

Physical activity and risks of breast and colorectal cancer: A Mendelian randomization analysis

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Abstract

Physical activity has been associated with lower risks of breast and colorectal cancer in epidemiological studies; however, it is unknown if these associations are causal or confounded. In two-sample Mendelian randomization analyses, using summary genetic data from the UK Biobank and GWA consortia, we found that a one standard deviation increment in average acceleration was associated with lower risks of breast cancer (odds ratio [OR]: 0.59, 95% confidence interval [CI]: 0.42 to 0.84, P-value=0.003) and colorectal cancer (OR: 0.66, 95% CI: 0.53 to 0.82, P-value= 2×10^{-4}). We found similar magnitude inverse associations by breast cancer subtype and by colorectal cancer anatomical site. Our results support a potentially causal relationship between higher physical activity levels and lower risks of breast cancer and colorectal cancer. Based on these data, the promotion of physical activity is probably an effective strategy in the primary prevention of these commonly diagnosed cancers.

Introduction

Breast and colorectal cancer are two of the most common cancers globally with a combined estimated number of 4 million new cases and 1.5 million deaths in 2018 ¹. Physical activity is widely promoted along with good nutrition, maintaining a healthy weight, and refraining from smoking, as key components of a healthy lifestyle that contribute to lower risks of several non-communicable diseases such as cardiovascular disease, diabetes, and cancer ².

Epidemiological studies have consistently observed inverse relationships between physical activity and risks of breast and colorectal cancer ²⁻⁵, but have generally relied on self-report measures of physical activity, which are prone to recall and response biases, and may attenuate ‘true’ associations with disease risk ⁶. More objective methods to measure physical activity, such as accelerometry, have seldom been used in large-scale epidemiological studies, with the UK Biobank being a recent exception, in which ~100,000 participants wore a wrist accelerometer for 7-days to measure total activity levels ⁷. Epidemiological analyses of these data will provide important new evidence on the link between physical activity and cancer, but these analyses remain vulnerable to other biases of observational epidemiology, such as residual confounding (e.g. low physical activity levels may be correlated with other unfavourable health behaviours) and reverse causality (e.g. preclinical cancer symptoms may have resulted in low physical activity levels).

Mendelian randomization (MR) is an increasingly used tool that uses germline genetic variants as proxies (or instrumental variables) for exposures of interest to enable causal inferences to be made between a potentially modifiable exposure and an outcome ⁸. Unlike traditional observational epidemiology, MR analyses, should be largely free of conventional confounding owing to the random independent assignment of alleles during meiosis ⁹. In

addition, there should be no reverse causation, as germline genetic variants are fixed at conception and are consequently unaffected by the disease process ⁹.

We used a two-sample MR framework to examine potential causal associations between objective accelerometer-measured physical activity and risks of breast and colorectal cancer using genetic variants associated with accelerometer-measured physical activity identified from a recent genome-wide association study (GWAS) ¹⁰. We examined the associations of these genetic variants with risks of breast cancer ¹¹ and colorectal cancer ¹².

Results

Mendelian randomization estimates for breast and colorectal cancer

We estimated that a 1 standard deviation (SD) (8.14 milli-gravities) increment in the genetically predicted levels of accelerometer-measured physical activity was associated with a 41% (Odds ratio [OR]: 0.59, 95% confidence interval [CI]: 0.42 to 0.84, P-value=0.003) lower risk of overall breast cancer (Table 2). Similar magnitude inverse associations were found for estrogen receptor positive (ER⁺) (OR: 0.53, 95% CI: 0.35 to 0.82, P-value=0.004) and estrogen receptor negative (ER⁻) (OR: 0.78, 95% CI: 0.51 to 1.22, P-value=0.27) breast cancer ($I^2=35\%$; P-heterogeneity by subtype=0.21). There was some evidence of heterogeneity based on Cochran's Q (P-value<0.05) for the breast cancer analyses; consequently, for these models random effects MR estimates were used (Table 2). MR estimates for each individual single nucleotide polymorphism (SNP) associated with accelerometer-measured physical activity in relation to breast cancer risk are presented in Figure 1.

For colorectal cancer, a 1 SD increment in accelerometer-measured physical activity level was associated with a 34% (OR: 0.66, 95% CI: 0.53 to 0.82, P-value= 1.9×10^{-4}) lower risk. The estimated effect size was stronger for women (OR: 0.54, 95% CI: 0.40 to 0.74, P-

value= 1.2×10^{-4}) than men (OR: 0.82, 95% CI: 0.61 to 1.11, P-value=0.21), although this heterogeneity did not meet the threshold of significance ($I^2=72\%$; P-heterogeneity by sex=0.06). For colorectal subsite analyses, accelerometer-measured physical activity levels were inversely associated with risks of colon cancer (OR per 1 SD increment OR: 0.61, 95% CI: 0.47 to 0.79, P-value= 2×10^{-4}) and rectal cancer (OR: 0.76, 95% CI: 0.55 to 1.07, P-value=0.12). MR estimates for each individual SNP associated with accelerometer-measured physical activity in relation to breast cancer risk are presented in Figure 2 and Supplementary fig 1.

Similar results were generally observed for all breast cancer and colorectal cancer endpoints when MR analyses were conducted with the two genome-wide significant accelerometer-measured physical activity SNPs only (Table 2).

Evaluation of assumptions and sensitivity analyses

The strength of the genetic instruments denoted by the F-statistic was ≥ 10 for all the accelerometer-measured physical activity variants and ranged between 29 and 56 (Table 1). The intercept test from the MR-Egger regression was statistically significant in the analysis of colorectal cancer in women denoting potential pleiotropy; however, the corrected estimate from MR-Egger replicated the initial finding (Table 2). The estimates from the weighted median approach were consistent with those of inverse variance weighted (IVW) models. The MR pleiotropy residual sum and outlier test (MR-PRESSO) method identified the SNPs rs11012732 and rs55657917 as pleiotropic for breast cancer, but similar magnitude inverse relationships were observed when these variants were excluded from the analyses (Supplementary Table 6).

After examining Phenoscanner and GWAS catalog, we found that several of the accelerometer-measured physical activity genetic variants were also associated with adiposity

related phenotypes (Supplementary Table 7). However, the results from the leave-one-SNP out analysis did not reveal any influential SNPs driving the associations (Supplementary Tables 8 – 10). Additionally, similar results were found when the five adiposity-related SNPs were excluded from the genetic instrument (Supplementary Table 11). Further, the results from the multivariable MR analyses adjusting for BMI were largely unchanged from the main IVW results (Supplementary Tables 12, 13).

The association of genetically predicted physical activity and colorectal cancer was similar (OR: 0.60, 95% CI: 0.47 to 0.76, P-value= 2.5×10^{-5}) after excluding UK Biobank participants from the GWAS for colorectal cancer.

Discussion

In this MR analysis, higher levels of genetically-predicted accelerometer-measured physical activity were associated with lower risks of breast cancer and colorectal cancer, with similar magnitude inverse associations found for breast cancer subtypes and by colorectal anatomical subsite. These findings indicate that population-level increases in physical activity may lower the incidence of these two commonly diagnosed cancers, and support the promotion of physical activity for cancer prevention.

A large body of observational studies has investigated how physical activity relates to risk of breast and colorectal cancer¹³. In a participant-level pooled analysis of 12 prospective studies, when the 90th and 10th percentile of leisure-time physical activity were compared, lower risks of breast cancer (Hazard ratio [HR]: 0.90, 95% CI: 0.87 to 0.93), colon cancer (HR: 0.84, 95% CI: 0.77 to 0.91), and rectal cancer (HR: 0.87, 95% CI: 0.80 to 0.95) were found³. These observational studies relied on self-report physical activity assessment methods that are prone to measurement error, which may attenuate associations towards the null. In

addition, causality cannot be ascertained from such observational analyses as they are vulnerable to residual confounding and reverse causality. Further, logistical and financial challenges prohibit randomized controlled trials of physical activity and cancer development. For example, it has been estimated that in order to detect a 20% breast cancer risk reduction, between 26,000 to 36,000 healthy middle-aged women would need to be randomized to a 5 year exercise intervention¹⁴. Several trials on cancer survivors are registered and underway, and these may provide evidence of potential causal associations between physical activity and disease free survival and cancer recurrence¹⁵; however, these interventions will not inform causal inference of the relationship between physical activity and cancer development. We conducted MR analyses to allow causal inference between accelerometer-measured physical activity and risks of developing breast and colorectal cancer. The inverse associations we found were consistent for breast cancer subtypes and across colorectal cancer subsites, and are strongly concordant with prior observational epidemiological evidence.

Being physically active is associated with less weight gain and body fatness, and lower adiposity is associated with lower risks of breast and colorectal cancer^{16,17}. Since body size/adiposity is likely on the causal pathway linking physical activity and breast and colorectal cancer, it is challenging to disentangle independent effects of physical activity on cancer development. This overlap between adiposity and physical activity is evident from 5 of the 10 SNPs in the genetic instrument for accelerometer-measured physical activity previously being associated with adiposity/body size traits. However, it is noteworthy that our results were unchanged when we excluded adiposity-related SNPs from the genetic instrument, and when we conducted multivariable MR analyses adjusting for body mass index (BMI). These results would therefore suggest that physical activity is also associated with breast and colorectal cancer independently of adiposity.

Multiple biological mechanisms are hypothesized to mediate the potential beneficial role of physical activity on cancer development. Greater physical activity has been associated with lower circulating levels of insulin and insulin-like growth factors, which promote cellular proliferation in breast and colorectal tissue and have also been linked to development of cancers at these sites¹⁸⁻²³. Higher levels of physical activity have also been associated with lower circulating levels of estradiol, estrone, and higher levels of sex hormone binding globulin²⁴⁻²⁶ which are strong risk factors for breast cancer development^{27,28}. Physical activity has also been associated with improvements in immune response, with increased surveillance and elimination of cancerous cells^{29,30}. Higher levels of physical activity may also reduce systemic inflammation by lowering the levels of pro-inflammatory factors, such as C-reactive protein (CRP), interleukin-6 (IL-6) and tumour necrosis factor-alpha (TNF- α)^{29,31,32}. Finally, emerging evidence suggests that the gut microbiome may play an important role in the physical activity and cancer relationship. Dysbiosis of the gut microbiome has been associated with increased risks of several malignancies, including breast and colorectal cancer³³. Changes in gut microbiome composition and derived metabolic products have been found following endurance exercise training, with short-chain fatty acid concentrations increased in lean, but not obese subjects^{34,35}.

A fundamental assumption of MR is that the genetic variants do not influence the outcome via a different biological pathway from the exposure of interest (horizontal pleiotropy). We conducted multiple sensitivity analyses to test for the influence of pleiotropy on our causal estimates, and our results were robust according to these various tests. A potential limitation of our analysis is that the genetic variants explained a small fraction of the variability of accelerometer-measured physical activity, which may have resulted in some of the breast cancer subtype and colorectal subsite analyses being underpowered. In addition, our use of summary-level data precluded subgroup analyses by other cancer risk factors (e.g.

BMI, exogenous hormone use). We were also unable to stratify breast cancer analyses by menopausal status; however, the majority of women in the source GWAS had postmenopausal breast cancer¹¹. Finally, 7-day accelerometer-measured physical activity levels of UK Biobank participants may not have been representative of usual behavioral patterns.

In conclusion, we found that genetically elevated levels of accelerometer-measured physical activity were associated with lower risks of breast and colorectal cancer. These findings strongly support the promotion of physical activity as an effective strategy in the primary prevention of these commonly diagnosed cancers.

Methods

Data on physical activity

Summary-level data were obtained from a recently published GWAS on accelerometer-measured physical activity conducted within UK Biobank¹⁰. In this GWAS, the regression models were adjusted for age, sex, the first ten genomic principal components, center, season (month), and genotyping chip. This GWAS identified 2 genome-wide-significant polymorphisms ($P\text{-value} < 5 \times 10^{-8}$) associated with accelerometer-measured physical activity. The estimated SNP-based heritability was 14% suggesting that additional SNPs contributed to its variation. Consequently, for our primary analyses, we used a larger number of 10 independent (linkage disequilibrium [LD] $r^2 \leq 0.001$) genetic variants by relaxing the significance threshold to $P\text{-value} < 1 \times 10^{-7}$. The expanded number of genetic variants in the accelerometer-measured physical activity instrument also allowed sensitivity analyses to be conducted to check for the influence of horizontal pleiotropy on the results. Data for the associations between the 8 additional SNPs and physical activity were obtained from a recent

MR study on physical activity and depression that used the data from the same UK Biobank GWAS³⁶. In secondary analyses, we used the two genome-wide significant SNPs only. Detailed information on the selected genetic variants is provided in Table 1.

Data on breast cancer and colorectal cancer

Summary data for the associations of the 10 accelerometer-measured genetic variants with breast cancer (overall and by estrogen receptor status: ER positive and ER negative) were obtained from a GWAS of 228,951 women (122,977 breast cancer [69,501 ER positive, 21,468 ER negative] cases and 105,974 controls) of European ancestry from the Breast Cancer Association Consortium (BCAC)¹¹. Genotypes were imputed using the 1000 Genomes Project reference panel and the regression models adjusted for the first ten principal components and country or study (Supplementary Table 1). For colorectal cancer, summary data from 125,915 participants (58,221 colorectal cancer cases and 67,694 controls) were drawn from a meta-analysis within the ColoRectal Transdisciplinary Study (CORECT), the Colon Cancer Family Registry (CCFR), and the Genetics and Epidemiology of Colorectal Cancer (GECCO) consortia¹². Imputation was performed using the Haplotype Reference Consortium (HRC) r1.0 reference panel and the regression models were further adjusted for age, sex, genotyping platform (whenever appropriate), and genomic principal components (from 3 to 13, whenever appropriate) (Supplementary Tables 2, 3).

Statistical power

The a priori statistical power was calculated using an online tool at <http://cnsgenomics.com/shiny/mRnd/>³⁷. The 10 accelerometer-measured physical activity SNPs collectively explained 0.4% of phenotypic variability. Given a type 1 error of 5%, we had sufficient power (>80%) when the expected OR per 1 SD was ≤ 0.83 and ≤ 0.77 for overall

breast cancer (122,977 cases and 105,974 controls) and colorectal cancer (58,221 colorectal cases and 67,694 controls), respectively. The power estimates for subtypes of breast cancer and by subsites of colorectal cancer are presented in Supplementary Table 4.

Statistical analysis

A two-sample MR approach using summary data and the fixed-effect IVW method was implemented. All accelerometer-measured physical activity and cancer results correspond to an OR per 1 SD increment (8.14 milli-gravities) in the genetically predicted overall average acceleration. The heterogeneity of causal effects by cancer subtype and sex was investigated by estimating the I^2 statistic assuming a fixed-effects model ³⁸.

For causal estimates from MR studies to be valid, three main assumptions must be met: 1) the genetic instrument is strongly associated with the level of accelerometer-measured physical activity; 2) the genetic instrument is not associated with any potential confounder of the physical activity – cancer association; and 3) the genetic instrument does not affect cancer independently of physical activity (i.e. horizontal pleiotropy should not be present) ³⁹. The strength of each instrument was measured by calculating the F statistic using the following formula: $F = R^2(N - 2)/(1 - R^2)$, where R^2 is the proportion of the variability of the physical activity explained by each instrument and N the sample size of the GWAS for the SNP-physical activity association ⁴⁰. To calculate R^2 we used the following formula: $(2 \times \text{EAF} \times (1 - \text{EAF}) \times \text{beta}^2) / [(2 \times \text{EAF} \times (1 - \text{EAF}) \times \text{beta}^2) + (2 \times \text{EAF} \times (1 - \text{EAF}) \times N \times \text{SE}(\text{beta})^2)]$, where EAF is the effect allele frequency, beta is the estimated genetic effect on physical activity, N is the sample size of the GWAS for the SNP-physical activity association and SE (beta) is the standard error of the genetic effect ⁴¹.

Sensitivity analyses

Several sensitivity analyses were used to check and correct for the presence of pleiotropy in the causal estimates. Cochran's Q was computed to quantify heterogeneity across the individual causal effects, with a $P\text{-value} \leq 0.05$ indicating the presence of pleiotropy, and that consequently, a random effects IVW MR analysis should be used^{38,42}. We also assessed the potential presence of horizontal pleiotropy using MR-Egger regression based on its intercept term, where deviation from zero denotes the presence of pleiotropy. Additionally, the slope of the MR-Egger regression provides valid MR estimates in the presence of horizontal pleiotropy when the pleiotropic effects of the genetic variants are independent from the genetic associations with the exposure^{43,44}. We also computed OR estimates using the complementary weighted-median method that can give valid MR estimates under the presence of horizontal pleiotropy when up to 50% of the included instruments are invalid³⁹. The presence of pleiotropy was also assessed using the MR-PRESSO. In this, outlying SNPs are excluded from the accelerometer-measured physical activity instrument and the effect estimates are reassessed⁴⁵. A leave-one-SNP out analysis was also conducted to assess the influence of individual variants on the observed associations. We also examined the selected genetic instruments and their proxies ($r^2 > 0.8$) and their associations with secondary phenotypes ($P\text{-value} < 5 \times 10^{-8}$) in Phenoscanner (<http://www.phenoscanner.medschl.cam.ac.uk/>) and GWAS catalog (date checked April 2019).

We also conducted multivariable MR analyses to adjust for potential pleiotropy due to BMI because the initial GWAS on physical activity reported several strong associations ($P\text{-value} < 10^{-5}$) between the identified SNPs and BMI⁴⁶. The new estimates correspond to the direct causal effect of physical activity with the BMI being fixed. The genetic data on BMI were obtained from a GWAS study published by The Genetic Investigation of

Anthropometric Traits (GIANT) consortium⁴⁷ (Supplementary Table 5). Additionally, we also conducted analyses with adiposity related SNPs (i.e. those previously associated with BMI, waist circumference, weight, or body/trunk fat percentage in GWAS studies at P-value $<10^{-8}$) excluded (n=5; rs34517439, rs6775319, rs11012732, rs1550435, rs59499656).

Finally, as the GECCO consortium includes 26,763 participants from the UK Biobank, we re-ran the colorectal cancer analyses using GWAS summary estimates with UK Biobank participants excluded in order to correct for any bias and inflated Type 1 errors this may have introduced into our results⁴⁸.

All the analyses were conducted using the MendelianRandomization package and R programming language⁴⁹.

Data availability

Data supporting the findings of this study are available within the paper and its supplementary information files.

References

- 1 Bray, F. *et al.* Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* **68**, 394-424, doi:10.3322/caac.21492 (2018).
- 2 World Health Organization. *Global status report on noncommunicable diseases 2014*, <<http://www.who.int/iris/handle/10665/148114>> (2014).
- 3 Moore, S. C. *et al.* Association of Leisure-Time Physical Activity With Risk of 26 Types of Cancer in 1.44 Million Adults. *JAMA Intern Med* **176**, 816-825, doi:10.1001/jamainternmed.2016.1548 (2016).
- 4 Kyu, H. H. *et al.* Physical activity and risk of breast cancer, colon cancer, diabetes, ischemic heart disease, and ischemic stroke events: systematic review and dose-response meta-analysis for the Global Burden of Disease Study 2013. *BMJ* **354**, i3857, doi:10.1136/bmj.i3857 (2016).
- 5 Morris, J. S., Bradbury, K. E., Cross, A. J., Gunter, M. J. & Murphy, N. Physical activity, sedentary behaviour and colorectal cancer risk in the UK Biobank. *Br J Cancer* **118**, 920-929, doi:10.1038/bjc.2017.496 (2018).
- 6 Prince, S. A. *et al.* A comparison of direct versus self-report measures for assessing physical activity in adults: a systematic review. *Int J Behav Nutr Phys Act* **5**, 56, doi:10.1186/1479-5868-5-56 (2008).
- 7 Doherty, A. *et al.* Large Scale Population Assessment of Physical Activity Using Wrist Worn Accelerometers: The UK Biobank Study. *PLoS One* **12**, e0169649, doi:10.1371/journal.pone.0169649 (2017).
- 8 Smith, G. D. & Ebrahim, S. 'Mendelian randomization': can genetic epidemiology contribute to understanding environmental determinants of disease? *Int J Epidemiol* **32**, 1-22 (2003).
- 9 Lawlor, D. A., Harbord, R. M., Sterne, J. A., Timpson, N. & Davey Smith, G. Mendelian randomization: using genes as instruments for making causal inferences in epidemiology. *Stat Med* **27**, 1133-1163, doi:10.1002/sim.3034 (2008).
- 10 Klimentidis, Y. C. *et al.* Genome-wide association study of habitual physical activity in over 377,000 UK Biobank participants identifies multiple variants including CADM2 and APOE. *Int J Obes (Lond)* **42**, 1161-1176, doi:10.1038/s41366-018-0120-3 (2018).
- 11 Michailidou, K. *et al.* Association analysis identifies 65 new breast cancer risk loci. *Nature* **551**, 92-94, doi:10.1038/nature24284 (2017).
- 12 Fred Hutchinson Cancer Research Center. *Genetics and Epidemiology of Colorectal Cancer Consortium (GECCO)*, <<https://www.fredhutch.org/en/labs/phs/projects/cancer-prevention/projects/gecco.html>> (
- 13 World Cancer Research Fund / American Institute for Cancer Research. Diet, Nutrition, Physical Activity and Cancer: a Global Perspective. Continuous Update Project Expert Report. (2018).
- 14 Ballard-Barbash, R. *et al.* Physical activity, weight control, and breast cancer risk and survival: clinical trial rationale and design considerations. *J Natl Cancer Inst* **101**, 630-643, doi:10.1093/jnci/djp068 (2009).
- 15 Friedenreich, C. M., Shaw, E., Neilson, H. K. & Brenner, D. R. Epidemiology and biology of physical activity and cancer recurrence. *J Mol Med (Berl)* **95**, 1029-1041, doi:10.1007/s00109-017-1558-9 (2017).

- 16 World Cancer Research Fund / American Institute for Cancer Research. Continuous Update Project Expert Report 2018. Diet, nutrition, physical activity and breast cancer.
- 17 World Cancer Research Fund / American Institute for Cancer Research. Continuous Update Project Expert Report 2018. Diet, nutrition, physical activity and colorectal cancer.
- 18 Bowers, L. W., Rossi, E. L., O'Flanagan, C. H., deGraffenried, L. A. & Hursting, S. D. The Role of the Insulin/IGF System in Cancer: Lessons Learned from Clinical Trials and the Energy Balance-Cancer Link. *Front Endocrinol (Lausanne)* **6**, 77, doi:10.3389/fendo.2015.00077 (2015).
- 19 Ulrich, C. M., Himbert, C., Holowatyj, A. N. & Hursting, S. D. Energy balance and gastrointestinal cancer: risk, interventions, outcomes and mechanisms. *Nat Rev Gastroenterol Hepatol* **15**, 683-698, doi:10.1038/s41575-018-0053-2 (2018).
- 20 Pollak, M. Insulin and insulin-like growth factor signalling in neoplasia. *Nat Rev Cancer* **8**, 915-928, doi:10.1038/nrc2536 (2008).
- 21 Endogenous, H. *et al.* Insulin-like growth factor 1 (IGF1), IGF binding protein 3 (IGFBP3), and breast cancer risk: pooled individual data analysis of 17 prospective studies. *Lancet Oncol* **11**, 530-542, doi:10.1016/S1470-2045(10)70095-4 (2010).
- 22 Shu, X. *et al.* Associations of obesity and circulating insulin and glucose with breast cancer risk: a Mendelian randomization analysis. *Int J Epidemiol*, doi:10.1093/ije/dyy201 (2018).
- 23 Murphy, N. *et al.* A Nested Case-Control Study of Metabolically Defined Body Size Phenotypes and Risk of Colorectal Cancer in the European Prospective Investigation into Cancer and Nutrition (EPIC). *PLoS Med* **13**, e1001988, doi:10.1371/journal.pmed.1001988 (2016).
- 24 McTiernan, A. *et al.* Effect of exercise on serum estrogens in postmenopausal women: a 12-month randomized clinical trial. *Cancer Res* **64**, 2923-2928 (2004).
- 25 Liedtke, S. *et al.* Physical activity and endogenous sex hormones in postmenopausal women: to what extent are observed associations confounded or modified by BMI? *Cancer Causes Control* **22**, 81-89, doi:10.1007/s10552-010-9677-4 (2011).
- 26 Bertone-Johnson, E. R., Tworoger, S. S. & Hankinson, S. E. Recreational physical activity and steroid hormone levels in postmenopausal women. *Am J Epidemiol* **170**, 1095-1104, doi:10.1093/aje/kwp254 (2009).
- 27 Key, T. *et al.* Endogenous sex hormones and breast cancer in postmenopausal women: reanalysis of nine prospective studies. *J Natl Cancer Inst* **94**, 606-616 (2002).
- 28 Endogenous, H. *et al.* Sex hormones and risk of breast cancer in premenopausal women: a collaborative reanalysis of individual participant data from seven prospective studies. *Lancet Oncol* **14**, 1009-1019, doi:10.1016/S1470-2045(13)70301-2 (2013).
- 29 Friedenreich, C. M., Neilson, H. K. & Lynch, B. M. State of the epidemiological evidence on physical activity and cancer prevention. *Eur J Cancer* **46**, 2593-2604, doi:10.1016/j.ejca.2010.07.028 (2010).
- 30 Zhang, X., Ashcraft, K. A., Betof Warner, A., Nair, S. K. & Dewhirst, M. W. Can Exercise-Induced Modulation of the Tumor Physiologic Microenvironment Improve Antitumor Immunity? *Cancer Research*, doi:10.1158/0008-5472.Can-18-2468 (2019).
- 31 McTiernan, A. Mechanisms linking physical activity with cancer. *Nat Rev Cancer* **8**, 205-211, doi:10.1038/nrc2325 (2008).
- 32 Woods, J. A., Vieira, V. J. & Keylock, K. T. Exercise, inflammation, and innate immunity. *Neurol Clin* **24**, 585-599, doi:10.1016/j.ncl.2006.03.008 (2006).

- 33 Helmkink, B. A., Khan, M. A. W., Hermann, A., Gopalakrishnan, V. & Wargo, J. A. The microbiome, cancer, and cancer therapy. *Nat Med* **25**, 377-388, doi:10.1038/s41591-019-0377-7 (2019).
- 34 Fernandez, D. M., Clemente, J. C. & Giannarelli, C. Physical Activity, Immune System, and the Microbiome in Cardiovascular Disease. *Front Physiol* **9**, 763, doi:10.3389/fphys.2018.00763 (2018).
- 35 Allen, J. M. *et al.* Exercise Alters Gut Microbiota Composition and Function in Lean and Obese Humans. *Med Sci Sports Exerc* **50**, 747-757, doi:10.1249/MSS.0000000000001495 (2018).
- 36 Choi, K. W. *et al.* Assessment of Bidirectional Relationships Between Physical Activity and Depression Among Adults: A 2-Sample Mendelian Randomization Study. *JAMA Psychiatry*, doi:10.1001/jamapsychiatry.2018.4175 (2019).
- 37 Brion, M. J., Shakhbazov, K. & Visscher, P. M. Calculating statistical power in Mendelian randomization studies. *Int J Epidemiol* **42**, 1497-1501, doi:10.1093/ije/dyt179 (2013).
- 38 Higgins, J. P., Thompson, S. G., Deeks, J. J. & Altman, D. G. Measuring inconsistency in meta-analyses. *BMJ* **327**, 557-560, doi:10.1136/bmj.327.7414.557 (2003).
- 39 Bowden, J., Davey Smith, G., Haycock, P. C. & Burgess, S. Consistent Estimation in Mendelian Randomization with Some Invalid Instruments Using a Weighted Median Estimator. *Genet Epidemiol* **40**, 304-314, doi:10.1002/gepi.21965 (2016).
- 40 Burgess, S., Thompson, S. G. & Collaboration, C. C. G. Avoiding bias from weak instruments in Mendelian randomization studies. *Int J Epidemiol* **40**, 755-764, doi:10.1093/ije/dyr036 (2011).
- 41 Shim, H. *et al.* A multivariate genome-wide association analysis of 10 LDL subfractions, and their response to statin treatment, in 1868 Caucasians. *PLoS One* **10**, e0120758, doi:10.1371/journal.pone.0120758 (2015).
- 42 Bowden, J. *et al.* A framework for the investigation of pleiotropy in two-sample summary data Mendelian randomization. *Stat Med* **36**, 1783-1802, doi:10.1002/sim.7221 (2017).
- 43 Bowden, J., Davey Smith, G. & Burgess, S. Mendelian randomization with invalid instruments: effect estimation and bias detection through Egger regression. *Int J Epidemiol* **44**, 512-525, doi:10.1093/ije/dyv080 (2015).
- 44 Burgess, S. & Thompson, S. G. Interpreting findings from Mendelian randomization using the MR-Egger method. *Eur J Epidemiol* **32**, 377-389, doi:10.1007/s10654-017-0255-x (2017).
- 45 Verbanck, M., Chen, C. Y., Neale, B. & Do, R. Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases. *Nat Genet* **50**, 693-698, doi:10.1038/s41588-018-0099-7 (2018).
- 46 Burgess, S. & Thompson, S. G. Multivariable Mendelian randomization: the use of pleiotropic genetic variants to estimate causal effects. *Am J Epidemiol* **181**, 251-260, doi:10.1093/aje/kwu283 (2015).
- 47 Locke, A. E. *et al.* Genetic studies of body mass index yield new insights for obesity biology. *Nature* **518**, 197-206, doi:10.1038/nature14177 (2015).
- 48 Burgess, S., Davies, N. M. & Thompson, S. G. Bias due to participant overlap in two-sample Mendelian randomization. *Genet Epidemiol* **40**, 597-608, doi:10.1002/gepi.21998 (2016).

- 49 Yavorska, O. O. & Burgess, S. MendelianRandomization: an R package for performing Mendelian randomization analyses using summarized data. *Int J Epidemiol* **46**, 1734-1739, doi:10.1093/ije/dyx034 (2017).

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Figure legends

Figure 1: Mendelian randomization analysis for individual SNPs associated with accelerometer-measured physical activity in relation to breast cancer risk. The x axis corresponds to a log OR per one unit increase in the physical activity based on the average acceleration (milli-gravities). The Mendelian randomization (MR) result corresponds to a random effects model due to heterogeneity across the genetic instruments. logOR = log odds ratio. 95% CI = 95% confidence interval. SNP = single nucleotide polymorphism.

Figure 2: Mendelian randomization analysis for individual SNPs associated with accelerometer-measured physical activity in relation to colorectal cancer risk (overall, colon, rectal). The x axis corresponds to a log OR per one unit increase in the physical activity based on the average acceleration (milli-gravities). The Mendelian randomization (MR) result corresponds to a random effects model due to heterogeneity across the genetic instruments. logOR = log odds ratio. 95% CI = 95% confidence interval. SNP = single nucleotide polymorphism.

Table 1: Summary information on accelerometer-measured physical activity for the 10 SNPs used as genetic instruments for Mendelian randomization analyses

SNP	Effect allele	Baseline allele	Chromosome	Position*	Gene	EAF	beta PA [†]	se PA	N [‡]	R	F statistic
rs12045968	G	T	1	33225097	ZNF362	0.22	0.239	0.044	91,084	0.0003	30
rs34517439	C	A	1	77984833	DNAJB4	0.91	0.308	0.056	91,084	0.0003	30
rs6775319	A	T	3	18717009	LOC105376976	0.3	0.225	0.041	91,084	0.0003	30
rs12522261	G	A	5	152675265	LINC01470	0.67	0.211	0.038	91,084	0.0003	31
rs9293503	T	C	5	88653144	LINC00461	0.88	0.329	0.059	91,084	0.0003	31
rs11012732	A	G	10	21541175	MLLT10	0.65	0.225	0.039	91,084	0.0004	33
rs148193266	C	A	11	104657953	RP11-681H10.1	0.02	0.51	0.092	91,084	0.0003	31
rs1550435	T	C	15	74039044	PML	0.53	0.2	0.037	91,084	0.0003	29
rs55657917	G	T	17	45767194	CRHR1	0.22	0.3	0.04	91,084	0.0006	56
rs59499656	T	A	18	43188344	RIT2/SYT4	0.34	0.228	0.038	91,084	0.0004	36

Abbreviations: EAF: effect allele frequency; PA, physical activity; se: standard error; SNP, single nucleotide polymorphism

* Position based on GRCh38.p12

[†] The beta coefficients are expressed in milligravities

[‡] N refers to the sample size of the initial GWAS from which the genetic variants were selected

Table 2. Mendelian Randomization estimates between accelerometer-measured physical activity and cancer risk

Methods	No. Cases	Extended number of SNPs (n=10)				Genome-wide SNPs only (n=2)			
		Estimates (OR)*	95% CI	P-value	P-value for pleiotropy [†] or heterogeneity [‡]	Estimates (OR)*	95% CI	P-value	P-value for pleiotropy [†] or heterogeneity [‡]
Breast Cancer									
Inverse-variance weighted [§]	122,977	0.59	0.42, 0.84	0.003	6.8×10 ⁻⁷	0.42	0.16, 1.16	0.09	9.4×10 ⁻⁴
MR-Egger		0.55	0.09, 3.20	0.50	0.9				
Weighted median		0.76	0.59, 0.98	0.03					
ER^{+ve} subset									
Inverse-variance weighted [§]	69,501	0.53	0.35, 0.82	0.004	0.004	0.43	0.20, 0.91	0.03	0.04
MR-Egger		0.61	0.07, 5.26	0.65	0.9				
Weighted median		0.66	0.48, 0.90	0.008					
ER^{-ve} subset									
Inverse-variance weighted [§]	21,468	0.78	0.51, 1.22	0.27	0.01	0.51	0.29, 0.89	0.02	0.11
MR-Egger		0.24	0.03, 1.81	0.17	0.24				
Weighted median		0.70	0.47, 1.04	0.08					
Colorectal Cancer									
Inverse-variance weighted	58,221	0.66	0.53, 0.82	1.9×10 ⁻⁴	0.56	0.64	0.42, 0.99	0.045	1
MR-Egger		0.29	0.11, 0.80	0.016	0.10				
Weighted median		0.67	0.50, 0.90	0.007					
Colorectal Cancer in men									
Inverse-variance weighted	30,933	0.82	0.61, 1.11	0.21	0.89	0.99	0.54, 1.81	0.98	0.31
MR-Egger		0.85	0.21, 3.42	0.82	0.97				
Weighted median		0.90	0.61, 1.33	0.60					
Colorectal Cancer in women									
Inverse-variance weighted	26,848	0.54	0.40, 0.74	1.2×10 ⁻⁴	0.08	0.45	0.24, 0.84	0.012	0.26
MR-Egger		0.10	0.02, 0.52	0.006	0.04				
Weighted median		0.56	0.36, 0.88	0.01					
Colon Cancer									
Inverse-variance weighted	30,621	0.61	0.47, 0.79	1.6×10 ⁻⁴	0.57	0.57	0.34, 0.95	0.032	0.31
MR-Egger		0.44	0.13, 1.45	0.18	0.59				

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Weighted median		0.55	0.39, 0.78	0.001					
Proximal Colon Cancer									
Inverse-variance weighted		0.69	0.50, 0.97	0.03	0.83	0.75	0.39, 1.45	0.39	0.45
MR-Egger	13,864	0.37	0.08, 1.75	0.21	0.42				
Weighted median		0.68	0.44, 1.04	0.08					
Distal Colon Cancer									
Inverse-variance weighted		0.48	0.34, 0.67	1.9×10^{-5}	0.75	0.36	0.18, 0.71	0.003	0.21
MR-Egger	14,940	0.39	0.08, 1.89	0.24	0.8				
Weighted median		0.5	0.31, 0.78	0.03					
Rectal Cancer									
Inverse-variance weighted		0.76	0.55, 1.07	0.12	0.64	0.91	0.47, 1.77	0.78	0.91
MR-Egger	15,859	0.37	0.08, 1.75	0.21	0.46				
Weighted median		0.91	0.58, 1.42	0.69					

Abbreviations: CI, confidence intervals; MR: Mendelian Randomization; OR: odds ratio; SNPs: Single nucleotide polymorphisms

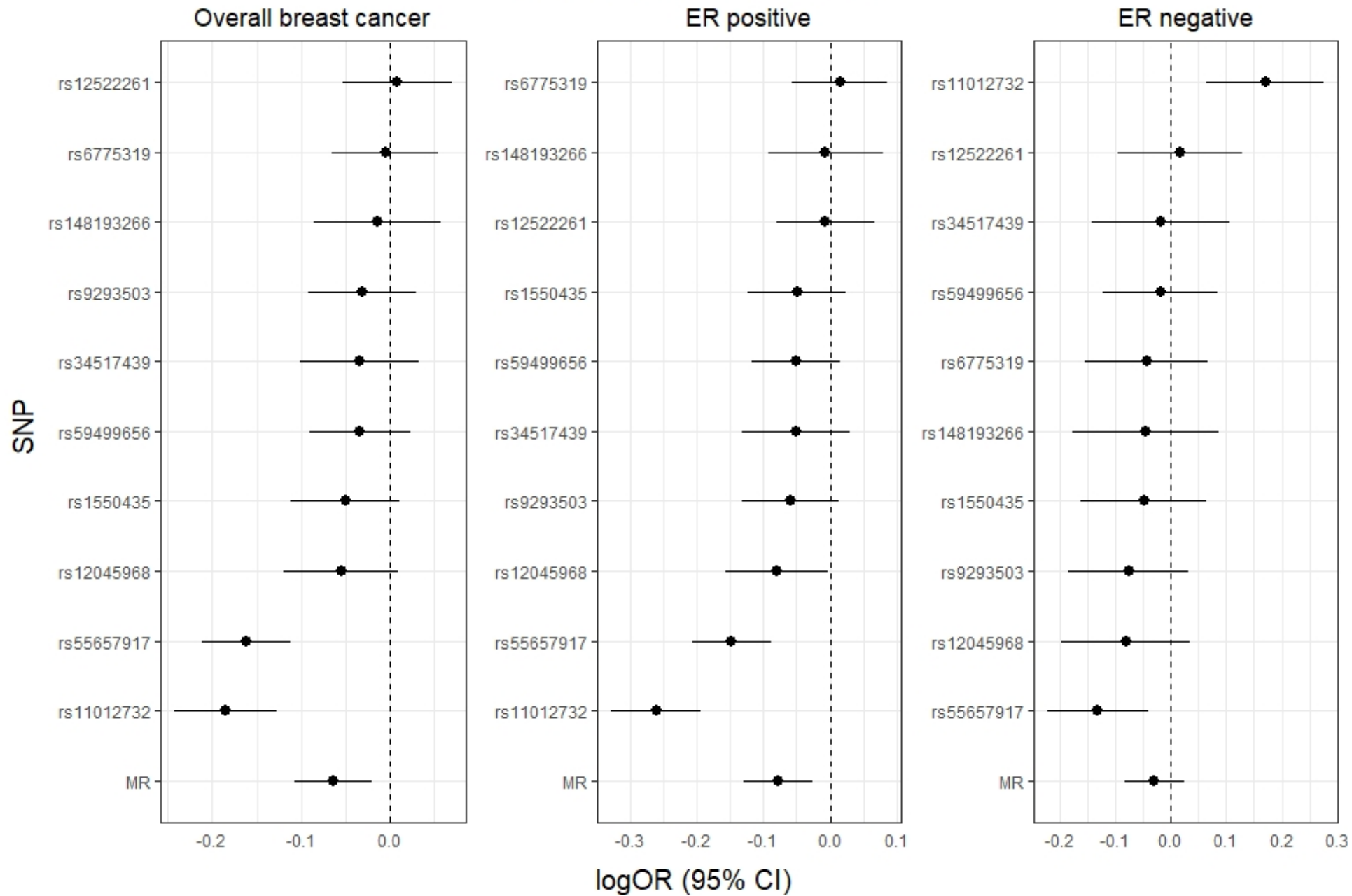
* The estimates correspond to a standard deviation increase in physical activity

† P-value or pleiotropy based on MR-Egger intercept

‡ P-value for heterogeneity based on Q statistic

§ The estimates were derived from a random-effects model due to the presence of heterogeneity based on Cochran's Q statistic

Accelerometer-Based Activity



Accelerometer-Based Activity

