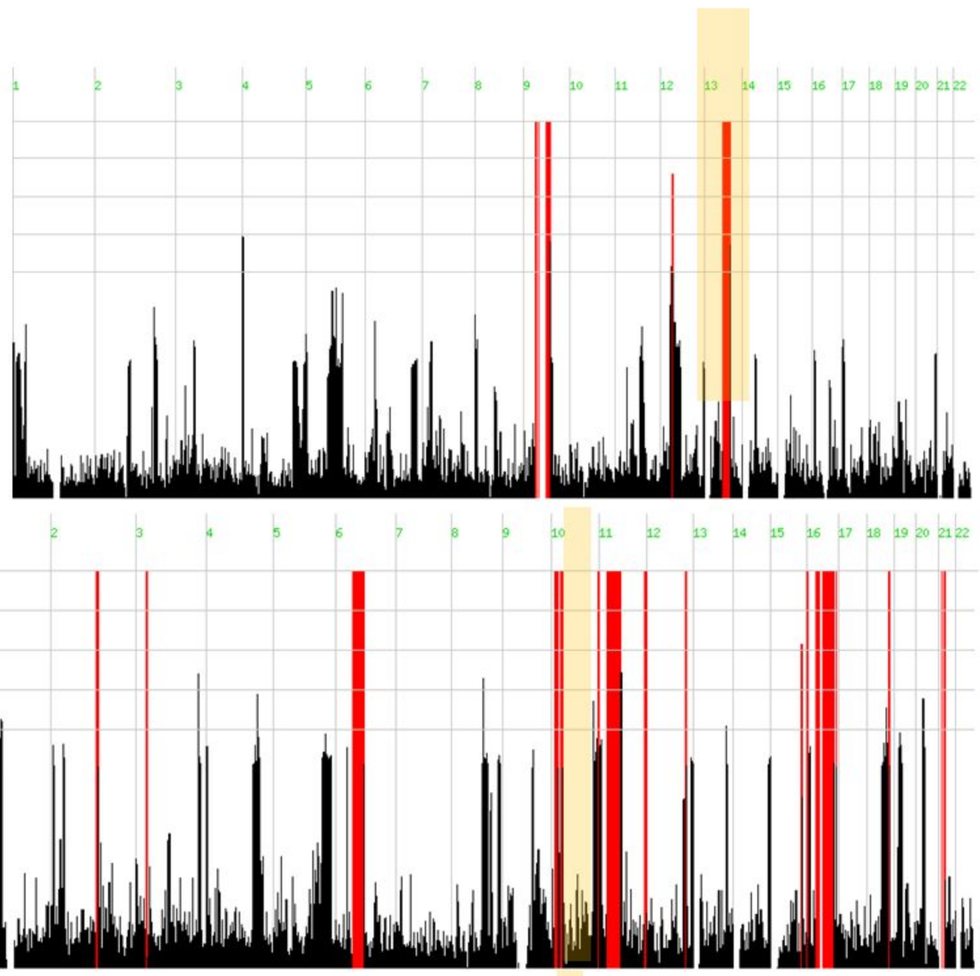

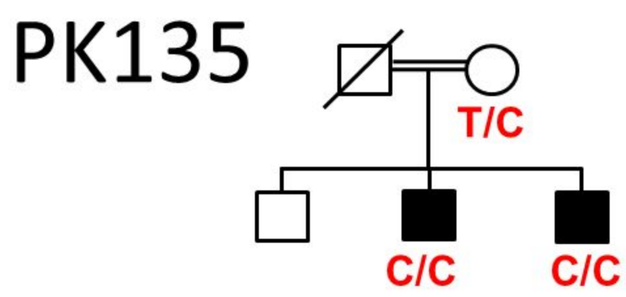

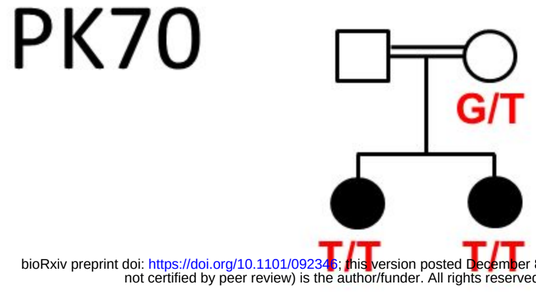

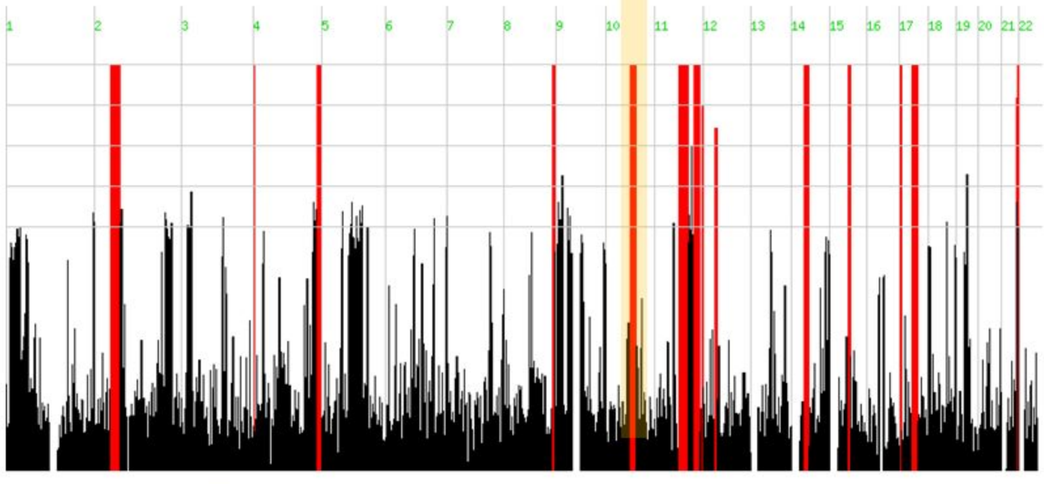

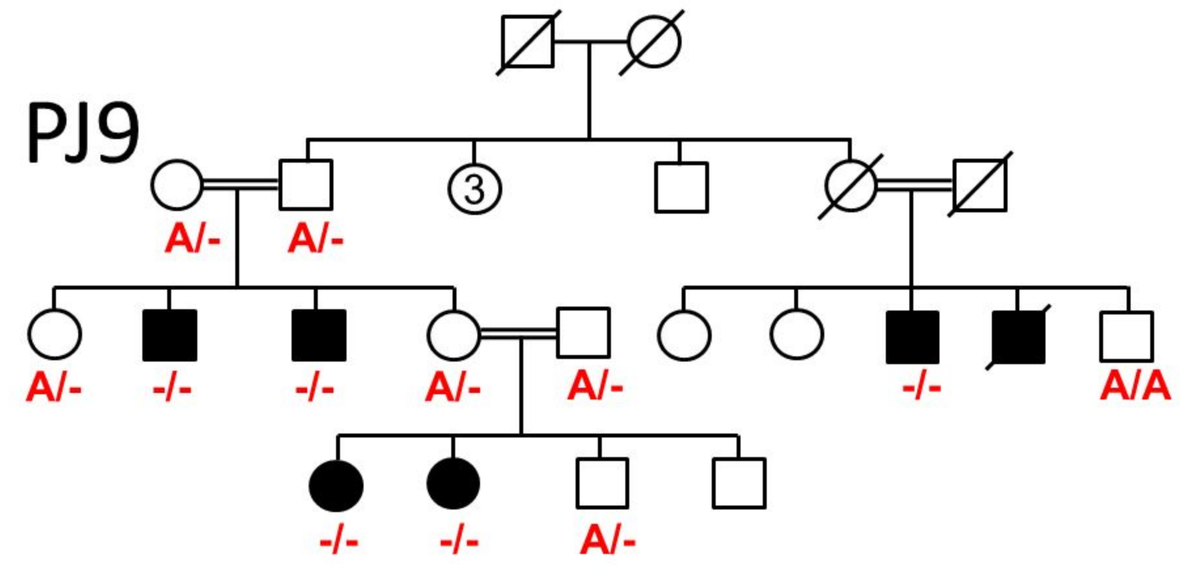

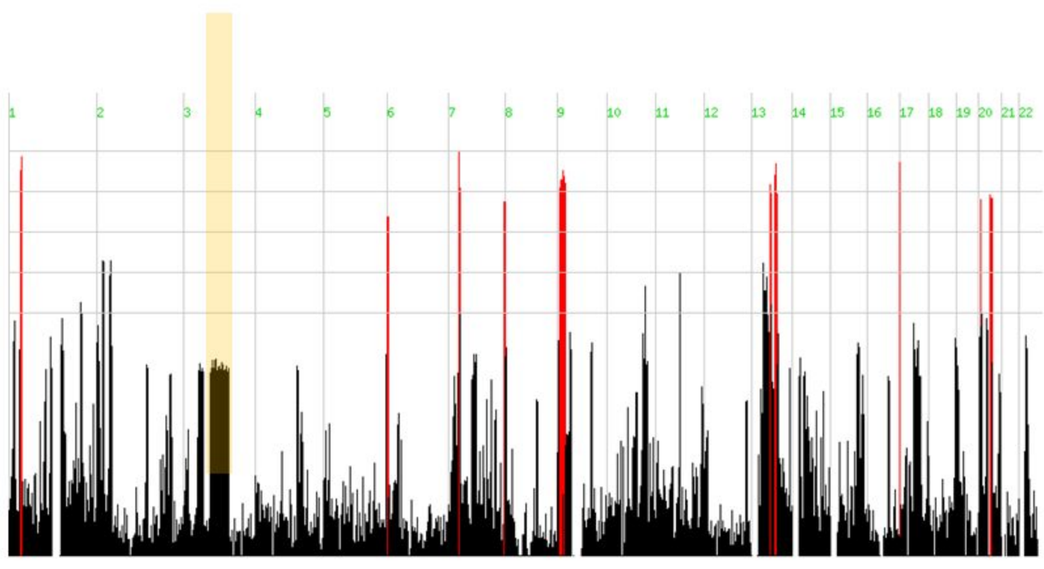

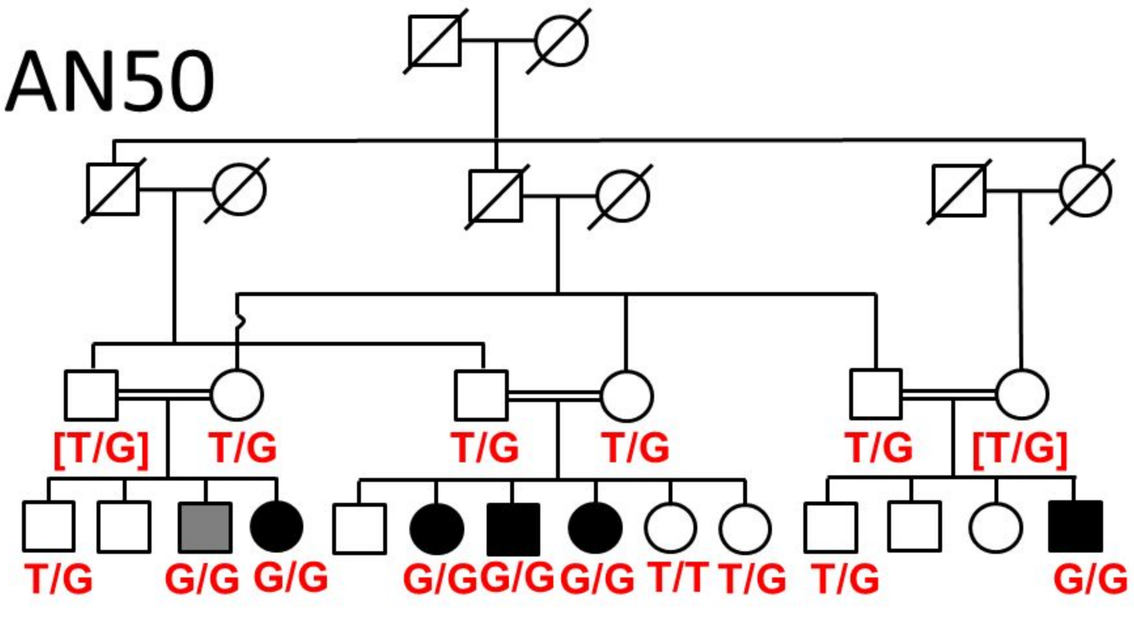

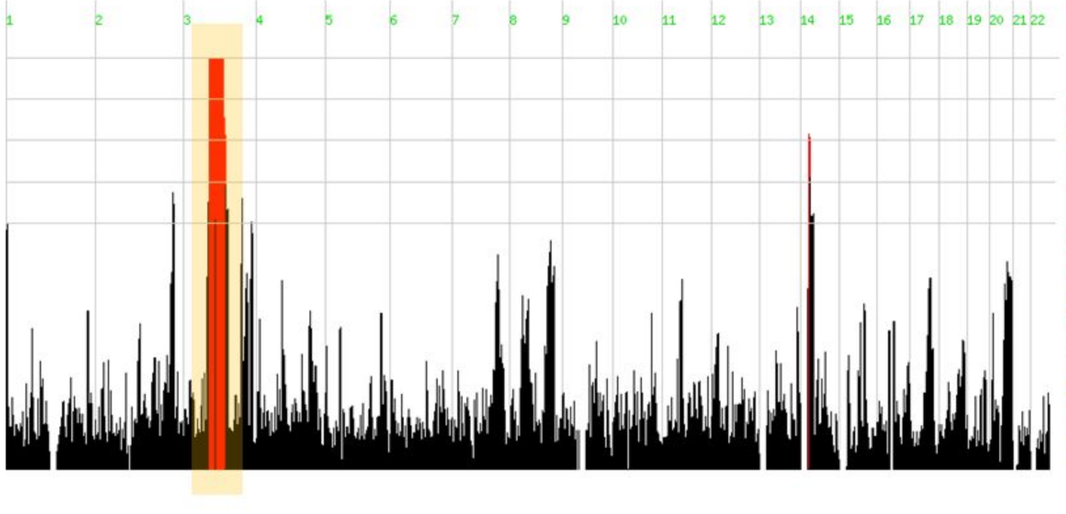

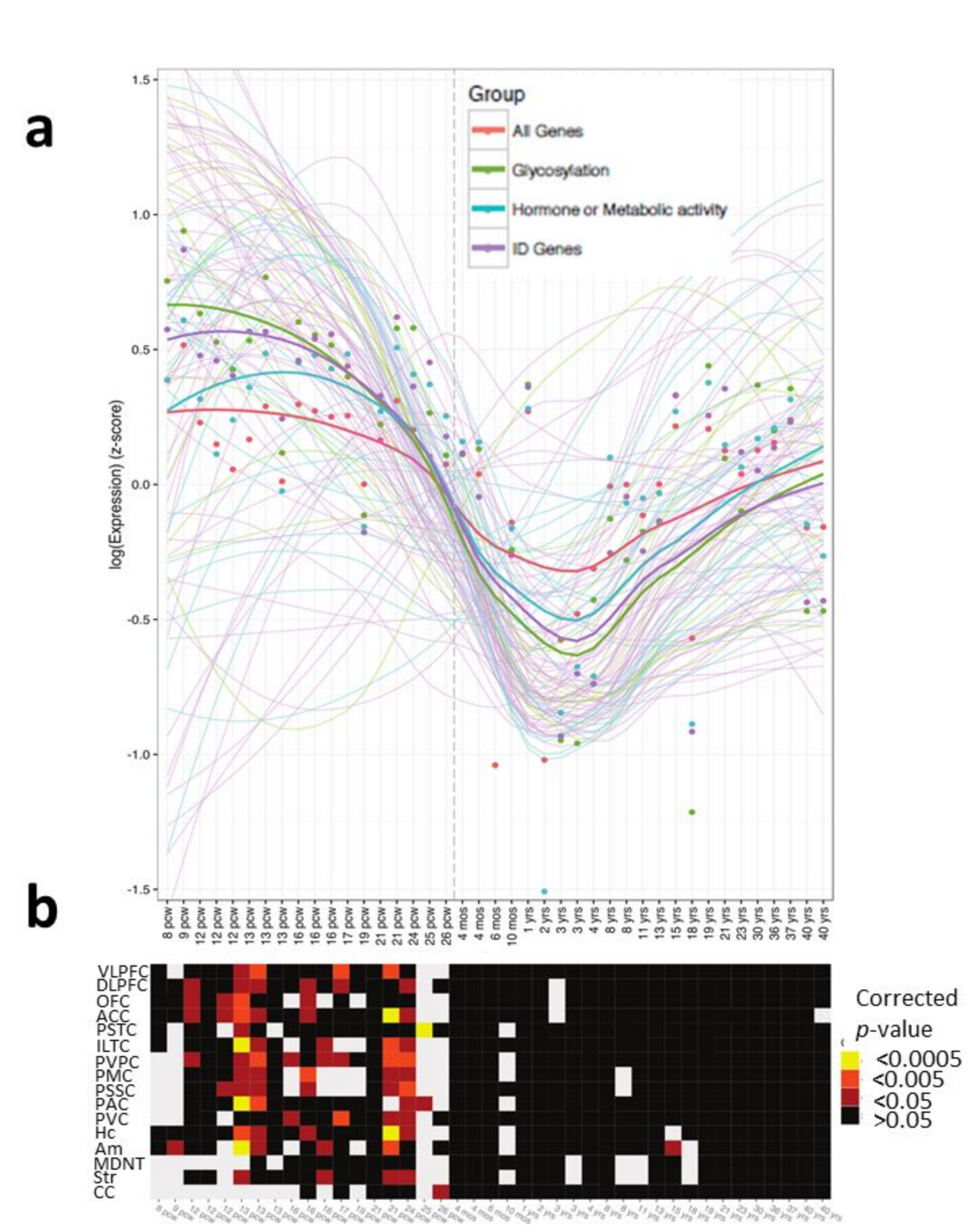
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Hormone or Metabolic activity
ID Genes
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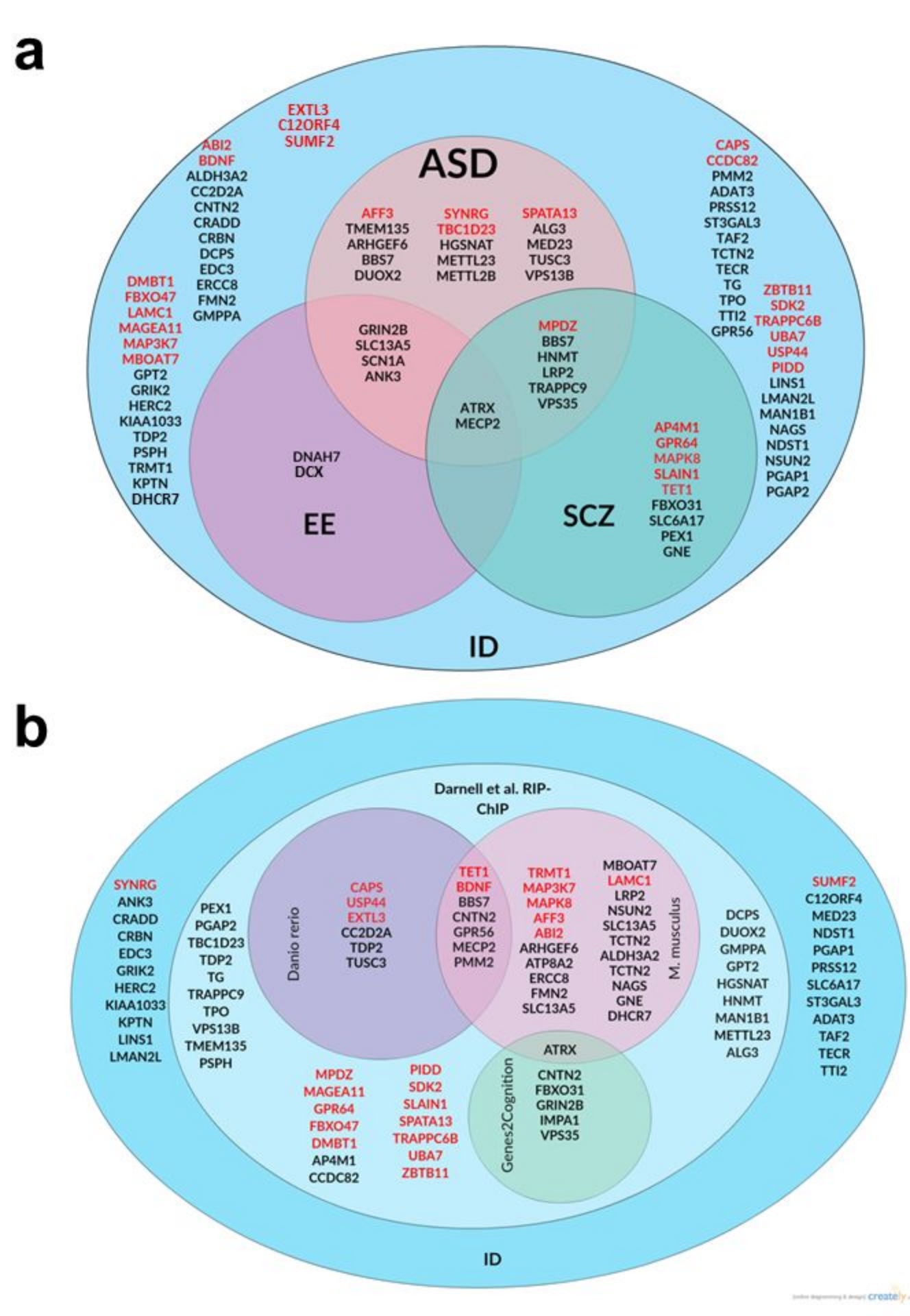
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