

1 **Title:** Ground truthing tip-dating methods using fossil Canidae reveals major
2 differences in performance

3

4 **Running head:** Ground truthing tip-dating with canids

5

6 **Authors:** Nicholas J. Matzke^{1,2,*}; April Wright³

7

8 ¹Discovery Early Career Researcher Award (DECRA) Fellow, Moritz Lab, Division of
9 Ecology, Evolution, and Genetics, Research School of Biology, Canberra, ACT 2601

10 AUSTRALIA

11

12 ²Work performed at: National Institute for Mathematical and Biological Synthesis
13 (NIMBioS, www.nimbios.org), 1122 Volunteer Blvd., Suite 106, University of
14 Tennessee, Knoxville, TN 37996-3410; and O'Meara Lab, 425a Hesler, Department
15 of Ecology and Evolutionary Biology, University of Tennessee, Knoxville, TN 37996

16

17 ³Heath Lab, 228 Bessey Hall, Iowa State University, 2200 Osborn Dr., Ames, Iowa,
18 50011-4009

19

20 *Corresponding author. Email: nick.matzke@anu.edu.au

21

22 **Abstract**

23

24 Tip-dating methods are becoming popular alternatives to traditional "node-dating."

25 However, they have not been extensively tested. We "ground-truth" the most

26 popular methods against a dated tree of fossil Canidae derived from monographs by

27 Wang and Tedford. Using a revised canid morphology dataset we compare MrBayes

28 3.2.5 to Beast 2.1.3 combined with *BEASTmaster* (phylo.wikidot.com/beastmaster),

29 an R package that automates the conversion of dates, priors, and NEXUS character

30 matrices into the complex Beast2 XML format. We find that unconstrained MrBayes

31 analysis under the uniform node age prior fails to retrieve reasonable results,

32 exhibiting extremely high uncertainty in dates. On the other hand, Beast2 inference

33 matches the ground-truth well, under both birth-death serially sampled (BDSS,

34 disallowing direct ancestors) and sampled ancestor (SABD) tree models, as does

35 MrBayes using BDSS. MrBayes using SABD seems to have difficulty converging in

36 some analyses. These results, on a high quality fossil dataset, indicate that while tip-

37 dating is very promising, methodological issues in tip-dating can have drastic

38 effects, and require close attention, especially on more typical datasets where the

39 distinction between "method problems" and "data problems" will be more difficult

40 to detect.

41 **Keywords:** Tip-dating, total evidence dating, Canidae, MrBayes,

42 BEASTmaster, uniform node age prior

43

44

45

46 **Main text** (2100 words)

47

48 Testing phylogenetic inference methods against an externally known truth is highly
49 desirable, but is rarely possible except when an experimenter manufactures a
50 known evolutionary history either with simulations [1] or splitting populations of
51 microbial/virus cultures [2]. Even when conducted, it is debatable to what extent
52 manufactured histories are comparable to the complexity of real evolutionary
53 histories, where heterogeneity of rates, environment, and data acquisition are likely
54 to be significant [3].

55

56 Our goal is to assess Bayesian total evidence (“tip-dating”) methods. These are
57 methods where the ages of fossil OTUs (operational taxonomic units / terminal
58 taxa) are used as the primary source of dating information, rather than priors on
59 node dates. The latter, while valuable, is subject to a number of well-known
60 criticisms [4-7] such as subjectivity and incomplete use of information. In addition,
61 node-dating weakens inferences to the extent that it essentially constrains *a priori*
62 some of the clade dates that we would prefer to infer.

63

64 A number of tip-dating methods and models recently have become available [5, 6, 8-
65 11] and are being applied to empirical datasets [5-7, 12-27]. However, tip-dating
66 results seem to vary widely between methods and datasets. Some papers conclude

67 that their tip-dating results are implausible (e.g. [14, 27, 28]), and some infer dates
68 that are wildly uncertain [17, 22, 29]. Evaluation of the methods against each other,
69 or against expectations based on the fossil record, is hampered by the complexity of
70 Bayesian analyses: differences in results might be produced by differences in clock
71 models, tree models, site models, priors (user-set or default) on any of the
72 parameters used in these models, issues in implementation (bugs in the code,
73 decisions about defaults, MCMC operators, etc.), user error in setting up the analysis
74 or post-analysis processing, or issues with the data itself. In addition, several of the
75 issues above could be in play and interacting in any particular study.

76

77 As tip-dating methods are still in development, it is useful to “ground-truth” the
78 major methods and models on an “ideal” empirical dataset, one where the fossil
79 record is of sufficiently high quality that the true evolutionary tree and dates are
80 broadly known even without complex computational methods. An ideal dataset
81 would also meet the assumptions made by the models (Table 1). When multiple
82 methods are run on the same ground-truth dataset, not only can differences in
83 inference be attributed to differences in the method, but an assessment can be made
84 about which methods are making inferences closer to the “known” truth. Methods
85 that fail on the ideal dataset are unlikely to provide useful inferences on typical,
86 non-ideal datasets.

87

88 Fossil datasets ideal for ground-truthing are few and far between, due to the
89 vagaries of preservation and description, but one for which a strong argument can
90 be made is the fossil Canidae (dog family; [30]). Canids avoid the challenges faced by
91 most datasets (Table 1). The bulk of canid evolution occurred in North America
92 from the Eocene to present, and their fossil record is approximately continuous,
93 with fossil diversity greater than extant diversity (approximately 35 living species,
94 at least 123 well-described fossil taxa). In addition, the group has been thoroughly
95 revised in three major monographs on the three subfamilies of Canidae: the extinct
96 Hesperocyoninae (~27 species, 40-15 Ma; [31], the extinct Borophaginae (~66
97 species, 34-2 Ma; [32], and the extinct and extant Caninae (>40 fossil species, 34 Ma-
98 present; [33]). All living dogs thus represent a small surviving branch, originating
99 10-12 Ma, of a much more massive tree of fossil Canidae. Thus, apart from utility for
100 ground-truthing, the Canidae serve as a useful group for comparing trait evolution
101 inferences made with living-only versus living+fossil datasets [24, 34, 35].

102

103 **Methods**

104

105 *Ground-truth tree and characters.* The ground-truth tree was digitized from the
106 monographs of Wang and Tedford, with judgment calls resolved in favor of
107 preserving the authors' depiction of divergence times (SM). Morphological
108 characters and dates came from the published matrix of Slater (2015) [24, 36].

109

110 *Tip-dating analyses.* MrBayes analyses were conducted by modification of Slater's
111 commands file. A large number of variant MrBayes analyses (35 total) were
112 constructed to investigate several issues that were noticed between MrBayes
113 versions, documentation, and Slater's commands file (Supplemental Material, SM).
114 Due to the many node date calibrations, these did not substantially change Slater's
115 inferred tree, but if left unchanged they would have a large impact on unconstrained
116 analyses. As the issues are subtle and due in part due to MrBayes' implementation
117 or documentation, they may cause problems for other MrBayes users (SM).
118
119 We focused on six analyses (four MrBayes 3.2.5 analyses and two Beast2.1.3) to
120 compare to the ground-truth tree, and to Slater's published analysis (mb1_orig)
121 analyses. These were: (1) *mb1*: Slater's original uniform node age prior analysis
122 including node date calibrations, with some corrections; (2) *mb8*: uniform node age
123 prior, no node dates, flat priors on clock parameters, uniform(45,100) prior on the
124 root age; (3) *mb9*: mb8 but with SABD tree prior and flat priors on speciation,
125 extinction, and sampling rate; (4) *mb10*: mb9 but BDSS, i.e. disallowing sampled
126 ancestors; (5) *r1*: Beast2 BDSS analysis with flat priors were used for each major
127 parameter (mean and SD of the lognormal relaxed clock; and birth, death, and serial
128 sampling rates); (6) *r2*: Beast2 SABD analysis with the same priors. Beast2 analyses
129 were constructed with BEASTmasterR [37]; full details on the analyses and post-
130 processing steps are presented in SM.
131

132 **Results**

133

134 The dated trees from the six focal analyses are compared in Figure 1 (plots of all
135 trees are available in SM), and key priors and results are shown in Supplemental
136 Table S1. The general picture is clear: the unconstrained MrBayes uniform node age
137 prior analysis (mb8) produces implausibly old ages and huge uncertainties, with the
138 age of Canidae overlapping the K-Pg boundary. This behavior was also noted by
139 Slater (2015). The ground-truth dates of crown *Canis* (which includes genera *Cuon*,
140 *Lycaon*, and *Xenocyon*) and crown Caninae are 3.2 and 11.7 Ma, but mb8 makes
141 mean estimates of 27.5 and 38.9 Ma, and even the very wide HPDs (width 22-25 my)
142 do not overlap the truth. More surprisingly, even Slater's highly constrained analysis
143 (mb1), although much closer, does not produce HPDs (5.1-9.6 Ma; 17.8-25.5 Ma)
144 that overlap the ground truth for these nodes. In contrast, both Beast2 analysis (r1
145 and r2) and MrBayes BDSS (mb10) produce mean estimates close to the truth, with
146 narrower HPD widths (2-3 my).

147

148 The MrBayes SABD analysis (mb9) wrongly estimated these node ages as identical
149 with the age of Canidae; this is due to mb9 misplacing *Lycaon pictus* (African wild
150 dog) and *Cuon javanicus* (Dhole) in the extinct Borophaginae. If this is ignored, the
151 estimates are much closer, although the crown *Canis* estimate still fails to overlap
152 (Supplemental Table S1, notes 5 and 6). A suggestion to repeat mb9 with 4 runs
153 instead of 2 (Mike Lee, personal communication) did produce an mb9 result that

154 placed these taxa in the conventional position (SM), but we present the
155 unconventional result to emphasize that it appears much greater care is required to
156 achieve convergence with MrBayes SABD than with other methods.
157
158 Comparing topological distances (Supplemental Table S1) provides a more
159 systematic assessment of topology differences between analyses. Randomly chosen
160 trees in the post-burnin posterior distribution of Slater's undated MrBayes analysis
161 (mb2; SM) have a mean symmetric distance of 31.0% (95% C.I.=±8.3%). The dated
162 MCC trees of all 8 focal analyses fall within this range, except for the MrBayes-
163 uniform and -SABD analyses.
164
165 A comprehensive view of the correlation between date estimates and ground truth
166 is shown in linear regression plots (SM) comparing the ages of nodes that are shared
167 between the ground truth tree and estimated tree (i.e., have the same descendant
168 OTUs after removing OTUs not shared by both trees; 127 OTUs were shared). While
169 all analyses have statistically significant correlation to ground truth ages, mb8 has a
170 lower R^2 (0.71) than the others (>0.9), and also has a systematic bias towards older
171 ages (intercept=6.67±2.94; for other analyses intercept is ~0-1). All of the BD
172 analyses have a bias towards underestimating dates near the base of the tree, where
173 the ground truth tree is dominated by Hesperocyoninae, with a lower density of
174 fossil OTUs and thus longer branches; the BD methods tend to infer shorter
175 branches here, leading to younger ages. This bias leads to an underestimation of

176 ages by about 2 my at age 20 Ma, to 4-5 my at 30 Ma. Prediction of tip dates
177 generally succeeded in overlapping the true value, although uncertainty is usually
178 high (5+ my) and extreme in the case of mb8 (17+ my). SABD analyses consistently
179 produce younger date estimates than BDSS analyses, but the effect is small (~1 my).

180

181 **Discussion**

182

183 The result of greatest interest is the poor performance of the MrBayes uniform node
184 age prior even in a “perfect-case” dataset. Whether or not this is surprising depends
185 on researcher background. We suggest that reasoning from first principles suggests
186 that effective tip-dating under the uniform node age prior will be difficult-to-
187 impossible without strongly informative priors on node dates and/or clock rate and
188 variability. Apart from such constraints, nothing in the tip dates or the uniform node
189 age prior restricts the age of nodes below the dated tips; thus the node ages are, in
190 effect, scaled up and down as the root age is sampled according to the root age prior
191 (a required setting for the MrBayes uniform node age prior). Without informative
192 priors, the clock rate and variability parameters will adjust along with the tree
193 height; highly uncertain node dates will result.

194

195 Despite what first principles suggest, we suspect may surprise some researchers.
196 The MrBayes uniform node age prior was the leading model in the early tip-dating
197 literature (11/16 papers as of mid-2015, 9 of them as the exclusive Bayesian tip-

198 dating method), and until recently (October 2014, v. 3.2.3) the uniform node age
199 prior was the only option available in MrBayes. Early tip-dating efforts in
200 Beast/Beast2 required tedious manual editing of XML and/or elaborate scripting
201 efforts (such as BEASTmasteR), whereas MrBayes was relatively easy to use.
202 Therefore, many early attempts at tip-dating used the uniform node age prior.
203
204 In contrast to the disappointing results with the uniform node age prior, analyses
205 using the BDSS or SABD tree prior (mb10, r1, r2) fared well against ground truth.
206 Given only the characters and tip-dates, and with uninformative priors on
207 parameters and the root age, these analyses were able to estimate node ages with
208 high accuracy. Surprisingly, these analyses outperformed the uniform node age
209 prior even when this analysis was given substantial additional information in the
210 form of many node calibrations (mb1). It seems that even well-constrained uniform
211 node age prior analyses have a tendency to space node ages unrealistically evenly
212 between calibrations and tip dates, regardless of morphological branch lengths. This
213 can be seen in our mb1 analysis, for example in the old age for crown *Canis*; we have
214 also observed the phenomenon with other datasets, and it seems to be a feature of
215 published uniform node age prior analyses as well. Admittedly we have not devised
216 a way to quantify the observation of “unrealistically even” node date estimates, but
217 the phenomenon does appear to be real, at least when uniform node age prior
218 results can be compared side-by-side to ground-truth and BDSS/SABD trees (e.g.,
219 mb1 vs. mb10, r1, r2), or to undated trees (e.g. mb2). The disagreement between the

220 MrBayes BDSS and SABD (m10 and m9) analyses about the position of *Lycaon+Cuon*
221 is puzzling and is discussed further in SM.

222

223 **Conclusions**

224

225 Tip-dating with the uniform node age prior was explicitly introduced [6] as an
226 alternative to node-dating, attractive precisely because tip-dating avoided various
227 undesirable compromises that researchers are forced to make to when constructing
228 node-age priors. Ronquist et al. [6] also critiqued Stadler's [38] BDSS prior as being
229 "complete but unrealistic," particularly due to assumptions about constant
230 birth/death/sampling rates and sampling in the Recent. They offered the uniform
231 prior as an alternative, free of these difficulties. If, however, strongly informative
232 node-age priors are required to produce reasonable results under the uniform node
233 age prior, the main appeal of this prior is lost. The exploration of birth-death-
234 sampling models for MrBayes [11] suggests that the future of tip-dating is likely to
235 lay in adding realism to the BDSS-like models, rather than in attempting to devise
236 wholly agnostic dating priors.

237

238 A great deal of work remains in the area of tip-dating in terms of methods testing
239 and implementing more realistic methods. We have shown that "ground-truth"
240 datasets, though rare and imperfect, are extremely useful in evaluating methods and
241 models, bringing to light issues that would be less noticeable with lower-quality

242 datasets and/or more complex setups (e.g., informative priors on parameters and
243 node dates).

244

245 **Data accessibility.** All scripts, data files, and results files are available via a zipfile
246 on Dryad (doi:XXX) [Note: Files also available at:
247 https://github.com/nmatzke/Matzke_Wright_2016]

248

249 **Competing interests.** We have no competing interests.

250

251 **Authors' Contributions.** NJM wrote *BEASTmasteR*, conducted the Beast2
252 computational analyses and drafted the manuscript. AW contributed to MrBayes
253 dating efforts and edited and corrected the manuscript.

254

255 **Acknowledgements.** We thank David Bapst, Graeme Lloyd, Jeremy Beaulieu,
256 Kathryn Massana, Brian O'Meara, and Mike Lee for helpful comments and
257 discussion, as well as the participants and attendees of the 2014 Society of
258 Vertebrate Paleontology workshop and symposium on tip-dating. We also thank the
259 BEAST developers and the *beast-users* Google Group, particularly Remco Bouckaert.

260

261 **Funding.** NJM was supported by NIMBioS fellowship under NSF Award
262 #EFJ0832858, and ARC DECRA fellowship DE150101773. Work on this topic began

263 under the NSF Bivalves in Time and Space grant (DEB-0919451). AW was supported
264 by NSF DEB-1256993.

265

266

267

268 **Captions for Figures and Tables**

269

270 **Figure 1.** Plots of (a) the ground-truth tree, (b) Slater’s original analysis, and (c-h)
271 the six focal analyses. Illustrative nodes (dates listed in Supplemental Table S1) are
272 (1) the common ancestor of crown (living) *Canis*, (2) the common ancestor of living
273 Caninae (2), and (3) the common ancestor of the total group Canidae. Panel (f)
274 shows only node (3) because all three nodes synonymize due to the misplacement
275 of living *Lycaon+Cuon* in the extinct Borophaginae. Note that the OTUs of the ground
276 truth tree and the Slater dataset do not overlap completely: the Slater dataset lacks
277 the living groups *Vulpes* (true foxes) and South American *Cerdocyonina* (e.g. crab-
278 eating fox). The ground-truth tree lacks Slater’s “outgroup” OTU (the branch below
279 node (3)). For high-resolution trees with node labels, see Supplementary
280 Information.

281

282 **Table 1.** Clade features that present challenges to tip-dating methods (or any dating
283 methods). Canidae exhibit few of the issues that may confound dating in other
284 clades (e.g. angiosperms, mammals, birds).

285

286 **Supplemental Figure 1.** Top: plot of the ground-truth tree, derived from
287 digitization of the phylogenies of Canidae published in the monographs of Wang and
288 Tedford, using TreeRogue. Bottom: Lineages through time plot of the ground-truth
289 tree.

290

291 **Supplemental Table S1.** Five Bayesian tip-dating analyses are compared to
292 (column 1) the conclusions of Tedford & Wang. The two Slater analyses (original,
293 and a modification repairing some unintended issues; cols. 2-3) represent MrBayes
294 analyses under a uniform node age prior, constrained by both tip dates and many
295 node-date priors. The third run (col. 4) shows the effect of removing the node-age
296 calibrations, and putting flat priors on the parameters for clock rate and variation.
297 The fourth run (col. 5) shows the drastic effect of switching to a fossilized birth-
298 death prior. Columns 5 and 6 show Beast2 analyses with flat clock priors and BDSS
299 and SABD tree priors, respectively. These five analyses are drawn from the 40
300 analyses shown in Supplemental Table S2.

301

302 **Supplemental Table S2.** Summary settings and results of all 40 tip-dating analyses.
303 As the table is large, it is presented as an Excel file.

304

305 **Supplemental Data Files**

306

307 *Canidae_traceLogs.pdf*-- Trace plots of key variables for all 40 analyses.

308

309 *Canidae_treeLogs.pdf*-- Plots of the MCC trees for all 40 analyses.

310

311 *Ground_truth_vs_estimated_node_ages.pdf*-- Linear regressions showing the

312 correlation between the ground truth and estimated node ages, for nodes shared

313 between the ground truth tree and estimated trees.

314

315 *Canidae_ground_truth.newick* – The “ground-truth” tree, derived from digitization of

316 the phylogenies of Canidae published in the monographs of Wang and Tedford,

317 using TreeRogue.

318

319 *Table_S2_TipDate_runs_v3.xlsx* – Summary of all 40 variant analyses (contains

320 Supplemental Table S2, and some associated notes and file locations)

321

322 *Matzke_Wright_SuppData.zip* – A zipfile of all inputs, outputs, and scripts for all

323 analyses.

324

325

326

327

328

329 **References**

330

331 [1] Huelsenbeck, J.P. 1995 Performance of phylogenetic methods in simulation.

332 *Systematic Biology* **44**, 17-48. (doi:10.1093/sysbio/44.1.17).

333 [2] Hillis, D., Bull, J., White, M., Badgett, M. & Molineux, I. 1992 Experimental

334 phylogenetics: generation of a known phylogeny. *Science* **255**, 589-592.

335 (doi:10.1126/science.1736360).

336 [3] Hillis, D.M. 1995 Approaches for assessing phylogenetic accuracy. *Systematic*

337 *Biology* **44**, 3-16. (doi:10.1093/sysbio/44.1.3).

338 [4] Parham, J.F., Donoghue, P.C.J., Bell, C.J., Calway, T.D., Head, J.J., Holroyd, P.A.,

339 Inoue, J.G., Irmis, R.B., Joyce, W.G., Ksepka, D.T., Patané, J.S.L., Smith, N.D., Tarver,

340 J.E., van Tuinen, M., Yang, Z., Angielczyk, K.D., Greenwood, J.M., Hipsley, C.A.,

341 Jacobs, L., Makovicky, P.J., Müller, J., Smith, K.T., Theodor, J.M., Warnock, R.C.M. &

342 Benton, M.J. 2012 Best practices for justifying fossil calibrations. *Systematic*

343 *Biology* **61**, 346-359. (doi:10.1093/sysbio/syr107).

344 [5] Pyron, R.A. 2011 Divergence time estimation using fossils as terminal taxa and

345 the origins of Lissamphibia. *Systematic Biology* **60**, 466-481.

346 (doi:10.1093/sysbio/syr047).

347 [6] Ronquist, F., Klopfstein, S., Vilhelmsen, L., Schulmeister, S., Murray, D.L. &

348 Rasnitsyn, A.P. 2012 A total-evidence approach to dating with fossils, applied to

349 the early radiation of the Hymenoptera. *Systematic Biology* **61**, 973-999.

350 (doi:10.1093/sysbio/sys058).

- 351 [7] Wood, H.M., Matzke, N.J., Gillespie, R.G. & Griswold, C.E. 2013 Treating fossils as
352 terminal taxa in divergence time estimation reveals ancient vicariance patterns
353 in the Palpimanoid spiders. *Systematic Biology* **62**, 264-284.
354 (doi:10.1093/sysbio/sys092).
- 355 [8] Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C.-H., Xie, D., Suchard, M.A.,
356 Rambaut, A. & Drummond, A.J. 2014 BEAST 2: a software platform for Bayesian
357 evolutionary analysis. *PLoS Comput Biol* **10**, e1003537.
358 (doi:10.1371/journal.pcbi.1003537).
- 359 [9] Gavryushkina, A., Welch, D., Stadler, T. & Drummond, A.J. 2014 Bayesian
360 inference of sampled ancestor trees for epidemiology and fossil calibration.
361 *PLoS Comput Biol* **10**, e1003919. (doi:10.1371/journal.pcbi.1003919).
- 362 [10] Heath, T.A., Huelsenbeck, J.P. & Stadler, T. 2014 The fossilized birth–death
363 process for coherent calibration of divergence-time estimates. *Proceedings of*
364 *the National Academy of Sciences* **111**, E2957-E2966.
365 (doi:10.1073/pnas.1319091111).
- 366 [11] Zhang, C., Stadler, T., Klopstein, S., Heath, T.A. & Ronquist, F. 2016 Total-
367 evidence dating under the fossilized birth-death process. *Systematic Biology* **65**,
368 228-249. (doi:10.1093/sysbio/syv080).
- 369 [12] Alexandrou, M.A., Swartz, B.A., Matzke, N.J. & Oakley, T.H. 2013 Genome
370 duplication and multiple evolutionary origins of complex migratory behavior in
371 Salmonidae. *Molecular Phylogenetics and Evolution* **69**, 514-523.
372 (doi:<http://dx.doi.org/10.1016/j.ympev.2013.07.026>).

- 373 [13] Arcila, D., Alexander Pyron, R., Tyler, J.C., Ortí, G. & Betancur-R, R. 2015 An
374 evaluation of fossil tip-dating versus node-age calibrations in tetraodontiform
375 fishes (Teleostei: Percomorphaceae). *Molecular Phylogenetics and Evolution* **82**,
376 **Part A**, 131-145. (doi:<http://dx.doi.org/10.1016/j.ympev.2014.10.011>).
- 377 [14] Beck, R.M.D. & Lee, M.S.Y. 2014 Ancient dates or accelerated rates?
378 Morphological clocks and the antiquity of placental mammals. *Proceedings of the*
379 *Royal Society of London B: Biological Sciences* **281**.
- 380 [15] Bracken-Grissom, H.D., Ahyong, S.T., Wilkinson, R.D., Feldmann, R.M.,
381 Schweitzer, C.E., Breinholt, J.W., Bendall, M., Palero, F., Chan, T.-Y., Felder, D.L.,
382 Robles, R., Chu, K.-H., Tsang, L.-M., Kim, D., Martin, J.W. & Crandall, K.A. 2014 The
383 emergence of lobsters: phylogenetic relationships, morphological evolution and
384 divergence time comparisons of an ancient group (Decapoda: Achelata,
385 Astacidea, Glypheidea, Polychelida). *Systematic Biology* **63**, 457-479.
386 (doi:10.1093/sysbio/syu008).
- 387 [16] Dornburg, A., Moore, J., Beaulieu, J.M., Eytan, R.I. & Near, T.J. 2015 The impact of
388 shifts in marine biodiversity hotspots on patterns of range evolution: Evidence
389 from the Holocentridae (squirrelfishes and soldierfishes). *Evolution* **69**, 146-
390 161. (doi:10.1111/evo.12562).
- 391 [17] Grimm, G.W., Kapli, P., Bomfleur, B., McLoughlin, S. & Renner, S.S. 2015 Using
392 more than the oldest fossils: dating Osmundaceae with three Bayesian clock
393 approaches. *Systematic Biology* **64**, 396-405. (doi:10.1093/sysbio/syu108).

- 394 [18] Huang, D., Goldberg, E.E. & Roy, K. 2015 Fossils, phylogenies, and the challenge
395 of preserving evolutionary history in the face of anthropogenic extinctions.
396 *Proceedings of the National Academy of Sciences* **112**, 4909-4914.
397 (doi:10.1073/pnas.1409886112).
- 398 [19] Lee, M.S.Y., Cau, A., Naish, D. & Dyke, G.J. 2014 Morphological clocks in
399 paleontology, and a mid-Cretaceous origin of crown Aves. *Systematic Biology* **63**,
400 442-449. (doi:10.1093/sysbio/syt110).
- 401 [20] Lee, M.S.Y., Cau, A., Naish, D. & Dyke, G.J. 2014 Sustained miniaturization and
402 anatomical innovation in the dinosaurian ancestors of birds. *Science* **345**, 562-
403 566. (doi:10.1126/science.1252243).
- 404 [21] Marx, F.G. & Fordyce, R.E. 2015 Baleen boom and bust: a synthesis of mysticete
405 phylogeny, diversity and disparity. *Royal Society Open Science* **2**.
- 406 [22] Sharma, P.P. & Giribet, G. 2014 A revised dated phylogeny of the arachnid order
407 Opiliones. *Frontiers in Genetics* **5**, 255. (doi:10.3389/fgene.2014.00255).
- 408 [23] Slater, G.J. 2013 Phylogenetic evidence for a shift in the mode of mammalian
409 body size evolution at the Cretaceous-Palaeogene boundary. *Methods in Ecology*
410 *and Evolution* **4**, 734-744. (doi:10.1111/2041-210X.12084).
- 411 [24] Slater, G.J. 2015 Iterative adaptive radiations of fossil canids show no evidence
412 for diversity-dependent trait evolution. *Proceedings of the National Academy of*
413 *Sciences* **112**, 4897-4902. (doi:10.1073/pnas.1403666111).
- 414 [25] Drummond, A.J. & Stadler, T. 2016 Bayesian phylogenetic estimation of fossil
415 ages. *arXiv*, 1-23. (doi:arXiv:1601.07447v1).

- 416 [26] Matzke, N.J. 2016 The evolution of antievolution policies after *Kitzmiller versus*
417 *Dover*. *Science* **351**, 28-30. (doi:10.1126/science.aad4057).
- 418 [27] Wood, H.M., Gillespie, R.G., Griswold, C.E. & Wainwright, P.C. 2015 Why is
419 Madagascar special? The extraordinarily slow evolution of pelican spiders
420 (Araneae, Archaeidae). *Evolution* **69**, 462-481. (doi:10.1111/evo.12578).
- 421 [28] Puttick, M.N., Thomas, G.H. & Benton, M.J. 2016 Dating placentalia:
422 morphological clocks fail to close the molecular fossil gap. *Evolution*.
423 (doi:10.1111/evo.12907).
- 424 [29] Tseng, Z.J., Wang, X., Slater, G.J., Takeuchi, G.T., Li, Q., Liu, J. & Xie, G. 2013
425 Himalayan fossils of the oldest known pantherine establish ancient origin of big
426 cats. *Proceedings of the Royal Society of London B: Biological Sciences* **281**.
- 427 [30] Wang, X.T., Richard H. 2008 *Dogs: their fossil relatives and evolutionary history*.
428 New York, Columbia University Press.
- 429 [31] Wang, X. 1994 Phylogenetic systematics of the Hesperocyoninae (Carnivora,
430 Canidae). *Bulletin of the American Museum of Natural History* **221**, 1-207.
- 431 [32] Wang, X.T., Richard H.; Taylor, Beryl E. 1999 Phylogenetic systematics of the
432 Borophaginae. *Bulletin of the American Museum of Natural History* **243**, 1-391.
- 433 [33] Tedford, R.H.W., Xiaoming; Taylor, Beryl E. 2009 Phylogenetic systematics of
434 the North American fossil Caninae (Carnivora, Canidae). *Bulletin of the American*
435 *Museum of Natural History* **325**, 1-218.
- 436 [34] Finarelli, J.A. & Flynn, J.J. 2006 Ancestral state reconstruction of body size in the
437 Caniformia (Carnivora, Mammalia): the effects of incorporating data from the

- 438 fossil record. *Systematic Biology* **55**, 301-313.
439 (doi:10.1080/10635150500541698).
- 440 [35] Jablonski, D. & Shubin, N.H. 2015 The future of the fossil record: Paleontology
441 in the 21st century. *Proceedings of the National Academy of Sciences* **112**, 4852-
442 4858. (doi:10.1073/pnas.1505146112).
- 443 [36] Slater, G.J. 2015. Data from: Iterative adaptive radiations of fossil canids show
444 no evidence for diversity-dependent trait evolution. Dryad. Accessed May 1,
445 2015. <http://dx.doi.org/10.5061/dryad.9qd51>
- 446 [37] Matzke, N.J. 2015 BEASTmaster: automated conversion of NEXUS data to
447 BEAST2 XML format, for fossil tip-dating and other uses. PhyloWiki.
448 <http://phylo.wikidot.com/beastmaster>
- 449 [38] Stadler, T. 2010 Sampling-through-time in birth-death trees. *Journal of*
450 *Theoretical Biology* **267**, 396-404. (doi:10.1016/j.jtbi.2010.09.010).
451

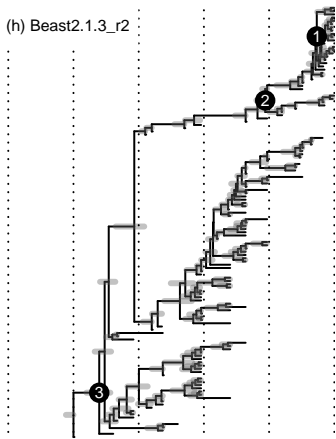
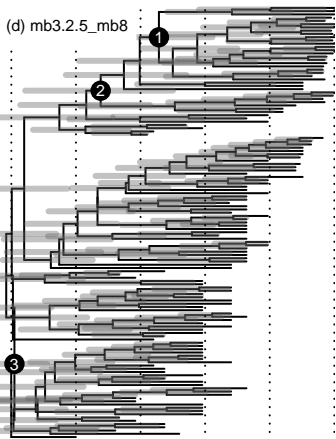
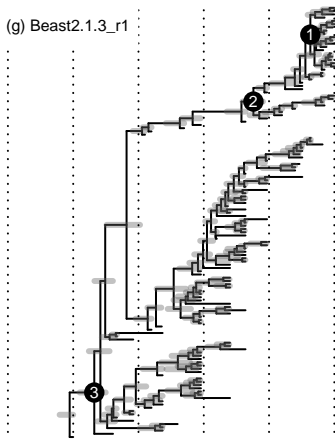
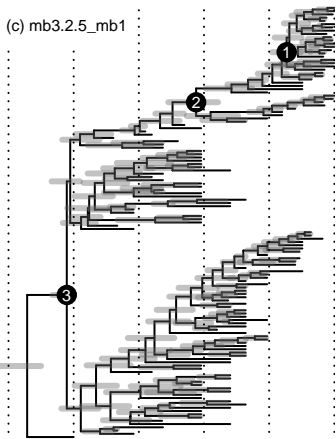
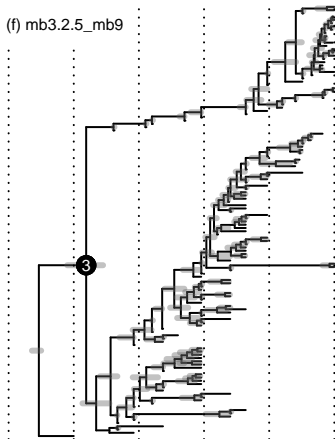
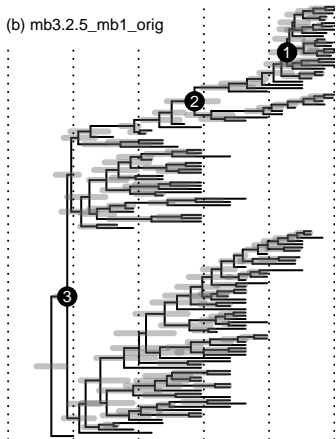
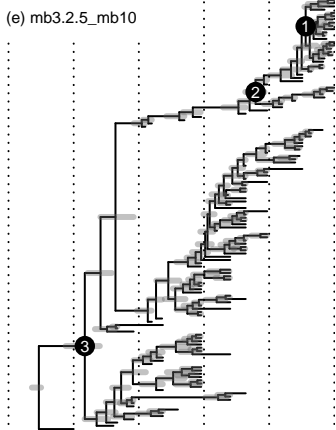
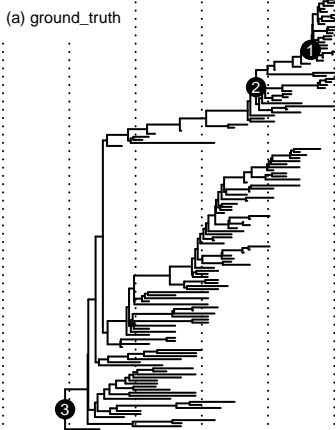


Figure 1

Ma

Ma

Table 1. Clade features that present challenges to tip-dating methods (or any dating estimation methods. Canidae exhibit few of the issues that may confound dating in other clades (e.g. angiosperms, mammals, birds).

Clade features that make tip-dating challenging	Examples	Canidae
Clade occupies widely disparate niches (habitats, body form, etc.)	angiosperms, mammals	Clade in about the same ecological niche (carnivore)
Clade spans a mass extinction and post-extinction diversification	mammals, birds	Approximately constant macroevolutionary regime
Clade has a massive worldwide radiation	angiosperms, mammals, birds	Mostly endemic to a single region (North America) for most of Canidae history
Fossils have few characters	angiosperms (pollen), bivalves	Fossils have many characters (100+)
Fossils episodic or scarce near possible clade origin	placentals, angiosperms, Cambrian arthropods	Fossils preserved continuously throughout clade history (40-0 Ma)
Possibly major changes in preservability	angiosperms (woody vs. herbaceous); Cambrian phyla (soft vs. hard parts); hominids (forest vs. savanna habitats)	Approximately constant preservability
Likely changes in molecular/ morphological rate (due to major changes in body size, population size, growth rate, etc.)	angiosperms (woody vs. herbaceous, annuals vs. perennials)	Moderate change
Available coded fossils represent only a small proportion of total known diversity	E.g. O'Leary et al. (2012) placental dataset	Coded fossil diversity greatly exceeds extant diversity

Supplemental Material for Matzke and Wright (2016), “Ground truthing tip-dating methods using fossil Canidae reveals major differences in performance”

Supplemental Methods

Ground-truth tree. The ground-truth tree was digitized using TreeRogue [1], with judgment calls resolved in favour of preserving Wang and Tedford’s depictions of divergence times. The source figures were, specifically, Figure 65 of Wang (1994) [2]; Figure 141 of Wang et al. (1999) [3]; and Figure 66 of Tedford et al. (2009) [4]. Digitization resolution was <1 my, undoubtedly more precise than either the ground-truth estimate or any Bayesian inference.

A plot of the tree, and a lineages-through-time plot, are available in Supplemental Figure 1. The Newick file is `Canidae_ground_truth.newick` (Supplemental Data).

Data. Morphological characters and dates came from the published matrix of Slater (2015) [5], specifically the Dryad repository [6] containing a NEXUS file with both morphology and MrBayes commands. Slater’s matrix synthesized and updated the matrices published in the monographs by Wang and Tedford.

The data (characters and tip-dates) were left unchanged, except that no non-North American species were removed, and taxon “outgroup” was removed. Slater used last-occurrence dates for the tip-date of each species; for the purposes of tip-dating, this decision might be suboptimal, because a specimen bearing characters may sample from anywhere in a species’ time-range. However, the nature of OTUs in tip-dating analyses is a complex question not yet addressed in the literature (Matzke and Irmis, this volume). Therefore, Slater’s tip dates were retained for purposes of simplicity and direct comparability. Slater also used extensive node calibrations to represent the stratigraphic first occurrences of many taxa; these are reasonable given Slater’s goal (fitting models of trait evolution), but node calibrations obscure the differences between tip-dating methods and so they were deleted from most analyses here.

MrBayes analyses. The “fossilized birth-death” (FBD) tree prior available in MrBayes starting with version 3.2.2 is closely analogous to the Birth-Death-Serial-Sampling (BDSS) tree prior available in Beast2, so we use BDSS to describe these runs. In MrBayes 3.2.5, sampling ancestors in BDSS became available

The 6 focal analyses presented in the main text are mb1 (Slater’s original uniform node age prior analysis including node date calibrations, with some corrections), mb8 (uniform node age prior, no node dates, flat priors on clock parameters, uniform(45,100) prior on the root age), mb9 (mb8 but with SABD tree prior and flat priors on speciation, extinction, and sampling rate), and mb10 (mb9 but BDSS, i.e. disallowing sampled ancestors via the command “prset samplestrat = fossiltips;”).

These focal analyses were selected out of a much larger collection of analyses (40 total) that were run while experimenting with modifications of the original

Slater (2015) NEXUS file. A summary of the variant inputs, and the results, is presented in Supplemental Table S2. Apart from the issues surrounding the uniform node age prior versus birth-death tree priors, which are the topic of the main text, Appendix 1 identifies other issues in the Slater (2015) NEXUS file, and the MrBayes implementation and documentation.

All MrBayes analyses ran for 5 million generations (sampling every 2500), with 2 runs, 4 chains, and default temperatures. The only exceptions were 3 analyses run with 4 runs and varying temperatures, aimed at improving the unsatisfactory topological result of the original mb9 run (see below). Typically this was sufficient for convergence as assessed in Tracer plots and MrBayes output statistics. For a few non-focal runs (Supplemental Table S2) it was not. Sometimes this was due to improper settings, and sometimes likely due to more fundamental issue with MrBayes's tree-searching abilities (notably in the case of BDSS analyses). As the purpose of many of the runs was to determine negative effects of certain versions, models, or settings (i.e. including problems with convergence), no effort was made to improve sampling further. The Perl scripts `burntrees` and `catmb` [7] were used to extract the last 50% of each tree sample and convert to dated trees.

Attempting to improve MrBayes SABD inference. The topological result of the original mb9 SABD run was unsatisfactory in that wild dogs () and the dhole () came out as a clade within the extinct Borophaginae. It was suggested that adding more runs and varying temperatures might help. We attempted this by changing the number of runs from the default 2 to 4, and then running three analyses with different temperatures: 9a (4 runs, temperature=0.05), 9b (4 runs, temperature=0.5, the default), and 9c (4 runs, temperature=1). These runs are numbered 36a, 36b, and 36c in Supplemental Table S2.

Beast2 analyses. BEASTmaster was used to construct the XML files for two Beast2 tip-dating analyses; the R scripts and Excel settings files are available in SM. The first analysis (r1) used a BDSS tree prior [8]; the second (r2) used SABD [9, 10]. Flat priors were used for each major parameter (mean and SD of the lognormal relaxed clock; and birth, death, and serial sampling rates). Rho (proportion of living species sampled) was fixed to 1, as it is not statistically identifiable if left free [11]. All runs in both programs used a single morphology partition, with an *Mkv* model correcting for the ascertainment bias against invariant characters [12, 13] and gamma-distributed rate variation with 4 rate categories.

The Beast2 analyses were run for 50 million generations (sampling every 25000). For all runs, TreeAnnotator was used to choose the Maximum Clade Credibility (MCC) tree and calculate node-date HPDs (95% highest posterior densities) and bipartition posterior probabilities (PP). Burntrees [7] was used to process MrBayes outputs for input into TreeAnnotator.

BEASTmaster [14] and custom R scripts (Supplemental Data) were used to plot all MrBayes and Beast2 MCC trees, as well as the trace plots for all key parameters (SI), and to extract parameter and node-date estimates of interest.

To assess the overall estimate of topology in each analysis, the topological symmetric distance (treedist function; phangorn R package; [15]) between the dated MCC tree and the MCC tree from the undated (mb2) analysis was calculated and compared to the distribution of distances between trees in the mb2 post-burnin treecloud [16].

For the six focal analyses, an additional test was run to see how well the methods could predict selected tip dates [17]. Four tips were selected from across the tree (*Canis ferax*, 3.5 Ma; *Epiacyon haydeni*, 5.3 Ma; *Leptocyon gregorii*, 23 Ma; *Hesperocyon gregarious*, 30.8 Ma). For each tip and focal analysis, the settings file was modified to change the tip date to a uniform(0,100) prior. After the MCMC run, the sampled tip date was extracted from the post-burnin tree sample and plotted as a histogram.

Supplemental Results

Rate parameters. Estimates of rate parameters in the focal analyses are consistent with the dating results, in that analyses with the youngest node age estimates have the highest clock, speciation, and sampling rates. The parameter describing the mean of relaxed clock branchwise rate variation (IGRvar for MrBayes, clockSD for Beast2) was inferred with similar precision across all analyses, despite uninformative priors, suggesting no special effort is needed to determine the prior for clock relaxation (clock models with autocorrelated rates may be different; [18]). The uncorrected Slater analysis does show the effect of the IGRvar prior used there (the intention was a diffuse prior, but the setting used forced a close-to-strict clock instead; see Appendix 1); however, the downstream effect on the analysis was minimal.

Summary results of all 40 variant analyses are presented in Supplemental Table S2 (supplemental Excel file). Trace plots of key variables for all 40 analyses are available in Supplemental Data (file *Canidae_traceLogs.pdf*). Plots of the MCC trees for all 40 analyses are also available (*Canidae_treeLogs.pdf*).

MrBayes SABD topology and convergence issues. The disagreement between the focal MrBayes BDSS and SABD (m10 and m9) analyses about the position of the wild dogs (*Lycaon pictus*) and dhole (*Cuon javanicus*) is puzzling, because both analyses place this clade with a sister with posterior probability 1.0; they just disagree on whether that sister is *Xenocyon texanus* (in Caninae) or *Desmocyon thomsoni* (in the extinct Borophaginae). It is true that in undated analyses (mb2), the clade has a relatively long morphological branch, and this may make it more difficult to place; however, the fact that Beast2 analyses under both BDSS and SABD place the clade unambiguously with *Xenocyon* suggest the primary issue may lie with MrBayes's implementation of the MCMC search for SABD. Design of the MCMC operators used to search tree and parameter space is very complex, and sampling ancestors adds another layer of complexity, e.g. because the tree may be changing dimensionality as fossils switch from being side-branches to direct ancestors. Bayesian estimation of direct ancestry is a very new (and potentially revolutionary) development in paleontology and phylogenetics.

Variant MrBayes mb9 analyses with 4 runs and altered temperatures did improve the situation for MrBayes SABD in some respects. Both mb9a and mb9b (4 runs each, temperatures 0.05 and the default 0.5, respectively) did retrieve *Lycaon+Cuon* inside the Caninae. The analysis mb9c (temperature 1) did not, again placing this clade in the Borophaginae. However, even the more successful MrBayes SABD analyses still exhibited difficulties (Supplemental Table S2). Convergence failed for mb9a and mb9c, with the standard deviation of split frequencies never declining below 0.1 (0.17 and 0.19, respectively). For mb9b, this metric was better, but still marginal (0.05). For comparison, mb10 (MrBayes BDSS) achieved 0.026 in the same number of generations.

In addition, the percentage Robinson-Foulds topological difference from the undated MrBayes analysis (mb2) also indicated issues. The average within-mb2 difference was 20.8%. The average difference between r1 and mb2 was 36.2%, and between r2 and mb2 it was 33.8%. For mb10, the difference was 33.1%. For mb9, mb9a, mb9b, and mb9c, by contrast, the differences were 56.2%, 43.8%, 41.5%, and 56.9%, respectively.

It is encouraging that the Beast2 implementation inferred the conventional relationship for *Lycaon+Cuon*, and also that it estimated several *Leptocyon* OTUs as direct ancestors of Caninae, confirming the conclusions of Tedford et al. (2009).

Supplemental References

- [1] Matzke, N.J. 2013 TreeRogue: R code for digitizing trees. <https://stat.ethz.ch/pipermail/r-sig-phylo/2010-October/000816.html>
- [2] Wang, X. 1994 Phylogenetic systematics of the Hesperocyoninae (Carnivora, Canidae). *Bulletin of the American Museum of Natural History* **221**, 1-207.
- [3] Wang, X.T., Richard H.; Taylor, Beryl E. 1999 Phylogenetic systematics of the Borophaginae. *Bulletin of the American Museum of Natural History* **243**, 1-391.
- [4] Tedford, R.H.W., Xiaoming; Taylor, Beryl E. 2009 Phylogenetic systematics of the North American fossil Caninae (Carnivora, Canidae). *Bulletin of the American Museum of Natural History* **325**, 1-218.
- [5] Slater, G.J. 2015 Iterative adaptive radiations of fossil canids show no evidence for diversity-dependent trait evolution. *Proceedings of the National Academy of Sciences* **112**, 4897-4902. (doi:10.1073/pnas.1403666111).
- [6] Slater, G.J. 2015. Data from: Iterative adaptive radiations of fossil canids show no evidence for diversity-dependent trait evolution. Dryad. Accessed May 1, 2015. <http://dx.doi.org/10.5061/dryad.9qd51>
- [7] Nylander, J.A.A. 2014. Burntrees v.0.2.2.: Perl script for manipulating MrBayes tree and parameter files. Accessed May 1, 2015. <https://github.com/nylander/Burntrees>
- [8] Stadler, T. 2010 Sampling-through-time in birth-death trees. *Journal of Theoretical Biology* **267**, 396-404. (doi:10.1016/j.jtbi.2010.09.010).

- [9] Gavryushkina, A., Welch, D., Stadler, T. & Drummond, A.J. 2014 Bayesian inference of sampled ancestor trees for epidemiology and fossil calibration. *PLoS Comput Biol* **10**, e1003919. (doi:10.1371/journal.pcbi.1003919).
- [10] Heath, T.A., Huelsenbeck, J.P. & Stadler, T. 2014 The fossilized birth–death process for coherent calibration of divergence-time estimates. *Proceedings of the National Academy of Sciences* **111**, E2957-E2966. (doi:10.1073/pnas.1319091111).
- [11] Stadler, T. & Yang, Z. 2013 Dating phylogenies with sequentially sampled tips. *Systematic Biology* **62**, 674-688. (doi:10.1093/sysbio/syt030).
- [12] Lewis, P.O. 2001 A likelihood approach to estimating phylogeny from discrete morphological character data. *Systematic Biology* **50**, 913-925. (doi:10.1080/106351501753462876).
- [13] Felsenstein, J. 1992 Phylogenies from restriction sites: a maximum-likelihood approach. *Evolution* **46**, 159-173. (doi:10.2307/2409811).
- [14] Matzke, N.J. 2015 BEASTmaster: automated conversion of NEXUS data to BEAST2 XML format, for fossil tip-dating and other uses. PhyloWiki. <http://phylo.wikidot.com/beastmaster>
- [15] Schliep, K.P. 2011 phangorn: phylogenetic analysis in R. *Bioinformatics* **27**, 592-593. (doi:10.1093/bioinformatics/btq706).
- [16] Matzke, N.J., Shih, P.M. & Kerfeld, C.A. 2014 Bayesian analysis of congruence of core genes in *Prochlorococcus* and *Synechococcus* and implications on horizontal gene transfer. *PLoS ONE* **9**, e85103. (doi:10.1371/journal.pone.0085103).
- [17] Drummond, A.J. & Stadler, T. 2016 Bayesian phylogenetic estimation of fossil ages. *arXiv*, 1-23. (doi:arXiv:1601.07447v1).
- [18] Ronquist, F., Klopfstein, S., Vilhelmsen, L., Schulmeister, S., Murray, D.L. & Rasnitsyn, A.P. 2012 A total-evidence approach to dating with fossils, applied to the early radiation of the Hymenoptera. *Systematic Biology* **61**, 973-999. (doi:10.1093/sysbio/sys058).

Table S1. Five Bayesian tip-dating runs are compared to (column 1) the conclusions of Tedford & Wang. The two Slater analyses (original, and a modification repairing some unintended issues; cols. 2-3) represent MrBayes analyses under a uniform node age prior, constrained by both tip dates and many node-date priors. The third run (col. 4) shows the effect of removing the node-age calibrations, and putting flat priors on the parameters for clock rate and variation. The fourth run (col. 5) shows the drastic effect of switching to a fossilized birth-death prior. Columns 5 and 6 shown Beast2 runs with flat clock priors and BDSS and SABD tree priors, respectively. These five runs are drawn from the 40 analyses shown in Supplemental Table 1.

Estimates	Analysis	Tedford & Wang, manually time-scaled cladogram	mb1_orig: Slater (2015) MrBayes analysis, original	mb1: Slater (2015) MrBayes analysis, corrected	mb8: MrBayes analysis, unconstrained, uniform node age prior	mb10: MrBayes analysis, unconstrained, fossilized-BDSS node age prior	mb9: MrBayes analysis, unconstrained, fossilized-SABD node age prior	r1: Beast2, unconstrained, BDSS tree prior	r2: Beast2, unconstrained, SABD tree prior	Notes
node ages	age(crown <i>Canis</i>)	3.2	7.3 (5.2,9.8)	7.4 (5.1,9.6)	27.5 (16.9,39.1)	4.4 (3.3,5.5)	37.1 (35.5,41.0)	3.7 (2.9,4.8)	2.7 (2.1,3.3)	4,6
	age(crown Caninae)	11.7	21.4 (17.9,25.8)	21.2 (17.8,25.5)	38.9 (29.0,54.0)	12.1 (10.7,13.7)	37.1 (35.5,41.0)	12.4 (10.9,14.1)	10.6 (10.3,12.0)	3,5
	age(Canidae)	40-36	41.0 (40.4,41.8)	41.0 (40.4,43.3)	49.0 (44.4,66.8)	38.3 (36.0,41.3)	37.1 (35.5,41.0)	36.8 (35.0,39.0)	36.1 (34.2,38.0)	2
	prior on root age	-	gamma(1,1) expect: 1 (0.024, 3.6)	offsetexp(45, 50) expect: 50 (45.1,63.6)	uniform(45,100)		none			
	age(root)	41.5	43.4 (41.2,45.5)	47.1 (45.0,52.0)	49.5 (45.0,67.7)	45.3 (45.0,46.4)	45.3 (45.0,46.5)	40.6 (40.0,42.1)	40.0 (40.0,41.2)	1
topology	Percent topological distance to undated MrBayes tree (mean betw. undated trees = 31.0% +/- 8.3%)		33.8%	32.3%	40.8%	33.1%	56.2%	36.2%	33.8%	
	crown Caninae monophyletic?	y	y	y	y	y	n	y	y	
	((Can., Boro.), Hesp.)?	y	n	n	y	y	n	y	y	
clock	model	-	IGR	IGR	IGR	IGR	IGR	ucl	ucl	
	prior	-	lognorm(-6,0.1) expectation: 0.0025 (0.002,0.003)		truncated normal(0.0025,0.1) expectation: 0.08 (0.003,0.23)		uniform(0,10)			
	clock rate	-	0.00309 (0.00201,0.00438)	0.0094 (0.0071,0.012)	0.0045 (0.0025,0.006)	0.019 (0.015,0.024)	0.024 (0.019,0.030)	0.038 (0.025,0.053)	0.052 (0.033,0.075)	
	variation prior	-	exp(126.887); expectation: 0.0079 (0.00019,0.029)		uniform(0.0001,200)		uniform(0,10)			
	among-branch variation parameter	-	0.00955 (0.00479,0.0154)	0.028 (0.018,0.039)	0.034 (0.021,0.046)	0.024 (0.015,0.033)	0.029 (0.018,0.043)	1.20 (0.98,1.42)	1.24 (1.03,1.47)	7
tip dates	among-site variation gamma parameter	-	1.03 (0.19, 1.92)	1.45 (0.87,2.02)	1.44 (0.86, 2.06)	1.38 (0.87,2.0)	1.335 (0.78,1.88)	1.16 (0.66,1.70)	1.14 (0.61,1.62)	
	<i>Canis ferax</i>	3.5	fixed(3.5)	4.4 (0.2,9.3)	10.4 (0.2,32.8)	2.8 (0.5,5.1)	3.9 (2.7,4.8)	2.9 (0.5,5.2)	2.3 (0.4,4.2)	8,9
	<i>Epicyon haydeni</i>	10.5-3	fixed(5.332)	3.5 (0.4,7.9)	4.7 (0.2,17.1)	4.9 (1.3,7.4)	4.1 (1.8,5.8)	5.4 (1.9,8.8)	5.0 (2.1,7.9)	
	<i>Leptocyon gregorii</i>	24.4-23	fixed(23)	23.3 (15.4,28.5)	32.3 (14.9,49.5)	21.8 (17.3,26.2)	23.6 (21.3,26.5)	20.1 (14.3,26.7)	15.4 (12,20.4)	
	<i>Hesperocyon gregarius</i>	37.2-30.8	fixed(30.8)	35.7 (28,40.5)	32.2 (11.4,43.7)	33.7 (30.4,36.2)	37 (30.9,43.0)	33.3 (30.1,35.7)	33.0 (30.3,34.8)	
tree	prior (all 3)	-	-	-	-	unif(0,10)	unif(0,10)	unif(0,10)	unif(0,10)	
	speciation	-	-	-	-	0.37 (0.041,0.79)	0.48 (0.043,0.96)	0.48 (0.29,0.73)	0.65 (0.36,1.08)	
	extinction	-	-	-	-	0.33 (0.037,0.71)	0.43 (0.39,0.48)	0.187 (0,0.53)	0.33 (0,0.90)	
	sampling	-	-	-	-	0.29 (0.033,0.63)	0.13 (0.08,0.18)	0.27 (0.16,0.38)	0.30 (0.15,0.44)	
Suppl. Table	run #	-	3	31	35	37	36	1	2	
	code	-	mb3.2.5, mb1_orig	mb3.2.5, mb1	mb3.2.5, mb8	mb3.2.5, mb10	mb3.2.5, mb9	r1	r2	

Notes

- 1 Wang & Tedford (2008) date from Fig. 6.6, Arctoidea outgroup 40 Ma; Wang & Tedford (2008), Fig. 6.6, origin of Hesperocyoninae;
- 2 36 Ma; Tedford & Wang (2008), Fig. 7.1, divergence of Hesperocyoninae and Borophaginae+Caninae
- 3 Tedford et al. (2009), Fig. 66
- 4 Tedford et al. (2009), Fig. 66
- 5 For mb9, excluding Cuon/Lyacon, crown Caninae date is 11.6 (10.3,12.8)
- 6 For mb9, excluding Cuon/Lyacon, crown *Canis* date is 5.9 (5.0,7.1)
- g The fixed() statements in column 2 are the tip ages used by Slater (2015) for these taxa. To estimate the tip date (next 4 columns), these were changed to uniform(0,100).
- 7 The branch-rate variation parameters are not equivalent between MrBayes IGR and Beast2 ucl relaxed clocks.
- 8 The time ranges in column 1 are the stratigraphic range of each species as indicated in Fig. 65 of Wang (1994; Hesperocyon), Fig. 141 of Wang et al. (1999; Epicyon), and Figure 66 of Tedford et al. (2009; Canis and Leptocyon).

Clock Rate Prior		Expected mean of clock rate prior (in expected changes/my)		Estimated clock rate		SD of estimated clock rate		Relaxed clock model		Prior on variance of branch rate variability (relaxed clock)		Expected mean of branch rate variance parameter		Estimated mean of branch rate variance parameter		SD of estimated mean of branch rate variance parameter		Relative rate setting for morphology partition		Crenelation observed in trace plots		outgroup setting		ingroup (Canidae) constrained to be monophyletic?		outgroup in correct position?		Wild dog+Dhole in correct position?		Modifications beyond 9 base models	
unif	flat	0.038	0.008	UCLD	unif	flat	1.202	0.11399	fixed	n	none	y	y	y	BEASTmasterR setup BDSS model, uniform priors, no node calibrations																
unif	flat	0.053	0.011	UCLD	unif	flat	1.236	0.11382	fixed	n	none	y	y	y	BEASTmasterR setup SABD model, uniform priors, no node calibrations																
lognorr	0.0025	0.003	8E-04	IGR	exp(12)	0.0079	0.01	0.00321	variabl	cor {3}	y	y	y	y	Slater original dating analysis; changed only to include non-North America living species																
NA	NA	NA	NA	NA	NA	NA	NA	NA	variabl	y	y	y	y	y	Slater original non-dating analysis; changed only to include non-North America living species																
NA	NA	NA	NA	NA	NA	NA	NA	NA	fixed	n	y	n	y	y	mb2, but relrate param m fixed to 1																
lognorr	0.0025	0.003	1E-03	IGR	exp(12)	0.0079	0.007	0.00246	variabl	y	y	y	y	y	mb1 but with fossilized-BD tree prior																
lognorr	0.0025	0.003	9E-04	IGR	exp(12)	0.0079	0.007	0.00266	variabl	y	y	y	y	y	mb3 but with priors on tree parameters explicitly set																
lognorr	0.0025	0.004	0.001	IGR	exp(12)	0.0079	0.005	0.00252	variabl	y	y	n	y	y	mb4 but with node calibrations removed, root date prior set to uniform(45,100)																
lognorr	0.0025	0.003	8E-04	IGR	exp(12)	0.0079	0.009	0.003	variabl	y	y	n	n?	y	mb5; node calibrations removed, root date prior set to uniform(45,100); tree prior back to uniform																
lognorr	0.0025	0.003	8E-04	IGR	uniform	flat	0.01	0.00352	variabl	y	y	n	n?	y	mb5; node calibrations removed, root date prior set to uniform(45,100), IGRprior set to igrvarpr=uniform(0.0001, 200)																
flattish	flat	0.373	0.554	IGR	uniform	flat	0.856	1.34177	variabl	y	y	n	n?	y	m7; vague prior on clockrate																
flattish	flat	0.493	0.433	IGR	uniform	flat	0.578	0.51918	variabl	y	y	n	y	y	m8; but with BDSS																
lognorr	0.0025	0.004	3E-04	IGR	exp(12)	0.0079	0.013	0.00206	variabl	cor	y	y	y	y	add treeagepr=uniform(45,50) to force non-gamma(1,1) tree age prior																
lognorr	0.0025	0.004	3E-04	IGR	exp(12)	0.0079	0.013	0.00235	fixed	n	y	y	y	y	"																
NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA															
lognorr	0.0025	0.004	4E-04	IGR	exp(12)	0.0079	0.009	0.00239	variabl	y	y	y	y	y	"																
lognorr	0.0025	0.004	4E-04	IGR	exp(12)	0.0079	0.008	0.00131	variabl	y	y	y	y	y	"																
lognorr	0.0025	0.005	3E-04	IGR	exp(12)	0.0079	0.008	0.00111	variabl	y	y	n	y	n	"																
lognorr	0.0025	0.004	3E-04	IGR	exp(12)	0.0079	0.015	0.00249	variabl	y	y	n	n	y	"																
lognorr	0.0025	0.004	3E-04	IGR	uniform	flat	0.015	0.0027	variabl	y	y	n	n	y	"																
flattish	flat	0.009	0.001	IGR	uniform	flat	0.027	0.00561	variabl	y	y	n	n	y	"																
flattish	flat	0.026	0.003	IGR	uniform	flat	0.034	0.00595	variabl	y	y	n	y	n	"																
lognorr	0.0025	0.003	3E-04	IGR	exp(12)	0.0079	0.04	0.00717	fixed	n	y	y	y	y	offsetexp(45,50) treeagepr, m fixed, ingroup monophyletic																
lognorr	0.0025	0.003	3E-04	IGR	uniform	flat	0.052	0.01145	fixed	n	y	y	y	y	"																
flattish	flat	0.006	9E-04	IGR	uniform	flat	0.067	0.015	fixed	n	y	y	y	y	"																
lognorr	0.0025	0.004	4E-04	IGR	exp(12)	0.0079	0.011	0.00262	fixed	n	y	y	y	y	unif treeagepr, m fixed, ingroup monophyletic																
lognorr	0.0025	0.003	2E-04	IGR	exp(12)	0.0079	0.047	0.00914	fixed	n	y	y	y	y	"																
lognorr	0.0025	0.003	2E-04	IGR	uniform	flat	0.063	0.01555	fixed	n	y	y	y	y	"																
flattish	flat	0.004	7E-04	IGR	uniform	flat	0.065	0.01447	fixed	n	y	y	y	y	"																
flattish	flat	0.02	0.002	IGR	uniform	flat	0.042	0.00883	fixed	n	y	y	y	y	"																
flattish	flat	0.009	0.001				0.028	0.00519						y	"																
lognorr	0.0025	0.005	3E-04	IGR	exp(12)	0.0079	0.007	0.00112	fixed	n	y	y	y	n	"																
lognorr	0.0025	0.003	2E-04	IGR	exp(12)	0.0079	0.029	0.00435	fixed	n	y	y	y	y	"																
lognorr	0.0025	0.003	3E-04	IGR	uniform	flat	0.033	0.00557	fixed	n	y	y	y	y	"																
flattish	flat	0.005	9E-04	IGR	uniform	flat	0.035	0.00653	fixed	n	y	y	y	y	"																
flattish	flat	0.024	0.003	IGR	uniform	flat	0.03	0.006	fixed	n	y	y	y	n	" prset samplestrat = random; [SABD tree prior]																
flattish	flat	0.019	0.002	IGR	uniform	flat	0.024	0.00473	fixed	n	y	y	y	y	" mb9 but prset samplestrat = fossiltips; (no sampled ancestors)																
<i>nct Borophaginae.)</i>																															
flattish	flat	0.026	0.003	IGR	uniform	flat	0.036	0.00701	fixed	n	y	y	y	y	unif treeagepr, m fixed, ingroup monophyletic; changed to 4 runs, same default temperature (0.5)																
flattish	flat	0.025	0.003	IGR	uniform	flat	0.032	0.00627	fixed	n	y	y	y	y	unif treeagepr, m fixed, ingroup monophyletic; changed to 4 runs, lower temperature (0.05)																
flattish	flat	0.028	0.004	IGR	uniform	flat	0.035	0.00709	fixed	n	y	y	y	n	unif treeagepr, m fixed, ingroup monophyletic; changed to 4 runs, higher temperature (1)																

Appendix 1: Issues with the MrBayes dating analysis of Slater (2015).

In setting up variant MrBayes analyses (Supplemental Table S2), a number of issues became apparent with the NEXUS file of the original Slater (2015) analysis. These are detailed below in order to help aid future MrBayes analyses, and in some cases to suggest improvements in the MrBayes code or documentation. These issues do not appear to greatly alter the dating results of Slater (2015), due to the large number of tip- and node-date constraints in that analysis (compare Figure 1b: mb2.3.5_mb1_orig; and Figure 1c: mb3.2.5_mb1), but they did cause major issues for analyses without node-date constraints.

The example NEXUS file being examined is canidae.nex, downloaded May 2015, and re-downloaded (unchanged) in April 2016 from: <http://datadryad.org/bitstream/handle/10255/dryad.73273/canidae.nex?sequence=1>.

A file correcting the issues identified below, but otherwise maintaining the intended analysis of Slater (2015) (uniform node age prior, node date constraints, etc.) is file “canidae_all_issues_fixed.nex”, located in directory mb_3.2.5b_add_ingroup/mb1/ of the Supplemental Data file “Matzke_Wright_SuppData.zip.”

Issue 1: Root node date calibration

The NEXUS file includes a variety of node-date calibrations, including an offsetexp(min=45, mean=50) calibration for the root node:

Line 433 of canidae.nex:

```
calibrate root=offsetexponential(45, 50); [mean = 50, median = 48.5, 95% upper = 60]
```

Unfortunately, this date prior on the root node appears to be ignored by MrBayes. This can be confirmed by inspecting Slater (2015)'s Figure S2, where the age of the root is approximately 42 Ma, despite the fact that the root node constraint has a hard minimum of 45 Ma.

The only hint that MrBayes is ignoring the root calibration is the following warning message:

```
=====  
WARNING: Constraint 'root' refers only to deleted taxa  
and will be disregarded  
=====
```

In the screen output of the MrBayes run, this warning is easy to miss, as it is hidden amongst many other warnings of this type:

=====
WARNING: There is one character incompatible with the specified
coding bias. This character will be excluded.
=====

The second warning is due to a character in the data matrix being invariant. Another reason that the first warning can be missed is that the warning is inaccurate (the taxa were not deleted).

It appears that, with the root node date calibration ignored, and with no tree age prior (treeagePr setting) given, the MrBayes dating analyses default to a tree height prior with a gamma(1,1) distribution. The message output to the screen at runtime is:

=====
Tree age has a Gamma(1.00,1.00) distribution
=====

This distribution is equivalent to an exponential(1) distribution. This suggests that the real prior being used on the age of the root is informative, and has a mean of 1 mya. This provides a ready explanation of the young date of the root node – the root node date is estimated to be just below the oldest tip (the outgroup, fixed to 40 Ma).

Issue 2: Prior on clock rate variability

The NEXUS file includes this prior for the igrvar parameter (igrvar = variance parameter for the gamma distribution on branchwise rate variability, for independent branch rates).

Line 468 of canidae.nex:

```
prset igrvarpr=exp(126.887) ; [a vague prior]
```

This parameter describes the expected variance given a branchlength in expected amounts of change. igrvar is multiplied by the each branchlength to give the expected variability.

The comment suggests the Exponential(126.887) prior as "a vague prior". We can see how a user could think this, given the language in the igrvar documentation:

MrBayes > help prset (in MrBayes 3.2.5) gives:

```
Igrvarpr -- This parameter allows you to specify a prior on the variance of the gamma distribution from which the branch lengths are drawn in the independent branch rate (IGR) relaxed clock model. Specifically, the parameter specifies the rate at
```

which the variance increases with respect to the base rate of the clock. If you have a branch of a length corresponding to 0.4 expected changes per site according to the base rate of the clock, and the igrvar parameter has a value of 2.0, then the effective branch length will be drawn from a distribution with a variance of 0.4×2.0 .

You can set the parameter to a fixed value, or specify that it is drawn from an exponential or uniform distribution:

```
prset igrvarpr = fixed(<number>)  
prset igrvarpr = exponential(<number>)  
prset igrvarpr = uniform(<number>,<number>)
```

For backward compatibility, 'ibrvarpr' is allowed as a synonym of 'igrvarpr'.

(This text is also found in commref_mb3.2.txt in the MrBayes 3.2.x download)

However, elsewhere in MrBayes, the exponential distribution is generally interpreted such that the input parameter for `exp()` is the exponential rate parameter, λ , and the expectation of the mean is $\beta = 1/\lambda$. Thus, the expectation of an Exponential(126.887) distribution is $1/126.887=0.00788$. Thus, instead of a vague prior on branchwise rate variation, this prior essentially mandates a strict clock.

Our interpretation is confirmed by examining the inference of the estimated mean of branch rate variance parameter under MrBayes runs where the igrvar parameter has been changed (Supplemental Table S2).

Issue 3. Relative rate prior (ratepr)

In the NEXUS file, the relative rate prior (ratepr) is set to "variable":

Line 458 of canidae.nex:

```
prset applyto=(all) ratepr = variable;
```

This setting creates a parameter, $m\{1\}$, representing the relative rate of the morphology partition compared to other partitions (DNA, RNA, etc.) under a common overall clock model. However, canidae.nex is a morphology-only dataset and only has 1 partition. MrBayes does not identify this situation and fix $m\{1\}$ to "fixed". Instead, it attempts to estimate this relative rate along with the clock rate and clock variability. This creates poor mixing due to non-identifiability, and "crenelations" in the MCMC trace of parameters. Page 3 (analysis mb1_orig) of the Supplemental Data file Canidae_traceLogs.pdf shows these crenelations: the MCMC trace jumps to one value, samples around that

value for a while, and then jumps to a much different value. Later in the chain, it discretely jumps towards the original value, and the cycle repeats. This behaviour leads to low ESS values and bimodal parameter estimates.

In Slater's highly-constrained original analysis, the effect on other inferences is not particularly noticeable and presumably makes little difference. However, it becomes a major issue for mixing and parameter estimation as node constraints are removed.

Issue 4. Rate prior on the morphological clock

In the NEXUS file, the relative rate prior (ratepr) is set to "variable":

Line 469 of canidae.nex:

```
prset clockratepr = lognorm(-6,0.1);
```

Slater set a tight prior on the morphology clock rate. The lognorm(-6, 0.1) distribution has a mean in real space of 0.0025 changes/my, and an SD of 0.00025. This is a user decision rather than a problem, and it is clearly mentioned in Slater (2015).

It may be, however, that the decision for a strongly informative prior on the clock rate was made in part in order to "make the analysis behave," due to problems caused by the uniform node age prior, and perhaps some of the other issues mentioned in this appendix. We note that tip-dating analyses with BDSS-type tree models function very well even with broad, uninformative priors on the rate of the morphological clock (Supplemental Table S2).

Issue 5. Outgroup, and specifying the outgroup

The outgroup taxon, named "outgroup" in Slater's analysis, is identified as the outgroup in canidae.nex:

Line 459 of canidae.nex:

```
outgroup 1;
```

Taxon 1 is the outgroup OTU. However, in MrBayes dating analyses, it appears that the outgroup setting is ignored. This highlights a fundamental difference between undated and dating analyses. In undated analyses, all trees are formally unrooted, and rooting via an outgroup can take place during or after the phylogenetic inference. Thus, in the original, non-dating versions of MrBayes, the "outgroup" option was simply a convenience for the user, unless the outgroup consisted of multiple OTUs, in which case it serves as a topology constraint.

However, in a dating analysis, all sampled trees are always rooted, whether or not the user has decided on an outgroup. Furthermore, the mechanics of specifying an outgroup are more complex. Merely declaring an OTU an outgroup, or declaring an outgroup clade to be monophyletic, will not necessarily do the job. After all, a clade that is forced to be monophyletic could still be deeply nested inside the ingroup, unless something prevents this.

The simplest way to force the outgroup to be the earliest-branching group in a dating analysis is to set up a node constraint specifying that the ingroup is monophyletic. This could be programmed into the MrBayes outgroup command, but at the time of writing, it was not. In the case of Slater (2015)'s *canidae.nex*, it happens that there is a node constraint named "Canidae" and includes all living and fossil Canidae in the analysis. This constraint is used in the original Slater analysis, so the effect of the MrBayes outgroup problem is not noticed until the node constraints are removed; in this situation, some uniform clock tip-dating analyses fail to put the outgroup in the outgroup position (Supplemental Table S2). Fossilized BD analyses seem to put the outgroup in the correct position even without any constraints (Figure 1).

Issue 6. Typos in some OTU names.

Comparison with the "ground truth" tree manually digitized from the monographs of Tedford and Wang identified several likely typos in *Canidae.nex* (in fairness, comparison also revealed a number of typos in the draft ground truth tree; these are corrected in the final version). The correct spellings were double-checked via google and comparison to the monographs.

Typo	Corrected
Cynarctoides_accridens	Cynarctoides_acridens
Phlaocyon_marshlandensis	Phlaocyon_marslandensis
Paracynarctus_sinclari	Paracynarctus_sinclairi
Rhizocyon_oreganensis	Rhizocyon_oregonensis
Cynarctoides_gawanae	Cynarctoides_gawnae
Protomarctus_opatus	Protomarctus_optatus
Urocyon_galushi	Urocyon_galushai
Urocyon_citronus	Urocyon_citrinus