

A robust role for motor cortex

Gonçalo Lopes^{1,2,*} Joana Nogueira^{1,2} Joseph J. Paton¹
Adam R. Kampff^{1,2}

¹Champalimaud Neuroscience Programme, Champalimaud Centre for the
Unknown, Lisbon, PT

²Sainsbury Wellcome Centre, University College London, London, UK

*Correspondence: Gonçalo Lopes, Champalimaud Neuroscience Programme,
Champalimaud Centre for the Unknown, Av. de Brasília s/n, Doca de Pedrouços,
1400-038, Lisbon, Portugal. *email*: goncalo.lopes@neuro.fchampalimaud.org

June 14, 2016

Abstract

1
2
3
4
5
6
7
8

The role of motor cortex in the direct control of movement remains unclear, particularly in non-primate mammals. More than a century of research using stimulation, anatomical and electrophysiological studies has implicated neural activity in this region with all kinds of movement. However, following the removal of motor cortex, or even the entire cortex, rats retain the ability to execute a surprisingly large range of adaptive behaviours, including previously learned

9 skilled movements. In this work we revisit these two conflicting views
10 of motor cortical control by asking what the primordial role of mo-
11 tor cortex is in non-primate mammals, and how it can be effectively
12 assayed. In order to motivate the discussion we present a new assay
13 of behaviour in the rat, challenging animals to produce robust re-
14 sponses to unexpected and unpredictable situations while navigating
15 a dynamic obstacle course. Surprisingly, we found that rats with motor
16 cortical lesions show clear impairments in dealing with an unexpected
17 collapse of the obstacles, while showing virtually no impairment with
18 repeated trials in many other motor and cognitive metrics of perfor-
19 mance. We propose a new role for motor cortex: extending the ro-
20 bustness of sub-cortical movement systems, specifically to unexpected
21 situations demanding rapid motor responses adapted to environmental
22 context. The implications of this idea for current and future research
23 are discussed.

24 1 Introduction

25 Since its discovery 150 years ago, the role of motor cortex has been a topic of
26 controversy and confusion [1, 2, 3, 4]. Here we report our efforts to establish
27 a teleology for cortical motor control. Motor cortex may play roles in “un-
28 derstanding” the movements of others [5], imagining one’s own movements
29 [6], or in learning new movements [7], but here we will focus on its role in
30 directly controlling movement.

31 **Stimulating motor cortex causes movement; motor cortex is active** 32 **during movement**

33 Motor cortex is broadly defined as the region of the cerebral hemispheres
34 from which movements can be evoked by low-current stimulation, following
35 Fritsch and Hitzig’s original experiments in 1870 [8]. Stimulating different
36 parts of the motor cortex elicits movement in different parts of the body, and
37 systematic stimulation surveys have revealed a topographical representation
38 of the entire skeletal musculature across the cortical surface [9, 10, 11]. Elec-
39 trophysiological recordings in motor cortex have routinely found correlations
40 between neural activity and many different movement parameters, such as
41 muscle force [12], movement direction [13], speed [14], or even anisotropic
42 limb mechanics [15] at the level of both single neurons [12, 16] and pop-
43 ulations [13, 17]. Determining what exactly this activity in motor cortex
44 controls [18] has been further complicated by studies using long stimulation

45 durations in which continuous stimulation at a single location in motor cor-
46 tex evokes complex, multi-muscle movements [19, 20]. However, as a whole,
47 these observations all support the long standing view that activity in motor
48 cortex is involved in the direct control of movement.

49 **Motor cortex lesions produce different deficits in different species**

50 What types of movement require motor cortex? In humans, a motor cortical
51 lesion is devastating, resulting in the loss of muscle control or even paraly-
52 sis; movement is permanently and obviously impaired [21]. In non-human
53 primates, similar gross movement deficits are observed after lesions, albeit
54 transiently [9]. The longest lasting effect of a motor cortical lesion is the
55 decreased motility of distal forelimbs, especially in the control of individual
56 finger movements required for precision skills [9, 22]. But equally impressive
57 is the extent to which other movements fully recover, including the ability
58 to sit, stand, walk, climb and even reach to grasp, as long as precise finger
59 movements are not required [9, 22, 23]. In non-primate mammals, the ab-
60 sence of lasting deficits following motor cortical lesion is even more striking.
61 Careful studies of skilled reaching in rats have revealed an impairment in paw
62 grasping behaviours [24, 25], comparable to the long lasting deficits seen in
63 primates, but this is a limited impairment when compared to the range of
64 movements that *are* preserved [24, 7]. In fact, even after complete decor-
65 tication, rats, cats and dogs retain a shocking amount of their movement
66 repertoire [26, 27, 28]. If we are to accept the simple hypothesis that motor

67 cortex is the structure responsible for “voluntary movement production”, then
68 why is there such a blatant difference in the severity of deficits caused by mo-
69 tor cortical lesions in humans versus other mammals? With over a century
70 of stimulation and electrophysiology studies clearly suggesting that motor
71 cortex is involved in many types of movement, in all mammalian species,
72 how can these divergent results be reconciled?

73 **There are anatomical differences in corticospinal projections be-**
74 **tween primates and other mammals**

75 In primates, the conspicuous effects of motor cortical lesion can also be pro-
76 duced by sectioning the pyramidal tract, the direct monosynaptic projection
77 that connects motor cortex, and other cortical regions, to the spinal cord [29,
78 30]. The corticospinal tract is thought to support the low-current movement
79 responses evoked by electrical stimulation in the cortex, as evidenced by the
80 increased difficulty in obtaining a stimulation response following section at
81 the level of the medulla [31]. In monkeys, and similarly in humans, this fibre
82 system has been found to directly terminate on spinal motor neurons respon-
83 sible for the control of distal muscles [9, 32]. However, in all other mammals,
84 including cats and rats, the termination pattern of the pyramidal tract in the
85 spinal cord largely avoids these ventral motor neuron pools and concentrates
86 instead on intermediate zone interneurons and dorsal sensory neurons [33,
87 34]. Furthermore, in humans, the rubrospinal tract—a descending pathway
88 originating in the brainstem and terminating in the intermediate zone—is

89 degenerated compared to other primates and mammals [35], and is thought
90 to play a role in compensating for the loss of the pyramidal tract in non-
91 human species [36, 23]. These differences in anatomy might explain the lack
92 of conspicuous, lasting movement deficits in non-primates, but leaves behind
93 a significant question: what is the motor cortex actually controlling in all
94 these other mammals?

95 **What is the role of motor cortex in non-primate mammals?**

96 In the rat, a large portion of cortex is considered “motor” based on anatomical
97 [37], stimulation [37, 11] and electrophysiological evidence [38]. However, the
98 most consistently observed long-term deficit following motor cortical lesion
99 has been an impairment in supination of the wrist and individuation of digits
100 during grasping, which in turn impairs reaching for food pellets through a
101 narrow vertical slit [24, 25]. Despite the fact that activity in rodent motor
102 cortex has been correlated with movements in every part of the body (not just
103 distal limbs) [39, 40], it would appear we are led to conclude that this large
104 high-level motor structure, with dense efferent projections to motor areas in
105 the spinal cord [33], basal ganglia [41, 42], thalamus [43], cerebellum [44]
106 and brainstem [45], as well as to most primary sensory areas [46, 47], evolved
107 simply to facilitate more precise wrist rotations and grasping gestures. Maybe
108 we are missing something. Might there be other problems in movement
109 control that motor cortex is solving, but that we may be overlooking with
110 our current assays?

111 **A role in modulating the movements generated by lower motor**
112 **centres**

113 A different perspective on motor cortex emerged from studying the neural
114 control of locomotion, suggesting that the corticospinal tract plays a role
115 in the *adjustment* of ongoing movements that are generated by lower motor
116 systems. In this view, rather than motor cortex assuming direct control over
117 muscle movement, it instead modulates the activity and sensory feedback in
118 spinal circuits in order to adapt a lower movement controller to challenging
119 conditions. This idea that the descending cortical pathways superimpose
120 speed and precision on an existing baseline of behaviour was also suggested
121 by lesion work in primates [36], but has been investigated most thoroughly
122 in the context of cat locomotion.

123 It has been known for more than a century that completely decerebrate
124 cats are capable of sustaining the locomotor rhythms necessary for walking
125 on a flat treadmill utilizing only spinal circuits [48]. Brainstem and midbrain
126 circuits are sufficient to initiate the activity of these spinal central pattern
127 generators [49], so what exactly is the contribution of motor cortex to the
128 control of locomotion? Single-unit recordings of pyramidal tract neurons
129 (PTNs) from cats walking on a treadmill have shown that a large proportion
130 of these neurons are locked to the step cycle [50]. However, we know from the
131 decerebrate studies that this activity is not necessary for the basic locomotor
132 pattern. What then is its role?

133 Lesions of the lateral descending pathways (containing corticospinal and

134 rubrospinal projections) produce a long term impairment in the ability of
135 cats to step over obstacles [51]. Recordings of PTN neurons during locomotion
136 show increased activity during these visually guided modifications to the
137 basic step cycle [52]. These observations suggest that motor cortex neurons
138 are necessary for precise stepping and adjustment of ongoing locomotion to
139 changing conditions. However, long-term effects seem to require complete
140 lesion of *both* the corticospinal and rubrospinal tracts [51]. Even in these
141 animals, the voluntary act of stepping over an obstacle does not disappear
142 entirely, and moreover, they can adapt to changes in the height of the obsta-
143 cles [51]. Specifically, even though these animals never regain the ability to
144 gracefully clear an obstacle, when faced with a higher obstacle, they are able
145 to adjust their stepping height in such a way that would have allowed them
146 to comfortably clear the lower obstacle [51]. Furthermore, deficits caused by
147 lesions restricted to the pyramidal tract seem to disappear over time [53],
148 and are most clearly visible only the first time an animal encounters a new
149 obstacle [53].

150 The view that motor cortex in non-primate mammals is principally re-
151 sponsible for adjusting ongoing movement patterns generated by lower brain
152 structures is appealing. What is this modulation good for? What does it
153 allow an animal to achieve? How can we assay its necessity?

154 **Towards a new teleology; new experiments required**

155 It should now be clear that the involvement of motor cortex in the direct
156 control of all “voluntary movement” is human-specific. There is a role for
157 motor cortex across mammals in the control of precise movements of the
158 extremities, especially those requiring individual movements of the fingers,
159 but these effects are subtle in non-primate mammals. Furthermore, what
160 would be a devastating impairment for humans may not be so severe for
161 mammals that do not depend on precision finger movements for survival.
162 Therefore, generalizing this specific role of motor cortex from humans to all
163 other mammals would be misleading. We could be missing another, more
164 primordial role for this structure that predominates in other mammals, and
165 by doing so, we may also be missing an important role in humans.

166 The proposal that motor cortex induces modifications of ongoing move-
167 ment synergies, prompted by the electrophysiological studies of cat locomo-
168 tion, definitely points to a role consistent with the results of various lesion
169 studies. However, in assays used, the ability to modify ongoing movement
170 generally recovers after a motor cortical lesion. What are the environmental
171 situations in which motor cortical modulation is most useful?

172 Cortex has long been proposed to be the structure responsible for inte-
173 grating a representation of the world and improving the predictive power of
174 this representation with experience [54, 55]. If motor cortex is the means by
175 which these representations can gain influence over the body, however subtle
176 and “modulatory”, can we find situations (i.e. tasks) in which this cortical

177 control is required?

178 The necessity of cortex for various behavioural tasks has been actively
179 investigated in experimental psychology for over a century, including the
180 foundational work of Karl Lashley and his students [56, 57]. In the rat, large
181 cortical lesions were found to produce little to no impairment in movement
182 control, and even deficits in learning and decision making abilities were diffi-
183 cult to demonstrate consistently over repeated trials. However, Lashley did
184 notice some evidence that cortical control may be involved in postural adap-
185 tations to unexpected perturbations [56]. These studies once again seem to
186 recapitulate the two most consistent observations found across the entire mo-
187 tor cortical lesion literature in non-primate mammals since Hitzig [8], Goltz
188 [26], Sherrington [58] and others [59, 28]. One, direct voluntary control over
189 movement is most definitely not abolished through lesion; and two, certain
190 aspects of some movements are definitely impaired, but only under certain
191 challenging situations. The latter are often reported only anecdotally. It
192 was this collection of intriguing observations in animals with motor cortical
193 lesions that prompted us to expand the scope of standard laboratory tasks
194 to include a broader range of motor control challenges that brains encounter
195 in their natural environments.

196 In the following, we report an experiment that was designed to provide
197 controlled exposure of animals to more naturally challenging environments.
198 The results of this experiment have led us to formulate a new teleology for
199 cortical motor control that we will present in the discussion.

200 **2 Experiment Introduction**

201 In the natural world, an animal must be able to adapt locomotion to any
202 surface, not only in anticipation of upcoming terrain, but also in response
203 to the unexpected perturbations that often occur during movement. This
204 allows animals to move robustly through the world, even when navigating a
205 changing environment. Testing the ability of the motor system to generate
206 a robust response to an unexpected change can be difficult as it requires
207 introducing a perturbation without cueing the animal about the altered state
208 of the world. Marple-Horvat and colleagues built a circular ladder assay for
209 cats that was specifically designed to record from motor cortex during such
210 conditions [60]. One of the modifications they introduced was to make one
211 of the rungs of the ladder fall unexpectedly under the weight of the animal.
212 When they recorded from motor cortical neurons during the rung drop, they
213 noticed a marked increase in activity, well above the recorded baseline from
214 normal stepping, as the animal recovered from the fall and resumed walking.
215 However, whether this increased activity of motor cortex was necessary for
216 the recovery response has never been assayed.

217 **3 Results**

218 To investigate whether the intact motor cortex is required for the robust
219 control of movement in response to unexpected perturbations, we designed a
220 reconfigurable dynamic obstacle course where individual steps can be made
221 stable or unstable on a trial-by-trial basis (Figure 1, also see Methods). In
222 this assay, rats shuttle back and forth across the obstacles, in the dark, in
223 order to collect water rewards. We specifically designed the assay such that
224 modifications to the physics of the obstacles could be made covertly. In this
225 way, the animal has no explicit information about the state of the steps until
226 it actually contacts them. Water deprived animals were trained daily for 4
227 weeks, throughout which they encountered increasingly challenging states of
228 the obstacle course. Our goal was to characterize precisely the conditions
229 under which motor cortex becomes necessary for the control of movement,
230 and this motivated us to introduce an environment with graded levels of
231 uncertainty.

232 We compared the performance of 22 animals: 11 with bilateral ibotenic
233 acid lesions to the primary and secondary forelimb motor cortex, and 11
234 age and gender matched controls (5 sham surgery, 6 wild-types). Animals
235 were given ample time to recover, 4 weeks post-surgery, in order to specifi-
236 cally isolate behaviours that are chronically impaired in animals lacking the
237 functions enabled by motor cortical structures. Histological examination of
238 serial coronal sections revealed significant variability in the extent of dam-

239 aged areas (Figure 2), which was likely caused by mechanical blockage of the
240 injection pipette during lesion induction at some sites. Nevertheless, volume
241 reconstruction of the serial sections allowed us to accurately quantify the size
242 of each lesion, identify each animal (from Lesion A to Lesion K; largest to
243 smallest), and use these values to compare observed behavioural effects as a
244 function of lesion size.

245 During the first sessions in the “stable” environment, all animals, both
246 lesions and controls, quickly learned to shuttle across the obstacles, achieving
247 stable, skilled performance after a few days of training (Figure 3). Even
248 though the distance between steps was fixed for all animals, the time taken to
249 adapt the crossing strategy was similar irrespective of body size. When first
250 encountering the obstacles, animals adopted a cautious gait, investigating
251 the location of the subsequent obstacle with their whiskers, stepping with
252 the leading forepaw followed by a step to the same position with the trailing
253 paw (Video 1: “First Leftwards Crossing”). However, over the course of only a
254 few trials, all animals exhibited a new strategy of “stepping over” the planted
255 forepaw to the next obstacle, suggesting an increased confidence in their
256 movement strategy in this novel environment (Video 1: “Second Leftwards
257 Crossing”). This more confident gait developed into a coordinated locomotion
258 sequence after a few additional training sessions (Video 1: “Later Crossing”).
259 The development of the ability to move confidently and quickly over the
260 obstacle course was observed in both lesion and control animals (Video 2).

261 In addition to the excitotoxic lesions, in three animals we performed larger

262 frontal cortex aspiration lesions in order to determine whether the remaining
263 trunk and hindlimb representations were necessary to navigate the elevated
264 obstacle course. Also, in order to exclude the involvement of other corti-
265 cospinal projecting regions in the parietal and rostral visual areas [61], we
266 included three additional animals which underwent even more extensive cor-
267 tical lesion procedures (Figure 4A,B, see Methods). These *extended* lesion
268 animals were identified following chronological order (from Extended Lesion
269 A to Extended Lesion F; where the first three animals correspond to frontal
270 cortex aspiration lesions and the remaining animals to the more extensive
271 frontoparietal lesions). In these extended cortical lesions, recovery was found
272 to be overall slower than in lesions limited to the motor cortex, and animals
273 required isolation and more extensive care during the recovery period.

274 Nevertheless, when tested in the shuttling assay, the basic performance of
275 these extended lesion animals was similar to that of controls and animals with
276 excitotoxic motor cortical lesions (Figure 4C). Animals with large frontopari-
277 etal lesions did exhibit a very noticeable deficit in paw placement throughout
278 the early sessions (Figure 4D). Interestingly, detailed analysis of paw place-
279 ment behaviour revealed that this deficit was almost entirely explained by
280 impaired control of the hindlimbs. Paw slips were much more frequent when
281 stepping with a hindlimb than with a forelimb (Figure 4E,F). In addition,
282 when a slip did occur, these animals failed to adjust the affected paw to
283 compensate for the fall (e.g. keeping their digits closed), which significantly
284 impacted their overall posture recovery. These deficits in paw placement are

285 consistent with results from sectioning the entire pyramidal tract in cats [53],
286 and reports in ladder walking following motor cortical lesion in rodents [62],
287 but surprisingly we did not observe deficits in paw placement in animals with
288 ibotenic acid lesions limited to forelimb motor cortex (Figure 4D). Further-
289 more, despite this initial impairment, animals with extended lesions were still
290 able to improve their motor control strategy up to the point where they were
291 moving across the obstacles as efficiently as controls and other lesioned ani-
292 mals (Figure 4C, Video 2). Indeed, in the largest frontoparietal lesion, which
293 extended all the way to rostral visual cortex, recovery of a stable locomotion
294 pattern was evident over the course of just ten repeated trials (Video 3). The
295 ability of this animal to improve its motor control strategy in such a short
296 period of time seems to indicate the presence of motor learning, not simply
297 an increase in confidence with the new environment.

298 In subsequent training sessions we progressively increased the difficulty of
299 the obstacle course, by making more steps unstable. The goal was to compare
300 the performance of the two groups as a function of difficulty. Surprisingly,
301 both lesion and control animals were able to improve their performance by
302 the end of each training stage even for the most extreme condition where
303 all steps were unstable (Figure 3, Video 4). This seems to indicate that the
304 ability of these animals to fine-tune their motor performance in a challenging
305 environment remained intact.

306 One noticeable exception was the animal with the largest ibotenic acid
307 lesion. This animal, following exposure to the first unstable protocol, was

308 unable to bring itself to cross the obstacle course (Video 5). Some other con-
309 trol and lesioned animals also experienced a similar form of distress following
310 exposure to the unstable obstacles, but eventually all these animals managed
311 to start crossing over the course of a single session. In order to test whether
312 this was due to some kind of motor disability, we lowered the difficulty of
313 the protocol for this one animal until it was able to cross again. Following a
314 random permutation protocol, where any two single steps were released ran-
315 domly, this animal was then able to cross a single released obstacle placed
316 in any location of the assay. After this success, it eventually learned to cross
317 the highest difficulty level in the assay in about the same time as all the
318 other animals, suggesting that there was indeed no lasting motor execution
319 or learning deficit, and that the disability must have been due to some other
320 unknown, yet intriguing, (cognitive) factor.

321 Having established that the overall motor performance of these animals
322 was similar across all conditions, we next asked whether there was any differ-
323 ence in the strategy used by the two groups of animals to cross the unstable
324 obstacles. We noticed that during the first week of training, the posture of
325 the animals when stepping on the obstacles changed significantly over time
326 (Figure 5B,C). Specifically, the centre of gravity of the body was shifted fur-
327 ther forward and higher during later sessions, in a manner proportional to
328 performance. However, after the obstacles changed to the unstable state, we
329 observed an immediate and persistent adjustment of this crossing posture,
330 with animals assuming a lower centre of gravity and reducing their speed

331 as they approached the unstable obstacles (Figure 5C,D). Interestingly, we
332 also noticed that a group of animals adopted a different strategy. Instead of
333 lowering their centre of gravity, they either kept it unchanged or shifted it
334 even more forward and performed a jump over the unstable obstacles (Fig-
335 ure 6A,B). These two strategies were remarkably consistent across the two
336 groups, but there was no correlation between the strategy used and the de-
337 gree of motor cortical lesion (Figure 5E,F, 6C). In fact, we found that the use
338 of a jumping strategy was best predicted by the body weight of the animal
339 (Figure 6C).

340 During the two days where the stable state of the environment was rein-
341 stated, the posture of the animals was gradually restored to pre-manipulation
342 levels (Figure 5B,C), although in many cases this adjustment happened at a
343 slower rate than the transition from stable to unstable. Again, this postu-
344 ral adaptation was independent of the presence or absence of forepaw motor
345 cortex.

346 We next looked in detail at the days where the state of the obstacle
347 course was randomized on a trial-by-trial basis. This stage of the protocol
348 is particularly interesting as it reflects a situation where the environment
349 has a persistent degree of uncertainty. For this analysis, we were forced to
350 exclude the animals that employed a jumping strategy, as their experience
351 with the manipulated obstacles was the same irrespective of the state of the
352 world. First, we repeated the same posture analysis comparing all the stable
353 and unstable trials in the random protocol in order to control for whether

354 there was any subtle cue in our motorized setup that the animals might be
355 using to gain information about the current state of the world. There was no
356 significant difference between randomly presented stable and unstable trials
357 on the approach posture of the animal (Figure 7A). However, classifying
358 the trials on the basis of past trial history revealed a significant effect on
359 posture (Figure 7B). This suggested that the animals were adjusting their
360 body posture when stepping on the affected obstacles on the basis of their
361 current expectation about the state of the world, which is updated by the
362 previously experienced state. Surprisingly, this effect again did not depend
363 on the presence or absence of frontal motor cortical structures (Figure 7C,D).

364 Finally, we decided to test whether general motor performance was af-
365 fected by the randomized state of the obstacles. If the animals do not know
366 what state the world will be in, then there will be an increased challenge to
367 their stability when they cross over the unstable obstacles, possibly demand-
368 ing a quick change in strategy when they learn whether the world is stable
369 or unstable. In order to evaluate the dynamics of crossing, we compared the
370 speed profile of each animal across these different conditions (Figure 8, see
371 Methods). Interestingly, two of the animals with the largest lesions appeared
372 to be significantly slowed down on unstable trials, while controls and the ani-
373 mals with the smallest lesions instead tended to accelerate after encountering
374 an unstable obstacle. However, the overall effect for lesions versus controls
375 was not statistically significant (Figure 8C).

376 Nevertheless, we were intrigued by this observation and decided to in-

377 vestigate, in detail, the first moment in the assay when a perturbation is
378 encountered. In the random protocol, even though the state of the world is
379 unpredictable, the animals know that the obstacles might become unstable.
380 However, the very first time the environment becomes unstable, the collapse
381 of the obstacles is completely unexpected and demands an entirely novel
382 motor response.

383 A detailed analysis of the responses to the first collapse of the steps re-
384 vealed a striking difference in the strategies deployed by the lesion and control
385 animals. Upon the first encounter with the manipulated steps, we observed
386 three types of behavioural responses from the animals (Video 6): investi-
387 gation, in which the animals immediately stop their progression and orient
388 towards, whisk, and physically manipulate the altered obstacle; compensa-
389 tion, in which the animals rapidly adjust their behaviour to negotiate the
390 unexpected instability; and halting, in which the ongoing motor program
391 ceases and the animals' behaviour simply comes to a stop for several sec-
392 onds. Remarkably, these responses depended on the presence or absence of
393 motor cortex (Figure 9). Animals with the largest motor cortical lesions,
394 upon their first encounter with the novel environmental obstacle, halted for
395 several seconds, whereas animals with an intact motor cortex, and those with
396 the smallest lesions, were able to rapidly react with either an investigatory
397 or compensatory response (Video 7,8).

398 The response of animals with extended lesions was even more striking.
399 In two of these animals, there was a failure to recognize that a change had

400 occurred at all (Video 9). Instead, they kept walking across the now unstable
401 steps for several trials, never stopping to assess the new situation. One
402 of them gradually noticed the manipulation and stopped his progression,
403 while the other one only fully realized the change after inadvertently hitting
404 the steps with its snout (Video 9: Extended Lesion A). This was the first
405 time we ever observed this behaviour, as all animals with or without cortical
406 lesions always displayed a clear switch in behavioural state following the first
407 encounter with the manipulation. In the remaining animals with extended
408 lesions, two of them clearly halted their progression following the collapse
409 of the obstacles, in a way similar to the large motor cortex ibotenic lesions
410 (Video 10). The third animal (Extended Lesion B) actually collapsed upon
411 contact with the manipulated step, falling over its paw and digits awkwardly
412 and hitting the obstacles with its snout. Shortly after this there was a switch
413 to an exploratory behaviour state, in a way similar to Extended Lesion A.

414 **4 Experiment Discussion**

415 In this experiment, we assessed the role of motor cortical structures by mak-
416 ing targeted lesions to areas responsible for forelimb control [7, 63]. Con-
417 sistent with previous studies, we did not observe any conspicuous deficits
418 in movement execution for rats with bilateral motor cortex lesions when
419 negotiating a stable environment. Even when exposed to a sequence of un-
420 stable obstacles, animals were able to learn an efficient strategy for crossing
421 these more challenging environments, with or without motor cortex. These
422 movement strategies also include a preparatory component that might reflect
423 the state of the world an animal expected to encounter. Surprisingly, these
424 preparatory responses also did not require the presence of motor cortex.

425 It was only when the environment did not conform to expectation, and
426 demanded a rapid adjustment, that a difference between the lesion and con-
427 trol groups was obvious. Animals with extensive damage to the motor cortex
428 did not deploy a change in strategy. Rather, they halted their progression
429 for several seconds, unable to robustly respond to the new motor challenge.
430 In an ecological setting, such hesitation could easily prove fatal.

431 **5 Extended Discussion**

432 Is “robust control” a problem worthy of high level cortical input? Recovering
433 from a perturbation, to maintain balance or minimize the impact of a fall,
434 is a role normally assigned to our lower level postural control systems. The

435 corrective responses embedded in our spinal cord [64, 65], brainstem [66] and
436 midbrain [49] are clearly important components of this stabilizing network,
437 but are they sufficient to maintain robust movement in the dynamic environ-
438 ments that we encounter on a daily basis? Some insight into the requirements
439 for a robust control system can be gained from engineering attempts to build
440 robots that navigate in natural environments.

441 In the field of robotics, feats of precision and fine movement control (the
442 most commonly prescribed role for motor cortex), are not a major source of
443 difficulty. Industrial robots have long since exceeded human performance in
444 both accuracy and execution speed [67]. More recently, using reinforcement
445 learning methods, they are now able to automatically learn efficient move-
446 ment strategies, given a human-defined goal and many repeated trials for
447 fine-tuning [68]. What then are the hard problems in robotic motor control?
448 Why are most robots still confined to factories, i.e. controlled, predictable
449 environments? The reason is that as soon as a robot encounters natural
450 terrain, a vast number of previously unknown situations arise. The result-
451 ing “perturbations” are dealt with poorly by the statistical machine learning
452 models that are currently used to train robots in controlled settings.

453 Let’s consider a familiar example: You are up early on a Sunday morning
454 and head outside to collect the newspaper. It is cold out, so you put on a robe
455 and some slippers, open the front door, and descend the steps leading down to
456 the street in front of your house. Unbeknownst to you, a thin layer of ice has
457 formed overnight and your foot is now quickly sliding out from underneath

458 you. You are about to fall. What do you do? Well, this depends. Is there
459 a railing you can grab to catch yourself? Were you carrying a cup of coffee?
460 Did you notice the frost on the lawn and step cautiously, anticipating a
461 slippery surface? Avoiding a dangerous fall, or recovering gracefully, requires
462 a rich knowledge of the world, knowledge that is not immediately available
463 to spinal or even brainstem circuits. This rich context relevant for robust
464 movement is readily available in cortex, and cortex alone.

465 Imagine now that you are tasked with building a robot to collect your
466 morning newspaper. This robot, in order to avoid a catastrophic and costly
467 failure, would need to have all of this contextual knowledge as well. It would
468 need to know about the structure of the local environment (e.g. hand railings
469 that can support its weight), hot liquids and their viscosities, and even the
470 correlation of frozen dew with icy surfaces. To be a truly robust movement
471 machine, a robot must *understand* the physical structure of the world.

472 Reaching to stop a fall while holding a cup of coffee is not exactly the
473 kind of feat for which we praise our athletes and sports champions, and
474 this might explain why the difficulty of such “feats of robustness” are often
475 overlooked. However, it would not be the first time that we find ourselves
476 humbled by the daunting complexity of a problem that we naively assumed
477 was “trivial”. Vision, for example, has remained an impressively hard task for
478 a machine to solve at human-level performance, yet it was originally proposed
479 as an undergraduate summer project [69]. Perhaps a similar misestimate has
480 clouded our designation of the hard motor control problems worthy of cortical

481 input.

482 Inspired by the challenges confronting roboticists, as well as our rodent
483 behavioural results, we are now in a position to posit a new role for motor
484 cortex.

485 **A primordial role for motor cortex**

486 We are seeking a role for motor cortex in non-primate mammals, animals
487 that do not require this structure for overt movement production. The strug-
488 gles of roboticists highlight the difficulty of building movement systems that
489 robustly adapt to unexpected perturbations, and the results we report in
490 this study suggest that this is, indeed, the most conspicuous deficit for rats
491 lacking motor cortex. So let us propose that, in rodents, motor cortex is pri-
492 marily responsible for extending the robustness of the subcortical movement
493 systems. It is not required for control in stable, predictable, non-perturbing
494 environments, but instead specifically exerts its influence when unexpected
495 challenges arise. This, we propose, was the original selective pressure for
496 evolving a motor cortex, and thus, its primordial role. This role persists in
497 all mammals, mediated via a modulation of the subcortical motor system (as
498 is emphasized in studies of cat locomotion), and has evolved in primates to
499 include direct control of the skeletal musculature. Our proposal of a “robust”
500 teleology for motor cortex has a number of interesting implications.

501 **Implications for non-primate mammals**

502 One of the most impressive traits of mammals is the vast range of environ-
503 mental niches that they occupy. While most other animals adapt to change
504 over evolutionary time scales, mammals excel in their flexibility, quickly eval-
505 uating and responding to unexpected situations, and taking risks even when
506 faced with challenges that have never been previously encountered [70]. This
507 success requires more than precision, it requires resourcefulness: the abil-
508 ity to quickly come up with a motor solution for any situation and under
509 any condition [71]. The Russian neurophysiologist Bernstein referred to this
510 ability with an unconventional definition of “dexterity”, which he considered
511 to be distinct from a simple harmony and precision of movements. In his
512 words, dexterity is required only when there is “a conglomerate of unex-
513 pected, unique complications in the external situations, [such as] in a quick
514 succession of motor tasks that are all unlike each other” [71].

515 If Bernstein’s “robust dexterity” is the primary role for motor cortex,
516 then it becomes clear why the effects of lesions have thus far been so hard
517 to characterize: assays of motor behaviour typically evaluate situations that
518 are repeated over many trials in a stable environment. Such repeated tasks
519 were useful, as they offer improved statistical power for quantification and
520 comparison. However, we propose that these conditions specifically exclude
521 the scenarios for which motor cortex originally evolved. It is not easy to
522 repeatedly produce conditions that animals have not previously encountered,
523 and the challenges in analysing these unique situations are considerable.

524 The assay reported here represents our first attempt at such an experi-
525 ment, and it has already revealed that such conditions may indeed be nec-
526 essary to isolate the role of motor cortex in rodents. We thus propose that
527 neuroscience should pursue similar assays, emphasizing unexpected perturba-
528 tions and novel challenges, and we have developed new hardware and software
529 tools to make their design and implementation much easier [72].

530 **Implications for primate studies**

531 In contrast to other mammals, primates require motor cortex for the direct
532 control of movement. However, do they also retain its role in generating
533 robust responses? The general paresis, or even paralysis, that results from
534 motor cortical lesions in these species obscures the involvement of cortex in
535 directing rapid responses to perturbations. Yet there is evidence that a role
536 in robust control is still present in primates, including humans. For example,
537 stroke patients with partial lesions to the distributed motor cortical system
538 will often recover the ability to move the affected musculature. However,
539 even after recovering movement, stroke patients are still prone to severe im-
540 pairments in robust control: unsupported falls are one of the leading causes
541 of injury and death in patients surviving motor cortical stroke [73]. We thus
542 suggest that stroke therapy, currently focused on regaining direct movement
543 control, should also consider strategies for improving robust responses.

544 Even if we acknowledge that a primordial role of motor cortex is still
545 apparent in primate movement control, it remains to be explained why the

546 motor cortex of these species acquired direct control of basic movements in
547 the first place. This is an open question.

548 **Some speculation on the role of direct cortical control**

549 What happens when cortex acquires direct control of movement? First, it
550 must learn how to use this influence, bypassing or modifying lower move-
551 ment controllers. While functional corticospinal tract connections may be
552 established prenatally [74], the refinement of corticospinal dependent move-
553 ments, which must override the lower motor system, takes much longer and
554 coincides with the lengthy maturation period of corticospinal termination
555 patterns [75]. Humans require years of practice to produce and refine ba-
556 sic locomotion and grasping [76, 77], motor behaviours that are available to
557 other mammals almost immediately after birth. This may be the cost of
558 giving cortex direct control of movement—it takes more time to figure out
559 how to move the body—but what is the benefit?

560 Giving motor cortex direct control over the detailed dynamics of move-
561 ment might simply have extended the range and flexibility of robust re-
562 sponses. This increased robustness may have been required for primates
563 to negotiate more difficult unpredictable environments, such as the forest
564 canopy. Direct cortical control of the musculature may have evolved be-
565 cause it allowed primates to avoid their less “dexterous” predators simply by
566 ascending, and robustly negotiating, the precarious branches of tree tops.
567 However, the consequences of this cortical “take-over” might be even more

568 profound.

569 With motor cortex in more direct control of overt movements, the be-
570 haviour of a primate is a more direct reflection of cortical state: when you
571 watch a primate move you are directly observing cortical commands. For
572 species that live in social groups, this would allow a uniquely efficient means
573 of communicating the state of cortex between conspecifics, a rather signif-
574 icant advantage for group coordination and a likely prerequisite for human
575 language. This novel role for motor cortex—communication—might have ex-
576 erted the evolutionary pressure to give cortex more and more control over
577 basic movements, ultimately obscuring its primordial, and fundamental, role
578 in robust control.

579 **Some preliminary conclusions**

580 Clearly our results are insufficient to draw any final conclusion, but that is
581 not our main goal. We present these experiments to support and motivate
582 our attempt to distil a long history of research, and ultimately suggest a
583 new approach to investigating the role of motor cortex. This approach most
584 directly applies to studies of non-primate mammals. There is now a host of
585 techniques to monitor and manipulate cortical activity during behaviour in
586 these species, but we propose that we should be monitoring and manipulating
587 activity during behaviours that actually require motor cortex.

588 This synthesis also has implications for engineers and clinicians. We sug-
589 gest that acknowledging a primary role for motor cortex in robust control,

590 a problem still daunting to robotics engineers, can guide the development of
591 new approaches for building intelligent machines, as well as new strategies
592 to assess and treat patients with motor cortical damage. We concede that
593 our results are still naïve, but propose that the implications are worthy of
594 further consideration.

595 6 Methods

596 All experiments were approved by the Champalimaud Foundation Bioethics
597 Committee and the Portuguese National Authority for Animal Health, Di-
598 recção-Geral de Alimentação e Veterinária (DGAV).

599 **Lesions:** Ibotenic acid was injected bilaterally in 11 Long-Evans rats
600 (ages from 83 to 141 days; 9 females, 2 males), at 3 injection sites with
601 2 depths per site (-1.5 mm and -0.75 mm from the surface of the brain).
602 At each depth we injected a total amount of 82.8 nL using a microinjector
603 (Drummond Nanoject II, 9.2 nL per injection, 9 injections per depth). The
604 coordinates for each site, in mm with respect to Bregma, were: $+1.0$ AP / 2.0
605 ML; $+1.0$ AP / 4.0 ML; $+3.0$ AP / 2.0 ML, following the protocol reported by
606 Kawai et al. for targeting forelimb motor cortex [7]. Five other animals were
607 used as sham controls (age-matched controls; 3 females, 2 males), subject to
608 the same intervention, but where ibotenic acid was replaced with physiologi-
609 cal saline. Six additional animals were used as wildtype, no-surgery, controls
610 (age-matched controls; 6 females).

611 For the frontal cortex aspiration lesions, the margins of the craniotomy
612 were extended to cover from -2.0 to $+5.0$ mm AP relative to Bregma and
613 laterally from 0.5 mm up to the temporal ridge of the skull. After removal
614 of the skull, the exposed dura was cut and removed, and the underlying
615 tissue aspirated to a depth of 2 to 3 mm with a fine pipette [78]. For the
616 frontoparietal cortical lesions, the craniotomy extended from -6.0 to $+4.0$

617 mm AP relative to Bregma and laterally from 0.5 mm up to the temporal
618 ridge. Two of these animals underwent aspiration lesions as described above.
619 In the remaining animal, the lesion was induced by pial stripping in order to
620 further restrict the damage to cortical areas. After removal of the dura, the
621 underlying pia, arachnoid and vasculature were wiped with a sterile cotton
622 swab until no vasculature was visible [79].

623 **Recovery period:** After the surgeries, animals were given a minimum
624 of one week (up to two weeks) recovery period in isolation. After this period,
625 animals were handled every day for a week, after which they were paired
626 again with their age-matched control to allow for social interaction during
627 the remainder of the recovery period. In total, all animals were allowed
628 at least one full month of recovery before they were first exposed to the
629 behaviour assay.

630 The three largest frontoparietal lesioned animals were originally prepared
631 for a study of behaviour in a dynamic visual foraging task, which they were
632 exposed to for one month in addition to the recovery period described above.
633 This task did not, however, require any challenging motor behaviours be-
634 sides locomotion over a completely flat surface. This period was also used
635 to monitor the overall health condition of the animals and to facilitate sen-
636 sorimotor recovery as much as possible. The animal with the largest lesion
637 (Extended Lesion F) was prevented from completing the behaviour protocol
638 due to deteriorating health conditions following the first two days of testing.

639 **Histology:** All animals were perfused intracardially with 4% paraformal-

640 dehyde in phosphate buffer saline (PBS) and brains were post-fixed for at
641 least 24 h in the same fixative. Serial coronal sections (100 μm) were Nissl-
642 stained and imaged for identification of lesion boundaries. In two of the
643 largest frontoparietal lesions (Extended Lesions D and E), serial sections
644 were taken sagittally.

645 In order to reconstruct lesion volumes, the images of coronal sections were
646 aligned and the outlines of both brain and lesions were manually traced in
647 Fiji [80] and stored as two-dimensional regions of interest. Lesion volumes
648 were calculated by summing the area of each region of interest multiplied by
649 the thickness of each slice. The stored regions were also used to reconstruct
650 a 3D polygon mesh for visualization of lesion boundaries.

651 **Behaviour assay:** During each session the animal was placed inside a
652 behaviour box for 30 min, where it could collect water rewards by shuttling
653 back and forth between two nose pokes (Island Motion Corporation, USA).
654 To do this, animals had to cross a 48 cm obstacle course composed of eight
655 2 cm aluminium steps spaced by 4 cm (Figure 1A). The structure of the assay
656 and each step in the obstacle course was built out of aluminium structural
657 framing (Bosch Rexroth, DE, 20 mm series). The walls of the arena were fab-
658 ricated with a laser-cutter from 5 mm thick opaque black acrylic and fixed
659 to the structural framing. A transparent acrylic window partition was po-
660 sitioned in front of the obstacle course in order to provide a clear view of
661 the animal. All experiments were run in the dark by having the behavioural
662 apparatus enclosed in a light tight box.

663 A motorized brake allowed us to lock or release each step in the obstacle
664 course (Figure 1B). The shaft of each of the obstacles was coupled to an
665 acrylic piece used to control the rotational stability of each step. In order
666 to lock a step in a fixed position, two servo motors are actuated to press
667 against the acrylic piece and hold it in place. Two other acrylic pieces were
668 used as stops to ensure a maximum rotation angle of approximately +/-
669 100°. Two small nuts were attached to the bottom of each step to work as a
670 counterweight that gives the obstacles a tendency to return to their original
671 flat configuration. In order to ensure that noise from servo motor actuation
672 could not be used as a cue to tell the animal about the state of each step, the
673 motors were always set to press against an acrylic piece, either the piece that
674 keeps the step stabilized, or the acrylic stops. At the beginning of each trial,
675 the motors were run through a randomized sequence of positions in order to
676 mask information about state transitions and also to ensure the steps were
677 reset to their original configuration. Control of the motors was done using a
678 Motoruino board (Artica, PT) along with a custom workflow written in the
679 Bonsai visual programming language [72].

680 **Data acquisition:** The behaviour of the animals was recorded with a
681 high-speed and high-resolution videography system (1280x680 @ 120 Hz) us-
682 ing an infrared camera (Flea3, PointGrey, CA), super-bright infrared LED
683 front lights (SMD5050, 850 nm) and a vari-focal lens (Fujinon, JP) positioned
684 in front of the transparent window partition. A top view of the assay was
685 simultaneously recorded with the same system at a lower frame-rate (30 Hz)

686 for monitoring purposes. All video data was encoded with MPEG-4 com-
687 pression for subsequent offline analysis. Behaviour data acquisition for the
688 nose poke beam breaks was done using an Arduino board (Uno, Arduino,
689 USA) and streamed to the computer via USB. All video and sensor data
690 acquisition was recorded in parallel using the same Bonsai workflow used to
691 control the behaviour assay.

692 **Behaviour protocol:** The animals were kept in a state of water depri-
693 vation for 20 h prior to each daily session. For every trial, rats were delivered
694 a 20 μ L drop of water. At the end of each day, they were given free access
695 to water for 10 min before initiating the next deprivation period. Sessions
696 lasted for six days of the week from Monday to Saturday, with a day of free
697 access to water on Sunday. Before the start of the water deprivation proto-
698 col, animals were run on a single habituation session where they were placed
699 in the box for a period of 15 min.

700 The following sequence of conditions were presented to the animals over
701 the course of a month (see also Figure 1A): day 0, habituation to the box;
702 day 1-4, all the steps were fixed in a stable configuration; day 5, 20 trials of
703 the stable configuration, after which the two center steps were made unstable
704 (i.e. free to rotate); day 6-10, the center two steps remained unstable; day
705 11, 20 trials of the unstable configuration, after which the two center steps
706 were again fixed in a stable state; day 12, all the steps were fixed in a stable
707 configuration; day 13-16, the state of the center two steps was randomized
708 on a trial-by-trial basis to be either stable or unstable. Following the end

709 of the random protocol, animals continued to be tested in the assay for a
710 variable number of days (up to one week) in different conditions. At the
711 end of the testing period, all animals were exposed to a final session where
712 all steps were made free to rotate in order to assay locomotion performance
713 under challenging conditions.

714 **Data analysis:** All scripts and custom code used for data analysis are
715 available online¹. The raw video data was first pre-processed using a custom
716 Bonsai workflow in order to extract features of interest. Tracking of the nose
717 was achieved by background subtraction and connected component labelling
718 of segmented image elements. First we compute the ellipse best-fit to the
719 largest object in the image. We then mark the tip of the nose as the fur-
720 thermost point, in the segmented shape of the animal, along the major axis
721 of the ellipse. In order to analyse stepping performance, regions of interest
722 were defined around the surface of each step and in the gaps between the
723 steps. Background subtracted activity over these regions was recorded for
724 every frame for subsequent detection and classification of steps and slips.

725 Analysis routines were run using the NumPy scientific computing package
726 [81] and the Pandas data analysis library [82] for the Python programming
727 language. Crossings were automatically extracted from the nose trajectory
728 data by first detecting consecutive time points where the nose was positively
729 identified in the video. In order for these periods to be successfully marked
730 as crossings, the starting position of the nose must be located on the opposite

¹<https://bitbucket.org/kampff-lab/shuttling-analysis>

731 side of the ending position. Inside each crossing, the moment of stepping with
732 the forelimb on the centre steps was extracted by looking at the first peak
733 above a threshold in the first derivative of the activation signal in the corre-
734 sponding region of interest. False positive classifications due to hindlimb or
735 tail activations were eliminated by enforcing the constraint that the position
736 of the head must be located before the next step. Visual confirmation of the
737 classified timepoints showed that spurious activations were all but eliminated
738 by this procedure as stepping with the hindlimb or tail requires the head to
739 be further ahead in space unless the animal turned around (in which case the
740 trajectory would not be marked as a crossing anyway). The position of the
741 nose at the moment of each step was extracted and found to be normally dis-
742 tributed, so statistical analysis of the step posture in the random condition
743 used an unpaired t-test to check for independence of different measurement
744 groups.

745 In order to evaluate the dynamics of crossing in the random condition,
746 we first measured for every trial the speed at which the animals were moving
747 on each spatial segment of the assay. To minimize overall trial-by-trial vari-
748 ation in individual animal performance, we used the average speed at which
749 the animal approached the manipulated step as a baseline and subtracted
750 it from the speed at each individual segment. To summarize differences in
751 performance between stable and unstable trials, we then computed the aver-
752 age speed profile for each condition, and then subtracted the average speed
753 profile for unstable trials from the average speed profile for stable trials. Fi-

754 nally, we computed the sum of all these speed differences at every segment in
755 order to obtain the speedup index for each animal, i.e. an index of whether
756 the animal tends to accelerate or decelerate across the assay on stable versus
757 unstable trials.

758 **Video classification:** Classification of paw placement faults (i.e. slips)
759 was performed in semi-automated fashion. First, possible slip timepoints
760 were detected automatically using the peak detection method outlined above.
761 All constraints on head position were relaxed for this analysis in order to
762 exclude the possibility of false negatives. A human classifier then proceeded
763 to manually go through each of the slip candidates and inspect the video
764 around that timepoint in order to assess whether the activation peak was a
765 genuine paw placement fault. Examples of false positives include tail and
766 head activations as well as paw activations that occur while the animal is
767 actively engaged in exploration, rearing, or other activities that are unrelated
768 to crossing the obstacles.

769 Classification of behaviour responses following first exposure to the unsta-
770 ble condition was done on a frame-by-frame analysis of the high-speed video
771 aligned on first contact with the manipulated step. The frame of first con-
772 tact was defined as the first frame in which there is noticeable movement of
773 the step caused by animal contact. Three main categories of behaviour were
774 observed to follow the first contact: compensation, investigation and halt-
775 ing. Behaviour sequences were first classified as belonging to one of these
776 categories and their onsets and offsets determined by the following criteria.

777 Compensation behaviour is defined by a rapid and adaptive postural correc-
778 tion to the locomotion pattern in response to the perturbation. Onset of
779 this behaviour is defined by the first frame in which there is visible rapid
780 contraction of the body musculature following first contact. Investigation
781 behaviour consists of periods of targeted interaction with the steps, often
782 involving manipulation of the freely moving obstacle with the forepaws. The
783 onset of this behaviour is defined by the animal orienting its head down to
784 one of the manipulated steps, followed by subsequent interaction. Halting
785 behaviour is characterized by a period in which the animal stops its ongoing
786 motor program, and maintains the same body posture for several seconds,
787 without switching to a new behaviour or orienting specifically to the manipu-
788 lated steps. This behaviour is distinct from a freezing response, as occasional
789 movements of the head are seen. Onset of this behaviour is defined by the mo-
790 ment where locomotion and other motor activities besides movement of the
791 head come to a stop. A human classifier blind to the lesion condition was
792 given descriptions of each of these three main categories of behaviour and
793 asked to note onsets and offsets of each behaviour throughout the videos.
794 These classifications provide a visual summary of the first response videos;
795 the complete dataset used for this classification is included as supplementary
796 movies.

797 **7 Acknowledgements**

798 We thank Lorenza Calcaterra for the extended frontoparietal cortical le-
799 sion preparations; João Frazão for invaluable help annotating the behaviour
800 videos and all the members of the Intelligent Systems Lab for constant feed-
801 back on the ideas, experiments and manuscript. G.L. is supported by the
802 PhD Studentship SFRH/BD/51714/2011 from the Foundation for Science
803 and Technology. The Champalimaud Neuroscience Programme is supported
804 by the Champalimaud Foundation.

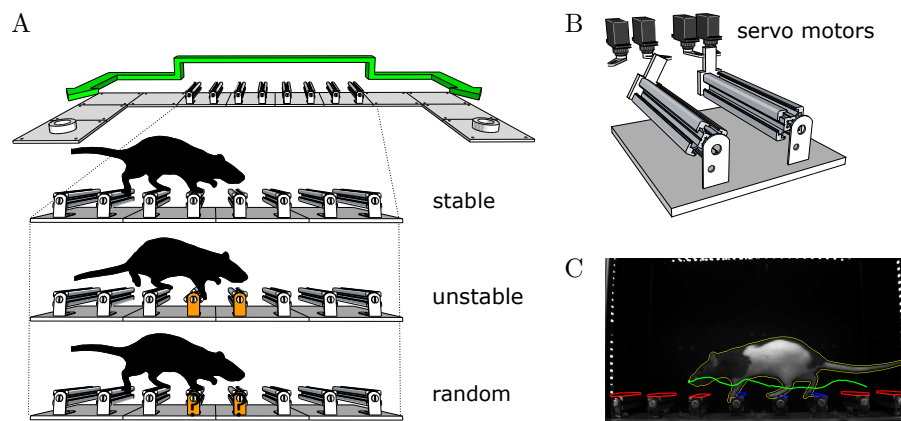


Figure 1: An obstacle course for rodents. (A) Schematic of the apparatus and summary of the different conditions in the behaviour protocol. Animals shuttle back and forth between two reward ports at either end of the enclosure. (B) Schematic of the locking mechanism that allows each individual step to be made stable or unstable on a trial-by-trial basis. (C) Example video frame from the behaviour tracking system. Coloured overlays represent regions of interest and feature traces extracted automatically from the video.

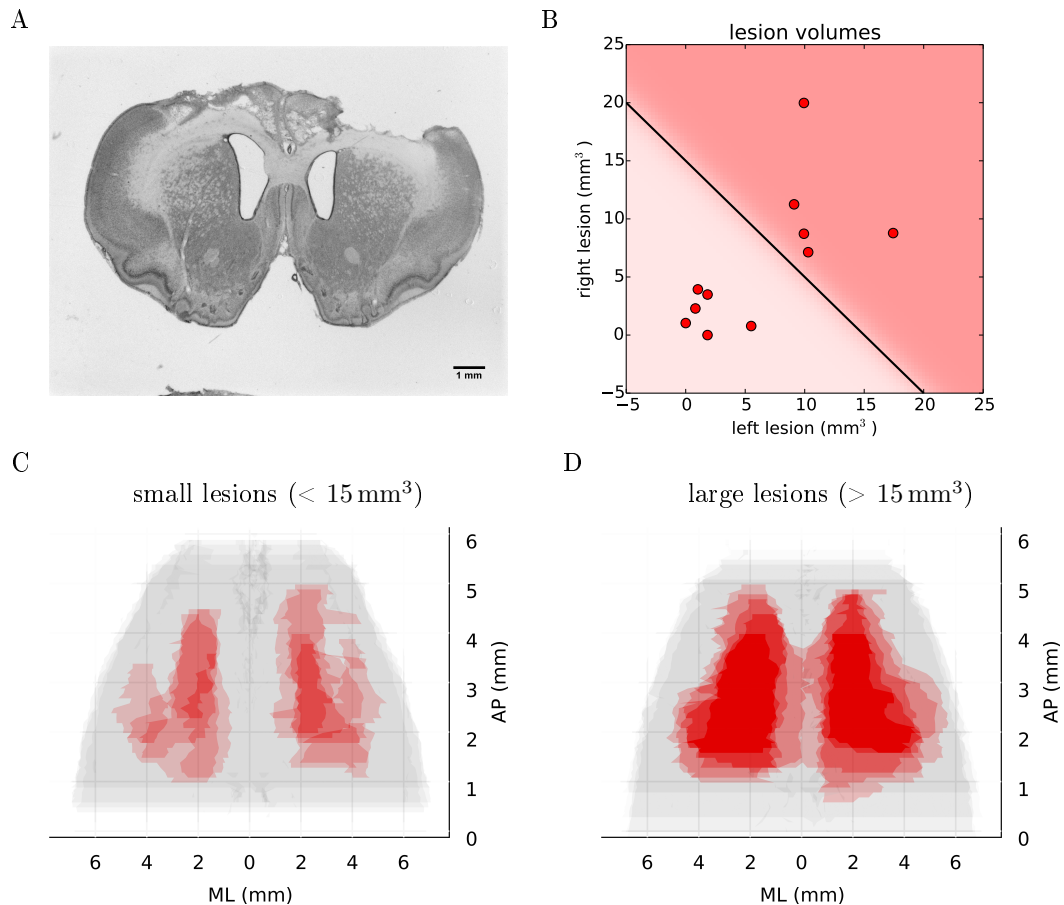


Figure 2: Histological analysis of lesion size. (A) Representative example of Nissl-stained coronal section showing bilateral ibotenic acid lesion of primary and secondary forelimb motor cortex. (B) Distribution of lesion volumes in the left and right hemispheres for individual animals. A lesion was considered “large” if the total lesion volume was above 15 mm³. (C) Super-imposed reconstruction stacks for all the small lesions ($n = 6$). (D) Super-imposed reconstruction stacks for all the large lesions ($n = 5$).

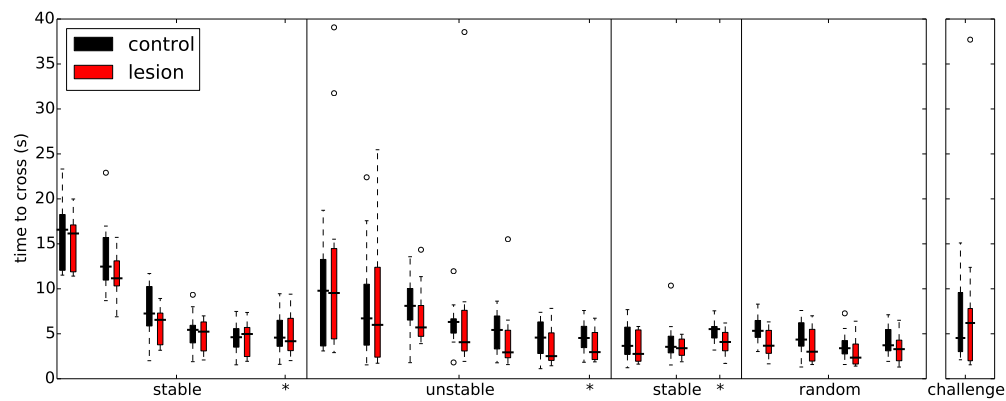


Figure 3: Overall performance on the obstacle course is similar for both lesion ($n = 11$) and control animals ($n = 11$) across the different protocol stages. Each set of coloured bars represents the distribution of average time to cross the obstacles on a single session. Asterisks indicate sessions where there was a change in assay conditions during the session (see text). In these transition sessions, the average performance on the 20 trials immediately preceding the change is shown to the left of the solid vertical line whereas the performance on the remainder of that session (after the change) is shown to the right.

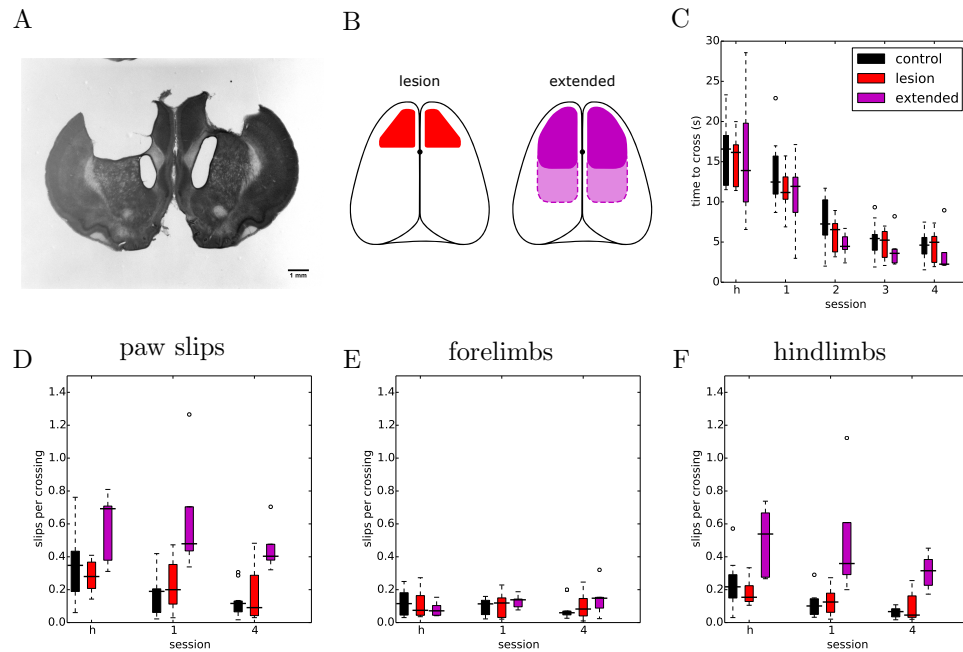


Figure 4: Extended frontoparietal cortex lesions perform as well as control animals despite impaired hindlimb control. (A) Representative example of Nissl-stained coronal section showing bilateral aspiration lesion of forelimb sensorimotor cortex. (B) Schematic depicting targeted lesion areas in the different animal groups. Left: outline of bilateral ibotenic acid lesions to the motor cortex. Right: outline of extended bilateral frontoparietal cortex lesions. Solid outline represents frontal cortex targeted lesions and dotted outline the more extensive frontoparietal lesions. (C) Average time required to cross the obstacles in the stable condition for extended lesions ($n = 5$). Performance of the other groups is shown for comparison. (D) Average number of slips per crossing in early versus late sessions of the stable condition. (E) Same data showing only forelimb slips. (F) Same data showing only hindlimb slips.

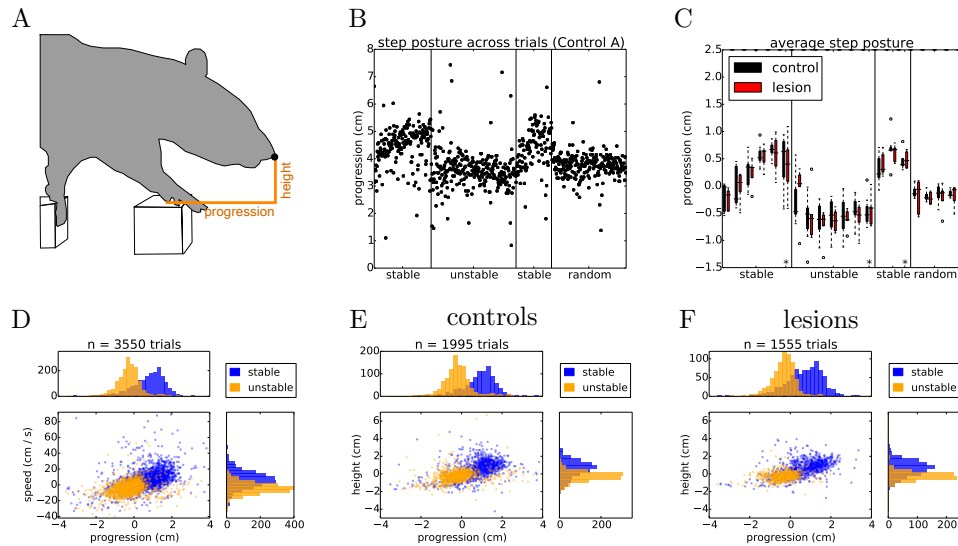


Figure 5: Rats adapt their postural approach to the obstacles after a change in physics. (A) Schematic of postural analysis image processing. The position of the animal's nose is extracted whenever the paw activates the ROI of the first manipulated step (see methods). (B) The horizontal position, i.e. progression, of the nose in single trials for one of the control animals stepping across the different conditions of the shuttling protocol. (C) Average horizontal position of the nose across the different protocol stages for both lesion and control animals. Asterisks indicate the average nose position on the 20 trials immediately preceding a change in protocol conditions (see text). (D) Distribution of horizontal position against speed for the last two days of the stable (blue) and unstable (orange) protocol stages. (E-F) Distribution of nose positions for control and lesion animals over the same sessions.

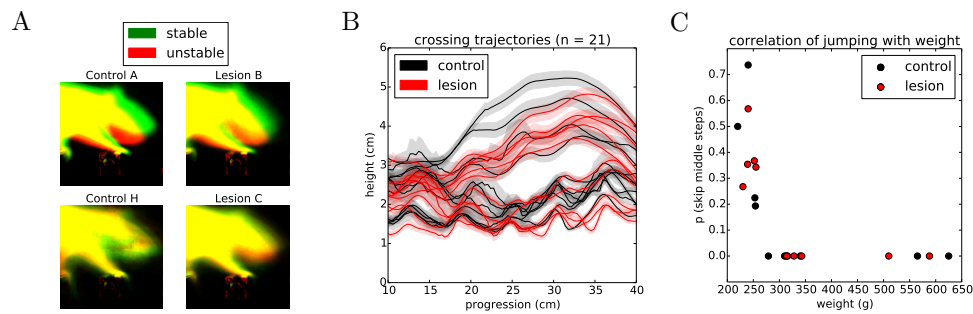


Figure 6: Animals use different strategies for dealing with the unstable obstacles. (A) Example average projection of all posture images for stable (green) and unstable (red) sessions for two non-jumper (top) and two jumper (bottom) animals. (B) Average nose trajectories for individual animals crossing the unstable condition. The shaded area around each line represents the 95% confidence interval. (C) Correlation of the probability of skipping the center two steps with the weight of the animal.

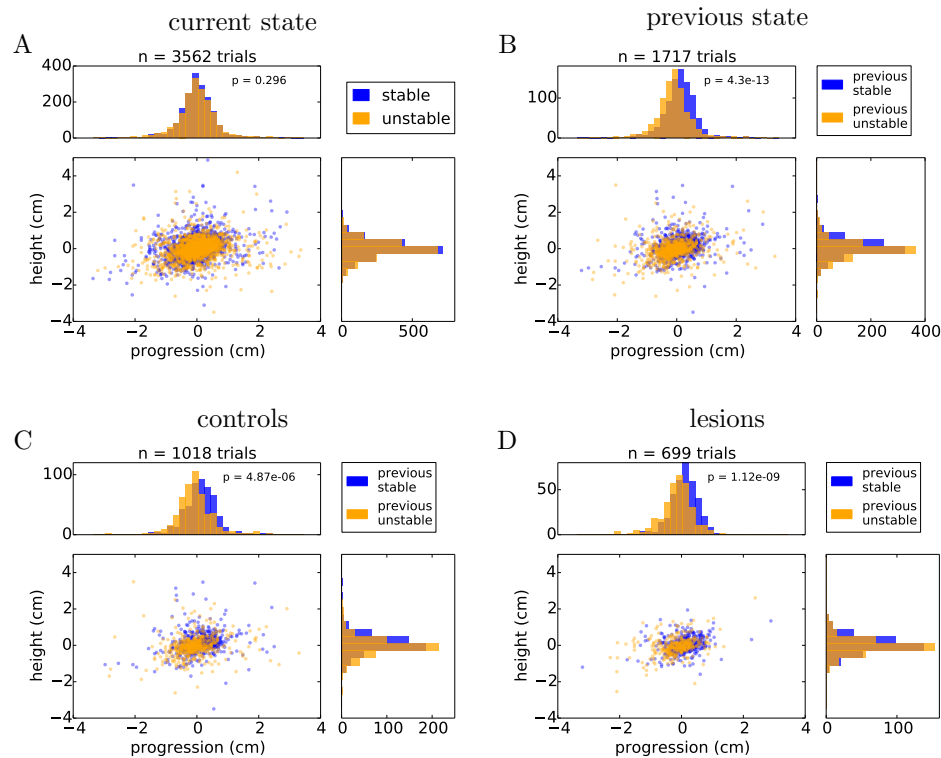


Figure 7: Animals adjust their posture on a trial-by-trial basis to the expected state of the world. **(A)** Distribution of nose positions on the randomized protocol when stepping on the first manipulated obstacle, for trials in which the current state was stable (blue) or unstable (orange). **(B)** Distribution of nose positions for trials in which the previous two trials were stable (blue) or unstable (orange). **(C-D)** Same data as in **(B)** split by the control and lesion groups. p values from Student's unpaired t -test are indicated.

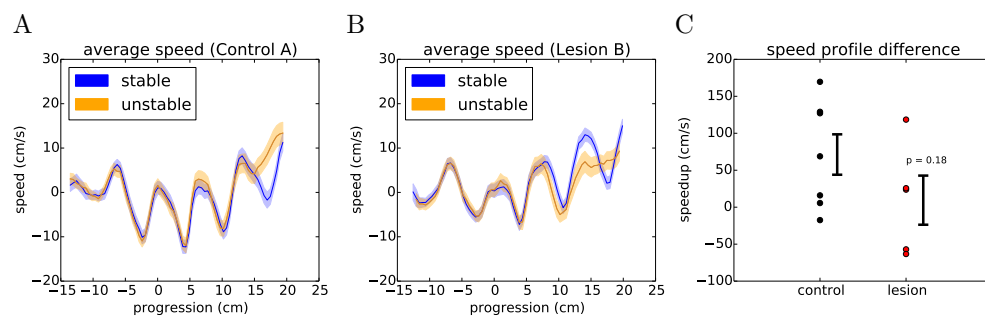


Figure 8: Encountering different states of the randomized obstacles causes the animals to quickly adjust their movement trajectory. (A) Example average speed profile across the obstacles for stable (blue) and unstable (orange) trials in the randomized sessions of a control animal (see text). The shaded area around each line represents the 95% confidence interval. (B) Respective for one of the largest lesions. (C) Summary of the average difference between the speed profiles for stable and unstable trials across the two groups of animals. Error bars show standard error of the mean. p value from Student's unpaired t-test is indicated.

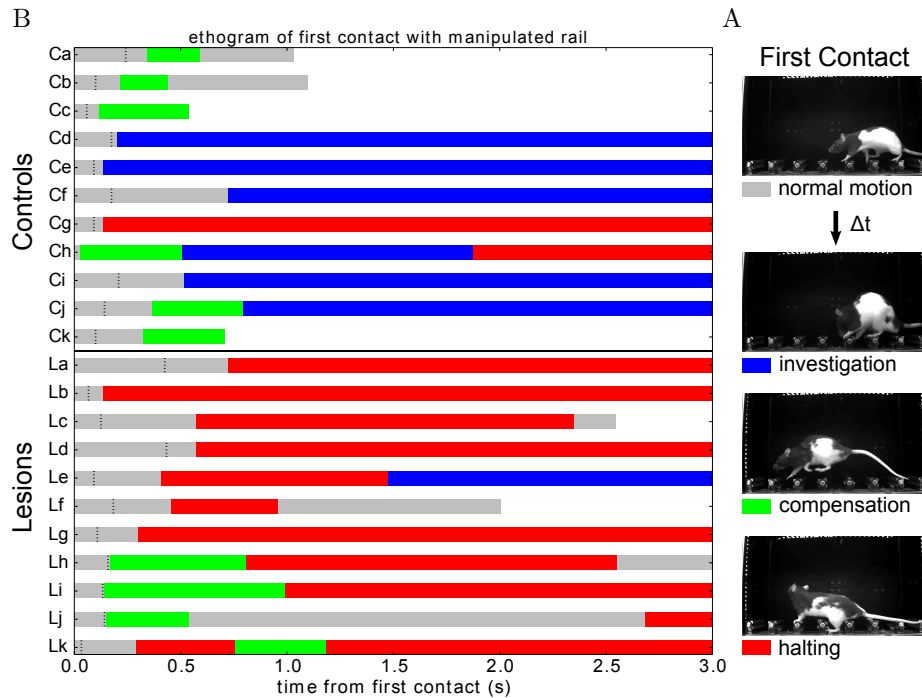


Figure 9: Responses to an unexpected change in the environment. (A) Response types observed across individuals upon first encountering an unpredicted instability in the state of the centre obstacles. (B) Ethogram of behavioural responses classified according to the three criteria described in (A) and aligned (0.0) on first contact with the newly manipulated obstacle. Black dashes indicate when the animal exhibits a pronounced ear flick. White indicates that the animal has crossed the obstacle course.

References

- [1] K. L. Tyler and R. Malessa. “The Goltz-Ferrier debates and the triumph of cerebral localizationalist theory.” In: *Neurology* 55 (2000), pp. 1015–1024. DOI: 10.1212/WNL.56.10.1424.
- [2] C. G. Gross. “The discovery of motor cortex and its background.” In: *Journal of the history of the neurosciences* 16.3 (2007), pp. 320–31. DOI: 10.1080/09647040600630160.
- [3] K. S. Lashley. “Studies of cerebral function in learning: V. The retention of motor habits after destruction of the so-called motor areas in primates”. In: *Archives of Neurology and Psychiatry* 12.3 (1924), pp. 249–276. DOI: 10.1001/archneurpsyc.1924.02200030002001.
- [4] J. G. D. de Barenne. “"Corticalization" of function and functional localization in the cerebral cortex”. In: *Archives of Neurology And Psychiatry* 30.4 (1933), pp. 884–901. DOI: 10.1001/archneurpsyc.1933.02240160196012.
- [5] G. Rizzolatti and L. Craighero. “The Mirror-Neuron System”. In: *Annual Review of Neuroscience* 27.1 (2004), pp. 169–192. DOI: 10.1146/annurev.neuro.27.070203.144230.
- [6] C. A. Porro et al. “Primary motor and sensory cortex activation during motor performance and motor imagery: a functional magnetic resonance imaging study.” In: *The Journal of neuroscience : the official journal of the Society for Neuroscience* 16.23 (1996), pp. 7688–7698.

- [7] R. Kawai et al. “Motor Cortex Is Required for Learning but Not for Executing a Motor Skill”. In: *Neuron* (2015), pp. 1–13. DOI: 10.1016/j.neuron.2015.03.024.
- [8] G. Fritsch and E. Hitzig. “Über die elektrische Erregbarkeit des Grosshirns”. In: *Archiv für Anatomie, Physiologie und Wissenschaftliche* 37 (1870), pp. 300–332.
- [9] A. S. F. Leyton and C. S. Sherrington. “Observations on the excitable cortex of the chimpanzee, orang-utan, and gorilla”. In: *Experimental Physiology* 11.2 (1917), pp. 135–222. DOI: 10.1113/expphysiol.1917.sp000240.
- [10] W. Penfield and E. Boldrey. “Somatic motor and sensory representation in the cerebral cortex of man as studied by electrical stimulation.” In: *Brain* 60.4 (1937), pp. 389–443.
- [11] E. Neafsey et al. “The organization of the rat motor cortex: a microstimulation mapping study”. In: *Brain Research Reviews* 11.1 (1986), pp. 77–96.
- [12] E. V. Evarts. “Relation of pyramidal tract activity to force exerted during voluntary movement.” In: *Journal of neurophysiology* 31.1 (1968), pp. 14–27.
- [13] A. Georgopoulos, A. Schwartz, and R. Kettner. “Neuronal population coding of movement direction”. In: *Science* 233.4771 (1986), pp. 1416–1419. DOI: 10.1126/science.3749885.

- [14] A. B. Schwartz. “Motor cortical activity during drawing movements: population representation during sinusoid tracing.” In: *Journal of neurophysiology* 70.1 (1993), pp. 28–36.
- [15] S. H. Scott et al. “Dissociation between hand motion and population vectors from neural activity in motor cortex”. In: *Nature* 413.6852 (2001), pp. 161–165.
- [16] M. M. Churchland and K. V. Shenoy. “Temporal complexity and heterogeneity of single-neuron activity in premotor and motor cortex”. In: *J Neurophysiol* 97.6 (2007), pp. 4235–4257. DOI: 10.1152/jn.00095.2007.
- [17] M. M. Churchland et al. “Neural population dynamics during reaching.” In: *Nature* 487.7405 (2012), pp. 51–6. DOI: 10.1038/nature11129.
- [18] E. Todorov. “Direct cortical control of muscle activation in voluntary arm movements: a model.” In: *Nature neuroscience* 3.4 (2000), pp. 391–8. DOI: 10.1038/73964.
- [19] M. S. A. Graziano, C. S. R. Taylor, and T. Moore. “Complex Movements Evoked by Microstimulation of Precentral Cortex”. In: *Neuron* 34 (2002), pp. 841–851.
- [20] T. N. Aflalo and M. S. A. Graziano. “Possible origins of the complex topographic organization of motor cortex: reduction of a multidimensional space onto a two-dimensional array.” In: *The Journal of neu-*

- rosience : the official journal of the Society for Neuroscience* 26.23 (2006), pp. 6288–97. DOI: 10.1523/JNEUROSCI.0768-06.2006.
- [21] D. Laplane et al. “Motor consequences of motor area ablations in man”. In: *Journal of the Neurological Sciences* 31.1 (1977), pp. 29–49. DOI: 10.1016/0022-510X(77)90004-1.
- [22] W. G. Darling, M. A. Pizzimenti, and R. J. Morecraft. “Functional recovery following motor cortex lesions in non-human primates: experimental implications for human stroke patients”. In: *Journal of Integrative Neuroscience* 10.3 (2011), pp. 353–384. DOI: 10.1142/S0219635211002737.
- [23] B. Zaaïmi et al. “Changes in descending motor pathway connectivity after corticospinal tract lesion in macaque monkey”. In: *Brain* 135.7 (2012), pp. 2277–2289. DOI: 10.1093/brain/aws115.
- [24] I. Q. Whishaw et al. “The impairments in reaching and the movements of compensation in rats with motor cortex lesions: an endpoint, videorecording, and movement notation analysis”. In: *Behavioural brain research* 42 (1991), pp. 77–91.
- [25] M. Alaverdashvili and I. Q. Whishaw. “Motor cortex stroke impairs individual digit movement in skilled reaching by the rat.” In: *The European journal of neuroscience* 28.2 (2008), pp. 311–22. DOI: 10.1111/j.1460-9568.2008.06315.x.

- [26] F. Goltz. “Über die Verrichtungen des Grosshirns”. In: *Pflügers Archiv gesamte Physiologie des Menschen und der Tiere* 42 (1888), pp. 419–467.
- [27] L. M. Bjursten, K. Norrsell, and U. Norrsell. “Behavioural repertory of cats without cerebral cortex from infancy”. In: *Experimental brain research* 130.2857 (1976), pp. 115–130.
- [28] P. Terry, B. A. Herbert, and D. A. Oakley. “Anomalous patterns of response learning and transfer in decorticate rats.” In: *Behavioural brain research* 33.1 (1989), pp. 105–9.
- [29] S. Tower. “Pyramidal lesion in the monkey”. In: *Brain* 63.1 (1940), pp. 36–90.
- [30] D. G. Lawrence and H. G.J. M. Kuypers. “The functional organization of the motor system in the monkey. I. The effects of bilateral pyramidal lesions”. In: *Brain* 91.1 (1968), pp. 1–14.
- [31] C. N. Woolsey et al. “Complete unilateral section of the pyramidal tract at the medullar level in *Macaca mulatta*”. In: *Brain Research* 40.1 (1972), pp. 119–123.
- [32] C. G. Bernhard and E. Bohm. “Cortical representation and functional significance of the corticomotoneuronal system”. In: *Archives of Neurology And Psychiatry* 72.4 (1954), pp. 473–502. DOI: 10.1001/archneurpsyc.1954.02330040075006.

- [33] H. G.J. M. Kuypers. “Anatomy of the descending pathways”. In: *Comprehensive Physiology* (1981), pp. 597–666.
- [34] H.-W. Yang and R. N. Lemon. “An electron microscopic examination of the corticospinal projection to the cervical spinal cord in the rat: lack of evidence for cortico-motoneuronal synapses.” In: *Experimental brain research* 149.4 (2003), pp. 458–69. DOI: 10.1007/s00221-003-1393-9.
- [35] P. W. Nathan and M. C. Smith. “The rubrospinal and central tegmental tracts in man”. In: *Brain* 105.2 (1982), pp. 223–269. DOI: 10.1093/brain/105.2.223.
- [36] D. G. Lawrence and H. G.J. M. Kuypers. “The functional organization of the motor system in the monkey. II. The effects of lesions of the descending brain-stem pathways”. In: *Brain* 91.1 (1968), pp. 15–36.
- [37] J. Donoghue and S. P. Wise. “The motor cortex of the rat: cytoarchitecture and microstimulation mapping.” In: *The Journal of comparative neurology* 212.1 (1982), pp. 76–88. DOI: 10.1002/cne.902120106.
- [38] B. Hyland. “Neural activity related to reaching and grasping in rostral and caudal regions of rat motor cortex”. In: *Behavioural Brain Research* 94.2 (1998), pp. 255–269. DOI: 10.1016/S0166-4328(97)00157-5.
- [39] D. N. Hill et al. “Primary motor cortex reports efferent control of vibration motion on multiple timescales”. In: *Neuron* 72.2 (2011), pp. 344–356. DOI: 10.1016/j.neuron.2011.09.020.

- [40] J. C. Erlich, M. Bialek, and C. D. Brody. “A cortical substrate for memory-guided orienting in the rat.” In: *Neuron* 72.2 (2011), pp. 330–43. DOI: 10.1016/j.neuron.2011.07.010.
- [41] R. S. Turner and M. R. DeLong. “Corticostriatal activity in primary motor cortex of the macaque.” In: *The Journal of neuroscience : the official journal of the Society for Neuroscience* 20.18 (2000), pp. 7096–7108.
- [42] J. H. Wu, J. V. Corwin, and R. L. Reep. “Organization of the corticostriatal projection from rat medial agranular cortex to far dorsolateral striatum”. In: *Brain Research* 1280 (2009), pp. 69–76. DOI: 10.1016/j.brainres.2009.05.044.
- [43] S. Lee, G. E. Carvell, and D. J. Simons. “Motor modulation of afferent somatosensory circuits.” In: *Nature neuroscience* 11.12 (2008), pp. 1430–8. DOI: 10.1038/nn.2227.
- [44] M. R. Baker, M. Javid, and S. A. Edgley. “Activation of cerebellar climbing fibres to rat cerebellar posterior lobe from motor cortical output pathways”. In: *Journal of Physiology* 536.3 (2001), pp. 825–839. DOI: 10.1111/j.1469-7793.2001.00825.x.
- [45] H. Jarratt and B. Hyland. “Neuronal activity in rat red nucleus during forelimb reach-to-grasp movements”. In: *Neuroscience* 88.2 (1999), pp. 629–642. DOI: 10.1016/S0306-4522(98)00227-9.

- [46] L. Petreanu et al. “Activity in motor–sensory projections reveals distributed coding in somatosensation”. In: *Nature* 489.7415 (2012), pp. 299–303. DOI: 10.1038/nature11321.
- [47] D. M. Schneider, A. Nelson, and R. Mooney. “A synaptic and circuit basis for corollary discharge in the auditory cortex”. In: *Nature* 513.7517 (2014), pp. 189–194. DOI: 10.1038/nature13724.
- [48] T. Graham Brown. “The Intrinsic Factors in the Act of Progression in the Mammal”. In: *Proceedings of the Royal Society B: Biological Sciences* 84.572 (1911), pp. 308–319. DOI: 10.1098/rspb.1911.0077.
- [49] S. Grillner and M. L. Shik. “On the descending control of the lumbosacral spinal cord from the "mesencephalic locomotor region"”. In: *Acta Physiol Scand.* 87.0001-6772 (1973), pp. 320–333.
- [50] D. M. Armstrong and T. Drew. “Discharges of pyramidal tract and other motor cortical neurones during locomotion in the cat.” In: *The Journal of Physiology* 346 (1984), pp. 471–495.
- [51] T. Drew, W. Jiang, and W. Widajewicz. “Contributions of the motor cortex to the control of the hindlimbs during locomotion in the cat.” In: *Brain research. Brain research reviews* 40.1-3 (2002), pp. 178–91.
- [52] T. Drew et al. “Role of the motor cortex in the control of visually triggered gait modifications.” In: *Canadian journal of physiology and pharmacology* 74.4 (1996), pp. 426–42.

- [53] E. Liddell and C. Phillips. “Pyramidal section in the cat”. In: *Brain* 67.1 (1944), pp. 1–9.
- [54] H. Barlow. “Cerebral cortex as model builder”. In: *Models of the visual cortex* 1985 (1985), pp. 37–46.
- [55] K. Doya. “What are the computations of the cerebellum, the basal ganglia and the cerebral cortex?” In: *Neural networks : the official journal of the International Neural Network Society* 12.7-8 (1999), pp. 961–974.
- [56] K. S. Lashley. “Studies of cerebral function in learning. III. The motor areas”. In: *Brain* 44 (1921), pp. 255–285. DOI: 10.1037/h0070668.
- [57] K. S. Lashley. “In search of the engram”. In: *Symposia of the Society for Experimental Biology* (1950).
- [58] C. S. Sherrington. “On Secondary and Tertiary Degenerations in the Spinal Cord of the Dog”. In: *Journal of Physiology* 6.4 (1885), pp. 177–191. DOI: 10.1113/jphysiol.1885.sp000195.
- [59] D. A. Oakley. “Cerebral cortex and adaptive behaviour”. In: *Brain, Behaviour and Evolution*. 1979, pp. 154–188.
- [60] D. Marple-Horvat et al. “Changes in the discharge patterns of cat motor cortex neurones during unexpected perturbations of on-going locomotion.” In: *Journal of Physiology* 462 (1993), pp. 87–113.
- [61] M. W. Miller. “The origin of corticospinal projection neurons in rat”. In: *Exp Brain Res* 67.2 (1987), pp. 339–351. DOI: 10.1007/BF00248554.

- [62] G. A. Metz and I. Q. Wishaw. “Cortical and subcortical lesions impair skilled walking in the ladder rung walking test: a new task to evaluate fore- and hindlimb stepping, placing, and co-ordination.” In: *Journal of neuroscience methods* 115.2 (2002), pp. 169–79.
- [63] T. M. Otchy et al. “Acute off-target effects of neural circuit manipulations”. In: *Nature* (2015). DOI: 10.1038/nature16442.
- [64] C. S. Sherrington. “Note on the Knee-jerk and the Correlation of Action of Antagonistic Muscles”. In: *Proceedings of the Royal Society of London* 52.315-320 (1893), pp. 556–564.
- [65] C. S. Sherrington. “Flexion-reflex of the limb, crossed extension-reflex, and reflex stepping and standing”. In: *The Journal of physiology* 40.1-2 (1910), pp. 28–121. DOI: 10.1113/jphysiol.1910.sp001362.
- [66] M. S. Arshian et al. “Vestibular nucleus neurons respond to hindlimb movement in the decerebrate cat”. In: *J Neurophysiol* 111.March 2014 (2014), pp. 2423–2432. DOI: 10.1152/jn.00855.2013.
- [67] T. Senoo et al. “Skillful manipulation based on high-speed sensory-motor fusion”. In: *2009 IEEE International Conference on Robotics and Automation* (2009), pp. 1611–1612. DOI: 10.1109/ROBOT.2009.5152852.
- [68] A. Coates, P. Abbeel, and A. Y. Ng. “Learning for control from multiple demonstrations”. In: *Proceedings of the 25th international conference*

- on Machine learning - ICML '08* (2008), pp. 144–151. DOI: 10.1145/1390156.1390175.
- [69] S. Papert. *The summer vision project*. 1966.
- [70] M. Spinka, R. C. Newberry, and M. Bekoff. “Mammalian Play: Training for the Unexpected”. In: *Source: The Quarterly Review of Biology* 76.2 (2001), pp. 141–168. DOI: 10.1086/393866.
- [71] N. A. Bernstein. *Dexterity and its Development*. Ed. by M. L. Latash and M. Turvey. L. Erlbaum Associates, 1996.
- [72] G. Lopes et al. “Bonsai: An event-based framework for processing and controlling data streams”. In: *Frontiers in Neuroinformatics* 9.7 (2015). DOI: 10.3389/fninf.2015.00007.
- [73] J. V. Jacobs. “Why we need to better understand the cortical neurophysiology of impaired postural responses with age, disease, or injury.” In: *Frontiers in integrative neuroscience* 8.August (2014), p. 69. DOI: 10.3389/fnint.2014.00069.
- [74] J. A. Eyre et al. “Functional corticospinal projections are established prenatally in the human foetus permitting involvement in the development of spinal motor centres.” In: *Brain : a journal of neurology* 123 (Pt 1 (2000), pp. 51–64. DOI: 10.1093/brain/123.1.51.
- [75] D. G. Lawrence and D. A. Hopkins. “The development of motor control in the rhesus monkey: evidence concerning the role of corticom-

- toneuronal connections.” In: *Brain : a journal of neurology* 99.2 (1976), pp. 235–54. DOI: 10.1093/brain/99.2.235.
- [76] E. Thelen. “Developmental origins of motor coordination: leg movements in human infants.” In: *Developmental psychobiology* 18.1 (1985), pp. 1–22. DOI: 10.1002/dev.420180102.
- [77] C. von Hofsten. “Mastering Reaching and Grasping: The Development of Manual Skills in Infancy”. In: *Advances in Psychology* 61.C (1989), pp. 223–258. DOI: 10.1016/S0166-4115(08)60023-0.
- [78] I. Q. Whishaw. “Loss of the innate cortical engram for action patterns used in skilled reaching and the development of behavioral compensation following motor cortex lesions in the rat”. In: *Neuropharmacology* 39.5 (2000), pp. 788–805. DOI: 10.1016/S0028-3908(99)00259-2.
- [79] T. D. Farr and I. Q. Whishaw. “Quantitative and qualitative impairments in skilled reaching in the mouse (*Mus musculus*) after a focal motor cortex stroke”. In: *Stroke* 33.7 (2002), pp. 1869–1875. DOI: 10.1161/01.STR.0000020714.48349.4E.
- [80] J. Schindelin et al. “Fiji: an open-source platform for biological-image analysis.” In: *Nature methods* 9.7 (2012), pp. 676–82. DOI: 10.1038/nmeth.2019.
- [81] S. van der Walt, S. C. Colbert, and G. Varoquaux. “The NumPy Array: A Structure for Efficient Numerical Computation”. In: *Computing in*

Science & Engineering 13.2 (2011), pp. 22–30. DOI: 10.1109/MCSE.2011.37.

- [82] W. McKinney. “Data Structures for Statistical Computing in Python”. In: *Proceedings of the 9th Python in Science Conference* (2010), pp. 51–56.