Impact of metabolic heterogeneity on tumor growth, invasion, and treatment outcomes

Mark Robertson-Tessi – *PSOC Member*, Robert J. Gillies – *PSOC Member*, Robert A. Gatenby – *PSOC Member*, and Alexander R. A. Anderson – *PSOC & ICBP Member*

Abstract— A hybrid multiscale mathematical model of tumor growth is used to investigate how tumoral and microenvironmental heterogeneity affect treatment outcomes. A key component of this model is normal and tumor metabolism and its interaction with microenvironmental factors. In early stages of growth, tumors are stratified, with the most aggressive cells developing within the interior of the tumor. Simulations suggest that in some cases chemotherapy may increase the metabolic aggressiveness of a tumor due to drug-mediated selection.

I. HETEROGENEITY: AN IMPORTANT DETERMINANT OF TUMOR PROGRESSION

Tumor heterogeneity at the genetic scale has been known for decades and until recently was largely viewed as a whole tumor metric. Historically, molecular techniques average genomic signals from large numbers of cells from single biopsies, thus smoothing and potentially hiding underlying variations. However, a potential issue with this approach was recently highlighted by Swanton and colleagues [1], who showed that multiple biopsies from the same tumor display distinct genetic profiles and yet are phenotypically similar. This genotypic divergence and phenotypic convergence has previously been hinted at theoretically [2] and may be a predictable evolutionary consequence of the tumor ecosystem [3, 4]. The intricate dialogue between tumor cells and environment selects for clones that are best adapted phenotypically to survive, regardless of specific mutations that may facilitate tumor progression. Furthermore, this environment is temporally and spatially heterogeneous largely due to variations in blood flow, resulting in local fluctuations of nutrients, growth factors and other cellular populations (e.g. normal cells, stromal cells and immune cells). These dynamics occurring within the cancer ecosystem are almost impossible to dissect via experimentation alone.

A serious effort is being put into quantifying this heterogeneity and understanding how it evolves as the tumor progresses and how it relates to overall outcome [5-7]. However, we are far from understanding how the microenvironment modulates this heterogeneity and drives the overall phenotypic behavior of the tumor cell population. Furthermore, the impact of heterogeneity on treatment outcomes remains poorly understood.

Research supported by the Moffitt Cancer Center PSOC, NIH/NCI U54CA143970.

M. Robertson-Tessi (813-745-6818; e-mail: mark.robertsontessi@ moffitt.org), R. J. Gillies (email: robert.gillies@moffitt.org), R. A. Gatenby (email: robert.gatenby@moffitt.org) and A. R. A. Anderson (alexander.anderson@moffitt.org) are with the Moffitt Cancer Center, Tampa, FL 33612 USA.

II. THERAPY RAPIDLY ALTERS THE SELECTION PRESSURES

Using a two-dimensional hybrid cellular automaton model, we have grown tumors in a vascularized tissue and applied therapy to them [8]. The heterogeneity was investigated on two metabolic axes: glycolytic capacity and resistance to extracellular acidosis. Absent treatment, a subset of tumor cells evolve under environmental selection pressure to become glycolytic and acid resistant. These properties cause increased tumor invasiveness under the right environmental conditions. Figure 1 shows the results of applying a chemotherapeutic agent in the model; in panels (a-f) the simulated tumor is shown, where the left column is the untreated case, the central column is an early application, and the right column is a late application. Panel (g) shows the growth curves for the three cases. The color of the tumor cells corresponds to the phenotype, and in all cases a heterogeneous mix is observed. Green tumor cells are metabolically normal, while purple cells are more aggressively glycolytic and acid-resistant. A segregated spatial structure is seen in the simulations; the aggressive cells develop in the center of the tumor, where hypoxia is more likely to occur due to dysfunctional vasculature.

This spatial heterogeneity has a profound impact on how the tumor responds to therapies given only months apart. The early application, in the central column, has the effect of delaying growth on the order of a year or two, compared to controls. On the other hand, applying the same therapy regimen just a few months later results in an acceleration of tumor growth, meaning that it would have been better to not treat at all. This occurs because the heterogeneity of the tumor at the time of the later treatment is poised for invasion, but not yet invasive. The therapy selects for these invasive cells, both spatially and temporally, and thus allows them to become actively invasive. This is compared to the early treatment case, where the invasive cells were not fully developed; the early treatment regimen has the effect of selecting for cells which are more resistant to invasion, due to their acid-resistant but low-glycolytic phenotype. In our publication [8], we have examined a number of therapies, showing how tumor heterogeneity has an impact on how therapies either succeed or fail.

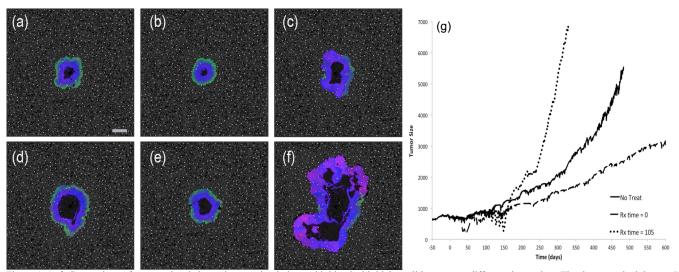


Figure 1. (a-f) Comparison of untreated and chemotherapy simulations with identical initial conditions at two different time points. The therapy schedule was 5 pulses, 2 weeks apart. The left column is an untreated simulation; the central column was pulsed with cytotoxic therapy starting at t_{rel}=0; the right column starts the identical treatment at t_{rel}=105. The top panels (a-c) show the state of the three simulations at t_{rel}=264, i.e. shortly after the tumor in the right column has finished the therapy. The bottom set of panels (d - f) shows the state of the tumors at t_{rel}=380. Scale bar is 400 microns. (g) Growth curves for untreated (solid), early (dashed), and late (dotted) chemotherapy from (a-f). Tumor size on the vertical axis is the diameter in microns.

III. QUICK GUIDE TO THE METHODS

A. Equations: The concentration of a molecule (C(x))across a tissue is described by

$$\frac{\partial c}{\partial t} = D\nabla^2 C + f(\mathbf{C}, \mathbf{p}),\tag{1}$$

with diffusion constant D, and f describing the production and consumption of the molecule depending on the concentrations of extracellular molecules (C(x)) and cellular parameters $(\mathbf{p}(x))$ at position x. Cells primarily produce ATP from glucose (G), using either an efficient aerobic pathway that requires oxygen (O), or using glycolysis, an inefficient anaerobic pathway that produces protons (H). The model assumes that cells meet a target level of ATP demand by preferentially using the aerobic pathway, and making up the difference by increasing flux through the glycolytic pathway in hypoxic regions. Oxygen consumption (f_O) and glucose consumption (f_G) are determined by the need to meet normal

ATP demand (A₀), given by
$$f_O = -V_O \frac{o}{o + k_O}, \ f_G = -\left(\frac{p_G A_O}{2} + \frac{27f_O}{10}\right) \frac{G}{G + k_G}$$
 (2) For tumor cells, if the coefficient $p_G > 1$, the tumor will

consume more glucose than needed to meet normal ATP demand, representing constitutively activated consumption seen in many tumors. The actual ATP production rate for the cell (f_A) is determined from nutrient consumption rates, given by

$$f_A = -\left(2f_G + \frac{27f_O}{5}\right). \tag{4}$$

 $f_A = -\left(2f_G + \frac{27f_O}{5}\right). \tag{4}$ Proton production (f_H) is linked to the amount of glycolysis that does not feed the aerobic pathway, given by $f_H = k_H \frac{p_G V_o + f_O}{5}$, where parameter k_H accounts for proton buffering.

$$f_H = k_H \frac{p_G V_O + f_O}{\epsilon},\tag{5}$$

This metabolic program is implemented into each cell of a hybrid cellular automaton (HCA) model. One cell type is permitted per grid point, either a normal cell, tumor cell, necrotic cell, or blood vessel. For each time step dt, Eq. (1) is solved over the domain of the HCA for the steady state. Then, cells in the grid are put through a decision process based on the metabolic state of each cell. Cells with enough

ATP to meet the threshold of proliferation will advance their cell cycle. Cells that have completed the cell cycle will proliferate if there is adjacent space. The cycle is not advanced if the cell is quiescent due to lowered ATP production. Cells with production less than a death threshold are removed.

Tumor cells have two heritable traits: excess glucose consumption, p_G from Eq. 2; and resistance to extracellular acidosis. These traits are passed from a parent cell to its daughters modified by a small equally-weighted variation, variation chosen at random. The model is agnostic with respect to specific biological mechanisms that underlie this drift, which could include gradual accumulation of mutations, regulation of gene transcription by epigenetics or aneuploidy, or changes in the number or structure of organelles.

A point-source vasculature is used to simulate blood vessels that spatiotemporally deliver nutrients and remove waste products. The field of vessels is seeded using a circlepacking algorithm based on vessel densities in vivo. This initial distribution can be altered by the creation of new vessels through angiogenesis, or by vessel degradation. For angiogenesis, new vessels are added to regions of hypoxia until there is enough oxygen delivery to remove the hypoxic state. Vessels are degraded over time due to surrounding tumor growth until they are lost from the tissue. These two opposing vascular forces impact the gradients of diffusible molecules.

Chemotherapy is pulsed through the vasculature, diffusing through the tissue subject to Eq. (1). Cell death depends on the concentration of the drug at the cell position.

B. Type of settings in which these methods are useful

HCA models have been used extensively to model cancer [9], including applications to angiogenesis [10-13], cell motility and invasion [14-18], tumor evolution and microenvironment [19-35], tumor-immune interactions [36], and metastasis [37, 38].

[22]

REFERENCES

- [1] M. Gerlinger, A. J. Rowan, S. Horswell, J. Larkin, D. Endesfelder, E. Gronroos, et al., "Intratumor heterogeneity and branched evolution revealed by multiregion sequencing," N Engl J Med, vol. 366, pp. 883-92, Mar 8 2012.
- [2] P. Gerlee and A. R. Anderson, "Modelling evolutionary cell behaviour using neural networks: application to tumour growth," *Biosystems*, vol. 95, pp. 166-74, Feb 2009.
- [3] K. J. Pienta, N. McGregor, R. Axelrod, and D. E. Axelrod, "Ecological therapy for cancer: defining tumors using an ecosystem paradigm suggests new opportunities for novel cancer treatments," *Transl Oncol*, vol. 1, pp. 158-64, Dec 2008.
- [4] D. Basanta and A. R. Anderson, "Exploiting ecological principles to better understand cancer progression and treatment," *Interface Focus*, vol. 3, p. 20130020, Aug 6 2013.
- [5] M. Greaves and C. C. Maley, "Clonal evolution in cancer," Nature, vol. 481, pp. 306-13, Jan 19 2012.
- [6] M. S. Lawrence, P. Stojanov, P. Polak, G. V. Kryukov, K. Cibulskis, A. Sivachenko, et al., "Mutational heterogeneity in cancer and the search for new cancer-associated genes," *Nature*, vol. 499, pp. 214-8, Jul 11 2013.
- [7] A. Sottoriva, I. Spiteri, S. G. Piccirillo, A. Touloumis, V. P. Collins, J. C. Marioni, et al., "Intratumor heterogeneity in human glioblastoma reflects cancer evolutionary dynamics," Proc Natl Acad Sci USA, vol. 110, pp. 4009-14, Mar 5 2013.
- [8] M. Robertson-Tessi, R. J. Gillies, R. A. Gatenby, and A. R. Anderson, "Impact of metabolic heterogeneity on tumor growth, invasion, and treatment outcomes," *Cancer Res*, vol. 75, pp. 1567-79, Apr 15 2015.
- [9] K. A. Rejniak and A. R. Anderson, "Hybrid models of tumor growth," Wiley Interdiscip Rev Syst Biol Med, vol. 3, pp. 115-25, Jan-Feb 2011.
- [10] A. R. Anderson and M. A. Chaplain, "Continuous and discrete mathematical models of tumor-induced angiogenesis," *Bull Math Biol*, vol. 60, pp. 857-99, Sep 1998.
- [11] J. L. Gevertz and S. Torquato, "Modeling the effects of vasculature evolution on early brain tumor growth," *J Theor Biol*, vol. 243, pp. 517-31, Dec 21 2006.
- [12] A. L. Bauer, T. L. Jackson, and Y. Jiang, "A cell-based model exhibiting branching and anastomosis during tumor-induced angiogenesis," *Biophys J*, vol. 92, pp. 3105-21, May 1 2007.
- [13] M. R. Owen, T. Alarcon, P. K. Maini, and H. M. Byrne, "Angiogenesis and vascular remodelling in normal and cancerous tissues," *J Math Biol*, vol. 58, pp. 689-721, Apr 2009.
- [14] A. R. Anderson, "A hybrid mathematical model of solid tumour invasion: the importance of cell adhesion," *Math Med Biol*, vol. 22, pp. 163-86, Jun 2005.
- [15] H. B. Frieboes, X. Zheng, C. H. Sun, B. Tromberg, R. Gatenby, and V. Cristini, "An integrated computational/experimental model of tumor invasion," *Cancer Res*, vol. 66, pp. 1597-604, Feb 1 2006.
- [16] M. Aubert, M. Badoual, and B. Grammaticos, "A model for short- and long-range interactions of migrating tumour cell," *Acta Biotheor*, vol. 56, pp. 297-314, Dec 2008.
- [17] P. Gerlee and A. R. Anderson, "Evolution of cell motility in an individual-based model of tumour growth," *J Theor Biol*, vol. 259, pp. 67-83, Jul 7 2009.
- [18] H. Hatzikirou, D. Basanta, M. Simon, K. Schaller, and A. Deutsch, "'Go or grow': the key to the emergence of invasion in tumour progression?," *Math Med Biol*, vol. 29, pp. 49-65, Mar 2012.
- [19] A. R. Kansal, S. Torquato, E. A. Chiocca, and T. S. Deisboeck, "Emergence of a subpopulation in a computational model of tumor growth," *J Theor Biol*, vol. 207, pp. 431-41, Dec 7 2000.
- [20] S. L. Spencer, R. A. Gerety, K. J. Pienta, and S. Forrest, "Modeling somatic evolution in tumorigenesis," *PLoS Comput Biol*, vol. 2, p. e108, Aug 18 2006.
- [21] A. R. Anderson, A. M. Weaver, P. T. Cummings, and V. Quaranta, "Tumor morphology and phenotypic evolution driven by selective pressure from the microenvironment," *Cell*, vol. 127, pp. 905-15, Dec 1 2006.

- K. Smallbone, R. A. Gatenby, R. J. Gillies, P. K. Maini, and D. J. Gavaghan, "Metabolic changes during carcinogenesis: potential impact on invasiveness," *J Theor Biol*, vol. 244, pp. 703-13, Feb 21 2007.
- [23] A. Bankhead, 3rd, N. S. Magnuson, and R. B. Heckendorn, "Cellular automaton simulation examining progenitor hierarchy structure effects on mammary ductal carcinoma in situ," *J Theor Biol*, vol. 246, pp. 491-8, Jun 7 2007.
- [24] P. Gerlee and A. R. Anderson, "A hybrid cellular automaton model of clonal evolution in cancer: the emergence of the glycolytic phenotype," *J Theor Biol*, vol. 250, pp. 705-22, Feb 21 2008
- [25] J. A. Engelberg, G. E. Ropella, and C. A. Hunt, "Essential operating principles for tumor spheroid growth," *BMC Syst Biol*, vol. 2, p. 110, 2008.
- [26] D. Basanta, D. W. Strand, R. B. Lukner, O. E. Franco, D. E. Cliffel, G. E. Ayala, et al., "The role of transforming growth factor-beta-mediated tumor-stroma interactions in prostate cancer progression: an integrative approach," Cancer Res, vol. 69, pp. 7111-20, Sep 1 2009.
- [27] S. H. Kim, J. Debnath, K. Mostov, S. Park, and C. A. Hunt, "A computational approach to resolve cell level contributions to early glandular epithelial cancer progression," *BMC Syst Biol*, vol. 3, p. 122, 2009.
- [28] P. Gerlee and A. R. Anderson, "Diffusion-limited tumour growth: simulations and analysis," *Math Biosci Eng*, vol. 7, pp. 385-400, Apr 2010.
- [29] A. S. Silva, R. A. Gatenby, R. J. Gillies, and J. A. Yunes, "A quantitative theoretical model for the development of malignancy in ductal carcinoma in situ," *J Theor Biol*, vol. 262, pp. 601-13, Feb 21 2010.
- [30] D. Basanta, B. Ribba, E. Watkin, B. You, and A. Deutsch, "Computational analysis of the influence of the microenvironment on carcinogenesis," *Math Biosci*, vol. 229, pp. 22-9, Jan 2011.
- [31] G. G. Powathil, K. E. Gordon, L. A. Hill, and M. A. Chaplain, "Modelling the effects of cell-cycle heterogeneity on the response of a solid tumour to chemotherapy: biological insights from a hybrid multiscale cellular automaton model," *J Theor Biol*, vol. 308, pp. 1-19, Sep 7 2012.
- [32] E. Kim, V. Rebecca, I. V. Fedorenko, J. L. Messina, R. Mathew, S. S. Maria-Engler, et al., "Senescent fibroblasts in melanoma initiation and progression: an integrated theoretical, experimental, and clinical approach," Cancer Res, vol. 73, pp. 6874-85, Dec 1 2013.
- [33] C. DuBois, J. Farnham, E. Aaron, and A. Radunskaya, "A multiple time-scale computational model of a tumor and its micro environment," *Math Biosci Eng*, vol. 10, pp. 121-50, Feb 2013.
- [34] J. G. Scott, A. B. Hjelmeland, P. Chinnaiyan, A. R. Anderson, and D. Basanta, "Microenvironmental variables must influence intrinsic phenotypic parameters of cancer stem cells to affect tumourigenicity," *PLoS Comput Biol*, vol. 10, p. e1003433, Jan 2014
- [35] D. Chen, Y. Jiao, and S. Torquato, "A cellular automaton model for tumor dormancy: emergence of a proliferative switch," *PLoS One*, vol. 9, p. e109934, 2014.
- [36] D. G. Mallet and L. G. De Pillis, "A cellular automata model of tumor-immune system interactions," *J Theor Biol*, vol. 239, pp. 334-50, Apr 7 2006.
- [37] H. Enderling, L. Hlatky, and P. Hahnfeldt, "Migration rules: tumours are conglomerates of self-metastases," *Br J Cancer*, vol. 100, pp. 1917-25, Jun 16 2009.
- [38] A. Araujo, L. M. Cook, C. C. Lynch, and D. Basanta, "An integrated computational model of the bone microenvironment in bone-metastatic prostate cancer," *Cancer Res*, vol. 74, pp. 2391-401, May 1 2014.