1 Landscape connectivity of a noxious invasive weed: 2 human-aided or natural dispersal? 3 4 Diego F. Alvarado-Serrano (corresponding author) 5 Department of Ecology and Evolutionary Biology 6 University of Michigan 7 E-mail: dalvarad@umich.edu 8 9 Megan Van Etten 10 Department of Ecology and Evolutionary Biology University of Michigan 11 12 E-mail: mvanette@umich.edu 13 14 Shu-Mei Chang 15 **Department of Plant Biology** University of Georgia 16 17 E-mail: smchang@uga.edu 18 19 Regina S. Baucom 20 Department of Ecology and Evolutionary Biology University of Michigan 21 22 E-mail: rsbaucom@umich.edu 23

Landscape connectivity of a noxious invasive weed: human-aided or natural dispersal?

# **ABSTRACT**

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

Examining how the landscape may influence gene flow is at the forefront of understanding population differentiation and adaptation. Such understanding is crucial in light of ongoing environmental changes and the elevated risk of ecosystems alteration. In particular, knowledge of how humans may influence the structure of populations is imperative to elucidate their role in shaping the evolution of other species, and specifically how humans may alter the balance between genetic drift and selection. Here we characterize the population genetic structure of *Ipomoea purpurea*, a noxious invasive weed, and assess the relative roles of natural and human-driven landscapes on genetic differentiation. By combining rigorous statistical analyses and a combination of different molecular markers, we detect both common and marker-specific patterns of genetic connectivity and identify human-aided migration as an important component shaping the evolutionary history of this species. In particular, we identified human population density as an important predictor of pairwise population differentiation, suggesting that the agricultural and/or horticultural trade may be involved in maintaining some level of connectivity across agricultural fields. Climatic variation, primarily temperature, appears as an additional predictor when considering agricultural fields in the northern United States. We discuss the implications of these results and the approach we followed in the context of understanding agricultural weed and invasive species' expansions, as well as the challenges and promises of current landscape genetics research.

**Keywords:** human-aided migration, landscape genetics, morning glory, population structure

# **INTRODUCTION**

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

Elucidating routes and levels of migration between subpopulations of a species is essential to understand the interplay between gene flow, adaptation, genetic drift, and selection, and hence the forces that shape its evolutionary trajectory (Barrowclough 1980; Slatkin 1985). Landscape features—such as rivers, mountain ranges, crop fields, and urban areas—can impact levels of gene flow between populations by determining dispersal rates and routes (McRae 2006; Cushman et al. 2006) as well as by influencing the likelihood of successful establishment of immigrants (Nosil et al. 2008; Sexton et al. 2014; Wang & Bradburd 2014). Landscape features can also indirectly condition the effect of gene flow through its effect on local effective population sizes since the actual role that migration plays in the evolution of a species is driven by the fraction of the local population size that correspond to immigrants (Wright 1949; Slatkin 1985). Consequently, the landscape, loosely defined as an area with spatially variable biotic and abiotic factors (Holderegger et al. 2010), creates the stage for spatially heterogeneous functional population connectivity, conditioned by species' specific physiological tolerances and behavioral preferences (Clobert et al. 2012). In this way, the landscape plays a pivotal role in the evolution of species. Previous studies have identified common responses of species to their surrounding landscape across a range of very different systems, including evidence of frequent long distance dispersal (Berthouly-Salazar et al. 2013), dispersal over large unsuitable areas (Manel & Holderegger 2013), and a prevalent mixed effect of local and intervening landscapes on structuring populations (Sexton et al. 2014). Such findings have led to novel emergent evolutionary hypotheses that better represent the complex interaction of processes determining

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

the evolutionary fate of natural populations (Dyer 2015a). However, our current knowledge of the interplay between landscape features and the genetic structure of populations comes mostly from human-avoider species (Blair 2001) facing indirect human impact (e.g., Culley et al. 2007; McRae & Beier 2007; Diniz-Filho et al. 2009; but see Harris et al. 2016). In contrast to species that almost exclusively depend on natural dispersal agents, species in heavily human-dominated ecosystems may exploit human activities to maintain gene flow among populations and expand their ranges (Everman & Klawinski 2013; Fountain et al. 2014). Such species may be capable of maintaining population connectivity over vast geographic ranges (Trakhtenbrot et al. 2005) by overcoming landscape features that would otherwise represent natural barriers and reach dispersal distances that could be orders of magnitude greater than those attain under natural agents or do it under much smaller time frames (Mack & Lonsdale 2001; Ricciardi 2007). In this way, by facilitating dispersal humans have the potential to: i) condition the balance between drift and selection (Slatkin 1985; Lenormand 2002), ii) introduce relevant genetic variation to local populations (Kolbe et al. 2004), iii) prevent local extinction or favor recolonization (Fountain et al. 2014), and alter the overall genetic constitution of populations (Bataille et al. 2011). Human-aided migration—intentional or unintentional— is particularly prevalent in plants (Hodkinson et al. 2007; Wichmann et al. 2009; Auffret & Cousins 2013), where it has had major impacts on the distribution of species and stability of communities (Simberloff 2013 and references therein). Yet, the open question remains: how important is human-aided migration relative to migration through other more natural means in maintaining population connectivity in plant populations? This question is further complicated in plants because in this group dispersal is a two-step process that involves the movement of both pollen and/or seeds (Holderegger et al.

2010), and these separate vectors of gene flow entail the movement of different genetic states (haploid vs. diploid) as well as different dispersal agents and mechanisms (i.e., wind, water, animal-mediated) (Levin & Kerster 1974).

A particularly amenable system to study the interaction between natural and human-aided dispersal comes from agricultural weed populations. Agricultural weeds face a highly dynamic landscape characterized by frequent spatial rearrangements (expansion of agricultural front, increased fragmentation) and a constantly changing environment (crop rotation, agricultural chemical use, climatic abnormalities) (Menchari *et al.* 2007; Meehan *et al.* 2011) that certainly impact their opportunities for survival and local adaptation through its effect on population connectivity (Margosian *et al.* 2009). Under these conditions, human-aided migration is expected to be critical for weeds' success, but knowledge on how or if weedy plant populations are able to maintain connectivity through such complex landscape matrix of croplands, grasslands, natural and urban areas is limited. This is a striking omission given that agricultural weeds impose severe economic costs (on the order of 33B USD per year in US agriculture alone, Pimentel *et al.* 2005).

As a first step into investigating the impact of human activities on structuring genetic diversity, we estimate the intensity and extent of migration from genetic data and evaluate how multiple landscape features influence genetic connectivity of a noxious agricultural weed, *I. purpurea*, using two different sets of molecular markers (nuclear microsatellites and a genomewide panel of SNPs). Specifically, we ask the following questions: 1) Which natural or human-influenced landscape features—soils, elevation, climate, landcover, crop types, human population density—promote or constrain genetic connectivity between populations of this agricultural

weed? 2) Do human-associated landscapes disproportionally influence genetic connectivity in this species, suggesting that this weed's evolutionary fate is strongly linked to human-aided dispersal?, and 3) what additional insights can we gain from a broader representation of the genome than traditionally used in landscape genetics studies (typically microsatellites and organelle DNA)? Taken together, the answers to these questions offer deeper knowledge of the interaction between human activities, landscape features, and population structure of noxious weeds and hence contribute to improve effective management and control of these damaging plants. More generally, these answers contribute to deepen our understanding of the interaction between geographic setting and population differentiation, adaptation, and persistence (Taylor *et al.* 1993).

# MATERIALS AND METHODS

# **Study system**

Ipomoea purpurea, the common morning glory, is an agricultural weed well suited for the study of human-aided migration on population connectivity because of its relatively well-characterized biology, its recent introduction into the United States, and its close association with agricultural crops. Its tight association with humans since its introduction allows for assessing the impact of recent landscapes on structuring genetic variance under the initial simplifying assumption of evolutionary equilibrium (Marko & Hart 2011). Ipomoea purpurea is a noxious weed with a widespread distribution that includes highly heterogeneous landscapes in the Eastern, South- and Mid-western regions of the United States (Culpepper 2006; Webster & Nichols 2012). It is a self-compatible annual bumblebee-pollinated vine, with heavy seeds, and is

found primarily in agricultural fields and disturbed areas (Tiffin & Rausher 1999; Baucom & Mauricio 2008). It is currently one of the most problematic weeds of agriculture (Webster & Nichols 2012) and is capable of infestations leading to substantial decline in crop (although not quantified for *I. purpurea*, related *Ipomoea* species may cause declines up to 80% of crop yield, Rogers *et al.* 1996). This species exhibits in general low-level resistance to the commonly used herbicide glyphosate (Baucom & Mauricio 2004, 2008), although the exact resistance level varies widely among populations of this species (Kuester *et al.* 2015). This species is also a major concern for conservation given its naturalization in multiple regions throughout the world and its aggressiveness as an invasive (Chaney & Baucom 2012; Fang *et al.* 2013). Previous analyses of its genetic diversity and differentiation using a panel of 15 nuclear microsatellite markers identify limited population structure and a less-than definitive isolation-by-distance pattern (Kuester *et al.* 2015). It remains to be seen, however, the role that landscape features, both natural and humandriven, play in structuring genetic variation within this species.

# **Data compilation**

To capture the plausible effect of both natural and disturbed landscapes on structuring genetic diversity in *I. purpurea*, we compiled a diverse set of GIS data for the continental US from a variety of sources (Table S1). These data encapsulate human activities (human population density, landcover, and planted crops) as well as the geographical setting of *I. purpurea* (elevation, climate—19 variables summarizing central tendencies and variability patterns in temperature and precipitation, soil—8 variables summarizing the texture, pH, and organic and inorganic content of the top 20cm of soil). We first processed all these data into landscape layers at a common spatial resolution of  $10 \text{km}^2$  and a common spatial extent around the US states with

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

available samples (Fig. 1). These spatial resolution and extent were chosen to maintain a practical balance between scale and analytical manageability given available computational resources. To reduce dimensionality, we opted to perform two separate Principal Component Analyses (PCAs) on the 19 climatic and 8 soil layers, respectively. For all subsequent analyses we kept the resulting first principal component of each of these analyses, which accounted for over 60% of the variance and primarily summarized temperature temporal gradients and soils' pH and sandiness, respectively (Table S2). With the objective of estimating the genetic connectivity of populations of *I. purpurea*, we compiled genetic data on an extensive panel of 15 previously optimized microsatellite loci (Molecular Ecology Resources Primer Development Consortium; et al. 2013). These data encompass a total of 597 individuals from 31 localities (with a minimum of 8 individuals per locality) (Fig. 1; Table S3). All individuals were collected in 2012 from farms across the range of I. purpurea in the United Sates (for further details and basic genetic variability analyses see Kuester et al. 2015). In addition, to obtain a more comprehensive representation of the genome of I. purpurea and assess the robustness of results in light of coalescent and mutational variance (Nielsen & Slatkin 2013), we generated a Next Generation Sequencing (NGS) dataset from a subsample of individuals. In conjunction with the microsatellite data, this complementary dataset, which offers a more widespread representation of this species' genome, is expected to lead to more comprehensive population genetic inferences (Bohonak & Vandergast 2011; Epps & Keyghobadi 2015).

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

To generate the NGS dataset, we constructed genome-wide Genotype By Sequencing (GBS) libraries for 80 individuals sampled across 8 of localities (10 individuals per locality; Fig 1). The GBS library was developed using 7ng of genomic DNA extracted from leaf or cotyledon tissue using SNPsaurus' (Oregon, USA) nextRAD technology, which uses a selective PCR primer to amplify consistent genomic loci among individuals. Similarly to RAD-Seq sequences (Rowe et al. 2011) in which the DNA flanking a restriction enzyme cut site is selected for amplification, nextRAD amplifies sequences that correspond to the DNA downstream of a short selective priming site. Samples were first fragmented and then ligated to short adapter and barcode sequences using a partial Nextera reaction (Illumina; California, USA) before being amplified using Phusion® Hot Start Flex DNA Polymerase (New England Biolabs; Massachusetts, USA). The 80 dual-barcoded PCR-amplified samples were pooled and the resulting libraries were purified using AMPure XP beads (Agencourt Bioscience Corporation; Massachusetts, USA) at 0.7x. The purified library was then size selected to 350-800 base pairs and sequenced using two runs of an Illumina NextSeq500 sequencer (Genomics Core Facility, University of Oregon). The resulting sequences were analytically processed by combining the reads of 16 randomly selected individuals (of the 80 sequenced) to create a pseudo-reference genome. This was done after removing loci with read counts above 20,000, which presumably corresponded to repetitive genomic material, and loci with read counts below 100, which presumably corresponded to offtarget or read errors. The filtered reads were aligned to each other using BBMap (http://sourceforge.net/projects/bbmap/). All parameters were set to default values with the exception of minimum sequence identity, which was set to 0.93 to identify alleles. A single read instance was chosen to represent the locus in the pseudo-reference. This resulted in a total of

263,658 loci. All reads from each of the 80 individuals were then aligned to the pseudo-reference using BBMap (http://sourceforge.net/projects/bbmap/) and converted to a vcf genotype table, using Samtools (Li *et al.* 2009) and bcftools (Li 2011), after filtering out nucleotides with a quality score of 10 or worst. The resulting vcf table was filtered using vcftools (Danecek *et al.* 2011) for SNPs with a minimum allele frequency of 0.02, a minimum read depth of 5, and a maximum 15% of missing data. This resulted in 9774 variable regions. Loci were further filtered using vcftools to exclude loci with less than 5 high quality base-calls and with more than 20% missing data or an average of less than 20 high quality base calls. This resulted in a final panel of 8210 Single Nucleotide Polymorphisms (SNPs) that we used in all subsequent analyses.

# **Population structure analyses**

We first conducted a series of preliminary analyses to characterize the overall genetic structure of I. purpurea. All analyses were run separately for the microsatellite (SSR, hereafter) and SNP datasets given their different sampling and geographic coverage (Fig. 1). First, we examined population differentiation by estimating  $F_{ST}$  using GenAlex v6.5 (Peakall & Smouse 2012) (because similar global  $F_{ST}$  and  $R_{ST}$  estimates were obtained for the SSR dataset, we opted to report  $F_{ST}$  values only to allow direct comparisons with the SNP dataset). We then estimated contemporary effective population size for each sampled locality in NeEstimator v2 using the excess heterozygous method (Do  $et\ al.\ 2014$ ). We performed this latter analysis to assess the possibility of whether effective rate of migration (Nm) may be asymmetric in response to differences in population size (e.g., greater migration into satellite populations).

230

231

232

233

234

235

236

237

238

239

240

241

242

243

244

245

246

247

248

249

250

251

In addition, we assessed population admixture and spatial genetic clustering using 2 spatially explicit Bayesian clustering algorithms: BAPS (Corander et al. 2004) and TESS (Chen et al. 2007). BAPS was run using its spatial clustering algorithm (Cheng et al. 2013) with 10 runs for each K value tested, whereas TESS was run using the admixture algorithm and a BYM model (i.e., using a spatial prior of admixture proportions; Durand et al. 2009b) with 10 runs per K value, and without using geographic weights. The TESS model, with the lowest DIC was chosen as the optimal model (Durand et al. 2009a). For both analyses K values ranging from 2 to the maximum number of sampled locations (31 and 10 for the SSR and SNP datasets, respectively) were tested. Results were generally consistent between BAPS and TESS analyses; thus, here we report exclusively the TESS results Additionally, following Wang et al. (2009), we complemented these analyses with an Analysis of Molecular Variance (AMOVA; Excoffier et al. 1992) run in GenAlex (Peakall & Smouse 2012) using 9999 permutation replicates. This analysis partitioned the variance into regions based on the spatial genetic clusters previously identified to quantify the fraction of the genetic variance explained by spatial genetic clusters, and hence the relative importance of spatial genetic structure in *I. purpurea*. Finally, we investigated population connectivity by estimating levels of recent migration between sampled localities through the identification of individuals of mixed ancestry using BayesAss (Wilson & Rannala 2003). BayesAss is a program that uses individual multilocus genotypes and a Markov Chain Monte Carlo (MCMC) algorithm to probabilistically distinguish between immigrants and long-term native individuals (Wilson & Rannala 2003). It provides posterior probability distributions of individual immigrant ancestries (i.e., the probability that an individual is a first or second generation migrant from each of the populations in the sample). We ran BayesAss for 6 million generations using default parameter settings, and discarded the first 2 million generations as burn-in (Dyer 2009). For each marker dataset, we repeated this analysis 3 times (for a total of 18 million generations) and combined the results from the 3 replicates for our final inference. Then, using a posterior probability cut-off of 0.75 we assign individuals' ancestry. It is important to note that because of computational limits we had to randomly subsample our set of SNPs to 400 SNPs for this analysis. We complemented this analysis with a spatial autocorrelation analysis run in GenAlex (Peakall & Smouse 2012) to evaluate the overall spatial scale of genetic turnover.

# Landscape genetics analyses

After assessing overall population structure of *I. purpurea*, we evaluated the association between landscape features and genetic differentiation. To do this, we first converted our pairwise F<sub>ST</sub> estimates into conditional genetic distances (Dyer *et al.* 2010) using GeneticStudio (Dyer 2009). Briefly, conditional distances are measures of pairwise genetic distance derived from population networks, constructed based on the degree of genetic similarity between sampled localities (Dyer & Nason 2004). Because these networks are pruned based on the principle of conditional independence of the total among population genetic covariance (the specific method of pruning used is edge deviance; Magwene 2001), conditional distances reflect genetic similarity between localities that better capture direct gene flow as opposed to connectivity driven by intervening localities (Dyer 2015b).

The climate, crops, elevation, landcover, population density, and soils landscape layers (Table S1) were converted into landscape resistance layers by assigning a resistance value to each

276

277

278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

297

landscape feature in these layers to reflect the difficulty that each feature offers to the movement of gametes or individuals (i.e., pollen or seeds). It is important to note that in contrast to previous studies that typically rely on expert opinion for resistance assignment, we utilized an unbiased statistical optimization to avoid the sensitivity of results to subjective resistance assignment (Spear et al. 2010). Specifically, resistance values were optimized through a genetic algorithm approach, which is a heuristic stochastic optimization algorithm that emulates the process of inheritance, mutation, and natural selection (Mitchell 1996). Briefly, in this search algorithm a population of individuals with traits encoded by unique combinations of model parameters (resistance assignment proposals in our case) is allowed to compete with each other based on the fitness associated with the traits it carries (Peterman et al. 2014). Specifically, in Peterman's (2014) implementation of this algorithm, which we followed here, individuals' fitness is estimated by the relative quality of a MLPE.lmm model (Maximum Likelihood Population Effects – Linear Mixture Model) that evaluates the association between pairwise genetic distance and landscape cumulative resistance between localities, estimated in Circuitscape (Shah & McRae 2008). Individuals with parameter settings (resistance assignments in our case) that result in better models, as measured by a Deviance Information Criterion (DIC) score, are preferentially represented in the following generation. Offspring modifications introduced by mutations (i.e., small resistance assignment perturbations) allow for exploration of the parameter space. The algorithm is stopped once a large number of generations have passed without significant improvement in fitness.

We implemented Peterman's (2014) algorithm in R (package ResistanceGA; Peterman 2014)

allowing for the independent optimization of each of our landscape layers. The optimal resistance

landscapes identified in this way were then used to run a final univariate MLPE.lmm model to identify the association between landscape features and conditional genetic distances between localities. In addition, we run separate simple and partial db-RDA (distance-based Redundancy Analyses; Legendre & Anderson 1999)—the latter uses the geographic distance between populations as a covariate to account for the concurrent increase in cumulative resistance with geographic distance. We opted for db-RDA instead of the most commonly used partial Mantel test given the statistical issues of the latter (Raufaste & Rousset 2001; Guillot & Rousset 2013). We run this latter analysis using the package vegan (Oksanen et al. 2015) in R and assessed significance with 9999 permutations. Finally, to identify the relative contribution of natural and human-driven landscape features we ran a Multiple Regression on Distance Matrices (MRDM; Legendre et al. 1994), which has been identified as one of the best performing methods for evaluating the interplay between landscape features and genetic connectivity (Balkenhol et al. 2009). Before running these regressions, we standardized all optimized resistance layers to mean of zero and variance of 1 (Dyer et al. 2010). These final regressions included geographic distance as a predictor and were run in R (using package ecodist; Goslee & Urban 2007) using 10,000 permutations to assess significance. In none of our analyses did we implement a Bonferroni correction for multiple testing because of the overly conservative nature of this correction (Nakagawa 2004; Glickman et al. 2014). Instead we applied a false recovery rate correction (Benjamini & Hochberg 1995) using the function *p.adjust* in R (R Core Development Team 2016).

# **RESULTS**

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

320

#### **Population structure**

The set of preliminary genetic analyses indicated that *I. purpurea* sampled localities were

321

322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

340

341

342

343

in no major violation of Hardy-Weinberg equilibrium, as judged by the small difference between expected and observed heterozygosity (mean He = 0.294±0.014 and 0.250±0.001; mean Ho = 0.291±0.009 and 0.260±0.001, respectively for SSR and SNP datasets). Levels of expected and observed heterozygosity for the SSR dataset were only slightly greater than those estimated for the SNP dataset (Table S3). Likewise, the estimated mean effective population size per sampled locality was only slightly greater and more variable for the SSR dataset than for the SNP dataset (13.71±5.59, 9.49±0.13, respectively; Table S3), but in neither case was there salient evidence of a plausible source-sink dynamic, as judged by the similar effective sizes among populations (Table S3; Diffendorfer 1998). Accordingly, spatial structure in this species was limited, as evidenced by the modest  $F_{ST}$  estimates (0.151 and 0.140, respectively for SSR and SNP datasets) and the admixed genetic composition of individuals (rather more structured in the SNP dataset; Fig. 2). Furthermore, although TESS results indicated the existence of partially discernable spatial genetic clusters (4 and 5 clusters for the SSR and SNP datasets, respectively; Fig. 2), variation among the inferred spatial genetic clusters explained less than 10% of the variance (Table 1). It is especially noteworthy that the inferred spatial genetic clusters were constrained to geographically contiguous areas in the SNP dataset, but not in the SSR dataset (Fig. 2). Despite these overall similarities between datasets, estimates of recent ancestry differed between them. The analysis on the SSR dataset indicated that migration among localities is widespread and hardly geographically constrained, with only four localities being primarily constituted of native individuals (Fig. 3a). Across localities, on average 73.65% of individuals were inferred to be 1st or 2nd generation immigrants. On the other hand, the analysis on the SNP

dataset showed that most populations have a limited number of recent immigrants, but that the relatively few inferred immigrants (on average 27.42% of individuals across localities) did not come exclusively from geographically proximate localities (Fig. 3c). Nevertheless, long distance migration events were estimated to be infrequent in this dataset relative to the SSR dataset, in which migration across geographically spread states was common (e.g., between North Carolina and Tennessee, Fig. 3a,c). Furthermore, of the 6 localities shared between datasets only 1 was similarly inferred to comprise mostly native individuals by both sets of markers; inferences for the other five localities differed between datasets. Accordingly, the SSR-based pruned conditional genetic network (Dyer *et al.* 2010) was significantly more interconnected (vertex connectivity = 5; White & Harary 2001) than the SNP-based network (vertex connectivity 0) (Fig. 3b,d). This relatively low complexity of the SNP-based network further confirmed the inference of locally constrained migration in this dataset compared to more widespread migration inferred with the SSR dataset.

In line with these findings, the spatial autocorrelation analyses indicated that genetic clustering extended over wider spatial scales for the SSR dataset than for the SNP dataset (Fig. S1). Specifically, nearby localities showed a significant tendency to be genetically similar to each other up to a considerably greater distance in the case of the SSR dataset relative to the SNP dataset (Fig. 3). Furthermore, at greater distances no significant association was recovered for the SNP dataset, further confirming their genetic isolation from each other, whereas a significant negative association (i.e., statistical tendency to be more different from each other than expected by chance) was recovered in the SSR dataset.

#### Landscape genetics

Unsurprisingly given the distinct geographical and environmental ranges covered by each dataset (Fig. 1), the optimization of landscape resistance layers resulted in alternative optimal solutions for the SSR and SNP datasets (Fig. S2). It is important to note, however, that a formal comparison is not possible as the associations recovered are statistical associations driven by the fit of the resistance parameterization to the data under the statistical model implemented (Martínez-Abraín 2008). While these associations are expected to recapitulate real biological properties of the study system, they are constrained to the data at hand. Nevertheless, association patterns that are robust to the data used are expected to better reflect the actual impact that landscape features have on gene flow, independently of possible biases introduced by expert opinion. Therefore, we focus below on the common biological findings between marker types, while also denoting the most relevant differences. Such differences likely reflect not only the different environmental ranges covered by each dataset, but most importantly, the particular genetic connectivity pattern captured by each one of them (Fig. 3).

In spite of the underlying differences in inferred population connectivity (Fig. 3), some of the resulting landscape resistance layers roughly resemble their counterpart in the other dataset in terms of their relative resistance allocation (Fig. S2). There were some landscape features that showed consistent resistance patterns between datasets. For example, cotton-dominated areas and woody savannas were recovered as highly permeable landscape features, whereas areas dominated with soybean fields, evergreen forests, open shrublands, and grasslands were recovered as highly resistant landscape features. This difference between the permeability of different crop and landscover features matches the prevalence and distribution of these features

across the study area. Soybean dominated areas occupied over 8.7% of the study area and were most prevalent in its northwestern portion, where *I. purpurea*, being primarily a subtropical vine (Fang *et al.* 2013), is less prevalent. On the other hand, cotton-dominated areas occupied 0.8% of the study area and were concentrated in its southern portion, including areas of high *I. purpurea* concentration. Likewise, woody savannas represented 10.8% of the study system and were constrained to the southern portion of it, whereas evergreen forests, open shrublands, and grasslands represent together 1.1% of the study system and are prevalent in areas where *I. purpurea*, being mainly found on agricultural fields, gardens, and roadsides (Defelice 2001), is absent. Hence, these landscape associations most likely reflected the environmental preferences of *I. purpurea*, as environments where this species thrives coincided with environments with a low resistance assigned, whereas environments unoccupied by this species were assigned greater resistances.

Given the differences in sampling between datasets (Fig. 1), the most revealing patterns were similar landscape variables identified as significant (or marginally significant) predictors of genetic differentiation in *I. purpurea* across datasets (Table 2). Both datasets pointed towards human-impacted landscapes playing an important direct role in shaping genetic connectivity in this species. While in both sets of MLPE.lmm models, natural (climate, elevation, and soils) and human-related landscapes (landcover and human population density) were identified as significant or marginally significant predictors of genetic similarity between localities, the variable with the greatest association coefficient and lowest AICc value in these models was in both cases a variable closely linked to human presence (landcover in the SSR dataset, and human population density in the SNP dataset; Table 2). In contrast, the dbRDA analyses showed that

none of these features were consistently recovered as a significant predictor of genetic similarity

413

414

415

416

417

418

419

420

421

422

423

424

425

426

427

428

429

430

431

432

433

434

435

in the SNP dataset, especially after accounting for geographic distance between populations even though geographic distance by itself was not a significant predictor of genetic structure in this species (i.e., no isolation by distance pattern was recovered). Yet, when considering all variables together in a multivariate manner while accounting for geographic distance, human population density was the only variable that remained a significant or marginally significant predictor of population structure across both SSR and SNP datasets (Table 2). Specifically, these results indicated that, after considering other landscape variables, human-population-density resistance was associated with population differentiation across datasets, with high and low populated areas identified as less favorable areas for migration (although conductance optimum for the SNP dataset was notably lower than for the SSR datasets; Fig. 4). Accordingly, the corresponding resistance-based conductance map (Shah & McRae 2008), showed local clusters of high permeability for both datasets (although slightly less constrained in the SSR dataset), surrounded by vast areas of high resistance (Fig. S3). It is important to note, however, that these multivariate regressions explained a small, non-significant proportion of the variance (MRDM R<sup>2</sup> for SSR and SNP dataset were 0.055 (F = 3.776, p-val. = 0.110) and 0.309 (F = 1.275, p-val. = 0.261), respectively). In addition to population density, climate was also recovered as a significant predictor of genetic dissimilarity across analyses on the SSR dataset but not for the SNP dataset (Table 3) most likely because of the lack of northern samples, from colder areas, for the latter dataset (Fig. 1). Specifically, the first component of the climate PCA (see methods above), which primarily summarized temperature variation, was consistently associated with genetic differentiation (Table 3), with intermediates values identified as a barrier to dispersal. Warmer areas were identified as the most conducive to dispersal, followed by colder areas—the latter pattern exclusively driven by connectivity between the northernmost localities (Fig. 4). The corresponding resistance-based conductance map showed a less spatially constrained distribution of permeable areas with connections between southern and between northern localities, and a strong barrier between them (Fig. S3).

# **DISCUSSION**

Taken together, the population structure analyses indicated limited to moderately high (depending on the molecular marker) global levels of genetic differentiation in *I. purpurea* and non-geographically structured migration. This non-geographically structured migration was inferred to be rather rare when using the SNP dataset (Fig. 3). These results suggest that broadly distributed populations of this agricultural weed are generally genetically distinct, but there is some indication of long-distance and putatively human-mediated migration between localities, as suggested by the recovered association between human population density and genetic similarity. Such levels of differentiation and long-distance migration strongly contrast with this species' patchy distribution, which is tightly linked to isolated agricultural patches that are surrounded by a complex matrix of natural and urbanized areas. As suggested by the landscape genetics analyses, this matrix does seem to impact connectivity in this species. Specifically, climate and human population density were robustly recovered as predictors of genetic connectivity in this species. Of these landscape variables, climate has a stronger effect, as judged by its greater MRDM coefficient, but only when considering the SSR dataset, which covers the northern

portion of the range (Table 3). Otherwise, population density was the only variable across datasets with a marginally significant effect—even after accounting for multiple tests. Along with the recovered pattern of landscape features of high and low permeability, in particular in regards to crop types (Fig. S2), this finding highlights the important role that humans play in structuring populations of this species. In addition, the results highlight how inferences about population structure and patterns of connectivity are dataset-dependent, with marked differences becoming apparent only after careful dissection of roughly similar F<sub>ST</sub> and heterozygosity estimates across molecular markers. Below, we detail each of these novel findings and place them in the context of agricultural weed movement across the landscape, invasive species, and landscape genetics practice.

# **Human impact**

Given that *I. pupurea* is a naturalized species in the United States that is found primarily associated with cultivated crops and horticultural gardens (Defelice 2001; Baucom & Mauricio 2004; Fang *et al.* 2013), the finding that human population density is a predictor of genetic similarity in this species is at first glance intuitive. Yet, because habitat requirements for establishment and migration are not always the same, especially for organisms with distinct migration stages (e.g., pollen or seeds in plants) and dormant stages (Murphy & Lovett-Doust 2004), this finding is not as straightforward as it seems. In particular, the fact that human population density is an informative predictor throughout the entire sampled distribution, whereas climate seems to mainly represent a gene flow barrier between southern and northern *I. purpurea* sites (Fig. S1) highlights the influence that humans have on structuring the populations of this

483

484

485

486

487

488

489

490

491

492

493

494

495

496

497

498

499

500

501

502

503

species and helps to discern the factors involved in the spread of this noxious weed. In this sense, the results point towards humans not only as likely responsible for the introduction of this weed into the United States (Fang et al. 2013), but also as responsible for facilitating its current spatial connectivity, and hence its opportunities for thriving in the complex landscapes it inhabits. As evidenced by the inferred landscape conductivity, human population density seems to primarily facilitate connectivity at local to intermediate scales (note the clusters of high conductivity around sampled localities; Figs. 4 and S2), which encompass proximate agricultural fields, suggesting that factors such as sharing of contaminated agricultural machines, trade between nearby farmers, or local distribution of contaminated crop seeds are at play (Dastgheib 1989; Thill & Mallory-Smith 1997; Benvenuti 2007; Boyd & White 2009). At the same time, considering that i) the horticultural trade has been recognized as the main source of invasive introductions and spread in the United States (Lehan et al. 2013), ii) that I. purpurea is an appreciated horticultural species (Fang et al. 2013), and that, given current agricultural practices, crop seed contamination is unlikely to be a major factor (Economic Research Service 1998), it is probable that ornamentals' trade between population centers may help explain both the long distance dispersal events recovered in both datasets (Fig. 3) and the local clustering (Fig. 4). Alternatively, the impact of human populations on the distribution and abundance of bumblebees (Martins et al. 2013; Jha 2015), which are *I. purpurea*'s predominant pollinators (Ennos 1981; Baucom & Mauricio 2008), could also be partially responsible for the connectivity patterns recovered as changes in the pollinators community would have strong effects on gene flow (Jha & Kremen 2013). In reality a combination of all these factors may be involved.

While further analyses are needed to elucidate the ultimate causes behind the recovered association between human population density and genetic dissimilarity in *I. purpurea*, our findings bring much needed information to limit the spread of this noxious weed. Our findings are not only relevant to *I. purpurea* and to the evolution of herbicide resistance in this species (i.e., is herbicide resistance evolving independently across populations or is it being disseminated through human-aided migration?), but also has important implications for other weeds of agricultural concern as well as other human-exploiter species (Blair 2001), such as other invasives. Specifically, in line with previous work (Bataille et al. 2011; Auffret et al. 2014; Banks et al. 2015), the results here point towards the need of better strategies to minimize the impact that humans have on the spread of species. In particular, our results further support that humans may not only facilitate the introduction of invasive species into non-colonized areas, but also contribute to the maintenance of gene flow among naturalized populations (Medley et al. 2015), which may be critical in providing relevant genetic variants for increased fitness as well as prevent inbreeding depression in these newly colonized areas (Kolbe et al. 2004; Edelaar & Bolnick 2012).

# Landscape genetics practice

504

505

506

507

508

509

510

511

512

513

514

515

516

517

518

519

520

521

522

523

524

525

526

Under the common landscape genetics' assumption of an equilibrium between migration and genetic drift (Marko & Hart 2011; Dyer 2015b), our results offer a valuable window into the role that environmental setting plays in structuring genetic diversity. Our analyses take advantage of recent methodological developments i) that surpass the need of arbitrary landscape resistance assignment that make inferences sensitive to subjectivity of expert opinion (Dyer 2015a), ii) that account for the indirect genetic similarity of populations (Dyer & Nason 2004), and iii) that use

528

529

530

531

532

533

534

535

536

537

538

539

540

541

542

543

544

545

546

547

548

549

rigorous statistical inferences (Balkenhol et al. 2009; Peterman et al. 2014). Furthermore, in contrast to the common practice in the field of using one or a few loci that are either uniparentally inherited (although a few exceptions exist; e.g. Perry et al. 2013), which prevents an assessment of common patterns across the genome (Bohonak & Vandergast 2011), our inferences are derived from common findings among two rather different sets of molecular markers. In doing so, we provide not only a better representation of the genome, and hence, less sensitive inferences to i) ascertainment bias (Brandström & Ellegren 2008), ii) molecular markers' idiosyncrasies (Buschiazzo & Gemmell 2006), and iii) coalescent and mutational variance (Nielsen & Slatkin 2013; Steiner et al. 2013), but also the ability to distinguish differences in the underlying population dynamics. Such differences have strikingly important implications. For example, when evaluating plausible approaches to the threat of an invasive species such as I. purpurea, recommendations would be quite different depending on whether gene flow is believed to be relatively widespread (as inferred by the SSR dataset) or whether it is believed to be minimal (as inferred by the SNP dataset). In this example, it is clear that evidence-based conservation would clearly benefit from recognizing the current uncertainty in regards to the exact population connectivity as opposed to automatically relying on a single-marker story. Given recent advances in next generation sequencing, it seems straightforward to focus on landscape genomics instead of few loci. Hence, development of methods for explicitly integrating inferences from multiple genome regions, as it is customary in population genetics, would be of great value.

Even more important, the differences identified between markers offer the opportunity to explore the underlying causes for such differences and hence a more in-depth understanding of

551

552

553

554

555

556

557

558

559

560

561

562

563

564

565

566

567

568

569

570

571

the landscape influences on species' genetic structure. While it is theoretically possible that the differences in inference between the two markers are exclusively driven by the particular geographic sampling of each dataset, the robust differences that we report between the localities and regions common to both datasets renders this possibility unlikely. A second alternative is that the differences are driven by the different mutation rates underlying the two type of markers (Wang 2010, but see Bohonak and Vandergast 2011). This time-differential hypothesis is based on the argument that SSR mutation rates are typically estimated to be in the  $10e^{-3}$ - $10e^{-4}$  range. whereas SNP mutation rates, although harder to estimate given their occurrence in multiple heterogeneous genomic regions (Lercher & Hurst 2002), are typically presumed to be on average much lower. Nonetheless, given mutational rates are still on the scale of thousand of years and up, most genetic variation seen in current populations would precede the temporal window of the majority of landscape genetics studies. It is then the sorting of standing genetic variation, rather than the mutation rate, what would primarily drive genetic dissimilarity between current subpopulations of a species. Such sorting is thus expected to be specific to different genomic regions. That is, because of the genome-wide coverage of SNPs and the independent evolutionary trajectories of SSR and SNP loci, recombination, effective population sizes, and other relevant evolutionary parameters are expected to be highly heterogeneous across loci, which can play an important role in the pace at which each loci segregates, making it unclear which temporal scale would be reflected by each dataset (Ennos 1994; Bohonak & Vandergast 2011). Of all the factors involved, effective population sizes of each locus, which directly affect the rate of genetic drift of each loci (Storz et al. 2001; Piganeau & Eyre-Walker 2009), is probably the most impacted by landscape conditions, and hence it is probably a major determinant of patterns of shared variants

across the landscape. Yet, traditionally estimates of effective population size are rarely incorporated into landscape genetics studies.

572

573

574

575

576

577

578

579

580

581

582

583

584

585

586

587

588

589

590

591

592

593

In reality, both datasets likely encompass a range of temporal scales, and presumably reflect both historical and contemporary processes. In this regard, working with an agricultural weed with a relatively well-known history offers the advantage of better accounting for the historical component of current genetic variation. Specifically, under the time-differential hypothesis (Wang 2010; but see Bohonak & Vandergast 2011), it is plausible that the spatial structure recovered by our SNP dataset is strongly influenced by ancestral subdivision and that the relative lack of structure recovered by our SSR dataset reflect more recent gene flow. However, considering the current understanding of the introduction of *I. purpurea* into the United Sates, this possibility seems unlikely. If this weed was indeed introduced through horticulture from a European bottlenecked population during the European colonization of North America (Fang et al. 2013), ancestral structure would be unlikely. Likewise, if the species expanded from the point of introduction, a nested pattern of genetic similarity driven by serial founder events would be expected (Ramachandran et al. 2005; Slatkin & Excoffier 2012)—not a rather distinct clustering of individual subpopulations as we recovered (Fig. 3). Instead, it seems more likely that the obvious structure recovered in the SNP dataset reflects the isolation driven by the complex agricultural matrix *I. purpurea* inhabits and the role that human intervention has had on its evolutionary trajectory (e.g., by facilitating long distance dispersal events; Fig. 3). Yet, further analyses (see below) are needed to test this hypothesis.

More importantly, the incorporation of both datasets into our analyses highlights a commonly overlooked issue in landscape genetics. That is, any pattern of genetic variation cannot be understood without explicit consideration of species' demographic history as current genetic variation is the result of multiple processes thorough the history of a species (Marko & Hart 2011), and different genomic regions reflect different coalescent histories (Nielsen & Slatkin 2013). Hence, the importance of incorporating coalescent-based simulations that explicitly model the presumed landscape effects on genetic population structure and that take into account its demographic history (Balkenhol & Landguth 2011; Hoban et al. 2012). Advances in this area are already being developed with promising perspectives (He et al. 2013; Alvarado-Serrano & Knowles 2014). In this regard, traditional landscape genetics results can be interpreted as a necessary first step towards a more comprehensive understanding of the interaction between landscapes and species evolutionary trajectories. By offering a working hypothesis of the effect of current landscapes on genetic differentiation, traditional landscape genetics results serve the purpose of identifying relevant hypotheses for further testing (Dyer 2015a) and pave the way for rigorous simulation-based assessments of the role of landscape features in promoting or deterring population differentiation.

# **ACKNOWLEDGEMENTS**

594

595

596

597

598

599

600

601

602

603

604

605

606

607

608

609

610

611

612

613

614

615

616

We thank Adam Kuester for seed collecting and for contributing valuable data for this study. We also thank Ariana Wilson, Eva Fall, and Dan York for tissue collection. This research was funded by USDA NIFA grants 04180 and 07191to R.S.B.

# **DATA ACCESSIBILITY**

All data generated is in the process of being archived in Dryad. The corresponding doi would be made available upon acceptance.

AUTHOR CONTRIBUTIONS

D.F.A.-S. and R.S.B conceived the study. D.F.A.-S. and M.V.E. generated and compiled the molecular and GIS data. D.F.A.-S. analyzed the data. D.F.A.-S., M.V.E., S.M.C., and R.S.B. wrote the manuscript. All authors read and approved the final submission.

627

628

629

630

631

632

633

634

635

636

637

638

639

640

641

642

643

644

645

646

647

LITERATURE CITED Alvarado-Serrano DF, Knowles LL (2014) Ecological niche models in phylogeographic studies: Applications, advances and precautions. *Molecular Ecology Resources*, **14**, 233–248. Auffret AG, Berg J, Cousins SAO (2014) The geography of human-mediated dispersal. *Diversity* and Distributions, **20**, 1450–1456. Auffret AG, Cousins SAO (2013) Humans as Long-Distance Dispersers of Rural Plant Communities. PLoS ONE, 8. Balkenhol N, Landguth EL (2011) Simulation modelling in landscape genetics: on the need to go further. *Molecular ecology*, **20**, 667–70. Balkenhol N, Waits LP, Dezzani RJ (2009) Statistical approaches in landscape genetics: an evaluation of methods for linking landscape and genetic data. *Ecography*, **32**, 818–830. Banks NC, Paini DR, Bayliss KL, Hodda M (2015) The role of global trade and transport network topology in the human-mediated dispersal of alien species. *Ecology Letters*, 18, 188–199. Barrowclough GF (1980) Gene flow, effective population sizes and genetic variance components in birds. Evolution, 34, 789–798. Bataille A, Cunningham AA, Cruz M, Cedeño V, Goodman SJ (2011) Adaptation, isolation by distance and human-mediated transport determine patterns of gene flow among populations of the disease vector Aedes taeniorhynchus in the Galapagos Islands. *Infection, Genetics and* Evolution, 11, 1996–2003. Baucom RS, Mauricio R (2004) Fitness costs and benefits of novel herbicide tolerance in a noxious weed. Proceedings of the National Academy of Sciences, 101, 13386–13390. Baucom RS, Mauricio R (2008) The evolution of novel herbicide tolerance in a noxious weed:

649

650

651

652

653

654

655

656

657

658

659

660

661

662

663

664

665

666

667

668

669

670

The geographic mosaic of selection. Evolutionary Ecology, 22, 85–101. Benjamini Y, Hochberg Y (1995) Controlling the false discovery rate [mdash] a practical and powerful approach to multiple testing. J. R. Stat. Soc. B, 57, 289–300. Benvenuti S (2007) Weed seed movement and dispersal strategies in the agricultural environment. Weed Biology and Management, 7, 141–157. Berthouly-Salazar C, Hui C, Blackburn TM et al. (2013) Long-distance dispersal maximizes evolutionary potential during rapid geographic range expansion. *Molecular Ecology*, 22, 5793-5804. Blair RB (2001) Birds and butterflies along urban gradients in two ecoregions of the US. In: Biotic homogenization (eds Lockwood JL, McKinney ML), pp. 33–56. Bohonak AJ, Vandergast AG (2011) The value of DNA sequence data for studying landscape genetics. *Molecular ecology*, **20**, 2477–9; authors reply 2480–2. Boyd NS, White S (2009) Impact of Wild Blueberry Harvesters on Weed Seed Dispersal within and between Fields. Weed Science, 57, 541–546. Brandström M, Ellegren H (2008) Genome-wide analysis of microsatellite polymorphism in chicken circumventing the ascertainment bias Genome-wide analysis of microsatellite polymorphism in chicken circumventing the ascertainment bias. Genome Research, 881– 887. Buschiazzo E, Gemmell NJ (2006) The rise, fall and renaissance of microsatellites in eukaryotic genomes. *BioEssays*, **28**, 1040–1050. Chaney L, Baucom RS (2012) The evolutionary potential of Baker's weediness traits in the common morning glory, Ipomoea purpurea (Convolvulaceae). American Journal of Botany, **99**, 1524–1530.

672

673

674

675

676

677

678

679

680

681

682

683

684

685

686

687

688

689

690

691

692

693

Chen C, Durand E, Forbes F, François O (2007) Bayesian clustering algorithms ascertaining spatial population structure: A new computer program and a comparison study. *Molecular Ecology Notes*, **7**, 747–756. Cheng L, Connor TR, Sir??n J, Aanensen DM, Corander J (2013) Hierarchical and spatially explicit clustering of DNA sequences with BAPS software. Molecular Biology and Evolution, **30**, 1224–1228. Clobert J, Baguette M, Benton TG, Bullock JM, Ducatez S (2012) Dispersal Ecology and Evolution. Oxford University Press, Oxford. Consortium; MERPD, Aksoy S, Almeida-Val VMF et al. (2013) Permanent Genetic Resources added to Molecular Ecology Resources Database 1 October 2012–30 November 2012. *Molecular Ecology Resources*, **13**, 341–343. Corander J, Waldmann P, Marttinen P, Sillanpää MJ (2004) BAPS 2: enhanced possibilities for the analysis of genetic population structure. *Bioinformatics (Oxford, England)*, **20**, 2363–9. Culley TM, Sbita SJ, Wick A (2007) Population genetic effects of urban habitat fragmentation in the perennial herb Viola pubescens (Violaceae) using ISSR markers. Annals of Botany, 100, 91–100. Culpepper AS (2006) Glyphosate-Induced Weed Shifts. Weed Technology, 20, 277–281. Cushman S a, McKelvey KS, Hayden J, Schwartz MK (2006) Gene flow in complex landscapes: testing multiple hypotheses with causal modeling. The American naturalist, 168, 486–99. Danecek P, Auton A, Abecasis G et al. (2011) The variant call format and VCFtools. Bioinformatics, 27, 2156–2158. Dastgheib F (1989) Relative importance of crop seed, manure and irrigation water as sources of weed infestation. Weed Research, 29, 113–116.

694 Defelice MS (2001) Intriguing World of Weeds Tall Morningglory, Ipomoea purpurea (L.) 695 Roth-Flower or Foe? Weed Technology, 15, 601–606. 696 Diffendorfer JE (1998) Testing Models of Source-Sink Dynamics and Balanced Dispersal. Oikos, 697 **81**, 417–433. 698 Diniz-Filho JAF, Nabout JC, Bini LM et al. (2009) Niche modelling and landscape genetics of 699 Caryocar brasiliense ("Pequi" tree: Caryocaraceae) in Brazilian Cerrado: an integrative 700 approach for evaluating central-peripheral population patterns. Tree Genetics & Genomes, 701 **5**, 617–627. 702 Do C, Waples RS, Peel D et al. (2014) NeEstimator v2: Re-implementation of software for the 703 estimation of contemporary effective population size (Ne) from genetic data. *Molecular* 704 *Ecology Resources*, **14**, 209–214. 705 Durand E, Chen C, François O (2009a) Tess version 2.3 - Reference Manual August 2009 \*. 706 Available at: memberstimc. imag. fr/Olivier. Francois/tess. html, 1–30. 707 Durand E, Jay F, Gaggiotti OE, François O (2009b) Spatial inference of admixture proportions 708 and secondary contact zones. *Molecular biology and evolution*, **26**, 1963–73. 709 Dyer RJ (2009) GeneticStudio: a suite of programs for spatial analysis of genetic-marker data. 710 *Molecular ecology resources*, **9**, 110–3. 711 Dyer RJ (2015a) Is there such a thing as landscape genetics? *Molecular Ecology*, **10**, 139–143. 712 Dyer RJ (2015b) Population Graphs and Landscape Genetics. *Annual Review of Ecology*, 713 Evolution, and Systematics, 46, annurev-ecolsys-112414-054150. 714 Dyer RJ, Nason JD (2004) Population Graphs: the graph theoretic shape of genetic structure. 715 Molecular ecology, 13, 1713–27.

717

718

719

720

721

722

723

724

725

726

727

728

729

730

731

732

733

734

735

736

737

738

Dyer RJ, Nason JD, Garrick RC (2010) Landscape modelling of gene flow: improved power using conditional genetic distance derived from the topology of population networks. *Molecular ecology*, **19**, 3746–59. Economic Research Service U (1998) Seed Industry Structure Is Characterized by Growth and Consolidation. *AIB*, **786**, 25–29. Edelaar P, Bolnick DI (2012) Non-random gene flow: An underappreciated force in evolution and ecology. Trends in Ecology and Evolution, 27, 659–665. Ennos RA (1981) Quantitative studies of the mating system in two sympatric species ofIpomoea(Convolvulaceae). Genetica, 57, 93–98. Ennos R a (1994) Estimating the relative rates of pollen and seed migration among plant populations. *Heredity*, **72**, 250–259. Epps CW, Keyghobadi N (2015) Landscape genetics in a changing world: disentangling historical and contemporary influences and inferring change. *Molecular Ecology*, **24**, 6021– 6040. Everman E, Klawinski P (2013) Human-facilitated jump dispersal of a non-native frog species on Hawai'i Island. Journal of Biogeography, 40, 1961–1970. Excoffier L, Smouse PE, Quattro JM (1992) Analysis of Molecular Variance Inferred From Metric Distances Among DNA Haplotypes: Application., **491**, 479–491. Fang Z, Gonzales AM, Durbin ML et al. (2013) Tracing the geographic origins of weedy Ipomoea purpurea in the Southeastern United States. *Journal of Heredity*, **104**, 666–677. Fountain T, Duvaux L, Horsburgh G, Reinhardt K, Butlin RK (2014) Human-facilitated metapopulation dynamics in an emerging pest species, Cimex lectularius. *Molecular* Ecology, 23, 1071–1084.

740

741

742

743

744

745

746

747

748

749

750

751

752

753

754

755

756

757

758

759

760

761

Glickman ME, Rao SR, Schultz MR (2014) False discovery rate control is a recommended alternative to Bonferroni-type adjustments in health studies. Journal of Clinical *Epidemiology*, **67**, 850–857. Goslee SC, Urban DL (2007) The ecodist Package for Dissimilarity-based Analysis of Ecological Data. Journal Of Statistical Software, 22, 1–19. Guillot G, Rousset F (2013) Dismantling the Mantel tests (L Harmon, Ed.). Methods in Ecology and Evolution, 4, 336–344. Harris SE, Xue AT, Alvarado-serrano D et al. (2016) Urbanization shapes the demographic history of a native rodent (the white-footed mouse, Peromyscus leucopus) in New York City. He Q, Edwards DL, Knowles LL (2013) Integrative testing of how environments from the past to the present shape genetic structure across landscapes. Evolution, 67, 3386–3402. Hoban S, Bertorelle G, Gaggiotti OE (2012) Computer simulations: tools for population and evolutionary genetics. Nature Reviews Genetics. Hodkinson DJ, Thompson K, Journal T, Dec N (2007) Plant Dispersal: The Role of Man., 34, 1484–1496. Holderegger R, Buehler D, Gugerli F, Manel S (2010) Landscape genetics of plants. Trends in Plant Science, 15, 675-683. Jha S (2015) Contemporary human-altered landscapes and oceanic barriers reduce bumble bee gene flow. Molecular Ecology, 24, 993–1006. Jha S, Kremen C (2013) Urban land use limits regional bumble bee gene flow. *Molecular* Ecology, 22, 2483–2495. Kolbe JJ, Glor RE, Rodríguez Schettino L et al. (2004) Genetic variation increases during

763

764

765

766

767

768

769

770

771

772

773

774

775

776

777

778

779

780

781

782

783

784

biological invasion by a Cuban lizard. *Nature*, **431**, 177–81. Kuester A, Chang S-M, Baucom RS (2015) The geographic mosaic of herbicide resistance evolution in the common morning glory, *Ipomoea purpurea*: Evidence for resistance hotspots and low genetic differentiation across the landscape. Evolutionary Applications, n/a-n/a. Legendre P, Anderson MJ (1999) Distance-based redundancy analysis: Testing multispecies responses in multifactorial ecological experiments. Ecological Monographs, 69, 1–24. Legendre P, Lapointe FJ, Casgrain P (1994) Modeling Brain Evolution from Behavior - a Permutational Regression Approach. *Evolution*, **48**, 1487–1499. Lehan NE, Murphy JR, Thorburn LP, Bradley BA (2013) Accidental introductions are an important source of invasive plants in the continental United States. American Journal of Botany, 100, 1287–1293. Lenormand T (2002) Gene flow and the limits to natural selection. Trends in Ecology and Evolution, 17, 183–189. Lercher MJ, Hurst LD (2002) Human SNP variability and mutation rate are higher in regions of high recombination. Trends in Genetics, 18, 337–340. Levin DA, Kerster HW (1974) Gene flow in seed plants. Evolutionary Biology, 7, 139–220. Li H (2011) A statistical framework for SNP calling, mutation discovery, association mapping and population genetical parameter estimation from sequencing data. *Bioinformatics*, 27, 2987-2993. Li H. Handsaker B. Wysoker A et al. (2009) The Sequence Alignment/Map format and SAMtools. Bioinformatics, 25, 2078–2079. Mack RN, Lonsdale WM (2001) Humans as global plant dispersers: Getting more than we

785 bargained for. BioScience, 51, 95. 786 Magwene PM (2001) New Tools for Studying Integration and Modularity. Evolution, 55, 1734— 787 1745. 788 Manel S, Holderegger R (2013) Ten years of landscape genetics. *Trends in Ecology & Evolution*, 789 **28**, 614–621. 790 Margosian ML, Garrett K a., Hutchinson JMS, With K a. (2009) Connectivity of the American 791 Agricultural Landscape: Assessing the National Risk of Crop Pest and Disease Spread. 792 *BioScience*, **59**, 141–151. 793 Marko PB, Hart MW (2011) The complex analytical landscape of gene flow inference. Trends in 794 Ecology and Evolution, 26, 448–456. 795 Martínez-Abraín A (2008) Statistical significance and biological relevance: A call for a more 796 cautious interpretation of results in ecology. Acta Oecologica, 34, 9–11. 797 Martins AC, Goncalves RB, Melo G a R (2013) Changes in wild bee fauna of a grassland in 798 Brazil reveal negative effects associated with growing urbanization during the last 40 years. 799 Zoologia, **30**, 157–176. McRae B (2006) Isolation by resistance. Evolution, 60, 1551–1561. 800 801 McRae BH, Beier P (2007) Circuit theory predicts gene flow in plant and animal populations. 802 Proceedings of the National Academy of Sciences of the United States of America, 104, 803 19885–90. 804 Medley KA, Jenkins DG, Hoffman EA (2015) Human-aided and natural dispersal drive gene 805 flow across the range of an invasive mosquito. Molecular Ecology, 24, 284–295. 806 Meehan TD, Werling BP, Landis D a, Gratton C (2011) Agricultural landscape simplification and 807 insecticide use in the Midwestern United States. Proceedings of the National Academy of

809

810

811

812

813

814

815

816

817

818

819

820

821

822

823

824

825

826

827

828

829

830

Sciences of the United States of America, 108, 11500–11505. Menchari Y, Délye C, Le Corre V (2007) Genetic variation and population structure in blackgrass (Alopecurus myosuroides Huds.), a successful, herbicide-resistant, annual grass weed of winter cereal fields. *Molecular Ecology*, **16**, 3161–3172. Mitchell M (1996) An introduction to genetic algorithms. MIT Press, Cambridge Mass. Murphy HT, Lovett-Doust J (2004) Context and connectivity in plant metapopulations and landscape mosaics: Does the matrix matter? *Oikos*, **105**, 3–14. Nakagawa S (2004) A farewell to Bonferroni: The problems of low statistical power and publication bias. *Behavioral Ecology*, **15**, 1044–1045. Nielsen R, Slatkin M (2013) An introduction to population genetics: theory and applications. Sinauer Associates, Sunderland Mass. Nosil P, Egan SP, Funk DJ (2008) Heterogeneous genomic differentiation between walking-stick ecotypes: "isolation by adaptation" and multiple roles for divergent selection. Evolution, 62, 316-36. Oksanen J, Blanchet FG, Kindt R et al. (2015) vegan: community ecology package. http://CRAN.R-project.org/package=vegan. Peakall R, Smouse PE (2012) GenAlEx 6.5: genetic analysis in Excel. Population genetic software for teaching and research--an update. Bioinformatics (Oxford, England), 28, 2537– 9. Perry GH, Louis EE, Ratan A et al. (2013) Aye-aye population genomic analyses highlight an important center of endemism in northern Madagascar. Proceedings of the National Academy of Sciences of the United States of America, 110, 5823–5828. Peterman WE (2014) ResistanceGA: An R package for the optimization of resistance surfaces

832

833

834

835

836

837

838

839

840

841

842

843

844

845

846

847

848

849

850

851

852

853

using genetic algorithms. bioRxiv. Peterman WE, Connette GM, Semlitsch RD, Eggert LS (2014) Ecological resistance surfaces predict fine-scale genetic differentiation in a terrestrial woodland salamander. *Molecular* Ecology, 23, 2402–2413. Piganeau G, Eyre-Walker A (2009) Evidence for variation in the effective population size of animal mitochondrial DNA. PLoS ONE, 4, 2–9. Pimentel D, Zuniga R, Morrison D (2005) Update on the environmental and economic costs associated with alien-invasive species in the United States. Ecological Economics, 52, 273– 288. R Core Development Team (2016) R: a language and environment for statistical computing, 3.2.4., **0**, [http://www.r-project.org]. Ramachandran S, Deshpande O, Roseman CC et al. (2005) Support from the relationship of genetic and geographic distance in human populations for a serial founder effect originating in Africa. Proceedings of the National Academy of Sciences of the United States of America, **102**, 15942–7. Raufaste N, Rousset F (2001) Are Partial Mantel Tests Adequate? *Evolution*, **55**, 1703–1705. Ricciardi A (2007) Are modern biological invasions an unprecedented form of global change? Conservation Biology, 21, 329–336. Rogers JOYB, Murray DONS, Verhalen LM, Claypool PL (1996) Ivyleaf Morningglory ( Ipomoea hederacea ) Interference with Cotton (Gossypium hirsutum)'., 10, 107–114. Rowe HC. Renaut S. Guggisberg a (2011) RAD in the realm of next-generation sequencing technologies. *Molecular ecology*, **20**, 3499–502. Sexton JP, Hangartner SB, Hoffmann A a (2014) Genetic isolation by environment or distance:

855

856

857

858

859

860

861

862

863

864

865

866

867

868

869

870

871

872

873

874

875

876

which pattern of gene flow is most common? Evolution; international journal of organic evolution, **68**, 1–15. Shah VB, McRae BH (2008) Circuitscape: A Tool for Landscape Ecology. *Proceedings of the* 7th Python in Science Conference, 62–65. Simberloff D (2013) *Invasive species*: what everyone needs to know. Slatkin M (1985) Gene flow in natural populations. Annual Review of Ecology and Systematicsystematics, 16, 393–430. Slatkin M, Excoffier L (2012) Serial founder effects during range expansion: a spatial analog of genetic drift. *Genetics*, **191**, 171–181. Spear SF, Balkenhol N, Fortin MJ, McRae BH, Scribner K (2010) Use of resistance surfaces for landscape genetic studies: Considerations for parameterization and analysis. *Molecular* Ecology, 19, 3576–3591. Steiner CC, Putnam AS, Hoeck PEA, Ryder OA (2013) Conservation genomics of threatened animal species. Annual Review of Animal Biosciences, 1, 261–281. Storz JF, Ramakrishnan U, Alberts SC (2001) Determinants of effective population size for loci with different modes of inheritance. The Journal of heredity, 92, 497–502. Taylor PD, Fahrig L, Henein K, Merriam G (1993) Connectivity Is a Vital Element of Landscape Structure. Nordic Society Oikos, 68, 571–573. Thill DC, Mallory-Smith CA (1997) The Nature and Consequence of Weed Spread in Cropping Systems., 45, 337–342. Tiffin P, Rausher M (1999) Genetic Constraints and Selection Acting on Tolerance to Herbivory in the Common Morning Glory Ipomoea purpurea. The American naturalist, 154, 700–716. Trakhtenbrot A, Nathan R, Perry G, Richardson DM (2005) The importance of long-distance

878

879

880

881

882

883

884

885

886

887

888

889

890

891

892

893

894

895

dispersal in biodiversity conservation. Diversity and Distributions, 11, 173–181. Wang IJ (2010) Recognizing the temporal distinctions between landscape genetics and phylogeography. *Molecular ecology*, **19**, 2605–8. Wang IJ, Bradburd GS (2014) Isolation by Environment. *Molecular Ecology*, n/a–n/a. Wang IJ, Savage WK, Bradley Shaffer H (2009) Landscape genetics and least-cost path analysis reveal unexpected dispersal routes in the California tiger salamander (Ambystoma californiense). *Molecular Ecology*, **18**, 1365–1374. Webster TM, Nichols RL (2012) Changes in the Prevalence of Weed Species in the Major Agronomic Crops of the Southern United States: 1994/1995 to 2008/2009. Weed Science, **60**, 145–157. White DR, Harary F (2001) The cohesiveness of blocks in social networks: node connectivity and conditional density. Sociological Methodology, **31**, 305–359. Wichmann MC, Alexander MJ, Soons MB et al. (2009) Human-mediated dispersal of seeds over long distances. Proceedings. Biological sciences / The Royal Society, 276, 523–532. Wilson GA, Rannala B (2003) Bayesian inference of recent migration rates using multilocus genotypes. *Genetics*, **163**, 1177–1191. Wright S (1949) The genetical structure of populations. *Annals of Eugenics*, **15**, 323–354.

# FIGURES AND TABLES - Landscape connectivity of a noxious weed

 Table 1. Analysis of Molecular Variance (AMOVA) of SSR and SNP data. The contribution of spatial clusters (region), individuals.

Effect	F-	Variance explained		F-value		P-value	
	statistic	SSR	SNP	SSR	SNP	SSR	SNP
Regions	$F_{RT}$	3.94 %	8.51%	0.039	0.085	0.001	0.001
Localities	$F_{SR}$	9.05 %	6.10 %	0.094	0.067	0.001	0.001
(Regions)							
Individuals	$F_{ST}$	0.67 %	24.85 %	0.130	0.146	0.001	0.001
(Localities)							
Individuals	$F_{IS}$	86.33%	60.54 %	0.008	0.291	0.252	0.001
Total	$F_{IT}$	100%	100 %	0.137	0.395	0.001	0.001

Table 2. Summary of landscape genetics models. Model coefficients are reported followed by associated p-value (in parenthesis) and, for MLPE.lmm models, followed by AIC difference and ranking (in square brackets). Because of multiple testing, p-values reported for MLPE.lmm and dbRDA analyses are adjusted using a false recovery rate correction. Significant coefficients after a false-recovery-rate correction are in bold; marginally significant ones are denoted with an asterisk.

Variable	MLPE.lmm		dbRDA		partial-dbRDA		MRDM			
	SSR	SNP	SSR	SNP	SSR	SNP	SSR	SNP		
Geographic distance	-	-	1.234	2.503	-	-	0.006	0.321		
			(0.173)	(0.202)			(0.969)	(0.542)		
Environmental layers										
Elevation	0.244	$0.840^{*}$	1.236	1.369	1.195	1.256	-0.629	-1.778		
	(0.028)	(0.054)	(0.104)	(0.348)	(0.206)	(0.438)	(0.303)	(0.685)		
	+10.742 [3]	+1.423 [4]								
PC1 – climate	0.242	0.967	1.454	1.513	1.402	1.031	1.645	-0.288		
	(0.034)	(0.050)	(0.009)	(0.302)	(0.099)	(0.438)	(0.002)	(0.847)		
	+10.356 [2]	+0.032 [2]								
PC1 – soil	0.208	1.044	1.599	3.847	1.315	2.644	-0.191	1.383		
	(0.031)	(0.050)	(0.009)	(0.194)	(0.146)	(0.340)	(0.651)	(0.670)		
	+11.378 [5]	+0.471 [3]								
Human-impact layers										
Crops	-0.226	0.858*	0.819	2.484	0.836	1.270	-0.240	1.414		
	(0.134)	(0.054)	(0.947)	(0.202)	(0.880)	(0.438)	(0.279)	(0.297)		
	+14.491 [6]	+15.371 [5]								
Landcover	0.582	1.358	1.272*	3.392	1.249	4.423	-0.222	-0.182		
	(<0.001)	(0.006)	(0.054)	(0.194)	(0.201)	(0.318)	(0.407)	(0.816)		
	0.000 [1]	+39.775 [6]								
Population density	0.227	0.912*	1.656	3.808	1.401*	2.873	-0.717*	-2.071*		
	(0.028)	(0.053)	(0.007)	(0.194)	(0.099)	(0.318)	(0.091)	(0.092)		
	+10.821 [4]	-0000 [1]								
911										

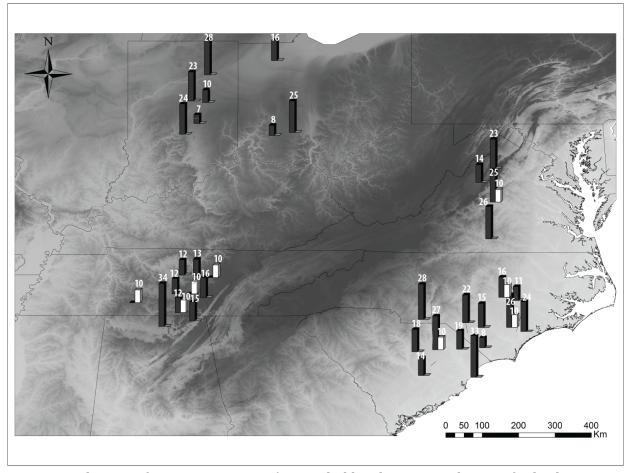


Fig. 1. Distribution of *Ipomoea purpurea*'s sampled localities. Sample sizes for both SSR (black bars) and SNP (white bars) datasets are indicated (numbers on top indicate the actual number of individuals used in our analyses). Elevation is provided as background.

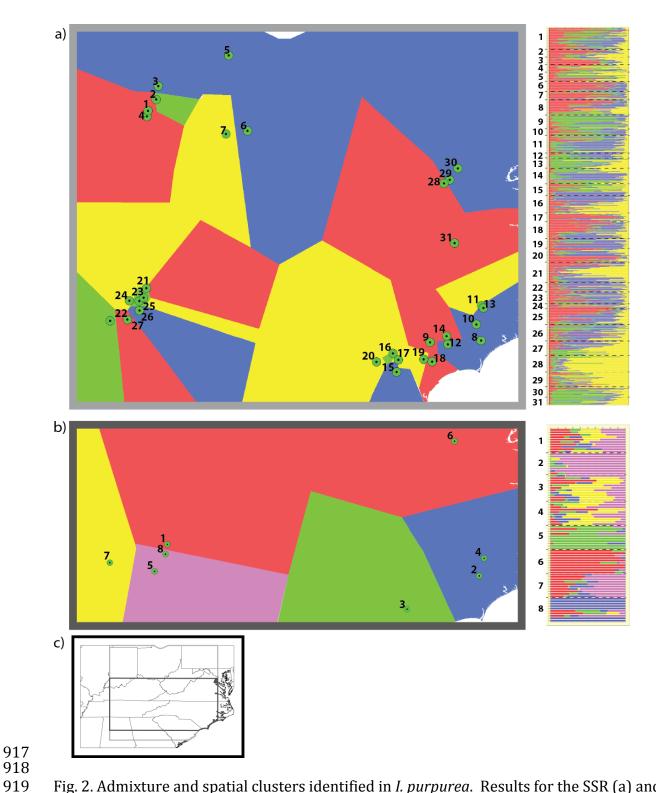


Fig. 2. Admixture and spatial clusters identified in *I. purpurea*. Results for the SSR (a) and SNP (b) datasets are shown together with an inset map (c) denoting the location of those clusters. Individuals in the admixture plot are sorted by sample locality.

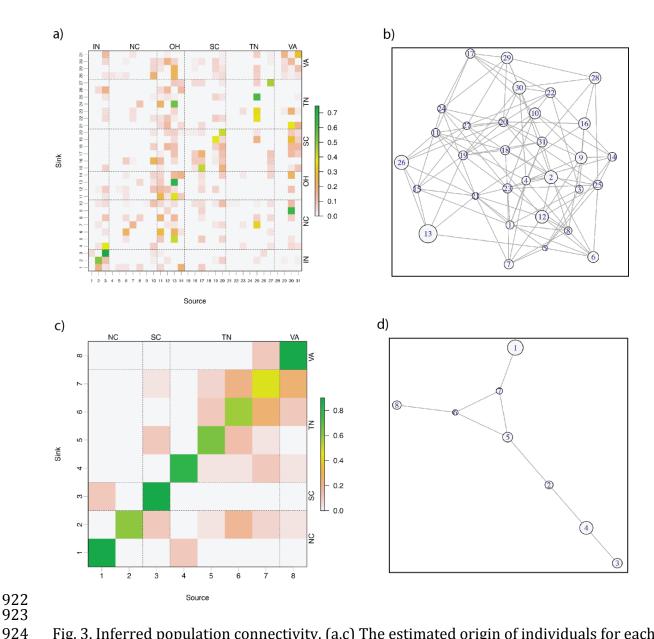


Fig. 3. Inferred population connectivity. (a,c) The estimated origin of individuals for each sampled locality (i.e., sink) is depicted according to the locality they were inferred to have originated from (source). The color of each cell in these plots depicts the proportion of individuals in the sink population that were estimated to be recent immigrants from each locality along the x-axis. Cells on the minor diagonal correspond to the proportion of native individuals. Localities are sorted by State. (b,d) Pruned conditional genetic networks. Note the great difference in complexity and interconnectedness between networks. The top row shows SSR-based results, the bottom, the SNP-based results. Locality numbers follow Fig. 2.

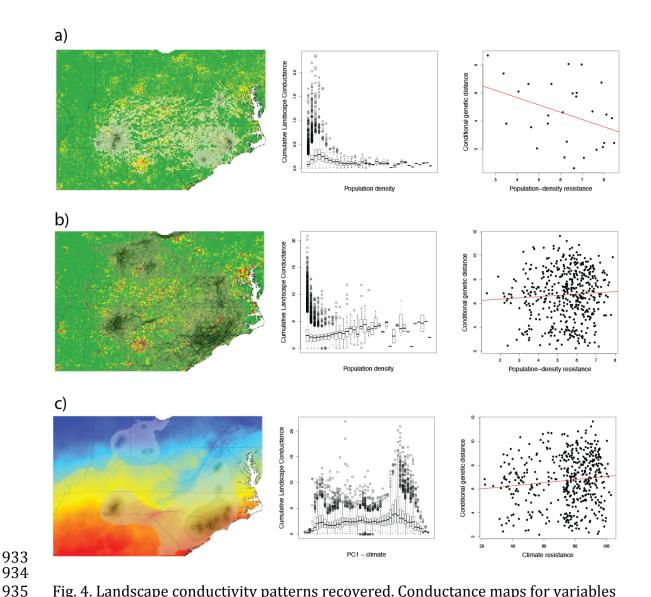


Fig. 4. Landscape conductivity patterns recovered. Conductance maps for variables consistently recovered as significant predictors of genetic dissimilarity are overlaid on maps of those variables to indicate the characteristics of the landscape that facilitate connectivity between sampled localities (left column). Boxplots showing the association between each variable values (binned into categories for simplicity) and conductance (center column), and regression between landscape resistance corresponding to each variable and genetic dissimilarity (right column) are also shown. The 3 associations portrayed are: (a) human population density – SNP differentiation association, (b) human population density – SSR differentiation association, and (c) PC1 climate and SSR differentiation.