Title: Hydroxychloroquine Proves Ineffective in Hamsters and Macaques Infected with SARS-CoV-2

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Short Title: Hydroxychloroquine in COVID-19 models

One Sentence Summary: Hydroxychloroquine prophylaxis/treatment showed no beneficial effect in SARS-CoV-2 hamster and macaque disease models.

We remain largely without effective prophylactic/therapeutic interventions for COVID-19. Although many human clinical trials are ongoing, there remains a deficiency of supportive preclinical drug efficacy studies. Here we assessed the prophylactic/therapeutic efficacy of hydroxychloroquine (HCQ), a drug of interest for COVID-19 management, in two animal models. When used for prophylaxis or treatment neither the standard human malaria dose (6.5 mg/kg) nor a high dose (50 mg/kg) of HCQ had any beneficial effect on clinical disease or SARS-CoV-2 kinetics (replication/shedding) in the Syrian hamster disease model. Similarly, HCQ prophylaxis/treatment (6.5 mg/kg) did not significantly benefit clinical outcome nor reduce SARS-CoV-2 replication/shedding in the upper and lower respiratory tract in the rhesus macaque disease model. In conclusion, our preclinical animal studies do not support the use of HCQ in prophylaxis/treatment of COVID-19.

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is the causative agent of coronavirus disease 2019 (COVID-19) (*1*). SARS-CoV-2 infections were initially reported in China near the beginning of December 2019 (*2*). Following early spread through Asia, and subsequently to European, American and African countries, the virus is responsible for the third pandemic of the 21st Century. With currently over 6.6 million confirmed cases and >390,000 deaths worldwide, health systems are stretched beyond limit with largely no proven treatment or

prophylaxis available to reduce the burden (*3*). Public health measures combined with increasingly severe restrictions on public life have been implemented in many countries to stop SARS-CoV-2 transmission. The goal of current public health strategies is to flatten the epidemiologic SARS-CoV-2/COVID-19 curve to ease the burden on health care systems challenged by the highly intensive care required for a significant proportion of COVID-19 cases. Over 1,000 clinical trials are currently open or being established in different countries testing drugs such as lopinavir/ritonavir, dexamethasone, hydroxychloroquine (HCQ) and inhaled interferon beta-1a (*4*). Yet, many of these treatments have not been empirically tested in relevant SARS-CoV-2 animal disease models to determine preclinical efficacy, and thereby provide valuable insight into prioritization of drugs to move forward in humans.

At the time this work was started, the US FDA had given emergency approval for the use of chloroquine and HCQ in COVID-19 patients (*5*). *In vitro* data on the inhibitory effect of chloroquine and HCQ on SARS-CoV-2 replication had been published (*6-8*) and HCQ alone or in combination with the macrolide antibiotic azithromycin had been used in early clinical trials to treat COVID-19 cases with varying effect (*9-11*). Despite ongoing clinical trials, preclinical efficacy data on the effect of HCQ in SARS-CoV-2 animal disease models were lacking. Herein, we assessed the efficacy of HCQ prophylaxis and treatment in two established animal disease models, the Syrian hamster and rhesus macaque (*12, 13*).

First, we confirmed the *in vitro* inhibitory effect of HCQ on SARS-CoV-2 replication in Vero E6 cells. Cells were pretreated with differing drug concentrations and the effect on viral RNA load in tissue culture supernatant was determined 72 hours after infection by quantitative reverse transcriptase polymerase chain reaction (qRT-PCR) (fig. S1). The half-maximal effective concentration (EC_{50}) value for HCQ was 164.7nM, consistent with low/sub-micromolar levels

previously reported for the established *in vitro* inhibitory effect of HCQ on SARS-CoV-2 replication (*6-8*).

Having confirmed *in vitro* efficacy, we next tested the ability of HCQ to alter the course of SARS-CoV-2 in the Syrian hamster disease model (*12*). Five groups of hamsters (n=6 per group) were prophylactically or therapeutically treated with an intraperitoneal infection of a standard (6.5 mg/kg in PBS; human dose for malaria prophylaxis/treatment) or high (50 mg/kg in PBS) dose HCQ regimen; control groups were treated with vehicle only. Hamsters were intranasally infected with SARS-CoV-2 using a dose of $1x10^4$ median tissue culture infectious doses ($TCID_{50}$). For prophylaxis, a single treatment was performed 24 hours prior to infection. The therapeutic treatment started 1 hour after SARS-CoV-2 infection and was continued for 3 consecutive days. Disease manifestation in this model is transient and clinical signs peak between days 3 and 5 post-infection with ruffled fur, increased respiration rate and reduced mobility (*12*). Virus replication and shedding was determined by qRT-PCR in swab samples (oral and rectal) collected on days 2 and 4, and lung tissue taken at necropsy on day 4 post-infection. Regardless of HCQ administration, all animals showed comparable high levels of genome copy numbers for oral swabs ($>10^7$ genome copies/mL) and comparable lower numbers for rectal swabs ($<10^6$ genome copied/mL) decreasing in all groups over time (Fig. 1, A and B). Like viral RNA loads in swabs, there was no significant difference in disease manifestation over the time of the study. Gross lung pathology was similar among the groups consisting of focally extensive areas of consolidation that failed to collapse upon removal (fig. S2). Viral lung loads on day 4 were high (10^{14} genome copies/g) but indistinguishable between all groups (Fig. 1C). Lung to body weight ratios were similar in all animals with no significant difference between groups (Fig. 1D). Overall, HCQ administered either as prophylaxis or treatment at standard or

high doses did not have any significant impact on SARS-CoV-2 replication and shedding, nor disease manifestation and progression in the Syrian hamster model.

Next, we assessed HCQ efficacy in the rhesus macaque; a recently developed nonhuman primate model displaying mild to moderate COVID-like disease upon SARS-CoV-2 infection (*13*). Similar to the hamster study, we investigated the effect of HCQ when administered either prophylactically or as a treatment after infection. For the prophylactic arm, 10 healthy rhesus macaques were randomly divided into vehicle control and HCQ prophylaxis groups (n=5 per group). Animals were treated by oral gavage with either vehicle (PBS) or HCQ (6.5mg/kg in PBS) three times one week apart (day -9, day -2 and day 5) (Fig. 2A). To test the efficacy of HCQ as a treatment, a separate group of 10 healthy rhesus macaques were randomly divided into vehicle control and HCQ treatment groups (n=5 per group). Animals were treated by oral gavage with either vehicle (PBS) or HCQ (6.5mg/kg in PBS) starting 12 hours post-infection followed by treatment at 18, 36, 60, 84 and 108 hours post-infection (Fig. 2B). Animals in all groups were infected on day 0 with SARS-CoV-2 (total dose 2.8 x10^6 $TCID_{50}$ by a combination of four routes (intratracheal, oral, intranasal and ocular) as previously described (*13, 14*). Animals were monitored at least twice daily using an established scoring sheet designed to assess clinical signs of disease (*13,15*). Multiple physical examinations were performed on different days pre- and post-inoculation including a clinical evaluation, radiographs, blood collection, and swabs (oral and nasal). Bronchoalveolar lavage (BAL) was performed on days 3, 5 and 7 (post-mortem) (Fig. 2, A and B). The endpoint for both studies was day 7 post-infection, at which time all animals were euthanized and necropsied.

To ensure that drug was present in therapeutic quantities plasma levels of HCQ and its secondary metabolites were measured. HCQ was detected in plasma samples post-administration in all

prophylactically or therapeutically treated animals with concentration ranging from 1.2 to 10.5ng/mL (3.6 nM to 31.3 nM) and 8 to 98 ng/mL (23.8 nM to 291.8 nM), respectively (Fig. 2, C and D). HCQ was also detected in lung tissue at time of necropsy in all prophylactically or therapeutically treated animals ranging from 0.85 to 4.18 ng/mg tissue and 1.39 to 11.54 ng/mg tissue, respectively. These numbers are in good agreement with the reported long half-life and large volume of distribution of HCQ (*16*). HCQ cytochrome p450 catalyzed secondary amine metabolites desethylchloroquine and desethylhydroxychloroquine, and the primary amine metabolite bisdesethylchloroquine are considered to be active forms of the drug in other disease models (*17*). Both desethylchloroquine and desethylhydroxychloroquine were detected in intermediate concentrations, while trace amounts of bisdesethylchloroquine were detected in both plasma and lung homogenate suggesting substantial persistence of active drug forms over the course of treatment (fig. S3). The plasma HCQ levels measured here fall within or near human therapeutically relevant ranges for other disease such as malaria and systemic lupus erythematosus (15 to 100 ng/mL plasma) (*18,19*). However, since SARS-CoV2 is a respiratory disease, levels of drug in lung tissue are a better indicator of therapeutic potential. Volume/concentration is difficult to estimate in tissue due to compartmentalization resulting in a non-homogenous distribution of the drug. However, using a water content of 80% by weight (*20*), day 7 levels in the lung indicated conservative estimates of at least 1 µg/mL (~3.0 uM) in all animals, which is above the cell culture EC_{50} which we determined to be ~ 0.2 uM (164.7 nM, 55 ng/mL here (fig. S1).

Macaques in both the prophylactic and treatment arms of the study first displayed clinical signs of SARS-CoV-2 infection on day 1, which peaked at day 2 and animals remained mildly to moderately ill until the study endpoint at day 7 (Fig. 2, E and F). Clinical signs included reduced

136 appetite and ruffled fur followed by pale appearance and irregular increased abdominal
137 respiration (table S1). Overall, animals in the vehicle treated groups appeared to have slightly
138 higher clinical scores throughout, but daily differences were not statistically significant.
139 Hematology and serum chemistry were unremarkable for all animals in both study arms.
140 Radiographic signs in the prophylaxis, treatment and control groups were minimal over the study
141 course (fig. S4). Pulmonary infiltrates, when seen, were noted to be of a mild unstructured
142 interstitial pattern. The pattern was rarely seen in the upper lung, being more commonly found in
143 middle and caudal lung lobes. No differences were noted in severity or appearance of
144 radiographic signs between HCQ prophylaxis, treatment or control groups.

145 Nasal and oropharyngeal swabs were positive for SARS-CoV-2 RNA in all animals of both
146 studies with the highest load on either day 1 or day 3, which then gradually decreased until the
147 end of the study (Fig. 3, A – D). Viral load in nasal swabs were consistently higher than in
148 oropharyngeal swabs. BAL samples were collected on days 3, 5 and 7 (post-mortem) and viral
149 loads were similar to nasal and oropharyngeal swabs with decreasing loads over time (Fig. 3, E
150 and F). Overall, there were no statistically significant differences in virus load and shedding
151 between HCQ- and vehicle-administered animals in the prophylaxis and treatment regimens.

152 At necropsy, gross pathology revealed consolidated lungs in animals of all groups with lesions
153 observed largely in the lower lung lobes, although some of the legions may have been the result
154 of the post-mortem BAL (Fig. 4, A and B). All other gross pathology was normal except for
155 enlarged cervical and mediastinal lymph nodes in several animals across the groups. Histological
156 analysis of the lungs of animals in the different prophylaxis and treatment groups determined a
157 comparable degree of pulmonary pathology when inoculated with SARS-CoV-2 similar to what
158 had been published previously (*13,14*) (Fig. 4C). Lesions were mild to moderate and

159 characterized as multifocal interstitial pneumonia frequently centered on terminal bronchioles.
160 The pneumonia was evident by a thickening of alveolar septae by edema fluid and fibrin and
161 small to moderate numbers of macrophages and fewer neutrophils. Infiltration of small numbers
162 of pulmonary macrophages and neutrophils were noticed in alveoli. Lungs with moderate
163 changes also had alveolar edema and fibrin with formation of hyaline membranes. There was
164 minimal to moderate type II pneumocyte hyperplasia. Occasionally, bronchioles had necrosis,
165 and loss and attenuation of the epithelium with infiltrates of neutrophils, macrophages and
166 eosinophils. Perivascular infiltrates of small numbers of lymphocytes forming perivascular cuffs
167 were noticed multifocally (Fig. 4C). Overall, there was no significant difference between vehicle
168 and HCQ treated animals in either of the regimens, prophylaxis or treatment.

169 Viral RNA loads were determined in several respiratory tissues using qRT-PCR (Fig. 5, A and
170 C). Highest genome copy numbers were found in lung tissue with a marginal but not significant
171 benefit for the HCQ- over the vehicle-treated group in the prophylaxis study arm when all lung
172 lobe samples were combined (Fig. 5, B and D). Virus isolation from tissues was inconsistent
173 among animals in the different groups, but at least one sample in each group showed infectious
174 virus for almost all respiratory tissues (Fig. 5, A and C). There was no difference between
175 animals of vehicle- and HCQ-treated groups in the prophylaxis and treatment study arms, which
176 is consistent with the lack of any observed effect of HCQ on virus shedding parameters.

177 In this study we used two established COVID-like animal models (*12,13*) and applied the
178 standard weight-based oral administration of HCQ prophylaxis and treatment of malaria in
179 humans (*21*). For the Syrian hamster model, we also included a high HCQ dose regimen (7.5
180 times the standard dose regimen) both prophylactically and as a treatment. For prophylaxis we
181 used a weekly dosing regimen. For treatment, we administered HCQ starting shortly after

infection and continued daily until study end. HCQ pharmacokinetic studies in humans and animal models have demonstrated a rapid blood bioavailability following oral administration with peak levels being reached in 2 to 4 hours followed by rapid absorption in various tissues including the lung (*22,23*). Samples for drug pharmacokinetics in plasma were collected when the drug levels were low, just before the administration of the next treatment. Nevertheless, the measurements taken during both studies are in good agreement with data from humans and animal models and suggest accumulation of drug in the lung at therapeutic levels (*18,19*).

The use of HCQ and chloroquine as treatment options for COVID-19 patients may have been partially rooted in early observations for their effect in impairing SARS-CoV-2 replication *in vitro* (*6-8*). These *in vitro* studies, which we confirmed herein, identified HCQ (and other 4-aminoquinolines) as potent inhibitors of coronaviruses, including SARS-CoV-2, with low EC_{50} values within the range of antivirals such as remdesivir (*6*); a drug that is now approved for COVID-19 cases by the FDA. The mechanism of action of 4-aminoquinolones against SARS-CoV-2 *in vitro* is not well defined, but increasing endosomal pH, inhibition of autophagosome-lysosome fusion, impairment of enzymes important for virus replication, and effects on protein glycosylation have been proposed, which may result in interference with SARS-CoV-2 entry/fusion, replication and spread (*24, 25*). However, despite the promising *in vitro* effect observed by us and others, we did not observe any significant prophylactic or therapeutic benefit of HCQ following *in vivo* infection in two animal disease models.

The use of HCQ to treat COVID-19 has been controversial since the results of the first clinical trials (*9-11*). Nevertheless, HCQ has been promoted as a COVID-19 treatment option and became part of multiple recent large-scale clinical trials including one of four initial treatment options in the multinational WHO “Solidarity” clinical trial for COVID-19 (*26*). However, HCQ

treatment does not come without risks as the 4-aminoquinolones are associated with multiple adverse effects such cutaneous adverse reactions, hepatic failure, and ventricular arrythmia; overdose is also difficult to treat (*21*). The US FDA recently updated its guidance by warning against use of HCQ outside of the hospital setting because of the potential for serious adverse effects (*27*). Over past weeks, several clinical trials, such as the WHO Solidarity study, have been stopped or have excluded HCQ arms due to a lack of evidence for therapeutic efficacy, and an increase level of adverse effects in COVID-19 patients (*26, 28, 29*). One influential study that had indicated a detrimental effect of HCQ in COVID-19 patients has subsequently been retracted by the authors due to their inability to confirm the veracity of the data (*29, 30)*, and the Solidarity HCQ arm has been resumed (*26*). Similarly, a multinational UK-based (COPCOV) HCQ prophylactic trial involving healthcare workers at high risk for SARS-CoV-2 infection was paused less than a week after starting due to safety concerns (*31*); the impact of the retraction on the status of this trial remains to be ascertained. Clearly, the effectiveness of HCQ to prevent or reduce infection and thereby impact the clinical course of COVID-19 remains highly contentious at this time.

In conclusion, HCQ prophylaxis and treatment had no beneficial effect in the two animal disease models tested. There is always the consideration as to what extent animal data can be extended to the situation in humans, but in general the nonhuman primate models are considered good indicators and the ultimate preclinical models before moving drugs into clinical trials. Independent of the safety issues associated with HCQ, the preclinical data presented here does not support HCQ and likely other 4-aminoquinolines as being either an effective prophylactic treatment to reduce SARS-CoV-2 infection or therapeutic for use in COVID-19 patients.

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235

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241 **References**

242 1. R. Lu *et al.*, Genomic characterisation and epidemiology of 2019 novel coronavirus:
243 implications for virus origins and receptor binding. *Lancet* **395**, 565-574 (2020).

244 2. N. Chen *et al*., Epidemiological and clinical characteristics of 99 cases of 2019 novel
245 coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet* **395**, 507-13 (2020).

246 3. World Health Organization (WHO), "Coronavirus disease (COVID-2019) Situation
247 Report - 129" (WHO, 2020); https://www.who.int/docs/default-source/coronaviruse/
248 situation-reports/20200528-covid-19-sitrep-129.pdf?sfvrsn=5b154880_2

4. National Institutes of Health (NIH), US National Library of Medicine (NIH, 2020); https://clinicaltrials.gov/ct2/results ?cond=COVID-19&age_v=&gndr=&type=Intr&rslt =&Search=Apply

5. Lenzer J COBID-19: US gives emergency approval to hydroxychloroquine despite lack of evidence. BMJ, **369**, m1335 (2020).

6. M. Wang *et al*., Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) *in vitro*. *Cell Research* **30**, 269-271 (2020).

7. J. Liu *et al*., Hydrochloroquine, a less toxic derivative of chloroquine, is effective in inhibiting SARS-CoV-2 infection *in vitro*. *Cell Discovery* ***6,*** 16 (2020).

8. X. Yao et al. *In vitro* antiviral activity and projection of optimized dosing design of hydroxychloroquine for the treatment of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). [published online ahead of print, 2020 Mar 9]. *Clin Infect Dis*. (2020), https://doi:10.1093/cid/ciaa237

9. Gao J, Tian Z, Yang X. Breakthrough: Chloroquine phosphate has shown apparent efficacy in treatment of COVID-19 associated pneumonia in clinical studies. *BioScience Trends* **14**, 72-73 (2020).

10. Z. Chen Z *et al*., Efficacy of hydroxychloroquine in patients with COVID-19; results of a randomized clinical trial. medRxiv (2020), https://www.medrxiv.org/content/10.1101/ 2020.03.22.20040758v3

11. P. Gautret *et a*l., Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial [published online ahead of print,

2020 Mar 20]. *International Journal of Antimicrobial Agents* (2020), https://doi.org/10.1016/j.ijantimicag.2020.105949

12. J.F. Chan *et al*., Simulation of the clinical and pathological manifestations of Coronavirus Disease 2019 (COVID-19) in golden Syrian hamster model: implications for disease pathogenesis and transmissibility [published online ahead of print, 2020 Mar 26]. *Clinical Infectious Diseases* (2020), https://doi.org/10.1093/cid/ciaa325

13. V.J. Munster *et al.*, Respiratory disease in rhesus macaques inoculated with SARS-CoV-2 [published online ahead of print, 2020 May 12]. *Nature* (2020), https://doi.org/10.1038/s41586-020-2324-7

14. B.N. Williamson et al., Clinical benefit of remdesivir in rhesus macaques infected with SARS-CoV-2. bioRxiv (2020), https://doi.org/10.1101/2020.04.15.043166

15. D.L. Brining et al., Thoracic radiography as a refinement methodology for the study of H1N1 influenza in cynomologus macaques (*Macaca fascicularis*). *Comparative Medicine* **60**, 389-395 (2010).

16. E. Schrezenmeier and T. Dörner. "Mechanisms of action of hydroxychloroquine and chloroquine: implications for rheumatology." *Nature Reviews Rheumatology* **16**, 155-166 (2020).

17. H. Fan et al. "Pharmacokinetics and bioequivalence study of hydroxychloroquine sulfate tablets in Chinese healthy volunteers by LC–MS/MS." *Rheumatology and Therapy* **2**, 183-195 (2015):

18. L. Durcan et al., Hydroxychloroquine blood levels in systemic lupus erythematosus: clarifying dosing controversies and improving adherence. *The Journal of Rheumatology* **42**, 2092-2097 (2015).

19. R.I. Fox, In Mechanism of action of hydroxychloroquine as an antirheumatic drug. *Seminars in Arthritis and Rheumatism*, Elsevier; pp 82-91 (1993).

20. N.R. Lange and D.P. Schuster. The measurement of lung water. *Critical Care* **3**, R19 (1999).

21. American Hospital Formulary Service (AHFS) Drug Information, 2020.

22. E.W. McChesney, Animal toxicity and pharmacokinetics of hydroxychloroquine sulfate. *American Journal of Medicine* **75**, 11-18 (1983).

23. H.S. Lim et al., Pharmacokinetics of hydroxychloroquine and its clinical implications in chemoprophylaxis against malaria caused by Plasmodium vivax. *Antimicrobial Agents and Chemotherapy* **53**, 1468-1475 (2009).

24. C. Salata *et al.,* Antiviral activity of cationic amphiphilic drugs. *Expert Review of Anti-infective Therapy* **15**, 483-492 (2017).

25. M.J. Vincent *et al*., Chloroquine is a potent inhibitor of SARS coronavirus infection and spread. *Virology Journal* **2**, 69 (2005).

26. World Health Organization (WHO). "Solidarity" clinical trial for COVID-19 treatments. (WHO, 2020); https://www.who.int/emergencies/diseases/novel-coronavirus-2019/global-research-on-novel-coronavirus-2019-ncov/solidarity-clinical-trial-for-covid-19-treatments

27. US Food and Drug Administration (US FDA). "Drug Safety and Availability" (US FDA, 2020); https://www.fda.gov/drugs/drug-safety-and-availability/fda-cautions-against-use-hydroxychloroquine-or-chloroquine-covid-19-outside-hospital-setting-or).

28. M.R. Mehra *et al.,* Hydroxychloroquine or chloroquine with or without a macrolide for treatment of COVID-19: a multinational registry analysis [published online ahead of print, 2020 May 22]. *Lancet,* https://doi.org/10.1016/S0140-6736(20)31180-6 (2020).

29. J. Magagnoli *et al*., Outcomes of hydroxychloroquine usage in United States veterans hospitalized with Covid-19. medRxiv (2020), https://doi.org/10.1101/2020.04.16.20065920

30. M.R. Mehra et al., Retraction– Hydroxychloroquine or chloroquine with or without a macrolide for treatment of COVID-19: a multinational registry analysis [published online ahead of print, 2020 June 4]. *Lancet,* https://doi.org/10.1016/S0140-6736(20)31324-6 (2020).

31. MORU Tropical Health Network. "COPCOV Trial" (2020); (https://www.tropmedres.ac/covid-19/copcov/copcov-news)

Figure Legends

Figure 1: Syrian hamster model - viral shedding, viral load and pathology. Hamsters were infected with SARS-CoV-2 by the intranasal route. HCQ was administered either prophylactically one time at 24 hours prior to infection (6.5mg/kg and 50mg/kg) or treatment started 1 hour post-infection for 3 consecutive days (6.5mg/kg and 50mg/kg). Hamsters were

scored for clinical signs daily and swabs (oral and rectal) were collected on day 2 and 4. Animals were euthanized on day 4 and lungs were harvested for pathology and virology. Swab and lung loads were determined by qRT-PCR. (A and B) Viral shedding. Oral and rectal swabs from day 2 and 4 were analyzed for viral genome copies by qRT-PCR. Swabs were analyzed as a correlate for viral shedding. (C) Viral load in lung tissue. Lung viral loads (genome copies) were determined by as a correlate for lower respiratory tract infection. No statistical significance was found among the groups presented in parts (A) to (C). (D) Lung to body weight ratio. Lung to body weight ratio was determined as an indicator for pneumonia with lung edema. Statistically significant differences were only found when compared to lung to body weight ratios of naïve hamsters. Multiple t tests were used to analyze differences among groups.

Figure 2: Rhesus macaque model – design, drug concentrations and clinical scoring. Macaques were infected with SARS-CoV-2 by the combined intratracheal, intranasal, oral and ocular routes. Animals were treated by oral gavage with either vehicle (PBS) or HCQ (6.5mg/kg in PBS). Administration was either one time per week for the prophylaxis arm or starting 12 hours post-infection followed by treatment at 18, 36, 60, 86 and 108 hours post-infection for the treatment arm. Animals were scored for clinical disease twice daily and examinations were performed as indicated. (A and B) Study design. The schematic depicts infection ('I'), HCQ or vehicle treatment ('T') and examinations ('E'). (C and D) Plasma levels of HCQ. HCQ levels were determined in both the prophylaxis and treatment study arms. Measurements reflect pre-dose levels of HCQ at each timepoint (limit of quantification = 0.5 ng/mL). (E and F) Clinical scores. Clinical scoring was performed twice daily by observation of non-anesthetized animals. The morning score is graphed here. Multiple t tests performed on individual days found no significance difference between groups. Area under the curve analysis was performed on each

355 individual animal in each study. This analysis found a significant difference (p=0.004) between
356 groups in the therapeutic study only. *Note:* red squares, vehicle-treated animals; blue circles,
357 HCQ-treated animals; PS, prophylaxis; TS, treatment.

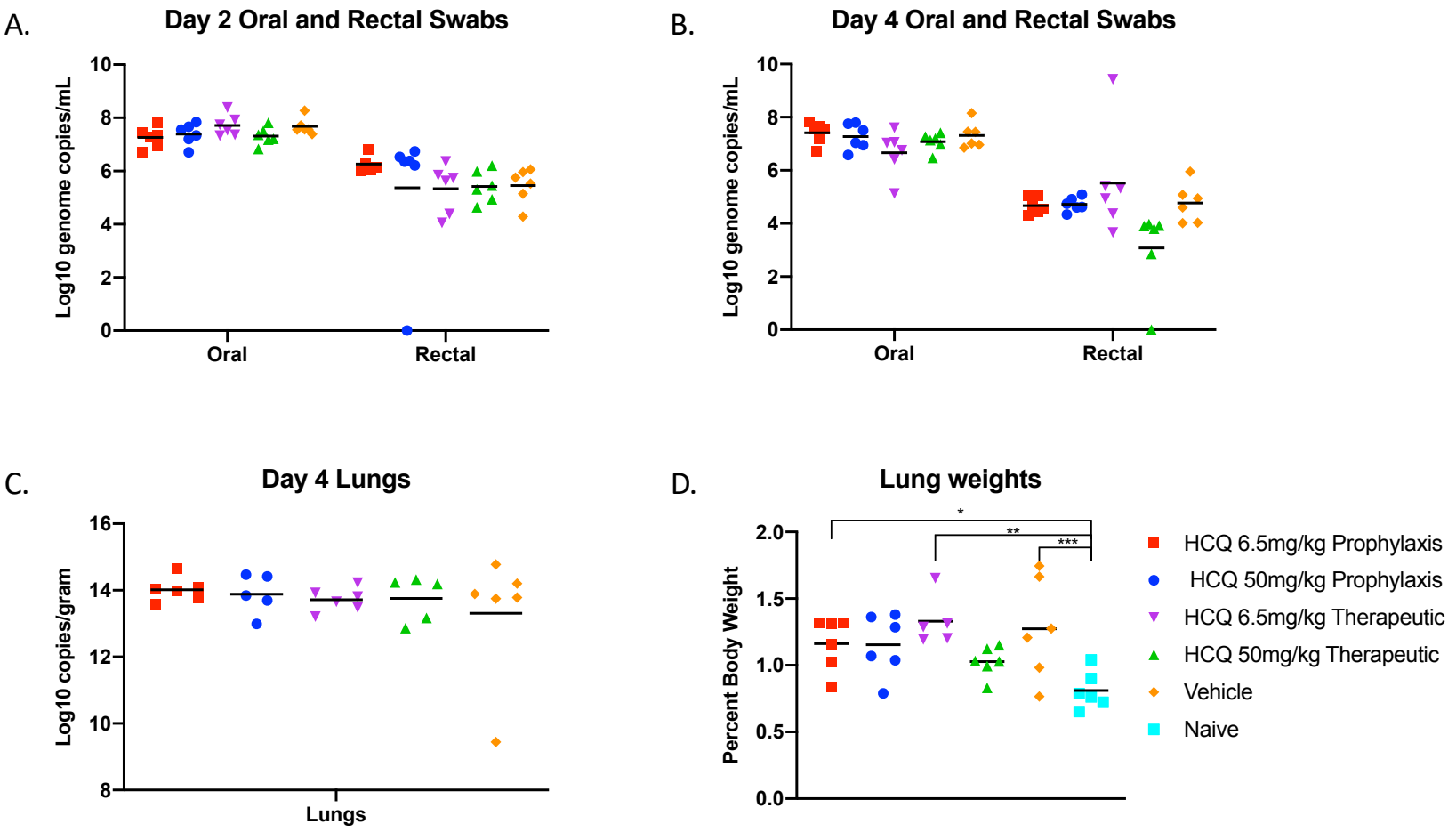
358 **Figure 3: Rhesus macaque model – viral loads in lower and upper respiratory tract.**
359 Macaques were infected with SARS-CoV-2 as described in the legend of Figure 2. Swab
360 samples (nasal and oropharyngeal) and bronchioalveolar lavage (BAL) were collected at all or
361 indicated examination time points. Viral loads were determined by qRT-PCR as genome copies.
362 (A and B) Nasal swabs. (C and D) Oropharyngeal swabs. (E and F) Bronchioalveolar lavage
363 (BAL). No statistical significance was found among the groups presented in (A) to (F). Multiple
364 t tests were used to analyze data and no significant difference was found. *Note:* red squares,
365 vehicle-treated animals; blue circles, HCQ-treated animals; PS, prophylaxis; TS, treatment.

366 **Figure 4: Rhesus macaque model – gross and histopathology.** Macaques were infected with
367 SARS-CoV-2 as described in the legend of Figure 2. Animals were euthanized on day 7 post-
368 infection for gross pathology and histopathology. (A and B) Gross pathology with consolidated
369 lower left lung lobe and area of post-mortem-BAL in the lower right lung lobe (asterisk). (C)
370 Hematoxylin and eosin (H&E) staining revealed multifocal, minimal to moderate, interstitial
371 pneumonia frequently centered on terminal bronchioles. Alveolar edema and fibrin with
372 formation of hyaline membranes was only seen in lungs with moderate changes. Multifocal
373 perivascular infiltrates of small numbers of lymphocytes that form perivascular cuffs. The left
374 panels show areas of unaffected lung tissue. *Note:* PS, prophylaxis; TS, treatment.

375 **Figure 5: Rhesus macaque model – viral loads in respiratory tissues.** Macaques were
376 infected with SARS-CoV-2 as described in the legend of Figure 2. Animals were euthanized on
377 day 7 post-infection for viral tissue load determination performed by qRT-PCR (genome copies)

and virus isolation (infectious virus). (A) Viral loads in lower and upper respiratory tissues and mediastinal lymph nodes for the prophylaxis study arm (PS). Virus isolation is indicated in numbers on top (n/5). (B) Viral lung loads (PS). All lung lobe genome copy data were combined. (C) Viral loads in lower and upper respiratory tissues and mediastinal lymph nodes for the treatment study arm (TS). Virus isolation frequency (number of animals per group) is indicated at top (n/5). (D) Viral lung loads (TS). All lung lobe genome copy data were combined. No statistical significance was found among groups presented in parts (A) to (D). A linear model was used to analyze viral RNA levels in tissues and lung lobes. No significant difference was found between groups in either study. *Note:* red squares, vehicle-treated animals; blue circles, HCQ-treated animals; PS, prophylaxis; TS, treatment.

Figure 1


A.
Day 2 Oral and Rectal Swabs
Log10 genome copies/mL
10
8
6
4
2
0
Oral
Rectal
B.
Day 4 Oral and Rectal Swabs
Log10 genome copies/mL
10
8
6
4
2
0
Oral
Rectal
C.
Day 4 Lungs
Log10 copies/gram
16
14
12
10
8
Lungs
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2.0
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**
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HCQ 6.5mg/kg Prophylaxis
HCQ 50mg/kg Prophylaxis
HCQ 6.5mg/kg Therapeutic
HCQ 50mg/kg Therapeutic
Vehicle
Naive

Figure 2

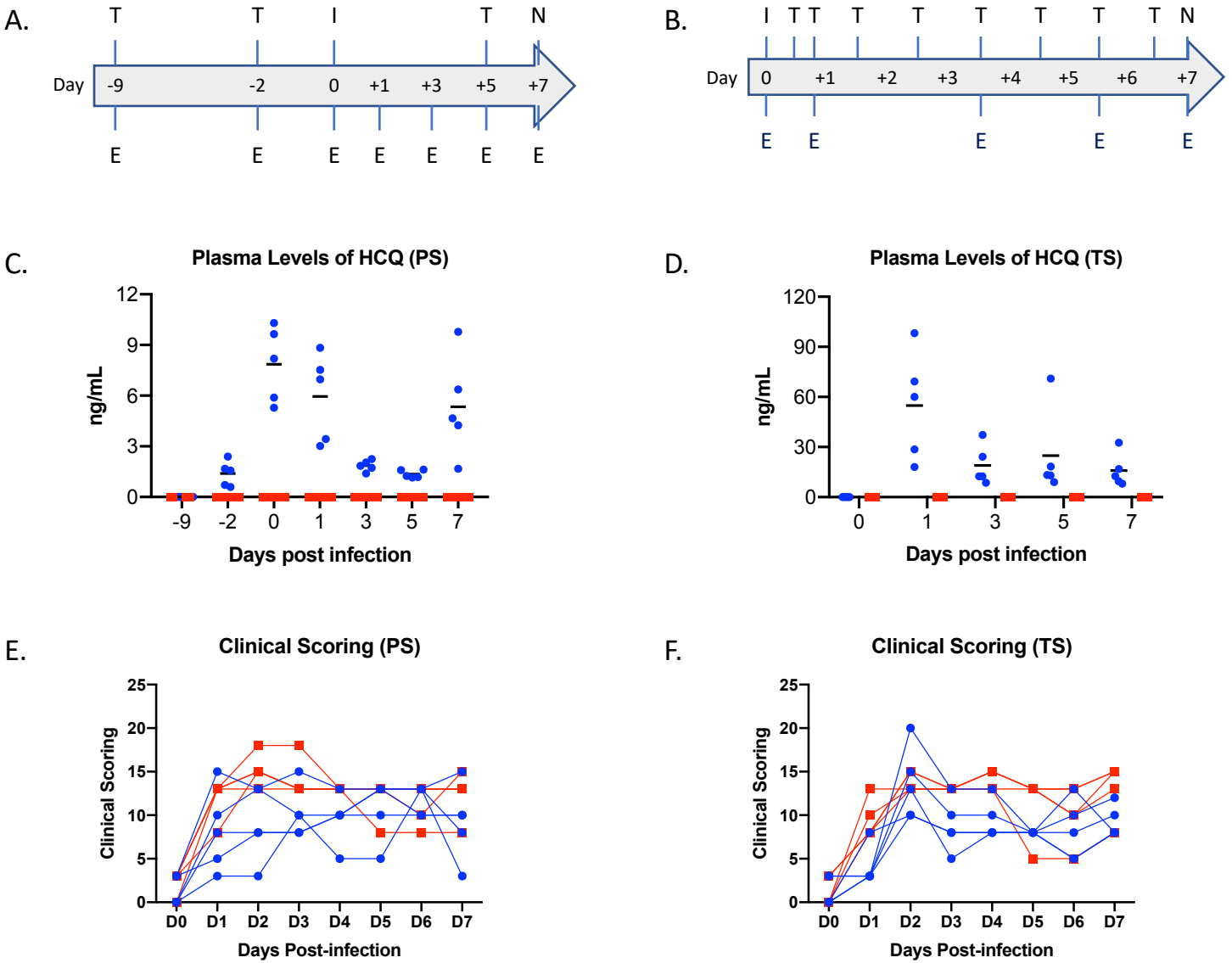

A.
T T I T N
Day -9 -2 0 +1 +3 +5 +7
E E E E E E E
B.
I T T T T T T T T N
Day 0 +1 +2 +3 +4 +5 +6 +7
E E E E E
C.
Plasma Levels of HCQ (PS)
ng/mL
12
9
6
3
0
-9 -2 0 1 3 5 7
Days post infection
D.
Plasma Levels of HCQ (TS)
ng/mL
120
90
60
30
0
0 1 3 5 7
Days post infection
E.
Clinical Scoring (PS)
Clinical Scoring
25
20
15
10
5
0
D0 D1 D2 D3 D4 D5 D6 D7
Days Post-infection
F.
Clinical Scoring (TS)
Clinical Scoring
25
20
15
10
5
0
D0 D1 D2 D3 D4 D5 D6 D7
Days Post-infection

Figure 3

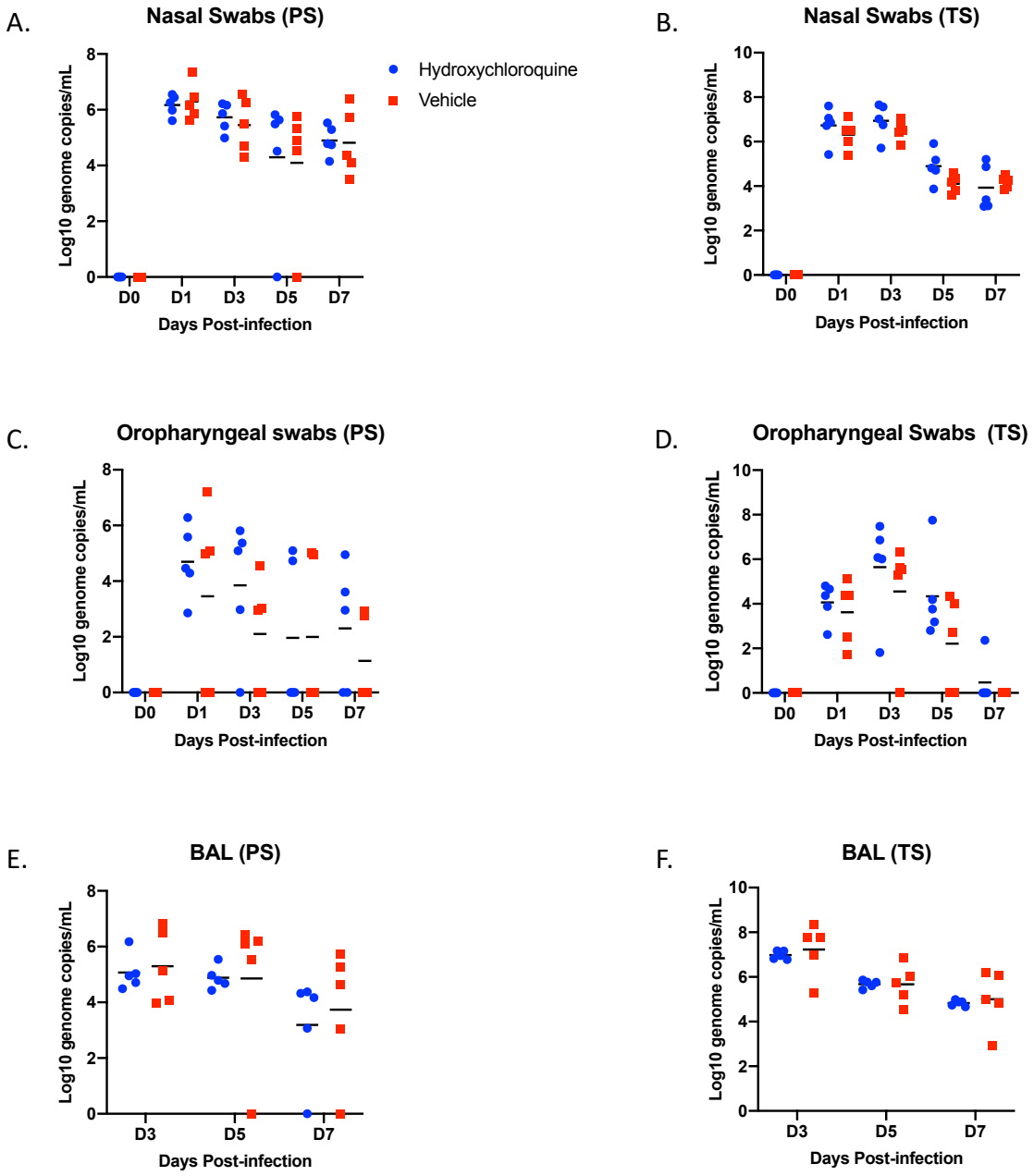

A.
Nasal Swabs (PS)
Hydroxychloroquine
Vehicle
Log10 genome copies/mL
Days Post-infection
D0
D1
D3
D5
D7
B.
Nasal Swabs (TS)
C.
Oropharyngeal swabs (PS)
D.
Oropharyngeal Swabs (TS)
E.
BAL (PS)
F.
BAL (TS)

Figure 4

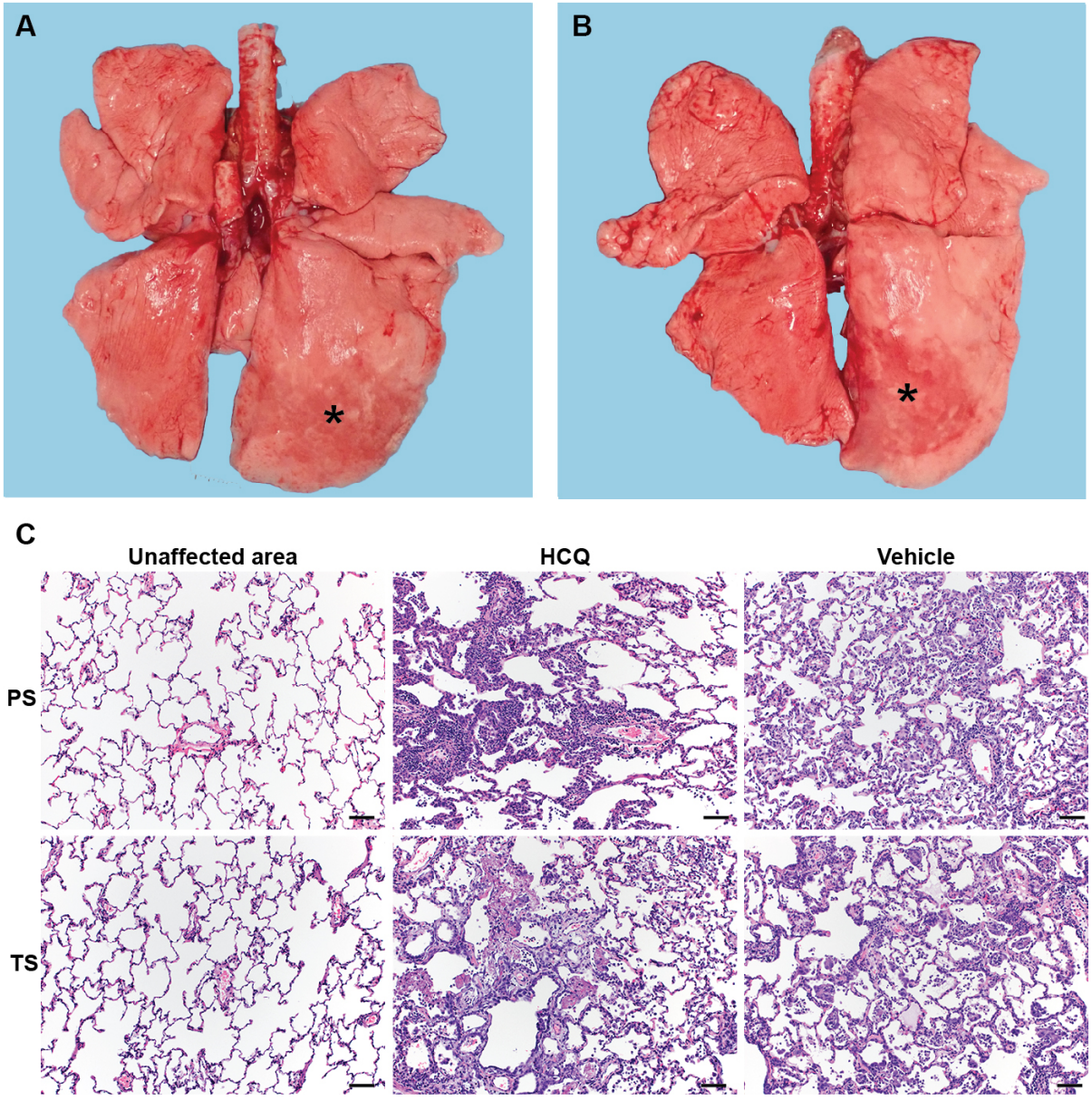
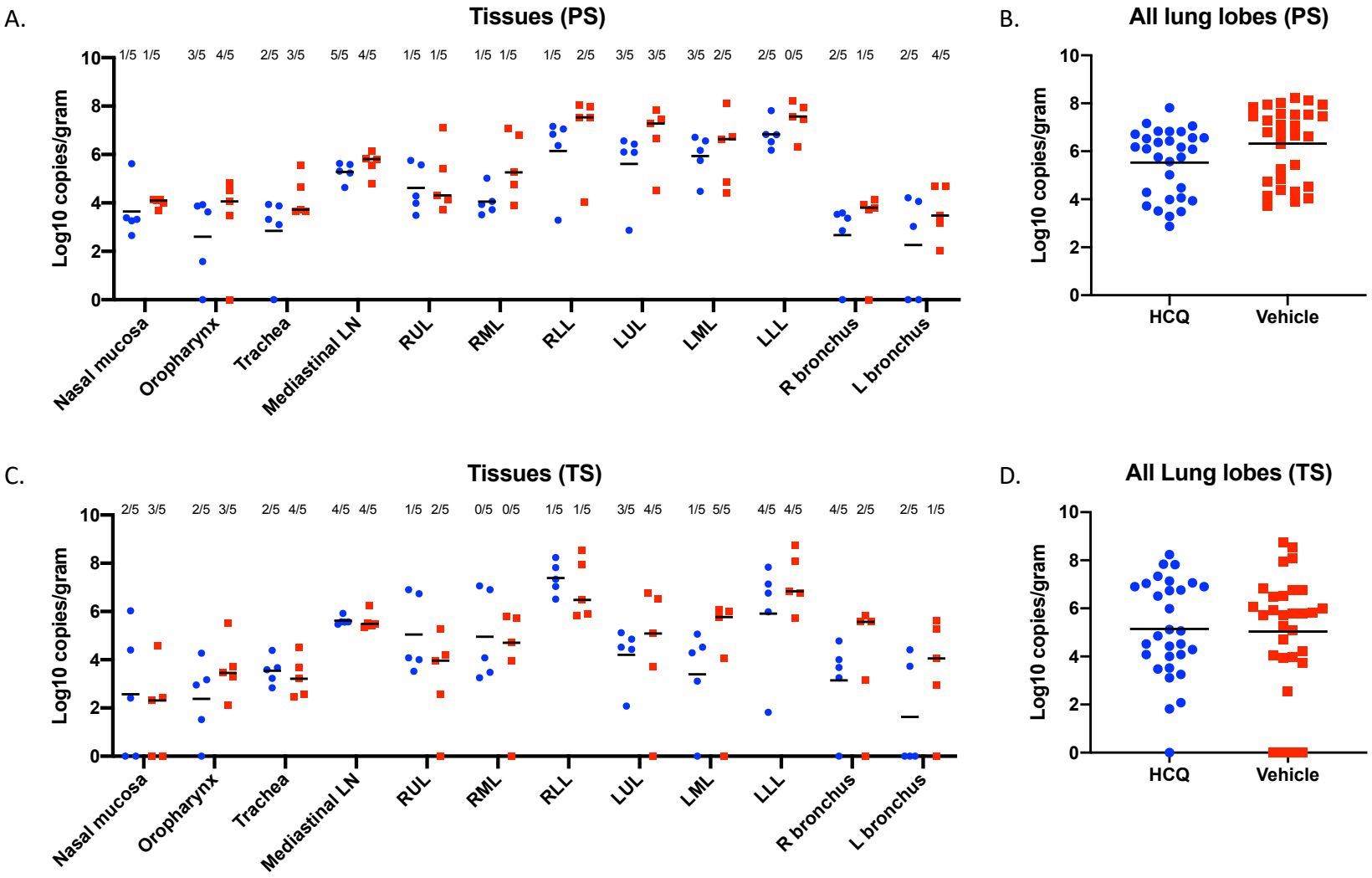


Figure 5


A.
Tissues (PS)
Log10 copies/gram
1/5 1/5
3/5 4/5
2/5 3/5
5/5 4/5
1/5 1/5
1/5 1/5
1/5 2/5
3/5 3/5
3/5 2/5
2/5 0/5
2/5 1/5
2/5 4/5
Nasal mucosa
Oropharynx
Trachea
Mediastinal LN
RUL
RML
RLL
LUL
LML
LLL
R bronchus
L bronchus
B.
All lung lobes (PS)
Log10 copies/gram
HCQ
Vehicle
C.
Tissues (TS)
Log10 copies/gram
2/5 3/5
2/5 3/5
2/5 4/5
4/5 4/5
1/5 2/5
0/5 0/5
1/5 1/5
3/5 4/5
1/5 5/5
4/5 4/5
4/5 2/5
2/5 1/5
Nasal mucosa
Oropharynx
Trachea
Mediastinal LN
RUL
RML
RLL
LUL
LML
LLL
R bronchus
L bronchus
D.
All Lung lobes (TS)
Log10 copies/gram
HCQ
Vehicle