

1   **Title:**

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3   Phylogenetic analysis supports a link between DUF1220 domain number and primate  
4   brain expansion

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## 35 Abstract

36 The expansion of DUF1220 domain copy number during human evolution is a  
 37 dramatic example of rapid and repeated domain duplication. However, the phenotypic  
 38 relevance of DUF1220 dosage is unknown. Although patterns of expression,  
 39 homology and disease associations suggest a role in cortical development, this  
 40 hypothesis has not been robustly tested using phylogenetic methods. Here, we  
 41 estimate DUF1220 domain counts across 12 primate genomes using a nucleotide  
 42 Hidden Markov Model. We then test a series of hypotheses designed to examine the  
 43 potential evolutionary significance of DUF1220 copy number expansion. Our results  
 44 suggest a robust association with brain size, and more specifically neocortex volume.  
 45 In contradiction to previous hypotheses we find a strong association with postnatal  
 46 brain development, but not with prenatal brain development. Our results provide  
 47 further evidence of a conserved association between specific loci and brain size across  
 48 primates, suggesting human brain evolution occurred through a continuation of  
 49 existing processes.

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## 69 Introduction

70 The molecular targets of selection favoring brain expansion during human evolution  
71 have been sought by identifying dramatic, lineage-specific shifts in evolutionary rate.  
72 The increase in DUF1220 domains during human evolution provides one of the most  
73 dramatic increases in copy number (Popesco et al., 2006; Dumas et al., 2012). A  
74 single copy of this protein domain is found in *PDE4DIP* in most mammalian  
75 genomes. In primates, this ancestral domain has been duplicated many times over,  
76 reaching its peak abundance in humans where several hundred DUF1220 domains  
77 exist across 20-30 genes in the Nuclear Blastoma Breakpoint Family (NBPF)  
78 (Vandepoele et al., 2005; Dumas et al., 2012). The majority of these map to 1q21.1, a  
79 chromosomal region with complex, and unstable genomic architecture (O'Bleness et  
80 al., 2012, 2014).

81 Interspecific DUF1220 counts show a pattern of phylogenetic decay with  
82 increasing distance from humans (Popesco et al., 2006; Dumas and Sikela, 2009;  
83 Dumas et al., 2012). In humans, DUF1220 dosage has also been linked to head  
84 circumference (Dumas et al., 2012), and severe neurodevelopmental disorders,  
85 including autism spectrum disorders (ASD) and microcephaly (Dumas et al., 2012;  
86 Davis et al., 2014). The severity of ASD impairments is also correlated with 1q21.1  
87 DUF1220 copy number suggesting a dosage effect (Davis et al., 2014). Taken  
88 together, these observations led to the suggestion that the expansion of DUF1220  
89 copy number played a primary role in human brain evolution (Dumas and Sikela,  
90 2009; Keeney et al., 2014a).

91 The strength of this hypothesis is difficult to assess given the paucity of  
92 information on the developmental function of NBPF genes and the DUF1220 domain.  
93 DUF1220 domains are highly expressed during periods of cortical neurogenesis,  
94 suggesting a potential role in prolonging the proliferation of neural progenitors by  
95 regulating centriole and microtubule dynamics to control key cell fate switches  
96 critical for neurogenesis (Keeney et al., 2014b). *PDE4DIP*, which contains the  
97 ancestral DUF1220 domain, does indeed associate with the spindle poles (Popesco et  
98 al., 2006) and is homologous to *CDK5RAP2*, a centrosomal protein essential for  
99 neural proliferation (Bond et al., 2005; Buchman et al., 2010), which co-evolved with  
100 brain mass across primates (Montgomery et al., 2011).

101 Two previous analyses reported a significant association between DUF1220  
102 copy number and brain mass, cortical neuron number (Dumas et al., 2012), cortical

gray and white matter, surface area and gyrification (Keeney et al., 2014b). However, several limitations in these analyses restrict confidence in the results. First, DUF1220 copy number was assessed across species using a BLAT/BLAST analysis with a query sequence from humans, which introduces a bias that may partly explain the observed phylogenetic decay. Second, counts were not restricted to those domains occurring in functional exonic sequence. Finally, the analyses were limited to a small number of species (4-8 primates), and did not correct for phylogenetic non-independence (Felsenstein, 1985) or autocorrelation between traits.

Here, we use nucleotide Hidden Markov Models (HMMER3; Eddy, 2011) to more accurately query the DUF1220 domain number of distantly related genomes. After filtering these counts to limit the analysis to exonic sequence, we use phylogenetic comparative methods that correct for non-independence to test whether DUF1220 copy number is robustly associated with brain size, whether this is due to an association with pre- or postnatal brain development, and whether the association is specific to the neocortex.

## Results

We find evidence that CM-associated exonic DUF1220 counts (Table 1) are associated with brain mass across primates ( $n = 12$ , posterior mean = 1.927, 95% CI = 0.800-3.040,  $p_{\text{MCMC}} = 0.001$ ). This association is robust to the exclusion of *Homo* (posterior mean = 1.271, 95% CI = 0.490-2.019,  $p_{\text{MCMC}} = 0.003$ ), and found when hominoids ( $n = 5$ , posterior mean = 3.679, 95% CI = 0.966-6.258,  $p_{\text{MCMC}} = 0.018$ ) or anthropoids ( $n = 9$ , posterior mean = 2.019, 95% CI = 0.352-3.684,  $p_{\text{MCMC}} = 0.010$ ) are analyzed alone, suggesting a consistent phylogenetic association. When body mass is included as a co-factor in the model, the positive association is restricted to brain mass (Table 2a).

Separation of pre- and postnatal development specifically links DUF1220 number to postnatal brain growth. Analysed separately, the association with prenatal brain growth is weaker ( $n = 11$ , posterior mean = 1.758, 95% CI = -0.039-3.543,  $p_{\text{MCMC}} = 0.023$ ) than with postnatal brain growth (posterior mean = 1.839, 95% CI = 0.895-2.808,  $p_{\text{MCMC}} = 0.001$ ). If both traits are included in the same model, only the positive association with postnatal brain growth remains (Table 2b). Multiple

136 regression analysis also confirms the association is specific to postnatal brain growth,  
137 rather than postnatal body growth (Table 2b).

138 Finally, we examined the hypothesized relationship with neocortex volume  
139 (e.g. Keeney et al., 2014a,b), but also consider cerebellum volume, as this region co-  
140 evolves with the neocortex (Barton and Harvey, 2000), has expanded in apes (Barton  
141 and Venditti, 2014), and shows high levels of NBPF expression (Popesco et al.,  
142 2006). When the rest-of-the-brain (RoB) is included as a co-factor, to account for  
143 variation in overall brain size, a positive association is found for neocortex volume  
144 but not cerebellum volume (Table 2c).

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## 146 **Discussion**

147 Our phylogenetic analyses support the hypothesis that the increase in DUF1220  
148 number co-evolves with brain mass, and contributes to the proximate basis of primate  
149 brain evolution. We also find evidence of specific associations with neocortex volume  
150 and postnatal brain growth. Previous hypotheses concerning the phenotypic relevance  
151 of DUF1220 domain number have focused on their possible contribution to  
152 neurogenesis (Dumas and Sikela, 2009; Keeney et al., 2014a; b). This is supported by  
153 homology to genes with known functions in cell cycle dynamics (Popesco et al.,  
154 2006; Thornton and Woods, 2009), relevant spatial and temporal expression patterns  
155 (Keeney et al., 2014b), and an effect on the proliferation of neuroblastoma cell  
156 cultures (Vandepoele et al., 2008). However, a direct effect of variation in DUF1220  
157 domain number on neural proliferation has not been demonstrated (Keeney et al.,  
158 2015).

159 If DUF1220 domains do regulate neurogenesis, we would expect them to co-  
160 evolve with prenatal brain growth, as cortical neurogenesis is restricted to prenatal  
161 development (Bhardwaj et al., 2006). Our results instead suggest a robust and specific  
162 relationship with postnatal brain development. Existing data on DUF1220 domain  
163 function suggest two potential roles that may explain this association: i) a contribution  
164 to axonogenesis via initiating and stabilizing microtubule growth in dendrites; and ii)  
165 a potential role in apoptosis during brain maturation. Both hypotheses are consistent  
166 with the reported association between variation in DUF1220 dosage and ASD (Davis  
167 et al., 2014). Indeed, an emphasis on postnatal brain growth is potentially more  
168 relevant for ASD, which develops postnatal, accompanied by a period of accelerated  
169 brain growth (Courchesne et al., 2001).

170 Microtubule assembly is essential for dendritic growth and axonogenesis  
 171 (Conde and Cáceres, 2009). *PDE4DIP*, which contains the ancestral DUF1220  
 172 domain, has known functions in microtubule nucleation, growth, and cell migration  
 173 (Roubin et al., 2013). There is also evidence NBPf1 interacts with a key regulator of  
 174 Wnt signaling (Vandepoele et al., 2010), which has important roles in neuronal  
 175 differentiation, dendritic growth and plasticity (Inestrosa and Varela-Nallar, 2014).  
 176 Consistent with this function, DUF1220 domains are highly expressed in the cell  
 177 bodies and dendrites of adult neurons (Popesco et al., 2006). A role for DUF1220  
 178 domains in synaptogenesis could potentially explain the association with ASD  
 179 severity (Davis et al., 2014). ASDs are associated with abnormalities in cortical  
 180 minicolumns (Casanova et al., 2002) and cortical white matter (Hazlett et al., 2005;  
 181 Courchesne et al., 2011), both of which suggest a disruption of normal neuronal  
 182 maturation (Courchesne and Pierce, 2005; Minshew and Williams, 2007).

183 Alternatively, NBPf genes are also known to interact with NF- $\kappa$ B (Zhou et al.,  
 184 2013), a transcription factor implicated in tumor progression, with a range of roles  
 185 including apoptosis and inflammation (Karin and Lin, 2002; Perkins, 2012). Postnatal  
 186 apoptosis has a significant influence on brain growth (Kuan et al., 2000; Polster et al.,  
 187 2003; Madden et al., 2007), including regulating neuronal density (Sanno et al.,  
 188 2010), and apoptotic genes may have been targeted by selection in relation to primate  
 189 brain expansion (Vallender and Lahn, 2006). Disruption of apoptosis causes  
 190 microcephaly (Poulton et al., 2011), potentially explaining the association between  
 191 DUF1220 dosage and head circumference (Dumas et al., 2012). The association of  
 192 NF- $\kappa$ B with inflammatory diseases (Tak et al., 2001) is also intriguing, given the  
 193 growing evidence that the inflammatory response is linked to the risk and severity of  
 194 ASD (Meyer et al., 2011; Depino, 2012).

195 If DUF1220 domain number does contribute to the evolution of postnatal  
 196 brain growth, this contrasts with results of previously studied candidate genes with  
 197 known roles in neurogenesis that co-evolve with prenatal brain growth (Montgomery  
 198 et al., 2011). This suggests a two-component model of brain evolution where selection  
 199 targets one set of genes to bring about an increase in neuron number (e.g.  
 200 Montgomery et al., 2011; Montgomery and Mundy, 2012a;b), and an independent set  
 201 of genes to optimize neurite growth and connectivity (e.g. Charrier et al., 2012).  
 202 NBPf genes may fall into the latter category. This two-component model is consistent  
 203 with comparative analyses that indicate pre- and postnatal brain development evolve

independently, and must therefore be relatively free of reciprocal pleiotropic effects (Barton and Capellini, 2011).

Finally, these results add further evidence that many of the genetic changes that contribute to human evolution will be based on the continuation or exaggeration of conserved gene-phenotype associations that contribute to primate brain evolution (Montgomery et al., 2011; Scally et al., 2012). Understanding the commonalities between human and non-human primate brain evolution is therefore essential to understand the genetic differences that contribute the derived aspects of human evolution.

## Materials and methods

### *Counting DUF1220 domains*

HMMER3.1b (Eddy, 2011) was used to build a Hidden Markov Model (HMM) from the DUF1220 (PF06758) seed alignment stored in the PFAM database (Finn et al., 2014). The longest isoforms for all proteomes of 12 primate genomes from Ensembl v.78 (Cunningham et al., 2014) (Figure 1A), were searched using the protein DUF1220 HMM (hmmsearch, E-value < 1e-10) (Table S1). We extracted the corresponding cDNA regions to build a DUF1220 nucleotide profile HMM (nHMM), allowing for more sensitive analysis across a broad phylogenetic range. The DUF1220 nHMM was used to search the complete genomic DNA for all 12 species. These counts were filtered to remove any DUF1220 domains not located in annotated exonic sequence, or located in known pseudogenes.

We next filtered our counts to limit them to exonic sequence in close proximity to the NBPF-specific Conserved-Mammal (CM) promoter (O’Bleness et al. 2012). To do so, we built a nucleotide HMM for the CM promoter based on a MAFFT (Kato et al., 2002) alignment of the 900bp CM region upstream of human genes NBPF4, NBPF6 and NBPF7. Using this CM promoter nHMM, we searched 1000bp up- and downstream of genes containing DUF1220 domains for significant CM promoter hits (nhmmer, E-value < 1e-10). This provided final counts for DUF1220 domains within exonic regions and associated with the CM promoter (Table 1). These counts were used in subsequent phylogenetic analyses. In the Supplementary Information we compare our counts with previous estimates and discuss possible sources of error. All scripts and data used in the analysis are freely available from: <https://github.com/qfma/duf1220>

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### 239 ***Phylogenetic gene-phenotype analysis***

240 Phylogenetic multivariate generalized mixed models were implemented using a  
 241 Bayesian approach in MCMCglmm (Hadfield, 2010), to test for phylogenetically-  
 242 corrected associations between DUF1220 counts and *log*-transformed phenotypic data  
 243 (Table S2). All analyses were performed using a Poisson distribution, as  
 244 recommended for count data (O’Hara and Kotze, 2010), with uninformative,  
 245 parameter expanded priors for the random effect (G:  $V = 1, n \nu = 1, \alpha.\nu = 0,$   
 246  $\alpha.V = 1000$ ; R:  $V = 1, \nu = 0.002$ ) and default priors for the fixed effects.  
 247 Phylogenetic relationships were taken from the 10k Trees project (Arnold et al.,  
 248 2010). We report the posterior mean of the co-factor included in each model and its  
 249 95% confidence intervals (CI), and the probability that the parameter value is  $>0$   
 250 ( $p_{MCMC}$ ) as we specifically hypothesize a positive association (Dumas et al., 2012).  
 251 Alternative data treatments lead to similar conclusions (Supplementary Information).

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417 **Tables**

418

419 **Table 1:** DUF1220 count data

Species	O'Bleness et al. (2012)	nHMM	
		whole genome	functional exonic with CM promoter
<i>Homo sapiens</i>	272	302	262
<i>Pan troglodytes</i>	125	138	34
<i>Gorilla gorilla</i>	99	97	32
<i>Pongo abelii</i>	92	101	27
<i>Nomascus leucogenys</i>	53	59	6
<i>Papio anubis</i>	-	75	15
<i>Chlorocebus sabaeus</i>	-	48	16
<i>Macaca mulatta</i>	35	74	10
<i>Callithrix jacchus</i>	31	75	9
<i>Tarsius syrichta</i>	-	47	2
<i>Microcebus murinus</i>	2	4	1
<i>Otolemur garnettii</i>	3	4	1

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**Table 2: MCMCglmm results of multivariate models**

**a) Brain mass and body mass**

Model	Posterior mean	95% CI	pMCMC
1. log(brain mass)	4.105	2.163 - 6.000	0.001
+ log(body mass)	-1.986	-3.544 - -3.900	0.988

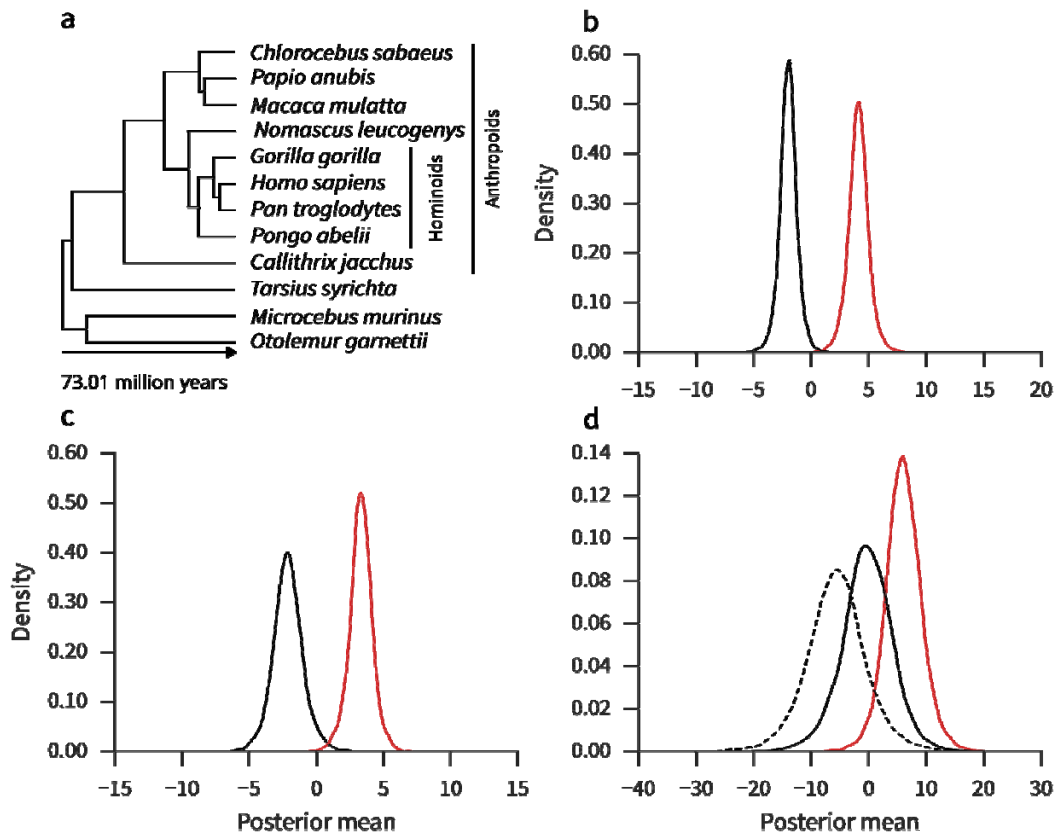
**b) Prenatal and postnatal growth**

Model	Posterior mean	95% CI	pMCMC
1. log(prenatal brain growth)	-2.158	-4.471 - 0.106	0.967
+ log(postnatal brain growth)	3.319	1.470 - 4.982	0.002
2. log(postnatal brain growth)	2.910	1.641 - 4.151	<0.001
+ log(postnatal body growth)	-1.241	-2.442 - -0.052	0.977

**c) Brain regions**

Model	Posterior mean	95% CI	pMCMC
1. log(neocortex volume)	5.961	0.720 - 11.173	0.014
+ log(RoB volume)	-5.817	-13.322 - 1.120	0.953
2. log(cerebellum volume)	3.699	-5.857 - 12.611	0.186
+ log(RoB volume)	-2.435	-13.869 - 10.132	0.681
3. log(neocortex volume)	6.076	-0.139 - 12.5712	0.025
+ log(cerebellum volume)	-0.369	-9.5128 - 8.961	0.526
+ log(RoB volume)	-5.494	-15.814 - 5.288	0.872

# Figures



**Figure 1:** A) Phylogeny of Ensembl primates. B) Posterior means of the association between DUF1220 count and brain mass (red) and body mass (black). C) Posterior means of the association between DUF1220 count and postnatal brain growth (red) and prenatal brain growth (black). D) Posterior means of the association between DUF1220 count and neocortex volume (red), cerebellum volume (solid black) and rest-of-brain volume (dashed black).