

1 How the FLP do nematodes jump?

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14 Abstract

15 Entomopathogenic nematodes (EPNs) are a guild of obligate insect parasites which share

16 many physiological and behavioural traits with mammalian strongylid and strongyloidid

17 parasites; including host-finding nictation behaviour. EPNs are also interesting from the

18 perspective of insect biocontrol. Like other parasitic nematodes, EPNs employ a

19 sophisticated chemosensory apparatus to detect potential hosts and communicate with

20 conspecifics. Understanding the underlying molecular basis of relevant host-finding

21 behaviours could facilitate improved EPN biocontrol approaches, and could lend insight to

22 similar behaviours in economically important animal parasites. FMRFamide-like peptides are

23 enriched and conserved across the Phylum Nematoda, and have been linked with motor and

24 sensory function, including dispersal and aggregating behaviours in the free living nematode

25 *Caenorhabditis elegans*. RNA interference (RNAi) was used to knockdown the expression

26 of the FMRFamide-like protein 21 (GLGPRPLRFamide) gene (*flp-21*) in *Steinernema*
27 *carpocapsae* infective juveniles (IJs). Our data show that *S. carpocapsae* is sensitive to
28 neuronal RNAi, and that knockdown of *flp-21* has a significant impact on dispersal, nictation
29 and jumping behaviours. Immunocytochemical localisation of FLP-21 to paired anterior
30 neurons corroborates the RNAi data, further suggesting a role in sensory modulation. This
31 study represents the first demonstration of a functional RNAi pathway in *S. carpocapsae*,
32 through which we have linked a single gene product to the coordination of nematode
33 jumping, nictation, and dispersal behaviours in IJs. These data can underpin efforts to study
34 these behaviours in other economically important parasites, and could facilitate molecular
35 approaches to EPN strain improvement for biocontrol.

36

37 **Introduction**

38 Entomopathogenic nematodes (EPNs) borrow their name from the entomopathogenic
39 bacteria (*Photorhabdus*, *Serratia* and *Xenorhabdus* spp.) with which they form a commensal
40 relationship. These nematodes provide a stable environment for the bacteria, and act as a
41 vector between insect hosts. Once the nematode has invaded an insect, the nematode
42 exsheaths (or 'recovers') and entomopathogenic bacteria are regurgitated into the insect
43 haemolymph; the bacteria then rapidly kill and metabolise the insect, providing nutrition and
44 developmental cues for the nematode. These entomopathogenic bacteria are then
45 transmitted between nematode generations (Dillman et al., 2012). The entomopathogenic
46 lifestyle has been found to arise independently in nematodes, at least three times, spanning
47 significant phylogenetic diversity. *Heterorhabditis* and *Oscheius* spp. (Torrini et al., 2015)
48 reside within clade 9 along with major strongylid parasites of man and animal (Holterman et
49 al., 2006); *Steinernema* spp. reside within clade 10 alongside strongyloidid parasites (Viney
50 and Lok, 2007).

51

52 Nictation is a dispersal and host-finding strategy, enacted by nematodes which stand upright
53 on their tails, waving their anterior in the air (Lee et al., 2012). This behaviour is shared
54 amongst many economically important animal parasitic and entomopathogenic nematodes,
55 alongside the model nematode *C. elegans*, for which nictation is a phoretic dispersal
56 behaviour of dauer larvae, used to increase the likelihood of attachment to passing animals.
57 Nictation is regulated by amphidial IL2 neurons in *C. elegans*, which occur in lateral triplets
58 either side of the pharyngeal metacarpus (Lee et al., 2012; Schroeder et al., 2013). IL2
59 neurons display significant remodelling from *C. elegans* L3 to dauer (the only life-stage to
60 enact nictation behaviours) such that connectivity with other chemosensory and cephalic
61 neurons is enhanced (Schroeder et al., 2013). It has been shown that IL2 neurons express
62 the DES-2 acetylcholine receptor subunit, and that cholinergic signalling is requisite for
63 nictation (Pereira et al., 2015; Zhang et al., 2014; Lee et al., 2012; Yassin et al., 2001).
64 Additionally, the central pair of IL2 neurons express the FMRFamide-like peptide (FLP)
65 receptor, NPR-1 (Coates and de Bono, 2002). To date there are two known NPR-1 agonists;
66 FLP-18 and FLP-21 (Rogers et al., 2003). However, there is also known redundancy of
67 FLP-18 and FLP-21 in signalling through other neuropeptide receptors (NPR-4, -5, -6 -10, -
68 11, and NPR-2, -3, -5, -6, 11, respectively) in heterologous systems (Ezcurra et al., 2016;
69 and reviewed by Li and Kim, 2014), making functional linkage difficult.

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71 *Steinernema* spp. also display a highly specialised jumping behaviour which is thought to
72 enhance both dispersal and host attachment. Jumping occurs when a nictating infective
73 juvenile (IJ) unilaterally contracts body wall muscles bringing the anterior region towards the
74 posterior region, forming a loop. This generates high pressure within the IJ pseudocoel, and
75 differential stretching and compression forces across the nematode cuticle. Release of this
76 unilateral contraction, in conjunction with the correction of cuticle pressure, triggers enough
77 momentum for an IJ to jump a distance of nine times body length, to a height of seven times
78 body length (Campbell and Kaya, 1999).

79

80 The recent publication of five *Steinernema* spp. genomes, along with stage-specific
81 transcriptomes (Dillman et al., 2015) represents a valuable resource, alongside the
82 previously published genome of *Heterorhabditis bacteriophora* (Bai et al., 2013). The
83 genome of *Steinernema carpocapsae* is the most complete, at an estimated 85.6 Mb, with
84 predicted coverage of 98% (Dillman et al., 2015). *S. carpocapsae* was selected as a test
85 subject for our study due to the quality of genome sequence, and the interesting biology
86 underpinning dispersal and host-finding behaviours in this species. The close phylogenetic
87 relationship between *Steinernema* spp. coupled with a diverse behavioural repertoire,
88 particularly in terms of dispersal and host-finding (Castelletto et al., 2014; Lee et al., 2016),
89 make this genus an extremely attractive model for comparative neurobiology. The aim of
90 this study was to examine RNAi functionality in *S. carpocapsae*, and to probe the
91 involvement of FLP-21 in coordinating nictation, jumping and dispersal phenotypes, as a
92 prelim to probing the neuronal and molecular underpinnings of host-finding behaviour in this
93 genus.

94

95 **Materials and Methods**

96 ***S. carpocapsae* culture**

97 *S. carpocapsae* (ALL) was maintained in *Galleria mellonella* at 23°C. IJs were collected by
98 White trap (White, 1927) in a solution of Phosphate Buffered Saline (PBS). Freshly emerged
99 IJs were used for each experiment.

100

101 **BLAST analysis of *S. carpocapsae* RNAi pathway**

102 BLAST analysis of RNAi pathway components was conducted as in Dalzell et al. (2011),
103 using a modified list of core RNAi pathway components from *C. elegans*, against predicted
104 protein sets and contigs of the *S. carpocapsae* genome, through the Wormbase Parasite
105 BLAST server (Howe et al., 2016; Altschul et al., 1990).

106

107 **dsRNA synthesis**

108 *Sc-flp-21* (Gene ID: L596_g19959.t1) dsRNA templates were generated from *S.*
109 *carpocapsae* IJ cDNA using gene-specific primers with T7 recognition sites (see Table 1).
110 Neomycin phosphotransferase (*neo*) and Green Fluorescent Protein (*gfp*) dsRNA templates
111 were generated from pEGFP-N1 (GenBank: U55762.1). Template PCR products were
112 generated as follows: [95 °C x 10 min, 40 x (95 °C x 30 s, 60 °C x 30 s, 72 °C x 30 s) 72 °C x
113 10 min]. PCR products were assessed by gel electrophoresis, and cleaned using the
114 Chargeswitch PCR clean-up kit (Life Technologies). dsRNA was synthesised using the T7
115 RiboMAX™ Express Large Scale RNA Production System (Promega), and quantified by a
116 Nanodrop 1000 spectrophotometer.

117

118 **RNAi**

119 1000 *S. carpocapsae* were incubated in 50 µl PBS with dsRNA and 50 mM serotonin across
120 four experimental regimes; (i) 24 h in 5 mg/ml dsRNA / serotonin / PBS; (ii) 24 h in 5 mg/ml
121 dsRNA / serotonin / PBS, followed by washes to remove the initial dsRNA, and 24 h
122 recovery in PBS only; (iii) 48 h in 5 mg/ml dsRNA / serotonin / PBS; and (iv) 48 h in 1 mg/ml
123 dsRNA and serotonin. Each experiment was conducted at 23 °C. We found that 50 mM
124 serotonin induced oral uptake of fluorescent dyes under all conditions tested; 50 mM
125 octopamine did not (data not shown).

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127 **RNA extraction, cDNA synthesis and quantitative (q)RT-PCR**

128 Total RNA was extracted from 1000 IJs using the Simply RNA extraction kit (Promega, UK)
129 and Maxwell 16 extraction system (Promega, UK). cDNA was synthesised using the High
130 Capacity RNA to cDNA kit (Applied Biosystems, UK). Each individual qRT-PCR reaction
131 comprised 5 µl Faststart SYBR Green mastermix (Roche Applied Science), 1 µl each of the
132 forward and reverse primers (10 µM), 1 µl water, 2 µl cDNA. PCR reactions were conducted
133 in triplicate for each individual cDNA using a Rotorgene Q thermal cycler under the following
134 conditions: [95 °C x 10 min, 40 x (95 °C x 20 s, 60 °C x 20 s, 72 °C x 20 s) 72 °C x 10 min].

135 Primer sets were optimised for working concentration, annealing temperature and analysed
136 by dissociation curve for contamination or non-specific amplification by primer–dimer as
137 standard. The PCR efficiency of each specific amplicon was calculated using the Rotorgene
138 Q software. Relative quantification of target transcript relative to two endogenous control
139 genes (*Sc-act* and *Sc-β-tubulin*) was calculated by the augmented $\Delta\Delta\text{Ct}$ method (Pfaffl,
140 2001), relative to the geometric mean of endogenous references (Vandesompele et al.,
141 2002). One way ANOVA and Fisher's LSD test were used to analyse data (GraphPad Prism
142 6). The most similar non-target gene (*L596_g5821.t1*) was identified using BLASTn against
143 the *S. carpocapsae* genomic contigs (supplemental figure S1), and primers *Sc-*
144 *L596_g5821.t1-f* and *Sc- L596_g5821.t1-r* were used to assess transcript abundance
145 relative to *Sc-act* across control and experimental conditions for the 48h dsRNA exposure
146 experiments only (Table 1).

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163 **Table 1.** dsRNA synthesis and qRT-PCR primer sequences.

Primer designation	Sequence (5' – 3')
Sc-flp-21-F	TTCTGAGCCGCTATCTGAGC
Sc-flp-21-Ft7	TAATACGACTCACTATAGGTTCTGAGCCGCTATCTGAGC
Sc-flp-21-R	AGTCGCAGGGAACAAACAAT
Sc-flp-21-Rt7	TAATACGACTCACTATAGGAGTCGCAGGGAACAAACAAT
neo-F	GGTGGAGAGGCTATTCGGCTA
neo-Ft7	TAATACGACTCACTATAGGGGTGGAGAGGCTATTCGGCTA
neo-R	CCTTCCCGCTTCAGTGACAA
neo-Rt7	TAATACGACTCACTATAGGCCTTCCCGCTTCAGTGACAA
gfp-F	GGCATCGACTTCAAGGAGGA
gfp-Ft7	TAATACGACTCACTATAGGGGCATCGACTTCAAGGAGGA
gfp-R	GTAGTGGTTGTCGGGCAGCA
gfp-Rt7	TAATACGACTCACTATAGGGTAGTGGTTGTCGGGCAGCA
Sc-act-Fq	ATGTTCCAGCCCTCTTTCT
Sc-act-Rq	GATGTCGCACTTCATGATCG
Sc- β tub-Fq	CTCGGAGGAGGAGATGACAG
Sc- β tub-Rq	ATCATAACGGCACGAGGAAC
Sc-flp-21-Fq	GCTGCCTTCTCGTACTCTTC
Sc-flp-21-Rq	TCAGATAGCGGCTCAGAAGC
Sc-L596_g5821.t1-Fq	GTGGGAAATCCGACACAAA
Sc-L596_g5821.t1-Rq	GTCACGTCGTCCACTATAAAC

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169 **Headspace Solid-Phase MicroExtraction (SPME) GC-MS**

170 Approximately 5 g of fresh waxworm (*Galleria mellonella*) and mealworm (*Tenebrio molitor*)
171 larvae were placed into 20 mL glass tubes and sealed. The holder needle was exposed to
172 the headspace of the tube over a 120 min timecourse (extraction time) at room temperature
173 (22 °C). After this time, the SPME syringe was directly desorbed in the GC injection port for
174 5 min. A fused silica fibre coated with a 95 µm layer of carboxen–polydimethylsiloxane
175 (CAR–PDMS; Supelco) was used to extract the volatile compounds from the samples.
176 Fibres were immediately thermally desorbed in the GC injector for 5 min (with this time we
177 desorb the analytes and re-activated the fiber for the next analysis) at 250 °C and the
178 compounds were analysed by GC-MS.

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180 A CTC Analytics CombiPal autosampler was coupled to a 7890N Agilent gas chromatograph
181 (Agilent, Palo Alto, California) and connected to a 5975C MSD mass spectrometer. The
182 manual SPME holder (Supelco, Bellefonte, PA, USA) was used to perform the experiments.
183 Chromatographic separation was carried out on 30 m x 0.25 mm I.D. ZB-semivolatiles,
184 Zebron column (Phenomenex, Macclesfield, UK). The oven temperature was set at 40 °C
185 for 3 min, temperature increased from 40 to 250 °C at 5 °C min⁻¹ and set at the maximum
186 temperature for 4 min. Helium was used as carrier gas at 1 ml min⁻¹. Mass spectra were
187 recorded in electron impact (EI) mode at 70 eV. Scan mode was used for the acquisition to
188 get all the volatile compounds sampled. Quadrupole and source temperature were set at 150
189 and 230 °C respectively. Compounds were identified using MS data from the NIST library
190 (>95% confidence).

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192 **Dispersal assays**

193 100 *S. carpocapsae* IJs were placed in the centre of a 90 mm PBS agar plate (1.5 % w/v) in
194 a 5 µl aliquot of PBS. Plates were divided into four zones; a central zone 15 mm in
195 diameter, and three further zones equally spaced over the remainder of the plate. Plates
196 were allowed to air dry for ~5 min. Evaporation of the PBS allowed the IJs to begin

197 movement over the agar surface. Lids were then placed back onto the Petri dishes, and
198 plates were incubated at 23 °C in darkness for one hour. IJs were counted across central
199 and peripheral zones and expressed as percentage of total worms. Our subsequent
200 analysis was conducted on total IJs found within the two central zones. Relative to those
201 found in the two peripheral zones.

202

203 **Nictation & jumping assays**

204 3.5 g of compost (John Innes No.2) was placed in a petri dish (55 mm), and dampened
205 evenly with 150 µl PBS. Approximately ten IJs were pipetted onto the compost in 5 µl of
206 PBS, and left for 5 minutes at room temperature; this enabled IJs to begin nictating. For the
207 waxworm volatile challenge, one healthy waxworm (UK Waxworms Ltd.) was placed inside a
208 1 mL pipette tip, without filter. For the mealworm volatile challenge, two mealworms
209 (Monkfield Nutrition, UK), weight-matched to the waxworm, were placed inside a 1 mL
210 pipette tip, without filter. Blank exposure data were captured using an empty 1 ml pipette tip,
211 without filter. In each case, the pipette was set to eject a volume of 500 µL, comprising air
212 and the corresponding insect volatiles. A binocular microscope was used to record IJ
213 behavioural responses following up to five volatile exposures each, on gentle ejection from
214 the pipette within a distance of ~1 cm of the *S. carpocapsae* IJs. A five second period was
215 allowed between each volatile exposure. Recording ended for any individual when jumping
216 was observed or the IJ abandoned a nictating stance (this always corresponded with
217 migration away from the stimulus). A jumping index was calculated for each treatment group
218 (Dillman et al., 2012). Additional behavioural observations were recorded, and subsequently
219 reported as percentage IJs displaying the behaviour over the course of up to five volatile
220 exposures, or until the IJ migrated / jumped out of the field of vision.

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222 **Immunocytochemistry**

223 Freshly emerged *S. carpocapsae* IJs were fixed in 4 % paraformaldehyde overnight at 4 °C,
224 followed by a brief wash in antibody diluent (AbD; 0.1 % bovine serum albumin, 0.1 %

225 sodium azide, 0.1 % Triton-X-100 and PBS pH 7.4). The fixed specimens were roughly
226 chopped on a glass microscope slide with a flat edged razor, and incubated in primary
227 polyclonal antiserum raised against GLGPRPLRFamide, N-terminally coupled to KLH, and
228 affinity purified (1:800 dilution in AbD) for 72 h at 4 °C. Subsequently, chopped IJs were
229 washed in AbD for 24 h at 4 °C, and then incubated in secondary antibody conjugated to
230 fluorescein isothiocyanate (1:100 dilution in AbD) for 72 h at 4 °C. A further AbD wash for
231 24 h at 4 °C was followed by incubation in Phalloidin–Tetramethylrhodamine B
232 isothiocyanate (1:100 dilution in AbD) for 24 h at 4 °C. Finally, chopped IJs were washed in
233 AbD for 24 h at 4 °C. Specimens were mounted onto a glass slide with Vectasheild
234 mounting medium and viewed with a Leica TCS SP5 confocal scanning laser microscope.
235 Controls included the omission of primary antiserum, and pre-adsorption of the primary
236 antiserum with ≥ 250 ng of GLGPRPLRFamide. Pre-adsorption in GLGPRPLRFamide did not
237 alter staining patterns.

238

239 **Statistical analysis**

240 Data pertaining to both qRT-PCR and behavioural assays were assessed by Brown-
241 Forsythe and Bartlett's tests to examine homogeneity of variance between groups. Were
242 variance differed statistically between groups a Kruskal-Wallis test was followed by Dunn's
243 multiple comparison test. In all other instances, One-way or two-way ANOVA was followed
244 by Fisher's Least Significant Difference (LSD) test. All statistical tests were performed using
245 GraphPad Prism 6.

246

247 **Results**

248 **The RNAi pathway of *S. carpocapsae***

249 As is the case for other parasitic nematode species, *S. carpocapsae* was found to encode a
250 less diverse RNAi pathway than that of *C. elegans*, in terms of gene for gene conservation

251 (Dalzell et al., 2011). However, the apparent reduction in AGO homologue diversity is offset
252 by significant expansions across several putative *ago* genes, to give a predicted overall
253 increase in the *S. carpocapsae* AGO complement (38 in total), relative to *C. elegans* (24, not
254 including pseudogenes) (Vasquez-Rifo et al., 2012); WAGO-1 (19), ALG-1 (three), ALG-3
255 (two), WAGO-5 (four), WAGO-10 (two), WAGO-11 (three) are all expanded relative to *C.*
256 *elegans*. Notably, no identifiable homologue of RDE-1, the primary AGO for exogenously
257 triggered RNAi events in *C. elegans*, could be identified (Supplementary table S2).

258

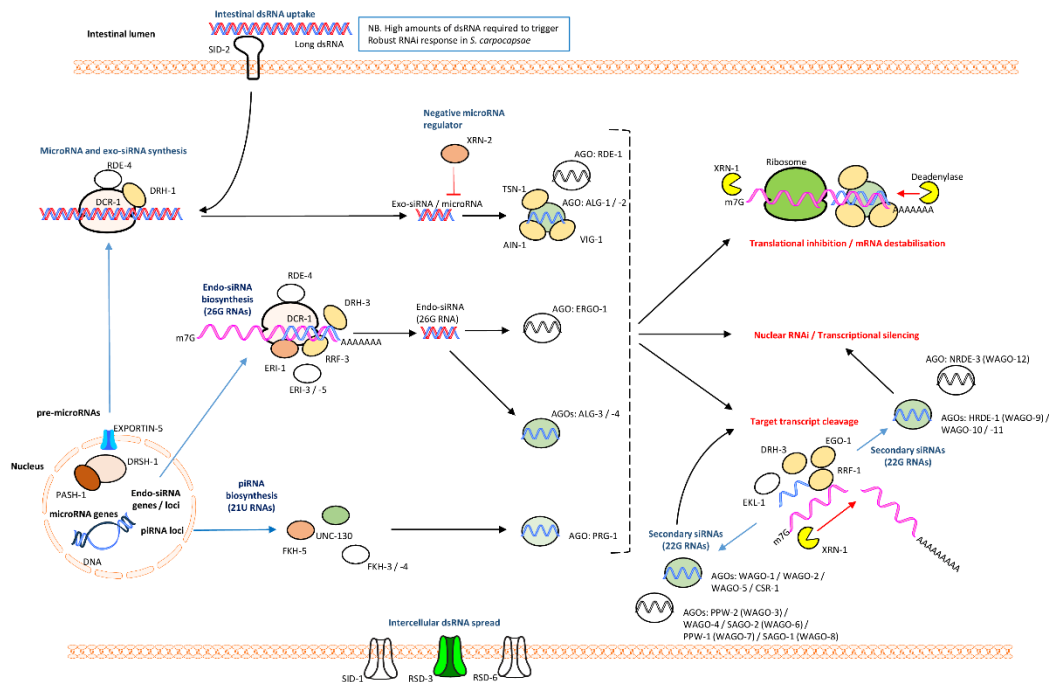
259 The presence of PRG-1 and components of the piwi interacting (pi)RNA biosynthetic
260 machinery suggests that a functional piRNA (or 21U RNA) pathway may be present. Whilst
261 ERGO-1 is not conserved, two putative ALG-3 orthologues suggest that a functional endo-
262 siRNA (26G RNA) pathway may also exist, which is supported by broad conservation of
263 associated proteins. MicroRNA-associated AGOs, ALG-1 and ALG-2 are conserved, with a
264 small apparent expansion of ALG-1 to three related proteins in *S. carpocapsae*. Further
265 understanding of how RNAi pathway complements influence functionality will require small
266 RNA sequencing efforts, and functional genomics approaches.

267

268 The RNA-dependent RNA Polymerase (RdRp), RRF-3 is conserved, and known to function
269 antagonistically to exogenously primed RNAi, through competing activity for pathway
270 components required for both exogenous RNAi, and the endo-siRNA (26G RNA) pathway
271 within which RRF-3 operates (Simmer et al., 2002; Sijen et al., 2001; reviewed by Billi et al.,
272 2014). The RdRps, RRF-1 and EGO-1, which are involved in the biosynthesis of secondary
273 siRNAs (22G RNAs) are also conserved. Loss of the argonaute ERGO-1 which functions
274 upstream of secondary siRNA biogenesis in the endo-siRNA (26G RNA) pathway in *C.*
275 *elegans*, also leads to an exogenous ERI phenotype (Enhanced RNAi), but is not conserved
276 in *S. carpocapsae*, suggesting that ALG-3 / -4 may be solely responsible for endo-siRNA
277 functionality (Conine et al., 2010; Han et al., 2009).

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279 The apparent absence of the intestinal dsRNA transporter, SID-2 is consistent with findings
 280 from other parasitic nematodes (Dalzell et al., 2011; Geldhof et al., 2007; Knox et al., 2005).
 281 SID-1 also appears to be absent, however CHUP-1, a putative cholesterol uptake protein
 282 which contains a SID-1 RNA channel is present, and may assist in the intercellular spread of
 283 dsRNA. RSD-3, which also effects intercellular spread of dsRNA is conserved (see Figure 1
 284 for pathway overview).
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286

287 **Figure 1. Core RNAi pathway components of *S. carpocapsae* relative to *C. elegans*.**

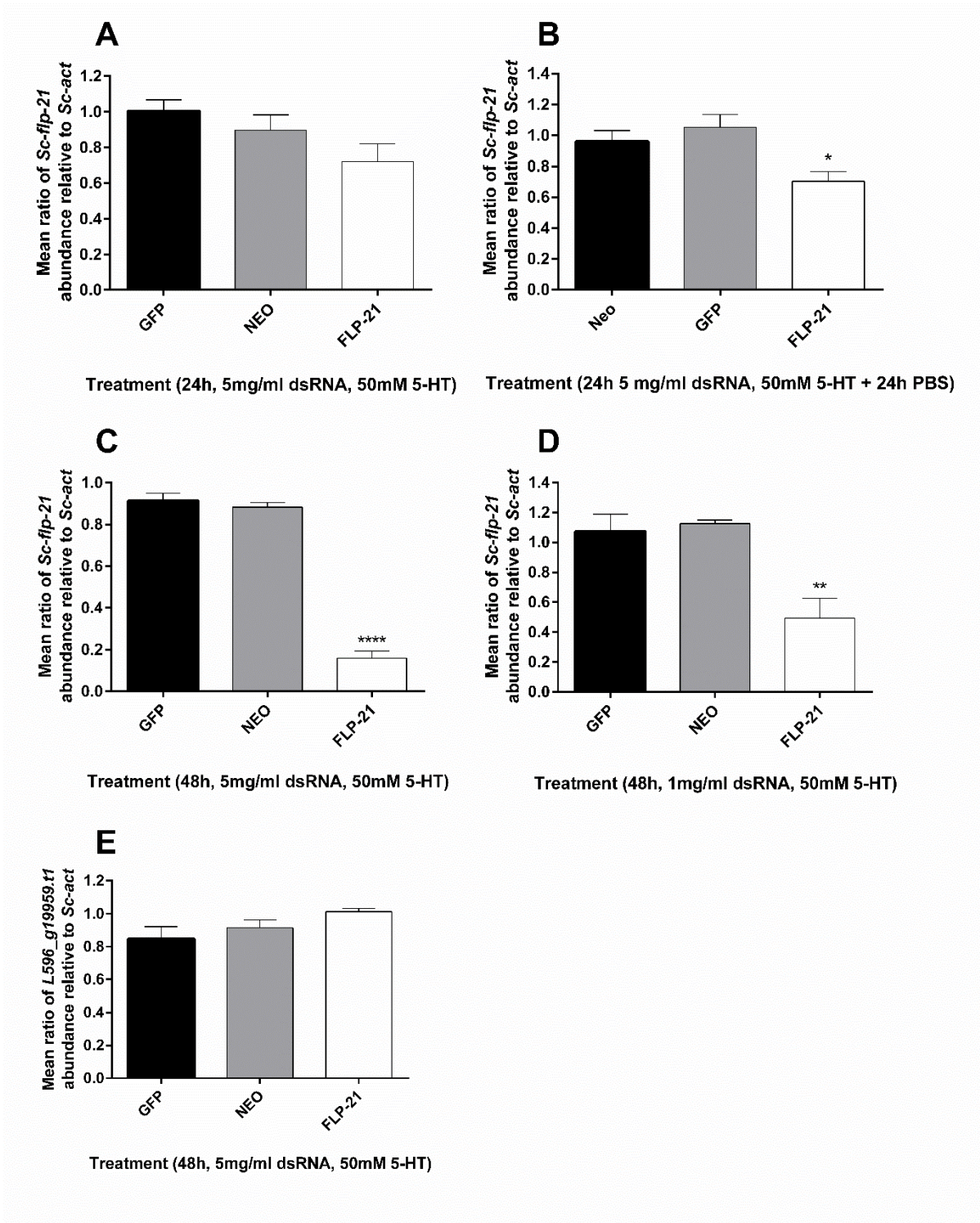
288 Proteins with at least one putative homologue are represented in colour; those without any
 289 identifiable homologues are colourless. See supplemental tables for information on number
 290 of putative related homologues.

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292 **Knockdown of *Sc-flp-21***

293 Various treatment regimens were employed in order to assess the responsiveness of *S.*
 294 *carpocapsae* IJs to exogenous dsRNA. 24h incubation in 5 mg/ml dsRNA, with 50 mM
 295 serotonin was not sufficient to trigger statistically significant *Sc-flp-21* knockdown (Fig. 2A),
 296 however a 24h dsRNA / serotonin incubation followed by a 24h recovery in PBS only, did
 297 trigger a small decrease in *Sc-flp-21* relative to *Sc-act* when compared to *gfp* and *neo*

298 dsRNA controls (0.70 ± 0.11 , $P < 0.05$) (Fig. 2B). Extended incubation of *S. carpocapsae* IJs
299 in 5 mg/ml dsRNA and 50 mM serotonin for 48 h triggered robust knockdown of *Sc-flp-21*
300 (0.16 ± 0.07 , $P < 0.0001$) (Fig. 2C). 48 h incubation in 1 mg/ml dsRNA, with 50 mM serotonin
301 also triggered significant levels of *Sc-flp-21* knockdown (0.49 ± 0.27 , $P < 0.01$), however this
302 was not as effective as the 5 mg/ml dsRNA treatment (Fig. 2D). A BLAST analysis identified
303 predicted *S. carpocapsae* transcript *L596_g5821.t* as the non-target gene with most
304 similarity to the *Sc-flp-21* dsRNA (supplemental figure S1). The relative expression level of
305 *L596_g5821.t1* was unaffected by a 48 h incubation in 5 mg/ml *Sc-flp-21* dsRNA with 50 mM
306 serotonin, relative to *neo* and *gfp* dsRNA (1.013 ± 0.04 , $P > 0.05$) (Fig. 2E).



307

308 **Figure 2. qRT-PCR expression analysis of *Sc-flp-21* and off-target control gene (A) *Sc-***
309 ***flp-21* transcript ratio relative to *Sc-act* following 24 h incubation in 5 mg/ml dsRNA, 50 mM**
310 **serotonin. (B) *Sc-flp-21* transcript ratio relative to *Sc-act* following 24 h incubation in 5 mg/ml**
311 **dsRNA, 50 mM serotonin, and 24 h in PBS; (C) *Sc-flp-21* transcript ratio relative to *Sc-act***

312 following 48 h incubation in 5 mg/ml dsRNA, 50 mM serotonin; (D) *Sc-flp-21* transcript ratio
313 relative to *Sc-act* following 48 h incubation in a reduced 1 mg/ml dsRNA, 50 mM serotonin;
314 (E) *L596_g5821.t1* transcript ratio relative to *Sc-act* following 48 h incubation in 5 mg/ml
315 dsRNA, 50 mM serotonin. *P<0.05; **P<0.01; ****P<0.0001.

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317 **Host insect volatiles**

318 Comprehensive volatile signatures were characterised, and significant differences noted
319 between *G. mellonella* and *T. molitor* larvae. In total, we identified 10 compounds unique to
320 *G. mellonella*, four compounds unique to *T. molitor*, and 14 compounds shared between
321 both species. These profiles vary significantly from headspace GC-MS data presented by
322 Hallem et al. (2011) for the same insect species (see Table 1).

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339 **Table 1. Headspace SPME GC-MS volatile profiles of *Galleria mellonella* and *Tenebrio***
 340 ***molitor* larvae**

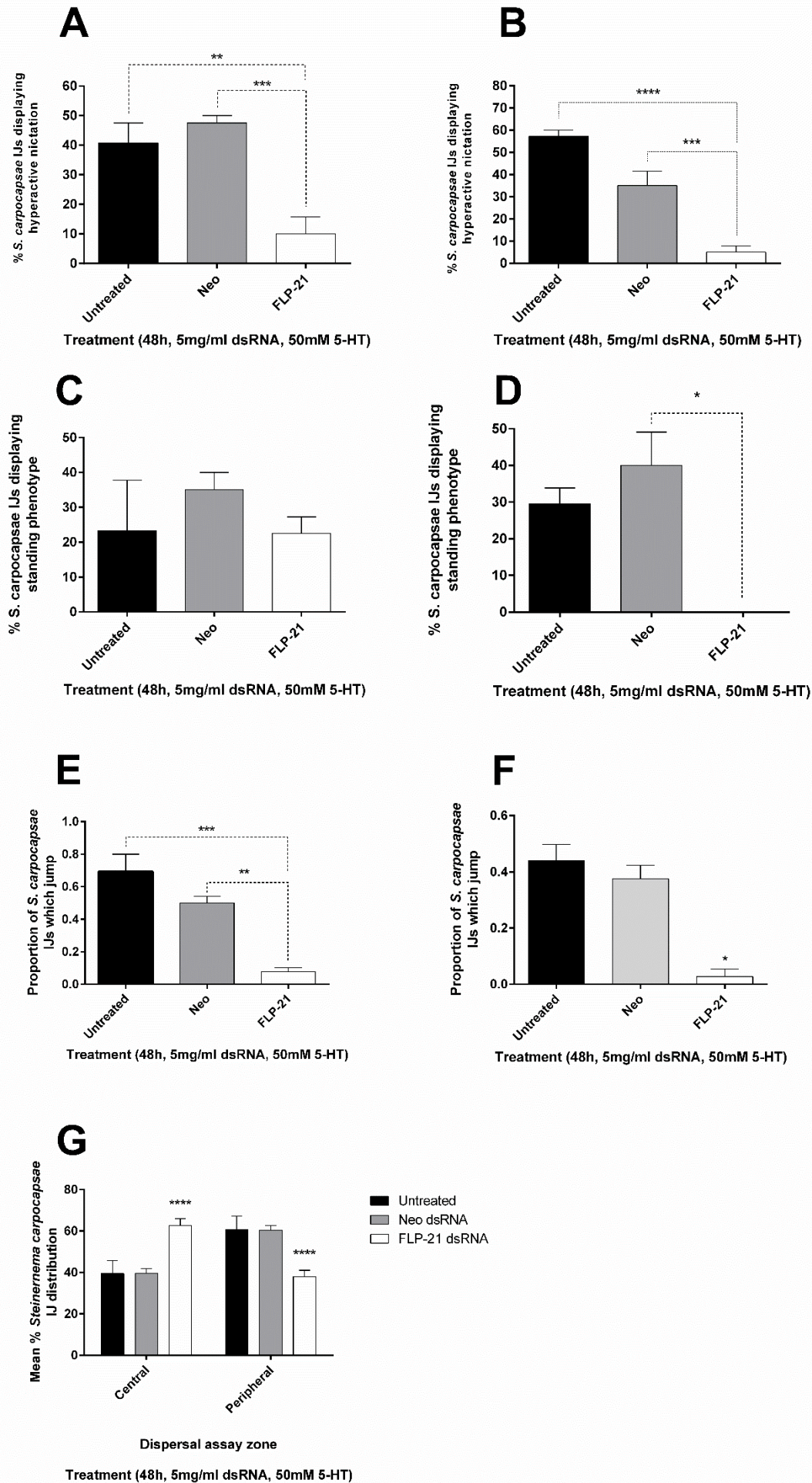
<i>Galleria mellonella</i>	<i>Tenebrio molitor</i>	Shared
α-pinene	decanal	nonanal
butanoic acid	1-tetradecanol-methyl ether	hexanoic acid
tetradecanal	1-hexadecanol	octanoic acid
tridecanol	cyclohexadecane	cyclododecane
nonanoic acid		decanoic acid
2-[phenyl methylene]-octanal		dodecanoic acid
tetracosanol		tetradecanoic acid
tridecane		pentadecanoic acid
1-heptacosanol		bis(2-ethylhexyl) ester-hexanedioic acid
cis-9-hexadecenoic acid		hexadecanoic acid
		heptadecanoic acid
		cis-10-heptadecenoic acid
		squalene
		octadecanoic acid

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342 **Behavioural impact of *Sc-flp-21* knockdown**

343 *S. carpocapsae* IJs were challenged by exposure to volatiles from *G. mellonella* or *T. molitor*
 344 following RNAi (48h 5 mg/ml dsRNA, 50 mM serotonin) and control treatments. A decrease
 345 in hyperactive nictation following *Sc-flp-21* knockdown was observed (10% ±5.774) relative
 346 to untreated (40.75% ±6.75; P<0.01) and *neo* dsRNA treatment (47.5% ±2.5; P<0.001)
 347 following *G. mellonella* volatile challenge (Fig. 3A). Likewise, a decrease in hyperactive
 348 nictation was observed following *T. molitor* volatile challenge to *Sc-flp-21* RNAi IJs (5.0%
 349 ±2.9), relative to untreated (57.25% ±2.8; P<0.0001) and *neo* dsRNA treatment (35.0% ±6.5;
 350 P<0.001) (Fig. 3B). A decrease in the jumping index of IJs following *Sc.flp-21* dsRNA

351 treatment was observed when challenged by *G. mellonella* volatiles (0.08 ± 0.03) relative to
352 untreated (0.69 ± 0.11 ; $P < 0.001$) and *neo* dsRNA treated (0.50 ± 0.04 ; $P < 0.01$) (Fig. 3E) .
353 Similarly, a decrease in jumping index as a response to *T. molitor* volatiles was observed
354 following *Sc-flp-21* RNAi (0.03 ± 0.03) relative to untreated (0.44 ± 0.06 ; $P < 0.001$) and *neo*
355 dsRNA treatment (0.38 ± 0.05 ; $P < 0.001$) (Fig 3F). However, it appears that *Sc-flp-21*
356 knockdown does result in a discrimination between *G. mellonella* and *T. molitor* volatiles in
357 triggering standing phenotypes (as described by Campbell and Gaugler, 1993; Ishibashi and
358 Kondo, 1990). No statistically significant difference in the percentage of IJs displaying a
359 standing phenotype is observed in response to *G. mellonella* volatiles between experimental
360 groups, however a significant decrease in standing phenotypes was observed in response to
361 *Sc-flp-21* knockdown following exposure to *T. molitor* volatiles (0.00%) relative to *neo*
362 dsRNA treated ($40.0\% \pm 9.13$; $P < 0.05$) but not untreated ($29.5\% \pm 4.4$; $P > 0.05$) using
363 Kruskal-Wallis and Dunn's multiple comparison tests. Undoubtedly, the significant difference
364 in variance across experimental groups in this instance has reduced statistical power (Fig.
365 3D). It was also found that *Sc-flp-21* RNAi resulted in significantly decreased lateral
366 dispersal, relative to both untreated and *neo* dsRNA treatment ($P < 0.0001$) (Fig. 3G). In all
367 instances, dsRNA treatment regimens which triggered lower levels of *Sc-flp-21* knockdown
368 relative to the 48h 5 mg/ml dsRNA, 50 mM serotonin approach, failed to trigger null
369 phenotypes (data not shown).



371 **Figure 3. *S. carpocapsae* IJ behavioural assays post-RNAi**

372 Behavioural impact of *Sc-flp-21* knockdown following IJ incubation in 5 mg/ml dsRNA, 50
373 mM serotonin: (A) Mean percentage of *S. carpocapsae* displaying hyperactive nictation upon
374 challenge by *G. mellonella* volatiles. (B) Mean percentage of *S. carpocapsae* displaying
375 hyperactive nictation upon challenge by *T. molitor* volatiles. (C) Mean percentage of *S.*
376 *carpocapsae* displaying standing upon challenge by *G. mellonella* volatiles. (D) Mean
377 percentage of *S. carpocapsae* displaying standing upon challenge by *T. molitor* volatiles. (E)
378 Mean jumping index of *S. carpocapsae* displaying upon challenge by *G. mellonella* volatiles.
379 (F) Mean jumping index of *S. carpocapsae* upon challenge by *T. molitor* volatiles. (G) Mean
380 percentage distribution of *S. carpocapsae* IJs across central and peripheral assay zones.

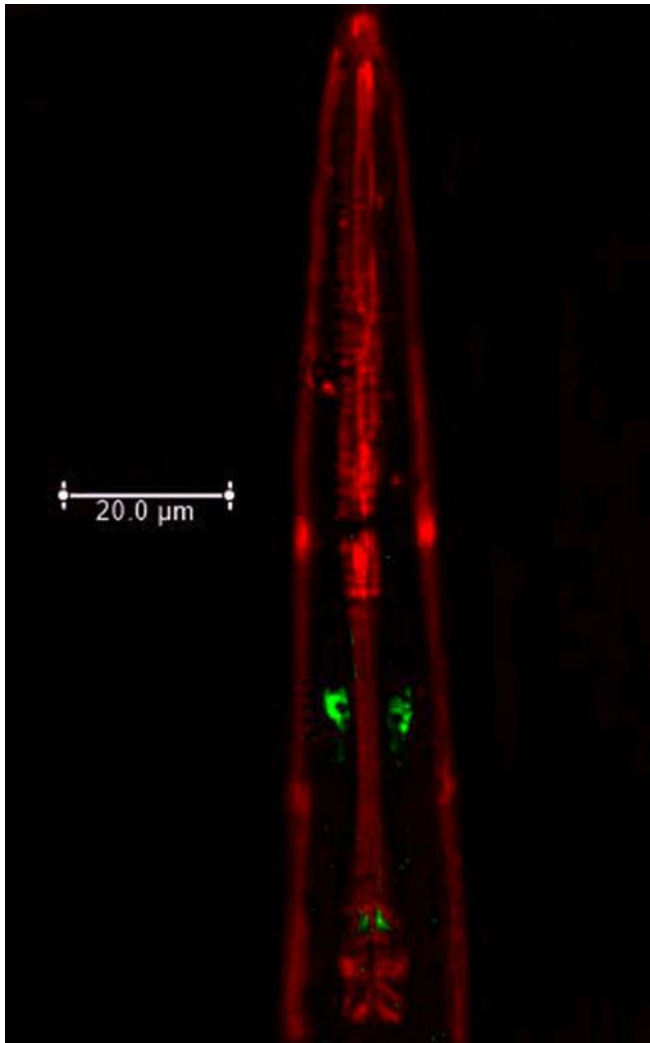
381 *P<0.05; **P<0.01; ****P<0.0001.

382

383 **Immunocytochemical localisation of FLP-21 in *S. carpocapsae* IJs**

384 FLP-21 was localised to paired neurons within the central nerve ring region of *S.*
385 *carpocapsae* IJs. Without additional neuroanatomical information on *S. carpocapsae* IJs it is
386 impossible to define these cells, however they do appear to project posteriorly, terminating in
387 synapses around the region of the pharyngeal metacarpal bulb (Fig. 4).

388



389

390 **Figure 4. Immunocytochemical localisation of FLP-21 (GLGPRPLRFamide) to paired**
391 **neurons within the central nerve ring of *S. carpocapsae* IJs.** Positive FLP-21
392 immunostaining is visible as green, muscle is counterstained red.

393

394 Discussion

395 RNA interference is an extremely important tool for the study of gene function in parasitic
396 nematodes (Dalzell et al., 2012; Maule et al., 2011). Three independent reports of a
397 functional RNAi pathway in the entomopathogenic nematode *Heterorhabditis bacteriophora*
398 have been published. Ciche and Sternberg (2007) assessed the efficacy of RNAi through
399 soaking egg / L1 stage *H. bacteriophora* in 5-7.5 mg/ml dsRNA targeting a number of genes
400 which had been selected on the basis of phenotypic impact on the model *C. elegans*.

401 Demonstrable phenotypes and target transcript knockdown signified an active pathway.
402 Moshayov, Koltai and Glazer (2013) employed the methodology of Ciche and Sternberg
403 (2007) to study the involvement of genes in the regulation of IJ exsheathment (or 'recovery').
404 Subsequently, Ratnappan et al. (2016) demonstrated that microinjection was also a suitable
405 method for introducing dsRNA into hermaphrodite gonads, effectively triggering the RNAi
406 pathway in F1 progeny. To date, no such assessment of a functional RNAi pathway has
407 been published for *Steinernema* spp.

408

409 The RNAi pathway of *S. carpocapsae* has been characterised by BLAST and validated
410 through silencing *Sc-flp-21* in IJs. Our data indicate that neuronal cells are sensitive to RNAi
411 in *S. carpocapsae* IJs, and that knockdown is highly sequence specific. Like other parasitic
412 nematodes *S. carpocapsae* encodes an expanded set of WAGO-1 (R06C7.1) family AGOs
413 (19 in total) which function primarily with secondary siRNAs (22G RNAs) in *C. elegans*,
414 along with CSR-1 which is also conserved. Whilst RDE-1 is primarily responsible for
415 triggering the onset of an exogenous RNAi response, acting upstream of secondary siRNAs
416 (22G RNAs), it is not conserved in *S. carpocapsae* (Billi et al., 2014; Dalzell et al., 2011).
417 Our observation of RNAi sensitivity in *S. carpocapsae* reveals that RDE-1 is not required to
418 trigger an exogenous RNAi response, however the functional significance of AGO
419 homologue expansions relative to *C. elegans* remains to be determined. The lack of SID-2
420 seems to correlate with our observation that relatively high amounts of dsRNA are required
421 to trigger the RNAi pathway by oral delivery.

422

423 The nearest non-target gene sequence within the *S. carpocapsae* genome represents an
424 uncharacterised predicted gene (*L596_g5821.t1*). The *Sc-flp-21* dsRNA shared high levels
425 of sequence similarity over a 21 bp stretch of *L596_g5821.t1* (20 of 21 bp shared), however
426 qRT-PCR indicates that *L596_g5821.t1* had not been silenced, which could suggest: (i) the
427 level of sequence similarity was either insufficient for gene knockdown; (ii) dsRNA was not
428 diced in the correct register to produce this exact 21 bp sequence within a significant

429 population of siRNAs; or (iii) the *L596_g5821.t1* gene is not expressed in cells / tissue which
430 is sufficiently susceptible to dsRNA delivered under the conditions tested. In order to trigger
431 significant knockdown of *Sc-flp-21*, 48h continuous exposure to dsRNA was required in the
432 presence of 50 mM serotonin. Reducing dsRNA exposure time lead to a corresponding
433 reduction in *Sc-flp-21* knockdown, as did a reduction of dsRNA amount from 5mg/ml to
434 1mg/ml over a 48h time-course. Phenotypes which developed following 48h dsRNA
435 exposure were not observed across any of the experimental variations which resulted in
436 decreased gene knockdown (shorter exposure timeframes / lower dsRNA amounts).

437

438 The neuronal RNAi sensitivity of *S. carpocapsae* IJs, and the ease of behavioural assays
439 makes these species ideal models for studying the neurobiology of nictation. Within the
440 Steinernematid EPNs, a number of species also display a highly specialised jumping
441 behaviour which can be triggered in nictating IJs on exposure to host Insect volatiles
442 (Castelletto et al., 2014). Silencing *Sc-flp-21* triggers pleiotropic effects on lateral dispersal,
443 hyperactive nictation and jumping phenotypes. The frequency of standing phenotypes
444 which is observed of nictating IJs which are stimulated by host volatile cues was found to
445 differ between *flp-21*-silenced IJs exposed to mealworm volatiles, but not waxworm volatiles
446 (Fig. 3C-D). Specifically, the standing phenotype is enacted by IJs which adopt a temporary,
447 upright torpid stance. The mechanistic significance of torpid standing nictation relative to
448 hyperactive waving nictation is not clear, however we observe that both can be triggered by
449 insect volatiles under the conditions tested here. The waxworm and mealworm headspace
450 SPME GC-MS profiles are expanded relative to those presented by Hallem et al. (2011), and
451 likely reflects the increased sensitivity of analysis in this study. These data could provide a
452 valuable tool for comparative analysis of neurobiology and host-finding behaviours across
453 EPN species.

454

455 Given the redundancy of FLP-21 and FLP-18 in activating NPR-1 (and other NPRs), further
456 work is required to understand the neuronal basis of nictation and jumping behaviours in *S.*

457 *carpocapsae* and other EPN species. Of particular interest is the function of FLP-21 in EPN
458 species which do not nictate or jump, such as *Steinernema glaseri* and *H. bacteriophora*.
459 Likewise, it is not yet known if, or how FLP-21 signalling is differentially regulated in
460 *Steinernema feltiae* which displays a less pronounced nictation behaviour involving partial
461 lifting of the IJ anterior from the substrate surface (Campbell and Gaugler, 1993).
462
463 Collectively, these data provide the first mechanistic insight to a behaviour which may have
464 implications for biocontrol efficacy. Through isolating genes and signalling pathways which
465 coordinate these behaviours, efforts to identify molecular markers of desired behaviours and
466 traits could facilitate the identification of more suitable isolates and strains for biocontrol use,
467 and the enhancement of current strains through selective breeding approaches. The
468 selection or manipulation of behavioural tendencies could lead to strains which are capable
469 of operating within new ecological niches, expanding their utility.

470

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650 Supplemental Data

651 Figure S1. Alignment of *Sc-flp-21* dsRNA against most similar non-target *S.* 652 *carpocapsae* gene (McWilliam et al., 2013).

```
653 CLUSTAL O(1.2.1) multiple sequence alignment
654
655
656
657 exon_14_L596g5821      CAAATTCAAGAGCGCAGCAGGGATAAAGGCTTGTAGCTGGCCGCAAAGCCATCAGACGC
658 Sc-flp-21_dsRNA      -----
659
660
661 exon_14_L596g5821      TAATTGCGCCAGAACTCGGTGACGCTAGTATTTCGGCGATCTTGCAAATGCATTGGACAAG
662 Sc-flp-21_dsRNA      -----
663
664
665 exon_14_L596g5821      TTTCTCCGTCAGGTCAGCCCTGAGAACTGCATCGCTGACATGCATCCTTTCCAGATGGC
666 Sc-flp-21_dsRNA      -----
667
668
669 exon_14_L596g5821      CAAGTATTGATCGATGATGTGGCAAACATCTACATAAGAAGTGTGCCGAATCTCCGA
670 Sc-flp-21_dsRNA      -----TTCTG
671                                     * * .
672
673 exon_14_L596g5821      AGACGCGATTTGGATTGTGGCGTCGAGTTGGCAAGCCAGTTCACGCGCCCGAACG--CC
674 Sc-flp-21_dsRNA      AGCCGCT-----ATCTGAGCCAGTTCACGCGCCCGAATACGAC
675 **.*.*                : .*****.*****.*.*
676
677 exon_14_L596g5821      TTCAGCTTCGCCGTGTCCATCCA---CTTCACG-----ACGACTCGGTCGAAAATCT
678 Sc-flp-21_dsRNA      CCCAGCGCTACATGTACTTCGATCAGCGCTCGATGAAGCGAGGCTCGGTCCCTC-----
679          **** * .*.***.*:* *      *:*      * *.***** :.
680
681 exon_14_L596g5821      GGCTCCAGTCGGATTTGTCCAGCCGGTATTTCTGCTTTGGAAGACGGTACTGACGAAATT
682 Sc-flp-21_dsRNA      -----GACCTCTCCGCTTTGGTTAACTGCTA-----GAAAT
683          *. .* ** *****:.* * :.      .**:*
684
685 exon_14_L596g5821      CTGCGCTGCCAGATGTACCTTCAGAATCGTGATCAAAAGTTCTTCATTGTGCGCTTCGC
686 Sc-flp-21_dsRNA      CGTGATATTTCAGAATTCTC-----CTCTTTGATCTCTTTGATTGTTTGTC
687 * *. . * *****: * . *      .***:::*** :* . * * *
688
689 exon_14_L596g5821      TCTTCGACGCGCTCATCAT
690 Sc-flp-21_dsRNA      CCTGCGACT-----
691          ** ****
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704 **Table S2. *Steinernema carpocapsae* argonaute proteins.**

<u>AGOs Protein List</u>	<u>Homologues</u>	<u>Accession *</u>
ALG-1	x2	L596_g7718.t1
ALG-2	x1	L596_g16709.t1
ALG-3 (T22B3.2)	x2	L596_g3457.t1
ALG-4		
CSR-1	x1	L596_g20174.t1
C04F12.1	x1	L596_g11107.t1
C14B1.7a		
ERGO-1		
HPO-24		
HRDE-1 (WAGO-9)	x1	L596_g11197.t1
NRDE-3 (WAGO-12)		
PPW-1 (WAGO-7)		
PPW-2 (WAGO-3)		
PRG-1	x1	L596_g25491.t1
RDE-1		
SAGO-2 (WAGO-6)		
SAGO-1 (WAGO-8)		
T23B3.2	x1	L596_g21112.t1
WAGO-1 (R06C7.1)	x19	L596_g19943.t1
WAGO-10 (T22H9.3)	x2	L596_g19923.t1
WAGO-11 (Y49F6A.1)	x3	L596_g17524.t1
WAGO-2 (F55A12.1)	x1	L596_g16917.t1
WAGO-4 (F58G1.1)		
WAGO-5 (ZK1248.7)	x4	L596_g12936.t1

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706 * Accession numbers presented are the top returning hits for each RNAi pathway
707 protein.

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717 **Table S3. *Steinernema carpocapsae* non-argonaute RNA interference pathway**
718 **proteins.**

<u>Non AGO Protein List</u>	<u>Homologues</u>	<u>Accession *</u>
AIN-1	x1	L596_g5788.t1
DCR-1	x1	L596_g26648.t1
DRH-1	x1	L596_g26385.t1
DRH-3	x1	L596_g25860.t1
DRSH-1	x4	L596_g13580.t1
EGO-1	X3	L596_g919.t1
EKL-1		
ERI-1	x5	L596_g16241.t2
ERI-3		
ERI-5		
FKH-3		
FKH-4		
FKH-5	x1	L596_g10448.t1
PASH-1	x2	L596_g24740.t1
RDE-4		
RRF-1	x1	L596_g7841.t1
RRF-3	x1	L596_g11915.t1
RSD-3	x1	L596_g5786.t2
RSD-6		
SID-1		
SID-2		
TSN-1	x1	L596_g28061.t1
UNC-130	x1	L596_g11813.t1
VIG-1	x1	L596_g22586.t1
XPO-5		
XRN-1	x1	L596_g18210.t1
XRN-2	x1	L596_g10155.t1

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733 RNAi pathway proteins

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735 >AIN-1 - L596_g5788.t1

736 MWGEETGAETMTLNPWAGVSQQPQVWQNMNSRGNGNPSVANSVGSFAPNGGSVRPGSQQW
737 DGGATPAGRSAWNPNPHAPRWTPSVNHGNASGWPNQQHWSVANSNGTYAEQTKKNMALG
738 RSSGPNLNMSSQRFDSQGWGSKVDQGT PWDGTGNSGGVSQQMHGEGKEQWSNPTRWTQPPGT
739 WPTPIVAPMVADWQQGQPRHDHWANHPTGGMVPSGGAWAQQALGVASSSEQGRVPYDPNP
740 QTPGPWVAPQPAEMNNDMMWHDPNPKQKKIQKDVGTGIWGDPTSQIEIRRWKDLEAEGGE
741 FPGGSDWGSNSSTQPTGWGDVGPAGQNDGTDWRGAQQALQGGWSDKVDQDRDLNNGGK
742 SDLNQIQRGIQNAIANGALCPDGQSRLQWKKQKRGIGVGSPEEKMGVASSSDSNVPSLI
743 AATKIMNIHGDWTPPSSVADDSAKSEDQVSSSGTNNEKESVKEASPTPPPTQLDDGPQ
744 EFVPGKKWEWRDPNKVAEDPNATPGNCKPNPLMAAGNNMNAFNSAVNTANPYGPEVPS
745 GNASTFWPNSQGFNVFGRDMYNSVRARLPSNGQFMPQRMSSGGYSNSNHNRMQTPPGK
746 GVFIVLNHQGANETQLNFSCTRAGQLLNVASLGGQTVLLRYADSSSEGLVQLKLKADFPN
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1238 LLERRRLRVTQHISLESVRAVVIENKIT TLDNIIHKLNVKNGGFNYTPLIEHVGN SRELE
1239 LASGNVLVIGLDVAHPAPMNASQRRMMHSVDAT IRSLEPSSIGIVANVIKNPHAFVGDYH
1240 FQTARREAVEPRILKERVKWIFDLLAQNRPEHQRPKHVVYLRDGVSEGQYTMTIRDELGA
1241 IREAVREIDPKYRPFALIIVTKRHNKRFFDS SKGVVGNPLPGTVVDHSVVRTDITEFFL
1242 QSHIPILGTVKIPQYDIPVNEG GFFMDELQAFANCLCHSHQIICTPVSLPEPVYAAHEVA

1243 KRGHNNFLEFRRSHPEMVPYVDGNVNIIDCEEVTAKLAYRGTPLEAIRFNA
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1245 >C04F12.1 - L596_g11107.t1
1246 MMGSKYPDKLPAHTKGNPKVPLVSNSEFEVSLGDKIIHVYDVSIIQDCESRGRKKVIDWST
1247 SSGDSAKRRIKLAVSKEIFKKALEVKKFASKEAALVDFDFSKILFSSEKLDHLCSAIIILT
1248 PEDFDVLPSEFEGNSKLCRGTYTVRIVPTKAGSHQFRANDLEKALAKNQEDHSLRQFLEVA
1249 TSFKPIQEGTHKFFRGILHDVRTSAPGRRDLQNGPFEIAYGISKGARIIGDMDNPKAALV
1250 MDSKRFAVYSESKETFLEDIQKVLGDKDLKIHLPRITKMFQGVTLGHCFNKAVEVKFSSSL
1251 SNVLAEKLTFFENSQKSFIVDYLEERYENYQCRARRWPVVVDKFPGRSGDVCYYPLDIL
1252 YVKEGQLVPLPLQQEFGITQELLKEVSKPHLRSAEIGRAPKDLELNARNAHLREHGINVK
1253 QAPIKLEGYRAQPPKLG YANGQTA AVDANRANWEAGRYIYPAKVDSFRYFVRQGC MGRDQ
1254 ANLFLNKFLDMCRSKGLDMPKPQIEFIKGPLALKNLLSAEDKAVGKKKSVTFVLFVDSEK
1255 SKTHDALKFYEAKYQILTQQVRSETTFKAGRQTMENIVAKTNEKCFGQNYAIHGDEFIST
1256 KDTLILAYDVWHPTGASAQKRILDIPDDTPSVVGMSEFNGGVHADGFIGFYAYQEPLQERV
1257 DVLKSYMQHILRIFKKTRGLLPKNIVVIRDGVSEGQFDMVCQHELASIRAGCRQFANA EK
1258 VGWNPKFMVVTVTKRHKRFFVQDGHVRLNPPPGTVVDCTVTRPDMTEIFLQPHRPFQGS
1259 AKAAAYSLLVNELEIPKQKSGSDLWLTNFLMKLCYSHQIAPSSISIPPEPVKQADEWAKRG
1260 AANLEFLKRENGDKSLDMHNFLKSSSEGSFYDWAALS DALGYHTKRLEGTRANA
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1262 >HRDE-1 (WAGO-9) - L596_g11197.t1
1263 MTSTSVNLNSFKIDISETVHRVQQYELVFVLYKSFKPGFTATPSRTRVPLGFKYKYNLDT
1264 FQNGDHWVGGYDIAYGPQNPVKIRMRKEMLFHFFFEFFKQQFSHTLAIGDVVPMMI FDCDR
1265 TIYSNQQLKVGRGFSHTWSYIEHLPEKVQDHIDILCGNVSDFCGLSVFLNRVGEVDMTDL
1266 GSEKNPNKGVIQFLETLLWQPLYTQFTDHVIYGKTFFDLAEKNRISVMQDGLFLATGYDI
1267 SVDLIPDYEKDMSKLMPPVRFPGSSVFYQNYSRMDALLMSLSGTNGRDTLYQLEYDCKH
1268 PSIVEKLSQTVRGATVQFIYDSDQIFEVDHLDPRTPHQITFLLQGRRRISVTDYFMAKYN
1269 IQITSELPCVARKTRYGMSFYFVEVLRVAPNTKAIHKYLPADV KDMIEKGTILLPSAVQQ
1270 RIKEAIGEMKLYSAGIRKHD MARINPFLFVFGIALVDKDRPVRIAAHVSESP LIEYCYRR
1271 GATSTLMKTEQNHGPGVWNKSMKFLPPFLKPADLGKPVKIRFVNTFVEEVH LINRFMDKTI
1272 HKLRLKGIDVMKERTISGSERF GKIRDARDIFGRNKDYRTMLKAAEDILTSSGADLFYII
1273 GSTDPMNPTRDIFKLAEITKPSDKRIVTQHIGCKTLCTTVSTGLSKRSPGILESIVMKTN
1274 MKLGGTNYTLREKDRSSGYRYGYPRSTHLIIAFDKVNPQHTDEKGEKLYHPKACAMTYMI
1275 PHTTGLITRGTYWFQTSNETNLTM MISAFEEALKFYHKDSRGYDPEKVVVYWDVSHGEKE
1276 VTQEMDAMMAIVAEQRGESVVRGPFLTFITVDQNHNTLLLPT EANVRDRLSLQNVTAGTWI
1277 QESTIGESFTMVSYESDDKMTRAVKYDVKVAETHMHSIQDLTHKLTYLQNSGWRSVSVPA
1278 PLKGATKLAKRAMKSYDIMDQIRGRSGPIEKQLFEERVKEISDWI AVKHLTNYWA
1279
1280 >PRG-1 - L596_g25491.t1
1281 MDARTGQLKSFRAASRQNRGPQTDES LICFAYQNHPRFIVKRTKSI SLLKTLILRRFDAL
1282 ASALMRQQKSI IYDFQMVI ALKTKF
1283
1284 >T23B3.2 - T23B3.2
1285 MRSRRIYSFNSSIEMSDGEIPIRVVDNRNRD DLLKILLVLIIVFP PAAVAVQANECNV
1286 HWWISLFLMLFFIIPSYIHAVWYVFIRKPKELTIA
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1288 >WAGO-1 (R06C7.1) #1 - L596_g19943.t1
1289 MSDFENPNVKRPVTRIVDDVEELFERLQLEPGKPMADKQLPGTTGRKVKLQTN IYGLSL
1290 QKIPVHRYDVNV IARLGEREVLF TKRSKEDAVSTDRKDKCRTAFELAVNTFGEQFFGQER
1291 YALYYDCQSILYSMKPIPALEDKMSHEL SLLPHHLTDFPAFTGLDAVVVEIKKVADTFNL
1292 NLGNLGFNLRELIEQDHS LQQFLELITSQH ALFTPADHICYGSGTSFLLNHKKYGF LDED
1293 CPDLGEGKYLA VGSHKSVRFVEGPGGRSGAHA AVVVDTKKAAFHACENLIQKAMAIINLT

1294 P QTHCRKNEVDKLRGQLKGLFVHTKHGRRQRLFP IASIT EDTAADKQFEDPDGNDVTLQI
1295 YFQQKYGITLRFPHAPLVVVQENKQTNYYPMEVCYVNDNQVALNQQTPLQIQKMIRACA
1296 T PPAQRVRQNKENMRALELNN SNRYLQAAEVRI SNNALILEGRVLQPPDVYYGNGVKAVV
1297 HPDKGSWRLQNKPHFLLA VEIHRWAIYVVG TGNRDI LDQPKFENFIRMYMAECSRSGIRI
1298 NDPCEHRILPADPEI IKDRIKAACEGECYFMYFITS DAVTDIHKIMKYAERE CGVITQDM
1299 RMSSANDVVAKGKRQ TLENVVNKTNIKLGGINYDIRFNSPDLNVLRNDRLF LFGFAMSHA
1300 PQTQHERNKG VAPRSPSVIGFSANMKSSPVDFV GDCVFQEPRRDEKIGVIRGVVNNVVTR
1301 FRDSRGYLPKELI IYRNGSSEGGYPLILKYEVPLIKKALEEVQCD AKVTLIVSQMHNVR
1302 LMLSQINERDRAPEQNIKPGTVLDVGAVHPVYNEFYLN SHVSLQGS AKTPRYTVLFDENN
1303 YNMDQLEYMTYHLSYGHQIVTLATSLPAPAYIASRYADRGRNLFNASNTNWNVLQDQGQLD
1304 YNQITRDL SYGVSDLRDRVNA

1305
1306 >WAGO-1 (R06C7.1) #2 - L596_g995.t1
1307 MDALKGHCESDETLYKIETALVSKSGTIRFY PQEAKKVLN WAVEPVMVEEDPYKEEDGAF
1308 YSLCHVNIEFLASGVHCIAKLLAPFSNFRRSSATKSSSRRPKTTANVENRSATEPMTNLR
1309 SKIAGPATKTASITVYLTMESVTTKMAHLTMAPKLGAGTGGRPVPLVTNMYQAKMMRAQP
1310 VYRYDVAMEMRFGAKSVSLVKKTIIDDMVAIDHKDKCRAAFRIAVRLHDKVLGSPVGLFYD
1311 LQSTLYAIDKIKDAKENTECKEIELCIPSDMLKNNDYFKDRNPDSVTITIKRVEADYQLS
1312 LGDLSFATDASKAHLNADLVQFLEIATTOYAYLKAGDVLTYSSGLIYFSEKRRKLN GGNL
1313 LIDGVQKSIKVI EGSQKGPQLAVVLDPKKTA FHAKDVAVSDKIYEAGFMEDDGRVHPAK
1314 LETVKNQVKNL FVEVRYGKKPTRFMINGIDKESARVKTFLSSTGETTVESY YLKQYQIEL
1315 QYPLAPLLAANKK VNGERTTIYFPMELCYVCDTQRVKNTQQTSKQISDMIRSCAMPLPADR
1316 VKEIKDCAKRLQLNGDAVSGSLRSAGLSVATQLT SVQGRALPPPEIVYKDNKKFSVDPNS
1317 GKWKATGAQKPKFLLGASINRWAMMCVADRPQRDDALMKNFAEKMVRECQGRGMRVNNP
1318 VFYQAVQGRPDSLES MFRRAKQDNMEFLFFVQDGR LQAHKDIKFFERKYEIITQDLNQQT
1319 CRSVVEQNKFLSLENIVNKTNVKLGGLNYSLIVNAPNTQH LFAKGRLYLGFQVSHPAPLS
1320 DDQKAKGMKPKQPTVVG VAGNITNQPAAFVGDVFFQEPRDDRMADAMETLVRDFALRYKD
1321 AVGVAPA EVIIYRNGASDGQYQTI LDVEVQEVIRAALTAAGAGS AKLTYMVVSKLHNVR
1322 MPAQITG IKAPEQNVKPGTVVDTNIVDPVFAEFYLN SHQTLQGS AKTPKYTVLYDQNNFP
1323 MSYLELMTYVLSYGHQIVGLPTSLPTPVYVAGRYAERGATLLSASRHDSMDCKFSELTEA
1324 LYGSTKIGKNRLNA

1325
1326 >WAGO-1 (R06C7.1) #3 - L596_g15757.t1
1327 MDTRSQAQGGFDTSIKVVYEMAPKQYAPENKAPVDLVCNAYRLRMPQPSSDYPARVYVYD
1328 VTLLTLTRADGSLTLVKTKMDDYTHYVSKQRCSALEAFS QKFPGFFRTNEQRLFYDLQS
1329 ILFTRVELPMRNLTEAMSIEASEFSRFGFAEGGQGMQRLNIVVQKTQSGT SIGLTD FSFL
1330 SSDIAEVGQRHDL SQFIEICSSQNAYMNPDQRVTFPGGVS YERGNPGDLVDGSKRVWNGV
1331 RKSRYRLTGAGNPVPAVVDARKSAFHKEGEMVSDKVYAMGLMNDG SVFDYNIDNITRQ
1332 LKGLFVMVKHLNNQRTFPI LKLDKKT PATYRFTSDDGVEMSVADYMSKKYGTALEYPKSP
1333 LVVVWMKQREVHYPMEMLYVC PNQRVTTNQQTSKLVSEMIKKCAI VPSERIKEIKHQATS
1334 LHLHDGALREVCIEVDPNMLHLQ GKAVEAPKLKYGSSALVSVERNTGKWRTSNTKFLHPV
1335 QIKKWAVIVLPTGGKLTGQDSQVPTKFVDLMIKALI IKGVQVGPPAYTGMSSGCQLEEDF
1336 KECARDGYDFMFFIQDSKLQFHKDIKMERRY EI VTDLNLNTARNVVQQRKHL SLENI I
1337 CKTNVKLGGVNYSIHIDRPEFADFFHRRRLYVGMQMSNSRVFELGDGSDVEKAGKPTIIG
1338 ISANVDRELSS FVGDFMCGPANIEDLSSV VTEIFKYYS EEFKMRGHFPPEI VVYIGGIS
1339 DGMPKLLRWHI PAIRHGLNTAKCTAKMTVI FTSKSHNVRLFPKNVTGERAPEQNVKPGT
1340 VVDTGIVHPEFTEFFVTSHQTLQGTAKVPKYTVMIDDNNLSLSYLETMTYVLA FGHQIVA
1341 LPTSLPTPLYVAGRYGDRGGVLYNAFDGKDDLYAVNDQLTFATS KKLCKGRVNA

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1343 >WAGO-1 (R06C7.1) #4 - L596_g16574.t1
1344 MPTGEDPVLAEPVASTEAE DNASPRTPHAIHEAGDAIGLERRLEASTSARDGNFYVALGL

1345 PILQGVVGNKKEALAVGVPIELSTNAFGFVLPALPVWQYEIEIEGLLAGSERRVFFTKRS
1346 PDDAFKIRKSQECRRLFQAIKHKYAHSFSEDNETYFYDNQGLLFAIAPLDLSLNEKMDCM
1347 LTAEELAALSPPTYASFSEVIVTIKNVAANPMTVGHVMTYTSQVLENNRRLQQFLEVLTL
1348 SQHMLANPNEFLTYGTRSAYLLDTSAHGLEAGHLTKDKEIKIGCEKSVKLVEGPMRGGQP
1349 DGKAVALIDVKKTPFHIPEGTVLDKARAILNREPKPSDAPRLKKDLAELVVYTKHTSKEH
1350 RYVVENVIADTAVSMTFPWTEEGREVTLSEFFVQKYRQNI SFPRTPLLVARFGREKLIH
1351 LPMELCYVARNQRVTSRQQEVDNISAKMIKACAIAPAERQLQIQETVNALQITSSNPYLR
1352 AARTKITAAPLIVTGHRLEAPKIAYANNEVLS PDARFGFWKPPNAHRRPKFFKPAVINSW
1353 AIVVLPSQAEFLQGDIISREILARFTDLFRSECRDRGMQIGQPVFTEFMKADVQQLRDLI
1354 KSLTRPDPSVCRPPLRYVIFITNGGITFCHQPMKYFERETE IITQDLKMQTVVNVVQQNK
1355 RLTLLENIVNKANIKNGGINYVVIRNMPGQRPI LKPGRLVIGLAMSYSVRRQAE EISTLPT
1356 AVGWAANITREEGELIGDFLLQESFKKDRVAVIQTI VDRVADAFKHPGGPKEVILYRSGE
1357 EGRFRAILEEDLAVLRATFDNMAKPKLTVIAVQKHHNLRLMPTKINRQDRPSLQNLVPG
1358 TVVDRYVTHPTFTEFYLN SHVAIQGTARTPKYTVLQDDANMSLEELEEMTFGLCFNHQIV
1359 SLPTSLPSPLYIAGRYAERGMTLYRQHQEHEEDQGTSASSPSSGSSGEPHLDVERLGTI
1360 SYGSSRKLKHLRVNA

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1362 >WAGO-1 (R06C7.1) #5 - L596_g20961.t1
1363 MEREGTELRPSTIVRQTQNA AASSAPAAQVSTSAGAPPAAFGIRGPATSAETS FYSQLVN
1364 IPTPPLEEKKQPGTTGREQFKLRTNVFGLSLPKDAQVFRYSVDASGTLQRNDRRIEFAKR
1365 VGDDITYLNRREKCRHVVDQVAKNPAIFGDRRELFWYDSQSILFSRNQLDIGSEAQFVL
1366 DQSDIGQNTLFEFGAHLKMVIRPAQTNFAVSIGDLEAYIQAE LFESDHALQQFLEILTAQ
1367 YAFNTPTEAMSFGTRTAYLLQPEKYGFKPADCADVGDGKFLGVGCDKSVRFIEGPGGAGG
1368 QRAALVVDLKKTA FHKVQSLYEKAREILNNRDPKST DASRLRMQLKGI VVETKHGSRRQE
1369 FAVDNVADTPATKKFKDLTGHEVTLQQYFQQKYNITLQH PDSPIVLTDRTKKFAAFPME
1370 VCWVVDGQRVALAQQT PVQIQKMIRQCAVPPADRQRQILGLVQGLQLNSENKYHKAAGVG
1371 ITPTALQVQARLLQNPIIVYGGKSTMRPDEKATWRLARQKPVYLPVKIDKWAMFVIRGG
1372 NRSDCVDQAILNQFSNMVQECRARGMTVPDRPTGLSFIGASREEVQETLEKAKTEGNQF
1373 CFFITNDVTHIHQFMKFQERKLSIVTQDLKMSSAFD VVKKGKRQTL ENVVNKTNIKNGG
1374 INYSLRFDDPVFNMDKLLPKDRMVI GLSTTHPKPIPGKKEQDQAPQDKKKQMHEQRTGPP
1375 VPSVVGVAANVLTESIEIVGDCLFQQPNREEKIAL LQPVIRSLMLQFMKHRGMPPVEIVV
1376 YRQGTSEGQFRNVMELEYKMKVAAALQOGLNPKITFIVVQKMHNVRLMPTDCKAGDRAPE
1377 QNVKPGTVVDTMVTHPKYNEFFLN SHVALQGSARTPRFTVLYDENRLPMDEIEALSHSLA
1378 FGHQIVNLTTSLPAPLYIANRYAERGHNIFIASQEDYTKSKSSLQSPHSTTIEDNLDFSR
1379 MMNELSYCNSELRDKRVNA

1380
1381 >WAGO-1 (R06C7.1) #6 - L596_g322.t1
1382 MSEELHAASKLKPTEGIASRKVKLTTNHFTLSFKSSKPVYRYDVAMVHYMMTKDGEKSRD
1383 MCKGERDDAAILERQRRCLVLM EA AKVAHFTPSKSCCVYDNSKTLFSSEKLENQCAQI
1384 RIEGEHIPDGFKNHPKLKQGYFFIEITPVSTNHKFTIDDLKSGVSDDLNTDHTLRQFYE
1385 ILTNAYAVTNDSHMVFGNLYDNDKGESGRKKLKEARNLIFGVNKGARIEGSSSRLVAA
1386 LVLDSKKSTFFDDANNKGLAGNIRDLLGSHFNAQPHEVVIGDHN RKTVVV TYLKDLRVYCR
1387 YQEDRDFVIAGVTKEPIEDLYFEYGSTKTSVM DYHKQAYNTKILYPHWPVIMQGPRSKN
1388 YFPIEVLGVSKGQRPVISKQTPGQMAETIHDCAARPHIRFAEILKKLDGLNLASNPNEF
1389 LQSFVVKIDCNPIQVQAHRR LAPKMVYAGNKEVDYDDIKGNWFANNAYILPAKIPKWFVI
1390 TDRIDVSIVRK FAGILKDTMKMKRMTVGE PQFLEMPVAQLDGFLGKMAKELKPEDQSPFI
1391 LFADTND DSHALLKLYEAKHQILTQH LRARTVIECLEPRKRLTVGNICNKL NCKNYGLVY
1392 AVNPQDHAKTMYLSKGDVMVVG YDVSHPEPQPAHERRLGIPTT P SVVGF SFNGGVHPGM
1393 FIGDYQFCAPRQERVDILEERIQWMLRVFTTNRKTLPSRIVIVRDGVSE GQMPMVLQHEL
1394 ESIRRGVKKLKAGYNPKFLLVTTTKRHAKRFFAETERGIDNPMLPSVIDHTVVRPDVTEF
1395 FMQAHKAIKGTAKMPAYTLLNELGMTLDQIQSFMMGLCFEHQIVNSPISIP EPVYQSDE

1396 WAKRGHSNILAFFRLMDCPDPQKPSQKMIQRYLKASENPTTGQPEVEGYDWVRISKMLCY
1397 RGRRLKTRANA
1398
1399 >WAGO-1 (R06C7.1) #7 - L596_g9675.t1
1400 MLEIRIPPIVDDQKRRETELENTVNVRTNVYEFELPTGTSEIYHYEVAVIGKLGKSGRTVD
1401 LTARTTNDVLTLERRDSCRKVLGIVSERYPAVFDHNLVIFYDQARLLFTSKNVEINTAAGI
1402 YSVILGCQDHLKGDDEMFAAFTSVEFRLARVSVVEVGNVQKYLNKNLRLVNHLSLEQYLNVL
1403 TSQHVANKTITTFGNESVYLEQDDQCGVRNNVDLGSGLKKTGFASRLIEGLTEQPAAA
1404 LVVDLKKAAFHVQQTVDKARLILQADRKRCCGSPRIEDTEDLSRELVLVETKHGSR
1405 VRKYKIAGFDKSTPASRSFEKDGRNVLIADYFAQQYKVKVENLDTPLVIVRGYGRDFHLP
1406 MELCYVKDQRVGLKQQTDPDIKMIKQCAVKPVVRIAEIEKIVKGLRLIDERKDI LGSKV
1407 GIKDKPLKVEGHLLPPPLIVYADKDTCNQRVPPSREPASNGTWSLTTANTTPAAFFKPAK
1408 IEKWAVIAIHTEDNGDPHDERNKRI LAQNTLETNIIHQFATVFTTEECRNRGMTLPEAEHV
1409 KFLDDTSTTVRDFIHQAKLSFVLFVCNNAITHIHQSMKCYERKFETVTQDVRMATVNDIM
1410 TKRRFQTLNIVAKTNVKNGLNYNVEMPSSKNGVRELMPKGRLLVIGLTVRVVVPKSPKE
1411 IEEDKKKAEEREKQHKKARDSKFARKESTTAKLKPIMTVGYAANFTDVPTEFIDGHLVQE
1412 YRETGDILGMQIIFERVVLEFKRARGMFPTEIVIYRECTENSDFIGQLQLEQMLLKSIAIK
1413 STASFRPGFEPKLTLLIAVQKRHHVRLMPIAMREAKAPEQNLQPGTVVDRMITHEPTEF
1414 YLNSHTTLQGTARI PRYTVLKDENNLSMHEVQLMTYGLSYAHQIVNSPTSLPTPVYVAVT
1415 CAERGMNLAKHNFRAMNIKFNANDDYSKLEDQHVDI SHLNEQLSFGNCKLSSIRNNA
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1417 >WAGO-1 (R06C7.1) #8 - L596_g18735.t1
1418 METLTKAVANMPPVNLASKREDAKPNAQERALPLQTNMFSLSMRDEVPVFMYSVDVFMK
1419 VRSKAISLVKHSRDDYIVIDRKNKCRAAFRFVVRANPAVFGNPGKIYYDLQAQLFTLEKL
1420 NMDNEDEGLELVIDGADARRSTDFAEIPLDGIVVQIKRAGPKFDLALGELQKLFVAQEPK
1421 SHELLQFLEVATSQYAFLTSPDFVTYPAGLSFIKQKDPKAGTELEGGKLLLDGAQKSVRL
1422 IEGEKKTDDGGKLAVIDAKKTAHFHKTQYQKVIKVNDFGFLQSDGTVHRMRI PDLAKLLK
1423 HIYVESRYRKRTQRFLINDVNPDNARNKMFTRNGVMI SVEQYYREVNITLRYPLAPLIV
1424 SKPMKSKDSEEMVCLFPMEVLFVCPDQRVKINQQTPRQISDMIKRCAMQP DVRVKETKN
1425 YAEKLQLN GPASQA CLNYAGVSIANEVVKVSGRKLVAPEIQYKDKKIAVDSNTGTWRSSG
1426 REKPKFLVGS SLKKWAVYLLGVRQPLDENLGRKFVMMLEEARARGMQWESPTAVKGV L
1427 GHVDNIEKLFKAAQKEGLEFLFFIGDQKVVHTELKFFERKYEVITQDLDLKTCRNVEQ
1428 GKFLTMENVI AKSNMKLGGVNYSIQVNDPTIKQFFKPGRLILGFQVSHAAPLKPDEIAAG
1429 VKPRVPTVVG VAGNVSKGDPACFVGDFFYQTPREDRMLEAMDTLIADFAQRYKRATGKDV
1430 AELI IYRNGASEAQYTDLVRDEI PSIRDALQSAGFRGVKLTVMVVTKLHNVRLMPVAIKT
1431 GAKASEQNIASGTVVDKGI THPKRAEFYLN GHVTLQGS AKTPKYTVLTDQDQFTLQQLER
1432 MTYALS YGHQIVSLPTSLPSPAYIAGRYAERGALIYQGFNHTVQHDADLQTLNGLTLYGK
1433 SKLGDKRVNA
1434
1435 >WAGO-1 (R06C7.1) #9 - L596_g18947.t1
1436 MSNPSHNGRRPGANGNGSQPGGLEMRAATHVRQIRSGASSGMSSEVRYQELTQAG
1437 QQLHLEEKKPPGTL CREELIVHTNVYGVGLPDVQVRYDVDACGFLERNDRRVEFAKRAV
1438 DDVADTTRRNK CRAVVQLVCQNYANIFGNHREYFWYDSQSILFSKNMLDGI PEREKKEFV
1439 LTQEMLAGNPMFRGFKNVRLNIQRVSN SFAINIGDLTQYISADLEENDHSLQQFLEILTS
1440 QYAFNTPSEAI CFGSRAAYLMQPDKHGFKAQDCADVGDGKYLGVGCSKSVRFIEGPRGDQ
1441 RAGLVVDLKKTA FHYESLLAKSRAI LNREPRGNDAGR LRQQLKGLVVETRHAQKPVRF T
1442 IEQVSD ETPRNK FMLQPDNREVTLLQYFQEKY NITLES PDSP IIVGDRLTKFAVFPLEA
1443 CFVVDGQRVSM DQQTPTQIQKMIRACAVPPADRQRQILSLVQGLQLTSSNVYHQAGIT I
1444 TEKPLLVRGRILPNPKI IYANNVVVSPDAQKATWR LDRQKPHYLI PAKIEKWAMYSIRAG
1445 GRSDVLDQPTLLRFAQTMVTECRM RGMQLGNPGEVSFIGCAQEEITATMEQAKKDGCTFC
1446 FFITNNDVTHIHQHMKLLERQ TGVITQDLKMS SAVDVIQKNKRQTLN IINKTNMKNGL

1447 NYTIRCEGLSNEQLLPSQRLIIGISTTHPKVAQTALEDRDRADKNE DPLTKKKPHHEKHH
1448 RSEHITPSVVGFAANFKKDPIEFIGDCLYQYPQRDEKIGVMQPLLRAVINEF SNNRGVPP
1449 NEIIVYRNGINSISTMELEFLMVKAVARCQGITPKVTMIACQKMHNMR LMPAKINPRDRA
1450 PDQNLKPGAVVDSNVTHPKFNEFYLN SHVCLQGSARTPRYTVLHDEGQFSMDELQALTYN
1451 LAFGHQIVNLTTSLPTPVYVAAAYADRGHNIYGV SQKDYTKSRTSDLNSSTLEGNLDFNR
1452 IMMDLSYSNSELR SKRVNA

1453

1454 >WAGO-1 (R06C7.1) #10 - L596_g1747.t1

1455 MASVDVELREKTKIRQEADRMFKEDYGIDV GSTEMPAKMMKPAAQNTMSLVTNLF PVKTH
1456 QQMPIYRWDVDIT IAGKNGKTFSLTKKSDSDAVAVNRK LKTRSIFKRLVAAHPEKFGKLE
1457 ENYYDLESMLF TTKDLSTDQEAQDFVVDGLDAKIIPGIASANVILKKVTDRYQVDLTD FR
1458 HLTRDVANVDLSHKQFLEV VTSQYPLMAEDYVCYPGGVSFSVKEKTTPLAEGKYLGHGVQ
1459 KSVRYIENDGRSPCAAVALDLKKT V FHSEINALEY LKMQNTYVKQLIESKQRAKDNDMTL
1460 KTACQRIKGLRAYTQHTKRQRVVTMDRVGPETANEARFQLQD GEEITVAA YFHRTYGAPL
1461 LFPNAPVAVERKPGSKDVNYHPLELLTIAENQRVTGELPQAVISDVIKQAAV VPLDRQQQ
1462 IMRAHRDLKISTNDYVDNVGTSIAPQMMQVKGRLLMTPKV VYGRNTQIEAREGKWRAERK
1463 TFLKPAGAARWTCMMLTNNRLG EQMMHNFLNKYVAVCRRNGMQMADPIEPFVVDWRRTDL
1464 QTEIDAFMKDCTQQYKLEFVLCIQDNNMHEHKY LKFLERQYGLITQDICTRTVERCIGNA
1465 SATIDNIVQKTNV KLGGLNYGLEYS PDGRHDVLSATTMFVGLGMSH SKPPKPDATGQEP
1466 SRPASVIGFAANVLAQSFAFVGDY YFQKADRDEKIFAIVPIVTR LLEQWCEHHGGQMPKN
1467 VVFFRNGGSEGFQMI LKYEVPLIKFAIEEF AKKSAVQQQFETKFCLLVANKRHNVRFFK
1468 SNITSGGRAAQNLQPGVVVDSGVTHPVFSEFYVNSHTTLQGTGKT PRYTVLHND CGLTL
1469 NQLEHFVFALSHGHQIVNLTTSLPTPAYIANDYAERGM AVLGQFLRASGAPVDEYETFNS
1470 KLTFAAMHPGSKFHHC RVNA

1471

1472 >WAGO-1 (R06C7.1) #11 - L596_g1696.t1

1473 MERFKKLPTTTLKARQVTLVTNHFR LNPTGRTVFRYDVAMSHTRMVKGTEKIRDLCCKGD
1474 RDDAAILERNRCLALMDAAFAA PFASTSAAVIYDNSKTLIAAEELDMRQCACIRLEGG
1475 TIPPGFINHPRFKEGYFTIQITPTSSNHRLVINDLEAALAGDEANPDRSLRQFLEILTNQ
1476 ETLKKN SFMAFYGNLYSREAADQRK LREARILKSGMSKGARIIGSSSDLVAALVLD SKKA
1477 TFFDDTNKNGLAGNIRELLNRAPNDAPSRVHINDFDRPDIVKYIKDLRVYCLNKPDNTFQ
1478 ISGLTREPLRNVFFDMGGEQLSVLEYHQ RDGARLAYSHWPAVIVQSPRGRNYFPPIEVLGV
1479 CEGQRVPI SKQTPGQMKVVVNDCAVLPNVRFAEIHQMLNALNLASTTPNRYLQAFGVTID
1480 VRPMKITAYRRQAPKIVYGGNIKVQYDDVKGSWFS SGPYVLPAKIPKWFVVDGIDQRTV
1481 QQFVGLVQQAMKDKRMEVAQPKYMEMKVAGMDAFLSSIVKSLKPGERSPFVLF TDANEDS
1482 HAFLKLQEAKHQIVTQHLR TKTVRECIEPRK KLTVGNILNKLNCKNFGLNHLVAPDHEKN
1483 YLQKADVMVLGYDVSHPEPQSPQDRRLGIPPSTPSVVGFSFNGGQNPEMFIGDYQFCPPR
1484 QERVDILESRIQWMLKVFNDHRK KLP ERIVVVRDGVSEGLDMVLQHEMASIRRGAA MIK
1485 EGYKPKFLLVVATKRHQKRFFIDKSNGEVDNSMPLTVIDHTVVRPDVTEFFMQSHKAIKG
1486 TAKVPAYTVLQNELGMSLDEIQAF LMGLCFEHQIVSSPISIP EPVYQADQWASRGHS NVL
1487 AFFRVMDIEDPEDPSKMINRYLKPVEDPLPGRPKFEYDWTRISKLLCYRGRRL EKTRAN
1488 A

1489

1490 >WAGO-1 (R06C7.1) #12 - L596_g18749.t1

1491 MPLITVEMSLFVAEGSVVIFFN AIVLLIIVTDSTLRESTELMFIGGLCFADMTDGIAYFY
1492 AGIHRLCNILSR TDDVMISRLECFQKPFMFFFYGYQLPAMMIFIVALDRFVAVFAPMWQ
1493 RKVERSSKLMVMVAIFLWVT LTYVINVFLFHSYSTGYTTAQCFAHDVFLSQLWDFIIVQR
1494 SILILLCVLLYVPI LIKTRRIFQSN DKSVQSN SFNVTIGLTMTC SVLLLFIPDVIYFDL
1495 IMDFHLILYLLGLNKCVANVFIYTLRQKEIRKKIELICRRVFCNLKGPFDLELSKRRQRS
1496 NITRTLGI AKRCQSLSAVAWSVVLSSAVPQETWHVISRPPLGEARFDGRIDKGF IAPLLY
1497 FYVSKMSAGPSNLQPDVELR PATKIRHAAEIKFKEMGIEVGIPEMPPKIPVPPNGNVKLT

1498 SNLYAVRTSSQIPIYRYDLDTISQRNGKQLALTKKSDSDAVSIDRKLKTKIIFFKMVQT
1499 YPELFSGVHHCYDLESTLFTLNDIMKDDPEAEPKTLVIEGLDGAKFQGTTSATVILKKC
1500 HDRFEVDLTNFSHLELNIESTDLSHKQFLELVT SQIPMMSDSFVTFPGGVSFMSNASTTE
1501 LPEGKYLGHGVQKSVRYVENPQTRKPMAAIALDLKKTAFHAVLTGLDYLSQIVDDRPLHN
1502 GQKLPNSVFGSFGALKMKGLRCCLQYGNRYREVVIHEISNKTAQEHYFDREGQQITVEN
1503 YFYQMYNLRRLKCPTAPLAGEKKPGQKALCYPLELIRVLDNQRVTGELPQKVIRDVIKHA
1504 AVVPALRIRQIQDSCADLGLFGNDYLNQVDTVIDSRPIGVEGRQLKNPKMVFVGKNTSVES
1505 RNGQWPNRGAMRPPFFIPSTIRKWSVLMISNLQNGFASETMHTFVSSFINECRARGMTLP
1506 APSDPWALNAREELKPQMEGFFSECPGLGIEFVVLVLDQDDCFHEHKFLKFIERKYNVISQD
1507 VNMKTVQKCLQRAAATLENIVQKTNIKLGGNYSIEMTNPAGTSNVFQPD TMYVGLAMSH
1508 SKPPKPDSTGREPPK PASVVGFAANVLPQNFAFVGDY YFQAADRDEKIDSIVPIMHRILD
1509 MWCQHHEGQMPKNILFFRNGASEGQYKSILKFEVPLIKHALEKFRNCCGVEQMTPETKFS
1510 LVVATKRHNVRIFKANIQAGRPNEQNLPPGIAVDRSIVHMPFAEFFVNSHTTLQGTGKTP
1511 RYTIMHNGADFKIGQLEHIVFGLAHGHQIVNLCTSLPTPSYIAGDYADRGMVVLHEFLKT
1512 TRAEADQYDTFNEQLTYAALQPESRFHYCRVNA

1513

1514 >WAGO-1 (R06C7.1) #13 - L596_g18732.t1

1515 MLGKWRTVRVEFEAAEELDFRWEFDHMAAIFREALQPHCRCLLGGFTRSTFQINTVYPKE
1516 LQSPAILFPDDPNGLCKRNLLVQVSLFLLLKSFQLHDFSSSQLSGRFLLLFRTCGESECE
1517 RKRPRFATFGLPNLRSSANVQKLRANSIAKTERAANRRSNPLQTNSSPGPARKTAPHRN
1518 PAANPKTCFAIGVRGQVADTVDRSFVPRGPLLTVAPPALSPSALPVFSPIRLFTPPHPAA
1519 YFRQSTMDLHTKLAELSVADNAMKPTPATEFKETVEVSSYFEISIGQNALAYRYEVDIL
1520 AISQRGQEKNLTRGPADDGAASLRRQLCHEVFNAALKKSNFGTKQGVRLPVVYDCRATL
1521 FLPAPLKMEDEVVILDKDADFAEMSKETLFTLDPTDVIRVRIAPTTQNAFEMDLRAELM
1522 KADFCEPDCEFARDRSFRTFLEMLTSVNAVRSQSHTQLGVGNFFDNDPSMIVDIGDAKCL
1523 RPGLSKGVRVVEKDDRPYPALVVDKSSCFFKAQNLAQS IMELGRQKGRPDMMWKTARFL
1524 FKDVRVISAPVDKGRVKSFP IRAITKMPATELNVKVKGFNGSLADYYARVLKIKLQHPN
1525 FMCVEADVPGPKKEFFPVEVLFVSPNQRVPIEKTEANQSSIVLKANAIKPDRRIKNIKDQ
1526 MSRMSLFANTPVMEAFKVFVCPDFIRLTAGVRVAPQISMGDRDVKIDQKKANWAKEANGA
1527 NYKQESFSLDSWAVLYANTEPGLIRQFVQRYVAAAQRRGFTVREPTILKFNQDFERTFSE
1528 CIDNDIRFLMLIDPKYVKTHESLKLFERLCHVLTQHVSLE RVFDVVQKNSRMTLDNILSK
1529 THMKLGGNLYVPIIENVGSRFALDSGEVLVIGYDVAHPTAMSPQERRLVRSNLNDVKSLE
1530 PSVVGITANCSEFNPHFIGDYHYQTARKESIDVSI LERRMVWIMRLLLEKNRPDQCRPKHV
1531 VILRDGVSEGQYDMARNEEMAALRDGLKLVDP EYNPTFTLVIATKRHNKRFFGQDGRNYV
1532 NTDPGTVIDKTVVRKDVPEFFLQSHYPLQGTVKI PQYNKLHDEANFSMDELQAFVNCLCH
1533 THQIVNLAVSIP EPIYQADELAKRGRNNYAELRRRFTSEVPRMNEAGVIDCNALTKILSY
1534 WDSPLEAVRFTA

1535

1536 >WAGO-1 (R06C7.1) #14 - L596_g20920.t1

1537 MEPSSQPPPTPSLPENMAPKIVGPRDIYERPVIASNLFPLEMKADVPIFMYHVQIHMKI
1538 GAHEINLVKRHTDDYMHIDHKDKCRAAFRFAVRSSPATFGDPKGLFYDLQGLYSDHKLK
1539 DVLGHDLIVREEIGIPGEEAGRAPEFKNL GVEYLRVEVEPTRENRP TMI LGEMMATRAKM
1540 QESVSRELTQFLEVATSQYAF LTPSKMVTYPSGVSYFKPTTAE PVQVFPAAKELLDGVHK
1541 VVKLIDGT KRGE MAVVDPKKA VFHKS NITVIDKTVEMGFLDRGSGTVLRETIPELAKKL
1542 KNLYVETRYGKKPIRF AVHDVVLDTARTSRFNKNGDMTSVEEHFKDEYNVILKHPHAPLV
1543 VSTPLKKNPESLQLLYFPMELLFVCPNQRVLFNQQTAKEAFAIKKASTILPENRLKEVVN
1544 SASKLRINGASVQGCFFKAKIEVGNAPLTV EGRSLVPPHIEYRAQQVQVDAVSGRWRSSS
1545 RRGKPYLVGGKIERWAMYVLSQAPSQAEEELGKNLLAKMEDEFQARGMQIAQAKFLATV
1546 KASPEYMKKIFDRAKAHQLEFLFFIQDTKICLHKEMKYERKYEIISQDLNMETAKAVTE
1547 EGKYHALDSLIAKLNKVG GTNYGLVGPSIPDLFKRGKLYIGFHASFSANPADAEDPTVI
1548 GSSANVTQTSAAFVGMFFQEKNDVDMNTAMAKATLKYVLRVYKNVHGHAPSEVVIYRSGS

1549 SEGQFGQILRDEVPALRNALQNAEAEDEAKLTLLMVNKQHSVRLMPSVIMPGSRAIEQNIK
1550 PGTVVDTKITHPRFAEFYLNHQVLHGHTAKTPKYTIIVDDSAHQIEYLERVITYALSYGHQ
1551 IVDNPTNLPSPLYIAGEYADRGLTLIKAKRKLGD TVDVKDLEKDLPYMASQVLADKRVNA

1552

1553 >WAGO-1 (R06C7.1) #15 - L596_g16096.t1

1554 MSNNERVKTIQNGMETISISATMPVKKKI ANAPGAPLQVATNIYKVGLSQVPIFRYDVDI
1555 TLKLPNGKDVKVVKKDRADHVI IQNKNKACKTFQKAVQKFPNVFGNVELFYDCQSILFSL
1556 RKLNISDKGQEF T LSPNDLPEVYGHVDSVNFIVKSVREKYQLTLNDLGFLSTQEVNGIKH
1557 DLAQFIEVATSQNALFNGKHSVYDGGVSYLMSTGEAVRNEPSKQLITGVQKSVRFIEGQG
1558 KPEATLVLDLAKTVFHKGGQONLYEKAISSVRNWRNDQVDRREIKSLSQQFKGLSIKTEH
1559 QKTKEYELAGLSEFSAHFKFDHNGKQMSVAEYFKKQYNKNLMHPNAPLAIKNFFMGKR
1560 NTMMLPLEICTVLPNQKVSQQETPLQTA AVIRYCAVLP SDRKEQIYTQVKELGFWGNKN
1561 LPITVDQQPIVVTGRQLPQPSIVFGGNQTVSVNPANGKWQATKGVNRNARLPFALPGNPM
1562 PWAVVLVGQGTQDTAKSFALAVKTECESRGLKMGDPSAVIQANYEDIEINARNNTFENLT
1563 KMTPRVKFCLVIENERSPAYIHALIKKNEQNWRITQVTD SATVQQFNFALGPAQGYGKG
1564 QTLNICLKTNVKFGGLNHF IKPPQGHEKVFASDRMYVGLGISHASPI SDAQRARNVKPS
1565 PSVIGISCNYLAHPQALAGTFVVFQEPRENKMVESLKKVFFDLATKFKNIRKVIPEVVIY
1566 RVGASEGQYATILEQEVPIRAGLKEAGCNAKLVLIVPSRTHNVRLFPQT IKKEDKASFQ
1567 NLKPGVVVDSVIVHPKFPEFFLNHCTLOGTANTPRYNVLVDDLQAPI SDHEMATYMWSF
1568 GHEIVGSPTSLPSPAYIADKYAERGRVLAVEARRKDEKFLNEEGEVDFTEMTAKLSFAGT
1569 PLANFRVNA

1570

1571 >WAGO-1 (R06C7.1) #16 - L596_g21757.t1

1572 MSEISVDELSSYISLESMSIGSSSAYHKKAPVPDSSLHHPVELLSSYLEIGIQEGSKAYR
1573 YDVEIEASKSGSLTRGPADDGHGAMRRKVCYDLLYAALAKSKGFGTGQDYHMLPVYNRQN
1574 HVFFAQPLPEDFITIDLDNADFTGITNYLYYMTGATESIQVKISKPQSEEHVLDLYESI
1575 FDTAPLDCDGNLEISSRDRSFRTFLELLTSGPASFTGSHVACGTSFYEATNGKDMKD GKT
1576 MRAGLKKGVCVIERDGILYPALVVDSKTGAFFKEQNLLKSMKEMNNGEVPRSAAPMWQK
1577 ARKLYKDVRVLVVS NVYKNSTQRRLTFPIQDFTREPASRQMMNMKGFRGTVEQYFRQHN
1578 LSLHHPHLP CAI HASNSKI PVPTFFFPIELLFVCGDQKVPLEKSDRFHSETLLRENAVDPK
1579 LRKERTEVQLKKLGLWREKREKMSVDDKKDLSEDGTNLLTAFGVSIINEFIPIRAGV RVA
1580 PTIEVANSEKISIVQKTANWEKKL TNKRYFSSVEITNWAVICSQISAENPMIVRFLKQMV
1581 NVSKRRGIRMDSPMKYSLKSGSREDFDDIFRHIAESGRHFV MYFSP LKEKQHDLVKWMEH
1582 HYSVVTQHVCLERIENVVTGRIQILDNI IHKANMKGGLNVI PRIEKLQRMEIESGDFL
1583 VIAYDVCHPAPMTSRERVLMSM TSFDP SIRS LDPSVVGIVANCVAHPHAFVGDYHYQAS
1584 RKESVDGRILVDRVKWIFELLAERRPNASRPKHIIILRDGVSEGQYKMAEHEELSAIRRA
1585 VAMIDPNYHPTFTLVIATKRHNKRFYDKNEGIAVNTEPGT IIDKDVVRGDVTEFFLQSHF
1586 PLKGTVKMPQYAILCDEADFSQDEIQAFVNCLCHSHQIVASAVSIPEPIYAADELAKRGS
1587 NNFAEHVKI HGRKSLRKDPMNPNLIDFEALTHELAYWKTNLEAIRFNA

1588

1589 >WAGO-1 (R06C7.1) #17 - L596_g18751.t1

1590 MSDQPSSSHQLDVQLRPATII RQDAEKMFKELGIEVPEP IMPARI PVPPNGNVRLVSNFY
1591 PAHVNGQVPIHRYDVEMTIARRNGSQLALTKKSTSDAVSIDRKAKTKAIFSKMVATHPEL
1592 FTNMYSCIYDLESMLFTLKEIEEVNLIIDGLDGEMFQGTTSATVNLKKCHDRYELDLTNY
1593 GFLQADVGSVDLSHKQFLELITSQIPMMSEEFVCFPGGISFGYHVDATPLSDAKNLYHGV
1594 QKSIRYVENPATKKPMAAVAVDLRKTTFHQ SINAFQYMCAQVQVDRNGCLTNASSLTKVS
1595 KAMKGI RCKLTYGRRYREMVIHAVVNKSPRSTFFKRGEQDISVEQYFYEVYQITLRYPNG
1596 LMAAEKKPGQRELCYFPMELLEILDNQRVSGELPQAVVSAVIKQAAVLP AQRRQEIQQAC
1597 NDIGLFDNEYLRNIQT TVEPRPIEIEGR LITNPKLIYGNNVLVDSRKGQWPNRGANKPKF
1598 FLPAVVRKWTFLMISEQRNGQATGTMNGFLKEFIRECQMRGMTLPHSPSPYVLD TNRNVE
1599 DQLEEFMRDCSEGDYEFVFLVLDQDGIKFKHKLKYLERYQVITQDLKTQTGQRCLQRAAA

1600 TLENIVQKTNMKLGGLNYSLQMTNPGGRKSVFDESTMFVGLSMTHGKPLKPDSTGELPPR
1601 PASVVGFAANTLPQQFAFIGDYFQAADRDEKIDSIVPIMTILLTKWSKHHDGQMPMNVV
1602 IFRNGASEGQYKNVLRFEIPLVKYALEKFRFAADVEQIHPETKLCMLVSNKHHSTRIFKT
1603 NVPVQGRAPEQNLEPGIAVDRAIVNPVFQEFYINSHTTLQGTGRVPRYAILNNDAGYALG
1604 HLEHIVFGLAHGHQIVNMTTSLPTPAYIASDYGDRGMVILTQFLREFEKLKGYEHPVDAY
1605 QEFNQSLTYASLDPDCKFNLCRVNA

1606
1607 >WAGO-1 (R06C7.1) #18 - L596_g4755.t1
1608 MLSVVEEQAIHPGRCGKRLQVNSNVLNVRLPASRIYHYHIDVVGHKRCGPKIVLSRSFFND
1609 SLGHERRTVLVNLFNWLAYFNEERIFAQPRNFLFYDAKSPLYTRKKNLCKCGEVVTISTA
1610 QLTEAQIEGLEEFLCVRVKFTEADPFEIVLNEPEEFRDLGGPLQNFLETLMQHSFLKKNY
1611 ESVMNPLIQVAADYEARGIPEVEASTLVKDGSRIFPAIRTKLMNMEGPQALTMIEGLHEA
1612 DHACLCFRWEPPFAVHSSVTLQKARRALEKEGFESFFPFELDALNRELCGIRVFTELSGT
1613 KKYFIVDHSV SERTAWTAVDQGTLKDFLKTEYEVTLKPKDLFLIVEKRGSLDIYHAMELCT
1614 VAPFQORPHKKNLNVPPRFKWDCHQMKNKASPSKHTDHAKRLRESVGLVNNVFLQSGSVTL
1615 ATEPMEVTARILESPLRVKREGLFAVKSKEGWRYPTTHFVHSAKIERWGVCLIFQERM
1616 WCDRYQAHIRLNNLMKVISDRAGGHGMRMAERFQEPYNVPIREGSSLSQIEQVRLCFE
1617 RCKETFDPQFLFFVIQEGIHGLRDCIEAFERKYQIAALDVNNNEALKMALAHCSSRQKSP
1618 MAQVLTSTNIEAEKALRTLMAKINTKMGGNFEVVPNHANRLLLQDGYLFIGIHAFAWY
1619 EQQRATVVGAAANLRYPMFSGDARVRKFDESLPEFLAKIVKRCCIQFNKVRGVDPOHVI
1620 IYQSQPSKDLCLIVDRTAILLGEIGISTELTYVFVDEHHDIRLTNTHLNNTSVIEEQNL
1621 LPGTVIDTHLVRKGASEFFLNSHIGLVGLSEVPKYTAHDFRSGLDGDNLQSLTYTLCYAV
1622 QSLNAPVSVPAPLYVAKNKARHGASCLKLTGKVGARDFVN

1623
1624 >WAGO-1 (R06C7.1) #19 - L596_g20173.t1
1625 MADEVIGRMRQLSICDDPAYALAPKLQRADKKSFVQHVDIVTSYVRINIVGPAKSYRYEV
1626 TIEALAEGREPNILTKGPADDGHAVMRRTVCYFLLYAALDKSGAFGTGLGHELPLVYNGQ
1627 TLVYFAKPLDRDVIQVTLQDQHDFANVPEELLYMVSGNDTVRISLQKALVDSEMDLQTM
1628 FETGQADVENLSTHRSFRFTFLEMATQQAHAHYAQSYSYTSIGNAFFEKDPSRTRSLGDGKII
1629 RPGLTKGVRLIERDGVVYPALVVDARSAAFYKEQNLFLSVKECMDSDGDPNMSMNDKWDR
1630 DNLFRGMSIFLAT

1631
1632 >WAGO-10 (T22H9.3) #1 - L596_g19923.t1
1633 MSFRIPKRRKPEDEGAGGTIPPKQNYSSSSSYASASSSSSRQTSNGGKNASVYMNGFELQI
1634 GRSVEIHKYEIKLFGVFRKRNGDEDKDLTQGSKEAKDDISIQKRRRGCWEIFRAVVREN
1635 NSLFGDRNHRFVYDCGLIFYSIDAI F PETETKTGTDFDVAI LSRECQDY YGQAMK MIEYSI
1636 KKVRDGTFTLGVAPEDKDDRSMQQFLEVLTSQGVYAEGLDNLI FRNHRYDVKYDTPVLRN
1637 KPEY PFCVRHGSSKSVFVAE IENS NKALQ TILQLEKKTSPFFPRMNVLQFVDQSKSDIGE
1638 RIVRGLFVTTTHLKKQKVFVRVADFSKMNCNDITFKMRDRQNEDEFREISVTEYYQEAHKF
1639 STRAGHRPCVVERKKRFNGEKEENNYPMDCLEIMDGQRIMDKKQSGDITAYLISEARVLP
1640 RQMGEIKEELNHRVLNQEAEKYLEAFGVRSRDLLRSDAKILQAPKIA YGDKEKYLTTD
1641 NGQKNAWKIDDALKFYRPGKISDAGGENEQNRWIFAVLNEFQSERDHAAAKRFLGKLQDR
1642 AKLRGMLMLNPEVRCLDVRDPDPTVNKKLSELCQYAKANKVKFIMFIQYERKDMSRDTM
1643 KQLETKFKLTTQQITMSTVNKGAGDKGDRMVLNLI LNKTNEKLGGINCIMKPSQIAQWF
1644 SGNV MYMGLDISHPGLGGNALSSVTP TAVGMTFTKNRDEVQGRYWFQEAREHMLRSLKKQ
1645 IVFAIEEFNRCSRRYPDKIVVYRGGVSEGEYDKVKTDEVEQFMEAFREINFPGKRKPALI
1646 IVIVQRNSGYRLIPTQDNDFRGNDAIVQNVLPGTCAEVI GEGGRKEFILLVPHQAIQGTAK
1647 PSKYVLIYDEAKCITLSELETLTNTLCYSHGIVTSPVSCPSILYQAGDLAKRAINNYRIH
1648 SSRRDFGAMPPIEEVDKRNEYFDQMCMDMLQVTL DTRFWA

1649
1650 >WAGO-10 (T22H9.3) #2 - L596_g15911.t1

1651 MLPNRAPQGNE LQRNRKVELQVNGYMRINESKVYMH EIKMEVAFNTLKG LNRNVDLLARP
1652 ANDVIRQKRRRLIWSIFHATRIKHPQQFLYNEYEYVYDCGGALFALHQIGDGNRIEFVMK
1653 MADFPEEAKGQLHRAEHVILRLAFTRILDLRQSNLYDGGEAGEQRCRFVQQFLDILTSQY
1654 VLGSDQHLVFNQNSRY SADPREDIEAHL SKVIKKGASKTINIVGDQKNQEALLFVEPRRSP
1655 FMADKKVLDI VEEVRRELGGRSSNAQLKKKLEDLLKGIVVETIHQKDAVQFPVKGFSAEP
1656 AGVLSFTMDAGESTTVADYFKRRYHLHVDRDMLCVVCERRQQKFYFPCV LVLVLPQQRVH
1657 CSRQTPKLVEQLIKESQQLPSRMKEEVNHEREVYGFHEL NQQLKAFNVTVDTELCTAVGK
1658 VLPPPVIQYEQRTVSADVDRSGQKITGRQWKVSGQRFVVRPAPTPEKVVLCVFENALESES
1659 TRTFARAYVNAARSHGLILGEP IIERLQEVNQSTIYARGQAYKSN DVKFI LFI FGGDRKN
1660 FERDIMKESETLFNYTTQAVNTKTAMKAI SDRGAFMVL DNLV MKTNLKLGGINHEL ANAQ
1661 DFPQGYLEKVLFKAGRVFI GLDLQSPGMLGGADEFTLDPTVVGMTFTLGS PADMRGTYWY
1662 QPAKKKYI SRLKDAIEDVLMVYQESVGS L PNDIVIYRAGVSEGDILMVVSEEI PAIKDYL
1663 TTLDNPDGCPYRPHL TVMVAQKNSTMRMLPLV VHQTGRAQDENVEPGTSVSTNIVSARHT
1664 EFVLA AQQALKGTARPTRYIVVHEEEGQFTVEQLENMTHQLCYLHGIVALPVS RPSPLYS
1665 ATDLVKRGRANWKVRMERKEGSHTPLGPVNDTFFDPINEERLRFLPVTLPKFWA

1666
1667 >WAGO-11 (Y49F6A.1) #1 - L596_g17524.t1
1668 MNKSSKIAAMP SFKKRPAENDDSPAPKRPAEKARRS QREEAPRRSHGGSKILMNGFRVE
1669 IEKAMTIHKYYIQLNGIFKKRGGEEIARDLTEGVHRDDVSMQRKRRCWDVFRQIVKENE
1670 SLFGGNTHKFVYDCGRLFCSMEEIFPKSETKTGTVDLATLPDRSQTFYRGALRIEWTIKK
1671 VEDGTFKLGVPPEARS SDRSAQQFLEILTSQGLYARGDDHLIFRNQRYDVQENAPVDKKS
1672 TYPFCVRQGVQKSVILCESAEKIATILQLEKKTSPFFPEMNLLDFVRQCSSDDKAMAVLK
1673 GLQVQTTHLKRQOKTFRISGYSNTACKDIHFENRDGEKISVPEYFLKHHA FRTAAGQLPC
1674 VEEKRKPQNNHYPMDC LKIVGGQRILS QKQPEIVEHLI STARILPLKMADEIGEQLGKH
1675 IILNREAE TFLRTFKVNVDRN LLETEATLIKAPQIQYGNQNGV NPAKTLTTERGQKNAWK
1676 MEDSLTFYRPGVVS DSGEAHRWMFVILNEPNQREHGDCRVFLEKFVARAGTRGIKIQFPH
1677 VEKKRIENSDPWPELQEI GQYAKSNGVKFVMFVYERTGDIRKAMK LLETTFALTTQHVSL
1678 KTISKAAGDKGAFMVL DNLMLKTNEKLGGLNTRVKAEP RVAEWF ERGTTMFMGFDVSHPG
1679 LGARNENGVTPTAVGMSFTKNSDLEVVGRLWYQEPREHLI PNMKDHI VEALETFKKHSGK
1680 FP ELVVVFRGGVSEGEYEVQTKEVQEFQDAFQQLKFSRKPILKIVVVQRNSGYRLMPAQ
1681 RNDFGYQKNEALVQNVVPGTCADAEIVDQKRTEFV LVP HQAIQGTAKPSKYVLLHDEAPK
1682 MSKEELSTIAHTLCFMHGIVTSPVSCPSILYQAGDLAKRASCNFKAFLNRKGGNVPIPPV
1683 EEKEKRKEFFDALCEK LKITLDTRFWA

1684
1685 >WAGO-11 (Y49F6A.1) #2 - L596_g17422.t1
1686 MPFPRGKGYHPYRGKSYHRGFGRNNFYRQTKREEYHSSDSSPTDNQPSTSRSHDRCDY
1687 KPKDEDDMKLYLSDRKPSPEPPEFQIRINAFPFNIDHAPKEVHTYELIFVMSQKLKKEKE
1688 PANHKLRTNAKL FSAEDIGLGFDAEEEGQVYHGTDMSCGPTDVVRRQRRKALLFQLFRH
1689 LINQSKEYFPGSKYVYAYDGQRILYSPEVLKMD EGMFMAQLTDLPE SVTQLLGP SDQENT
1690 EINAYIRKAEDVIDLHDFGTEARPIHGIEGFLETLTWQHAFEGFEEHIVYGTRYFALNAG
1691 KKLKHCTGFKSIVGF EKRIELLPDYTC SNILLERSLRPVMRMTPKFDLFFDNSTAI SLDD
1692 FAVLFFRVEVDDL SATLQKKENLQKLNQVFKNAVVRTIHRDDGRQDTFMI DHLDERNPFE
1693 ITLKEEDEETVADYLYGVYGYKVDNRDLPCVARKFKNEFAYYPLRTLVL LPNQKVASKF
1694 LPEDMRKSYQEACQNLPSDALNNIFTAMSQNLNLSRASEDYNND EKVIHVNNKYMENFKIT
1695 LESNQLIQMPAQRICEPRIAYRESTH LAQDGAWDYEKAAVFIDAVIKVRRIGLVNTCEDV
1696 GEDDLGEFISKLIGWLRNKTIDLKLTPEDI FWWPDAWNEFKAGKSKKEM LATAEKI IETS
1697 EVKLMFVICSGSADDEIHDIWKLA EVTNELVKV NKKDFVTTQCITPATLSNLV LKSTYQD
1698 EILTNIIMKMN LKLG GTNYVLSKSPSNEMSIPH IHS SRMFV GIDVLQPEKTQLNGEPTN
1699 NPTVVGISFTDATSKFYLRGTYWYQQSPTVSLCLLQKHFDEALYWDCHKMDSVPREVFV
1700 YWRDSRIQKNMEEVKMVLEDVICNRAKDVRRDASKLFLIMVDTKPKTR LFTWETQFSGNA
1701 QTQNVQAGTFVRESYRKRQFTMINHKS GAGLAHPVRFTMINDDVNEKDYVEAELEKTTNA

1702 LCFLQNTSTRSTSIPAPLYSAMD LAKRGMKNYETMDAVMREEERDEDRKKREARTPEAWH
1703 RYYKQLVKTHMSVMPIRD SKFWA
1704
1705 >WAGO-11 (Y49F6A.1) #3 - L596_g18655.t1
1706 MSPPRNGSNFN RGNHGRPPHGHNSQAKRVKWEHSSDSSPIYNQPSTSRSYDRQPKEEN
1707 DVKPLFSSRERS PRLRLDLQIRINAFPFNLDHAPKEVYTYELIFVMSQK LKREKEPENYKI
1708 RTNAKLLAEADVGLGWFEAE EPGQFYHGTDMSCGPM DVVRRQKRKALLFQLFRHLINQSK
1709 EYFPGSKYVYAYD GQRILYSPEVLKMDEGMFMAQLADLPESVTQFLGLDDQKNTEINAYI
1710 KKSDEVIDLHDFG TESKPIHGIEGFLETLTWQHAFEGFDKHIVYGTRDFALNAGK LKHC
1711 TGFKSIVGFEKKI ELLPDYTSNMI LERRIHPVMRMTPKFDLFFDNST AISLEDFAVLFF
1712 RVEVENLPATLQ KEENLQKLNQVFKNAVVRTNHRDDGRQDTF MIDHLDKRNPF EIIVTEE
1713 DQETVADYLYDV YSYKVDRNDRLPCVARRFRNELAYYPLRTLVL LPNQKVASKFLSEDMR
1714 KSFQEACQNLPS DALNNIFTAMSQNLNLSRASEDYNND EEVTHVNNEYMENFKITLESNQL
1715 IQISANRVHEPQ IAYQKSVKPAQDGAWAYEKGAFFVHPKKGVRK IGLVNTCEDVKEDVLG
1716 EFISKLIGWLRNK DIDLKLTKEDI FWWPDAFSDFGVMKSVKEMLAKAEKI LETSAVNHMI
1717 VICNGSAEDKTH DVWKLAEVTNGLAKKDRTNFVTTQCITPTTLG SILVKSTYQDEILTSV
1718 IMKMNLKLG GTNYVLTNSNRNYMSVPHITQDRMFVGI AVLQPEKTRLNGDATYNPTVVGL
1719 SYSEGSPNFYLRG TYWYQQSPSVDLSILKKTFAEALYRFDKFHLIPKEIFVFWRS GKFQR
1720 SMEEEKIALQEV IDKRVKDVNAKSPKLFII SVNTKPKTRFF TWETKFSGNAQTQNVQAGT
1721 FVQESFLRREFT MINHKSQAGLAHPVRFTMLNDEIGE QDYAEAEIERTTNALCFLQNNST
1722 RSTAVPAPLYSAMD LAKRGMKNYETMDAVIREEESE EERKKRETRTPEGWQKY YGNLMER
1723 HMPVPIKSSKFWA
1724
1725 >WAGO-2 (F55A12.1) - L596_g16917.t1
1726 MHAHRLQQLTDG INRNLNDEGIARRTPMSPTPSVGRHDKEITL TSNLYELHLAHGKVRVY
1727 HYTIKLSDYTGK TAEVHG DYGRVNLGSKLLLAADTFM KRNGFRDHDFFCDLAGMRLYTIK
1728 PIPRSEGNPRFYV SEATKDG NFAVCRDDGRPFELNIRDVFKEVRI PNLQDATLDAFIN TA
1729 INKACLAKSDLY FPIFETCYPKTGAPVARKDGRSLILG TKTSVERVEGRYGSSTTPVVAL
1730 NVSAEFWYMKKNL LDFCKQNIELNEKSVPVDAASFQL LSDSMQGVTVRLICSKNPLLFTI
1731 KSLANKNVSM LKYPLPNIKGTGLIKFLWETYSVRLTY PESFAAEVKPDFHVTDPRPIFY P
1732 VELLEIMPMQRAL KGKSCDGS AVDREKKIQEKVHQMERHVQKAGLGLSMDDAPVEVEAG
1733 VLDLPKIVFADEK VVEVNPSSASWRFGSGE CPERFARPAEVKELPWCVMLVSDVPPTKGM
1734 SKKAKFFADLLK KQAAERGLQMK EPMYHPTKQ GKVEIERFFNTTAVEFFVFLMAKSLDYH
1735 DFTKILERKYQV ITQTVKMENAFDVVEDPNSTKSRKIVEHIVMKMNLKLG VNSTVKPSR
1736 HLF SHEEVRRL LIGFTLIEGPKITGRDAAAFNRFGGKI PAVVGFSANMG SLEHEFLGDF T
1737 FQYLDHTNIVQD IKKIVATILERFRKTRRGRDPAE VVIYRKLDEFHFPRV IQNEIAPLKN
1738 LLEEKCHGFVSLI YIAMTKTHNIRFFPKGPPPEG DERKPNLVPGTVIDSGAVGGGLKQFF
1739 IASYSASTGTT RPPRFTILENSKEAKIRDLEKLTLELTF AHQTSTRSLGVPAPLVVAKNY
1740 ARRGVNLAKNEAEKIS DVDTSVG VETMIDRLPYADAPVLRDKRVTA
1741
1742 >WAGO-5 (ZK1248.7) #1 - L596_g12936.t1
1743 MEGEGTEL RPSTVVRQARNAAASSAGAPPAAPGSRAATASVEKSFYSKLVNVPTPPPEEK
1744 KPHGTAGRQLKLR TNVYGLSLPKDVQVFRYSVDASGTLQRNDLRIEFAKRVANDITYLNR
1745 REKCRVIDQV VAKYSAIFGDRRELFWYDSQSILFSRNQLDISSEGQFVLDQSDIGQNPL
1746 FEGFAHLKMVIRPA QTNFAVSIGDLEAYIQAEV FESDHALQQFLEILTAQYAFNTPLEAM
1747 SFGSRTAYLLN PEKYGFKPADCADVGDGKFLGVGCDKSVRFIEGPGGAGSQR AALVVDLK
1748 KTAFHKDQSLY EKAREILNNRDPKST DASRLRMQLKGIVVETKHGSR RQEFVNDVVADT
1749 PATKKFKDLTGQ EVTLQQYFQQKYNITLQHPD SPIVLTDRTKKFAAFPMEVCSVVDGQRV
1750 TLAQQTPVQIQMIRQC AVPPADRQRQILGLVQGLQLNSENKYHKAASV GITPTALQVQA
1751 RLLQNPTIVYGRNSTMKPDEKATWRLARQKPVYLKPAKV DKWAMFVICGGNRSDCVDQDI
1752 LNQFSNMVQECRARGMTVSDPTGFSFIGASREVVQETLEKAKTEGNQFCFFITNNDVTN

1753 IHQFMKFQERKLSIVTQDMKMSAFAFDVVRKGRQTLNENVNKTNMKNNGGVNYSLRFDDPA
1754 FSMEKLLPKDRLVIGLATTHPKPIVVKKEQDEGPHDKKKQMHQQRTPPVPSVVGVAANA
1755 LTESIEIVGDCLFQQPNREEKIALLOPVIRSLMLQFMKHRGMPPAEIVVYRQGTSEGOFR
1756 DVMELEYKMOVKAAALQQGLNPKITFIVVQKMHNVRLMPMDSKAGDKAPEQNVKPGTVVDT
1757 MVTHPKYNEFFLNHVALQGSARTPRYTPLYDENRLLPMDEIEALSHSLAFGHQIVNLTTT
1758 LPAPLYIANRYAERGHNIFIASQEDYTKSKTSFQSPHSTTIEGNLDFGRMMNELSYCNSE
1759 LKDKRVNA

1760

1761 >WAGO-5 (ZK1248.7) #2 - L596_g12936.t1

1762 MEGEGTELRPSTVVRQARNAAASSAGAPPAAPGSRAATASVEKSFYSKLVNVPPTPPPEEK
1763 KPHGTAGRQLKLRTNVYGLSLPKDVQVFRYSVDASGTLQRNDLRIEFAKRVANDITYLNR
1764 REKCRVIDQVVAKYSAIFGDRRELFWYDSQSILFSRNQLDISSEGQFVLDQSDIGQNPL
1765 FEGFAHLKMOVIRPAQTNFAVSIGDLEAYIQAEVFESDHALQQFLEILTAQYAFNTPLEAM
1766 SFGSRTAYLLNPEKYGFKPADCADVGDGKFLGVGCDKSVRFIEGPGGAGSQRAALVVDLK
1767 KTAFHKDQSLYEKAREILNNRDPKSTDASRLRMQLKGIIVVETKHGSRRQEFVADNVVADT
1768 PATKKFKDLTGQEVTLQQYFQQKYNITLQHPDSPIVLTDRTKKFAAFPMEVCSVVDGQRV
1769 TLAQQTTPVQIQKMIRQCAVPPADRQRQILGLVQGLQLNSENKYHKAASVGITPTALQVQA
1770 RLLQNPTIVYGRNSTMKPDEKATWRLARQKPVYLLKPAKVDKWAMFVICGGRNSDCVDQDI
1771 LNQFSNMVQECRARGMTVSDPTGFSFIGASREVVQETLEKAKTEGNQFCFFITNNDVTN
1772 IHQFMKFQERKLSIVTQDMKMSAFAFDVVRKGRQTLNENVNKTNMKNNGGVNYSLRFDDPA
1773 FSMEKLLPKDRLVIGLATTHPKPIVVKKEQDEGPHDKKKQMHQQRTPPVPSVVGVAANA
1774 LTESIEIVGDCLFQQPNREEKIALLOPVIRSLMLQFMKHRGMPPAEIVVYRQGTSEGOFR
1775 DVMELEYKMOVKAAALQQGLNPKITFIVVQKMHNVRLMPMDSKAGDKAPEQNVKPGTVVDT
1776 MVTHPKYNEFFLNHVALQGSARTPRYTPLYDENRLLPMDEIEALSHSLAFGHQIVNLTTT
1777 LPAPLYIANRYAERGHNIFIASQEDYTKSKTSFQSPHSTTIEGNLDFGRMMNELSYCNSE
1778 LKDKRVNA

1779

1780 >WAGO-5 (ZK1248.7) #3 - L596_g17875.t1

1781 MVCKGQATSEIELTTNAYGFLPFLSAKVQYDVEIVGILSGTGRTVNFTKCSKHDAFRAA
1782 RIEECRDLFEMVKQKYPEVFNAPDSNYFYDNGRRLFTKESLLPPLVSKQEFPLNEFDTVY
1783 LDRDYSRFDKILFSVEKAAEDPMDVKEVLKHVKDSGKSIRLNQFLNVLTSQHALSNPVKF
1784 ATYRIGSAFFINADRYDHNHGVEDLTDDEKIRVGCESVKLIAGSSDDGSAIALVDIKRT
1785 AFHKSGGTLKAREVLGKWPQPCDANRLKPHFIDLAVYTQHGSKDRRYVIDDVIAETPA
1786 TLKFAWVRGSKELTLVEYFHQAYSIEIKFPRTPLAVAMAKEGRTIYLPLELCFVSPHQRV
1787 TTAQQQATEEMIKRCSIPPLDRQKRIGRIVEAMKISDNPFLOIADTGCKLLSTIPLSVTG
1788 RVIAPPKIRYGHSEIQKSAFLKPAKINSWAIIVLTAQDDPELARGDILSPSVLTKFARLF
1789 RKECKARGMQLPDPFLKEFMKADVDQLGELMRCLAQGDSTECRPPFLRFIFVTNEKLTDL
1790 HHPMKYFERQCGIITQDMKMQTVVDVVLHKKRQILENIVSKANIKNGGLNYSVVIIPDVPG
1791 KRPILGSGRLIVGLFTSPTFKWQFEDSPSRPTALGYAANTTPNEGEFIGDCLIQKGRASA
1792 IQTILRRVLAEFKKQRRCDPADVIIYRSCEEGREKKTLEEDLAAVRSILKSSNPMPTLTV
1793 IAVQKRHGLRLMPTAIQRQGEPOSENLPKPGTVLDSCLTDPALTEFYLNHSHATLQGTARTP
1794 KYTVVHNDVGLSLEEMETLTALSFSHQIVPLPTSLPSPLYIAGTYAERGVSLYQQDRER
1795 GREGSLESCFTYGSPPGLKHLRITA

1796

1797 >WAGO-5 (ZK1248.7) #4 - L596_g24399.t1

1798 MDVLTEAMSKMLPMNIAPKIVGAQDPYERVVPLTANMFPLHMRAEVPIFMYNVQVFMKVG
1799 FREVNLVKRNTDDFTIIDHKNKCRSAFRFAVRAAPQVFGHPSGLFYDIIQAQLYSVRELKD
1800 VLGNDLKKKEEIIIVPGEDARKAYDFQDIDLEYLRLVIEPVNGTNP SINL GELVLKQKNFS
1801 DEVPCCELLQFLDVACSQHAFLLPTKFTTYPGGFAYFNPTSEEPARELPDAARLHNGVHKS
1802 VKVIEGCTAGRCGELAVVLDPKKAAFHKPDI TVVQKI QEMGFLQMASENVAPHRIPELA
1803 EALKNVFEVETRHGKRRSRFAIHSVVAESARTNRFTKDDGQVTVEEHFKKEYDIALKYPHL

1804 PLVVSAMPLRKKTPSNGRAPPRLFFPMEVLFICPNQRVLRNQOSAKQNNEVIKSCAVAPE
1805 HRLKDVIASGQKMRINGPNVHGCLNSAQIQVESEPLKVEGRTLVPNNIDYKGCQVQVDSF
1806 TGKWRNFGRNKPHYLEGGKIGRWGLYVLSKAPSSEEEQLALKFKDKMLVEFQSRGMQIDL
1807 PMFLATVKATPLYLRTIFEKARKERLEFLFFIQDKDLALHNEMKFYERAYEVITQDLRTD
1808 TARAVIEQGKNLSLENI IAKLNVKVGGTNYSVNGPSVPDLFKKGRLYIGLQASTNGPPAA
1809 GAHLPTVVGSAANVTTAPSSFVGDIFYQKFGEMDLQGAMASTTEGYVKRYAAVHGRAPDE
1810 VFIYRSGTANTNIGQMLRDEVPAIRCALKNSGASRARLTLVMVTKQHNVRMLPTNMTLGG
1811 RAIDQNIKPGTVVDQKITHPRFAEFYLNHQALHGSAKTPKYVVVADDCSNPIQYLERVT
1812 YALSYGHQIVGMPTSLPSPVYIAGKYAERGAALLQTKRNLGGALDVDALAEELAYANSKV
1813 LGFKRINA
1814