

Modelling the impact of curtailing antibiotic usage in food animals on antibiotic resistance in humans

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Abstract

Consumption of antibiotics in food animals is increasing worldwide and is approaching, if not already surpassing, the volume consumed by humans. It is often suggested that reducing the volume of antibiotics consumed by food animals could have a public health benefits. Although this notion is widely regarded as intuitively obvious there is a lack of robust, quantitative evidence to either support or contradict the suggestion.

As a first step towards addressing this knowledge gap, we develop a simple mathematical model for exploring the relationship between antibiotic consumption by food animals and levels of resistant bacterial infections in humans. We use the model to investigate the impact of restricting antibiotic consumption by animals and to identify which model parameters most strongly determine that impact.

Our results suggest that for a wide range of scenarios curtailing the volume of antibiotics consumed by food animals has, as a stand-alone measure, little impact on the level of resistance in humans. We also find that reducing the rate of transmission of resistance from animals to humans may often be more effective than an equivalent reduction in the consumption of antibiotics in food animals. Moreover, the response to any intervention is strongly determined by the rate of transmission from humans to animals, an aspect which is rarely considered.

Keywords: agriculture; food animals; antibiotic usage; antimicrobial resistance; mathematical model; one health

Introduction

Heightened concern about increasing levels of antimicrobial resistance worldwide has led to renewed calls to reduce substantially the use of antibiotics in food animals [1]. The notion that such a reduction could have public health benefits arises primarily from the observation that the volume of antibiotics consumed by food animals worldwide is approaching, and may have already overtaken, the volume consumed by humans [2]. This situation is expected to worsen as the transition to intensive animal production systems continues in many regions, especially China, India and other Asian countries [3, 4].

Antibiotic consumption by food animals occurs for the purposes of herd health, prophylaxis and growth promotion. Growth promotion, often involving sub-therapeutic doses, is particularly controversial. It has been banned in European Union (EU) countries since 2005 and is the subject of a more recent voluntary ban in the USA. Currently, 51% of the OIE member countries have banned antimicrobials as growth promoters, and a further 19% have a partial ban [5, 6]. The agricultural industry has adapted to these measures in ways such that there have been only modest net impacts on net consumption of antibiotics by food animals and levels of antibiotic resistance therein, such that any consequent benefits to human health are not easily discerned [7].

A key challenge for understanding the expected impact of reducing drug usage is that the relationship between antibiotic consumption by food animals and levels of resistant bacteria in humans is complex. First, food animals are far from the only source of human exposure to antibiotic resistant bacteria: high levels of antibiotic use in hospitals, clinics and the general population are also major drivers of resistance in humans [8]. Quantifying the specific contribution of the food animal route is not straightforward and has yet to be attempted. Second, there will be many different answers to the quantification question: there are numerous combinations of different antibiotics, bacterial strains and farm animal species, each with their own dynamics, and these are likely to vary between different countries with different health care systems and agricultural production systems.

As a first step towards addressing this knowledge gap, here we develop a simple mathematical model for exploring the relationship between antibiotic consumption by food animals and levels of resistant bacteria in humans. Our objective is to better understand the dynamics of AMR moving between food animal and human populations and to identify which model parameters have the greatest influence on levels of resistance in humans and for which parameter combinations we expect to see the greatest impact of reducing antibiotic consumption by food animals.

Methods

Mathematical model

Our mathematical model is intended to be as simple as possible while capturing the non-linearities inherent in infectious agents spreading between two host populations. To achieve this we consider two variables: R_H , the fraction of humans with antibiotic resistant bacteria (so a measure of the ‘level of resistance’); and R_A , the fraction of food animals with antibiotic resistant bacteria. The dynamics of R_H and R_A are given by the coupled ordinary differential equations:

$$R_H/dt = \Lambda_H(1 - R_H) + \beta_{HH}R_H(1 - R_H) + \beta_{AH}R_A(1 - R_H) - \mu_H R_H; \quad \text{Equation 1a}$$

$$R_A/dt = \Lambda_A(1 - R_A) + \beta_{AA}R_A(1 - R_A) + \beta_{HA}R_H(1 - R_A) - \mu_A R_A \quad \text{Equation 1b}$$

where: Λ_H is the per capita rate at which humans acquire antibiotic resistant bacteria through direct exposure to antibiotics and Λ_A is the equivalent in food animals; β_{HH} is the per capita rate at which humans acquire antibiotic resistant bacteria as a result of exposure (directly or indirectly via the wider environment) to other humans harbouring resistant bacteria and β_{AA} is the equivalent for animals; β_{AH} is the per capita rate at which humans acquire antibiotic resistant bacteria as a result of exposure (directly or, more frequently, indirectly via food products or the environment) to food animals harbouring resistant bacteria and β_{HA} is the reverse; μ_H is the per capita rate at which humans with resistant bacteria revert to having only susceptible bacteria (as a combination of clearance of resistance bacterial infections and demographic replacement) and μ_A is the equivalent in food animals. The time unit is arbitrary and does not affect the equilibrium values.

To obtain an equation for the equilibrium value, R_H^* , Equation 1 was set to 0 and solved for R_H in terms of the eight model parameters.

Although we do not consider the system to be at equilibrium, largely because antibiotic consumption patterns have changed considerably in recent decades (from zero before 1932) and continue to do so, we regard R_H^* as a useful indication of where the system is tending, and the approach to R_H^* will be relatively rapid if μ_H and μ_A are high, i.e. the mean duration of resistant infections is short ($\ll 1$ year).

As an indication of the potential impact of curtailing antibiotic usage in farm animals we set $\lambda_A = 0$ and define impact, ω , as the ratio of R_H^* with $\lambda_A = 0$ to R_H^* with $\lambda_A > 0$ (baseline $\lambda_A = 0.1$).

All analyses were carried out in Wolfram Mathematica, version 10.3 [9].

Scenarios

We consider two scenarios, a low impact scenario and a high impact scenario. The difference between these two scenarios is the baseline value chosen for β_{HA} . For the low impact scenario $\beta_{HA}=0.1$ and for the high impact scenario $\beta_{HA}=0.001$. These values were chosen to maximise the differences between the scenarios whilst minimising changes in the baseline levels of resistance between the two scenarios.

Sensitivity analysis

We determine which model parameters have most influence on the outcome value by computing the total sensitivity index D_{Ti} using the extension of Fourier amplitude sensitivity test (FAST) as described in Saltelli *et al.* [10]. The extended FAST method is a variance-based, global sensitivity analysis technique that has been largely used for studying complex agricultural, ecological and chemical systems (see [11, 12] for examples). Independently of any assumption about the model structure (such as linearity, monotonicity and additivity of the relationship between input factors and model output), the extended FAST method quantifies the sensitivity of the model output with respect to variations in each input parameter by means of spectral analysis. It provides measures of the amount of variance of the prevalence that arise from variations of a given parameter in what is called a total sensitivity index, D_{Ti} . It therefore captures the overall effect of parameter variations on equilibrium levels of resistance over a pre-specified range (i.e., including first- and higher-order interactions between model parameters). For example, a value of $D_{Ti}=0.10$ indicates that 10% of the total recorded variation of the prevalence is explained by the parameter under consideration. The sensitivity analysis was carried out using R [13]. For the sensitivity analysis we used a parameter range of 0.0 to 1.0 for all parameters under investigation.

Results

For the two scenarios considered here Figure 1 shows the trajectory of R_H and R_A in time. Figure 1 shows that the long term prevalence of resistance in the human population stabilises to a value of 0.71 for the low impact scenario and 0.70 for the high impact scenario. The long term prevalence of resistance in the animal population stabilises to a value of 0.71 for the low impact scenario and 0.62 for the high impact scenario. These differences between the scenarios are due to the lower value of β_{HA} in the high impact scenario (Table 1).

The sensitivity analysis on the equilibrium equation for R_H^* shows that this equilibrium is most sensitive to changes in μ_H , followed by λ_H , β_{AH} and β_{HH} , (Figure 2). Furthermore, the system is minimally sensitive to changes in “animal” parameters (β_{AA} , β_{HA} , λ_A and μ_A).

Figure 3 shows that, in accordance with the sensitivity analysis, but covering a wider range of parameter space (0.0 to 1.0), the equilibrium R_H^* is relatively insensitive to changes in λ_A , but is more sensitive to changes in β_{AH} , suggesting that reducing the former without addressing the latter may have limited impact on the prevalence of antibiotic resistance in the human population. Furthermore, the figure shows that the change in R_H^* due to β_{AH} is non-linear, which suggests that partial reductions of β_{AH} may only have limited impact when R_H^* is already high. Comparing panel A with panel B in Figure 3 shows that the effect of reducing the rate at which animals acquire antibiotic resistant bacteria as a result of exposure of animals to antibiotics (λ_A) is also strongly influenced by the per capita rate at which food animals acquire antibiotic resistant bacteria as a result of exposure to humans carrying resistant bacteria (β_{HA}). For the higher value of β_{HA} (low impact scenario, panel B) lowering λ_A by itself has little influence.

Figures 4 and 5 show the results for the impact of curtailing antibiotic use in food animals, quantified as the variable ω (defined above). Figure 4 shows the sensitivity of the impact to the different parameters. From Figure 4 it is clear that the impact is most sensitive to λ_H , the rate at which humans acquire antibiotic resistant bacteria as a result of exposure of humans to antibiotics, followed by μ_H , μ_A and β_{HA} . λ_A and β_{AH} have less influence. Figure 5 shows the effects of varying β_{AH} and λ_A on the

impact, ω , of curtailing antibiotic use in food animals. The difference between the low impact scenario and the high impact scenario is immediately clear from these graphs as there is virtually no impact of reducing λ_A in the low impact scenario (with relatively high per capita rate of transmission from humans to food animals, β_{HA}) while in the high impact scenario (with relatively low β_{HA}) there is a much more obvious benefit, though impact is still more sensitive to changes in β_{AH} over some regions of parameter space.

Discussion

It is often implied that the high levels of consumption of medically important antibiotics by food animals is contributing significantly towards the global public health problem of antibiotic resistance. Therefore we tested the (potential) impact of curtailing the use of antibiotics in food animals on the (long term) prevalence of humans carrying resistant bacteria using a mathematical model designed to capture the non-linearities inherent in the transmission of infectious agents between two populations as simply as possible. Our results show that, as expected, the system is particularly sensitive to changes in per capita rate at which humans acquire antibiotic resistant bacteria as a result of direct exposure to antibiotics (λ_H). Of much greater interest is the importance of the per capita rate of transmission of antimicrobial resistance from humans to animals (β_{HA}) (see Figures 4 and 5). For this reason, we compared two scenarios, a low impact scenario (high β_{HA}) and a high impact scenario (low β_{HA}). If β_{HA} is high (Figure 3, panel B; Figure 5, panel B) then the effects of reducing the rates at which animals acquire resistance as a result of antibiotic usage (λ_A) and humans acquire antibiotic resistant bacteria from animals (β_{AH}) are limited (Figure 3 and Figure 5, panel A, when β_{AH} or λ_A approaches 0). This contrasts with the situation where β_{HA} is low (Figure 3, Figure 5, panel A). This indicates that whenever the rate of transmission of antibiotic resistant bacteria from humans to animals is high it is more difficult to curb the antibiotic resistance problem, a rather counterintuitive result and often overlooked in discussion about this topic. Also of interest is that a failure to address the agricultural usage of antibiotics severely limits what we can achieve by tackling the problem from the human side, i.e. even if no resistance is required via direct exposure to antibiotics in humans (achieved by reducing λ_H to 0), we can only reduce the long term prevalence of antibiotic resistance in humans to 0.56 for the low impact scenario and 0.54 for the high impact scenario (see Figure S1 in the Supplementary information). In other words, if resistance dynamics in human and animal populations are coupled, as is generally thought to be the case in practice, substantial impacts on levels of resistance requires coordinated interventions across both populations.

Our study has several limitations that should be addressed. The first limitation is the simplicity of the model used. As indicated in the introduction, antibiotic resistance is a highly complex problem with

numerous routes of transmission and that will vary qualitatively and quantitatively for different drug-bug-animal combinations. Accounting for all these different routes and combinations separately is challenging. However, by taking the simplest possible mathematical model as a starting point, we are able to make a first step in trying to understand this highly complex system and gain some robust and useful insights into its behaviour. These findings can then be used as stepping stones for the development of more complicated (and perhaps more precise) models. Future models should include information on geographical distance and connections (i.e. networks) between and within populations and different modes of inheritance of resistance, and relate rates of gain of resistance to historical data on antibiotic consumption.

As with all models, parameterisation is an important issue. In this study we chose our baseline parameter values such that the long term prevalence of the fraction of the human population that is affected by resistant bacteria is roughly 70%. This mimics a situation where antibiotic resistance is well established but still leaves room for improvement or deterioration of the levels of resistance. Ideally the parameters would have been either measured directly in the population(s) or estimated from (good quality) datasets from experiments or from the real world. There is, however, a general lack of data on antibiotic resistance and antibiotic consumption, especially in the food animal population, making it notoriously difficult to obtain accurate point estimates and ranges for the parameters. Moreover, the many different combinations of antibiotic, bacteria strain, food animal species and setting will represent many different points in parameter space, each of which would need to be determined individually – a significant challenge.

To conclude, we have shown in this study that we can obtain useful insights into a highly complex problem like antibiotic resistance by using a simple mathematical model. Although it is widely regarded as intuitively obvious that reducing antibiotic consumption in animals would decrease levels of antibiotic resistance in humans this is, in fact, not the case for a wide range of scenarios (i.e. parameter space), especially if this intervention is made in isolation. Reducing the rate of transmission of resistance from animals to humans may often be more effective. In addition, the behaviour of the system, and so the response to any intervention, is strongly determined by the rate of transmission

from humans to animals, but this has received almost no attention in the literature. It is thus not enough to only lower the consumption of antibiotics in food animals, the transmission both from and to food animals should also be limited in order to maximise the impact of this and other interventions. We recommend that formal, quantitative analyses are needed to assess the expected benefits to human health of reducing antibiotic consumption by food animals. In some circumstances these benefits will be very small and other measures will be needed to reduce the public health burden of antibiotic resistance.

Acknowledgement

We thank colleagues from Edinburgh Infectious Diseases for their helpful discussions during the drafting of this manuscript.

Competing interests

We have no competing interests.

Authors' contributions

BvB participated in the design of the study, performed the analysis and drafted of the manuscript. MW conceived of the study, designed the study and helped draft the manuscript. All authors gave final approval for publication.

Funding

This study was financially supported by the *European Union's Horizon 2020 research and innovation programme* under grant agreement number 643476 (www.compare-europe.eu). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

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Tables

Table 1. Baseline parameters for the two scenarios.

Parameter	Description	Value used	
		Low impact	High impact
β_{AA}	per capita rate at which animals acquire antibiotic resistant bacteria as a result of exposure to other animals harbouring resistant bacteria	0.1	0.1
β_{HH}	per capita rate at which humans acquire antibiotic resistant bacteria as a result of exposure to other humans harbouring resistant bacteria	0.1	0.1
β_{AH}	per capita rate at which humans acquire antibiotic resistant bacteria as a result of exposure to food animals carrying resistant bacteria	0.1	0.1
β_{HA}	per capita rate at which food animals acquire antibiotic resistant bacteria as a result of exposure to humans carrying resistant bacteria	0.1	0.001
λ_H	per capita rate at which humans acquire antibiotic resistant bacteria as a result of direct exposure to antibiotics	0.1	0.1
λ_A	per capita rate at which food animals acquire antibiotic resistant bacteria as a result of direct exposure to antibiotics	0.1	0.1
μ_A	per capita rate at which humans with resistant bacteria revert to having only susceptible bacteria	0.1	0.1
μ_H	per capita rate at which food animals with resistant bacteria revert to having only susceptible bacteria	0.1	0.1

Figures

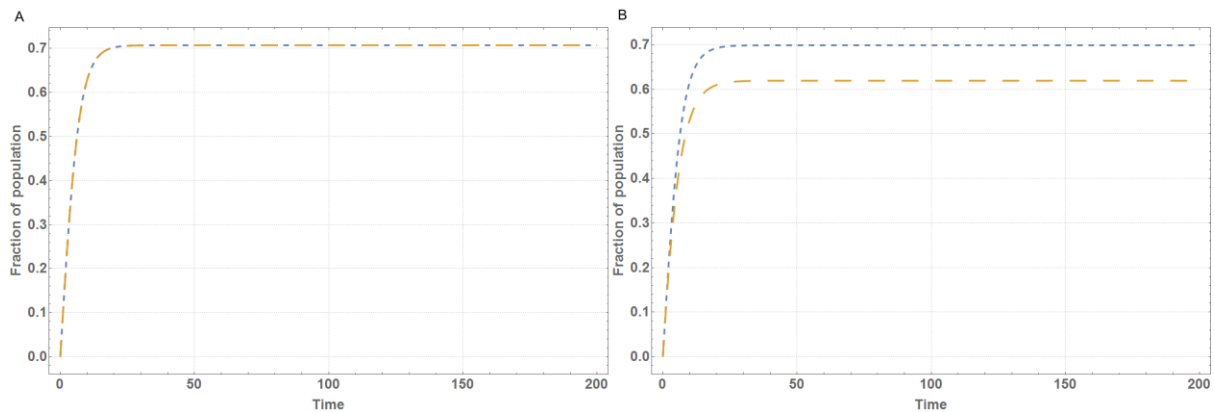


Figure 1. Trajectory of the fraction of the human population with antibiotic resistant bacteria (R_H) and the fraction of food animals with antibiotic resistant bacteria (R_A) in time for the low impact scenario (panel A) and the high impact scenario (panel B). Blue curves represent R_H , orange curves represent R_A .

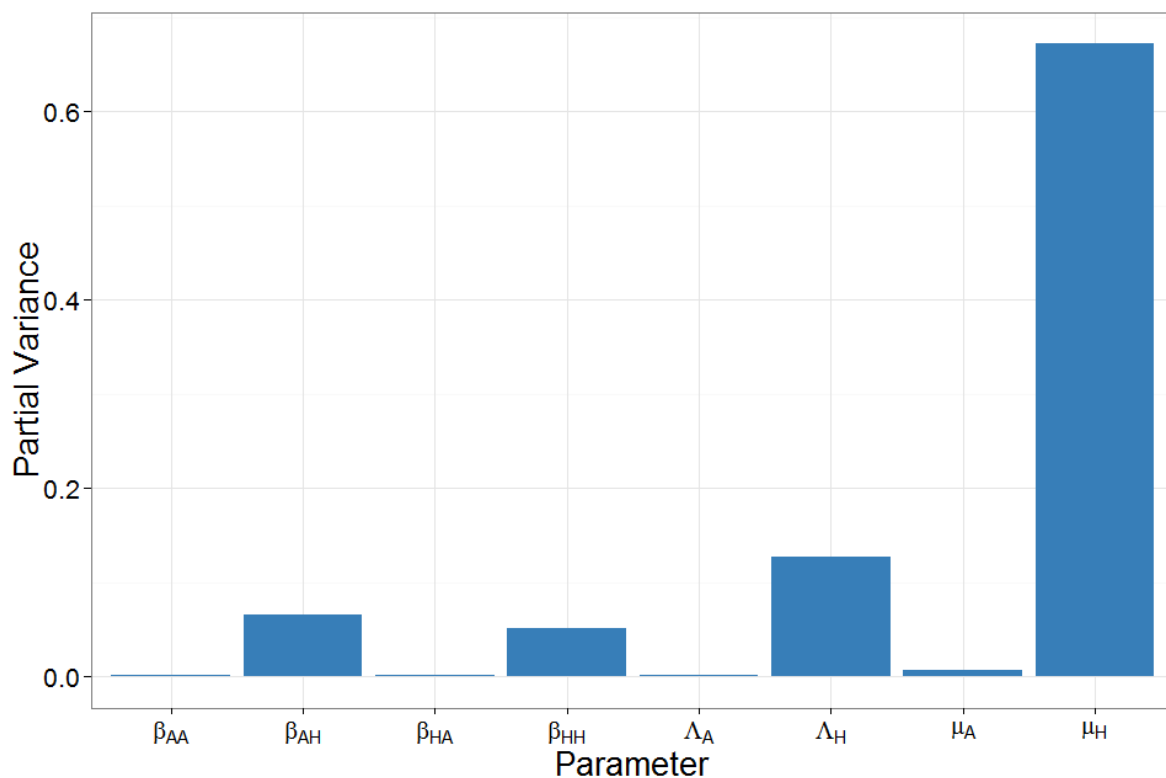


Figure 2. Result from a global sensitivity analysis on the equilibrium equation showing the partial variance of the individual model parameters. Higher bars indicate greater sensitivity of the model to that parameter. See Methods section for details about the sensitivity analysis and parameter ranges used.

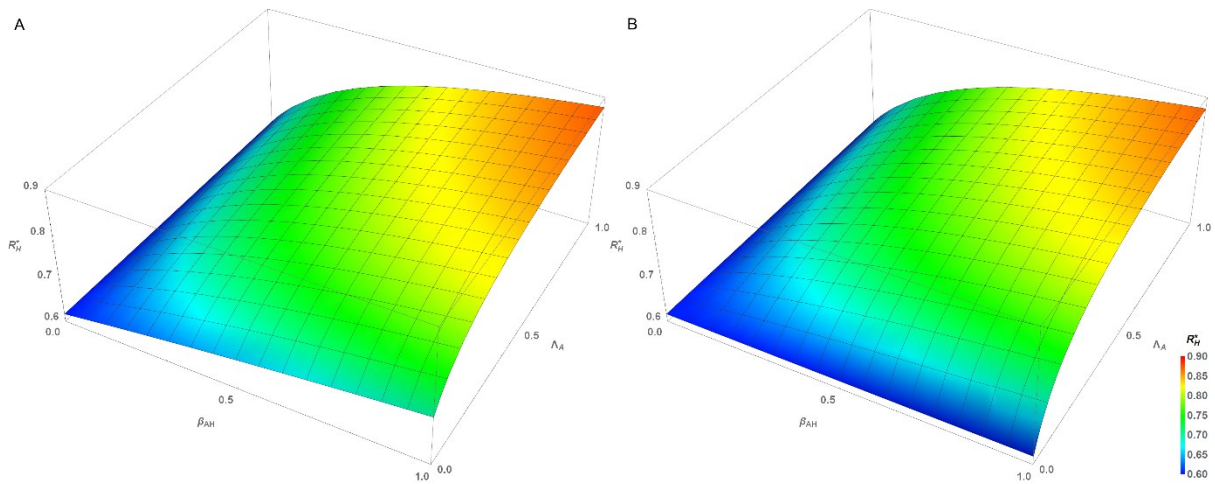


Figure 3. Surface of R_H^* as a function of β_{AH} and λ_A for the low impact scenario (panel A) and the high impact scenario (panel B).

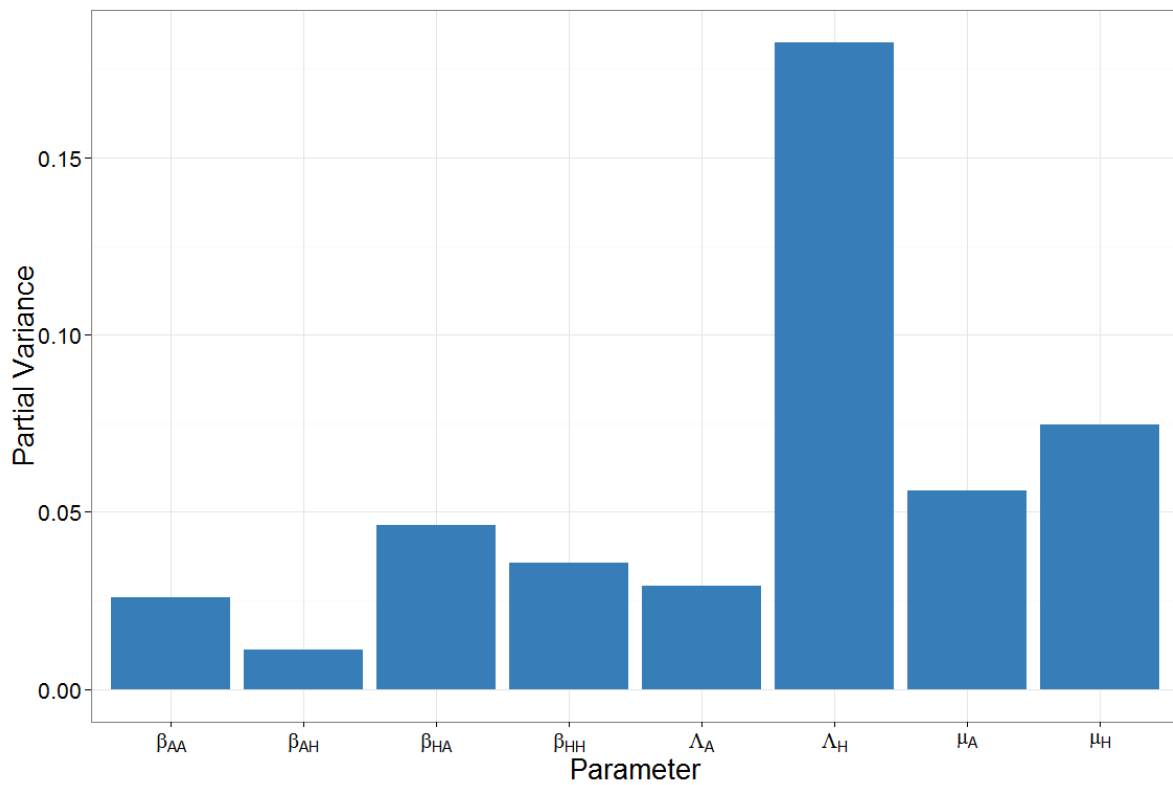


Figure 4. Result from a global sensitivity analysis on the impact (ω , defined in the Methods section) showing the partial variance of the individual model parameters. Higher bars indicate greater sensitivity of the model to that parameter. See Methods section for details about the sensitivity analysis and parameter ranges used.

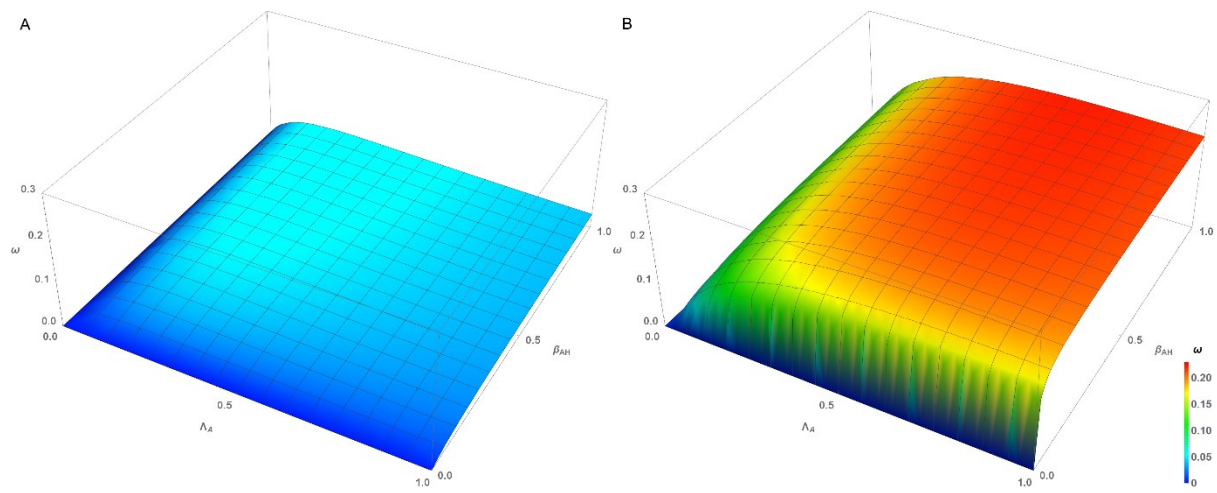


Figure 5. Surface of impact, ω , as a function of β_{AH} and λ_A for the low impact scenario (panel A) and the high impact scenario (panel B).