Selection against maternal microRNA target sites in maternal transcripts

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ABSTRACT

In animals, before the zygotic genome is expressed, the egg already contains gene products deposited by the mother. These maternal products are crucial during the initial steps of development. In *Drosophila melanogaster* a large number of maternal products are found in the oocyte, some of which are indispensable. Many of these products are RNA molecules, such as gene transcripts and ribosomal RNAs. Recently, microRNAs – small RNA gene regulators – have been detected early during development and are important in these initial steps. The presence of some microRNAs in unfertilized eggs has been reported, but whether they have a functional impact in the egg or early embryo has not being explored. I have extracted and sequenced small RNAs from *Drosophila* unfertilized eggs. The unfertilized egg is rich in small RNAs and contains multiple microRNA products. Maternal microRNAs are often encoded within the intron of maternal genes, suggesting that many maternal microRNAs are the product of transcriptional hitch-hiking. Comparative genomics and population data suggest that maternal transcripts tend to avoid target sites for maternal microRNAs. A potential role of the maternal microRNA mir-9c in maternal-to-zygotic transition is also discussed. In conclusion, maternal microRNAs in *Drosophila* have a functional impact in maternal protein-coding transcripts.

INTRODUCTION

In animals, the initial steps of embryonic development are driven by the gene products deposited by the mother in the egg. For instance, in *Drosophila melanogaster*, the anteroposterior axis is determined by the presence of maternal transcripts from genes such as *bicoid* and *nanos* (LAWRENCE 1992). Traditionally, maternal genes have been identified by genetic analysis (LAWRENCE 1992). More recently, the roles of microRNAs during development have become a major research area. MicroRNAs are small RNA molecules that regulate gene expression by targeting gene transcripts by sequence complementarity. MicroRNAs are expressed during early development (ARAVIN *et al.* 2003; ABOOBAKER *et al.* 2005), and they target other embryonic expressed gene transcripts (ENRIGHT *et al.* 2003; LAI *et al.* 2003). As a matter of fact, a number of homeotic genes detected by genetic analysis were later shown to be microRNA encoding genes (reviewed in (MARCO 2012)). The characterization of maternal microRNAs is particularly difficult as they are too short for standard genetic analyses. However, it is now possible to isolate small RNAs from egg extracts and characterize their products with RNAseq or microarray technologies. For instance, the microRNA content of mouse (TANG *et al.* 2007) and cow (TESFAYE *et al.* 2009) oocytes have been characterized with this high-throughput approach. In other cases, such as in zebrafish (CHEN *et al.* 2005) and *Xenopus* (WATANABE *et al.* 2005), microRNAs appear to have a minor presence in oocytes.

Several lines of evidence suggested that, in *Drosophila*, maternally transmitted microRNAs are important. First, some microRNAs are highly abundant during early development (RUBY *et al.* 2007). Also, the enzymes responsible for microRNA biogenesis are present in the ovaries (ROBINSON *et al.* 2013) and microRNAs may have a role in oocyte maturation (NAKAHARA *et al.* 2005). Indeed, mature microRNAs have been identified in *Drosophila* unfertilized eggs (LEE *et al.* 2004, 2014; VOTRUBA 2009). Recently, it has been shown that maternally transmitted microRNAs are adenylated during the maternal-to-zygotic transition (MZT) (LEE *et al.* 2014) .Whether maternal

microRNAs have a functional impact in *Drosophila* eggs is still unknown. To identify which microRNAs are maternally transmitted I extracted and sequenced small RNAs from *Drosophila* unfertilized eggs. To explore their potential function I predicted their targets in maternal and non-maternal gene products. The evolutionary impact of maternal microRNAs was estimated by using comparative genomics and population data.

MATERIALS AND METHODS

Flies and egg collection: Fly stocks used in this study, with Bloomington reference number in square brackets, were: w^{1118} [#3605] and Oregon-R-modENCODE [#25221]. All flies were kept at 25ºC on cornmeal based media, with 12 hours light/dark cycles. Virgin females were sorted at the pupae stage to avoid any unwanted fertilization. (Previous attempts selecting for <6 hours females produced a small yet significant number of fertilized eggs). In a population cage I let 80-100 females to lay eggs in apple juice agar plates for 8 hours, collecting 1 hour after dawn. Eggs were collected with a sieve and washed with saline solution. Eggs from virgin females do not degenerate even several hours after laying (TSIEN and WATTIAUX 1971).

RNA extraction, sequencing and profiling: Total RNA was extracted from eggs or early embryos with TRIzol reagent (Life Technologies), following instructions given by the manufacturer, and dissolving the RNA in RNase-free water. For RNA sequencing, a cDNA library was generated with TruSeq Small RNA Sample Preparation Kit (Illumina). Amplified cDNA constructs were size selected in a 6% PAGE gel for 145 to 160 bp (fragments including RNA-derived sequences of size ~20-30 bp plus adapters). Size selected cDNAs were purified and precipitated with ethanol, and DNA integrity was checked with TapeStation (Agilent). Samples were sequenced with Illumina MiSeq in the Genomics Core Facility at the University of Manchester. A total of 4,507,291 reads

were sequenced, most of them (95.5%) deriving from ribosomal RNAs which is expected in *Drosophila* where the majority of small RNAs are 2S rRNA (SEITZ *et al.* 2008). 13,114 reads were identified as microRNA products. Sequence reads are available from Gene Expression Omnibus (GEO) at NCBI (accession number: GSE63488).

Illumina MiSeq produces 50 bp sequence reads. Hence, I removed adapters with Cutadapt (https://code.google.com/p/cutadapt/) and mapped the processed reads of size 18-26 bp to known microRNAs from miRBase v.20 (KOZOMARA and GRIFFITHS-JONES 2014), using Bowtie v.0.12.7 (LANGMEAD *et al.* 2009), allowing no mismatches and considering reads mapping to up to five positions. Other RNA collections from embryos and ovaries were also analysed: 0-1h embryos, 2-6h embryos, 6-10 h embryos (RUBY *et al.* 2006) and ovaries (CZECH *et al.* 2008). Expression profiling in Figure 1 was done with R, scaling the Z-scores of the heatmap across rows, and generating a hierarchical tree of microRNAs with complete linkage clustering.

Validations were made with Mir-X first-strand synthesis and SYBR qRT-PCR assays manufactured by Clontech Laboratories, Inc. MicroRNA cDNA llibraries were constructed for unfertilized eggs and 2-6 hours old embryos from Oregon-R flies, following manufacturer's indications. Primers for microRNA-specific amplification during qPCRs were: let-7-5p (5'-TGAGGTAGTAGGTTGTATAGT-3'), miR-34-5p (5'-TGGCAGTGTGGTTAGCTGGTTGTG-3'), miR-311-3p (5'-TATTGCACATTCACCGGCCTGA-3'), mir-92b-3p (5'-AATTGCACTAGTCCCGGCCTGC-3'), miR-184-3p (5'-TGGACGGAGAACTGATAAGGGC-3'), miR-9c-5p (5'-TCTTTGGTATTCTAGCTGTAGA-3'), bantam-3p (5'-TGAGATCATTTTGAAAGCTGATT-3'), miR-995-3p (5'-TAGCACCACATGATTCGGCTT-3'), and miR-14-3p (5'-TCAGTCTTTTTCTCTCTCCTAT-3'). Fluorescent quantification was done in a LightCycler 96 Real-Time PCR System (Roche) for 50 cycles, Cts were estimated with the software provided by the manufacturer with default parameters, and ΔCts calculated using U6 spliceosomal rRNA as a normalization standard. Relative expression values in Figure 2 for unfertilized eggs were

calculated with respect to the average bantam level. That is: $[miR]/[bantam]=2^{-\Delta Ct(miR)}/2^{-\Delta Ct(bantam)}$. For 2-6 hour embryos, the relative levels are calculated with respect to the levels in egg samples. Each amplification was performed for three biological replicated (independent egg/embryo collections) with two technical replicates each.

MicroRNA target analysis and polymorphisms: Target analysis was based on the presence of canonical seeds in the transcripts (BARTEL 2009). Canonical seed predictions have the advantage that only primary sequence information is used, so populations models (see below) can be easily fitted. Maternally deposited gene transcripts are listed in the Berkeley Drosophila Genome Project webpage at http://insitu.fruitfly.org (TOMANCAK *et al.* 2007). Which transcripts are destabilized during the maternal-to-zygotic transition were identified from Tadros et al. microarray experiments (Gene Expression Omnibus accession number GSE13287), detecting probes with a >1.5 fold change in their expression level between 4-6 h embryos and oocytes (TADROS *et al.* 2007). To assess whether maternal microRNAs target transcripts that are destabilized during the MTZ transition I calculated the proportion of unstable transcripts targeted by each microRNA and compared it to the expected proportion (0.146) with a cumulative binomial test. False Discovery Rate was accounted by calculating q-values associated to the p-values (BENJAMINI and HOCHBERG 1995; STOREY 2002).

For the population analyses, I first mapped the single-nucleotide polymorphisms (SNPs) from the Drosophila Genetic Reference Panel (MACKAY *et al.* 2012; HUANG *et al.* 2014), available at http://dgrp2.gnets.ncsu.edu/, against the 3'UTR of *Drosophila melanogaster* release 5.13 (http://flybase.org). For each microRNA I defined a target sequence (sixmer) and its 18 non-target neighbours, that is, the 18 one-nucleotide variations of the target site (Figure 3A). Every SNP that connects a target with a non-target sixmer was further considered. 3'UTRs with introns were discarded. For each polymorphic target site, the allele frequency distribution was calculated as the proportion of the target allele with respect to the total number of sampled individuals (isogenic

lines). In this analysis, both target and non-target alleles are considered in the reference genome. To identify alleles conserved in *Drosophila sechellia* I first map polymorphic target sites from *D. melanogaster* genome release 5.13 onto release 6 using the coordinate converter in FlyBase, and then finding the conserved sites in *D. sechellia* by parsing the genome sequence alignment files available at UCSC Genome Browser (ftp://hgdownload.cse.ucsc.edu/goldenPath/dm6/multiz27way; SIEPEL *et al.* 2005) using custom-made PERL scripts.

RESULTS

Mature microRNAs are maternally deposited in the egg: To characterize maternal microRNAs in *Drosophila* I first characterized RNAs from unfertilized eggs with high-throughput sequencing (see Materials and Methods). The most abundant microRNAs in unfertilized eggs were produced by *mir-92b*, *mir-184*, the *mir-310/mir-311/mir-312/mir-313* cluster and *bantam* genes, which account for over a half of the microRNA reads. Table 1 shows microRNA loci producing more than 13 reads (1‰ of the microRNA-associated reads). A full list of detected microRNAs with their read counts is in Table S1. The dataset was screened for new microRNAs as previously described (MARCO *et al.* 2010, 2013b; MARCO and GRIFFITHS-JONES 2012), but no new microRNAs were found. This tell us that maternal microRNAs are already known in *Drosophila*.

In a recent report, Narry Kim and collaborators identified maternally transmitted microRNAs in *Drosophila* and demonstrated that they are targeted to degradation during maternal-to-zygotic transition (MTZ) by adenylation via Wispy (LEE *et al.* 2014). Their set of maternal microRNAs is virtually identical to the set here described. Overall, the read counts from both datasets are highly correlated (R^2=0.62; p<0.001, Supplementary Figure S1A). The overlap for the top N-th most abundant microRNAs between both datasets is highly significant (Supplementary Figure S1B). Specifically, the microRNAs here described as maternal in Table 1 (more than 13

reads) are the top 35 mature sequences, which overlaps with 26 microRNAs from Lee et al's top 35 microRNAs (74.3%; p=0.00031, Supplementary Figure S1B). Additionally, the read counts form this study and a recent report by NINOVA *et al.* (2015) , which uses the same protocol for RNA extraction and sequencing, are highly correlated (R^2=0.86; p<0.001; Supplementary Figure S1C). All these observations support the high confidence of the maternal microRNA set here described.

Figure 1 compares the relative expression of maternal microRNAs in the ovary, unfertilized eggs and early stages of development. From this comparison three types of maternal microRNAs can be distinguished. First, some maternal microRNAs are highly expressed in the ovary. A second class consists on microRNAs that are found primarily in the unfertilized egg. Third, a large proportion of maternal microRNAs is also transcribed later on during development. These groups are referred hereinafter as 'high in ovary', 'high in egg' and 'high in zygote' maternal microRNAs. Some of these microRNAs were detected at very low levels, and whether they are bona fide maternal microRNAs may need further evidence.

To further confirm the presence of maternal microRNAs in unfertilized eggs, I validated the presence of highly abundant mature products by qPCR (see Materials and Methods). Figure 2 shows the relative abundance of selected microRNAs (with respect to the average level of *bantam* mature product). Although the microRNA level varies substantially across biological replicates, the presence of 7 of the maternal microRNAs here described is validated (bantam-3p, mir-311-3p, mir-92b-3p, mir-184-3p, mir-14-3p, mir-995-3p and mir-9c-5p), although the levels of the latter two were relatively low. Furthermore, the level of mir-34-5p, which has been reported to be maternally transmitted (SONI *et al.* 2013), was very low, in agreement with this and other investigations (see Discussion). The conserved microRNA let-7-5p was used as a negative control, as it was not detected in unfertilized eggs. In the qPCR analysis, let-7-5p was not detected in unfertilized eggs (Figure 2). I further measured the relative levels of maternal microRNAs in later stage embryos (2-6 hours). In concordance to the deep sequencing analysis presented in Figure 1, bantam-3p, mir-311-

3p and mir-92-3p were more abundant in the unfertilized egg than in the developing embryo. On the other hand, mir-14-3p was higher expressed in the embryo than in the egg. However, for mir-184-3p and mir-995-3p the pattern was not consistent between RNAseq and qPCR. The differences were not significant. Both mir-34-5p and let-7-5p were highly abundant in developing embryos, further supporting that they are virtually absent from the unfertilized egg.

Intronic maternal microRNAs hosted in maternal protein-coding genes: In a previous work I observed that female biased microRNAs tend to be produced from introns of female biased protein coding transcripts (MARCO 2014). For instance, mir-92a is highly expressed in females, and it is encoded within the *jigr1* gene, which is maternally deposited in the egg. Here I show that mir-92a is also maternal. To further explore the relationship between maternal microRNAs and the maternal deposition of overlapping genes, I compared the expression pattern of intronic maternal microRNAs and the host protein coding gene. Table 2 lists 12 maternal microRNA clusters hosted in protein coding genes. For nine of these host genes there are in situ hybridization experiments (TOMANCAK *et al.* 2002, 2007), and eight of them are maternally loaded. Since 55.8% of genes in this dataset are shown to produce maternally deposited transcripts, our set of host genes is statistically enriched for maternal products ($p \sim 0.044$; binomial test). There is no information from high-throughput in situ hybridization analyses for *grp*, but it is known to be present in unfertilized oocytes (FOGARTY *et al.* 1997). The other two host genes have no expression information available at FlyBase. From this analysis I conclude that intronic maternal microRNAs are frequently produced from introns of maternally deposited gene transcripts.

Maternal microRNAs in the Maternal-to-Zygotic Transition: As shown in Figure 1, a significant fraction of maternal microRNAs have a lower expression when zygotic transcription starts. One possibility is that some of these maternal microRNAs have a role in destabilizing

maternal transcripts during the MZT. A similar role has been described for early expressed zygotic microRNAs in *Drosophila* (BUSHATI *et al.* 2008) and other species such as zebrafish (GIRALDEZ *et al.* 2006). I predicted target sites for each maternal microRNAs in stable and unstable maternal transcripts during MZT (TADROS *et al.* 2007). Table 3 shows maternal microRNAs targeting more unstable maternal transcripts than expected by chance (FDR < 10%). Two of the microRNAs, mir-283 and mir-277, were detected at very low levels in unfertilized eggs (Table S1) and have a higher expression level later on during embryonic development (Figure 1). It is possible that these microRNAs contribute to the destabilization of maternal transcripts, but probably as zygotic microRNAs. A third set of microRNAs which may contribute to transcript clearance during MZT are the mir-310 and mir-92 families. They both share the same seed sequence (which defines the targeted transcripts). Last, the microRNA-9 family also target unstable maternal transcripts. These microRNAs are abundant in the *Drosophila* unfertilized egg, and may be contributing to the MZT. In summary, some maternally deposited microRNAs have a potential role in destabilizing maternal transcripts.

Maternal protein-coding transcripts avoid target sites for maternal microRNAs: If maternal microRNAs have a functional impact on maternal transcripts, these transcripts should have a different target repertoire compared to non-maternal (zygotic) transcripts. Thus, I estimated how many maternal and zygotic transcripts are targeted by maternal microRNAs. Overall, 73% of maternal transcripts and 63% of zygotic transcripts have canonical seed target sites for maternal microRNAs. However, for transcripts from genes with a recent evolutionary history, that is, that they originated in the *Drosophila melanogaster* lineage, maternal transcripts were less likely to be targeted by maternal microRNAs than zygotic transcripts: 50.7±0.5% of maternal transcripts have canonical target sites for maternal microRNAs, whilst this percentage is 52.6±0.4% for zygotic transcripts ($p \sim 0.004$; t-test). Although the difference is small, the observation that evolutionarily

young genes have a relatively lower proportion of targets for maternal microRNAs than the overall set of genes, suggest purifying selection against microRNA targets.

To test whether there is a selection against maternal microRNA target sites we should evaluate population data. To do so, I first constructed a model of microRNA target mutation as follows (see Figure 3A): 1) a target site is defined as any 6 nucleotide sequence (sixmer) in a 3'UTR complementary to the seed region (BARTEL 2009) of a microRNA; 2) any target site has 18 mutant neighbours, which are one nucleotide mutation apart from the canonical target, and are not themselves targets; 3) only polymorphic sites in which one of the alleles is a target site and the other a non-target are further considered in this analysis. Allele frequency is defined as the proportion of the target allele (*p* in Figure 3B). For instance, an allele frequency of 0.8 means that 80% of the sampled individuals have the target site at a given position and 20% have a non-target mutant neighbour. Conversely, an allele frequency of 0.3 will show that the non-target neighbour is more frequent (70%) than the target allele (30%). Population genetics theory (CROW and KIMURA 1970; NEI 1975) predicts that, in a finite population, two alleles neutral to each other will have a U-shaped distribution, that is, most individuals will be homozygous for one of the alleles. However, if there is a selection pressure to conserve a target site, the distribution will be shifted to the right. Conversely, if selection is against a target site allele, the distribution will be shifted to the left (see Figure 3C). A U-shape distribution is not expected if other evolutionary forces are in place (for instance, mutation bias, or background selection). Hence, in other to estimate the selective pressure for, or against, a microRNA target site in maternal transcripts we need an empirical expected distribution of allele frequencies. Therefore, I calculated the frequency of target/non-target pairs of alleles in zygotic transcripts, in which maternal microRNAs have no (or little) influence. By comparing the allele frequency distribution of target sites between maternal and zygotic transcripts we can estimate the relative selective pressure on microRNA target sites in maternal with respect to zygotic transcripts.

To do so, polymorphic sites for target/non-target seed sequences in *Drosophila* 3'UTRs are

identified and then the allele frequency is calculated. For each pair of alleles, both target and non-target sites were searched in the reference genome. This way, we also account for non-target alleles in the reference genome sequence that may be *bona fide* microRNA target sites. The distribution of allele frequencies between maternal and zygotic gene transcripts for each microRNA can then be compared. Figure 3D shows the case for *mir-995* microRNA products. One of them, mir-995-3p is abundant in unfertilized eggs whilst the alternate arm, mir-995-5p, is virtually absent in eggs. By comparing the allele frequency distributions for both arms I observe that the degree of non-target conservation in maternal transcripts with respect to zygotic transcripts is is higher in mir-995-3p than in mir-995-5p. That is, the dark grey bar is higher than the light grey bar for allele frequency less than 0.1 in mir-995-3p in Figure 3D. In other words, there is a preference for alleles that avoid being targeted by the maternal mir-995-3p, but no such a preference for non-maternal mir-995-5p. Both arms of mir-305 are present at high levels in unfertilized eggs. Figure 4A shows the allele frequency distribution for their targets, and both arms show evidence of target avoidance. As a counterexample, Figure 4B shows the allele frequency distribution of a microRNA for which none of the arms was detected in unfertilized eggs: mir-4986. Consistently, none of the microRNA products showed evidence of target avoidance by maternal transcripts.

To explore whether this pattern is a general feature of maternal microRNAs I defined a measure of target avoidance at the population level as the log2 ratio of the number of non-target sites (allele frequency <0.1) between maternal and zygotic transcripts. Positive values indicate that targets for a specific microRNA tend to be avoided by maternal transcripts. Figure 5A is a bar plot of target avoidance log2 ratios for different levels of microRNA abundance in the egg. Maternally deposited coding transcripts tend to avoid some target sites for highly abundant maternal microRNAs (with respect to zygotic transcripts). Differences were statistically significant (Figure 5A). In a similar manner I defined a target conservation measure as the log2 ratio of the number of target sites (allele frequency >0.9) between maternal and zygotic transcripts. A positive value

indicates that target-sites are preferentially conserved in maternal transcripts. Figure 5B shows these values for different microRNA abundances. Overall, maternal transcripts conserve some target sites, but there is no distinctive pattern between maternal and non-maternal microRNAs (Figure 5B). To further validate the observed target avoidance, we can compare the whole allele frequency distribution and evaluate whether the distribution for maternal transcripts is shifted to the left with respect to that of zygotic transcripts (as suggested in Figure 3C, right panel). Figure 5C plots the p-value for a one-tail Kolmogorov-Smirnov test, the lower the p-value, the more shifted to the left, that is, the more evidence for target avoidance. For highly abundant maternal microRNAs, the allele frequency distribution of target/non-target sites is shifted to the left, that is, there is a preference for the non-target allele.

The population genetic and comparative genomics analyses suggest that whenever there is selection against a target site, the non-target allele is eventually fixed in the population. To further explore selection on conserved sites I first catalogued, among all target/non-target pair of alleles analysed in this study, those sites that were conserved in *Drosophila sechellia*, which diverged from the *D. melanogaster* lineage about 2 million years ago. Figure 6A plots, for sites that are conserved as target sites in *D. sechellia*, the allele frequency distribution of target sites (as described for Figures 3 and 4) of targets in maternal transcripts for maternal microRNAs (dark grey bars) and zygotic microRNAs (light grey bars). Both distributions are heavily skewed to the right, meaning that conserved (target) sites in *D. sechellia* most often have the conserved allele in *D. melanogaster* populations. Additionally, the proportion of the target allele in populations is higher for maternal microRNAs than for zygotic microRNAs. That indicates that there is ongoing selection to preserve some conserved maternal microRNA target sites. On the other hand, Figure 6B shows the allele frequency distributions in sites where the non-target allele is conserved in *D. sechellia*. Like above, the conserved allele is more frequent than the non-conserved allele, but in this case the conserved allele is the non-target. Likewise, the proportion of non-target alleles is higher for maternal than for

zygotic microRNA target/non-target sites. This indicates ongoing selection in conserved non-target sites against target sites for maternal microRNAs. From the analyses here described I conclude that maternal transcripts, compared to non-maternal zygotic transcripts, have a mild yet significant tendency to avoid maternal microRNA target sites.

DISCUSSION

This study characterizes microRNA products from *Drosophila* unfertilized eggs. I validated seven of these microRNAs by qPCR. The presence of microRNAs in unfertilized oocytes have been described in mice (TANG *et al.* 2007). However, it has been shown that microRNA activity is suppressed in mice oocytes, indicating that maternally deposited microRNAs may not have a defined function in this species (MA *et al.* 2010; SUH *et al.* 2010). Here I show evidence for *Drosophila* maternal microRNA activity as they have an impact in the evolution of potential target sites in maternal microRNAs (Figures 3, 4, 5 and 6).

One of the most abundant maternal microRNAs, mir-184, has been already described in freshly laid *Drosophila* eggs (IOVINO *et al.* 2009). Also, the *mir-184* gene has an important role during oocyte development as well as in early development (IOVINO *et al.* 2009). Another maternal microRNA gene, *mir-14*, seems to be involved in transcriptional silencing of transposable elements in the germline (MUGAT *et al.* 2015). On the other hand, the conserved microRNA mir-34 has been also described as a maternal microRNA (SONI *et al.* 2013) but it has only one read copy in our dataset, and it has not been detected in two other independent high-throughput screens (LEE *et al.* 2014; NINOVA *et al.* 2015). The level of mir-34-5p was also very low in specific qPCR assays (Figure 2). All these suggest that either mir-34 is a very low copy maternal microRNA, or that it is rapidly degraded after egg deposition/activation.

Another maternal microRNA gene, *mir-9c*, has been shown to be crucial to regulate the

number of germ cells (KUGLER *et al.* 2013). Indeed, is the maternal loss of *mir-9c* what produces this phenotype (KUGLER *et al.* 2013). This microRNA is hosted within a maternally deposited gene, *grapes* (Table 2). Here I show that mir-9c-5p targets more unstable transcript during the MZT than expected by chance (Table 3), which indicates that mir-9c-5p may have a role during maternal transcript clearance during the initial steps of development. A similar role has been described for zygotically transcribed microRNAs (BUSHATI *et al.* 2008).

Other maternally deposited microRNAs derive from the *mir-310/mir-313* cluster. This cluster is highly conserved in the *Drosophila* lineage (MARCO *et al.* 2013a), although it may have originated in insects (NINOVA *et al.* 2014), and is evolutionarily related with the (also maternal) *mir-92a/mir-92b* cluster (LU *et al.* 2008; NINOVA *et al.* 2014). Mature products from the orthologous *mir-310/311/312/313* and *mir-92a/92b* clusters in *Drosophila virilis* has been detected at high levels during the first two hours of development, suggesting that this cluster is also maternally deposited in this species (Supplementary Table 2 in (NINOVA *et al.* 2014)). Interestingly, some maternal microRNAs have other functions later on during development. MicroRNAs from the *mir-310/311/312/313* cluster are known to be involved in male gonads development (PANCRATOV *et al.* 2013). Recently, Ranz and collaborators found that mir-310/mir-313 microRNAs show male biased expression pattern at the onset of metamorphosis (YEH *et al.* 2014). On the other hand, *mir-92a* is expressed in the adult, and it is involved in leg morphology (ARIF *et al.* 2013). Some other maternal microRNAs have roles unrelated with embryonic development such as *mir-14*, which regulates insulin production (XU *et al.* 2003); *mir-279*, involved in the circadian clock (LUO and SEHGAL 2012 p. 279); or *mir-8*, associated to abdominal pigmentation (KENNELL *et al.* 2012), to name but a few cases. Altogether, these examples show that maternal microRNAs frequently have other functions in different developmental stages and/or tissues.

MicroRNA target avoidance has been observed in *Drosophila* (STARK *et al.* 2005), as well as in mice (FARH *et al.* 2005) and humans (SOOD *et al.* 2006; CHEN and RAJEWSKY 2006). Here I

observe a similar pattern in *Drosophila* eggs, in which maternal transcripts tend to avoid target sites for maternal microRNAs. Comparative genomics and population genetic data independently suggest that maternal transcripts tend to avoid being targeted by maternal microRNAs. The main advantage of working with microRNAs to study evolution at the population level is that we can predict the impact of single point mutations in both microRNAs and their targets, something that is not yet possible with other regulators (such as transcription factors). Further analyses, including the theoretical expectations of the proposed target-site-mutation model, will shed light on how microRNA function diversify and, more generally, how gene regulation evolves.

Alternatively, a lower number of target sites in maternal transcripts could be explained as an early degradation of transcripts with conserved target sites, and therefore not detected in early embryos. However, in *Drosophila*, microRNA-mediated transcript degradation happens a few hours after microRNA-mediated repression (DJURANOVIC *et al.* 2012). Maternal transcripts are detected from 0-2 hour old embryos, and they are unlikely to have had microRNA-mediated transcript degradation. The microRNA genes studied in that paper were: *mir-9b*, *mir-279* and *bantam*, all of which were detected in this study as maternal.

If microRNAs are likely to have a function in maternal transcripts, why we observe selection against target sites? I suggest the following explanation. A microRNA that is maternally deposited and targets several maternal microRNAs may have a function, for instance, induce their programmed degradation during MZT. However, there are hundreds of other maternal transcripts that should not be targeted. This situation creates a conflict in which functional interactions must be conserved, but new interactions that potentially impair existing regulatory networks should be avoided. In this context, most maternal transcripts will be selected against target sites for maternal microRNAs. It is likely that this conflict also happens in other tissues and species, and probably will also affect transcription factor mediated regulation. The extend to which selective avoidance of targets for gene regulators affects genome evolution is not yet known, and more studies need to be

done.

Overall, this paper describes three features of maternally transmitted microRNAs: 1) they are often produced from introns of maternally deposited transcripts; 2) they can be zygotically transcribed and have other functions during development; 3) maternal transcripts tend to avoid target sites for maternal microRNAs. Additionally, I suggest that mir-9c may be involved in maternal transcript clearance during MZT. These observations indicate that some maternal microRNAs may have a function during development but are potentially damaging to the normal function of other maternal genes, and therefore selective pressures favour maternal transcripts to avoid being targeted by maternal microRNAs.

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Table 1. Maternal microRNAs in *Drosophila melanogaster*.

MicroRNA transcript*	**Reads per miRNA**	**Total reads (%)****
mir-310/311/312/313	356/2012/1661/82	4111 (31.3)
mir-92a/92b	172/2109	2281 (17.4)
mir-184	1377	1377 (10.5)
mir-9c/306/79/9b	1064/154/4/132	1354 (10.3)
bantam	1204	1204 (9.2)
mir-995	624	624 (4.8)
mir-14	411	411 (3.1)
mir-275/305	90/269	359 (2.7)
mir-998/11	5/205	210 (1.6)
mir-8	209	209 (1.6)
mir-2b-2/2a-1/2a-2	85/25/19	129 (1.0)
mir-279/996	61/56	117 (0.9)
mir-2b-1	92	92 (0.7)
mir-281-2/281-1	66/13	79 (0.6)
mir-4969/999	0/69	69 (0.5)
mir-33	62	62 (0.5)
mir-263a	59	59 (0.4)
mir-10	28	28 (0.2)
mir-2c/13a/13b-1	0/0/27	27 (0.2)
mir-13b-2	26	26 (0.2)
mir-970	26	26 (0.2)
mir-1012	25	25 (0.2)
mir-31a	23	23 (0.2)
mir-9a	21	21 (0.2)
mir-309/3/286/4/5/6-1/6-2/6-3	0/1/15/1/1/1/1/1	21 (0.1)
mir-956	20	20 (0.2)
mir-276a	18	18 (0.1)
mir-994/318	2/14	16 (0.1)
mir-1010	14	14 (0.1)

* MicroRNAs clustered in the genome (<10kb).
** Percentage over total number of reads mapping to microRNAs.

Table 2. Maternal microRNA loci within protein-coding genes.

MicroRNA cluster	Host gene	Protein-coding gene maternal?
mir-995	cdc2c	yes
mir-11/998	E2f	yes
mir-92a	jigr1	yes
mir-999	CASK	yes
mir-281-1/281-2	Oda	yes
mir-970	Tomosyn	yes
mir-2b-2/2a-1/2a-2	spi	yes
mir-13b-2	CG7033	yes
mir-9c/306/79/9b	grp	yes*
mir-33	HLH106	no expression information
mir-1012	Lerp	no expression information
mir-1010	SKIP	no

* Detected in the oocyte.

Table 3. Maternal microRNAs targeting unstable transcripts during maternal-to-zygotic transition

MicroRNA	Unstable targets	Stable targets	Proportion unstable transcripts*	q-value**
mir-283-5p	179	805	0.182	0.018
mir-277-3p	116	497	0.189	0.026
mir-9a/b/c-5p	50	166	0.232	0.036
mir-92a/b-3p; mir-310/311/312/313-3p	74	313	0.191	0.096

* Expected proportion is 0.146 **Binomial test

Figure legends

Figure 1. Expression profile of maternal microRNAs in *Drosophila melanogaster*. The hierarchical tree is split into three categories of microRNAs: high abundance in ovaries compared to the other stages; those which are mainly present in the unfertilized eggs and those which have a higher expression level later during development.

Figure 2. Expression of maternal microRNAs. Levels of microRNA mature products in unfertilized eggs with respect with average bantam-3p levels from qPCR assays (grey boxes; see Materials and Methods), and levels of microRNA mature products detected in 2-6 hours embryos with respect to levels in unfertilized eggs. Error bars are for three biological replicates. Dashed line indicates the levels of bantam-3p as a reference.

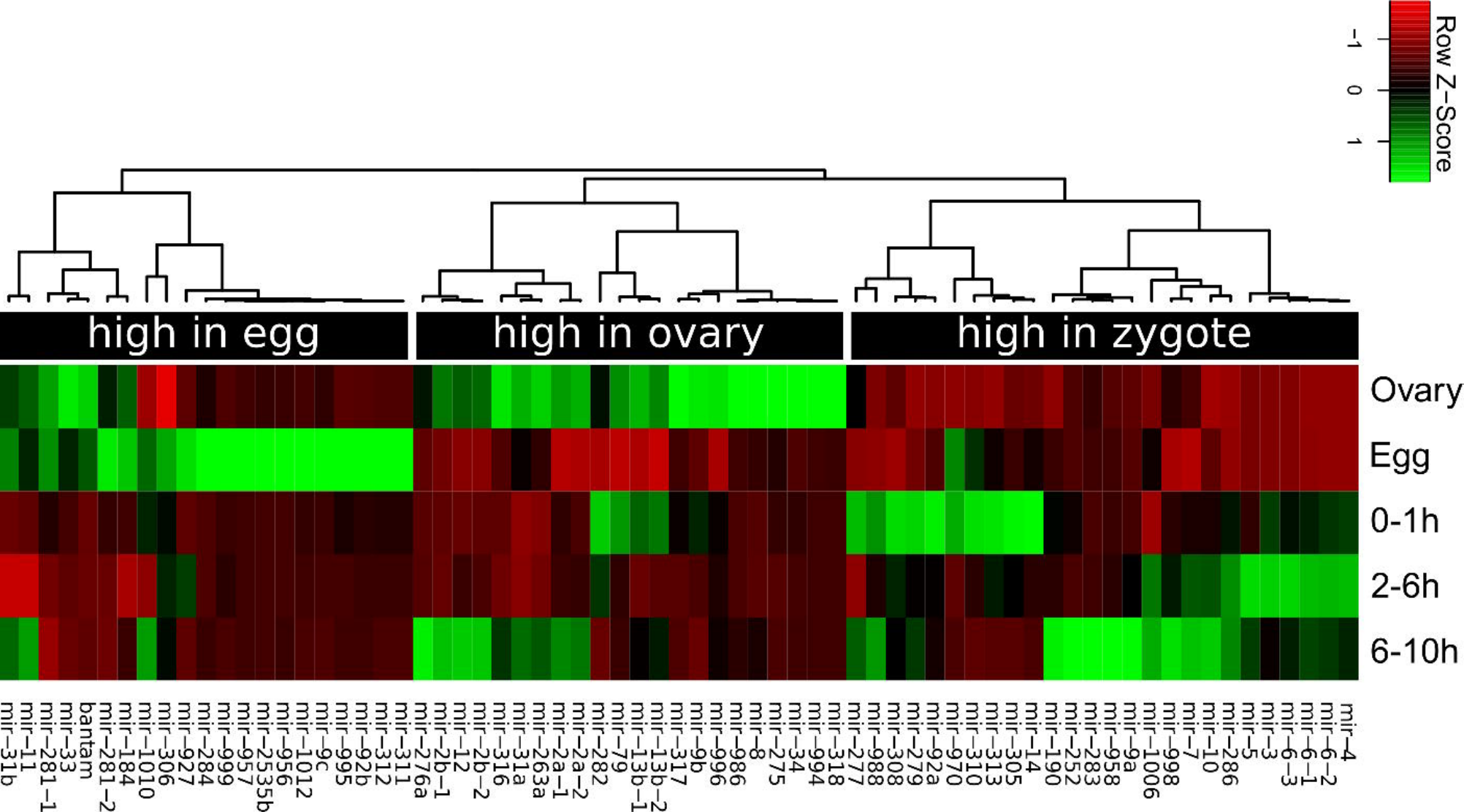
Figure 3. Polymorphic target sites in *Drosophila* populations. (A) Each microRNA sixmer target site has 18 one-nucleotide mutant neighbours which are themselves not target sites. (B) The allele frequency for each pair of target/non-target site is calculated as the proportion of target site alleles with respect to the total number of alleles in the pair. (C) Allele frequency distribution in a finite population is U-shaped for pairs of alleles neutral to each other. If there is selection favouring target sites, distributions are expected to be shifted to the right. Conversely, if there is selection against target sites, distributions will be shifted to the left. (D) Allele frequency distribution for target sites for maternal microRNAs in maternal (dark grey) and non-maternal zygotic (light grey) transcripts. Left and right panel shows the distributions for 5' and 3' arms of mir-995 respectively. Grey box in left panel indicates that mir-995-5p is virtually absent from the unfertilized egg.

Figure 4. Allele frequency distribution for target sites of maternal microRNAs. Distribution of

targets for maternal and zygotic transcripts for mir-305 (both arms are highly present in the egg) and mir- 4986 (grey box, neither of the arms was detected in the egg).

Figure 5. Maternal microRNA target avoidance. (A) Target avoidance ratio (see main text for details) for microRNAs with differences abundances in the unfertilized egg (NULL – not detected, LOW – less than 0.1% of the set, MID – between 0.1% and 1%, HIGH – more than 1%). Error bars represent the Standard Error of the Mean. Asterisks show statistically significant differences ($p<0.01$) for t-test with unequal variances. (B) Target conservation ratio. (C) Distribution shifting to the left (see main text for details) of allele frequency distribution in maternal transcripts with respect to non-maternal zygotic transcripts.

Figure 6. Conserved target/non-target sited in *Drosophila sechellia*. Allele frequency distribution of maternal microRNAs target/non-target sites (dark grey bars) and for zygotic microRNAs (light grey bars), and p-values for Kolmogorov-Smirnov test of differences between distributions. (A) Sites conserved in *D. sechellia* as a target site. (B) Sites conserved in *D. sechellia* as a non-target site.


Row Z-Score
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0
1
high in egg
high in ovary
high in zygote
Ovary
Egg
0-1h
2-6h
6-10h
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mir-6-2
mir-6-1
mir-6-3
mir-3
mir-5
mir-286
mir-10
mir-7
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mir-1006
mir-9a
mir-958
mir-283
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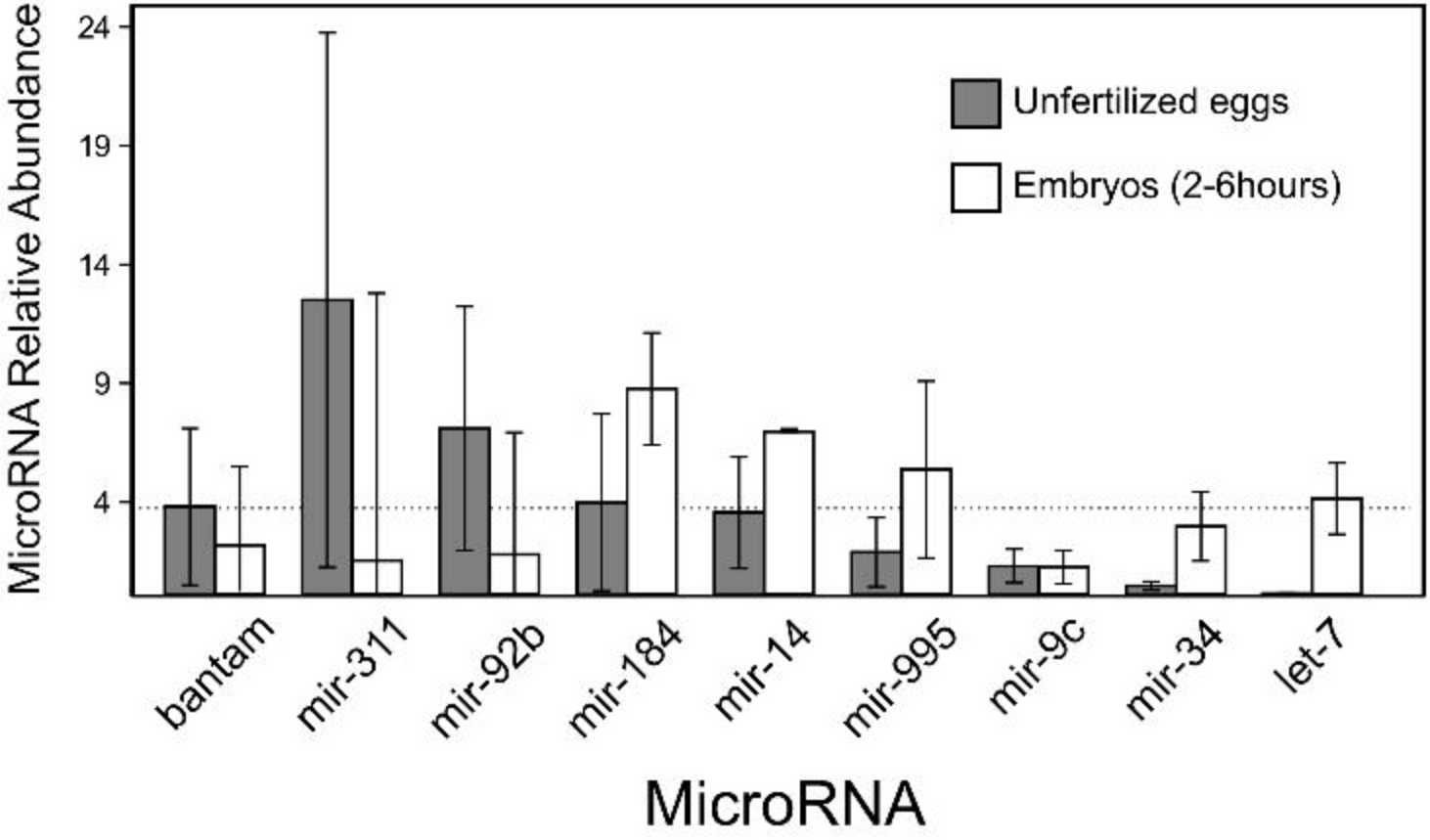
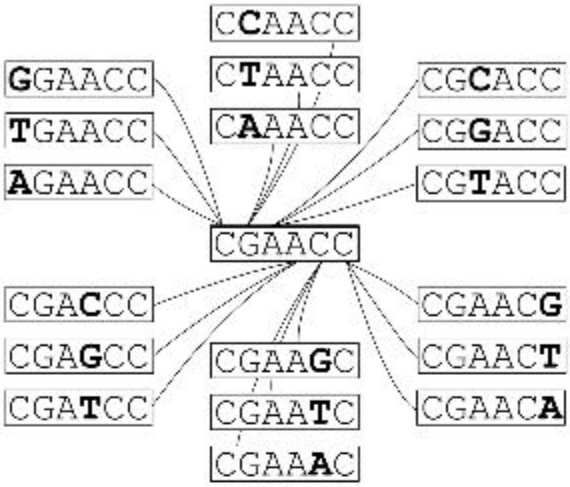

MicroRNA Relative Abundance
24
19
14
9
4
Unfertilized eggs
Embryos (2-6hours)
bantam
mir-311
mir-92b
mir-184
mir-14
mir-995
mir-9c
mir-34
let-7
MicroRNA

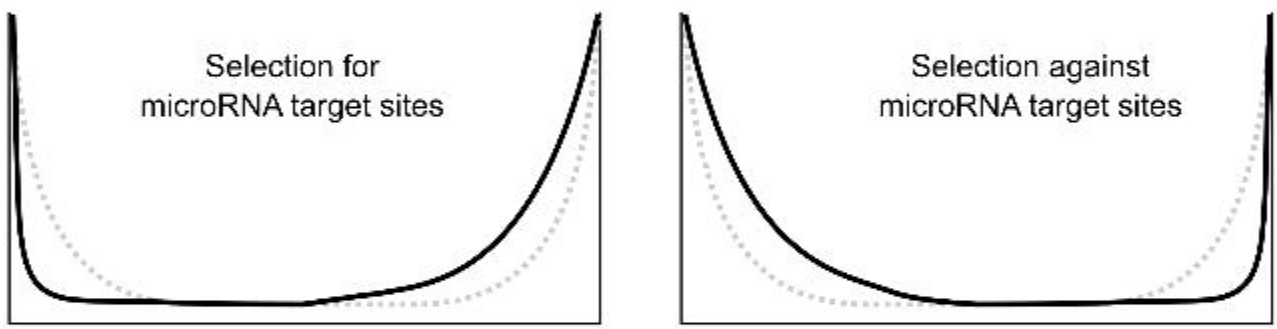
(A)



(B)

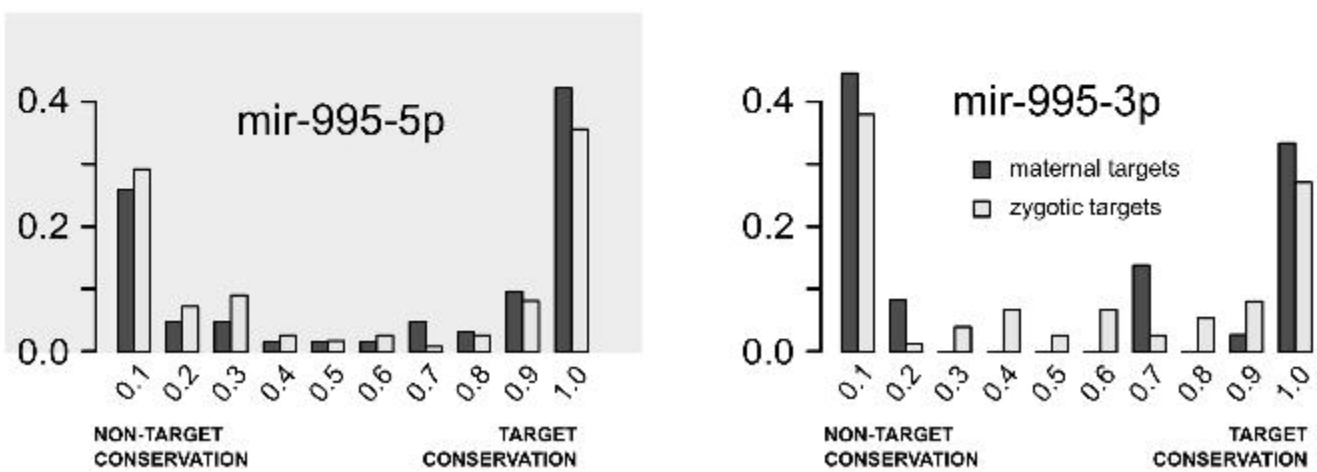
$$p = \frac{\#\ \text{CGAAC}\mathbf{C}}{\#\ \text{CGAAC}\mathbf{C}\ +\ \#\ \text{GGAAC}\mathbf{G}}$$

(C)

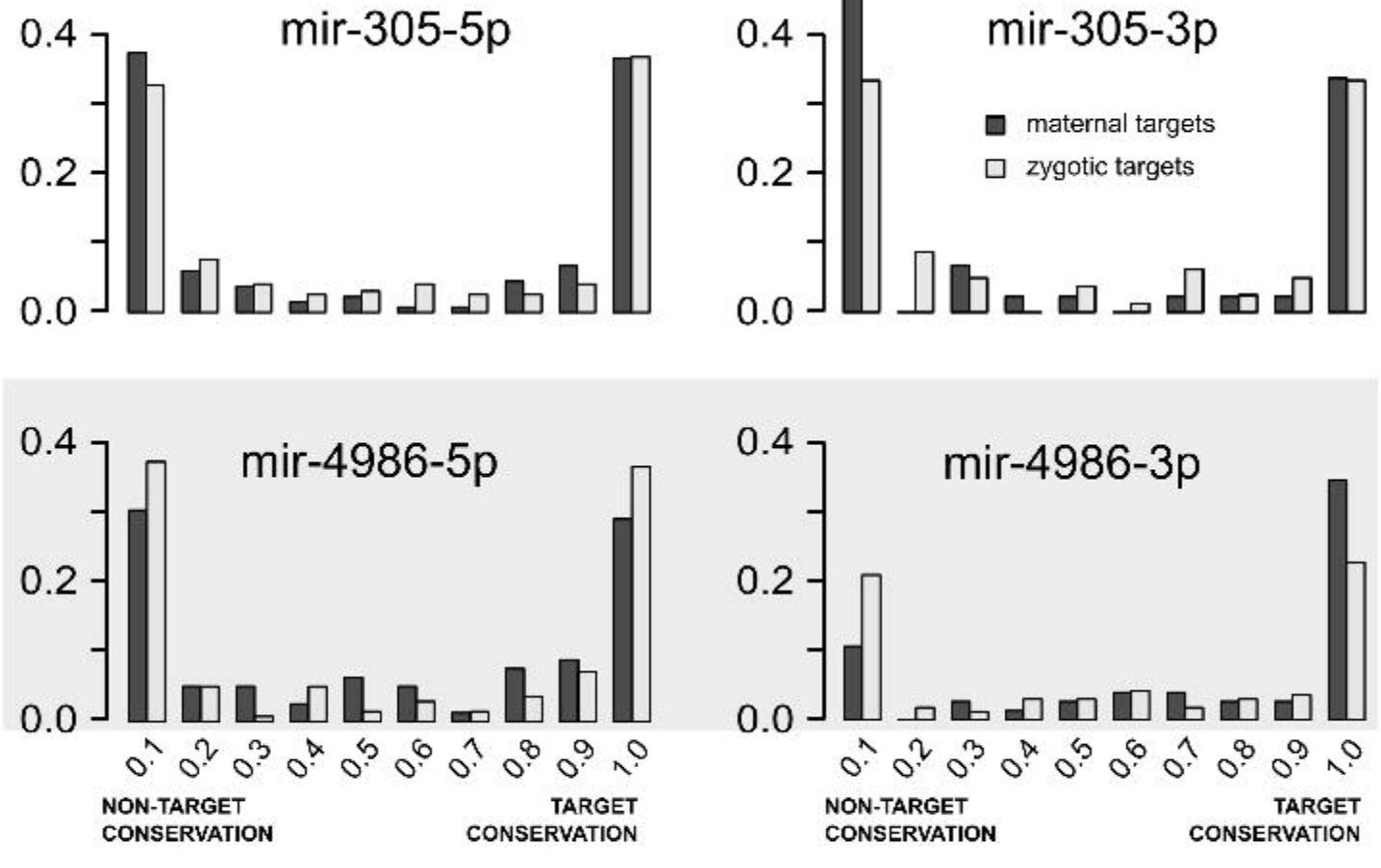


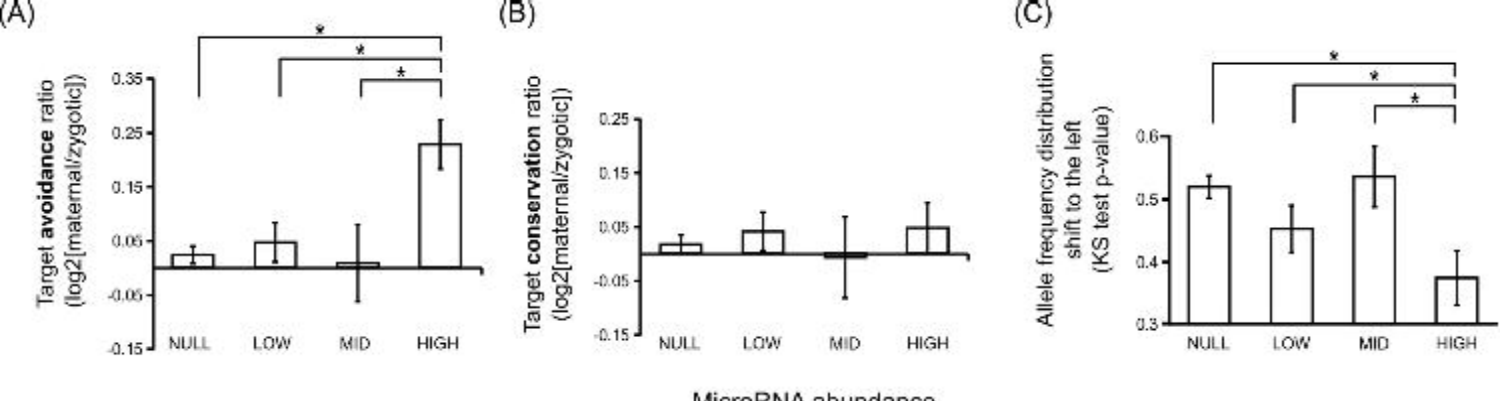
Allele frequency (p)

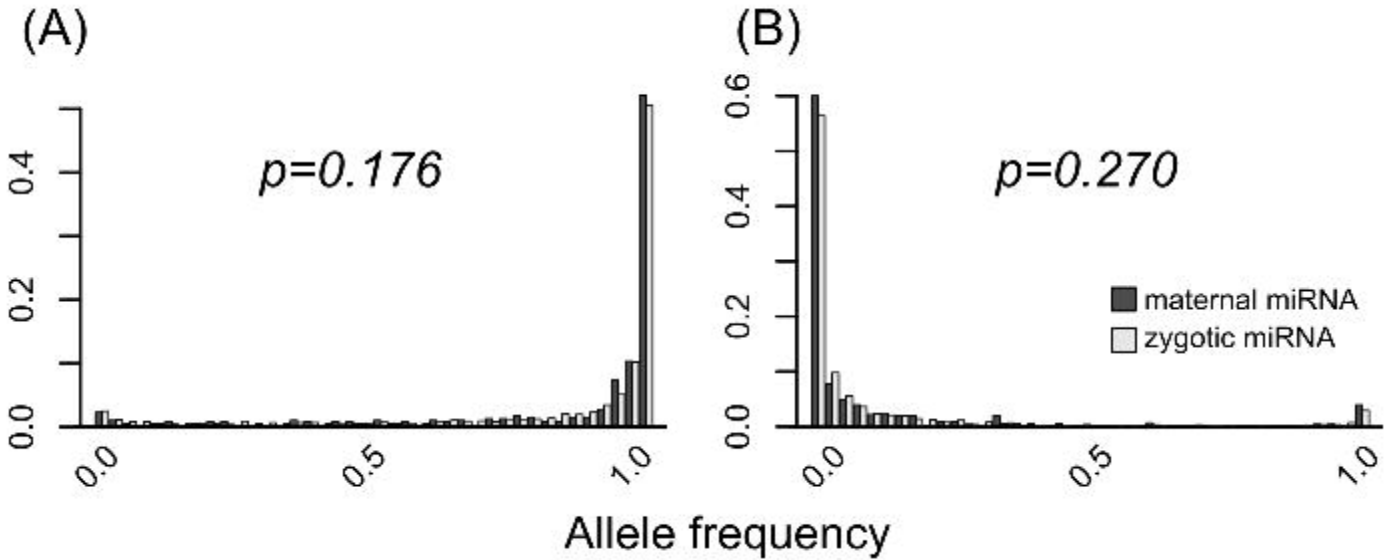
(D)



Allele frequency (p)


mir-305-5p
mir-305-3p
mir-4986-5p
mir-4986-3p
maternal targets
zygotic targets
0.4
0.2
0.0
0.1
0.2
0.3
0.4
0.5
0.6
0.7
0.8
0.9
1.0
NON-TARGET CONSERVATION
TARGET CONSERVATION
Allele frequency


(A)
(B)
(C)
Target avoidance ratio (log2[maternal/zygotic])
Target conservation ratio (log2[maternal/zygotic])
Allele frequency distribution shift to the left (KS test p-value)
0.35
0.25
0.15
0.05
-0.05
-0.15
0.6
0.5
0.4
0.3
NULL
LOW
MID
HIGH
*
MicroRNA abundance


(A)
p=0.176
0.0
0.2
0.4
0.0
0.5
1.0
(B)
p=0.270
0.0
0.2
0.4
0.6
maternal miRNA
zygotic miRNA
0.0
0.5
1.0
Allele frequency