

1    **Estimating the effect of competition on trait evolution using maximum likelihood inference**

2    Jonathan Drury<sup>1\*</sup>, Julien Clavel<sup>1</sup>, & Hélène Morlon<sup>1</sup>

3    <sup>1</sup>*Département de Biologie, École Normale Supérieure, 46 rue d'Ulm, 75005 Paris, France*

4    \*Correspondence: [drury@biologie.ens.fr](mailto:drury@biologie.ens.fr), +33 (7) 83 11 43 21

5    Running head: INTERSPECIFIC COMPETITION & TRAIT EVOLUTION

6

## Abstract

Many classical ecological and evolutionary theoretical frameworks posit that competition between species is an important selective force. For example, in adaptive radiations, resource competition between evolving lineages plays a role in driving phenotypic diversification and exploration of novel ecological space. Nevertheless, current models of trait evolution fit to phylogenies and comparative datasets are not well designed for incorporating the effect of competition. The most advanced models in this direction are diversity-dependent models where evolutionary rates depend on lineage diversity. However, these models still treat changes in traits in one branch as independent of the value of traits on other branches, thus ignoring the effect of species similarity on trait evolution. Here, we consider a model where the evolutionary dynamics of traits involved in interspecific interactions are influenced by species similarity in trait values and where we can specify which lineages are in sympatry. We develop a maximum-likelihood based approach to fit this model to combined phylogenetic and phenotypic data. Using simulations, we demonstrate that the approach accurately estimates the simulated parameter values across a broad range of parameter space. Additionally, we develop tools for specifying the biogeographic context in which trait evolution occurs. In order to compare models, we also apply these biogeographic methods to specify which lineages interact sympatrically for two diversity-dependent models. Finally, we fit these various models to morphological data from a classical adaptive radiation (Greater Antillean *Anolis* lizards). We show that models that account for competition and geography perform better than other models. Given the importance of interspecific interactions, in particular competition, in many ecological and evolutionary processes, this model is an important new tool for using comparative datasets to study phenotypic evolution.

*Keywords:* interspecific competition, phylogenetic comparative methods, adaptive radiation, trait evolution, *Anolis*, maximum likelihood

Interactions between species can be strong selective forces. Indeed, many classical evolutionary theories assume that interspecific competition has large impacts on fitness. Character displacement theory (Brown and Wilson 1956; Grant 1972; Pfennig and Pfennig 2009), for example, posits that interactions between species, whether in ecological or social contexts, drive adaptive changes in phenotypes. Similarly, adaptive radiation theory has been a popular focus of investigators interested in explaining the rapid evolution of phenotypic disparity (Grant and Grant 2002; Losos 2009; Mahler et al. 2013; Weir and Mursleen 2013), and competitive interactions between species in a diversifying clade are a fundamental component of adaptive radiations (Schluter 2000; Losos and Ricklefs 2009; Grant and Grant 2011).

Additionally, social interactions between species, whether in reproductive (Gröning and Hochkirch 2008; Pfennig and Pfennig 2009) or agonistic (Grether et al. 2009, 2013) contexts, are important drivers of changes in signal traits used in social interactions. Several evolutionary hypotheses predict that geographical overlap with closely related taxa should drive divergence in traits used to distinguish between conspecifics and heterospecifics (Wallace 1889; Fisher 1930; Dobzhansky 1940; Mayr 1963; Gröning and Hochkirch 2008; Ord and Stamps 2009; Ord et al. 2011). Moreover, biologists interested in speciation have often argued that interspecific competitive interactions are important drivers of divergence between lineages that ultimately leads to reproductive isolation. Reinforcement, or selection against hybridization (Dobzhansky

1937, 1940), for example, is often thought to be an important phase of speciation (Grant 1999; Coyne and Orr 2004; Rundle and Nosil 2005; Pfennig and Pfennig 2009).

Despite the preponderance of classical evolutionary processes that assume that interspecific interactions have important fitness consequences, existing phylogenetic models of trait evolution treat trait evolution within a lineage as independent from traits in other lineages. For example, in the commonly-used Brownian motion and Ornstein-Uhlenbeck models of trait evolution (Cavalli-Sforza & Edwards 1967, Felsenstein 1988), once an ancestor splits into two daughter lineages, the trait values in those daughter lineages do not depend on the trait values of sister taxa. Some investigators have indirectly incorporated the influence of interspecific interactions by fitting models where evolutionary rates at a given time depend on the diversity of lineages at that time (e.g., the “diversity-dependent” models of Mahler et al. 2010, Weir and Mursleen 2013). While these models capture some parts of the interspecific processes of central importance to evolutionary theory, such as the influence of ecological opportunity, they do not explicitly account for trait-driven interactions between lineages, as trait values in one lineage do not vary directly as a function of trait values in other evolving lineages.

Recently, Nuismer and Harmon (2015) proposed a model where the evolution of a species’ trait depends on other species’ traits. In particular, they consider a model, which they refer to as the model of phenotype matching, where the probability that an encounter between two individuals has fitness consequences declines as the phenotypes of the individuals become more dissimilar. The consequence of the encounter on fitness can be either negative if the interaction is competitive, resulting in character divergence (matching competition, e.g. resource competition), or positive if the interaction is mutualistic, resulting in character convergence (matching mutualism, e.g. Müllerian mimicry). Applying Lande’s formula (Lande 1976) and

given a number of simplifying assumptions—importantly that all lineages evolve in sympatry—this model yields a simple prediction for the evolution of a population’s mean phenotype.

Here, we develop inference tools for fitting the matching competition model (i.e., the phenotype matching model of Nuismer and Harmon incorporating competitive interactions between lineages) to combined phylogenetic and trait data. We begin by showing how to compute likelihoods associated with this model. Next, we use simulations to explore the statistical properties of maximum likelihood estimation of the matching competition model (parameter estimation as well as the power and type I error rates). While the inclusion of interactions between lineages is an important contribution to quantitative models of trait evolution, applying the matching competition model to an entire clade relies on the assumption that all lineages in the clade are sympatric. However, this assumption will be violated in most empirical cases, so we also developed a method for incorporating data on the biogeographical overlap between species for this model and for the linear and exponential diversity-dependent trait models of Weir & Mursleen (2013), wherein the evolutionary rate at a given time in a tree varies as a function of the number of lineages in the reconstructed phylogeny at that time (see also Mahler et al. 2010).

We then fit the model to data from a classical adaptive radiation: Greater Antillean *Anolis* lizards (Harmon et al. 2003; Losos 2009). Many lines of evidence support the hypothesis that resource competition is responsible for generating divergence between species in both habitat use (e.g., Pacala and Roughgarden 1982) and morphology (Schoener 1970; Williams 1972; see review in Losos 1994). Thus, we can make an *a priori* prediction that model comparison will uncover a signature of competition in morphological traits that vary with habitat and resource use. However, in spite of the importance of interspecific competition to adaptive radiation

theory, the role of competition in driving adaptive divergence has been historically difficult to measure (Losos 2009) because trait divergence resulting from competition between lineages during their evolutionary history should have the effect of eliminating competition between those lineages at the present. Nevertheless, the signature of such competition may be detectable in the contemporary distribution of trait values and their covariance structure (Hansen and Martins 1996; Nuismer and Harmon 2015), and as such, the matching competition model should provide a good fit to morphological data collected on anoles. Given the well-resolved molecular phylogeny (Mahler et al. 2010, 2013) and the relatively simple geographical relationships between species (i.e., species are restricted to single islands, Rabosky and Glor 2010; Mahler and Ingram 2014), the Greater Antillean *Anolis* lizards provide a good test system for exploring the effect of competition on trait evolution using the matching competition model.

## METHODS

### *Likelihood Estimation of the Matching Competition Model*

We consider the evolution of a quantitative trait under the matching competition model of Nuismer & Harmon (2015) wherein trait divergence between lineages will be favored by selection. In our version of the model, we remove stabilizing selection to focus on the effect of competition. The evolution of a population's mean phenotype is thus given by (Eq. S38 in Nuismer & Harmon 2015 with  $\psi = 0$ ):

$$\bar{z}'_i = \bar{z}_i + S(\mu - \bar{z}_i) + \delta \quad (\text{Eq. 1})$$

where drift is incorporated as a Gaussian variable  $\delta$  with mean = 0 and variance =  $\sigma^2$  (the Brownian motion rate parameter) and  $S$  measures the strength of interaction (more intense competitive interactions are represented by larger negative values),  $\bar{z}_i'$  is the mean trait value for lineage  $i$  after an infinitesimally small time step, and  $\mu$  is the mean trait value for the entire clade at the beginning of that time step. Hence, if a species trait value is greater (respectively smaller) than the trait value average across species in the clade, the species' trait will evolve towards even larger (respectively smaller) trait values.

Like several other models of quantitative trait evolution (Harmon et al. 2010, Weir & Mursleen 2013), the expected distribution of trait values on a given phylogeny under the matching competition model follows a multivariate normal distribution with mean vector made of terms each equal to the character value at the root of the tree and variance-covariance matrix determined by the parameter values and phylogeny. Nuismer & Harmon (2015) provide the system of ordinary differential equations describing the evolution of the variance and covariance terms through time (their Eqs. 10b and 10c). These differential equations can be integrated numerically from the root to the tips of phylogenies to compute expected variance-covariance matrices and the associated likelihood values given by the multivariate normal distribution.

Additionally, to relax the assumption that all of the lineages in a clade coexist sympatrically, we included a term to specify which lineages co-occur at any given time-point in the phylogeny, which can be inferred, e.g., by biogeographical reconstruction. The resulting system of ordinary differential equations describing the evolution of the variance and covariance terms through time are:

$$\frac{dv_i^2}{dt} = -\left(\frac{2(n_i-1)}{n_i}S\right)v_i^2 + \frac{2S}{n_i}\left(\sum_{\substack{j=1 \\ (j \neq i)}}^n \mathbf{A}_{i,j}v_{i,j}\right) + \sigma^2 \quad (\text{Eq. 2a})$$

144

$$\frac{dv_{i,j}}{dt} = -\left(\frac{2(n_i-1)}{n_i}S\right)v_{i,j} + \frac{S}{n_i}\left(\sum_{\substack{k=1 \\ k \neq j}}^n \mathbf{A}_{i,k}v_{i,k} + \sum_{\substack{l=1 \\ l \neq i}}^n \mathbf{A}_{j,l}v_{j,l}\right) \quad (\text{Eq. 2b})$$

146

147 where  $v_i$  is the variance for each species  $i$  at time  $t$  and  $v_{i,j}$  is the covariance for each species pair

148  $i,j$  at time  $t$ . Eq. 2b describes the evolution of the covariance at time  $t$  if species  $i$  and  $j$  are in

149 sympatry at time  $t$ . If they are not,  $\frac{dv_{i,j}}{dt} = 0$  such that the covariance between species  $i$  and  $j$

150 remains fixed. The terms  $\sigma^2$  and  $S$  are as specified above,  $\mathbf{A}_{i,j}$  equals 1 at time  $t$  if  $i$  and  $j$  are

151 sympatric at that time, and 0 otherwise (Fig. 1), and  $n_i = \sum_{j=1}^n A_{ij}$  is the number of lineages

152 interacting with lineage  $i$  at time  $t$  (equal to the number  $n$  of lineages in the reconstructed

153 phylogeny at time  $t$  if all species are sympatric). Here, we consider the matrix  $\mathbf{A}$  to be a block

154 diagonal matrix, such that sets of interacting species are non-overlapping, as in the case of *Anolis*

155 lizards (Fig. 1). This implies that if species  $i$  and  $j$  are sympatric, then  $n_i = n_j$  and  $v_{i,j} = v_{j,i}$ .

156 Note that when  $S = 0$  or  $n = 1$  (i.e., when a species is alone), this model reduces to Brownian

157 motion. This model has three free parameters:  $\sigma^2$ ,  $S$  and the ancestral state  $z_0$  at the root. As with

158 other models of trait evolution, the maximum likelihood estimate for the ancestral state is

159 computed through GLS using the estimated variance-covariance matrix (Grafen 1989; Martins

160 and Hansen 1997).

161 We used the ode function in the R package deSolve (Soetaert et al. 2010) to perform the

162 numerical integration of the differential equations using the “lsoda” solver, and the Nelder-Mead

163 algorithm implemented in the optim function to perform the maximum likelihood optimization.



Codes for these analyses are freely available on github (<https://github.com/hmorlon/PANDA>) and included the R package RPANDA (Morlon 2014).

### *Incorporating Geography into Diversity-Dependent Models*

Using the same geography matrix **A** described above for the matching competition model (Fig. 1), we modified the diversity-dependent linear and exponential models of Weir & Mursleen (2013) to incorporate biological realism into the models, because ecological opportunity is only relevant within rather than between biogeographical regions. The resulting variance-covariance matrices, **V**, of these models have the elements:

$$\mathbf{V}_{ij} = \sum_{m=2}^M (\sigma_0^2 + bn_i) (\max(s_{ij} - t_{m-1}, 0) - \max(s_{ij} - t_m, 0)) \quad (\text{Eq. 3})$$

for the diversity-dependent linear model, and

$$\mathbf{V}_{ij} = \sum_{m=2}^M (\sigma_0^2 \times e^{rn_i}) (\max(s_{ij} - t_{m-1}, 0) - \max(s_{ij} - t_m, 0)) \quad (\text{Eq. 4})$$

for the diversity-dependent exponential model, where  $\sigma_0^2$  is the rate parameter at the root of the tree,  $b$  and  $r$  are the slopes in the linear and exponential models, respectively,  $s_{ij}$  is the shared path length of lineages  $i$  and  $j$  from the root of the phylogeny to their common ancestor,  $n_i$  is the number of sympatric lineages (as above) between nodes at times  $t_{m-1}$  and  $t_m$  (where  $t_1$  is 0, the time at the root, and  $t_M$  is the total length of the tree) (Weir & Mursleen 2013). When  $b$  or  $r = 0$ , these models reduce to Brownian motion.

# *Simulation-based Analysis of Statistical Properties of the Matching Competition Model*

To verify that the matching competition model can be reliably fit to empirical data, we simulated trait datasets to estimate its statistical properties (i.e., parameter estimation, power, and type I error rates). For all simulations, we began by first generating 100 pure-birth trees using TreeSim (Stadler 2014). To determine the influence of the number of tips in a tree, we ran simulations on trees of size  $n = 20, 50, 100$ , and 150. We then simulated continuous trait datasets by applying the matching competition model recursively from the root to the tip of each tree (Paradis 2011), following Eq. 1. For these simulations, we set  $\sigma^2 = 0.05$  and systematically varied  $S$  (-1.5, -1, -0.5, -0.1, or 0). Finally, we fit the matching competition model to these datasets using the ML optimization described above.

To determine the ability of the approach to accurately estimate simulation parameter values, we first compared estimated parameters to the known parameters used to simulate datasets under the matching competition model ( $S$  and  $\sigma^2$ ). We also quantified the robustness of these estimates in the presence of extinction by estimating parameters for datasets simulated on birth-death trees; in addition, we compared the robustness of the matching competition model to extinction to that of the diversity-dependent models. These two latter sets of analyses are described in detail in the Supplementary Appendix 1.

To assess the power of the matching competition model, we compared the fit of this model to other commonly used trait models on the same data (i.e. data simulated under the matching competition model). Specifically, we compared the matching competition model to (1) Brownian motion (BM), (2) Ornstein-Uhlenbeck/single-stationary peak model (OU, Hansen & Martin 1996), (3) exponential time-dependent ( $TD_{exp}$ , i.e., the early burst model, or the ACDC model with the rate parameter set to be negative, Blomberg et al. 2003; Harmon et al. 2010), (4)

linear time-dependent evolutionary rate ( $TD_{lin}$ , Weir and Mursleen 2012), (5) linear rate diversity-dependent ( $DD_{lin}$ , Mahler et al. 2010; Weir and Mursleen 2013), and (6) exponential rate diversity-dependent ( $DD_{exp}$ , Weir and Mursleen 2013). These models were fitted using *geiger* (Harmon et al. 2008) when available there (BM, OU,  $TD_{exp}$ ,  $TD_{lin}$ ), or using our own codes, available in RPANDA (Morlon 2014) when they were not available in *geiger* ( $DD_{lin}$ ,  $DD_{exp}$ ). Support for each model was determined using Akaike weights (Burnham and Anderson 2002).

We assessed the type I error rate of the matching competition model by calculating the fit of this model to datasets simulated under the same trait models mentioned above. For BM and OU models, we generated datasets from simulations using parameter values from the appendix of Harmon et al. 2010 scaled to a tree of length 400 (BM,  $\sigma^2 = 0.03$ ; OU,  $\sigma^2 = 0.3$ ,  $\alpha = 0.06$ ). For both the linear and exponential versions of the time- and diversity-dependent models, we simulated datasets with starting rates of  $\sigma^2 = 0.6$  and ending rates of  $\sigma^2 = 0.01$ , declining with a slope determined by the model and tree (e.g., for time-dependent models, the slope is a function of the total height of the tree; for the  $TD_{exp}$  model, these parameters result in a total of 5.9 half-lives elapsing from the root to the tip of the tree, Slater and Pennell 2014). These simulations were performed using our own codes, available in RPANDA (Morlon 2014). As above, we calculated the Akaike weights for all models for each simulated dataset.

### *Fitting the Matching Competition Model of Trait Evolution to Caribbean Anolis Lizards*

To determine whether the matching competition model is favored over models that ignore interspecific interactions in an empirical system where competition is known to have influenced character evolution, we fit the matching competition model to a morphological dataset on adult

males of 100 species of Greater Antillean *Anolis* lizards and the time calibrated, maximum clade credibility tree calculated from a Bayesian sample of molecular phylogenies (Mahler et al. 2010, 2013; Mahler and Ingram 2014). We included the first four size-corrected phylogenetic principal components from a set of 11 morphological measurements, collectively accounting for 93% of the cumulative variance explained (see details in Mahler et al. 2013). Together, the shape axes quantified by these principal components describe the morphological variation associated with differences between classical ecomorphs in Caribbean anoles (Williams 1972). In addition to the matching competition model, we fit the six previously mentioned models (BM, OU,  $TD_{exp}$ ,  $TD_{lin}$ ,  $DD_{exp}$ , and  $DD_{lin}$ ) to the *Anolis* dataset.

For the matching competition model and diversity-dependent models, to determine the influence of designating clades as sympatric and allopatric, we fit the model for each trait using two sets of geography matrices (i.e.,  $\mathbf{A}$  in Eq. 1b, 2, & 3, see Fig. 1): one where all lineages were set as sympatric, and another following the biogeographical reconstruction presented in Fig 18.3 of Mahler and Ingram (2014). To simplify the ML optimization, we restricted  $S$  to take negative values while fitting the matching competition model including the biogeographical relationships among taxa.

## RESULTS

### *Statistical Properties of the Matching Competition Model*

Across a range of  $S$  values, maximum likelihood optimization returns reliable estimates of parameter values (Fig. 2). As the number of tips increases, so does the reliability of maximum likelihood parameter values (Fig. 2). Parameter estimates remain reliable in the presence of extinction, unless the extinction fraction is very large (Supplementary Appendix 1). When

datasets are simulated under the matching competition model, model selection generally picks the matching competition model as the best model (Figs. 3, S1); the strength of this discrimination depends on both the  $S$  value used to simulate the data and the size of the tree (Figs. 3, S1).

We found that the matching competition model behaves similarly to a model of declining evolutionary rates through time, as the rate parameter values of the  $TD_{exp}$  model estimated on data generated under the matching competition model became increasingly negative under increasing levels of competition (Fig. S2). The dynamics of this declining rate were better described by an exponential decline in rates than by a linear one (Table S1).

Simulating datasets under BM, OU,  $DD_{exp}$ , and  $DD_{lin}$  generating models, we found that there is a reasonably low type I error rate for the matching competition model (Fig. 4a,b,e,f). When character data were simulated under a  $TD_{lin}$  model of evolution, the matching competition and/or the diversity-dependent models tended to have lower AICc values than the  $TD_{lin}$  model, especially among smaller trees (Figure 4d). For data generated under a  $TD_{exp}$  model, model selection always favored the matching competition model over the  $TD_{exp}$  model (Fig. 4c).

### *Competition in Greater Antillean Anolis Lizards*

For the first four phylogenetic principal components describing variation in *Anolis* morphology, we found that models that incorporate species interactions fit the data better than models that ignore them (Table 1). PC1 and PC3, which describe variation in hindlimb/hindtoe and forelimb/foretoe length (Mahler et al. 2013), respectively, are fit best by the matching competition model. PC2 and PC4, which describe variation in body size (snout vent length) and lamellae number, respectively, are fit best by the exponential diversity-dependent model.

Additionally, in every case, models that incorporated the geographic relationships among species in the tree outperformed models that assumed that lineages interact in sympatry at the clade level (Table 1).

## DISCUSSION

The inference methods we present here represent an important new addition to the comparative trait analysis toolkit. Whereas previous models had not accounted for the influence of trait values in other lineages on character evolution, the matching competition model takes these into account. Furthermore, extending both the matching competition model and two diversity-dependent trait evolution models to incorporate geographic networks of sympatry further extends the utility and biological realism of these models.

We found that the matching competition property has increasing power and accuracy of parameter estimation with increasing tree sizes and competition strength. We also found that, for most of the generating models we tested (but see below), the matching competition model is not erroneously favored using model selection (i.e., there is a reasonably low type-I error rate). As with all other models, the statistical properties of the matching competition model will depend on the size and shape of a particular phylogeny as well as specific model parameter values. Future investigators can employ other approaches, such as phylogenetic Monte Carlo and posterior predictive simulations directly on their empirical trees (Boettiger et al. 2012, Slater & Pennell 2014), to assess the confidence they can have in their results

We found that data generated under time-dependent models were often fit better by models that incorporate interspecific interactions (i.e., density-dependent and matching competition models) (Fig. 4c,d). This was especially true for the  $TD_{exp}$  model, often referred to

as the early-burst model—the matching competition model nearly always fit data generated under the  $TD_{exp}$  model better than the  $TD_{exp}$  model (Fig. 4c). We do not view this as a major limitation of the model for two reasons. First, the  $TD_{exp}$  model is known to be statistically difficult to estimate on neontological data alone (Harmon et al. 2010; Slater et al. 2012; Slater and Pennell 2014). Accordingly, since the matching competition model also describes declining evolutionary rates from the root to the tip of the tree (Fig. S2, Table S1), it is perhaps not surprising that the matching competition model fits data generated under the  $TD_{exp}$  model well. Secondly, and more importantly, time-dependent models are not process-based models, but rather incorporate time since the root of a tree as a proxy for ecological opportunity or available niche space (Harmon et al. 2010; Mahler et al. 2010; Slater 2015). The matching competition and density-dependent models explicitly account for the interspecific competitive interactions that time-dependent models purport to model, thus we argue that these process-based models are more biologically meaningful than time-dependent models (Moen and Morlon 2014).

Because the matching competition model depends on the mean trait values in an evolving clade, maximum likelihood estimation is robust to extinction, whereas the diversity-dependent models are less so (Appendix S1, Figs. S3-S6). Nevertheless, given the failure of maximum likelihood to recover accurate parameter estimations at high levels of extinction ( $\mu: \lambda \geq 0.6$ ), we suggest that these models should not be used in clades where the extinction rate is known to be particularly high. In such cases, it would be preferable to adapt the inference framework presented here to include data from fossil lineages (Slater et al. 2012), which could be easily done by integrating the ordinary differential equations described in Eq. 2a and 2b on non-ultrametric trees.

For all of the traits we analyzed, we found that models incorporating both the influence of other lineages and the specific geographical relationships among lineages were the most strongly supported models (though less strikingly for PC3 and PC4). The matching competition model is favored in the PCs describing variation in relative limb size. Previous research demonstrates that limb morphology explains between-ecomorph variation in locomotive capabilities and perch characteristics (Losos 1990, 2009; Irschick et al. 1997), and our results suggest that the evolutionary dynamics of these traits have been influenced by the evolution of limb morphology in other sympatric lineages. These results support the assumption that interspecific interactions resulting from similarity in trait values are important components of adaptive radiations (Losos 1994, Schluter 2000), a prediction that has been historically difficult to test (Losos 2009, but see Mahler et al. 2010). In combination with previous research demonstrating a set of convergent adaptive peaks to which lineages are attracted (Mahler et al. 2013), our results suggest that competition likely played an important role in driving lineages toward these distinct peaks.

We imagine that the matching competition model and biogeographical implementations of diversity-dependent models will play a substantial role in comparative studies of interspecific competition. There are many possible extensions of the tools developed in this paper. In the current implementation of interspecific geographic overlap, the matrices of sympatry/allopatry are assumed to be block diagonal matrices. In the future, the model can be extended to more complex geography matrices that are more realistic for mainland taxa (e.g., using ancestral biogeographical reconstruction, Ronquist and Sanmartín 2011; Landis et al. 2013), and can also specify degrees of sympatric overlap (i.e., syntopy). Additionally, although the current version of the model is rather computationally expensive with larger trees (with 100 or more tips), future work to speed up the calculation of the variance-covariance matrix under the matching



competition model would allow the inclusion of uncertainty in biogeographical reconstruction (e.g., from samples of stochastic maps). The current form of the model assumes that the degree of competition is equal for all interacting lineages. Future modifications of the model, such as applications of stepwise AICc algorithms (Alfaro et al. 2009; Thomas and Freckleton 2012; Mahler et al. 2013) or reversible-jump Markov Chain Monte Carlo (Pagel and Meade 2006; Eastman et al. 2011; Rabosky 2014; Uyeda and Harmon 2014), may be useful to either identify more intensely competing lineages or test specific hypotheses about the strength of competition between specific taxa. Improvements could also be made on the formulation itself of the evolution of a species' trait as a response to the phenotypic landscape in which the species occurs. The current formulation (Eq. 1) corresponds to a scenario in which the rate of phenotypic evolution on a lineage gets higher as the lineage deviates from the mean phenotype, although character displacement theory, for example, posits that selection for divergence should be the strongest when species are most ecologically similar (Brown & Wilson 1956). Finally, a great array of extensions will come from modeling species interactions not only within clades, but also among interacting clades, as in the case of coevolution in bipartite mutualistic or antagonistic networks, such as plant-pollinator or plant-herbivore systems (Bartosek et al. 2012).

In addition to uncovering a signature of competitive interactions in traits used in exploitative competition for resources (e.g., trait-axes along which adaptive radiations proceed, Schluter 2000), the tools presented here can be used to test several hypotheses that have historically been difficult to test using phylogenetic comparative methods. For example, various hypotheses about the dynamics of traits involved in social interactions that posit that interspecific interactions have played an important role in shaping contemporary trait values (e.g., Pfennig and Pfennig 2012; Grether et al. 2013) are beginning to be tested at comparative scales (Seddon

et al. 2013; Tobias et al. 2013; Martin et al. 2015). However, the tools available for such analyses have been limited (e.g., sister-taxa comparisons) relative to those available to test macroevolutionary hypotheses about within-taxa processes. By comparing the fits of the matching competition model with other models that do not include competitive interactions between lineages, biologists can directly test hypotheses that make predictions about the role of interspecific interactions in driving trait evolution.

## Acknowledgements

We thank M. Manceau for his help troubleshooting the inference approach, J. Weir for providing R code for diversity-dependent models, and E. Lewitus and O. Missa for helpful comments on the manuscript. This research was funded by the Agence Nationale de la Recherche (grant CHEX-ECOEVOBIO); and the European Research Council (grant 616419-PANDA to HM).

## References

- Alfaro M.E., Santini F., Brock C., Alamillo H., Dornburg A., Rabosky D.L., Carnevale G.,  
Harmon L.J. 2009. Nine exceptional radiations plus high turnover explain species diversity  
in jawed vertebrates. *Proc. Natl. Acad. Sci.* 106:13410–13414.
- Bartosek K., Pienaar J., Mostad P., Andersson S., Hansen T.F. 2012. A phylogenetic  
comparative method for studying multivariate adaptation. *J. of Theor. Biol.* 314:204-215.
- Blomberg S.P., Garland T., Ives A.R. 2003. Testing for phylogenetic signal in comparative data:  
behavioral traits are more labile. *Evolution.* 57:717–745.
- Boettiger C., Coop G., Ralph P. 2012. Is Your Phylogeny Informative ? Measuring the Power of  
Comparative Methods. *Evolution* 66:2240–2251.
- Brown W.L., Wilson E.O. 1956. Character Displacement. *Syst. Zool.* 5:49-62.
- Burnham K.P., Anderson D.R. 2002. Model selection and multimodel inference: a practical  
information-theoretic approach. New York, NY: Springer.
- Cavalli-Sforza, L.L., Edwards, A.W.F. 1967. Phylogenetic analysis: Models and estimation  
procedures. *Amer. J. of Hum. Gen.* 19:233-257.
- Coyne J.A., Orr H.A. 2004. Speciation. Sunderland, MA: Sinauer Associates.
- Dobzhansky T. 1937. Genetics and the Origin of Species. New York, NY: Columbia University  
Press.

- 403 Dobzhansky T. 1940. Speciation as a stage in evolutionary divergence. *Am. Nat.* 74:312–321.
- 404 Eastman J.M., Alfaro M.E., Joyce P., Hipp A.L., Harmon L.J. 2011. A novel comparative  
405 method for identifying shifts in the rate of character evolution on trees. *Evolution* 65:3578–  
406 3589.
- 407 Felsenstein, J. 1988. Phylogenies and quantitative characters. *Ann. Rev. of Ecol., Evol., &*  
408 *Systematics.* 19:445-471.
- 409 Fisher R.A. 1930. *The Genetical Theory of Natural Selection.* Oxford, UK.: Oxford University  
410 Press.
- 411 Grafen A. 1989. The phylogenetic regression. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.*  
412 326:119–157.
- 413 Grant P.R., Grant B.R. 2002. Adaptive radiation of Darwin’s finches: Recent data help explain  
414 how this famous group of Galápagos birds evolved, although gaps in our understanding  
415 remain. *Am. Sci.* 90:130–139.
- 416 Grant P.R., Grant B.R. 2011. *How and Why Species Multiply: the Radiation of Darwin’s*  
417 *Finches.* Princeton, NJ: Princeton University Press.
- 418 Grant P.R. 1972. Convergent and divergent character displacement. *Biol. J. Linn. Soc.* 4:39–68.
- 419 Grant P.R. 1999. *Ecology and Evolution of Darwin’s finches.* Princeton, NJ: Princeton  
420 University Press.

421 Grether G.F., Anderson C.N., Drury J.P., Kirschel A.N.G., Losin N., Okamoto K., Peiman K.S.  
 422 2013. The evolutionary consequences of interspecific aggression. *Ann. N. Y. Acad. Sci.*  
 423 1289:48–68.

424 Grether G.F., Losin N., Anderson C.N., Okamoto K. 2009. The role of interspecific interference  
 425 competition in character displacement and the evolution of competitor recognition. *Biol.*  
 426 *Rev.* 84:617–635.

427 Gröning J., Hochkirch A. 2008. Reproductive interference between animal species. *Q. Rev. Biol.*  
 428 83:257–282.

429 Hansen T.F., Martins E.P. 1996. Translating between microevolutionary process and  
 430 macroevolutionary patterns: The correlation structure of interspecific data. *Evolution*  
 431 50:1404–1417.

432 Harmon L.J., Losos J.B., Jonathan Davies T., Gillespie R.G., Gittleman J.L., Bryan Jennings W.,  
 433 Kozak K.H., McPeck M.A., Moreno-Roark F., Near T.J., Purvis A., Ricklefs R.E., Schluter  
 434 D., Schulte J.A., Seehausen O., Sidlauskas B.L., Torres-Carvajal O., Weir J.T., Mooers  
 435 A.T. 2010. Early bursts of body size and shape evolution are rare in comparative data.  
 436 *Evolution* 64:2385–2396.

437 Harmon L.J., Schulte J.A., Larson A., Losos J.B. 2003. Tempo and mode of evolutionary  
 438 radiation in iguanian lizards. *Science* 301:961–964.

439 Harmon L.J., Weir J.T., Brock C.D., Glor R.E., Challenger W. 2008. GEIGER: investigating  
 440 evolutionary radiations. *Bioinformatics.* 24:129–131.

- 441 Irschick D.J., Vitt L.J., Zani P.A., Losos J.B. 1997. A comparison of evolutionary radiations in  
442 mainland and Caribbean *Anolis* lizards. *Ecology*. 78:2191–2203.
- 443 Landis M.J., Matzke N.J., Moore B.R., Huelsenbeck J.P. 2013. Bayesian analysis of  
444 biogeography when the number of areas is large. *Syst. Biol.* 62:789–804.
- 445 Losos J.B., Ricklefs R.E. 2009. Adaptation and diversification on islands. *Nature*. 457:830–836.
- 446 Losos J.B. 1990. The evolution of form and function: morphology and locomotor performance in  
447 West Indian *Anolis* lizards. 44:1189–1203.
- 448 Losos J.B. 1994. Integrative Approaches to Evolutionary Ecology: *Anolis* Lizards as Model  
449 Systems. *Annu. Rev. Ecol. Syst.* 25:467–493.
- 450 Losos J.B. 2009. *Lizards in an Evolutionary Tree: Ecology and Adaptive Radiation of Anoles*.  
451 Los Angeles, CA: University of California Press.
- 452 Mahler D.L., Ingram T., Revell L.J., Losos J.B. 2013. Exceptional convergence on the  
453 macroevolutionary landscape in island lizard radiations. *Science* 341:292–5.
- 454 Mahler D.L., Ingram T. 2014. Phylogenetic Comparative Methods for Studying Clade-Wide  
455 Convergence. In: Garamszegi L., editor. *Modern Phylogenetic Comparative Methods and*  
456 *Their Application in Evolutionary Biology*. New York, NY: Springer. p. 425–450.
- 457 Mahler D.L., Revell L.J., Glor R.E., Losos J.B. 2010. Ecological opportunity and the rate of  
458 morphological evolution in the diversification of greater Antillean anoles. *Evolution*  
459 64:2731–2745.

- 460 Martin P.R., Montgomerie R., Loughheed S.C. 2015. Color patterns of closely related bird species  
461 are more divergent at intermediate levels of breeding-range sympatry. *Am. Nat.* 185:443–  
462 451.
- 463 Martins E.P., Hansen T.F. 1997. Phylogenies and the comparative method: a general approach to  
464 incorporating phylogenetic information into the analysis of interspecific data. *Am. Nat.*  
465 149:646–667.
- 466 Mayr E. 1963. *Animal Species and Evolution*. Cambridge, MA: Harvard University Press.
- 467 Moen D., Morlon H. 2014. Why does diversification slow down? *Trends Ecol. Evol.* 29:190–  
468 197.
- 469 Morlon H. 2014. RPANDA: Phylogenetic ANalyses of DiversificAtion. R package version 1.0.  
470 <https://github.com/hmorlon/PANDA>
- 471 Nuismer S.L., Harmon L.J. 2015. Predicting rates of interspecific interaction from phylogenetic  
472 trees. *Ecol. Lett.* 18:17–27.
- 473 Ord T.J., King L., Young A.R. 2011. Contrasting theory with the empirical data of species  
474 recognition. *Evolution* 65:2572–2591.
- 475 Ord T.J., Stamps J.A. 2009. Species identity cues in animal communication. *Am. Nat.* 174:585–  
476 593.
- 477 Pacala S., Roughgarden J. 1982. Resource partitioning and interspecific competition in two two-  
478 species insular *Anolis* lizard communities. *Science* 217:444–446.

- 479 Pagel M., Meade A. 2006. Bayesian analysis of correlated evolution of discrete characters by  
480 reversible-jump Markov chain Monte Carlo. *Am. Nat.* 167:808–825.
- 481 Paradis E. 2011. *Analysis of Phylogenetics and Evolution with R*. New York, NY: Springer.
- 482 Pfennig D.W., Pfennig K.S. 2012. *Evolution's Wedge: Competition and the Origins of Diversity*.  
483 Los Angeles, CA: Univ of California Press.
- 484 Pfennig K.S., Pfennig D.W. 2009. Character displacement: ecological and reproductive  
485 responses to a common evolutionary problem. *Q. Rev. Biol.* 84:253–276.
- 486 Rabosky D.L., Glor R.E. 2010. Equilibrium speciation dynamics in a model adaptive radiation of  
487 island lizards. *Proc. Natl. Acad. Sci. U. S. A.* 107:22178–22183.
- 488 Rabosky D.L. 2014. Automatic detection of key innovations, rate shifts, and diversity-  
489 dependence on phylogenetic trees. *PLoS One*. 9:e89543.
- 490 Ronquist F., Sanmartín I. 2011. Phylogenetic methods in biogeography. *Annu. Rev. Ecol. Evol.*  
491 *Syst.* 42:441–464.
- 492 Rundle H.D., Nosil P. 2005. Ecological speciation. *Ecol. Lett.* 8:336–352.
- 493 Schluter D. 2000. *The Ecology of Adaptive Radiation*. Oxford, UK: Oxford University Press.
- 494 Schoener T.W. 1970. Size Patterns in West Indian Anolis Lizards. II. Correlations with the Sizes  
495 of Particular Sympatric Species-Displacement and Convergence. *Am. Nat.* 104:155–174.



496 Seddon N., Botero C.A., Tobias J.A., Dunn P.O., Macgregor H.E., Rubenstein D.R., Uy J.A.,  
 497 Weir J.T., Whittingham L.A., Safran R.J. 2013. Sexual selection accelerates signal  
 498 evolution during speciation in birds. *Proc. R. Soc. B Biol. Sci.* 280:20131065.

499 Slater G.J., Harmon L.J., Alfaro M.E. 2012. Integrating fossils with molecular phylogenies  
 500 improves inference of trait evolution. 66:3931–3944.

501 Slater G.J., Pennell M.W. 2014. Robust regression and posterior predictive simulation increase  
 502 power to detect early bursts of trait evolution. *Syst. Biol.* 63:293–308.

503 Soetaert K., Petzoldt T., Setzer R.W. 2010. Solving Differential Equations in R: Package  
 504 deSolve. *J. Stat. Softw.* 33:1–25.

505 Stadler T. 2014. TreeSim: Simulating trees under the birth-death model. R package version 2.1.  
 506 <http://CRAN.R-project.org/package=TreeSim>.

507 Thomas G.H., Freckleton R.P. 2012. MOTMOT: models of trait macroevolution on trees.  
 508 *Methods Ecol. Evol.* 3:145–151.

509 Tobias J.A., Cornwallis C.K., Derryberry E.P., Claramunt S., Brumfield R.T., Seddon N. 2013.  
 510 Species coexistence and the dynamics of phenotypic evolution in adaptive radiation. *Nature*.  
 511 506:359–363.

512 Uyeda J.C., Harmon L.J. 2014. A novel Bayesian method for inferring and interpreting the  
 513 dynamics of adaptive landscapes from phylogenetic comparative data. *Syst. Biol.* 63:902–  
 514 918.

- 515 Wallace A.R. 1889. Darwinism. 2007 facsimile, Cosimo, Inc.
- 516 Weir J.T., Mursleen S. 2013. Diversity-dependent cladogenesis and trait evolution in the
- 517 adaptive radiation of the auks (Aves: Alcidae). *Evolution* 67:403–416.
- 518 Williams E.E. 1972. The Origin of Faunas. Evolution of Lizard Congeners in a Complex Island
- 519 Fauna: A Trial Analysis. *Evol. Biol.* 6:47–89.
- 520
- 521

## Table & Figure legends

**Table 1.** Comparison of model fits for the first four phylogenetic principal components of a morphological dataset of Greater Antillean anoles. Models run incorporating geography matrices are indicated by “+ GEO”, and models with the lowest AICc for each trait are shaded and written in bold text. Parameter values presented follow the nomenclature of Eqs. 2-4 in the main text, and  $k$  represents the number of parameters estimated for each model. Note that  $TD_{exp}$  is the ACDC model (or the early-burst model when  $r < 0$ ). OU model weights were excluded because  $\alpha$  was estimated to be 0 for each trait, and thus the OU model was equivalent to BM.

*Figure 1.* Illustration of geography matrices (defined between each node) delineating which lineages interact in sympatry in an imagined phylogeny. These matrices were used to identify potentially interacting lineages for the matching competition and both diversity-dependent models of character evolution (see Eqs. 2-4 in the main text). *Anolis* outline courtesy of Sarah Werning, licensed under Creative Commons (<http://creativecommons.org/licenses/by/3.0/>).

*Figure 2.* Parameter estimation under the matching competition model. As tree size increases and/or the magnitude of competition increases (i.e., the  $S$  parameter in the matching competition model becomes more negative), so does the accuracy of ML parameter estimates of (A)  $S$  ( $n = 100$  for each tree size and  $S$  value combination; red horizontal lines indicate the simulated  $S$  value) and (B) the accuracy of ML parameter estimates of  $\sigma^2$  ( $n = 500$  for each tree size; red horizontal lines indicate the simulated  $\sigma^2$  value). The numbers below the violin plots in (B) show the number of outliers removed for plotting.

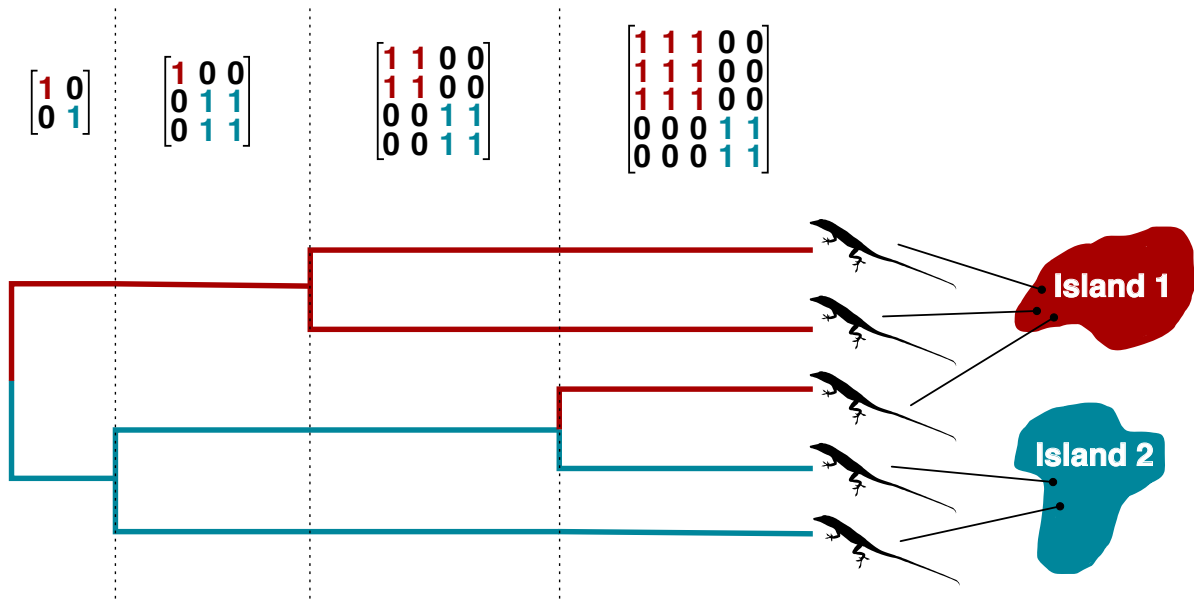
*Figure 3.* The statistical power of the matching competition increases with tree size and with increasing levels of competition (i.e., increasingly negative  $S$  values).

*Figure 4.* Type-I error simulation results for the matching competition model. When the generating model is either (A) BM, (B) OU, (E)  $DD_{exp}$  (for larger trees) or (F)  $DD_{lin}$ , the generating model is largely favored by model selection. However, both (C)  $TD_{exp}$  and (D)  $TD_{lin}$  (for smaller trees) are erroneously rejected as the generating model.

**Table 1.**

Trait	Model	$k$	$\sigma^2$	$b$	$r$	$S$	$\ln(\mathcal{L})$	$\Delta AICc$	Akaike weights
pPC1	BM	2	0.0033	—	—	—	-13.68	22.40	<0.01
	OU	3	0.0033	—	—	—	-13.68	24.52	—
	TDexp	3	0.0324	—	-0.068	—	-5.20	7.55	0.02
	TDlinear	3	0.0113	-0.019	—	—	-4.88	6.92	0.03
	LDexp	3	0.0184	—	-0.028	—	-4.37	5.90	0.04
	LDexp + GEO	3	0.0086	—	-0.043	—	-7.28	11.72	<0.01
	LDlin	3	0.0089	-0.00008	—	—	-4.89	6.94	0.02
	LDlin + GEO	3	0.0042	-0.000042	—	—	-10.34	17.84	<0.01
	MC <sub>sym</sub>	3	0.0010	—	—	-0.037	-3.67	4.50	0.08
<b>MC + GEO</b>		<b>3</b>	<b>0.0010</b>	—	—	<b>-0.037</b>	<b>-1.42</b>	<b>0</b>	<b>0.80</b>
pPC2	BM	2	0.0027	—	—	—	-4.69	8.55	0.01
	OU	3	0.0027	—	—	—	-4.69	10.67	—
	TDexp	3	0.0046	—	-0.014	—	-4.30	9.89	<0.01
	TDlinear	3	0.0047	-0.011	—	—	-4.23	9.75	<0.01
	LDexp	3	0.0041	—	-0.006	—	-4.27	9.84	<0.01
	<b>LDexp + GEO</b>	<b>3</b>	<b>0.0067</b>	—	<b>-0.039</b>	—	<b>0.65</b>	<b>0</b>	<b>0.84</b>
	LDlin	3	0.0041	-0.00002	—	—	-4.21	9.72	<0.01
	LDlin + GEO	3	0.0035	-0.000035	—	—	-1.57	4.44	0.09
	MC <sub>sym</sub>	3	0.0021	—	—	-9.9e-3	-3.95	9.19	<0.01
<b>MC + GEO</b>		<b>3</b>	<b>0.0019</b>	—	—	<b>-0.014</b>	<b>-2.90</b>	<b>7.10</b>	<b>0.02</b>
pPC3	BM	2	0.0010	—	—	—	45.57	1.4	0.11
	OU	3	0.0010	—	—	—	45.57	3.5	—
	TDexp	3	0.0020	—	-0.019	—	46.30	2.06	0.08
	TDlinear	3	0.0019	-0.013	—	—	46.41	1.85	0.09
	LDexp	3	0.0017	—	-0.008	—	46.40	1.86	0.09
	LDexp + GEO	3	0.0015	—	-0.017	—	46.67	1.33	0.11
	LDlin	3	0.0017	-0.000009	—	—	46.46	1.74	0.09
	LDlin + GEO	3	0.0013	-0.000014	—	—	46.58	1.5	0.10
	MC <sub>sym</sub>	3	0.0007	—	—	-0.012	46.75	1.17	0.12
<b>MC + GEO</b>		<b>3</b>	<b>0.0007</b>	—	—	<b>-0.015</b>	<b>47.33</b>	<b>0</b>	<b>0.22</b>
pPC4	BM	2	0.0006	—	—	—	69.07	1.96	0.07
	OU	3	0.0006	—	—	—	69.07	4.09	—
	TDexp	3	0.0015	—	-0.015	—	70.55	1.12	0.10
	TDlinear	3	0.0012	-0.011	—	—	70.45	1.32	0.09
	LDexp	3	0.0012	—	-0.0061	—	70.52	1.18	0.10
	<b>LDexp + GEO</b>	<b>3</b>	<b>0.0010</b>	—	<b>-0.019</b>	—	<b>71.11</b>	<b>0</b>	<b>0.17</b>
	LDlin	3	0.0011	-0.00002	—	—	70.39	1.45	0.08
	LDlin + GEO	3	0.0008	-0.000035	—	—	70.66	0.92	0.11
	MC	3	0.0004	—	—	-0.015	71.1	0.02	0.17
<b>MC + GEO</b>		<b>3</b>	<b>0.0004</b>	—	—	<b>-0.013</b>	<b>70.6</b>	<b>1.03</b>	<b>0.10</b>

Figure 1.



562 Figure 2.

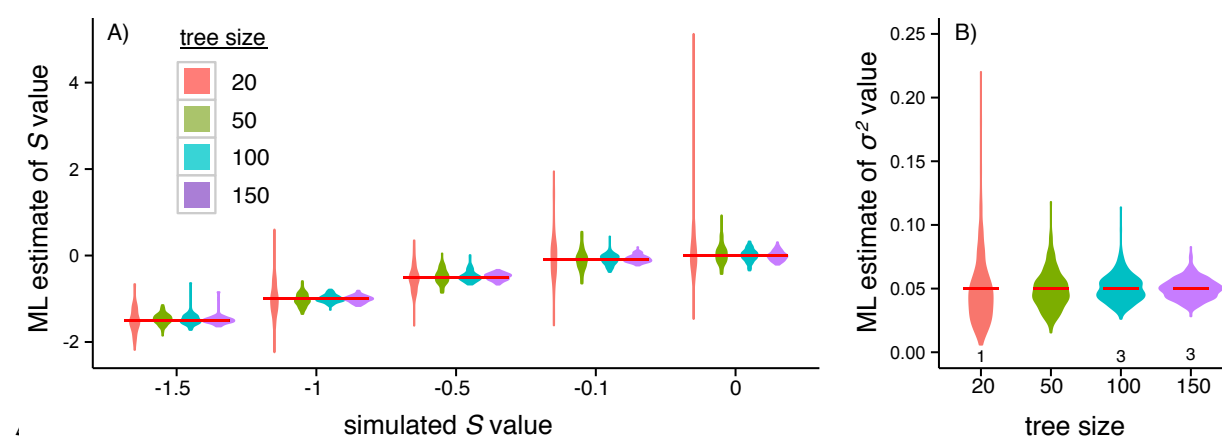


Figure 3

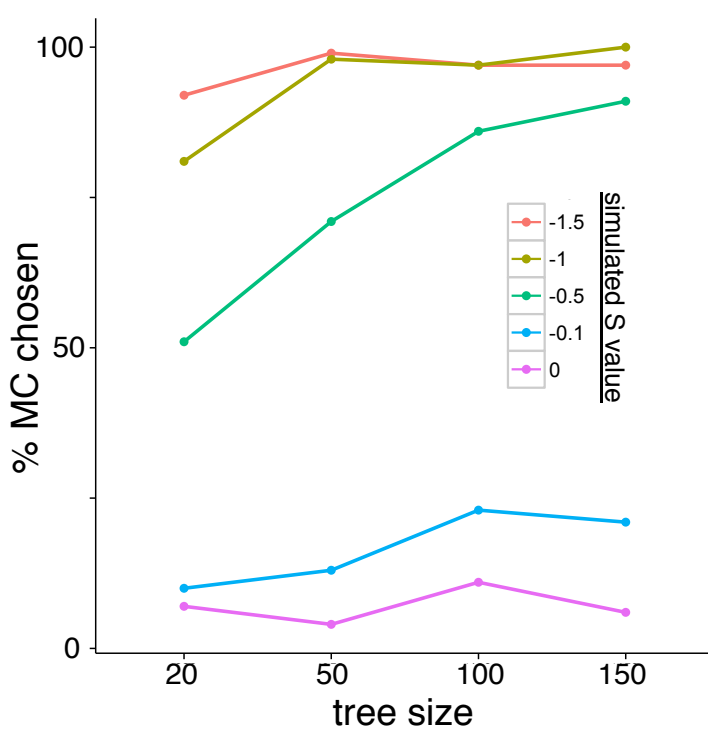
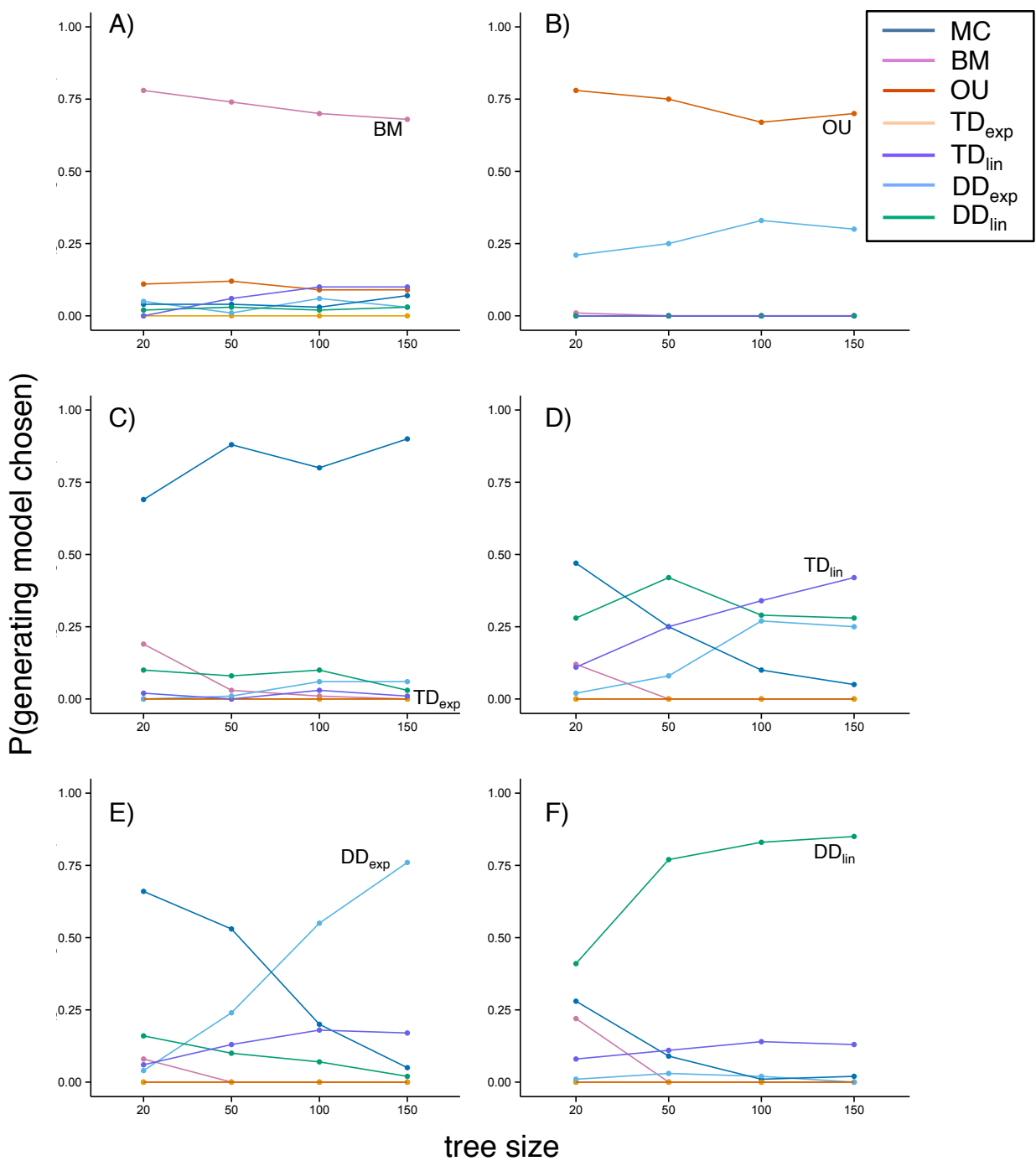




Figure 4.



574 **Supplementary Material**

575

576 Appendix S1

577

578 Table S1

579

580 Figure S1-S6

581

582

583

584

585

# *Supplementary Appendix 1: Estimating the effect of extinction on parameter estimation for the matching competition and density-dependent models.*

Given that the matching competition and diversity-dependent models take into account the number of interacting lineages, extinction may affect our ability to recover true parameter values. To estimate the impact of extinction, we simulated 100 trees with 100 extant species, varying the extinction fraction ( $\mu$ :  $\lambda = 0.2, 0.4, 0.6$ , and  $0.8$ ). As above, we recursively simulated traits using the matching competition model with  $\sigma^2 = 0.05$  and  $S = -1.5, -1, -0.5, -0.1$ , or  $0$ , and the linear and exponential diversity-dependent models with starting rates of  $\sigma^2 = 0.6$  and ending rates of  $\sigma^2 = 0.01$ . We then estimated the maximum likelihood parameter estimates for the generating models by fitting the models to the trait values for extant species and the tree with extinct lineages removed. In the case of the matching competition model, because many simulated birth-death trees with high extinction rates have substantially older root ages, the simulated trait datasets for some trees had very large variances. For these biologically unrealistic trait datasets (i.e., variance in trait values  $\geq 1 \times 10^8$ ), ML does not yield reliable parameter estimates, so we removed them from further analyses (the sample size of included simulations is reported in Fig. S3, S4).

Parameter estimates are quite robust to extinction under the matching competition model (Fig. S3, S4), and much more so than under both diversity-dependent models (Fig. S5, S6). Under the matching competition model, the maximum likelihood optimization returns reliable estimates of  $S$  and  $\sigma^2$  values used to simulate datasets on trees with extinct lineages (Fig. S3, S4), although the estimates become much less reliable with larger extinction fractions, likely because simulations under the matching competition model were unbounded, resulting in trait datasets

609 with biologically unrealistic variances. Under both diversity-dependent models, the magnitude of  
 610 both the slope and  $\sigma^2$  parameter values are increasingly underestimated with increasing  
 611 extinction fractions (Fig. S5, S6).

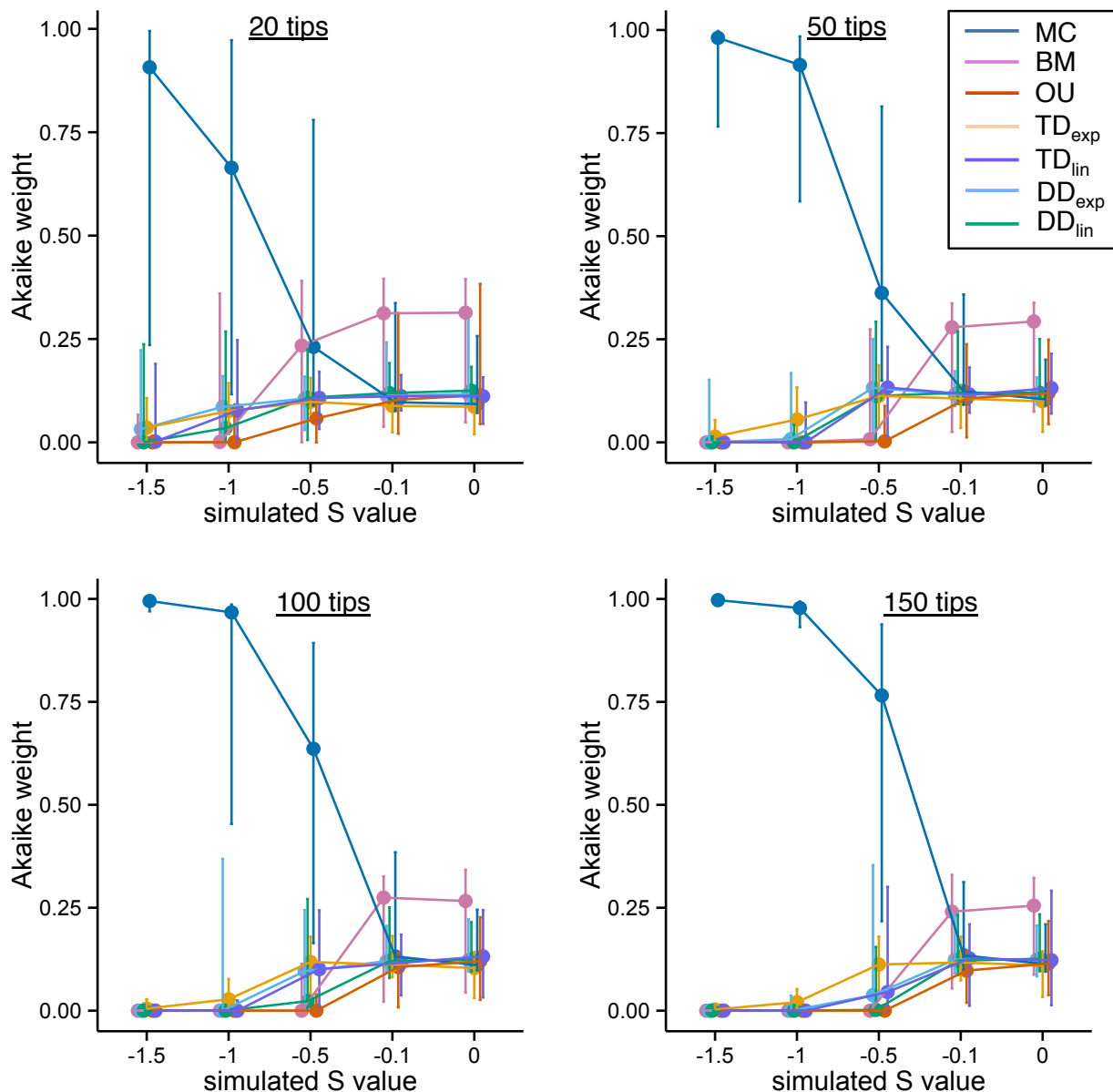
612

613

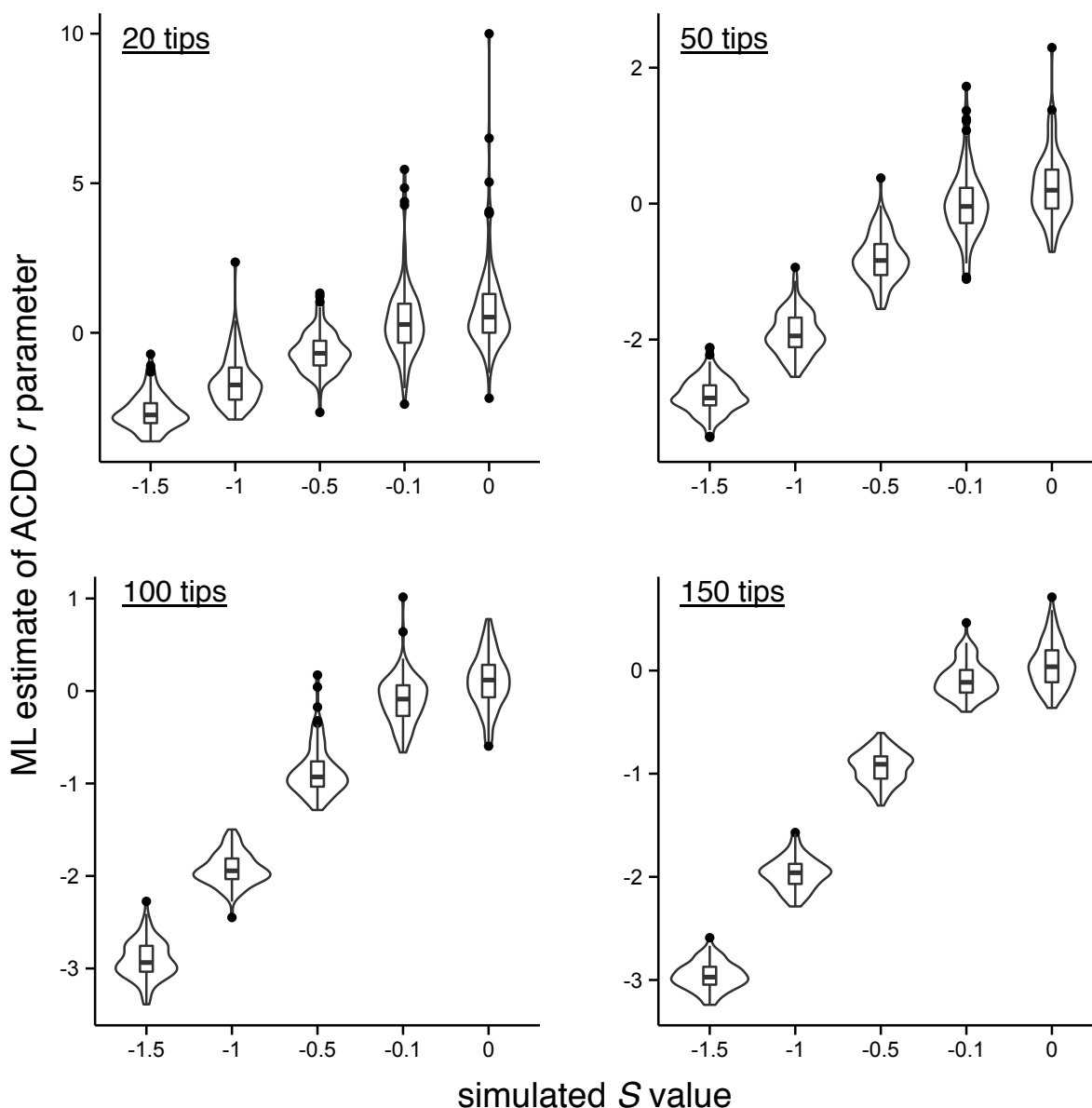
**Table S1.** Data generated under the matching competition model are better fit by the TD<sub>exp</sub> model than by the TD<sub>lin</sub> model.  $\Delta$ AICc [90% CI] presented for the matching competition (MC) and time dependent models.

tips	<i>S</i> value	MC	TD <sub>exp</sub>	TD <sub>lin</sub>
20	-1.5	0 [0, 0.68]	6.04 [3.20, 10.53]	12.17 [1.27, 67.92]
	-1	0 [0, 2.17]	3.96 [1.87, 7.47]	4.18 [0.78, 25.03]
	-0.5	0 [0, 2.76]	2.74 [1.70, 4.06]	2.42 [0.97, 6.36]
	-0.1	2.37 [0, 2.79]	2.79 [1.83, 5.04]	2.1 [0.85, 3.11]
	0	2.47 [0, 2.98]	2.79 [2.17, 6.0]	2.06 [0.88, 3.62]
50	-1.5	0 [0,0]	8.46 [5.57, 12.06]	63.24 [11.75, 216.11]
	-1	0 [0,0]	5.51 [3.45, 8.49]	17.26 [3.50, 67.67]
	-0.5	0 [0, 1.57]	2.53 [1.36, 5.01]	1.87 [0.51, 8.29]
	-0.1	1.83 [0, 2.37]	2.27 [1.44, 4.09]	1.86 [0.51, 3.2]
	0	1.98 [0.09, 2.45]	2.27 [1.37, 4.62]	1.61 [0.05, 3.39]
100	-1.5	0 [0,0]	10.56 [7.11, 13.92]	208.76 [61.32, 442.7]
	-1	0 [0,0]	7.02 [4.93, 8.84]	45.69 [6.44, 149.18]
	-0.5	0 [0, 1.43]	3.34 [1.61, 4.62]	3.67 [0.63, 15.55]
	-0.1	1.60 [0, 2.12]	2.04 [0.99, 2.45]	1.95 [0.61, 4.36]
	0	1.83 [0, 2.11]	2.13 [0.98, 4.41]	1.52 [0, 4.77]
150	-1.5	0 [0,0]	11.77 [8.74, 14.23]	392.29 [177.79, 835.42]
	-1	0 [0,0]	7.63 [5.80, 10.14]	80.58 [27.51, 232.06]
	-0.5	0 [0, 1.0]	3.50 [2.35, 6.11]	5.71 [0.42, 29.24]
	-0.1	1.45 [0, 2.17]	1.83 [0.81, 2.61]	1.6 [0.19, 6.49]
	0	1.81 [0, 2.18]	2.08 [1.02, 4.45]	1.67 [0, 5.98]

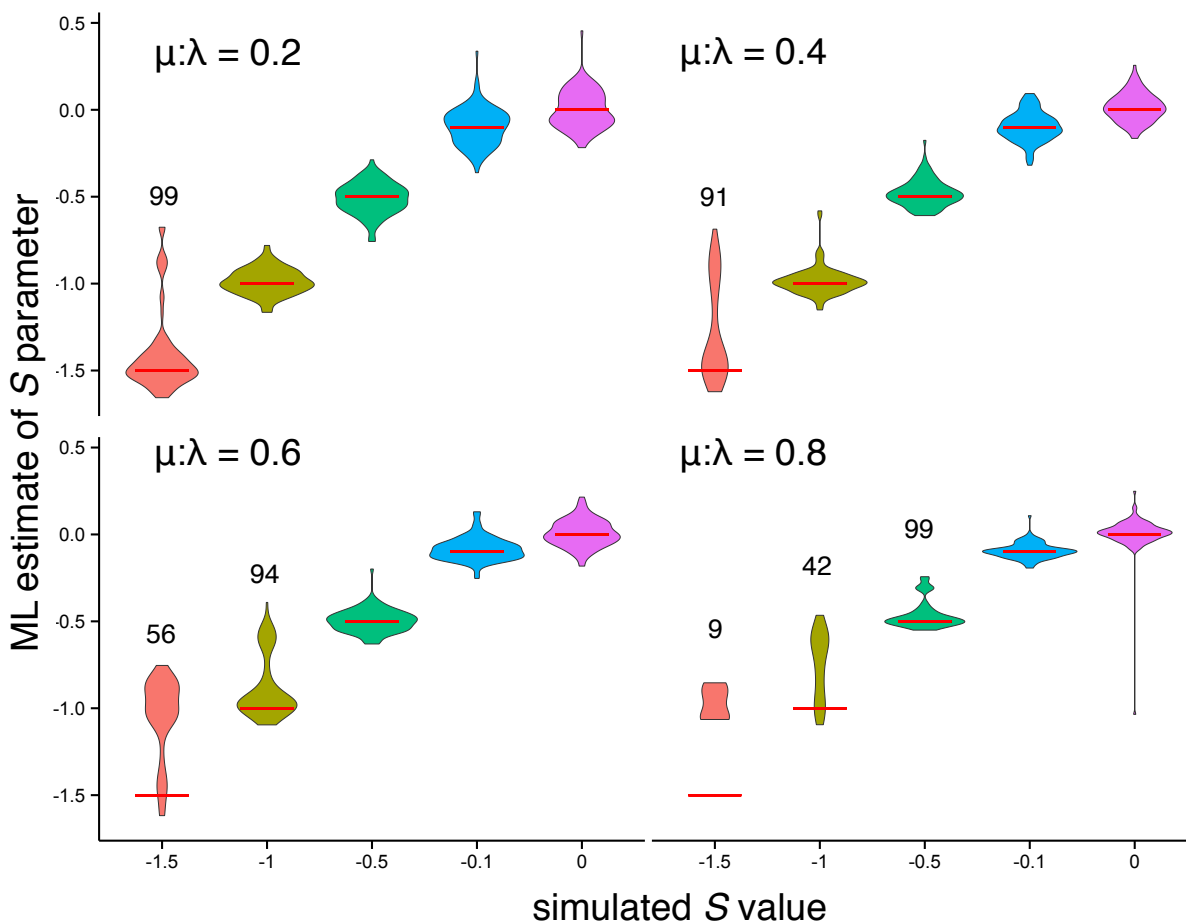
*Supplementary Figure 1.* As tree size and/or the degree of competition ( $S$ ) increases, model selection becomes more reliable. Comparison of Akaike weights (median & 90% CIs) for NH, BM, OU, and EB models when simulated under various levels of competition ( $S = -1.5, -1, -0.5, -0.1$ , and 0) for trees with 20, 50, 100, and 150 tips.



*Supplementary Figure 2.* Maximum likelihood estimates of ACDC rate parameters (i.e., TDexp where rate parameters can take both positive and negative values) fit to data generated under the matching competition model. As the level of competition increases (i.e., as  $S$  becomes increasingly negative), the estimated rate parameter of the ACDC model becomes increasingly negative.

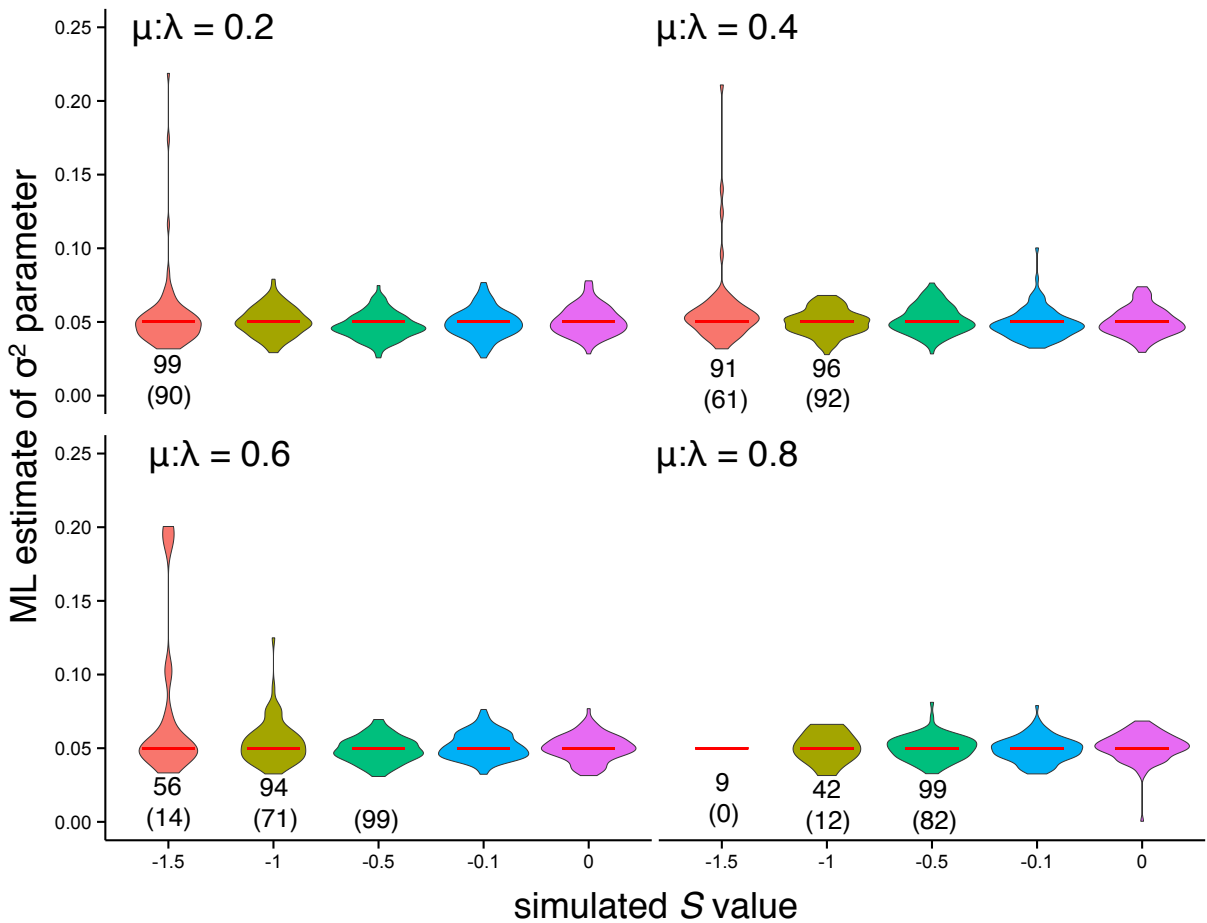


*Supplementary Figure 3.* Simulation results showing the effect of varying the extinction fraction on estimation of the  $S$  parameter for the matching competition model. Red horizontal lines indicate the simulated  $S$  values, and numbers above sets of simulations indicate the sample size of included simulations under those scenarios (see main text for more details).

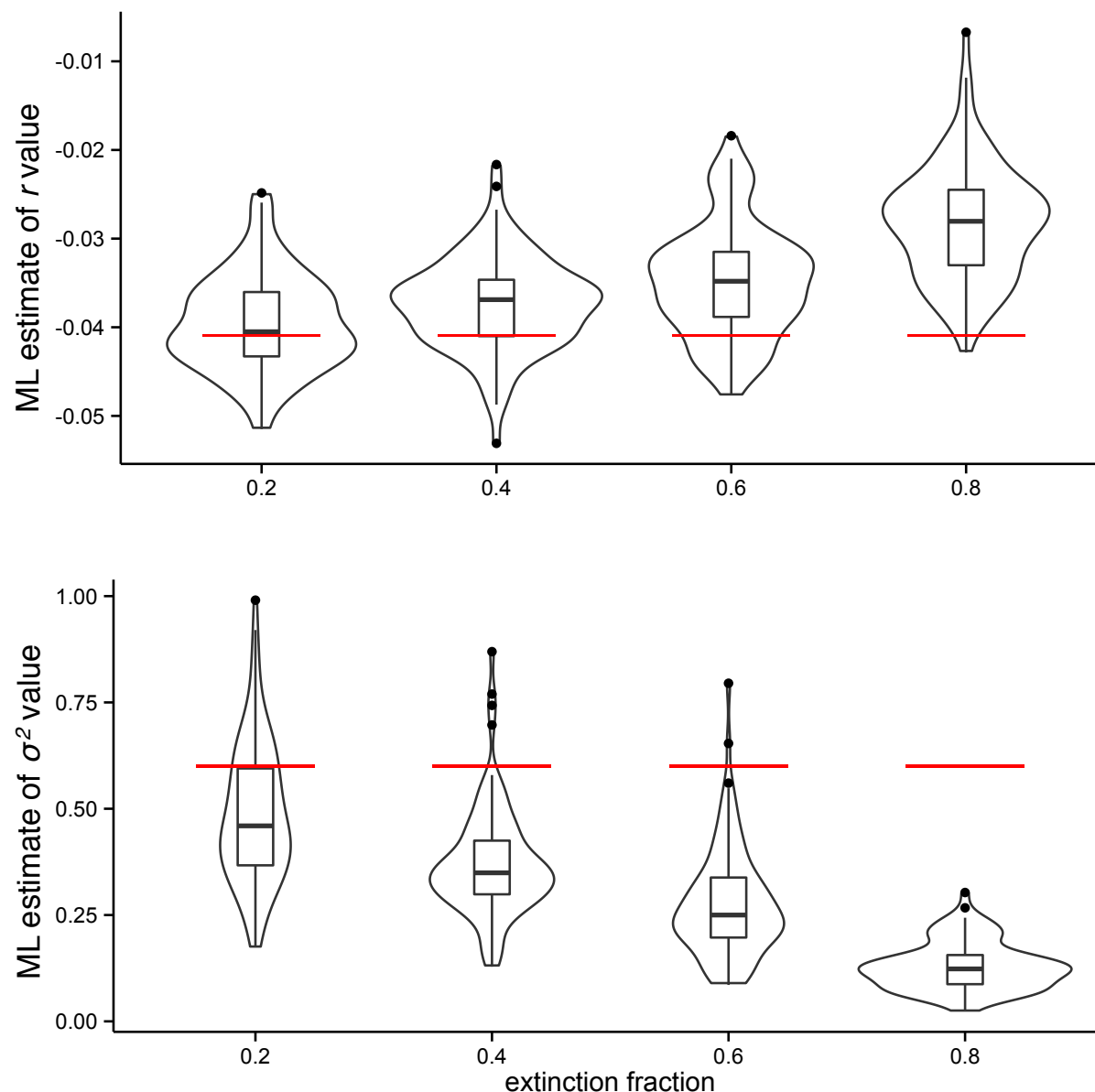




*Supplementary Figure 4. Simulation results showing the effect of varying the extinction fraction on estimation of the  $\sigma^2$  parameter for the matching competition model. Red horizontal lines indicate the simulated  $\sigma^2$  value (0.05), the numbers below sets of simulations indicate the sample size of included simulations under those scenarios (see main text for more details), and the number in parentheses indicate sample size after  $\sigma^2$  values  $> 0.25$  re removed.*



*Supplementary Figure 5.* Simulation results showing the effect of varying the extinction fraction on slope (top) and  $\sigma^2$  (bottom) parameters for the exponential diversity-dependent model. Increasing extinction levels result in increasingly underestimated slope values and  $\sigma^2$  parameters. Red horizontal lines indicate the simulated parameter values.



*Supplementary Figure 6.* Simulation results showing the effect of varying the extinction fraction on slope (top) and sigma-squared (bottom) parameters for the linear diversity-dependent model. Increasing extinction levels result in increasingly underestimated slope values and  $\sigma^2$  parameters. Red horizontal lines indicate the simulated parameter values.

