# 1 Colonization and diversification of aquatic insects on three Macaronesian

# 2 archipelagos using 59 nuclear loci derived from a draft genome

- 3 Sereina Rutschmann<sup>a,b,c,\*</sup>, Harald Detering<sup>a,b,c</sup>, Sabrina Simon<sup>d,e</sup>, David H. Funk<sup>f</sup>, Jean-Luc
- 4 Gattolliat<sup>g</sup>, Samantha J. Hughes<sup>h</sup>, Pedro M. Raposeiro<sup>i</sup>, Rob DeSalle<sup>4d</sup>, Michel Sartori<sup>g</sup>, and
- 5 Michael T. Monaghan<sup>a,b</sup>
- <sup>a</sup>Leibniz-Institute of Freshwater Ecology and Inland Fisheries (IGB), Müggelseedamm 301,
- 7 12587 Berlin, Germany
- 8 Berlin Center for Genomics in Biodiversity Research, Königin-Luise-Straße 6-8, 14195
- 9 Berlin, Germany
- <sup>c</sup>Department of Biochemistry, Genetics and Immunology, University of Vigo, 36310 Vigo,
- 11 Spain
- <sup>d</sup>Sackler Institute for Comparative Genomics, American Museum of Natural History, Central
- 13 Park West and 79<sup>th</sup> St., New York, NY 10024, USA
- <sup>e</sup>Biosystematics Group, Wageningen University, Droevendaalsesteeg 1, 6708 PB
- 15 Wageningen, The Netherlands
- 16 <sup>f</sup>Stroud Water Research Center, Avondale, Pennsylvania 19311, USA
- 17 <sup>g</sup>Musée cantonal de zoologie, Palais de Rumine, Place de la Riponne 6, 1014 Lausanne,
- 18 Switzerland
- 19 <sup>h</sup>Centro de Investigação e de Tecnologias Agro-Ambientais e Biológicas (CITAB),
- 20 Universidade de Trás-os-Montes e Alto Douro, Quinta de Prados, Apartado 1013, 5001-801
- 21 Vila Real, Portugal
- <sup>i</sup>Research Centre in Biodiversity and Genetic Resources (CIBIO)-Açores and the Biology
- 23 Department, University of Azores, Rua Mãe de Deus 13A, 9501-855 Ponta Delgada, Portugal
- \*Correspondence: Sereina Rutschmann, Phylogenomics Lab, Department of Biochemistry,
- 25 Genetics and Immunology, University of Vigo, 36310 Vigo, Spain, E-mail:
- sereina.rutschmann@gmail.com

## **Abstract**

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

The study of processes driving diversification requires a fully sampled and well resolved phylogeny. Multilocus approaches to the study of recent diversification provide a powerful means to study the evolutionary process, but their application remains restricted because multiple unlinked loci with suitable variation for phylogenetic or coalescent analysis are not available for most non-model taxa. Here we identify novel, putative single-copy nuclear DNA (nDNA) phylogenetic markers to study the colonization and diversification of an aquatic insect species complex, Cloeon dipterum L. 1761 (Ephemeroptera: Baetidae), in Macaronesia. Whole-genome sequencing data from one member of the species complex were used to identify 59 nDNA loci (32,213 base pairs), followed by Sanger sequencing of 29 individuals sampled from 13 islands of three Macaronesian archipelagos. Multispecies coalescent analyses established six putative species. Three island species formed a monophyletic clade, with one species occurring on the Azores, Europe and North America. Ancestral state reconstruction indicated at least two colonization events from the mainland (Canaries, Azores) and one within the archipelago (between Madeira and the Canaries). Using random subsets of the 59 loci showed a positive linear relationship between number of loci and node support. In contrast, node support in the multispecies coalescent tree was negatively correlated with mean number of phylogenetically informative sites per locus, suggesting a complex relationship between tree resolution and marker variability. Our approach highlights the value of combining coalescent-based phylogeography, species delimitation, and phylogenetic reconstruction to resolve recent diversification events in an archipelago species complex.

49 **Keywords:** Baetidae, island radiation, multispecies coalescent, phylogeny, phylogeography

## 1. Introduction

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

Any inference about the ecological and evolutionary processes driving diversification requires a well sampled and fully resolved phylogeny upon which traits can be mapped. Molecular phylogenetic studies historically have been limited to a small number of loci. The majority of studies are based largely on mitochondrial DNA (mtDNA) loci (Avise et al., 2000; Garrick et al., 2015) which have the benefit of small population size and high levels of polymorphism but suffer from several characteristics that can limit their suitability to reconstruct the evolutionary process. These include an inability to detect processes that confound gene trees and species trees such as hybridization and introgression, the inference of oversimplified or unresolved evolutionary relationships based on their matrilineal history, underestimated genetic diversity (Zhang and Hewitt 2003), and overestimation of divergence times (Zheng et al., 2011). Another major drawback is the presence of mtDNA genes that have been transposed to the nuclear genome, forming nuclear mitochondrial DNA (Numt; Lopez et al., 1994) which may appear homologous but give very different evolutionary signals from those of the real mtDNA. Phylogenetics has begin to benefit from more widespread use of single-copy nuclear DNA (nDNA) loci, and several recent studies have applied greater numbers of nDNA loci with success at the species (e.g. Ambystoma tigrinum (O'Neill et al., 2013); Triturus cristatus (Wielstra et al., 2014)), genus (e.g. Takydromus (Tseng et al., 2014); Heliconius (Kozak et al., 2015)), and higher taxonomic levels (e.g. Plethodontidae (Shen et al., 2016)). The phylogenetic resolution of closely related taxa enables crucial insights in studies of evolution. In particular, the investigation of recent or ongoing species radiations helps to explain how components such as adaptation and hybridization are involved in the diversification process (e.g. Monaghan et al., 2006; Morvan et al., 2013; Giarla and Esselstyn 2015; Toussaint et al., 2015). A number of model systems in evolutionary biology come from

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

93

94

95

96

97

98

99

closely related species groups that have diversified in island archipelagos (Schluter 2000; Gillespie and Roderick 2002, and references therein). Examples include Darwin's finches (Grant and Grant 2008), Anolis lizards (Losos and Ricklefs 2009), or Hawaiian spiders (Gillespie et al., 1994). While a robust phylogeny is needed to study diversification and adaptation in such groups, phylogenetic analysis of close relatives can be problematic. Discordance between gene trees and species trees is more likely when speciation is recent and the effective population size of the ancestral population is large relative to the age of the species (Kubatko and Degnan 2007; Degnan et al., 2012). This discordance can arise through hybridization, gene duplication and loss, and incomplete lineage sorting (Maddison 1997; Degnan and Rosenberg 2009; Knowles and Kubatko 2010; Nakhleh 2013). Increasing arrays of methods exist for examining multilocus data that account for these processes (Rannala and Yang 2003; Edwards 2009; Heled and Drummond 2010; Knowles and Kubatko 2010). Unfortunately, the appropriate data for these analyses can be lacking because it is difficult to generate sequence data for a sufficient number of suitable nDNA loci from non-model systems. Most nDNA loci exhibit low levels of polymorphism and therefore many loci are needed, whereas identification of novel nDNA loci that are suitable as phylogenetic markers is generally not straightforward. Here we use a whole-genome draft of a non-model species to develop nDNA markers suitable for phylogenetic reconstruction. Macaronesia consists of four archipelagos (Azores, Madeira, Canary Islands, and Cape Verde) whose flora and fauna have been used in several studies as model systems for evolutionary research. Their distances to the adjacent continental mainland vary from 110 km (Fuerteventura in Canary Islands to Morocco) to more than 2000 km (Flores in the Azores to Portugal). Several colonization pathways have been identified (Juan et al., 2000; Emerson 2002; Emerson and Kolm 2005), including a single colonization event followed by steppingstone dispersal (Juan et al., 1997; Emerson and Oromi 2005; Illera et al., 2007; Arnedo et al.,

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

2008; Dimitrov et al., 2008), or multiple independent colonization events within the Canary Islands (Nogales et al., 1998; Ribera et al., 2003a; Díaz-Pérez et al., 2012; Rutschmann et al., 2014; Gohli et al., 2015; Stervander et al., 2015; Faria et al., 2016). While much research has been carried out on island evolution and endemism of terrestrial organisms, comparatively limited information exists for aquatic invertebrates (e.g. Stauder 1995; Drotz 2003; Ribera et al., 2003b, 2003c; Jordal and Hewitt 2004; Hughes and Malmqvist 2005). This is a large discrepancy considering that aquatic insects contribute a disproportionally large amount of global biodiversity despite the relatively small extent of their habitat (Dijkstra et al., 2014). Mayflies are well suited for phylogeographic studies considering their ancient origins (300 million years (Ma)), global distribution, and limited dispersal ability due to the strict water habitat fidelity of larvae and very short life of the winged adults (Monaghan et al., 2005; Barber-James et al., 2008). Several studies have pointed out their unusual potential for dispersion, reporting mayfly species on remote islands such as the Azores (Brinck and Scherer 1961; Raposeiro et al., 2012), trans-oceanic dispersal between Madagascar and continental Africa (Monaghan et al., 2005; Vuataz et al., 2013), and recent colonization processes of several lineages on the Canary Islands and Madeira ≈ 14 Ma, including a close link to the African mainland (Rutschmann et al., 2014). The species complex of Cloeon dipterum L. 1761 is one of the most common and abundant species of freshwater insects in European standing water. The taxonomic classification and phylogenetic relationships within the C. dipterum s.l. species complex, including its complicated synonymy, remain largely unknown. The species complex belongs to the subgenus Cloeon Leach, 1815. In Europe, Cloeon consists of C. dipterum, two other currently recognized species (C. peregrinator Gattolliat and Sartori, 2008, and C. saharense Soldán and Thomas, 1983), and three species with unclear status (species inquirenda; C. cognatum Stephens, 1836, C. inscriptum Bengtsson, 1914, and C. rabaudi Verrier, 1949) that are often

considered to be synonyms of C. dipterum. Its distribution ranges from North America, across Europe to Northern Asia (excluding China), making it one of the largest known distributions among mayflies (Bauernfeind and Soldán 2012, and references therein). Larvae are found in a variety of aquatic habitats, including natural standing or slow-flowing waters, brackish water, intermittent watercourses, and artificial biotopes across a wide range of climatic zones (Bauernfeind and Soldán 2012, and references therein). For this study we used a draft genome sequence of Cloeon to develop 59 nDNA loci suitable for phylogenetic reconstruction of closely related members of the C. dipterum s.l. species complex of mayflies. We identified target genes and designed primer pairs for them. Standard PCR and Sanger sequencing were used to generate sequences. We then applied Bayesian phylogenetic inference using concatenated sequence alignments and multispeciescoalescent approaches to delineate species, examine their colonization from the mainland, and understand their diversification throughout Atlantic oceanic islands (Fig. 1, Azores, Madeira, and Canary Islands). Additionally, we quantitatively examined the effect of increasing numbers of nDNA loci on tree resolution. Our analyses show how marker development can proceed efficiently from draft whole genomes and that large numbers of nDNA loci can produce fully resolved trees in closely related taxa, revealing the evolution and diversification of the geographically widespread C. dipterum s.l. species complex. The disentangled colonization routes of the three species occurring on the Macaronesian Islands highlight transoceanic dispersal abilities of aquatic insects as an important driver of allopatric speciation, including sympatric occurring sister-species on the islands and the mainland.

## 2. Material and methods

125

126

127

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

### 2.1 Development of nuclear DNA loci

To develop a set of nuclear loci we sequenced a newly created whole-genome library of C. dipterum (see also Rutschmann et al., accepted). Libraries were generated from laboratoryreared subimagos of C. dipterum specimens (full siblings). DNA was extracted from pooled specimens (5-20) after removing eyes and wings using the Invisorb® Spin Tissue Mini kit (STRATEC, Berlin, Germany). Extracted DNA was precipitated using Isopropanol and pooled in order to obtain higher DNA yield. We prepared one 454 shotgun and one 454 paired-end library according to the manufacturer's guidelines (Rapid Library Preparation Method Manual, GS FLX+ Series - XL+, May 2011; Paired End Library Preparation Method Manual – 20 kb and 8 kb Span, GS FLX Titanium Series, October 2009). The fragments were amplified with an emulsion PCR (emPCR Method Manual - Lib-L SV, GS FLX Titanum Series, October 2009; Rev. Jan 2010). Four lanes per library were sequenced on a Roche (454) GS FLX machine). The sequence reads were trimmed and de novo assembled using NEWBLER v. 2.5.3 (454 Life Sciences Corporation) under the default settings for large datasets. We made two different assemblies, one with the reads from the shotgun library and one with the reads from both shotgun and paired-end libraries. The newly sequenced draft whole genome was combined with 4,197 expressed sequence tag (EST) sequences from Baetis sp. (GenBank Acc. no. FN198828-FN203024). Cloeon and Baetis belong to the Baetidae subfamilies Cloeoninae and Baetinae. Primer pairs were designed in the conserved regions of orthologous sequences from included taxa. The above analysis procedures have since been incorporated into the DISCOMARK pipeline for marker discovery and primer design (Rutschmann at al., accepted; see Supplementary File 2).

### 2.2 Taxon sampling and DNA extraction

148

149

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

We sampled individuals of the *C. dipterum* s.l. species complex from larval aquatic habitats at 38 sampling sites on 13 islands including the Azorean archipelago, the Canary Islands, Madeira (Fig. 1), and 32 sampling sites on the European and North American

mainland (Supplementary Tables 1 and 2). All samples were preserved in 99% ethanol in the field and stored at 4°C until analysis. DNA was extracted from 107 individuals using NucleoSpin® 96 tissue kits (Macherey-Nagel, Düren, Germany). Our analysis included multiple populations of all currently recognized taxa (based on both morphological and molecular data) on the islands (Brinck and Scherer 1961; Gattolliat et al., 2008; Rutschmann et al., 2014).

### 2.3 PCR amplification, sequence alignment, and sequence heterogeneity

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

We sequenced 60 loci for the study: the mtDNA barcoding gene (cox1) and 59 newly developed nDNA loci. The cox1 locus was amplified and sequenced using the procedure described by Rutschmann et al. (2014). Based on a general mixed Yule-coalescent (gmvc) model analysis (Fujisawa and Barraclough 2013) of cox1, we selected a representative set of 29 individuals for which we obtained nDNA sequences, using the 59 newly designed primer pairs (Supplementary Table 3). Nuclear loci were amplified using standard polymerase chain reaction (PCR) protocols with an annealing temperature of 55°C. The PCR products were custom purified and sequenced at Beckman Coulter Genomics (Essex, UK) or Macrogen (Amsterdam, The Netherlands). Forward and reverse sequences were assembled and edited using Generous R7 v.7.1.3 (Biomatters Ltd.). Length variation (i.e. heterozygous indels) was decoded using CODONCODE ALIGNER v.3.5.6 (CodonCode Corporation, Centerville MA, USA). Additionally, we included previously published sequences from four individuals of 6 nDNA loci (KU971838-KU971840, KU971851, KU971919-KU971921, KU971933, KU972490-KU972492, KU972503, KU972568-KU972570, KU972583, KU972653-KU972654, KU972666, KU973060-KU973062, KU973074). Multiple sequence alignments were made for each locus using MAFFT v.7.050b (L-INS-I algorithm with default settings; Katoh and Standley 2013). The predicted orthologous sequences of Baetis sp. were used to infer the correct exon-intron splicing boundaries

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

(canonical and non-canonical splice site pairs) of each alignment. Exon-intron boundaries of locus 411912 could not be fully reconstructed and thus we used the exon sequence predicted from tblastx searches for subsequent analyses. Locus alignments were split into coding and non-coding parts using custom script (https://github.com/srutschmann/python\_scripts/blob/master/extract\_introns.py). All coding alignments were checked for indels and stop codons using MESQUITE v.2.75 (Maddison and Maddison 2011). Genotypes of the coding alignments were phased using the probabilistic Bayesian algorithm implemented in PHASE v.2.1.1 (Stephens et al., 2001; Stephens and Donnelly 2003) with a cutoff value of 0.6 (Harrigan et al., 2008; Garrick et al., 2010). Multiple runs were performed for each alignment and phase calls checked for consistency. Input and output files were formatted using the scripts from SEQPHASE (Flot 2010). Heterozygous sites that could not be resolved were coded using ambiguity codes for subsequent sequence analyses. All alignments were re-aligned after phasing with MAFFT. We excluded introns for the haplotype phasing because the noncoding alignments contained many gaps and missing data and thus the results of the sequence phasing were not satisfactory. For the subsequent analyses, we prepared three alignment sets (Table 1), whereby we used all nDNA sequences (all data), all coding genotypes (exon all data) and all coding haplotypes (exonhap all data). Because data matrices were not 100% complete (see 3.1. Development of nuclear DNA loci), we made a second set of matrices that were 100% complete using only the 17 loci that were sequenced for all 29 individuals (complete matrix, exon complete matrix, exonhap complete matrix). The most appropriate substitution model for each locus was determined according to Bayesian Information Criterion in the program JMODELTEST v.2.1 (Guindon and Gascuel 2003; Darriba et al., 2012) (Supplementary Table 3).

To investigate the heterogeneity among the newly developed loci, we reconstructed reticulation-free haplotype genealogies based on Fitch distances (Fitch 1970), using the program FITCHI (Matschiner 2015). We used the exonhap\_all\_data matrix and calculated a gene tree for each locus using RAXML v.8 (Stamatakis 2014) under the GTRCAT model with 1,000 bootstrap replicates using the rapid bootstrap algorithm. The number of variable sites, informative sites, and Tajima's D for each locus was assessed using the package DENDROPY (Sukumaran and Holder 2010; <a href="https://github.com/srutschmann/python\_scripts/blob/master/alignment\_stats.py">https://github.com/srutschmann/python\_scripts/blob/master/alignment\_stats.py</a>) and a custom script.

# 2.4 Species assignment and population structure analysis

Most analyses that use phylogenetic or multilocus species tree approaches require *a priori* species assignment. Because of the partly unknown and largely incomplete taxonomy of the group, we used two approaches to first assign the 29 *C. dipterum* individuals to putative species: the gmyc approach (Fujisawa and Barraclough 2013) and a Bayesian clustering algorithm to assign individuals to 'populations' (STRUCTURE, Pritchard et al., 2000; Falush et al., 2003). The gmyc approach was carried out using *cox1* from 147 specimens that included all newly sequenced *Cloeon* individuals, published sequences that were available as of February 2016 (Supplementary Table 2), six newly sequenced individuals of *C. simile* Easton, 1870, and *Baetis rhodani* (GenBank Acc. no. KF438126) as an outgroup. The analysis followed that of Rutschmann et al., (2014) except that we used BEAST v.2.3.2 (Bouckaert et al., 2014) and a 2-partition scheme in which the first two codon positions were modeled with HKY + I and the third codon position with HKY +  $\Gamma$ . For the Bayesian clustering approach we used the exon\_all\_data matrix. We assumed 1-10 genotypic clusters (K) and ran nine replicate analyses for each K, using 1 × 10<sup>6</sup> MCMC generations with a burn-

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

267

268

269

270

in of 10%. All individuals were assigned probabilistically without a priori knowledge to genetic clusters. We applied an admixture model with default settings (Supplementary File 3). 2.5 Phylogenetic reconstruction We performed Bayesian phylogenetic reconstructions using all data (exon all data) and the complete matrix (exon complete matrix) using MRBAYES v.3.2.2 (Ronquist et al., 2012). As outgroup we used *Baetis* sp.. All individual locus alignments were concatenated using a Python custom script (https://github.com/srutschmann/python\_scripts/blob/master/fasta\_concat.py). For the tree reconstruction, we implemented the best-fit models for each locus, and unlinked the nucleotide frequencies, gamma distributions, substitution rates and the proportion of invariant sites across partitions. Two independent analyses of four MCMC chains, each with  $1 \times 10^7$ generations and 25% burn-in were run. To investigate how the number of loci analyzed affected node support values, we performed phylogenetic reconstructions based on concatenated sets of varying numbers of randomly selected loci (Supplementary Table 4). The analyses were performed and summarized as above. Linear regressions were used to predict the number of supported nodes for Bayesian posterior probability (PP)  $\geq$  0.95 and PP = 1 as a function of the number of loci used in the analysis. The Pearson correlation between the number of loci and number of supported nodes was calculated separately for both PP, using the stats package in R (R Development Core Team, 2016). Species tree reconstructions were carried out under a multispecies coalescent framework (Drummond and Rambaut 2007; Heled and Drummond 2010) as implemented in the program \*BEAST v.2.1.3 (Bouckaert et al., 2014). All analyses were performed using exons, one analysis using all data and one using only the complete matrix, as above (Table 1;

exonhap\_all\_data, exonhap\_complete\_matrix). All individuals were *a priori* assigned to species based on the gmyc and Bayesian clustering analyses described above. In the Bayesian clustering analysis, one Russian individual was considered to be admixed based on PP assignment values > 0.05 for more than one cluster (Supplementary File 3, Supplementary Fig. 1, Supplementary Table 5). This individual was therefore excluded from further analysis. We used a relaxed uncorrelated lognormal clock for gene tree estimation at each locus and a Yule speciation-process prior. We conducted six independent runs of 8 × 10<sup>8</sup> million generations each. Runs were combined in LOGCOMBINER v.2.1.3 (Bouckaert et al., 2014), whereby all parameters reached effective sample sizes (ESS) > 600. Maximum clade credibility trees for each species trees were obtained using TREEANNOTATOR v.2.1.3 (Bouckaert et al., 2014). As for the concatenated phylogenetics (above), we examined how the number of loci included in the multilocus species tree analysis affected node support by re-running the analysis using subsets of differing numbers of randomly selected loci (Supplementary Table 4).

#### 2.5 Ancestral state reconstruction

An ancestral state reconstruction approach was used to test the direction of the radiation (i.e. Continental to Island or Island to Continental). Ancestral range patterns of each individual were defined into four geographic areas: (1) a broadly defined Continental referring to the European and North American mainland, (2) Canary Islands, (3) Madeira, and (4) Azores. As input tree, we used the concatenated tree based on the exon\_all\_data inferred with MRBAYES. A chronogram was fit to the tree using the chronos function in the ape v.3.4 (Paradis et al., 2004) package in R. Ancestral states were estimated under an equal-rates (ER) model using the function ace, and the scaled likelihoods of each ancestral state were calculated using the function lik.anc in ape v.3.4. A MCMC approach was used to sample character histories from their PP distribution generating 1,000 stochastic character maps with

the function make.simmap of the phytools v.0.4.98 (Revell 2012) package in R (Supplementary File 4).

# 3. Results

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

### 3.1 Development of nuclear DNA loci

Whole-genome sequencing resulted in 1,109,684 raw reads, including 651,306 reads for the shotgun library and 458,378 reads for the paired-end library, with an average large contig length of 1,187 and 736 bp, respectively (BioSample SAMN03202660, BioProject PRJNA268073, Sequence Read Archive SRP050093). All reads were assembled into 68,473 contigs with an N50 of 1,116 bp. The reads of the shotgun library were assembled into 31,827 contigs with an N50 of 1,260 bp. We detected 918 putative orthologous gene sequences for C. dipterum from the contigs derived from the shotgun library, 1,298 putative orthologous gene sequences from the contigs of the combined assembly, and 416 for *Baetis* sp. (Supplementary Table 6). We successfully designed primer pairs for 59 sequence alignments (Supplementary Table 3), mostly based on orthologous sequences from both taxa. Total fragment length per sequenced locus ranged from 210 - 1,007 bp with a mean of 545 bp. Exon sequence length ranged from 210 - 710 bp with a mean of 410 bp (Supplementary Table 3) (KF438124-KF438125, KU757080-KU757184, and KU971616-KU973191). The full data matrix of all 29 individuals and all 59 loci including exons and introns (all data) was 32,213 bp in length when concatenated and when introns were removed (exon all data) it was 24,168 bp (Table 1). All individuals were successfully sequenced for at least 44 loci, and the above matrices were >75% complete. The 100% complete matrix included 17 loci that were sequenced successfully for all 29 individuals. All heterozygous indels were located in the intron sequences. However, 100 heterozygous sites could not been resolved and remained in the exonhap alignments.

All haplotype genealogies showed clear structuring (Supplementary Fig. 2). For 33 loci, we found haplotypes shared between putative species. The number of variable sites per locus ranged from six to 65 (mean: 18.95). These values were lower than those reported above because ambiguous sites were not considered variable in the haplotype analysis. In the exon\_all\_data matrix, there was one SNP per every 21.62 nucleotides sequenced (i.e. total length per total number of variable sites). The loci included between six and 54 informative sites (mean: 16) and one to 26 ambiguous sites (mean: 8.4). Nucleotide diversity ranged from 0.007 to 0.04 (mean: 0.017), and Tajima's D varied between -0.85 and 1.97 (mean: 0.29) (Supplementary Table 7).

# 3.2 Species assignment and population structure

There were 62 unique coxI haplotypes of Cloeon and the gmyc model was a significantly better fit to the data than the null ( $\chi^2 = 31.00$ , p < 0.001). There were seven putative species delineated within C. dipterum s.l. (Fig. 2a): One occurred only in Asia (South Korea) while the remaining six included three species with distributions that included the Macaronesian Islands (IS1- IS3) and three species only occurring on the European and North American continents (CT1- CT3). The population assignments from the Bayesian clustering analyses of nDNA (Supplementary File 3, Supplementary Fig. 1, and Supplementary Table 5) agreed completely with the results from the gmyc analysis. Among these six, one widespread species (IS1) was found on all Azorean islands, in Greece and Italy, and in North America, one on the Canary Islands and Madeira (IS2), and one only on four of the Canary Islands (IS3). The model recognized all seven C. dipterum gmyc species even when using the most conservative estimate (95% CI based on two log likelihood units: 16-19 gmyc species). The two C. cognatum specimens from the North American DNA barcoding project (Webb et al., 2012) had coxI haplotypes identical to our gmyc species IS1.

#### 3.3 Phylogenetic reconstruction

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

366

367

Analyses based on both exon matrices (exon all data; exon complete matrix) recovered the same tree topology with strong node support, resolving each of the three species occurring on Macaronesia (IS1-IS3) as monophyletic and members of a monophyletic 'Island clade' (Fig. 3a). The geographically widespread species IS1 was sister taxon to the two others. Species CT2 and CT3 were both monophyletic and sister group to the Island clade (Fig. 3a). All individuals in CT1 were monophyletic except for a single individual that was sister taxon to the entire C. dipterum s.l. lineage. There were 27 resolved (PP  $\geq 0.95$ ) nodes in the full matrix (all data) tree; the only unresolved node was between the two Azorean individuals (Fig. 3a). In contrast, the complete matrix tree contained only 19 resolved nodes, with lack of resolution most pronounced in IS2 (Supplementary Fig. 3). All species tree phylogenies had identical topologies and these matched the Bayesian phylogenies (Figs. 2b, 3a) in that Island and Continental clade both were monophyletic, with IS1 sister taxon to IS2 + IS3, and with CT1 sister taxon to CT2 + CT3. Using the exon complete matrix, all nodes were highly supported (PP  $\geq$  0.99; Table 2). All individuals clustered into six species in the same way in both the multilocus nDNA tree and the singlelocus (cox 1) mtDNA tree (PP = 1), but the relationships among the species were different. The mtDNA tree did not support the sister relationship of IS2 + IS3 or the monophyly of the Continental clade (Fig. 2a; Table 2). There was a strong positive relationship ( $R^2 = 0.83$ , p < 0.001 for  $PP \ge 0.95$ , and  $R^2 = 0.75$ , p < 0.001 for PP = 1) between the number of loci employed and the number of nodes resolved for the concatenation approach (Fig. 3b). The relationship was less clear in the multispecies coalescent analysis, with node support in the species tree varying more widely with the number of loci employed. The highest overall support came from analysis of 17 and 40 loci,

although only the analysis using 20 loci failed to recover either node in the Macaronesian clade and resulted in no resolution other than continental monophyly (Fig. 2b, Table 2).

#### 3.4 Ancestral state reconstruction

The ancestral state reconstruction identified four nodes showing marginal states with less than 0.9 Bayesian PP for one character, including sister relationship between individual CH010\_SR21B07 and the remaining species, the ancestral node of IS2 + IS3 (Canary Islands and Madeira), and the nodes separating Madeiran from Canarian individuals within IS2. The Island clade had a continental origin, further a Canarian origin was estimated for IS2 + IS3. The clade IS2 was estimated to have an ancestral state of 0.59 for Madeira and 0.4 for the Canary Islands (Supplementary File 4).

### 4. Discussion

### 4.1 Number of loci for phylogenetics

A recent study by O'Neill et al., (2013) examined how multilocus species tree inferences varied with differing number of loci. In their study, analysis based on the 20 and 30 most informative loci (using a parsimony criterion) resulted in high PPs, whereas node support values were lower and likelihoods failed to converge when less informative loci were added to the analysis. They concluded this was the result of the increasing number of parameters while adding loci with decreasing levels of information. Our results are not directly comparable to those of O'Neill et al. (2013) for the species tree reconstruction, because we did not explicitly order loci by parsimony-informative sites in our tests. Nonetheless, we found a strong negative correlation (Pearson R = -0.95) between the mean number of informative sites per locus and mean node support in the coalescent species tree (data not shown). This suggests that the number of informative sites was not able to explain variation in support alone, and that multiple characteristics of individual loci play an important role in

whether or not analyses achieve convergence and tree resolution. For the concatenation approach, we found a positive linear correlation between number of loci and node support. This was despite the larger number of parameters. Simulations based on 200 to 300 loci showed that the divergence time estimation using 50 loci are robust (Shen et al., 2016). In our study we observed that the reduction in node support when using a reduced set of loci (exon\_complete\_matrix *vs.* exon\_all\_data, see section 2.3) primarily affected the most derived clade (IS2), which highlights the importance of large nDNA marker sets for the reconstruction of shallow phylogenies.

# 4.2 Species delineation

392

393

394

395

396

397

398

399

400

401

402

403

404

405

406

407

408

409

410

411

412

413

414

415

416

The perfect agreement of the Bayesian clustering and mitochondrial gmyc approaches for a priori species delineation support the use of cox1 as barcoding gene for the taxa studied (e.g. Lucentini et al., 2011; Pereira-da-Conceicoa et al., 2012; Webb et al., 2012; Rutschmann et al., 2014). The distant clustering of one individual (CH010 SR21B07) in the concatenated tree analyses might be explained by incomplete lineage sorting since the species tree inferences using \*BEAST did result in a clear clustering of CT1 with low frequency of different topology (Fig. 3a). Moreover, when incomplete lineage sorting is present, standard methods for estimating species trees, such as concatenation and consensus methods, can be statistically inconsistent (Degnan et al., 2009; Roch and Steel 2014), and produce highly supported but incorrect trees (Kubatko and Degnan 2007). The majority of gene trees could support an incorrect species tree if the phylogeny is in the anomaly zone (Degnan and Rosenberg 2006). However, here this does not seem to be the case, otherwise one would expect the concatenation and coalescent approach to support different topologies (Kubatko and Degnan 2007; Liu and Edwards 2009). The inferred haplotype networks illustrate the necessity of using several individuals per species. For example, the individuals of IS2 shared several haplotypes with other species, indicating incomplete lineage sorting between the

different species.. Originally, it was thought that \*BEAST analyses would be quite robust in the presence of gene flow while migration is problematic (Heled et al., 2013). However, Leaché et al., (2014) have shown that gene flow can alter species trees, ranging from decreasing PPs for low gene flow up to altering the species tree topology when high levels of gene flow occur.

# 4.3 Species diversity

417

418

419

420

421

422

423

424

425

426

427

428

429

430

431

432

433

434

435

436

437

438

439

440

441

The use of nDNA and geographically extensive sampling uncovered a largely underestimated species diversity for C. dipterum s.l. species complex, supporting the existence of six geographically relevant species from our study (with a seventh in Asia). Recent evidence from the study of another mayfly species found fine-scale ecological differences among cryptic species detected with molecular methods (Macher et al., 2016), lending support to the ecological and evolutionary significance of these and other DNA-based findings. Another widespread species, Baetis harrisoni, was also found to consist of several cryptic species (Pereira-da-Conceicoa et al., 2012). In light of the unusually broad ecological tolerance (among mayflies) observed for C. dipterum, we also conclude that the lineage clearly consists of multiple independent species, as has been recognized by morphological taxonomy for some of the members (e.g. C. peregrinator). All of our analyses grouped two specimens of C. peregrinator from Madeira with individuals from several Canary Islands into species IS2. Gattolliat et al., (2008) described C. peregrinator as an endemic Madeiran species based on morphological characters and support from mtDNA cytochrome-oxidase b sequences. At the time, there were no nDNA sequences of Canarian C. dipterum s.l. specimens available. Rutschmann et al., (2014) assigned all Madeiran Cloeon individuals to C. peregrinator for their mtDNA phylogeny, but the specimens were not included in their gmyc analysis because there were no cox1 sequences. Based on our findings here, there is no endemic Cloeon species on Madeira.

The focus of our study was Macaronesia and therefore nDNA results are only applicable to these taxa, but the mtDNA gene tree provides evidence for broad cryptic diversity within the subfamily Cloeoninae. Although these are single-locus data and must therefore be considered preliminary, we note that the mtDNA and nDNA species delineation results were fully congruent. *Cloeon simile* included two geographically widespread European gmyc species, and *C. smaeleni* Lestage 1924 was two gmyc species, one with Saudi Arabian and one with Afrotropical distribution. The species *C. praetexum* was clearly distinct from all other examined European specimens, which was surprising because it is thought to belong to *C. simile* s.l.. The two specimens of *C. cognatum*, which is thought to be a junior synonym of *C. dipterum* by some authors, were nested within the IS1 clade. All of the above findings must be considered preliminary because they are based on mtDNA, although we note that mtDNA and nDNA markers agreed in all of the *Cloeon* species that were directly compared. Further studies on these taxa with additional molecular markers, using morphological characteristics, and including comparisons with previously described species that are now considered junior synonyms or *species inquirenda* would be a valuable complement to the work presented here.

### 4.4 Evolution, colonization, and diversification

For the species occurring in the Macaronesian region, one species appeared widely distributed on all Canary Islands and Madeira (IS2), one species was found only on the western group of the Canarian islands (IS3), and one species was found on five islands of the Azores, in Italy, in Greece and in North America (IS1). The short branches and occurrence of shared haplotypes of individuals from IS1 support very recent or perhaps ongoing gene flow. Other studies have found evidence for recent or ongoing dispersal in *Cloeon* (e.g., Monaghan et al., 2005) including a recent introduction of African *Cloeon* to South America (*C. smaeleni*, Salles et al., 2014). This long-distance dispersal ability is probably at least partly related to their reproductive flexibility including ovivipary and their ability to survive in anthropogenic

468

469

470

471

472

473

474

475

476

477

478

479

480

481

482

483

484

485

486

487

488

489

490

491

habitats. Our ancestral state reconstruction indicated that IS2 may have first colonized Madeira and then the Canaries from west to east. Colonization routes between these two archipelagos have been suggested for several taxa (Emerson et al., 2000a; Emerson et al., 2000b; Trusty et al., 2005; Illera et al., 2007; Dimitrov et al., 2008; Amorim et al., 2012), IS3 seems not to have reached La Palma and the two most eastern Canarian islands of Fuerteventura and Lanzarote. The dispersal of IS3 appears to have followed the progression rule, in which older islands are inhabited by older clades, which is further supported by stepping-stone dispersal along an east-western gradient. Our data confirm at least three and possibly four independent colonization events of the islands studied, with a European origin for the Macaronesian C. dipterum s.l.; however, long branches between Continental clades and the Island clade suggest there may be missing intermediates. These may occur in the Iberian Peninsula or North Africa. Several studies have proposed a North African origin for both the Canarian and Madeiran fauna (Brunton and Hurst 1998; Kvist et al., 2005; Weingartner et al., 2006; Gohli et al., 2015; Stervander et al., 2015). The Continental clades are also distantly related to one another and the long branches within both clades, compared to the Island clades, suggest there may be additional European species that are not included here. A strong effect of different habitat preferences between the two Canarian species, which might impact their colonization success, is evidenced. Although we recognize that our dataset was not quantitative, we observed that species IS3 generally occurred on islands with more potential habitats in comparison to IS2, which seems to have better dispersal abilities and might therefore be able to more successfully colonize islands with very little water occurrence. This pattern may be linked with the occurrence of suitable water habitats on the Canarian Islands, including the four islands of Gran Canaria, Tenerife, La Gomera, and La Palma, which all have permanent natural water sources, and the island of El Hierro where

493

494

495

496

497

498

499

500

501

502

503

504

505

506

507

508

509

510

511

512

513

514

515

several artificial standing water habitats exist due to mostly temperate climatic conditions, whereas there are only very few habitats on Fuerteventura and Lanzarote due to the arid climatic conditions. The effect of habitat use on species richness has been shown for aquatic beetles (Ribera et al., 2003c), whereby running water bodies contain more species than standing ones. This pattern also applies to the Macaronesian mayflies. The genus Baetis occurs in running waters and is species-rich, including eight island endemic species on five islands of Madeira and the Canary Islands (Rutschmann et al., 2014). In contrast, the genus Cloeon comprises three species none of which are restricted to a single island. The impact of agriculture and tourism on natural habitats (Malmqvist et al., 1995; Nilsson et al., 1998) has clearly threatened the occurrence of species living in lotic habitats (Baetis canariensis and B. pseudorhodani, Rutschmann et al., 2014), but it may have had less of an effect on C. dipterum. The recent records of mayflies from El Hierro indicate a recent anthropogenic import of the species, moreover because it is the youngest island of the Canarian archipelago and its remote geographical position. Interestingly, there were eight sampling sites (out of 32 examined sites, i.e. 25%) in which both species IS2 and IS3 occurred sympatrically. Four of these localities were natural habitats. However, more work needs to be done to make quantitative assessments on species occurrence and local abundance of the two distinct species occurring on the same habitats. A wider geographic sampling, focusing on the specimens from the European mainland and North Africa will be needed to clarify the origin and distribution of the C. dipterum s.l. species complex. We expect to find more individuals from distinct geographic localities belonging to the species IS1, since this species seems to exhibit long distance trans-oceanic dispersal abilities.

## 5. Conclusion

516

517

518

519

520

521

522

523

524

525

526

527

528

529

530

531

532

533

534

535

536

537

538

539

540

Our aims were to delineate the species boundaries within the *C. dipterum* species complex and place these lineages within a phylogenetic framework, in order to better understand their evolution on the Macaronesian Islands. Robust phylogenetic reconstruction of such closely related species can be challenging, but is a necessary step in the understanding of evolutionary processes of diversification and adaptation. Most readily available nDNA loci (e.g. rRNA) do not exhibit suitable polymorphism, resulting in a large dependence on mtDNA for phylogenetics (Garrick et al., 2015). A distinct advantage of using multiple nDNA loci comes from the advent of multilocus species tree reconstruction methods. These are important tools in the reconstruction of relationships between close relatives, which is often intractable based on single-locus (i.e. mtDNA) data. The difficulty in developing large numbers of nDNA loci remains one of the primary reasons that there are few model systems available for detailed studies of speciation and diversification processes. This is particularly true for freshwater insects, despite their overwhelming contribution to global biodiversity (Dijkstra et al., 2014). Here we developed a large set of nDNA loci using draft whole-genome sequencing combined with sequences from a published EST library that was constructed using a different subfamily. All of the procedures we used have since been incorporated into a single analysis pipeline (Rutschmann et al., accepted). Our results show that even for taxa with very limited available genomic resources, it is possible to develop sets of nuclear loci that produce fully resolved and supported coalescent-based species trees and single-matrix phylogenetic trees. Using these results, we were able to infer species boundaries within the largely cryptic C. dipterum s.l. species complex and reconstruct the diversification and island colonization history of these species with confidence.

#### **Author contributions**

541

546

- S.R., M.S. and M.T.M. conceived the study. S.R., H.D., D.H.F., J.-L.G., S.J.H., and P.M.R.,
- provided samples. S.R., H.D., S.S., and R.D. contributed analytical tools. S.R., and M.T.M
- performed and interpreted the analyses. S.R. and M.T.M wrote the manuscript. All authors
- provided comments and approved the final manuscript.

# Acknowledgements

- We are grateful to B. Ortiz Crespo, S. Mbedi, K. Preuß, and L. Wächter for laboratory work;
- D. Murányi, A. Wagner, A. Przhiboro, P. Manko, M. F. Geiger, K. Kurzrock, L.F. Pires Braz,
- K. C. Gritzalis, and M. Alp for field work; P. Rutschmann and the HPC Service of ZEDAT,
- Freie Universität Berlin for access to high-performance computing resources. We are greatly
- 551 indebted to M. Báez for obtaining sampling permits for the Canary Islands and to the
- authorities who provided us with the collection permissions. We are very grateful to our
- research groups, especially to I. Lucas Lledó and M. Gamboa for their constructive comments
- on this work. This is publication number ### of the Berlin Center for Genomics in
- 555 Biodiversity Research. This work was supported by the Leibniz Association (PAKT für
- Forschung und Innovation) project FREDIE (SAW-2011-ZFMK-3 to M.T.M.) and by a travel
- award from the Leibniz-Institute of Freshwater Ecology and Inland Fisheries to S.R..
- Individual support was provided by the Japan Society for the Promotion of Science (Long-
- Term Research Fellowship L15543 to M.T.M.), the Swiss National Science Foundation
- 560 (Early PostDoc.Mobility fellowship P2SKP3 158698 to S.R.), the European Investment
- Funds by FEDER/COMPETE/POCI Operacional Competitiveness and Internationalisation
- Programme (POCI-01-0145-FEDER-006958 to S.J.H.), and the National Funds by FCT -
- Portuguese Foundation for Science and Technology (UID/AGR/04033/2013 to S.J.H. and
- 564 SFRH/BPD/99461/2014 to P.M.R.).

#### References

- Amorim, I.R., Emerson, B.C., Borges, P.A.V., Wayne, R.K., 2012. Phylogeography and molecular phylogeny of Macaronesian island *Tarphius* (Coleoptera: Zopheridae): why are there so few species in the Azores? J. Biogeogr. 39, 1583-1595.
- Arnedo, M.A., Oromi, P., De Abreu, S.M., Ribera, C., 2008. Biogeographical and evolutionary patterns in the Macaronesian shield-backed katydid genus *Calliphona* Krauss,
- 571 1892 (Orthoptera: Tettigoniidae) and allies as inferred from phylogenetic analyses of multiple mitochondrial genes. Syst. Entom. 33, 145-158.
- Avise, J.C., Nelson, W.S., Bowen, B.W., Walker, D., 2000. Phylogeography of colonially nesting seabirds, with special reference to global matrilineal patterns in the sooty tern (*Sterna fuscata*). Mol. Ecol. 9, 1783-1792.
- Barber-James, H.M., Gattolliat, J.-L., Sartori, M., Hubbard, M.D., 2008. Global diversity of mayflies (Ephemeroptera, Insecta) in freshwater. Hydrobiologia 595, 339-350.
- Bauernfeind, E., Soldán, T. 2012. The Mayflies of Europe. Ollerup, Apollo Books.
- Bouckaert, R., Heled, J., Kuhnert, D., Vaughan, T., Wu, C.H., Xie, D., Suchard, M.A., Rambaut, A., Drummond, A.J., 2014. BEAST 2: a software platform for Bayesian evolutionary analysis. PLoS Comp. Biol. 10, e1003537.
- Brinck, P., Scherer, E. 1961. On the Ephemeroptera of the Azoreas and Madeira. Boletim do Museu Municipal do Funchal 47, 55-66.
- Brunton, C.F.A., Hurst, G.D.D., 1998. Mitochondrial DNA phylogeny of Brimstone butterflies (genus *Gonepteryx*) from the Canary Islands and Madeira. Biol. J. Linn. Soc.
- 586 Lond. 63, 69-79.

- Darriba, D., Taboada, G.L., Doallo, R., Posada, D., 2012. jModelTest 2: more models, new heuristics and parallel computing. Nat. Methods 9, 772.
- Degnan, J.H., DeGiorgio, M., Bryant, D., Rosenberg, N.A., 2009. Properties of consensus methods for inferring species trees from gene trees. Syst. Biol. 58, 35-54.
- Degnan, J.H., Rosenberg, N.A., 2006. Discordance of species trees with their most likely gene trees. PLoS Genet. 2, e68.
- Degnan, J.H., Rosenberg, N.A., 2009. Gene tree discordance, phylogenetic inference and the multispecies coalescent. Trends Ecol. Evol. 24, 332-340.
- Degnan, J.H., Rosenberg, N.A., Stadler, T., 2012. A characterization of the set of species trees that produce anomalous ranked gene trees. IEEE/ACM Trans. Comput. Biol. Bioinform. 9, 1558-1568.
- Díaz-Pérez, A.J., Sequeira, M., Santos-Guerra, A., Catalán, P., 2012. Divergence and biogeography of the recently evolved Macaronesian red *Festuca* (Gramineae) species inferred from coalescence-based analyses. Mol. Ecol. 21, 1702-1726.
- Dijkstra, K.D., Monaghan, M.T., Pauls, S.U., 2014. Freshwater biodiversity and aquatic insect diversification. Annu. Rev. Entomol. 59, 143-163.
- Dimitrov, D., Arnedo, M.A., Ribera, C., 2008. Colonization and diversification of the spider genus *Pholcus* Walckenaer, 1805 (Araneae, Pholcidae) in the Macaronesian archipelagos: evidence for long-term occupancy yet rapid recent speciation. Mol. Phylogenet. Evol. 48, 596-614.
- Drotz, M.K., 2003. Speciation and mitochondrial DNA diversification of the diving beetles

  Agabus bipustulatus and A. wollastoni (Coleoptera, Dytiscidae) within Macaronesia. Biol.

  J. Linn. Soc. Lond. 79, 653-666.
- Drummond, A.J., Rambaut, A., 2007. BEAST: Bayesian evolutionary analysis by sampling trees. BMC Evol. Biol. 7, 214.
- Edwards, S.V., 2009. Is a new and general theory of molecular systematics emerging? Evolution 63, 1-19.
- Emerson, B.C., 2002. Evolution on oceanic islands: molecular phylogenetic approaches to understanding pattern and process. Mol. Ecol. 11, 951-966.
- Emerson, B.C., Kolm, N., 2005. Species diversity can drive speciation. Nature 434, 1015-1017.
- Emerson, B.C., Oromí, P., 2005. Diversification of the forest beetle genus *Tarphius* on the Canary Islands, and the evolutionary origins of island endemics. Evolution 59, 586-598.
- Emerson, B.C., Oromí, P., Godfrey, M.H., 2000a. Interpreting colonization of the *Calathus* (Coleoptera: Carabidae) on the Canary Islands and Madeira through the application of the parametric bootstrap. Evolution 54, 2081-2090.
- Emerson, B.C., Oromí, P., Hewitt, G.M., 2000b. Tracking colonization and diversification of insect lineages on islands: mitochondrial DNA phylogeography of *Tarphius canariensis* (Coleoptera: Colydiidae) on the Canary Islands. Proc. Biol. Sci. 267, 2199-2205.
- Falush, D., Stephens, M., Pritchard, J.K., 2003. Inference of population structure using multilocus genotype data: linked loci and correlated allele frequencies. Genetics 164, 1567-1587.
- Faria, C.M.A., Machado, A., Amorim, I.R. Gage, M.J.G., Borges, P.A.V., Emerson, B.C., 2016. Evidence for multiple founding lineages and genetic admixture in the evolution of species within an oceanic island weevil (Coleoptera, Curculionidae) super-radiation. J. Biogeogr., 43, 178-191.
- Fitch, W.M., 1970. Distinguishing homologous from analogous proteins. Syst. Zool. 19, 99-113.
- Flot, J.F., 2010. seqphase: a web tool for interconverting phase input/output files and fasta sequence alignments. Mol. Ecol. Resour. 10, 162-166.

- Fujisawa, T., Barraclough, T.G., 2013. Delimiting species using single-locus data and the Generalized Mixed Yule Coalescent approach: a revised method and evaluation on simulated data sets. Syst. Biol. 62, 707-724.
- Garrick, R.C., Bonatelli, I.A., Hyseni, C., Morales, A., Pelletier, T.A., Perez, M.F., Rice, E.,
   Satler, J.D., Symula, R.E., Thomé, M.T., Carstens, B.C., 2015. The evolution of
   phylogeographic data sets. Mol. Ecol. 24, 1164-1171.
- Garrick, R.C., Sunnucks, P., Dyer, R.J., 2010. Nuclear gene phylogeography using PHASE: dealing with unresolved genotypes, lost alleles, and systematic bias in parameter estimation. BMC Evol. Biol. 10, 118.
- Gattolliat, J.-L., Hughes, S.J., Monaghan, M.T., Sartori, M., 2008. Revision of Madeiran mayflies (Insecta, Ephemeroptera). Zootaxa 1957, 52-68.
- 648 Giarla, T.C., Esselstyn, J.A., 2015. The challenges of resolving a rapid, recent radiation: empirical and simulated phylogenomics of philippine shrews. Syst. Biol. 64, 727-740.
- 650 Gillespie, R.G., Croom, H.B., Palumbi, S.R., 1994. Multiple origins of a spider radiation in 651 Hawaii. Proc. Natl. Acad. Sci. U.S.A. 91, 2290-2294.
- 652 Gillespie, R.G., Roderick, G.K., 2002. Arthropods on islands: colonization, speciation, and conservation. Annu. Rev. Entomol. 47, 595-632.
- Gohli, J., Leder, E.H., Garcia-Del-Rey, E., Johannessen, L.E., Johnsen, A., Laskemoen, T.,
   Popp, M., Lifjeld, J.T., 2015. The evolutionary history of Afrocanarian blue tits inferred
   from genome wide SNPs. Mol. Ecol. 24, 180-191.
- Grant, P., Grant, R., 2008. How and why species multiply: the radiation of Darwin's finches.
   New Jersey, Princeton University Press.
- Guindon, S., Gascuel, O., 2003. A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. Syst. Biol. 52, 696-704.
- Harrigan, R.J., Mazza, M.E., Sorenson, M.D., 2008. Computation vs. cloning: evaluation of two methods for haplotype determination. Mol. Ecol. Resour. 8, 1239-1248.
- Heled, J., Bryant, D., Drummond, A.J., 2013. Simulating gene trees under the multispecies coalescent and time-dependent migration. BMC Evol. Biol. 13, 44.
- Heled, J., Drummond, A.J., 2010. Bayesian inference of species trees from multilocus data.
   Mol. Biol. Evol. 27, 570-580.
- Hughes, S.J., Malmqvist, B., 2005. Atlantic Island freshwater ecosystems: challenges and considerations following the EU Water Framework Directive. Hydrobiologia 8, 289-297.
- Illera, J.C., Emerson, B.C., Richardson, D.S., 2007. Population history of Berthelot's pipit: colonization, gene flow and morphological divergence in Macaronesia. Mol. Ecol. 16, 4599-4612.
- Jordal, B.H., Hewitt, G.M., 2004. The origin and radiation of Macaronesian beetles breeding in Euphorbia: the relative importance of multiple data partitions and population sampling. Syst. Biol. 53, 711-734.
- Juan, C., Oromí, P., Hewitt, G.M., 1997. Molecular phylogeny of darkling beetles from the Canary Islands: comparison of inter island colonization patterns in two genera. Biochem. Syst. Ecol. 25, 121-130.
- Juan, C., Emerson, B.C., Oromí, P., Hewitt, G.M., 2000. Colonization and diversification: towards a phylogeographic synthesis for the Canary Islands. Trends Ecol. Evol. 15, 104-109.
- Katoh, K., Standley, D.M., 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Mol. Biol. Evol. 30, 772-780.
- Knowles, L.L., Kubatko, L.S., 2010. Estimating Species Trees: Practical and Theoretical Aspects. Wiley-Blackwell.

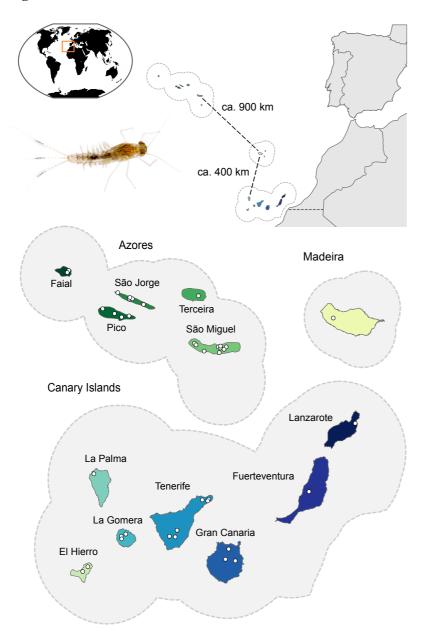
- Kozak, K.M., Wahlberg, N., Neild, A.F., Dasmahapatra, K.K., Mallet, J., Jiggins, C.D., 2015.
- Multilocus species trees show the recent adaptive radiation of the mimetic *Heliconius* butterflies. Syst. Biol. 64, 505-524.
- Kubatko, L.S., Degnan, J.H., 2007. Inconsistency of phylogenetic estimates from concatenated data under coalescence. Syst. Biol. 56, 17-24.
- Kvist, L., Broggi, J., Illera, J.C., Koivula, K., 2005. Colonisation and diversification of the blue tits (*Parus caeruleus* teneriffae-group) in the Canary Islands. Mol. Phylogenet. Evol. 34, 501-511.
- Leaché, A.D., Fujita, M.K., Minin, V.N., Bouckaert, R.R., 2014. Species delimitation using genome-wide SNP data. Syst. Biol. 63, 534-542.
- Liu, L., Edwards, S.V., 2009. Phylogenetic analysis in the anomaly zone. Syst. Biol. 58, 452-460.
- Lopez, J.V., Yuhki, N., Masuda, R., Modi, W., O'Brien, S.J., 1994. Numt, a recent transfer and tandem amplification of mitochondrial DNA to the nuclear genome of the domestic cat. J. Mol. Evol. 39, 174-190.
- Losos, J.B., Ricklefs, R.E., 2009. Adaptation and diversification on islands. Nature 457, 830-836.
- Lucentini, L., Rebora, M., Puletti, M.E., Gigliarelli, L., Fontaneto, D., Gaino, E., Panara, F.,
   2011. Geographical and seasonal evidence of cryptic diversity in the *Baetis rhodani* complex (Ephemeroptera, Baetidae) revealed by means of DNA taxonomy. Hydrobiologia
   673, 215-228.
- Macher, J.N., Salis, R.K., Blakemore, K.S., Tollrian, R., Matthaei, C.D., Leese, F., 2016.
  Multiple-stressor effects on stream invertebrates: DNA barcoding reveals contrasting responses of cryptic mayfly species. Ecol. Indic. 61, 159-169.
- Maddison, W.P., 1997. Gene trees in species trees. Syst. Biol. 46:523.
- Maddison, W.P., Maddison, D.R., 2011. Mesquite: a modular system for evolutionary analysis. http://mesquiteproject.org (accessed 25.03. 2016).
- Malmqvist, B., Nilsson, A.N., Báez, M., 1995. Tenerife's freshwater macroinvertebrates: status and threats (Canary Islands, Spain). Aquat. Conserv. 5, 1-24.
- Matschiner, M., 2015. Fitchi: Haplotype genealogy graphs based on the Fitch algorithm. Bioinformatics 32, 1250-1252.
- Monaghan, M.T., Balke, M.M., Pons, J.J., Vogler, A.P., 2006. Beyond barcodes: complex DNA taxonomy of a South Pacific Island radiation. Proc. Biol. Sci. 273, 887-893.
- Monaghan, M.T., Gattolliat, J.L., Sartori, M., Elouard, J.M., James, H., Derleth, P., Glaizot,
   O., de Moor, F., Vogler, A.P., 2005. Trans-oceanic and endemic origins of the small
   minnow mayflies (Ephemeroptera, Baetidae) of Madagascar. Proc. Biol. Sci. 272, 1829-
- 721 1836.
- Morvan, C., Malard, F., Paradis, E., Lefebure, T., Konecny-Dupre, L., Douady, C.J., 2013.
- Timetree of Aselloidea reveals species diversification dynamics in groundwater. Syst. Biol. 62, 512-522.
- Nilsson, A.N., Malmqvist, B., Báez, M., Blackburn, J.H., Armitage, P.D., 1998. Stream insects and gastropods in the island of Gran Canaria (Spain). Ann. Limnol.-Int. J. Lim.. 34, 413-435.
- Nogales, M., Delgado, J.D., Medina, F.M., 1998. Shrikes, lizards and *Lycium intricatum* (Solanaceae) fruits: a case of indirec seed dispersal on an oceanic island (Alegranza, Canary Islands). J. Ecol. 86, 866-871.
- O'Neill, E.M., Schwartz, R., Bullock, C.T., Williams, J.S., Shaffer, H.B., Aguilar-Miguel, X.,
- Parra-Olea, G., Weisrock, D.W., 2013. Parallel tagged amplicon sequencing reveals major
- lineages and phylogenetic structure in the North American tiger salamander (Ambystoma
- tigrinum) species complex. Mol. Ecol. 22, 111-129.

- Paradis, E., Claude, J., Strimmer, K., 2004. APE: Analyses of Phylogenetics and Evolution in R language. Bioinformatics 20, 289-290.
- Pereira-da-Conceicoa, L.L., Price, B.W., Barber-James, H.M., Barker, N.P., de Moor, F.C.,
- Villet, M.H., 2012. Cryptic variation in an ecological indicator organism: mitochondrial
- and nuclear DNA sequence data confirm distinct lineages of *Baetis harrisoni* Barnard (Ephemeroptera: Baetidae) in southern Africa. BMC Evol. Biol. 12, 26.
- Pritchard, J.K., Stephens, M., Donnelly, P., 2000. Inference of population structure using multilocus genotype data. Genetics 155, 945-959.
- R Core Team. 2016. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. https://www.R-project.org (acessed 26.03.2016).
- Rannala, B., Yang, Z., 2003. Bayes estimation of species divergence times and ancestral population sizes using DNA sequences from multiple loci. Genetics 164, 1645-1656.
- Raposeiro, P.M., Cruz, A.M., Hughes, S.J., Costa, A.C., 2012. Azorean freshwater invertebrates: Status, threats and biogeographic notes. Limnetica 31, 13-22.
- Revell, L., 2012. phytools: An R package for phylogenetic comparative biology (and other things). Methods Ecol. Evol. 3, 217-223.
- Ribera, I., Bilton, D.T., Balke, M., Hendrich, L., 2003a. Evolution, mitochondrial DNA phylogeny and systematic position of the Macaronesian endemic *Hydrotarsus* Falkenström (Coleoptera: Dytiscidae). Syst. Entomol. 28, 493-508.
- Ribera, I., Bilton, D.T., Vogler, A.P., 2003b Mitochondrial DNA phylogeography and population history of Meladema diving beetles on the Atlantic Islands and in the Mediterranean basin (Coleoptera, Dytiscidae). Mol. Ecol. 12, 153-167.
- Ribera, I., Foster, G.N., Vogler, A.P., 2003c. Does habitat use explain large scale species richness patterns of aquatic beetles in Europe? Ecography 26, 145-152.
- Roch, S., Steel, M., 2014. Likelihood-based tree reconstruction on a concatenation of aligned sequence data sets can be statistically inconsistent. Theor. Popul. Biol. 100C, 56-62.
- Ronquist, F., Teslenko, M., van der Mark, P., Ayres, D.L., Darling, A., Hohna, S., Larget, B.,
   Liu, L., Suchard, M.A., Huelsenbeck, J.P., 2012. MrBayes 3.2: efficient Bayesian
   phylogenetic inference and model choice across a large model space. Syst. Biol. 61, 539 542.
- Rutschmann, S., Detering, H., Simon, S., Fredslund, J., Monaghan, M.T. (accepted)
  DiscoMark: Nuclear marker discovery from orthologous sequences using draft genome
  data. Mol. Ecol. Resour.
- Rutschmann, S., Gattolliat, J.-L., Hughes, S.J., Sartori, M., Monaghan, M.T. 2014. Evolution and island endemism of morphologically cryptic *Baetis* and *Cloeon* species (Ephemeroptera, Baetidae) on the Canary Islands and Madeira. Freshwater Biol. 59, 2516-2527.
- Salles, F.F., Gattolliat, J.-L., Angeli, K.B., De-Souza, M.R., Goncalves, I.C., Nessimian, J.L., Sartori, M., 2014. Discovery of an alien species of mayfly in South America (Ephemeroptera). ZooKeys 1-16.
- Schluter, D., 2000. The Ecology of Adaptive Radiation. New York, Oxford University Press.
- Shen, X.X., Liang, D., Chen, M.Y., Mao, R.L., Wake, D.B., Zhang, P., 2016. Enlarged multilocus data set provides surprisingly younger time of origin for the Plethodontidae, the largest family of salamanders. Syst. Biol. 65, 66-81.
- Soldán, T., Thomas, A., 1983. New a little-known species of mayflies (Ephemeroptera) from
   Algeria. Acta Entomol. Bohemos. 80, 356-376.
- Stamatakis, A., 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics 30, 1312-1313.

- Stauder, A., 1995. Survey of the Madeiran limnological fauna and their zoogeographical distribution. Boletim do Museu Municipal do Funchal 4, 715-723.
- Stephens, M., Donnelly, P., 2003. A comparison of bayesian methods for haplotype reconstruction from population genotype data. Am. J. Hum. Genet. 73. 1162-1169.
- Stephens, M., Smith, N.J., Donnelly, P., 2001. A new statistical method for haplotype reconstruction from population data. Am. J. Hum. Genet. 68, 978-989.
- Stervander, M., Illera, J.C., Kvist, L., Barbosa, P., Keehnen, N.P., Pruisscher, P., Bensch, S., Hansson, B., 2015. Disentangling the complex evolutionary history of the Western Palearctic blue tits (*Cyanistes* spp.) - phylogenomic analyses suggest radiation by multiple colonization events and subsequent isolation. Mol. Ecol. 24, 2477-2494.
- Sukumaran, J., Holder, M.T., 2010. DendroPy: a Python library for phylogenetic computing. Bioinformatics 26, 1569-1571.
- Toussaint, E.F., Condamine, F.L., Hawlitschek, O., Watts, C.H., Porch, N., Hendrich, L., Balke, M., 2015. Unveiling the diversification dynamics of australasian predaceous diving beetles in the cenozoic. Syst. Biol. 64, 3-24.
- Trusty, J.L., Olmstead, R.G., Santos-Guerra, A., Sa-Fontinha, S., Francisco-Ortega, J., 2005.

  Molecular phylogenetics of the Macaronesian-endemic genus *Bystropogon* (Lamiaceae):
  palaeo-islands, ecological shifts and interisland colonizations. Mol. Ecol. 14, 1177-1189.
- Tseng, S.-P., Li, S.-H., Hsieh, C.-H., Wang, H.-Y., Lin, S.-M., 2014. Influence of gene flow on divergence dating implications for the speciation history of *Takydromus* grass lizards. Mol. Ecol. 23, 4770-4784.
- Vuataz, L., Sartori, M., Gattolliat, J.-L., Monaghan, M.T., 2013. Endemism and diversification in freshwater insects of Madagascar revealed by coalescent and phylogenetic analysis of museum and field collections. Mol. Phylogenet. Evol. 66, 979-991.
- Webb, J.M., Jacobus, L.M., Funk, D.H., Zhou, X., Kondratieff, B., Geraci, C.J., DeWalt,
  R.E., Baird, D.J., Richard, B., Phillips, I., Herbert, P.D., 2012. A DNA barcode library for
  North American Ephemeroptera: progress and prospects. PLoS ONE 7, e38063.
- Weingartner, E., Wahlberg, N., Nylin, S., 2006. Speciation in *Pararge* (Satyrinae: Nymphalidae) butterflies North Africa is the source of ancestral populations of all Pararge species. Syst. Entom. 31, 621-632.
- Wielstra, B., Duijm, E., Lagler, P., Lammers, Y., Meilink, W.R., Ziermann, J.M., Arntzen,
   J.W., 2014. Parallel tagged amplicon sequencing of transcriptome-based genetic markers
   for *Triturus* newts with the Ion Torrent next-generation sequencing platform. Mol. Ecol.
   Resour. 14, 1080-1089.
- Zhang, D.-X., Hewitt, G.M., 2003. Nuclear DNA analyses in genetic studies of populations: practice, problems and prospects. Mol. Ecol. 12, 563-584.
- Zheng, Y., Peng, R., Kuro-o, M., Zeng, X,. 2011. Exploring patterns and extent of bias in estimating divergence time from mitochondrial DNA sequence data in a particular lineage: a case study of salamanders (order Caudata). Mol. Biol. Evol. 28, 2521-2535.

### 824 Figures



**Fig. 1.** Overview of the sampling localities on the Macaronesian region. The three archipelagos of the Azores, Madeira, and the Canary Islands are shown in detail, whereby the 38 sampling sites are indicated by white dots. For the Azores and Madeira only islands with sampling sites are shown in the detailed view. Photo of *Cloeon dipterum* s.l. larvae by Amanda44 / CC BY 3.0.

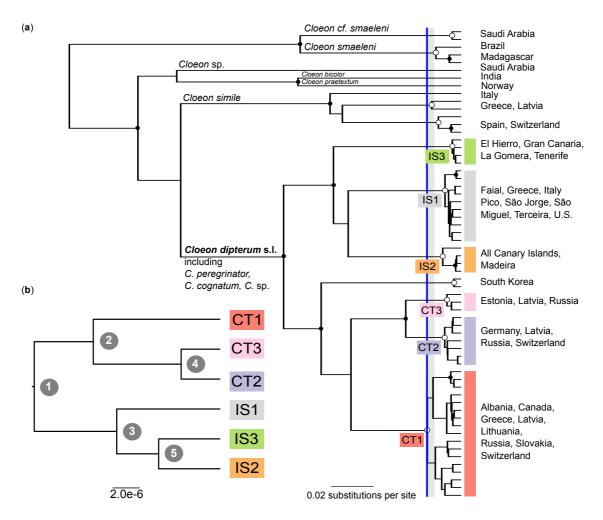
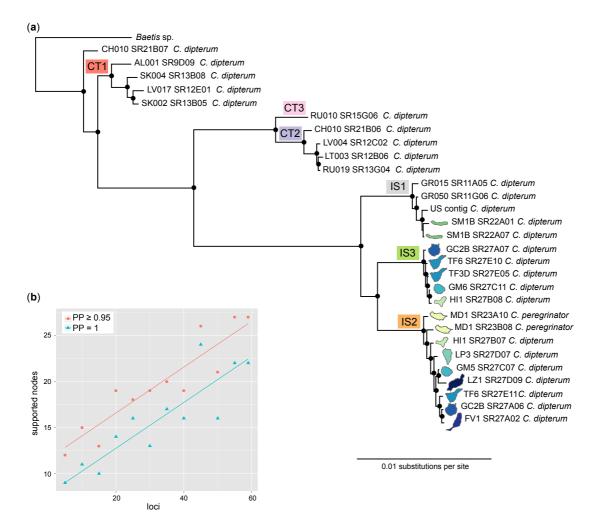


Fig. 2. Mitochondrial gene tree and nuclear species tree topology. Species delimitation from (a) the general mixed Yule-coalescent (gmyc) approach based on mitochondrial data (single-locus), and (b) the multispecies coalescent approach based on nuclear data (59 loci). (a), the ultrametric mitochondrial cox1 gene tree was used as input for the gmyc analysis of Cloeon sp. All circles indicate well supported nodes (Bayesian posterior probability (PP)  $\geq 0.95$ ). Open circles at subtending nodes indicate sequence clusters corresponding to single gmyc species. Colors and alphanumeric codes indicate the six putative gmyc species. The outgroup  $Baetis\ rhodani$  is not shown. The blue line indicates the point of maximum-likelihood fit of the single-threshold gmyc model with 95% confidence interval in grey shading. Terminal labels indicate sampling regions (Supplementary Table 1). (b), the species trees of C.  $dipterum\ s.l.$  inferred by multispecies coalescent approach based on the exonhap\_all\_data matrix. Posterior probabilities of the five nodes are indicated in Table 2.



**Fig. 3.** Bayesian inference reconstruction of concatenation approach. (a), the phylogenetic relationships among *Cloeon dipterum* s.l. (including *C. peregrinator*) based on concatenated exon\_all\_data matrix. Filled circles indicate well supported nodes (Bayesian posterior probability (PP)  $\geq$  0.95). (b), the positive linear relationship between number of loci analyzed and number of supported nodes. Red circles represent nodes with PP  $\geq$  0.95 and blue triangles represent nodes with PP = 1. Sets of loci are reported in Supplementary Table 4. The R<sup>2</sup> for PP  $\geq$  0.95 were 0.83 (p < 0.001), and 0.75 (p < 0.001) for PP = 1.

**Table 1.** Overview of seven sequence alignments, including one based on mitochondrial sequences and six based on nuclear sequences. The mitochondrial sequence alignment including only the species group of *Cloeon dipterum* s.l. comprised 130 taxa with 148 variable sites. Matrices containing all taxa and all loci were >75% complete; the matrices containing only loci sequenced for all taxa were 100% complete. Exon matrices refer to exon sequence alignments and the exonhap matrices refer to exon haplotype sequences.

Data Matrix	Concatenated	Number	Number	Number of	Description	
	Length [bp]	of Taxa	of Loci	Variable Sites		
mitochondrial	658	148	1	240	All cox1 sequences used for putative species assignment	
all_data	32,213	29	59	2,481	All taxa and loci; introns and exons	
exon_all_data	24,168	29	59	1,118	All taxa and loci; exons	
complete_matrix	8,565	29	17	648	Only loci sequenced for all taxa; introns and exons	
exon_complete_matrix	6,485	29	17	293	Only loci sequenced for all taxa; exons	
exonhap_all_data		29	59	1,390	All taxa and loci; haplotypes of exons	
exonhap_complete_matrix		29	17	361	Only loci sequenced for all taxa; haplotypes of exons	

**Table 2.** Node support values of the species tree analysis (Fig. 2b) using six different sets of loci (Supplementary Table 4). Support values are given as Bayesian posterior probability (PP).

	Node Support								
Number of Loci	All (1)	Continental (2)	Island (3)	CT2+CT3 (4)	IS2+IS3 (5)	Mean			
17	1.00	0.99	1.00	1.00	1.00	1.00			
20	1.00	1.00	-	1.00	-	0.60			
30	1.00	0.85	0.87	0.87	0.99	0.92			
40	1.00	1.00	1.00	0.67	0.83	0.70			
50	1.00	0.83	0.83	0.83	1.00	0.87			
59	1.00	0.83	0.83	0.83	1.00	0.87			