1Universal metabarcoding of pico- to mesoplankton reveals seasonal 2dynamics and a bacterial bloom

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4Christian Wurzbacher^{1, 2, 3}, Katrin Attermeyer^{2, 4}, Marie Therese Kettner², Clara ⁵Flintrop³, Norman Warthmann^{3, 5, 6}, Sabine Hilt⁶, Hans-Peter Grossart^{2, 7}, Michael 6T. Monaghan^{3, 6}

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81 University of Gothenburg, Department of Biological and Environmental Sciences, Box 461, 940530 Göteborg, Sweden 102 Leibniz-Institute of Freshwater Ecology and Inland Fisheries, Alte Fischerhütte 2, 16775 11Stechlin, Germany 123 Berlin Center for Genomics in Biodiversity Research, Königin-Luise-Str. 6-8, 14195 Berlin, 13Germany 144 Uppsala University, Department of Ecology and Genetics, Limnology, Norbyvägen 18D, 75236 15Uppsala, Sweden 165 The Australian National University, Research School of Biology, Linnaeus Way, Canberra, ACT 172601, Australia 186 Leibniz-Institute of Freshwater Ecology and Inland Fisheries, Müggelseedamm 301, 12587 19Berlin, Germany 207 Potsdam University, Institute of Biochemistry and Biology, Maulbeerallee 2, 14469 Potsdam, 21Germany 22 23Corresponding authors: Christian Wurzbacher (christian@wurzbacher.cc), Katrin 24Attermeyer (katrin.attermeyer@ebc.uu.se) 25

26The authors declare no conflict of interest

27**Abstract**

28Most studies of biodiversity focus on either macroscopic or microbial 29communities, with little or no simultaneous study of eukaryotes and 30prokaryotes. We tested whether a universal metabarcoding approach could be 31used to study the total diversity and temporal dynamics of aquatic pico- to 32mesoplankton communities in a shallow temperate lake. The approach 33revealed significant changes in the relative abundance of eukaryotic and 34prokaryotic plankton communities over a period of three months. These 35patterns, based on sequencing reads, fit with counts using traditional methods. 36We also witnessed the bloom of a conditionally rare bacterial taxon belonging 37to *Arcicella*, a genus that has been largely overlooked in freshwaters. Our data 38demonstrate the potential of universal metabarcoding as a complement to 39traditional studies of plankton communities, and for long-term monitoring 40across a broad range of organisms.

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42Introduction

⁴³Microbial communities play key roles in ecosystems and knowledge on their ⁴⁴composition and dynamics is critical for understanding ecosystem functions ⁴⁵and services. Recently, Barberán et al. (2014) emphasized the importance of ⁴⁶studying microbial diversity as an integral part of total biodiversity. In aquatic ⁴⁷ecosystems, plankton communities are a mixture of prokaryotes and ⁴⁸eukaryotes of a wide range of sizes, and the traditional separation into size ⁴⁹classes and domains, with their largely independent research disciplines, ⁵⁰impairs a full integration of microbial communities into ecological concepts. ⁵¹Simultaneous studies of diversity of all three domains are rare (< 1% of all ⁵²studies, SI1) in environmental sequencing surveys (e.g., Steele *et al.*, 2011,

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⁵³Lima-Mendez *et al.*, 2015) and size pre-selection steps are common in most ⁵⁴analyses. Nonetheless, bacterioplankton should exhibit temporal dynamics that ⁵⁵are similar to those of macroorganisms (Shade et al. 2013), suggesting that the ⁵⁶monitoring of temporal dynamics of all organisms has great potential for ⁵⁷identifying ecological interactions (Fuhrman *et al.*, 2015). In addition, metadata ⁵⁸on organismal ratios such as eukaryotes : prokaryotes or heterotrophs : ⁵⁹phototrophs could be useful for monitoring ecosystem changes. Here we used a ⁶⁰universal metabarcoding approach to simultaneously study the dynamics of ⁶¹both prokaryotic and eukaryotic plankton (pico- to mesoplankton) in a lake that ⁶²exhibits pronounced seasonal plankton dynamics.

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64 Methods

65We applied universal metabarcoding to water samples of the eutrophic, shallow 66temperate Lake Gollin (Kleiner Gollinsee) in northwestern Germany. Lake Gollin 67was sampled monthly as part of a study of bacteria, phyto- and zooplankton 68from April 2010 to December 2011 (Brothers *et al.*, 2013; see Sl1 for 69parameters and methods). Briefly, we took monthly samples in spring 2010 70(April – June) from littoral water, pelagic water, and water above the pelagic 71sediment. DNA was extracted from filters (0.22 μm Sterivex; Millipore, 72Germany) using commercial kits and mechanical bead-beating. We used the V9 73region of the ribosomal small subunit for taxa detection (Engelbrektson *et al.*, 742010). Methodological discussions on a related integrative single marker can 75be found in Parada *et al.* (2015). Amplicons were constructed and sequenced 76following the conditions described for pyrosequencing by Hölker *et al.* (2015) 77with the modification that we employed the AccuPrime High Fidelity Polymerase 78(Invitrogen, Carlsbad, USA). Sequences were processed in Mothur (version

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⁷⁹1.24.1; Schloss *et al.*, 2009) and classified with SINA aligner (version 1.2.11; ⁸⁰Pruesse *et al.*, 2012) against the SILVA SSU reference database (115 Ref NR 99, ⁸¹www.arb-silva.de). After detection of a bacterial bloom (see below) we designed ⁸²a PCR test for *Arcicella* bacteria for the monthly samples. PCR products were ⁸³quantified by gel electrophoresis assigning them to four intensities.

85Results & Discussion

86There was a pronounced shift in dominance from mainly eukaryotes in April to 87predominantly prokaryotes in June (Fig. 1a; SI2 for taxa browsing). This was 88accompanied by an increased heterotrophs : phototrophs ratio (SI3). Archaea 89were not abundant in Lake Gollin, despite their high abundance in some 90meromictic lakes (Gies et al., 2014). In the winter prior to our study, a 91significant fish-kill led to a bloom of herbivorous ciliates in April 2010 that was 92confirmed by our data (Fig. 1d). This likely exerted a very high grazing pressure 93upon the small plankton (<5 μm; Lischke *et al.*, 2016). Potentially, the abrupt 94disappearance of ciliates in May – June (Fig. 1d) following increasing crustacean 95abundances (Hilt et al., 2015) reduced the abundance of algae and bacterial ⁹⁶grazers. This may have opened a niche for the detected bacteria, in particular 97of the genera Aricella and Variovorax (see below). Our molecular data 98complement traditional counting data, following the trend of the sum 99parameters of phyto-, zoo- and bacterioplankton (SI3). The approach may 100therefore have the potential to follow dynamics of entire plankton communities 101in a single analysis.

102Bacterial OTUs of the genera *Arcicella* and *Variovorax* were major 103representatives of the bacterial "bloom" that dominated the lake in June. 104*Arcicella* was more abundant in the upper water layers and potentially

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105colonized the water from the neuston, whereas Variovorax was more abundant 106above the sediment suggesting colonization from the sediment (Fig. 1b, c). 107Interestingly, Arcicella was rare in April and May (<0.2%). There are few reports 1080f blooms of rare bacterial taxa (e.g., Gilbert et al., 2012) and causes for single 109taxa bacterial blooms are often connected to phytoplankton events (Bizic-110 lonescu et al. 2014). Arcicella re-occurred the following year (Fig. 2) and we 111were able to relate its appearance to the phyto- and zooplankton dynamics 112(SI1). In contrast to the Vibrio sp. bloom described by Gilbert et al. (2012), our 113Arcicella OTU can be classified as conditional/recurrent rare taxa (Lynch and 114Neufeld, 2015). Searching through existing freshwater data sets, we found 115Arcicella to be a prevalent freshwater lineage on a global scale, especially in 116lotic ecosystems. Arcicella was reported as the second most abundant OTU 117(6.8%) in all large circumpolar streams (Crump et al., 2009). In other studies, 118Arcicella was detected but not explicitly reported. For example, a single 119Arcicella OTU was among the dominant taxa in the Danube River, comprising 1202% ± 2% (Savio pers. comm.; Savio et al., 2015) and a single Arcicella OTU 121 reached 12 ± 4% in a small turbid glacial lake (Peter pers. comm.; Peter and 122Sommaruga, 2016). Despite those previous appearances, their autecology has 123never been discussed, suggesting that we have probably revealed a thus far 124unrecognized major freshwater lineage.

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126Using universal metabarcoding we could monitor major changes in freshwater 127pico- to mesoplankton with a single water sample analysis. Changes in relative 128abundance of OTUs matched the seasonal dynamics that have previously been 129reported for this lake. The discovery of a conditionally rare taxon supports the 130observed plankton dynamics for the bacterioplankton, since such taxa are of a

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131disproportional significance for the overall community dynamics (Shade et al., 1322014). Integrative analysis may help to answer broader ecological guestions 133related to the role of keystone species, ecosystem resilience and resistance, 134and cross-domain interactions of species (Lima-Mendez et al., 2015). Universal 135metabarcoding represents a valuable means of studying plankton dynamics in 136aguatic systems and shows promise for long-term monitoring.

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138Acknowledgements

139We would like to thank Uta Mallock for chemical analysis, Susan Mbedi for 140assistance in sequencing and Marie Jeschek for assistance in data processing. 141This is publication #027 of the Berlin Center for Genomics in Biodiversity 142Research. Financial support was provided by two projects of the Pact for 143Innovation and Research of the Gottfried Wilhelm Leibniz Association "Terralac" 144and "TEMBI". R. Tischbier (Stiftung Pro Artenvielfalt) kindly provided access to 145the lake.

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147**Conflict of interest**

148The authors declare no conflict of interest.

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150Supplementary information is available

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153**References**

154Barberán A, Casamayor EO, Fierer N. (2014). The microbial contribution to

155macroecology. Front Microbiol 5: 203.

156Bižić-Ionescu M, Amann R, Grossart, HP. (2014). Massive regime shifts and high

157activity of heterotrophic bacteria in an ice-covered lake. *PloS one* 9(11): 158e113611.

¹⁵⁹Brothers SM, Hilt S, Attermeyer K, Grossart HP, Kosten S, Lischke B, Mehner T, ¹⁶⁰Meyer N, Scharnweber K, Köhler J. (2013). A regime shift from macrophyte to ¹⁶¹phytoplankton dominance enhances carbon burial in a shallow, eutrophic lake. ¹⁶²Ecosphere 4(11): 137. <u>http://dx.doi.org/10.1890/ES13-00247.1</u>

163Crump BC, Peterson BJ, Raymond P., Amon RM, Rinehart A, McClelland JW,
164Holmes RM. (2009). Circumpolar synchrony in big river bacterioplankton. *Proc*165*Natl Acad Sci* 106(50): 21208-21212.

¹⁶⁶Engelbrektson A, Kunin V, Wrighton KC, Zvenigorodsky N, Chen F, Ochman H, ¹⁶⁷Hugenholtz P. (2010). Experimental factors affecting PCR-based estimates of ¹⁶⁸microbial species richness and evenness. *ISME J* 4(5): 642-647.

¹⁶⁹Fuhrman JA, Cram JA, Needham DM. (2015). Marine microbial community
¹⁷⁰dynamics and their ecological interpretation. *Nat Rev Microbiol* 13(3): 133-146.
¹⁷¹Gies EA, Konwar KM, Beatty JT, Hallam SJ. (2014). Illuminating microbial dark
¹⁷²matter in meromictic Sakinaw Lake. *Appl Environ Microbiol* 80(21): 6807-6818.
¹⁷³Gilbert JA, Steele JA, Caporaso JG, Steinbrück L, Reeder J, Temperton B et al.
¹⁷⁴(2012). Defining seasonal marine microbial community dynamics. *ISME J* 6:
¹⁷⁵298–308.

176Hilt S, Wanke T, Scharnweber K, Brauns M, Syväranta J, Brothers S et al. (2015). 177Contrasting response of two shallow eutrophic cold temperate lakes to a partial 178winterkill of fish. *Hydrobiologia* 749: 31-42.

179Hölker F, Wurzbacher C, Weißenborn C, Monaghan MT, Holzhauer SI, Premke K. 180(2015). Microbial diversity and community respiration in freshwater sediments 181influenced by artificial light at night. *Philos T Roy Soc B* 370(1667): 20140130. 182Lima-Mendez G, Faust K, Henry N, Decelle J, Colin S, Carcillo F. et al. (2015).

183Determinants of community structure in the global plankton interactome.184Science 348(6237): 1262073.

185Lischke B, Weithoff G, Wickham SA, Attermeyer K, Grossart HP, Scharnweber K 186et al. (2016). Large biomass of small feeders: ciliates may dominate herbivory 187in eutrophic lakes. *J Plankton Res* 38(1): 2–15.

188Parada AE, Needham DM, Fuhrman JA. (2015). Every base matters: assessing 189small subunit rRNA primers for marine microbiomes with mock communities, 190time series and global field samples. *Environ Microbiol* doi: 10.1111/1462-1912920.13023.

192Peter H, Sommaruga R. (2016). Shifts in diversity and function of lake bacterial
193communities upon glacier retreat. *ISME J* doi: 10.1038/ismej.2015.245
194Pruesse E, Peplies J, Glöckner FO. (2012). SINA: accurate high-throughput
195multiple sequence alignment of ribosomal RNA genes. *Bioinformatics* 28(14):
1961823-1829.

197Savio D, Sinclair L, Ijaz UZ, Parajka J, Reischer GH, Stadler P, et al. (2015).
198Bacterial diversity along a 2600 km river continuum. *Environ Microbiol* 17(12):
1994994-5007.

200Schloss PD, Westcott SL, Ryabin T, Hall JR, Hartmann M, Hollister EB, et al. 201(2009). Introducing mothur: open-source, platform-independent, community-202supported software for describing and comparing microbial communities. *Appl* 203*Environ Microbiol* 75(23): 7537-7541.

204Shade A, Caporaso JG, Handelsman J, Knight R, Fierer N. (2013). A meta-205analysis of changes in bacterial and archaeal communities with time. *ISME J* 2067(8): 1493-1506.

207Shade A, Jones SE, Caporaso JG, Handelsman J, Knight R, Fierer N, Gilbert JA. 208(2014). Conditionally rare taxa disproportionately contribute to temporal

15 16

209changes in microbial diversity. *MBio* 5(4): e01371-14.

210Steele JA, Countway PD, Xia L, Vigil PD, Beman JM, Kim DY, et al. (2011). Marine 211bacterial, archaeal and protistan association networks reveal ecological 212linkages. *ISME J* 5(9): 1414-1425.

213Figure Legends

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215**Figure 1.** Spatial and temporal changes of microorganisms in Lake Gollin from 216three samplings (April, May, June) and locations (littoral, pelagic, and above 217sediment) displaying abundances (mean ± standard deviation) based on a 218rarefied OTU matrix (1744 reads/sample) of (a) all three domains with a total of 2191307 bacteria, 388 eukaryote, and 190 archaea OTUs in the dataset; (b) the 220dominant *Arcicella* OTU; (c) the dominant *Variovorax* OTU and (d) the sum of all 221ciliate OTUs (70 OTUs).

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223**Figure 2.** Seasonal appearance of *Arcicella* assessed via a PCR assay and 224evaluated based on the band intensity in four categories: 0 = no product, 1 =225very weak product, 2 = weak product, 3 = medium product, 4 = strong 226product. *Arcicella* exhibited pronounced maxima and minima over the course of 227the 2 years and appeared in the particle-attached (> 5 µm) and free-living 228fraction (0.2 - 5 µm).



