Characterization by Next Generation Sequencing Reveals the Molecular Mechanisms Driving the Faster Evolutionary rate of Cassava brown streak virus Compared with Ugandan cassava brown streak virus Titus Alicai¹, Joseph Ndunguru², Peter Sseruwagi², Fred Tairo², Geoffrey Okao-Okuja¹, Resty Nanvubya¹, Lilliane Kiiza¹, Laura Kubatko³, Monica A. Kehoe⁴ and Laura M. Boykin^{5,*} ¹National Crops Resources Research Institute, P.O. Box 7084, Kampala, Uganda ²Mikocheni Agricultural Research Institute, Coca cola Road, Box 6226, Dar es Salaam, Tanzania ³The Ohio State University, 154W 12th Avenue, Columbus, Ohio 43210, USA 4 Crop Protection Branch, Department of Agriculture and Food, Western Australia, Bentley Delivery Centre, Perth, 6983, Western Australia, Australia. 5 The University of Western Australia, ARC Centre of Excellence in Plant Energy Biology and School of Chemistry and Biochemistry, Crawley, Perth 6009, Western Australia, Australia. * Corresponding author: laura.boykin@uwa.edu.au Keywords: cassava, Uganda, Manihot esculenta, smallholder farmer, Cassava brown streak virus, species tree estimation, SVD Quartets, nonsynonomous mutations

Abstract

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Cassava is a major staple food for 800 million people. Cassava brown streak disease (CBSD), is caused by Cassava brown streak virus (CBSV) and Ugandan cassava brown streak virus (UCBSV) is suppressing cassava yields in East Africa at an alarming rate. Previous studies have documented CBSV is more devastating than UCBSV. This is because CBSV is harder to breed resistance for, causes more infections and yield losses in cassava, and its species delimitation is more challenging. We set out to characterize the CBSV and UCBSV whole genomes from the 26 previously published genomes and three new from Uganda, using NGS data with the goal of uncovering genetic patterns that explain the observed biological differences. In this paper, we report phylogenetic relationships, rates of synonymous and non-synonymous substitutions, and whole genome-based evolutionary rates for CBSV and UCBSV. Using the whole genome sequences we produced the first coalescent based species tree estimation for CBSV and UCBSV which supports previously published studies pointing to multiple species of both CBSV and UCBSV. This new species framework led to the finding that CBSV has a faster rate of evolution when compared with UCBSV. The genes responsible for CBSV's rapid rate of evolution are NIa, 6K2, NIb and P1. Furthermore, we have discovered that for CBSV, rates of nonsynonomous substitutions are more predominant than synonymous substitution and occur across the entire genome. All comparative analyses between CBSV and UCBSV presented suggests CBSV is outsmarting the cassava immune system, thus is more devastating and harder to control.

Introduction

Cassava (*Manihot esculenta* Crantz) is a major staple food crop for 800 million people in over 100 tropical and sub-tropical countries ¹. In sub-Saharan Africa, it is the main source of dietary calories for approximately 300 million people ². The tuberous storage roots of cassava are rich in carbohydrates and can be cooked or processed for human food, animal feeds and a wide range of industrial products. The crop is relatively drought tolerant and can yield well even in less fertile soils, hence, its importance to poor families farming marginal lands ³. Cultivation of cassava is most adversely affected by two viral diseases; cassava mosaic disease (CMD) and cassava brown streak disease (CBSD) ⁴, which together were reported to cause production losses of more than US\$1 billion every year ⁵ in Africa.

Serious yield losses due to CMD were first observed on mainland East Africa in the 1920s ⁶. Recorded epidemics of CMD later occurred in the 1930s, 1940s and from 1990s to date ^{7,8}. By contrast, for about 70 years since it was first described ⁹, CBSD was confined to low altitudes (below 1000 meters above sea level) along coastal eastern Africa in Kenya, Tanzania and Mozambique. However, in the early 2000s, outbreaks of CBSD were reported over 1000 km inland at mid-altitude locations (above 1000m) in multiple countries all around Lake Victoria in Uganda ¹⁰, western Kenya ¹¹ and northern Tanzania ⁴. Where it is already established in eastern Africa, the current CBSD epidemic prevails as the main cause of losses in cassava production. Over the last 10 years, the CBSD epidemic has expanded to other countries in East and Central Africa such as Rwanda, Burundi, Congo, DR Congo and South Sudan ¹²⁻¹⁴. This has significantly increased the risk to countries in central and west Africa which are among the world's leading cassava producers, and where CBSD does not occur.

CBSD is caused by *Cassava brown streak virus* (CBSV) and *Ugandan cassava brown streak virus* (UCBSV). Both viruses are (+) ssRNA viruses in the genus *Ipomovirus* and family *Potyviridae* ¹⁵⁻¹⁸, and are often together referred to as cassava brown streak viruses (CBSVs). The CBSVs have genomic organization of 10 segments, total size of approximately 8.9 to 10.8 kb, and coding for a polypeptide with about 2,900 amino acid residues ^{15,17,18}. The complete genome of a CBSD causal virus was first sequenced in 2009 ¹⁸, and to date there are only 26 publicly available ¹⁹. Currently there are two species recognized by the ICTV, but Ndunguru et al. ¹⁹ have suggested further speciation in the UCBSV clade. Both viruses are transmitted in a semi-persistent manner by the whitefly *Bemisia tabaci* ²⁰ and mechanically ²¹. Symptoms of CBSD on cassava vary with cultivar, virus or plant age, but typically include leaf veinal choloris, brown stem lesions, as well as constrictions, fissures and necrosis of the tuberous storage roots ^{22,23}.

Although CBSD has become established in eastern Africa, there is limited knowledge on the diversity of causal viruses, their distribution and evolutionary potential. Therefore, it is necessary to obtain several full genome sequences of CBSD viral isolates, better understand the causal viruses and design long term control approaches for the disease.

In contrast to the growing knowledge on the causal agents of CBSD, host-pathogen interactions are less clear. As such, little is known about specific responses of different cassava varieties to prevailing species or strains of CBSD viral pathogens. Development and dissemination of CBSD-tolerant varieties has been the main means adopted for CBSD control in eastern Africa. With significant efforts geared at breeding for CBSD-resistant varieties, it is of great interest to know if such resistance protects cassava against one or both CBSVs. Such resistance may be expressed as several related features including restricted infection, systemic spread or recovery

of infected plants from disease and the possibility that stem cuttings taken from these may give rise to progeny that are virus-free (reversion). Recent studies have shown CBSV to be the more aggressive virus, infecting both tolerant and susceptible cultivars as single or mixed infections with UCBSV ^{15,24,25}. In contrast, tolerant varieties were infected with only CBSV, but free of UCBSV, suggesting their resistance to the latter. Compared with UCBSV, CBSV isolates have been reported to be more detectable, having higher infection rates by graft inoculation and inducing more severe symptoms ²⁶. It has also been shown that plants of CBSD tolerant or resistant cultivars graft-inoculated with UCBSV developed milder symptoms and a significantly higher proportion of the progenies were virus-free (reverted) compared to those infected with CBSV ²⁷. To date, the underlying reasons for this more aggressive nature of CBSV compared with UCBSV are not known.

In this study, CBSV and UCBSV molecular diversity was investigated by using next generation sequencing to understand new complete genomes of three isolates from Uganda. The sequences obtained were analyzed to determine species composition, CBSV and UCBSV evolutionary rates, role of such changes in virus-host interactions, resulting into cassava cultivar susceptibility or resistance. We set out to answer the following questions:

- 1) How do the three new complete genomes from Uganda compare to those already published ¹⁹?
- 2) Are CBSV and UCBSV distinct species and is there further speciation?
- 3) Why is CBSV more aggressive and harder to breed resistance for than UCBSV?

Results

CBSD Field Symptoms Associated with CBSV and UCBSV Isolates

Categorisation of CBSD foliar symptom distribution on symptomatic plants assessed revealed that the most frequently encountered type was LL - symptoms only on lower leaves (68.4%), followed by SW - systemic and on the whole plant (26.3%), and SL - systemic but localized (5.3%) (table 1). Based on CBSVs detected and CBSD leaf symptom severity scores for 57 sampled plants, whereas the majority of plants infected by UCBSV alone as determined by RT-PCR had mild chlorosis (severity score 2), CBSV infections (single or mixture with UCBSV) tended to have moderate to severe symptoms (scores 3-4) in same proportion to those exhibiting score 2 (fig. 1, table 1). Regarding the three isolates used here for whole genome sequencing, U8 (UCBSV) was from a plant with CBSD score 3 and LL symptom type. Both CBSV isolates (U1 and U4) were from plants with severity scores 2 and 3, symptom types LL and SL, respectively.

Next Generation Sequencing

The three samples from Uganda produced raw reads ranging from 21,844,716 to 23,648,990. After trimming for quality using CLCGW, these numbers were reduced to 21,582,374 to 23,373,606 (table 2). Following *de novo* assembly of the trimmed reads using CLCGW, the numbers of contigs produced were 621-1,008. The contigs of interest from *de novo* assembly were of lengths 2,214 to 8,954nt, with average coverage 24 to 366. After mapping to a reference genome in Geneious, the lengths of the consensus sequences were 8,893 to 9,563 with average coverages of 25 to 393. The final sequences consisted of a consensus between the *de novo* and the mapped consensus with lengths of 8,700 to 8,748.

Genomic Variability and Positive Selection

The CBSV genomes included in this study were more variable when compared with those of UCBSV (supplementary figs. S1 and S2). Characterizing amino acid usage at each position in the

whole genome revealed that CBSV genomes have non-synonymous substitutions present across their entire genome (fig. 2), and predominating when compared to synonymous substitutions. In contrast, UCBSV had near equal non-synonymous and synonymous substitutions across the entire genome. Genes in the UCBSV genomes with non-synonymous substitutions at a higher frequency were; P1, NIb and HAM1 (fig. 2).

CBSV had 68 positively selected sites and 66 negatively selected sites, UCBSV had zero positively selected sites (codons) and 558 negatively selected sites (table 3). Analyzed together there are 3 positively selected sites and 1383 negatively selected sites. The coat protein (CP) of CBSV had the highest number of positively selected sites (16) while 6K2 had zero.

Rates of Evolution

CBSV and UCBSV have different rates of evolution (table 4). We tested two hypothesis using CODEML. The null hypothesis tested was CBSV and UCBSV have equal rates of evolution while the null hypothesis was that CBSV and UCBSV have different rates of evolution (two omegas; model = 2). The Likelihood Ratio Test was used to test for significance if the difference in likelihood was greater than 3.84 (based on the Chi-squared distribution and one degree of freedom) we rejected the null hypothesis that the rates between CBSV and UCBSV are equal.

CBSV whole genome sequences showed it is evolving 5 times faster than UCBSV. The genes contributing to this accelerated rate of evolution for CBSV are NIa (D=29.95), followed by 6K2 (D=6.74), NIb (D=5.18) and P1 (4.61) (table 4 and fig. 4). The transition/transversion ratios were also estimated using CODEML and show the 6K1 (19.6) and CP (13.2) genes have the highest estimates while the remaining 8 genes ranged from 5.05 – 9.93.

Species Tree Estimation - SVDQ

The species phylogeny (fig. 3) shows strong support for a split into two primary viral clades, one consisting of CBSV (fig. 3 clades A and B) and the other consisting of UCBSV (fig. 3 clades E-G), with 100% bootstrap support separating the two clades. Figure 3 shows clades labeled A–G which correspond to; 1) labels A-F from Ndunguru et al ¹⁹, and 2) a new clade G defined in this study. Within the CBSV clade, there are several additional clades with 100% bootstrap support, including the two new CBSV whole genomes from Uganda (U1 and U4). These are the first CBSV whole genomes sequences from Uganda. The other CBSV grouping with 100% bootstrap support labeled B in Figure 3 contains 4 Tanzania samples KoR6, Tan 79, Tan 19 1 and Nal 07. In the UCBSV clade there are 6 nodes supported with a 100% bootstrap, including the new UCBSV whole genome added from this study (U8) which is sister to Kab 07 from Uganda. In addition, the CBSV clade had all samples from a given country grouping together while the UCBSV clade had monophyletic clades from different countries (the multi-colored lines in fig. 3).

Comparison of Gene Trees to Species Tree

Clades A and B, which partition the CBSV isolates into two groups, are consistently present with high support in all genes except HAM1 and CP (table 5). Clades D and G, which each consist of a pair of UCBSV isolates, have high support across all genes, while clades C and E have relatively high support across a majority of genes. Clade F is strongly supported by the CI gene, which is relatively long, but is not found in the phylogenetic tree estimated for any of the other genes.

The whole genome concatenated analysis using MrBayes shows strong support (posterior probability 1.0) for all clades (table 5). However, this analysis does not take into account the

possibility of variation in the evolutionary processes across the individual genes. The SVDQ analysis, on the other hand, uses a coalescent-based method to estimate the overall species tree, and properly accounts for variation in the evolutionary history for each gene. In viewing the bootstrap support values for each of the clades from the SVDQ analysis, we see that the level of support for each clade across the genome is more accurately represented by the corresponding bootstrap proportion. For example, clade F, which was found only in the phylogeny of the CI gene, shows a bootstrap proportion of 0.44 for the SVDQ analysis (as compared to 1.0 for the MrBayes concatenated analysis) (table 5). Similarly, the SVDQ analysis gives a bootstrap proportion of 0.87 for clade E, which showed posterior probabilities below 0.8 for 3 of the 10 genes, as compared to a posterior probability of 1.0 for the concatenated analysis with MrBayes. All other clades are supported with bootstrap values of 1.0, consistent with the MrBayes analysis.

Sliding Window SVD Score

The SVD Score Sliding Window analysis (fig. 4) shows several interesting patterns. First, note that the gene boundaries track well with shifts in the magnitude of the SVD Score, indicating that individual genes are subject to specific evolutionary processes that vary from gene to gene. In particular, several genes show strong support for the primary CBSV/UCBSV split, as indicated by their low scores, while other genes show variation from this basic process, as indicated by increases in the scores. In addition, fig. 4 shows the test statistic associated with the hypothesis test of a shift in the rate of evolution between the two groups, with '*' indicating that the rate difference between the two groups is statistically significant. It is readily apparent from the graph that genes that show strong support (low SVD Score) for the primary CBSV/UCBSV split also show strong evidence for statistically significant differences in evolutionary rate. These

- 224 results support the overall hypothesis that certain genes in CBSV have accelerated rates of
- evolution that contribute to the increased aggressiveness of the virus.

Discussion

In this study we analyzed the molecular mechanisms underlying the field and laboratory observations that CBSV more readily infects cassava plants and tends to display severe symptoms when compared with those infected with UCBSV. Our analyses included characterizing three new complete CBSV (2) and UCBSV (1) genomes, which were combined with the 26 previously published. Our major findings show further speciation of CBSV and UCBSV, a larger genetic landscape for CBSV, including many nonsynonomous sites, and that CBSV has a faster rate of evolution compared with UCBSV (table 4 and fig. 4).

Genes with Accelerated Rates of Evolution in CBSV

We have identified P1, 6K2, NIb and NIa as the genes with accelerated rates of evolution in CBSV. The function of P1 is as an RNA silencing suppressor (RSS), and there is also the suggestion that it may be involved in virion binding to the whitefly stylet via a "bridge" formation by a virus-encoded P1 protein for both CBSV and UCBSV. 6K2 is associated with cellular membrane and is responsible for systemic infection and viral long distance movement ²⁸. The NIb encodes for a nuclear inclusion polymerase and the NIA for a nuclear inclusion protease ^{18,29}.

In Potyviruses generally, when NIa and VPg are associated together they are located in the cytoplasm and nucleus of infected cells. When 6K2-VPg-NIa forms a larger product, the VPg plays a role in viral RNA replication ³⁰. Even though VPg is not one of the genes with a higher evolution rate, both 6K2 and NIa are a part of the complex which affects replication, and this may go some way to explaining their apparent accelerated evolution rate. Is it possible that the

accelerated rates of evolution for genes involved in replication could even be a response to the relatively recent interaction of the viruses and cassava? These viruses are not present in South America where cassava originates so the viruses must be native to Africa. It would appear that the adaptation is still occurring and the cassava immune system does not know how to fight these infections yet. Cassava was introduced to East Africa in the 18th century through oceanic movement. The first reports of brown streak disease in Tanzania occurred 1936 ^{9,13}. There has been little opportunity for the co-evolution of the viruses and the host, therefore a natural resistance would be hard prospect. This raises the possibility of the original host of these viruses, a non-cassava host which may be harboring these viruses or the most recent common ancestor of these viruses. This in turn leads us to wonder just how old these viruses and their ancestors are, and the best way to answer that is to sequence more virus genomes from both cassava and non-cassava hosts wherever they are found.

How Can CBSV Still Function with Such a Large Genetic Landscape?

CBSV and UCBSV have different evolutionary patterns as observed by characterizing the whole genome sequences of CBSV and UCBSV separately. CBSV is genetically more diverse when compared with UCBSV, as evident by the greater amino acid usage (supplementary fig. 1), the faster rates of evolution across the entire genome (table 4), and greater number of nonsynonymous sites across the entire genome (Figure 2). How can CBSV still function with such a large genetic landscape? RNA viruses walk a very fine line of having the genetic arsenal to overcome the host immune system and diverging to a point that key functions of genes are lost ³¹. Recent studies ^{32,33} have shown that viruses with a large genetic landscape adapt to host changes much quicker and can overcome the host immune system faster. Viruses that occupy a large portion of the possible sequence space might be less fit but they outcompete the fitter

strain when the host immune system shifts and hence these viruses have been described as adapted to "survival of the flattest" ^{34,35}. This means that a virus that covers the most sequence space will be able to adapt to host immune system faster than those with smaller spaces. Viruses that are adapted in this category ("survival of the flattest") are going to be harder to breed resistance for because the virus has a larger ability to adapt to changes. It is clear that in our case, CBSV is the virus that has a larger sequence space (Supplemental Figure 1) when compared to that of UCBSV, which is clearly smaller (Supplemental Figure 2). CBSV is one of the RNA viruses that can be described as adapted to "survival of the flattest", while UCBSV is not. Therefore, CBSV is more devastating because it has a larger genetic arsenal which it uses overcome the changes breeders are introducing into cassava.

Not only are the CBSV genomes more genetically diverse, but are also characterized by a large number of nonsynoymous changes in the genome (Figure 2). An excess of nonsynonymous over synonymous substitutions at individual amino acid sites is signifies that positive selection has affected the evolution of a protein between the extant sequences under study and their most recent common ancestor ³⁶. Positive selection is the process by which new advantageous genetic variants sweep a population and is the mechanism Darwin described to drive evolution. This is further evidence that CBSV has a greater capacity to evade the cassava immune system as compared with UCBSV. CBSV had 66 sites under positive selection (Table 4) while UCBSV had none. The CBSV sites under positive selection are found not only in the regions that have gained the most attention, CP and HAM1-like ¹³, but are also found in all other genes except 6K2. This is further support for CBSV's ability to outsmart the cassava immune system. Every gene in the CBSV genome (except 6K2) has sites under positive selection indicating effective RNA silencing of the virus will need to encompass many loci.

Using computational methods combined with field observations we have concluded that CBSV is more devastating than UCBSV. This assertion is also supported by two recent biological studies. The first was a test of reversion in three different cassava varieties (Albert, Kaleso and Kiroba) infected with CBSV and UCBSV. Reversion is a type of resistance mechanism where by virus-infected plants will naturally recover from infection over time, and their progeny from stem cuttings are virus-free. A reversion event infers the host immune system was able to clear or restrict the virus from systemic movement. It was shown that UCBSV infected cassava had a higher rate of reversion when compared to plants infected with CBSV ²⁷ indicating the plants infected with UCBSV recovered more often than those infected with CBSV. This is another line of evidence that CBSV is more devastating and the cassava immune systems of the three varieties tested are struggling to resist the virus.

The second study that supports the hypothesis that CBSV is more aggressive than UCBSV analyzed virus-derived small RNAs within three cassava varieties (NASE 3, TME204 and 60444). Plants infected with viruses are known to trigger RNAi antiviral defense that can be measured by quantifying the abundance of 21-24 nucleotide (nt) segments produced by the dicer enzyme ³⁷. Cassava varieties were infected with either CBSV or UCBSV, NGS was used to detect virus-derived small RNAs ²⁴, and the 21-24 nt dicer fragments were mapped to either CBSV or UCBSV depending on which virus was used to infect the plant. The results showed that CBSV infection triggered a stronger immune response as measured by greater abundance of virus derived small RNA fragments across the entire CBSV genome compared with UCBSV. In addition, across all three genotypes they observed that cassava grafted with CBSV-infected buds showed more severe symptoms compared to UCBSV-infected plants ²⁴. This is further evidence that CBSV is a

more aggressive virus and breeding for resistance to CBSV and UCBSV will require different experimental approaches.

Implications of the Species Tree for CBSV and UCBSV

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We have produced the first species tree estimation of the CBSD causal virus species using whole genome sequences and the coalescent based SVD Quartets species tree estimation algorithm. Differences in the evolutionary history of the two viruses are seen in the branching patterns in Figure 3. CBSV has diverged into two main clades A and B, while UCBSV has several well supported clades but the backbone is still unresolved, indicating more sampling is needed to fully understand the diversity and evolutionary history of UCBSV. The species tree (Figure 3) is similar to the concatenated whole gene tree reported in Ndunguru et al. ¹⁹, except addition of the clade labeled "G", and lack of support for clades E and F in the UCBSV species. It is well documented that concatenating genes without using the coalescent based models can produce misleading results ^{38,39}. In our case, only CI supports clade F, and it is also the longest gene (1,883 bp), therefore swamps signals of other genes. The whole genome concatenation recovers clade F with a posterior probability of 1.00 (Table 4). With regards to clade E, the SVDQ tree was more reflective of the individual gene tree signal by producing a bootstrap value of 0.87 versus 1.00 for the whole genome concatenated tree (Table 4). These results suggest that the topology in the UCBSV species will change as more samples are added.

Our integrative approach of species tree estimation coupled with analyzing rates of evolution has lead to a new framework for CBSV and UCBSV, which includes analyzing and treating these two groups of viruses as separate species. Multiple putative species of both CBSV and UCBV have been identified which means cassava needs to be resistant to the virus species that are

prevalent if farmers' fields. We argue that this genomic diversity and faster rate is what is causing the breeders to struggle with breeding resistant varieties and also why the diagnostic primers are not working consistently. CBSV also has more positively selected sites than UCBSV. It was first thought that CBSD was restricted to the coastal areas and below 1000 m ²³ but as more genetic data is gathered CBSV and UCBSV are found at all elevations in many ecozones throughout East Africa ^{4,10,13,15,19,40}. We are still in the discovery phase with CBSV and UCBSV species as there are only 29 (now with the three new included here) whole genome sequences and other new species of both viruses are likely to be discovered. As we move forward it is important to include all known samples and use appropriate species tree estimation methods such as SVDQ.

Finally, the traditional gene regions (CP and HAM1-like) used to delimit species and are the targets for diagnostic primers do not recover the species tree (Table 4). We recommend designing new diagnostic regions for other genes that recover the species tree and also do not have an accelerated rate of molecular evolution (Figure 4), such as CI or P3 for species level diagnoses. It is possible that the spread of CBSV and UCBSV could have been exacerbated through dissemination of infected cuttings, as virus indexing with primers targeting CP may have misleadingly returned negative results.

Implications of the Results for Cassava Breeding

During the last three decades worldwide, agricultural production has been compromised by a series of epidemics caused by new variants of classic viruses that show new pathogenic and epidemiological properties. An important determinant of the fitness of a virus in a given host is its ability to overcome the defenses of the host. Overcoming plant resistance by changes in the

pathogenicity of viral populations represents a specific and important case of emergence, with tremendous economic consequences since it jeopardizes the success and durability of resistance factors in crops as an anti-viral control strategy. In this study, we found CBSV to be more variable, have more positively selected sites and evolving five times faster than UCBSV. These findings have huge implications for cassava improvement efforts in Africa where CBSV is widely present. Field and laboratory results have proven CBSV to be more virulent and more devastating than UCBSV. Knowledge of specific virus species an improved cassava variety is resistant to will determine where to screen, multiply and deploy such varieties. Cassava breeders have to take into consideration the evolutionary and biological differences between CBSV and UCBSV in the breeding programs. For example, cassava breeders can breed varieties that are resistant to CBSV that can be strategically deployed in areas where CBSV is more prevalent, and similarly for UCBSV. Furthermore, it becomes more appropriate to always screen cassava materials against CBSV as a minimum, even if UCBSV is the more prevalent virus. Such strategy will in effect ensure durable resistance as opposed to the indiscriminate screening and distribution of the improved CBSD resistant cassava varieties, without knowledge of the virus species in the area.

Methods

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Field Plant Sample Collection

Farmers' fields in Uganda with cassava plants 3-6 months old were surveyed for CBSD in 20 districts. In each field, cassava plants were visually assessed to confirm typical CBSD symptoms on leaves and stems. CBSD leaf symptom severity was scored on a 1-5 scale 41,42 ; 1 = no visible symptoms, 2 = mild vein yellowing or chlorotic blotches on some leaves, 3 = pronounced/extensive vein yellowing or chlorotic blotches on leaves, but no lesions or streaks

on stems, 4 = pronounced/extensive vein yellowing or chlorotic blotches on leaves and mild lesions or streaks on stems, 5 = pronounced/extensive vein yellowing or chlorotic blotches on leaves and severe lesions or streaks on stems, defoliation and dieback. CBSD symptoms were also categorized based on distribution of leaf chlorosis and stem lesions on the plant; systemic and on the whole plant (SW), systemic on leaf or stem parts but localized (SL), only on lower leaves (LL). On selected symptomatic plants, portions of the third fully expanded leaf on a shoot were picked as samples, air-dried by pressing between sheets of newsprint and stored pending RNA extraction.

RNA Extraction

About 0.25 g cassava leaf samples were frozen in liquid nitrogen, then ground using a mortar and pestle. 2 ml CTAB lysis buffer (2% CTAB; 100 mM Tris–HCl, pH 8.0; 20 mM EDTA; 1.4 M 134 NaCl; 1% sodium sulphite; 2% PVP) was added and samples homogenized. The 1 ml of the homogenate was incubated at 65° C for 15 min, an equal volume of chloroform: isoamyl alcohol (24:1) was added, and the sample was centrifuged for 10 min at approximately 14,500 rpm. 800µl of the aqueous layer was transferred to a new tube with an equal volume of 4 M LiCl and incubated at -20°C for 2 hrs. The samples were centrifuged for 25 min at 14,500 rpm and the supernatant was poured off. The pelleted RNA was re-suspended in 200 µl TE buffer containing 1% SDS, 100 µl of 5M NaCl. 300 µl of ice-cold isopropanol were added and incubated at -20°C for 30 min. The sample was centrifuged at 13,000 rpm for 10 min and the aqueous layer was decanted and RNA pellets washed in 500 µl of 70% ethanol by centrifuging at 13,000 rpm for 5 min. The ethanol was decanted off and RNA pellet dried to remove residual ethanol. The RNA was re-suspended in 50 µl nuclease-free water and stored at -80 °C prior to testing.

CBSV and UCBSV Detection by RT-PCR

All samples were tested for presence of CBSV and UCBSV by a two-step RT-PCR assay 43 . The PCR mixture consisted of 16.0 μ l nuclease free water, 2.5 μ l PCR buffer, 2.5 μ l MgCl₂ (2.5 mM), 0.5 μ l dNTPs (10 mM), 1.0 μ l of each primer (10mM) [forward CBSDDF2 5'-GCTMGAAATGCYGGRTAYACAA-3' and reverse CBSDDR 5'-GGATATGGAGAAAGRKCTCC-3'], 0.5 μ l Taq DNA polymerase and 1.0 μ l of cDNA. The PCR thermo profile consisted of: 94°C for 2 min followed by 35cycles of 94°C (30 s), 51°C (30 s) and 72°C (30 s) for denaturation, annealing and extension, respectively. PCR products were analysed by electrophoresis in a x1 TAE buffer on a 1.2% agarose gel, stained with ethidium bromide, visualized under UV light and photographed using a digital camera.

Sample Selection for Sequencing

From the data obtained in the diagnostic tests, samples for sequencing were selected to represent different geographical regions, symptom types and severities. Three samples that tested positive for either CBSV (2) or UCBSV (1) were selected for this study. The two samples for which presence of CBSV was confirmed (U1 and U4) had been collected from different farmer's fields in Mukono district, central Uganda. The sample with UCBSV (U8) selected for further analysis originated was collected from a field in Mayuge district, eastern Uganda.

Generation of the Transcriptomes

The three samples were transported to the laboratory and extracted as detailed above. Total RNA was blotted on to FTA cards and later extracted using methods previously described ⁴⁴. Total RNA from each sample was sent to the Australian Genome Research Facility (AGRF) for

library preparation and barcoding before 100 bp paired-end sequencing on an Illumina HiSeq2000.

De novo Sequence Assembly and Mapping

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For each sample, reads were first trimmed using CLC Genomics Workbench 6.5 (CLCGW) with the quality scores limit set to 0.01, maximum number of ambiguities to two and removing any reads with <30 nucleotides (nt). Contigs were assembled using the de novo assembly function of CLCGW with automatic word size, automatic bubble size, minimum contig length 500, mismatch cost two, insertion cost three, deletion cost three, length fraction 0.5 and similarity fraction 0.9. Contigs were sorted by length and the longest subjected to a BLAST search (blastn and blastx) ⁴⁵. In addition, reads were also imported into Geneious 6.1.6 ⁴⁶ and provided with reference sequences obtained from Genbank (KR108828 for CBSV and KR108836 for UCBSV). Mapping was performed with minimum overlap 10%, minimum overlap identity 80%, allow gaps 10% and fine tuning set to iterate up to 10 times. A consensus between the contig of interest from CLCGW and the consensus from mapping in Geneious was created in Geneious by alignment with MAFFT ⁴⁷. Open reading frames (ORFs) were predicted and annotations made using Geneious. Finalized sequences were designated as "complete" based on comparison with the reference sequences used in the mapping process, and "coding complete" if some of the 5' or 3' UTR was missing but the coding region was intact 48,49, and entered into the European Nucleotide Archive (WEBIN ID number Hx2000053576).

Genome Alignment and Annotation

Twenty-six whole genomes (12 CBSV and 14 UCBSV) were downloaded from GenBank and imported into Geneious ⁴⁶, and the MAFFT plugin ⁴⁷was used to align them with the 3 new

whole genome sequences obtained in this study. Nucleotide alignments were translated into protein using the translate align option in Geneious and then visually inspected for quality. Annotations were transferred to the 3 new genomes from the 26 previously published genomes using the live annotation option in Geneious.

Characterizing the Genetic Diversity in CBSV and UCBSV Genomes

CBSV and UCBSV are distinct species (Figure 2) therefore the genomes were treated separately in the analyses in characterizing the genomes. Characterizing the genetic diversity of CBSV and UCBSV was done using the Synonymous Non-synonymous Analysis Program (SNAP v2.1.1) implemented in the Los Alamos National Laboratory HIV-sequence (http://www.hiv.lanl.gov) 50. SNAP calculates synonymous and non-synonymous substitution rates based on a set of codon-aligned nucleotide sequences. This program is based on the simple methods for estimating the numbers of synonymous and nonsynonymous nucleotide substitutions of ⁵¹, and incorporating a statistic developed for computing variances and covariances of dS's and dN's 52. An application of the SNAP package in HIV-1 research has also been developed ⁵³.

Estimating Rates of Evolution

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To further characterize the CBSV and UCBSV genomes, we estimated the rates of molecular evolution using CODEML implemented in PAML (Phylogenetic Analysis by Maximum Likelihood) ⁵⁴. PAML is a package of programs for analysis of DNA or protein sequences by using maximum likelihood methods in a phylogenetic framework. The null hypothesis tested was CBSV and UCBSV have equal rates of evolution (one omega; model = 0) while the alternative hypothesis was that CBSV and UCBSV have different rates of evolution (two omegas; model = 2). The

Likelihood Ratio Test was used to test for significance if the difference in likelihood was greater than 3.84 (based on the Chi-squared distribution and one degree of freedom) we then rejected the null hypothesis that the rates between CBSV and UCBSV are equal. Initial analyses were carried out for the entire genome and showed CBSV has a higher rate of evolution (Table 4; Figure 4). To identify which gene or genes were contributing to the faster rate of evolution we analyzed the individual genes separately testing the hypotheses and parameters utilized for the complete genome.

Testing for Positive Selection

Sites under positive selection were identified using SLAC ⁵⁵ implemented on the http://www.datamonkey.org web server ⁵⁶. The settings used to run SLAC were as follows: the best fitting model (GTR) was specified global dN/dS value was estimated and the significance level was set to 0.01.

Gene Tree Estimation

Individual gene trees were estimated using MrBayes 3.2.1 ⁵⁷ run in parallel on Magnus (Pawsey Supercomputing Centre, Perth, Western Australia) utilizing the BEAGLE library ⁵⁸. MrBayes 3.2.1 was run utilizing 4 chains for 30 million generations and trees were sampled every 1000 generations. All runs reached a plateau in likelihood score, which was indicated by the standard deviation of split frequencies (0.0015), and the potential scale reduction factor (PSRF) was close to one, indicating the MCMC chains converged.

Species Tree Estimation

The SVDQ method ⁵⁹ implemented in PAUP* ⁶⁰ was used to analyze the whole-genome data. This method allows analysis of multi-locus data in a coalescent framework that allows for variation in the phylogenetic histories of individual genes. The method was run with all possible quartets (23,751) sampled in each of 100 bootstrap replicates, and the consensus across all bootstrap replicates was used as the estimate of the species tree. Bootstrap support values for each node were used to quantify uncertainty in the species tree estimate. The entire analysis took approximately 2.5 minutes on a MacBook Pro running OSX 10.11.2 with a 2.2 GHz Intel Core i7 processor.

Comparison of Gene Trees to Species Tree

We compared the single-gene phylogenies constructed using MrBayes with the overall species tree phylogeny estimated using SVDQ and the concatenated phylogeny estimated by MrBayes. For each tree, we evaluated presence or absence of the clades identified by Ndunguru et al.¹⁹ labeled A-F in Figure 3. We identified an additional clade (clade G, Figure 3) that we noticed to be consistently present across genes and methods. For each of these clades present in a particular tree, we recorded the posterior probability (for trees constructed by MrBayes) or the bootstrap proportion (for the tree estimated by SVDQ) in Table 5.

Sliding Window SVD Score

The SVD Score ⁶¹ was used to quantify support for two viral clades for portions of the genome in a sliding window analysis. Briefly, the SVD Score measures the extent to which the data support a phylogenetic "split" – a division of the taxa into two groups with specified group membership. Low values of the SVD Score indicate strong support for the split of interest, while larger values indicate *either* a lack of support for the split or a shift in the underlying evolutionary process

(see Allman et al. (2016) for details and examples). We computed the SVD Score with the split defined by CBSV vs. UCBSV across the genome in windows of 500 bp, sliding in increments of 100 bp, and plotted the resulting SVD Scores across the genome, with boundaries between genes marked with vertical lines. The computations took less than one minute on a MacBook Pro running OSX 10.11.2 with a 2.2 GHz Intel Core i7 processor.

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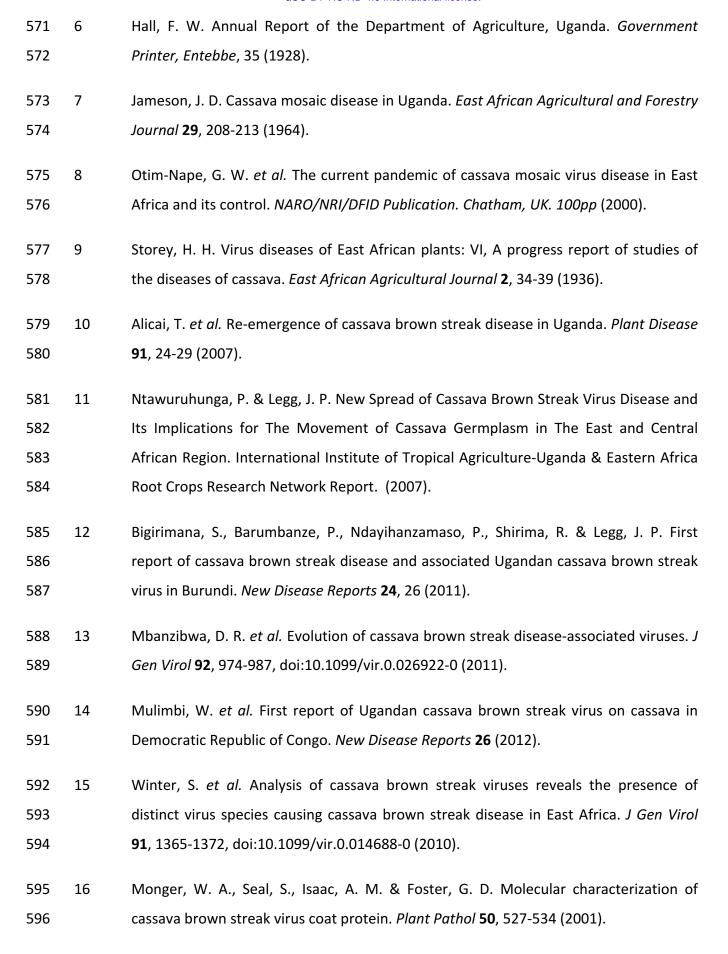
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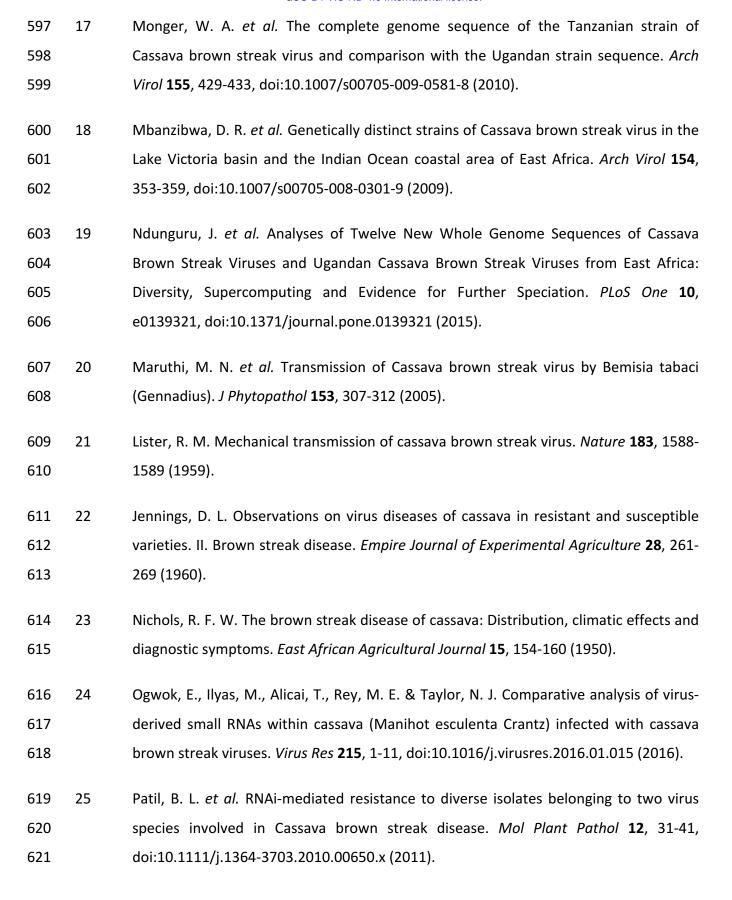
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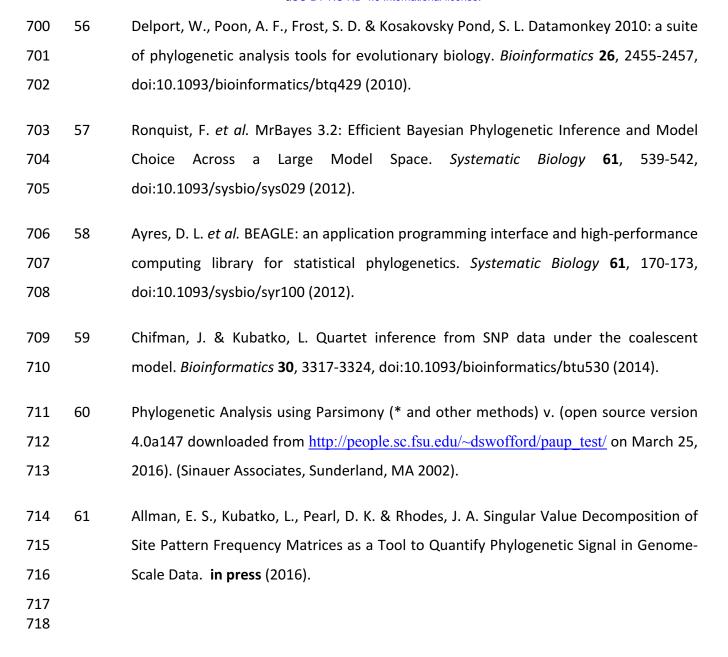


Table 1: CBSD leaf symptom severities and types on plants infected by *Cassava brown streak* virus and *Ugandan cassava brown streak* virus

Virus species		er of plar symptom			Number of plants with CBSD foliar symptom type ²		
	2	3	4	5	SW	LL	SL
CBSV	5	7	1	0	3	8	2
UCBSV	24	5	0	0	7	21	1
CBSV + UCBSV	9	6	0	0	5	10	0
Total	38	18	1	0	15	39	3
Percentage	66.7	31.6	1.7	0.0	26.3	68.4	5.3

¹Foliar CBSD symptom severity score based on 1-5 scale; 1 = no visible symptoms, 2 = mild vein yellowing or chlorotic blotches on some leaves, 3 = pronounced/extensive vein yellowing or chlorotic blotches on leaves, but no lesions or streaks on stems, 4 = pronounced/extensive vein yellowing or chlorotic blotches on leaves and mild lesions or streaks on stems, 5 = pronounced/extensive vein yellowing or chlorotic blotches on leaves and severe lesions or streaks on stems, defoliation and dieback.

²Types of foliar CBSD symptoms based on distribution of leaf chlorosis and stem lesions on the plant; systemic and on the whole plant (SW), systemic on leaf or stem parts but localized (SL), only on lower leaves (LL).

Table 2. Next generation sequencing data for samples from cassava brown streak disease symptomatic plants collected in Uganda

Sample ID	Accession number	Virus	No. of reads obtained	No. of reads after trimming	Number of contigs produced (CLC)	Contig length (CLCGW, nt)	Average coverage (CLCGW)	Number of reads mapped to contig of interest	Ref seq. used for mapping	Length of consensus sequence from mapping (Geneious)	No. reads mapped to ref. sequence	Average coverage (Geneious)	Final sequence length (Coding region only)
U1		CBSV	23,335,344	23,053,082	726	3919, 2214	31, 24	1264, 549	KR108828	8,893	2,233	25	8,748
U4		CBSV	21,844,716	21,582,374	621	8,949	255	23,658	KR108828	8,949	22,987	256	8,748
U8		UCBSV	23,648,990	23,373,606	1,008	8,954	366	33,778	KR108836	9,563	178,117	393	8,700

Table 3. Cassava brown streak virus (CBSV) amino acid (AA) sites under positive selection (analyses method: SLAC Hy-Phy). There were no sites under positive selection for Ugandan cassava brown streak virus (UCBSV).

Gene	CBSV AA site under positive selection
P1	44, 46, 50, 174, 224, 230, 250, 283, 288, 349, 358
P3	415, 455, 467, 472, 499, 525, 618
6K1	658, 678
CI	735, 761, 820, 827, 848, 852, 894, 935, 1218
VPg	1465
NIa	1620, 1645, 1704, 1754, 1785
Nib	1879, 1880, 1890, 1907, 1929, 2109, 2145, 2156, 2161, 2285
HAM1	2320, 2345, 2404, 2432, 2453, 2475, 2519
СР	2550, 2555, 2588, 2611, 2631, 2635, 2640, 2659, 2728, 2745, 2783, 2818, 2843, 2860, 2877, 2884

Table 4. Rates of evolution tested using CODEML implemented in PAML. H_0 was CBSV and UCBSV have equal rates of evolution (one omega; model = 0), while H_1 was that CBSV and UCBSV have different rates of evolution (two omegas; model = 2).

Gene	Assumptions	K (ts/tv rate ratio)	w (omega Dn/Ds) 0	w (omega Dn/Ds)1	Likelihood Ratio Test (if greater than 3.84 reject H ₀) H ₀ =equal rates
WGS	UCBSV and CBSV equal rates	5.90944	0.06358		
WGS	UCBSV and CBSV different rates	5.9598	0.05518	0.07622	26.29*
P1	UCBSV and CBSV equal rates	5.07336	0.10394		
PI	UCBSV and CBSV different rates	5.05456	0.09047	0.12203	4.61*
Р3	UCBSV and CBSV equal rates	5.17197	0.08635		
P3	UCBSV and CBSV different rates	5.20559	0.0764	0.10198	2.22
6K1	UCBSV and CBSV equal rates	19.54143	0.00969		
PKI	UCBSV and CBSV different rates	19.69676	0.01527	0.00316	3.01
CI	UCBSV and CBSV equal rates	9.9388	0.01722		
Ci	UCBSV and CBSV different rates	8.06276	0.0155	0.01977	0.73
6K2	UCBSV and CBSV equal rates	8.40649	0.04684		
ONZ	UCBSV and CBSV different rates	8.82738	0.02354	0.11057	6.74*
VPg	UCBSV and CBSV equal rates	5.852	0.05759		
VPg	UCBSV and CBSV different rates	5.87009	0.054	0.06323	0.29
Nla	UCBSV and CBSV equal rates	8.01105	0.02932		
INIA	UCBSV and CBSV different rates	8.68283	0.01408	0.06719	29.95*
NIb	UCBSV and CBSV equal rates	6.07872	0.05329		
NID	UCBSV and CBSV different rates	6.14047	0.0452	0.06508	5.18*
HAM1	UCBSV and CBSV equal rates	7.25177	0.16144		
ПАІЛІ	UCBSV and CBSV different rates	7.2343	0.17929	0.14007	2.23
СР	UCBSV and CBSV equal rates	13.09297	0.06075		
CF	UCBSV and CBSV different rates	13.26752	0.05606	0.07155	1.29
			faster rate UCBSV	faster rate CBSV	* Rates are different

Table 5. Support for Clades A – G (Figure 3) in individual gene trees and whole genome analyses. Table entries represent posterior probabilities from analysis with MrBayes, except values reported for SVDQ, which are bootstrap proportions. Support values below 95% are indicated in bold, and '--' indicates that the clade was not present.

Genomic region	Clade A	Clade B	Clade C	Clade D	Clade E	Clade F	Clade G
P1	0.9998	0.9998	0.9698	0.9972	0.9954		0.9998
Р3	0.9998	0.9998	0.9857	0.9998	0.9998		0.9998
6K1	0.9466	0.9985	0.9887	0.9995			0.9842
CI	0.9998	0.9996	0.9976	0.9998	0.9998	0.9999	0.9999
6K2	0.9765	0.9998	0.7152	0.9170	0.6646		0.9875
Vpg	0.9998	0.9999		0.9918	0.6455		0.9999
Nla	0.9996	0.9999	0.9998	0.9941	0.7631		0.8934
NIb	0.9999	0.9998	0.9998	0.9999	0.9998		0.9998
HAM1-like				0.9989	0.9983		0.9855
СР				0.9959	0.9950		0.9989
Whole genome	1.00	1.00	1.00	1.00	1.00	1.00	1.00
SVDQ	1.00	1.00	1.00	1.00	0.87	0.44	1.00

Figure 1: Cassava brown streak disease symptoms on leaves and stems of sampled plants; (a) Chlorosis along secondary and tertiary leaf veins of CBSV-infected plant of cultivar TME 204 (severity score 3), (b) Cultivar TME 14 plant with dual CBSV+UCBSV infection showing chlorosis on secondary or tertiary veins, reverse chlorosis (general chlorosis and green area along veins) (severity score 3), (c) UCBSV-infected plant of cultivar TME 204 exhibiting chlorosis on secondary veins, reverse chlorosis, chlorotic spots and mild stem lesions (severity score 3), (d) Very severely diseased plant (severity score 5) of cultivar TME 14 infected with both CBSV and UCBSV, and having chlorosis on leaves, severe stem lesions/brown streaks, defoliation, stem dieback.



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Avg. Behavior of Each Codon 1.5 СР CI NIb P1 Р3 Vpg Nla HAM1 0.1 **UCBSV UCBSV** 0.5 500 1000 2000 3000 1500 2500 Non-synonymous substitutions **Synonymous substitutions** Avg. Behavior of Each Codon 0. CBSV **CBSV** 0.5 1000 500 2000 2500 1500 3000 Genome Position (AA)

Figure 3. Species tree generated from SVD Quartets using the whole genome sequences. Colors at the tips are based on country of origin. Branches with mixed colors indicate a clade that contains samples with mixed country of origin. For example, the ancestral branch of UCBSV TZ Tan 23 KR108839 and UCBSV UG MI B3 FJ039520 is colored red and orange to indicate a clade with sampled with mixed country of origin.

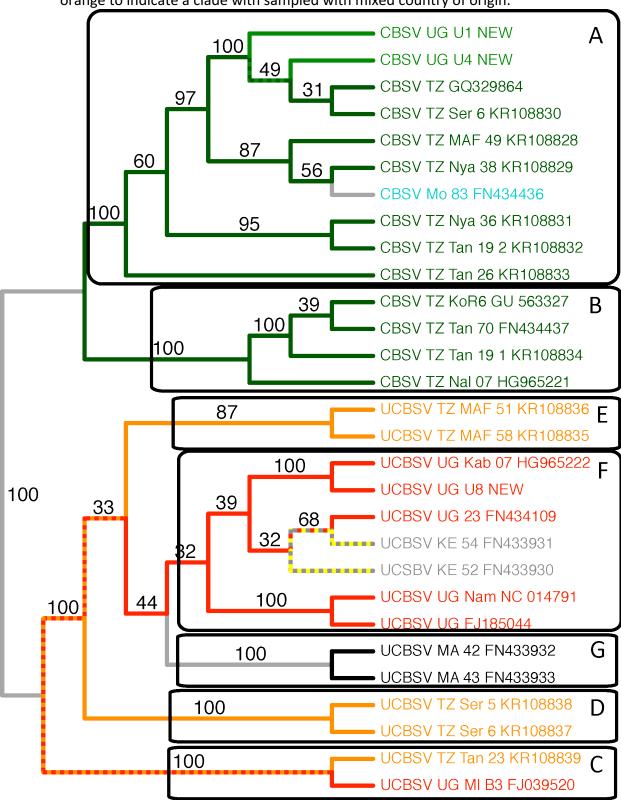
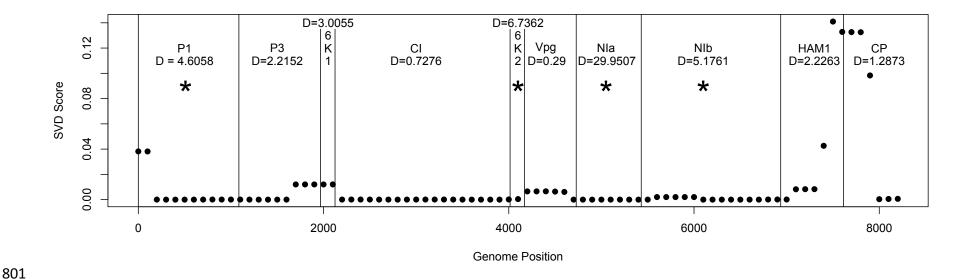
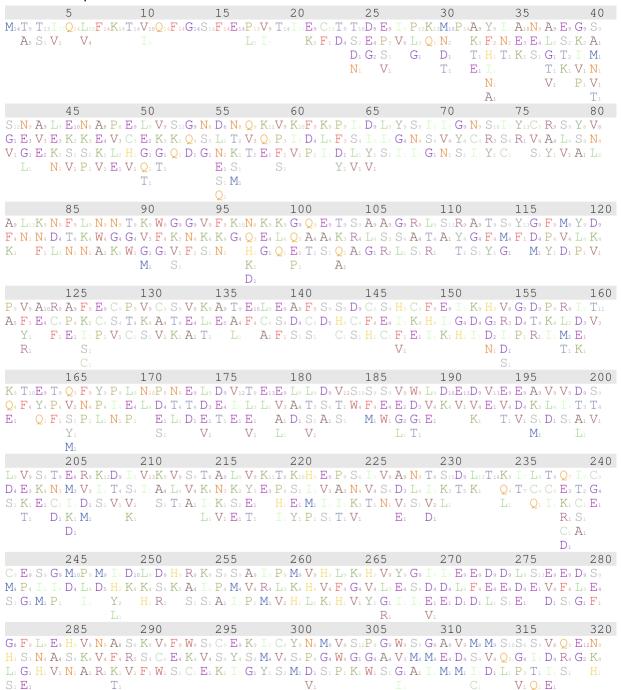
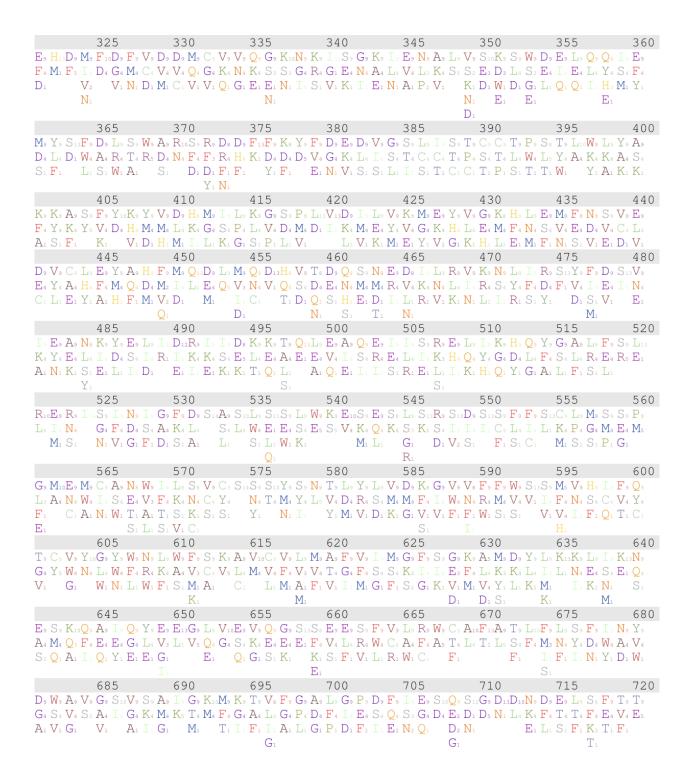


Figure 4. Computed SVD Score with the split defined by CBSV vs. UCBSV across the genome in windows of 500 bp, sliding in increments of 100 bp, and resulting SVD Scores plotted across the genome. Boundaries between genes are marked with vertical lines to further characterize the CBSV and UCBSV genomes. Rates of molecular evolution were estimated using CODEML implemented in PAML (Phylogenetic Analysis by Maximum Likelihood) ⁵⁴. The results are shown for each gene and D represents the difference in likelihoods from the null hypothesis (CBSV and UCBSV have equal rates) and the alternative hypothesis (CBSV and UCBSV have different rates).

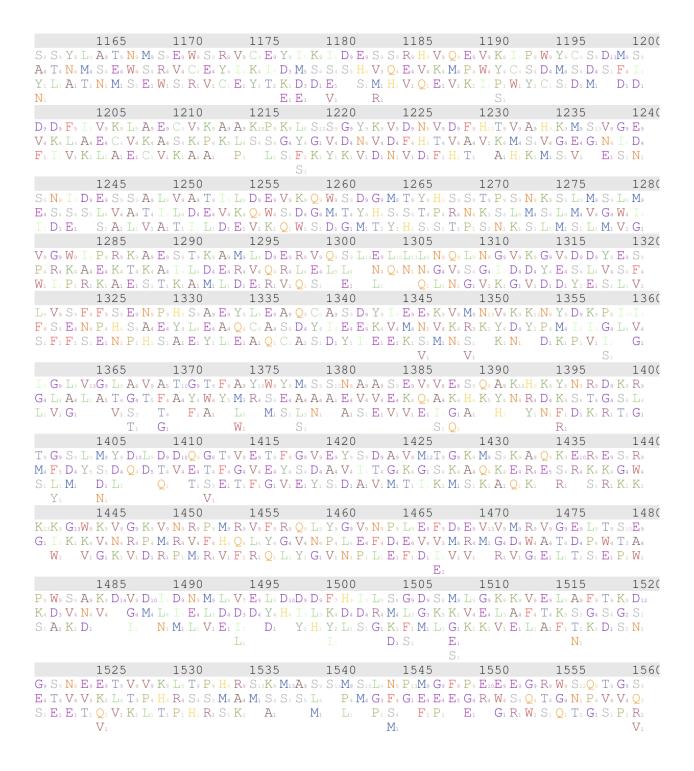


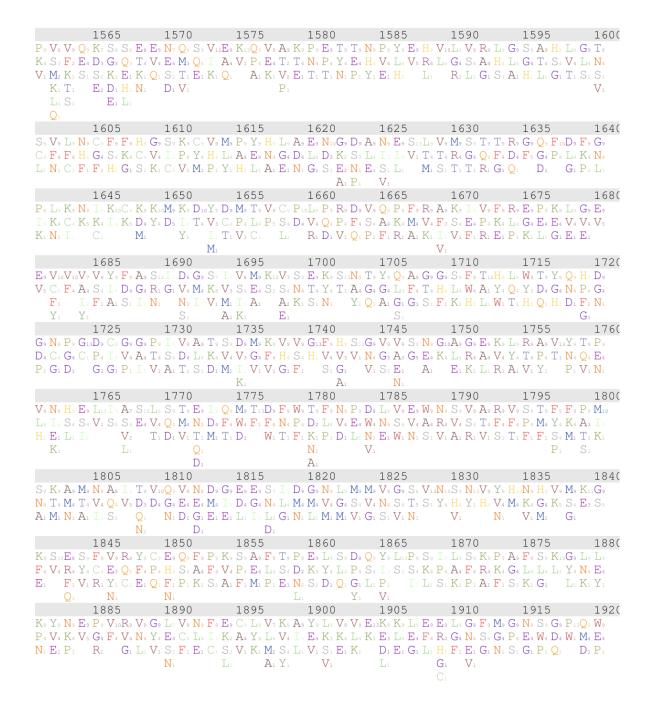
Supplemental Figure 1. CBSV Amino Acid variability obtained using datamonkey.org. Once files are uploaded to the site, the images below are obtained from the "Information from upload" tab and the pdf is downloaded.

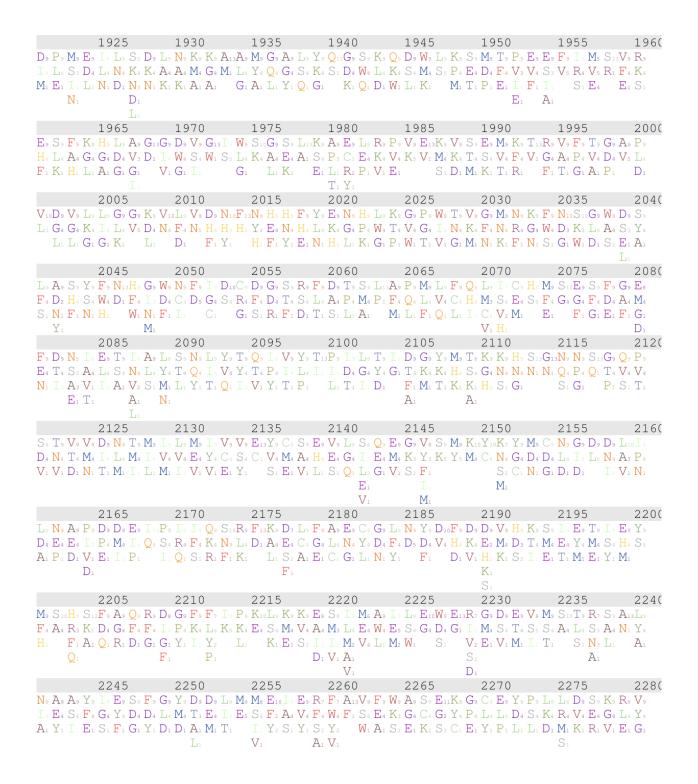


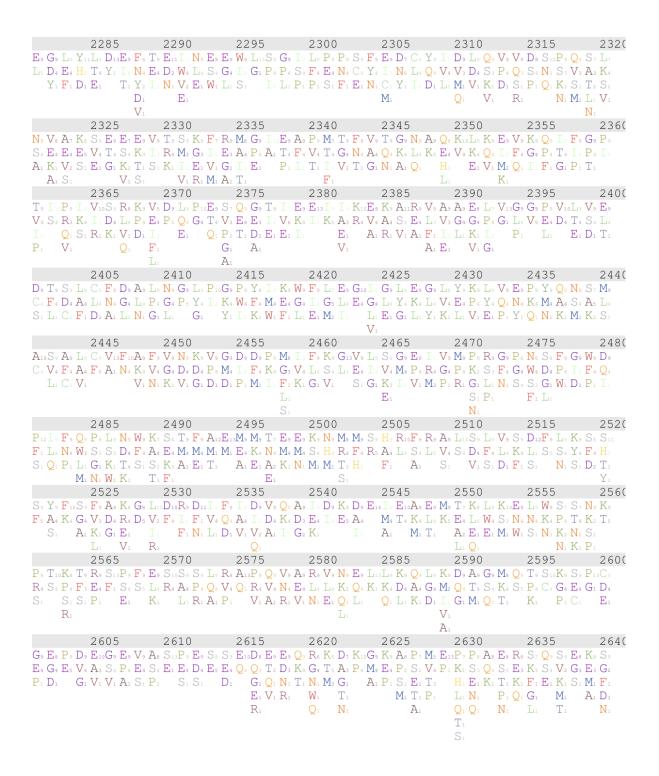


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M ₄ P ₄ G ₄ P ₁ A ₄ G ₄ T ₄ S ₄ D ₄ S ₅ Q ₄				
			1 L1 T1 A1 I1 E1 P1 T1	
	~			
765 770	775	780 785	790	795 800
L10T9 L9 V9 K10D9 K9 A13 <mark>N</mark> 9 E9 L9	A, D, K, M, Q, S, L, N	110A8 N9 D9 I8 R9 V12H	9 G9 G14V9 G10T13G9 K9 S	S9 T9 A9 L13P9 R9 E9
K4 D3 S4 A4 G4 E4 L4 E1 D4 K4 M4	Q4 S4 L4 N4 A4 C4 D4 I	3 R4 V4 H4 G4 G4 G1 G	5 T4 K4 S4 K1 A4 L4 I	P4 S3 E4 R1 I3 S4 F4
$E_1 \ K_1 \ D_1 \qquad A_1 \ {\color{red}N_1} \qquad L_1 \ A_1 \ D_1$	$K_1\; \boldsymbol{M_1}\; \boldsymbol{Q_1}\; S_1\; L_1\; \boldsymbol{N_1}\; \boldsymbol{A_1}\; \boldsymbol{V}$	V_1 D_1 I_1 R_1 V_2 H_1 I_1	V_1 T_1 $S_1 T_1 Z_2$	$A_1 L_1 P_1 = E_1 L_1 M_1$
V_1		Т1		R_1 M_1
805 810	815	820 825	830	835 840
L9 M9 R9 F8 G9 A9 V13L9 M9 C9 V13				
G4 S4 V4 L4 I4 C4 M1 P4 T4 R4 T1				
$R_1 \mathbf{F}_1 \mathbf{G}_1 \mathbf{A}_1 \mathbf{V}_1 \mathbf{L}_1 = C_1 \mathbf{V}_1 \mathbf{P}_1$	$R_1 V_1 L_1 \qquad L_1 H$	lı Eı Sı Fı Mı Aı Lı Y	ı Gı Fı Dı Vı Sı Lı Aı Y	I_1 G_1 V_1 T_1
Y ₁	٥٢٢	0.00	070	875 880
845 850	855	860 865	870	
R ₉ T ₉ G ₉ S ₈ K ₉ P ₉ I ₉ T ₉ V ₉ M ₉ T ₉ K ₄ P ₄ M ₄ T ₄ I ₅ M ₄ T ₄ Y ₄ G ₄ Y ₅ A ₄				
	A ₁ L ₁ N ₁ F ₁ H			14
885 890	895	900 905	910	915 920
P ₉ V ₉ H ₉ L ₉ N ₉ P ₉ L ₉ F ₉ S ₉ L ₁₃ M ₉				
N ₄ P ₄ L ₄ F ₄ S ₄ L ₄ I ₄ R ₄ E ₄ S ₁ S ₄				
H ₁ L ₁ N ₁ P ₁ L ₁ F ₁ S ₁ L ₁ M ₁ E ₁				
				Mı
925 930	935	940 945	950	955 960
D9 I9 H9 T9 L10G9 L8 M9 D9 V9 K9	K9 W9 A9 E9 M9 Q9 G10 T	9 G9 V9 F9 G9 D13T9 T	9 K9 D9 T9 G9 N9 V14L9 V	710 F 9 V9 A9 S9 Y9 K9
				S4 Y4 K4 D4 V4 D5 A4
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$A_1 \ E_1 \ M_1 \ Q_1 \qquad T_1 \qquad V$		$_{1}$ T_{1} G_{1} N_{1} L_{1} F_{1}	$A_1 S_1 Y_1 K_1 V_1$
		T1 T1		
965 970	975	980 985	990	995 1000
D ₁₀ V ₉ D ₉ T ₉ C ₉ A ₉ E ₉ K ₁₃ L ₉ S ₉ D ₉				
C4 A4 E4 K4 L4 S4 D4 S1 G4 L4 P4				
$T_1 C_1 A_1 E_1 K_1 L_1 \qquad D_1 K_1 G_1$	$L_1 P_1 V_1 L_1 I_1 V_1 D_1 G$ K_1	Rı SıNıFı KıNıI		C1 L1 V1 M2 D1 T1 K1 A1 G1
1005 1010		1020 102		1035 1040
To Ko Fo Io Io Ao To NiaMo Ma Eo				
V ₄ A ₄ T ₄ N ₄ M ₄ V ₄ E ₄ M ₁ G ₄ V ₄ T ₄				
F ₁ I ₁ V ₁ M ₁ V ₃ N ₁ M ₁ E ₁ I ₁ G ₁			1 S1 P1 G1 L1 F1 S1 E1 H	~
$A_1 T_1$ N_1			D_1 S_1	
1045 1050	1055	1060 106	5 1070	1075 1080
L9 H9 S13Q13R9 I9 S13Q9 A9 E9 S9	K ₉ Q ₉ R ₇ F ₉ G ₉ S ₈ V ₉ G	s S M K K G T V	9 Y9 K9 F9 G 9 R9 E9 T9]	L9 P9 D9 S10M13S9 N9
S5 I4 R1 I1 A4 E4 A1 K4 Q4 S4 F4	G4 S4 V4 G4 S4 M4 K4 K	4 G4 T3 I3 Y4 K4 F 4 G	4 K4 E4 T4 L4 P4 D4 S4 I	14 S5 N4 R4 N1 G4 S4
Q_1 $S_1 Q_1$ $E_1 S_1 K_1 Q_1$	$R_1 \; \textbf{F}_1 \; S_2 \; S_1 \; \textbf{V}_1 \; \textbf{G}_1 \; S_1 \; \textbf{M}$	I1 K1 K1 G1 T1 V1 Y1 K	1 F 1 G 1 R 1 E 1 T 1 L 1 P 1 I	$N_1 = M_1 = R_1 M_1$
	G_1 R_1	$V_1 M_1$		
1085 1090		1100 110		1115 1120
R ₉ M ₉ G ₉ S ₉ T ₉ E ₉ S ₉ A ₉ L ₉ L ₉ C ₉				
Ta Ea Sa Aa La La Ca Fa Aa Ya Ga				
$G_1 S_1 T_1 E_1 S_1 A_1 L_1 L_1 C_1 F_1 A_1$	$Y_1 \mathbf{G}_1 \mathbf{L}_1 \mathbf{K}_1 \mathbf{P}_1 \mathbf{V}_1 \mathbf{V}_1$	V_2 $S_1 K_1 V_2$	1 K1 K1 S1 K1 Q1 A	$A_1 L_1 T_1 A_1 S_1 M_1 F_1$
1105 1100	1125	A ₁	1150	1155 1166
1125 1130		1140 114		1155 116(
M ₉ F ₉ E ₉ A ₉ N ₉ Y ₉ M ₉ F ₈ T ₉ A ₈ H ₉ N ₄ Y ₄ M ₄ F ₄ T ₄ A ₄ H ₄ L ₄ V ₄ D ₄ S ₄				
E ₁ A ₁ N ₁ Y ₁ M ₁ F ₁ T ₁ A ₁ H ₁ L ₁ V ₁	~			
	ט מון דוו ועונען ען דער זע		1V1 11 111 11 11 11 11 11 11 11 11 11 11	VI VI GI TI CI DI DI

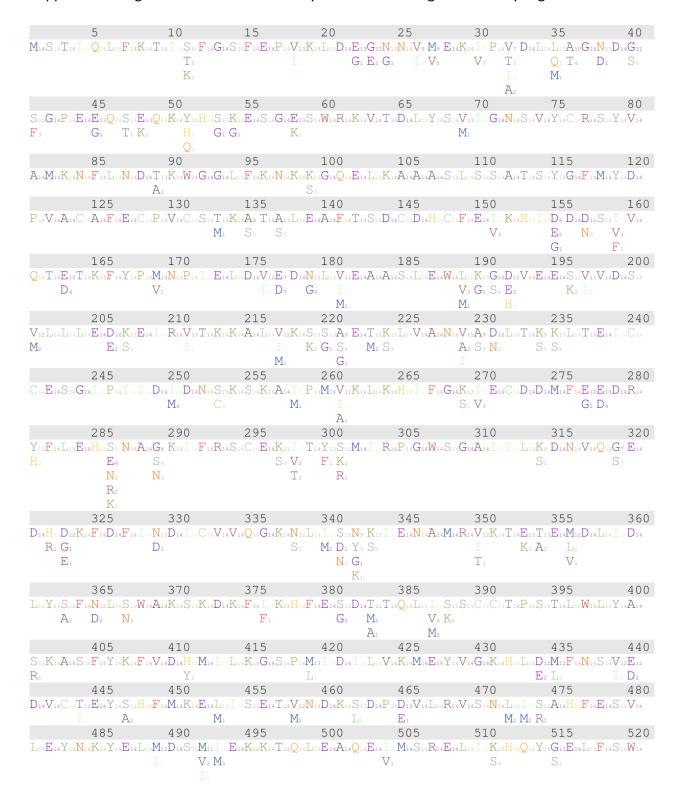








Supplemental Figure 2. UCBSV AA variability determined using datamonkey.org.



	560
$R_{14}E_{14}R_{14}L_{11}C_{12}L_{12}K_{12}L_{14}G_{14}M_{14}G_{14}S_{14}S_{14}N_{14}L_{14}Y_{14}T_{14}Y_{14}W_{14}V_{12}E_{14}R_{14}E_{14}E_{14}K_{10}Q_{12}S_{13}E_{14}K_{13}S_{14}S_{16}S_{14}C_{12}L_{14}L_{14}S_{14}K_{14}C_{12}L_{14}C_{1$	P14
$M_3 Y_3 M_2 S_2$ E_2 $S_2 H_2 L_1 S_1 N_4 V_3 F_2$	
	600
565 570 575 580 585 590 595 G16M16E14M16L14I16N12W16V13S16E14V13C16K13S13K14Y16H16S16L16V14N14C16V14D16S16G14I16L16F14L16W14S16S16M11V13H16L16A12	600
Ma S2 A2 A1 S1 N1 V2 II M2 V2 M1 V2 II M2 V2 II	
FM 52 A2 A1 51 N1	LX4
605 610 615 620 625 630 635	640
V12S14V13Y14G14Y14W14N14L14W14F14S14Q14A14M11C14V14L14F14L14V12S14N14F14S14S1 K14M12V14S11Y11L14K14K12L14L14V13	
A_2 I_1 V_2 V_1 I_2 G_5 I_2 N_2 F_3 T_2 I_1	
L_1 C_2 G_1	
645 650 655 660 665 670 675	680
$E_{14}K_{12}K_{14}L_{14}A_{14}M_{12}K_{14}N_{14}E_{14}E_{12}G_{14}F_{14}V_{13}E_{14}V_{14}Q_{14}G_{14}K_{14}E_{14}E_{14}S_{13}F_{14}V_{14}L_{14}K_{14}W_{14}C_{14}A_{14}A_{14}A_{14}L_{14}F_{14}L_{14}S_{14}F_{14}L_{14}N_{14}A_{1$	Y ₁₄
S_2 I_2 A_2 I_1 P_1 V_3	
	720
$D_{14}W_{14}A_{14}V_{14}G_{14}C_{14}V_{13}S_{14}A_{14}I_{14}G_{14}K_{14}M_{14}K_{14}I_{14}M_{12}F_{14}S_{12}A_{14}I_{14}G_{14}P_{14}D_{14}F_{14}M_{12}E_{14}K_{10}Q_{14}D_{14}G_{14}D_{14}D_{14}I_{14}K_{13}F_{14}I_{14}F_{14}F_{14}G_{14}F_{14}G_{14}F_{14}G_{14}F_{14}G_{14}F_{14}G_{1$	E14
M_1 I_2 G_2 I_2 S_4 S_1	
	760
$ V_{14}E_{14}I_{15}P_{14}G_{14}D_{14}S_{14}S_{14}S_{14}S_{14}S_{14}A_{14}Q_{14}T_{14}F_{14}G_{14}E_{14}W_{14}I_{15}E_{14}H_{14}C_{14}I_{14}K_{14}F_{15}N_{14}I_{14}V_{14}S_{14}I_{15}E_{14}P_{14}T_{14}T_{14}S_{14}G_{14}P_{14}M_{14}I_{15}\\ M_2 \qquad \qquad Y_1 $	I 14
	800
100 100 100 100 100 100 100 100 100 100	
I 2 I S 2	-114
T_1	
805 810 815 820 825 830 835	840
$S_{14}Y_{14}G_{14}A_{14}V_{14}L_{14}V_{13}C_{14}V_{14}P_{14}T_{14}S_{14}V_{14}L_{14}A_{14}N_{14}A_{14}L_{16}H_{16}E_{14}S_{16}F_{14}M_{14}S_{14}L_{16}F_{14}G_{16}F_{14}D_{14}V_{14}S_{16}L_{16}A_{14}Y_{14}R_{12}G_{13}R_{14}V_{14}S_{16}L_{16}H_{16}E_{14}S_{16}F_{14}M_{14}S_{16}L_{16}F_{14}G_{16}F_{14}D_{14}V_{14}S_{16}L_{16}A_{16}Y_{14}R_{12}G_{13}R_{16}V_{14}S_{16}L_{16}F_{16}G_{16}G_{16}F_{16}G_{16}G_{16}F_{16}G_{1$	Γ_{14}
M_1 $S_2 S_1$	
845 850 855 860 865 870 875	880
$G_{14}S_{14}K_{14}P_{14}I_{14}T_{14}Y_{14}G_{14}Y_{14}G_{14}Y_{14}A_{14}I_{14}N_{14}H_{14}F_{14}H_{14}H_{14}P_{14}S_{10}N_{14}I_{14}A_{14}Q_{14}F_{14}Q_{14}F_{14}M_{14}M_{12}M_{14}D_{14}E_{14}V_{14}H_{14}F_{14}P_{14}F_{14}P_{14}F_{14}P_{1$	V14
K_4 $V_2 L_2$	
I ₁	0.00
	920
H14L14N14P14L14F14S14L14L14R14E14L14S14P14D14K14K14M12I14K14T14S14A14T14H14V14G14H14N13V14D14L14S14T14N14H14K14V14D14L1 L2 S1	M ₁₂
	960
925 930 935 940 945 935 930 935 935 935 935 935 935 935 935 935 935	
V_3 S_2 H_1 I_1 I_2 E_1	V 14
L_2	
	1000
$D_{14}V_{14}C_{12}A_{13}E_{12}K_{14}L_{14}K_{14}D_{14}K_{14}G_{14}F_{14}P_{14}V_{14}I_{12}K_{14}V_{14}D_{14}G_{14}S_{12}N_{14}F_{14}S_{14}K_{14}N_{14}T_{14}E_{14}V_{14}Q_{14}K_{14}M_{14}V_{14}D_{14}G_{8}M_{13}Q_{10}G_{14}E_{14}V_{14}G_{14$	K14
$S_1D_2 \hspace{1.5cm} M_2 \hspace{1.5cm} R_2 \hspace{1.5cm} E_4L_1R_2$	
A_2 K_1	
$ m M_{i}$	
	1040
F14I12V14A14T14N14I14I14E14N14G14V14T14L14D14V14D14V14V14V13D14F14G14E14R14M14S14P14N14L14C14S14E14D13S14C14I14L14M14	Q 14
$egin{array}{cccccccccccccccccccccccccccccccccccc$	1000
1045 1050 1055 1060 1065 1070 1075 SisQi4Si4IisSi4Qi4Ai4Ei4Si4Ki4Qi4Si4Fi4Gi4Si4Vi4Gi4Si4Mi4Ki4Si4Gi4Si4Vi4Yi4Ki4Fi4Gi4Si4Ei2Ti4Li4Pi4Di4Si4Mi4Si4Si4Si4Si4Si4	1080
K_1 — D_2 — N_5	V 14
	1120
$G_{14}S_{14}T_{14}E_{14}S_{14}A_{14}L_{14}C_{14}F_{14}A_{14}Y_{14}G_{14}L_{14}K_{14}P_{14}V_{14}V_{14}D_{14}M_{14}G_{14}S_{14}V_{14}S_{14}S_{14}S_{14}V_{14}Q_{14}S_{14}Q_{14}A_{14}L_{14}T_{14}A_{14}S_{14}M_{14}S_{1$	
G_1	

	1125	1130	1125	1140	11/5	1150	1155	1160
F.A.N.V	1125 14 M 14 F 14 T 14 A 13 H 1							
15142A14IN14 1	T ₁	141114 V 1411114Q1		14E 14V14E 14E14E11				14 L 14
	1165	1170		1180				1200
V T 7\ T	1105 14 <mark>N14</mark> M14S14G14W1							
I 14L114A14 I	14IN14IM14014G14W1		V ₁ K ₂		4 V 14Q14 L 14 V 13Q1 I 1	4 I 14P 14W14 I 14C1	4014D14M14014D1	14 D 14
	1005					1000	1005	1240
T T T7 T7	1205	1210	1215	1220	1225	1230	1235	
	14L14 A 14 E 14C14 V 1	14K14A14A14K14P1	4K14S14Q14C14G1					14 IV 14
	1045	1050	1055			1070		1000
M D T G					1265		1275	1280
M14D14E14S	14S13A14L14V14A1 R1 S1		.4V14K14Q14W14S1	14D14G14L14T14Y1	4H14S14S14T14P1	4S14N14K14S14L1	4M14S14L14M14V1	14 G 14
	1285		1205	1300	1305	1310	1215	1320
M T D C	1285 14K14A14E14K14T1							
W14 1 14 F 14 O	14[X14 H 14 E 14[X14 I 1	14K14E14		.4114 E 1411411411	4IV14Q14L114IV14G1	V ₁	4D14 I 14E14 O14L1	14 V 14
	1325	1330		1340	12/5		1355	1360
DFFC	1323 14 E 14 N 14 P 14 H 14 S 1							
IX14E 14E 14D	14E14IN14E 14II14O1	14 A 14 L 14 L 14L014 L 1	4514 Q 14C14 A 1451	14D14 I 14 I 14B14B1		S ₂		14 G 14
	1365	1370	1375	1380		1390		1400
M13V14G14L	14A14V14A14T14G1							
V_1			D				S2	
	1405	1410			1425	1430	1435	1440
S14L114M12F	14D12M14D14D14Q1							
		S		M ₃			E ₂ S ₁	
			S ₂				2. 0.	
	1445	1450		1460	1465	1470	1475	1480
G14W14K14A	14 G 14 K 14 M 10 N14 R1							
		I ₁			Iz		A ₁	
	1485		1495	1500			1515	1520
D ₁₄ V ₁₄ K ₁₄ E	14L14 <mark>N14</mark> V13D13A1							
214 7 141 (142	I1 G1 G2				S ₂ Y ₁		S ₃	S ₂
	1525		1535	1540	1545		1555	1560
D.,E.,E.,T	14 I 14 V 14 N 13 L 14 T 1							
H ₂	S ₁	INT INTINITUO INTO	T ₆		TOTAL TALLALIA	4 L 14 C 14C14V14C1	4 2 14 1 14 0 142 1141 1	V_3
G ₁			10					M_1
N_1								1.11
141	1565	1570	1575	1580	1585	1590	1595	1600
MKKT	13K8 K10E14D12E1							
	13 C 8 1 C 10 E 14 D 12 E 1					R ₁		14 114
	S ₂ T ₂		Si		V Z	IXI		
11 Q1	1605	1610	1615	1620	1625	1630	1635	1640
N.C.F.F	14H14G14S12K14C1							
IN14C14F 14F		I1 V 13 ± 14 E 14 ± 14 11 1	4 L14A14L14IX14G1		I 4	4 I 145 14 G 14 Q 14 F 1		E ₁
	T_1			E 2			111	121
		1650	1655	1660	1665	1670	1675	1600
N. T. K. C	1045 14S12K14I7 T14D1							
IN14 ± 14IX14 C	K ₂ V ₇	V ₃	S ₁	14D14 V 14Q14E 14E 1	A1	41*1141X141514E 141X1	41114 G 141114114 V 1	14 V 14
	1\(\frac{1}{2}\)	M ₁	51		Ai			
	1685	1690	1695	1700	1705	1710	1715	1720
I14V14C12F	14T12S14I14N14G1							
Y_2	A_2		A_1	A_2	~		N_3	
		1730	1735	1740	1745	1750	1755	1760
G14D14C14G	1725 14G14P14I14V14A1		1735 4V14V14G14 <mark>F14H</mark> 1				1755 4Y14T14P14V14N1	
G14D14C14G	1725							

1765 E14L14L12N14C12I10S14G12D1			1780			1795	1800
M ₂ Y ₂ V ₄ S ₂ 1805		1815		1825	1830	1835	1840
I 14 N 14 T 14 I 12 T 14 V 14 Q 14 A 14 N 1 M2	1E14G14E14E14L1	3 I 14D14G14 <mark>N</mark> 14L	14 M 14 I 12 V 14 G 14 Y 2		4Y14 <mark>H14N14H14</mark> V1		
1845 S14 F1 4M10S13Y14C14E14Q14 F 1	1850 4P14 <mark>N12</mark> C14A14 F 1	1855 4T14K14E12L114R	1860 12D14Q13Y14L14P:	14S14I11L14S14K1	1870 4P14A14 <mark>F14</mark> S14K1	1875 4G14L14L14K14Y1	1880 4N14
T ₄ K ₁ 1885 E ₁₄ P ₁₄ V ₁₄ R ₁₃ V ₁₄ G ₁₄ S ₁₀ V ₁₁ N ₁		1895	1900	V ₃ 1905 4E ₁₃ N ₅ I ₁₁₄ G ₁₄ F ₁		1915	1920
Q1 L4 M2 D4				$egin{array}{c} K_1 \ D_4 \ T_2 \ A_2 \end{array}$	R ₂ E ₅ G ₂		
1925 E14I14L14E9 D14L14 <mark>N</mark> 14K14K1	1930	1935	1940	1945	1950	1955	1960
D_5			F		V1 M4		
1965 Ki4Hi4Li4A14G14G14D14V14G1			1980 14S14P12V12E10K1 T1 I2 K4 S1				200(₄ L ₁₄
2005 L14L14G14G14K14I12L14V14D1 M2	2010 4 <mark>N14F14N14H14F</mark> 1	2015 4F14Y14F14N14H	2020 14L14K14G14P14W	2025 14T14V14G14I14 <mark>N</mark> 1	2030 4K14 F 14 N 14K14 G 1	2035 4 <mark>W</mark> 14D14S14L14 A 1	204(₄ S ₁₂ K ₂
2045 Y ₁₃ F ₁₄ N ₁₂ H ₁₄ S ₁₂ W ₁₄ N ₁₄ F ₁₄ M ₁ N ₁ D ₂ N ₂ T ₁		2055 4S14 <mark>F14</mark> D14T14S		2065 14 F 14 Q 14 L 14 V 14 C 1		2075 4F14G14N14F14D1	208(4D14
2085	2090	2095	2100	2105	2110	2115	2120
I 11 E 14S12 A 12 A 14L14R14 N 14L1 V1 K2 T2	4 Y 14 T 14 Q 14 M 14 V 1	4Y14T14P14I14L	14 T 14 I 14 D 14 G 14 Y :	14 I 14 T 8 K 14 K 14 H 1 A 4 M 1 V 1	4R14G14N14N14S1	4 G 14 Q 14 P 14 S 14 T 1	4V14
2125 V14D14N14T14T12M11L14M14T1 M2 I3	2130 4V14V14E14Y14C1			M ₁₄ Q ₁₄ F ₁₄ K ₁₄ Y ₁	2150 4M14C14 <mark>N1</mark> 4G14D1		216(₄ V ₁₄
2165 P14D12D14E14V14S12M14T11Q1	2170 ₄ S14S14 F 14S14E1 D2			2185 13 F 14D14D14 V 14 H 1		2195 4I:4E14Y14M14S:	2200 4H14
2205	N ₁			2225	2230	2235	2240
S14F14M14L13K14D14G10V12Y1 M1 D4 L1 M1						2S14 A 14L14 <mark>N14</mark> A1	
2245	2250	2255	2260		2270	2275	2280
$\begin{array}{c} Y_{14} I_{14} E_{14} S_{14} Y_{14} G_{14} Y_{14} E_8 \ D_7 \\ \\ D_6 \end{array}$	4L13M14V11E14I1 P1 I1 T1 M1	4E14R14Y14A14V	14 F 14 W 14 A 14 T 8 E 1 A 6 G 1		4P14L14L14D14S1	4K14R14V14E14G1	4L14

2285 Y14K10D14D14H11T14D14I16			2300 P14S14 F 14 E 14 H 14			2315 Si3Ga Sa EizKid	232(P ₁₂ E ₁₄
S_4 Y_2 N_1	\ [2	73 [2			I1 E 2 V2 I		
2325	2330	2335	2340	2345	2250	2355	2360
Z3Z3 L13S13I5 E12S13H10D13G11							
F ₁ G ₁ V ₃ G ₂ N ₁ C ₂ N ₁ S ₃		1112012 112 11 0 11.	142 14 0 14 2 14 0 14	F ₂	S ₁	S ₁ S	
T ₃ Y ₂	$A_1 L_2$						
P_2	M_1 T_2						
A ₁	0070	0275	0000	0205	0200	0205	0.406
2365 V ₁₄ M ₉ A ₁₄ K ₈ N ₁₄ I ₁₄ D ₁₄ L ₁₄	2370 P. F. V. O. G. J	2375			2390 M. T. N. S. P. V		
V14M9 A14IX8 IN14 L14 D14 L14 V5 S5		E ₂ V ₁ I		F ₁			L 13 C 14 A1
R ₁		V ₁	M ₁		G ₂	-	
2405	2410	2415	2420	2425	2430	2435	2440
L14C14F14N14A14F14N14G14							Y14A14
	Ŋ	\mathbf{I}_1	1	D ₆ S ₂		Ξ_4	
2445	2450	2455	2460	2165		S ₂ 2475	2480
L14C14T14F14A14Y14V13H14							
	S ₁ S ₇ E ₁		K ₁		K ₂	-	I ₅
2485	2490	2495	2500	2505	2510	2515	2520
$K_{14}P_{14}D_{14}G_9$ $C_{14}G_7$ $C_{14}T_{14}$							
E5 S6	S ₅		Ε1	K_1	$S_3 Y_4$	M_4	S_2
E ₁ 2525	2530	G ₁ N ₁ 2535	2540	2545	2550	2555	2560
Q14E14E12K12K11A13K7 V13							
K ₂ Q ₁ S ₃ T ₁ S ₆ M ₁						55	G1
N_1 G_1			I1	Iı Sı			
2565	2570				2590		
T14R14S14P12F14E14A14S11					G14L14Q14T14K14l	K14S14P14C14 G 14I	E14P14
S ₂ K ₁ 2605	S ₂			Q ₂ N ₁ 2625	2630	2635	2640
D ₁₄ D ₁₄ G ₁₄ E ₁₄ V ₁₃ V ₁₃ D ₁₃ D ₁₄							
	N ₂ N ₁ D ₁ I						
	\mathbf{E}_{1}	D_1	1	N_1 G_1	Sı	L_1	L ₁
						Q ₁	
2645	2650		2660			2675	2680
G14D14G13S11A10I9 S12R14 E1 N3 T4 V5 K2	1D14D14 L 14D14K13 J S1	L 14 P 14 L 14 N 13A 14J S1	L14 L 14 L 14 L 14 L 14		N10 V 14 D 14 <mark>V 14</mark> D 14 <i>1</i> S 4	1 14 1 14 1 14 W 14 1 14 £	7 14K14
2685	2690	2695	2700	2705	2710	2715	2720
S14Q14S14D14N14L114T14P14	D14V14I13Q14N13F						
	M 1 S1	V_4	Y ₁		V_1		J ₂ M ₁
2725	2730	2735	2740	2745	2750	2755	2760
V14A14K14A14Y14G14V14N14	F						
2765	2770	2775	2780	2785	2790	2795	2800
A14V14E14L14N14S14Q14G14	1E14D14V14D14D14F	¹ 14E14Y14P14M14I		F 14A14L14P 14T14		514N14F14S14S14	214A14
			S_1				